



# The soil microbiome – from metagenomics to metaphenomics

Janet K Jansson<sup>1</sup> and Kirsten S Hofmockel<sup>1,2</sup>

Soil microorganisms carry out important processes, including support of plant growth and cycling of carbon and other nutrients. However, the majority of soil microbes have not yet been isolated and their functions are largely unknown. Although metagenomic sequencing reveals microbial identities and functional gene information, it includes DNA from microbes with vastly varying physiological states. Therefore, metagenomics is only predictive of community functional potential. We posit that the next frontier lies in understanding the metaphenome, the product of the combined genetic potential of the microbiome and available resources. Here we describe examples of opportunities towards gaining understanding of the soil metaphenome.

## Addresses

<sup>1</sup> Earth and Biological Sciences Directorate, Pacific Northwest National Laboratory, Richland, WA 99352, United States

<sup>2</sup> Department of Ecology, Evolution and Organismal Biology, Iowa State University, Ames, IA 50010, United States

Current Opinion in Microbiology 2018, 43:162–168

This review comes from a themed issue on **Environmental microbiology**

Edited by **Alberto Scoma** and **Julia Vorholt**

For a complete overview see the [Issue](#) and the [Editorial](#)

Available online 15th February 2018

<https://doi.org/10.1016/j.mib.2018.01.013>

1369-5274/© 2018 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

## Introduction

Soil microbial communities carry out key ecosystem services that are vital for life on our planet, including cycling of carbon (C) and other nutrients and sustaining plant growth. Unfortunately, many beneficial functions carried out by the soil microbiome are currently threatened due to changing climate and precipitation patterns, soil degradation and poor land management practices [1]. Recently there has been increased interest in manipulation of soil microbiomes to restore ecosystem function [2]. The opportunity for managing ecosystem services and bioprospecting soil microbial metabolism will be possible with a greater comprehension of how soil microbiomes interact under different conditions. Exploration and management of soil microbiomes remains a daunting task, however, because the majority of soil microbes have not

yet been isolated and molecular details underlying their functions are largely cryptic.

Here we will focus on one of the biggest enigmas facing soil microbiologists; namely understanding how soil C is transformed by soil microbes. Ultimately, the soil microbiome, together with plants, determines whether C is released to the atmosphere as CO<sub>2</sub> or CH<sub>4</sub>, or retained in soil [3]. Although molecular interactions between microbial species and their environment strongly influence the fate of soil C, details of these interactions are largely unknown. Unlike other microbial habitats (e.g. gut, water), both microbial communities and substrates in the soil are highly diverse and subject to physical protection and chemical stabilization [4]. Therefore, identifying and measuring the network of active microbial metabolic interactions in soils requires approaches adapted to the heterogeneous soil environment. Advancing soil microbiome research thus depends on identifying dominant heterotrophic pathways for C metabolism and how microbial physiology influences the relative importance of C cycling pathways in response to environmental conditions. Furthermore, as one of the most diverse habitats on the planet, soil microbiomes provide rich opportunities to commandeer metabolic interactions for industrial applications, such as biofuel, and mining for novel bio-products, including new antibiotics [5].

## From soil metagenomes to metaphenomes

High throughput sequencing studies have succeeded in illuminating the previously unknown compositions and diversities of soil microbial communities across a variety of soil habitats without the necessity for cultivation [6]. Deep metagenome sequencing has also started to reveal the functional potential of soil communities, for example, genes involved in C cycling [7] and links between community genes and functions [8]. A current challenge is to go beyond predictive understanding of gene function based on the genome/metagenome to understanding of actual functions carried out by the soil microbiome *in situ*. This is especially important in soil environments, where metagenomes include relic DNA extracted from dead and dormant cells [9,10] and DNA that is trapped in biofilms [11]. Even viable cells that are actively growing only regulate gene expression as needed, and not all genes are expressed at any given time. For example, representatives of the *Verrucomicrobia* are often present in soil metagenomes, but in a recent study they were shown to have low levels of gene expression based on metatranscriptome data [12]. Therefore, a soil

metagenome provides an overview of potential microbial function and other methods are needed to determine the actual functions that are carried out by viable and active cells under given environmental conditions.

