Package 'octopus'

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2 auto.library

auto.library

Load package, install automatically if missing.

Description

```
Load package, install highest version from bioconductor or rstudio if missing.
```

Usage

```
auto.library(package)
```

Arguments

package the name of a package

Value

invisible TRUE

Examples

}

```
{
  auto.library(stringr) #use package name
  auto.library("stringr") #use package name string
  auto.library("package_no_exist") # Print warning message when package is not found

tutorial("auto.library") # print tutorial to console
```

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create.info

Print info with timestamp

Description

Print debug message, progress reports... with timestamp.

- · Default on
- Turn off by info.enable(FALSE)
- Turn on by info.enable(TRUE)

Usage

```
create.info(prefix = "")
```

Arguments

prefix prefix of message.

Value

object of class info

```
info.self <- create.info() #create object of info class
print(info.self,"print message without prefix.")

info.self <- create.info("tutorial.info") #create object of info class with "tutorial.info" as prefix
print(info.self,"print message with prefix, use identifier for prefix usually.")

# turn it off
info.enable(FALSE) #turn information printing off all together
print(info.self,"will print nothing from now on.")
print(info.self,"conform print nothing.")

# turn it on
info.enable(TRUE) #turn information printing on all together
print(info.self,"start printing again.")</pre>
```

octopus.glm.nb

```
tutorial("info") # print tutorial to console
```

}

octopus.glm.nb

Fit a Negative Binomial Generalized Linear Model

Description

Fit glm.nb, add anova to result, add lsm to result.

Before fitting, data and factors will be merged by row.names, mismatching rows will be removed.

If formula start with ~, will fit model with each data column of data as respond y.

Usage

```
octopus.glm.nb(formula, data, factors, specs4lsmeans = NULL)
```

Arguments

formula formula

data normalized reads, expecting gene as column, sample as row factors factors, expecting factor name as column, sample as row

specs41smeans specs for Ismeans

Value

model if formula has respond y; flattened anova table, lsm table and se table if formula starts with \sim .

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Examples

}

```
{
    data(sample_keys) # facotrs
  for(cur_col in colnames(sample_keys)){ sample_keys[,cur_col] <- as.factor(sample_keys[,cur_col]) }</pre>
    data(sample_reads) # reads
    # reads normalization
   norm.reads <- octopus.normalize(sample_reads)</pre>
   print(row.names(sample_keys))
   print(row.names(norm.reads))
    # transpose it to meet input requirements of founction octopus.glm.nb()
    norm.reads <- data.frame(t(norm.reads))</pre>
    print(row.names(norm.reads))
    # fit model with first gene column, column name is Bcin01g00040.1
  result <- octopus.glm.nb(Bcin01g00040.1 ~ Experiment + Experiment/GrowingFlat + Experiment/GrowingFlat/AgarFla
                              , norm. reads
                              ,sample_keys)
    print(result$anova)
    # calculate LSMean
    result$lsmeans <- lsmeans(result,~ Isolate | HostGenotype)</pre>
    summary(result$lsmeans)
    # fit model and pull LSMean in one call
  result <- octopus.glm.nb(Bcin01g00040.1 ~ Experiment + Experiment/GrowingFlat + Experiment/GrowingFlat/AgarFla
                              ,norm.reads
                              ,sample_keys
                              ,specs4lsmeans = ~ Isolate | HostGenotype)
    print(result$anova)
    summary(result$lsmeans)
    # iterate through all data
  result <- octopus.glm.nb( ~ Experiment + Experiment/GrowingFlat + Experiment/GrowingFlat/AgarFlat + Isolate + Ho
                               ,norm.reads
                               ,sample_keys
                               ,specs4lsmeans = ~ Isolate | HostGenotype)
   print(result$anova)
   print(result$1sm)
   print(result$se)
```

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octopus.normalize

Normalize

Description

NA will be treated as 0

Usage

```
octopus.normalize(x, dist_dir = "results/")
```

Arguments

```
x number matrix, expecting gene as row, sample as column dist_dir
```

Value

number matrix

See Also

octopus.glm.nb

Examples

```
tutorial("octopus.glm.nb") # to print tutorial
```

octopus.short_reads

Extract Short Reads

Description

Extract short reads from seq_files using bowtie.

Output 2 files, dist_dir/orig.reads.csv and dist_dir/orig.reads.info.csv

Remove dist_dir/reusing/ folder if don't want reuse data from that folder

Setup ./tmp/ as an RAM disk folder will avoid lots of disk IOs, speed things up and protect you SSD.

Example for linux:

• cd to curent folder.

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• create ram disk in console : mount -t tmpfs -o size=4g tmpfs ./tmp/

Example for mac:

- cd to curent folder.
- mkdir -p tmp
- sudo mount -t tmpfs -o size=4096M tmpfs ./tmp/

For windows, there are a number of RAM disk softerwares you can use.

Usage

```
octopus.short_reads(seq_files, references, ..., type = "single",
 dist_dir = "results/")
```

Arguments

seq_files Sequencing files, accepts .fastq or .gz format for files.