Here we define the *metaphenome* as the product of expressed functions encoded in microbial genomes (metagenome) and the environment (resources available; spatial, biotic and abiotic constraints). To our knowledge this term has only been reported once previously for microbial communities, when describing metagenomes as genomes of communities with expression through the community 'metaphenome' [13]. The soil metaphenome is dependent on the combined genetic potential encoded by the soil member genomes, the physiological status of the member populations, their access to resources, contact with other organisms and signaling molecules, combined with their genetic capacity to respond to environmental cues. The metaphenome thus encompasses the entire 'omics' field, including the metagenome, metatranscriptome (expressed genes), metaproteome (proteins resulting from translation) and the metabolome (metabolic products) [14]. The soil metaphenome is also ultimately governed by the highly structured soil environment, resulting in a very heterogeneous availability of electron acceptors and redox chemistry, and both strong spatial and temporal variability. Therefore, the soil metaphenome remains a considerable challenge to measure and predict. Here we will review the current state-of-the-science, knowledge gaps and discuss future opportunities for understanding the soil metaphenome.

### Influence of soil structure and connectivity on the soil metaphenome

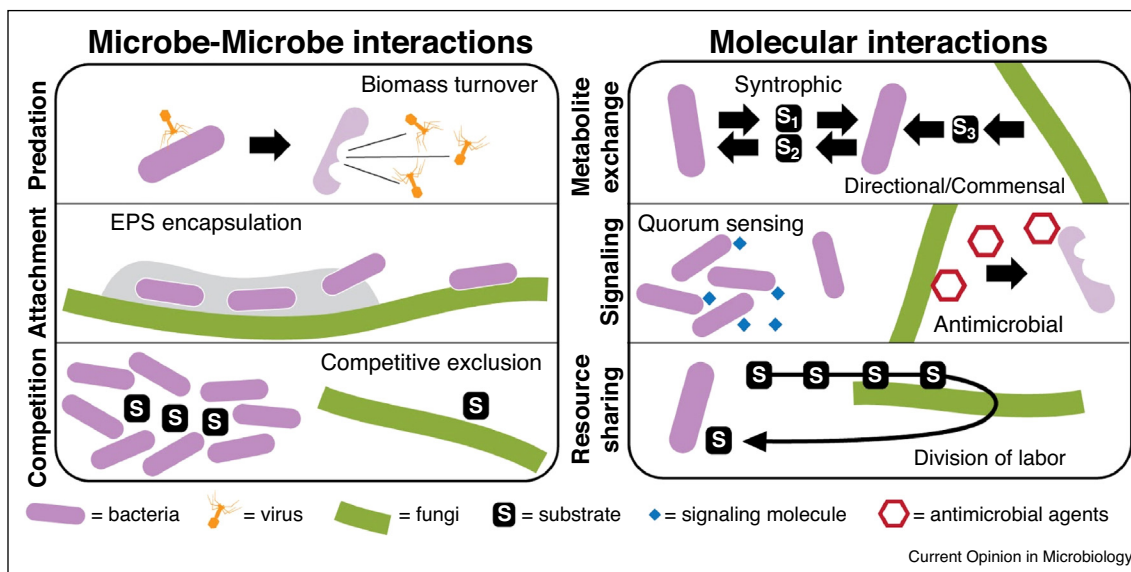
Although we recognize that understanding of metaphenomes is important and relevant for understanding functions of microbial communities in a variety of ecosystems, ranging from water to humans to soil, soil presents unique challenges. For example, physical protection of substrates may prevent their utilization by the resident soil microbiome [15]. Soil is also spatially complex with a highly dynamic and patchy distribution of C and other resources that results in distributed hot spots suitable for growth of microbial consortia, for example within microaggregates or the rhizosphere [16,17]. The spatial constraints imposed upon distinct consortia residing in individual soil microaggregates (~50–200 µm in diameter) presumably constrain the types of cross-species interactions that can occur in a given soil habitat. Soil aggregates have recently been considered to be analogs of evolutionary incubators for soil microbial life [18<sup>•</sup>]. Because they are isolated and tremendously abundant, soil aggregates can allow for massively parallel evolution of distinct microbial consortia. We propose that this spatial isolation could be one of the contributing factors underlying the high microbial diversity found in most soil habitats [6<sup>•</sup>].

Understanding the fine scale distribution of microbes and resources is required to predict species physiology and metabolic interactions among community members, that comprise the collective soil metaphenome. Given that life in soil is concentrated within micro-spatial 'islands', soil microbes have evolved to interact with each other through a variety of mechanisms that deal with spatial constraints [19<sup>•</sup>]. Currently broad scale process measurements, such as soil respiration, mask details of the molecular reactions occurring by interacting members in discrete, spatially isolated soil consortia (Figure 1). For example, as soils become drier, microbial dispersion becomes more limited and microbial life more constrained within physically protected soil pores (Figure 1). Little is known about how resulting subpopulations or consortia distribute metabolic functions among themselves or regulate/signal other populations in response to changing environmental conditions and how this relates to the soil metaphenome.

### Influence of physiological status on the soil metaphenome

The collection of physiological responses of individual microorganisms to the environment results in a community metaphenomic response, including genetic regulation and cell–cell interactions, that underlie which community genes are expressed in response to resource availability. Depending on the step in the chain of expression, different information is obtained about the physiological status of the soil microbiome. Metatranscriptomics captures transient responses to environmental conditions, whereas metaproteomics provides a more stable indication of the overall state of the environment [20]. The responses of individual soil microbes to changes in environmental conditions are highly regulated at the genetic level, resulting in a range of physiological alterations, including shifts in fatty acids making up the cell membrane, production of specific proteins (e.g. heat shock or cold shock proteins), and reduction in respiratory activity [21]. In addition, many metabolic pathways are regulated so that the genes are only transcribed when needed. Sequencing of genes by metagenomics will include genes that are not being transcribed and therefore not translated into proteins. Should conditions change, other genes will be induced, transcribed and translated. For example, prolonged survival under subzero conditions, results in a range of physiological responses across community members [21,22]. Some microbes accumulate C storage reserves and osmolytes as a resource to stay viable over extended periods of low nutrient conditions or low soil moisture levels. Another adaptive response is to decrease genome copy numbers to cope with low nutrients, such as observed during a long-term soil warming experiment [23<sup>•</sup>]. In general, as microbes enter a state of low activity or dormancy, their contribution to the metaphenome decreases, relative to those that maintain an active metabolic state.

Figure 1



Illustrative overview of biotic and environmental factors contributing to the soil metaphenome. A cross section of a field is shown with different soil moisture levels. On the right side, plant growth is constrained due to low soil moisture levels. An example of a measurable phenotype is shown ( $\text{CO}_2$ , corresponding to soil respiration), which is the result of combined metabolic interactions between soil microbes and plants. Call out circles correspond to a microscale view of soil consortia residing in spatially discrete soil aggregates. Connectivity between consortia is determined by the extent of the pore volume that is water filled and available for diffusion of chemical signals and metabolites. Bacterial (purple symbols) interactions within consortia are designated with white arrows. Fungal hyphae (green filaments) may bridge spatially discrete consortia. Soil viruses (orange symbols) also play a yet undefined role in regulating the soil metaphenome. Lower panel illustrates different types of models applicable to defining the soil metaphenome; from left to right: biochemical reaction networks squares correspond to bacterial (purple) or fungal (green) metabolites, interspecies interaction networks, and interkingdom interactions [26\*,36].