If seq_files is a List or Vector, index of seq_files is assumed to be sampe

names of sequencing files.

If seq_files is a data. frame, row.names is assumed to be sampe names of sequencing files, the first column is assumed to be sequencing files, when type is paired the second column is assumed to be the second mate pair sequences.

references A comma-separated list of FASTA files containing the reference sequences to

be aligned to

Additional arguments to be passed on to the binaries. See ... of bowtie

Could be one of c("single", "paired", "crossbow"). type

If single, the input sequences are interpreted as single reads.

If paired, they are supposed to be mate pair reads.

If crossbow, they are considered to be Crossbow-style reads.

dist_dir folder for result file orig.reads.csv and orig.reads.info.csv 8 octopus.short_reads

```
{
        # single
        references <- "seq_data/cdna/Arabidopsis_thaliana.TAIR10.25.cdna.all.fa"
     seq_files <- data.frame(seq_file=c("seq_data/1_AACGTGAT_L003_R1_001.fastq.gz","seq_data/1_AACGTGAT_L007_R1_0
                                                          ,sample_name=c("sample1","sample2","sample3")
                                                          ,stringsAsFactors = FALSE)
        row.names(seq_files) <- seq_files$sample_name</pre>
        octopus.short_reads(seq_files,references
                                                  ,p=3 # number of alignment threads to launch
                                                  ,`phred33-quals`=TRUE # input quals are Phred+33
                                                  ,t=TRUE # print wall-clock time taken by search phases
                                                  ,quiet=TRUE # print nothing but the alignments
                                                  ,trim5=10 # trim <int> bases from 5' (left) end of reads
        )
        # paired
        references <- "seq_data/cdna/Arabidopsis_thaliana.TAIR10.25.cdna.all.fa"
     seq\_files <- \ data.frame (seq\_file=c("seq\_data/A9\_S1\_L001\_R1\_001.fastq.gz", "seq\_data/xxx\_R1\_001.fastq.gz", "seq\_data/xxx_R1\_001.fastq.gz", "seq_data/xxx_R1\_001.fastq.gz", "seq_data/xxx_R
                                      ,seq_file_pair=c("seq_data/A9_S1_L001_R2_001.fastq.gz","seq_data/xxx_R2_001.fastq.gz","seq_
                                                          ,sample_name=c("sample1","sample2","sample3")
                                                          ,stringsAsFactors = FALSE
        row.names(seq_files) <- seq_files$sample_name</pre>
        octopus.short_reads(seq_files,references
                                                  ,p=3 # number of alignment threads to launch
                                                  , `phred33-quals`=TRUE # input quals are Phred+33
                                                  ,t=TRUE # print wall-clock time taken by search phases
                                                  ,quiet=TRUE # print nothing but the alignments
                                                  ,trim5=10 # trim <int> bases from 5' (left) end of reads
                                 # ,y=TRUE # more sensitive but much slower, see http://bowtie-bio.sourceforge.net/manual.shtml#bo
                                                  ,type="paired"
                                                  ,dist_dir="results_paired/"
       )
        # multiple referencing files
     references <- c("seq_data/ref_sequences/Botrytisfusarivirus1.txt", "seq_data/ref_sequences/BotrytisHypovirus
     seq_files <- data.frame(seq_file=c("seq_data/1_AACGTGAT_L003_R1_001.fastq.gz","seq_data/1_AACGTGAT_L007_R1_0
                                                          ,sample_name=c("sample1", "sample2", "sample3")
                                                          ,stringsAsFactors = FALSE)
        row.names(seq_files) <- seq_files$sample_name</pre>
        octopus.short_reads(seq_files,references
                                                  ,p=3 # number of alignment threads to launch
                                                  , 'phred33-quals'=TRUE # input quals are Phred+33
                                                  ,t=TRUE # print wall-clock time taken by search phases
```

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```
,quiet=TRUE # print nothing but the alignments
,trim5=10 # trim <int> bases from 5' (left) end of reads
)
```

slice

Slice list into sublists by n

Description

Slice list into sublists by n

Usage

```
slice(x, n)
```

Arguments

```
x list
```

n desired length of sublist

Value

A list of sublists

```
{
    x <- 5:505
    slice(x,100) # 5 sublists
    slice(x,99) # 6 sublists
    slice(x,101) # 4 sublists</pre>
```

10 tutorial

```
tutorial("slice") # print tutorial to console
}
```

tutorial

Print Tutorial

Description

Print tutorial for function/class and return tutorial code as a function. It is a generic function.

Usage

```
tutorial(topic)
```

Arguments

topic

tutorial topic, it is usually a function name.

Value

tutorial code as a function

```
# access tutorial code for class info
tutorial("info")  # use class name string
tutorial(create.info()) # use object

tutorial(create.info)  # use function name
tutorial("create.info") # use function name string

tutorial_func <- tutorial("info")
tutorial_func() # run tutorial code, suggest read it first</pre>
```

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```
tutorial("tutorial") # print tutorial to console
```

}

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