### Influence of microbial community interactions on the soil metaphenome

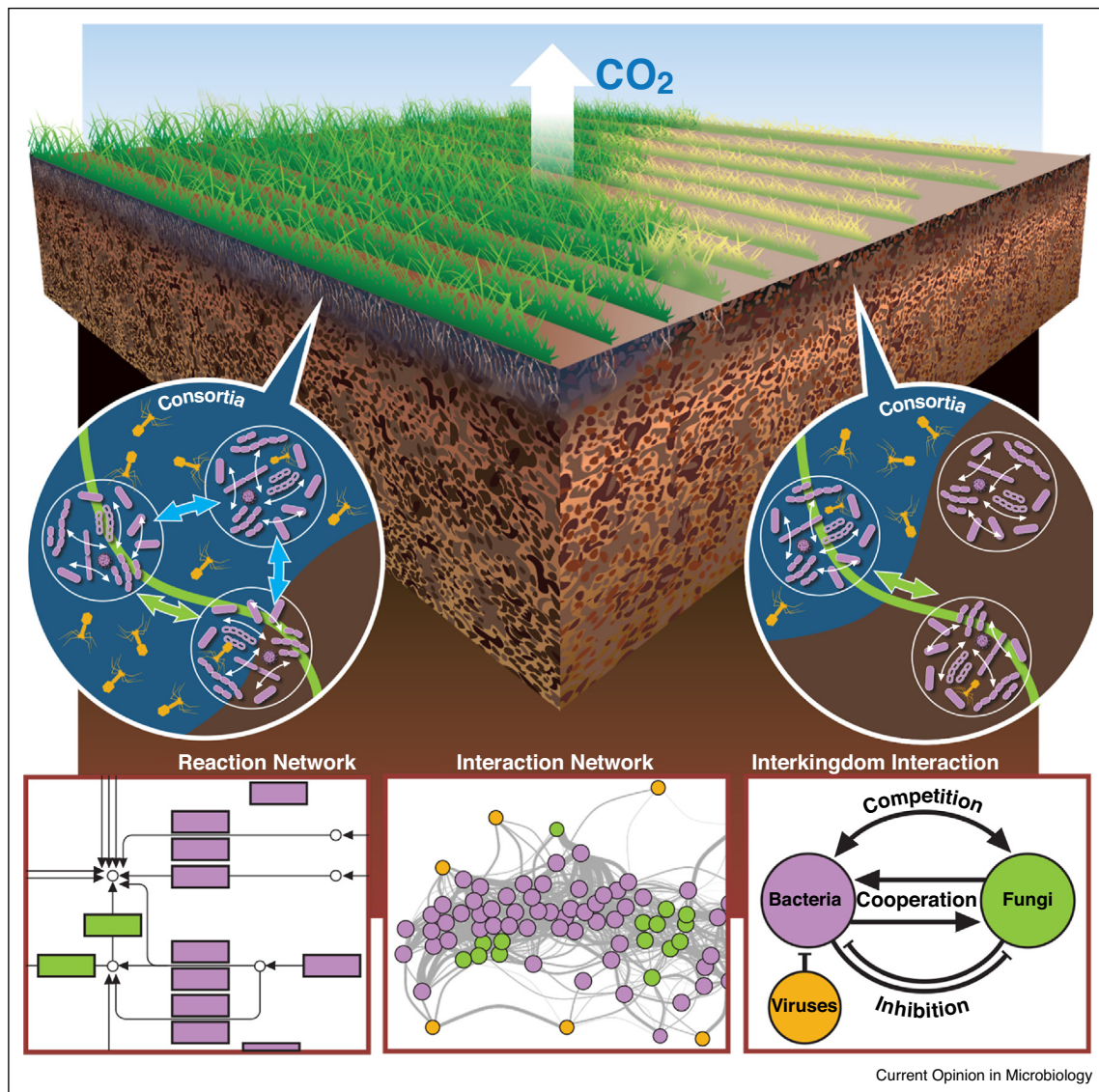
At the community level the soil metaphenome includes the combined metabolic outputs of the community members. An example is cellulose degradation. Cellulose is a complex polymer that is degraded by different microorganisms with complementary metabolic traits. For example, some microbes possess glycoside hydrolases, others have transporters, and so on [24\*]. Different microbial community interactions might affect the fitness of microbes that have all of the enzymes to degrade cellulose to cellobiose to glucose, while others must compete for products of exocellulase enzymes [25]. However, details of soil microbial metabolic interactions during degradation of cellulose and other C compounds, as well as the myriads of other soil processes that result in a given metaphenome, are not well known. Even with increasing access to soil metagenomes, we are still challenged to understand how the physiology of the interacting organisms respond to environmental conditions to define the response surface — the possible metaphenomes — given the genetic potential and range of environmental conditions (temperature, moisture) for a given ecosystem. Interpreting bulk dynamics of microbial genes and gene-products involved in the resulting soil metaphenome thus remains challenging due to high diversity

and complexity that confound our ability to link mechanistic details to emergent properties.

Ecological network theory has been employed to predict species interactions and the stability of simple microbial communities [26\*]. However, the concept of interacting microbial networks has been elusive to test in complex, heterogeneous soil ecosystems. Nutritional interactions in soil involve interconnected metabolic webs between species and across kingdoms [27\*]. These interactions include complex nutritional interactions, involving interconnected metabolic pathways, with cross-feeding and metabolite exchange between species. The types of interactions range from metabolic cooperation between microbes in syntrophic relationships, to competition for access to limiting nutrients (Figure 2). Soil microbes communicate with each other and their environment through a variety of chemical signals [28]. Few studies, however, have determined specific metabolic and signaling interactions between members of soil microbial communities [29], including interactions across trophic levels. This knowledge is important because soil is home to highly diverse and complex communities of organisms, including bacteria, archaea, fungi, virus [30\*] and higher organisms (plants, insects, protozoa, and so on) [31]. Together these different soil organisms interact in trophic



Figure 2



Overview of types of soil microbial community and molecular interactions.

food webs to break down complex organic compounds and exchange nutrients. For soil microbiome research to advance, innovative approaches are required to reveal the details underlying the myriad of interactions carried out by naturally complex soil microbiomes, and the interplay between and within different kingdoms that result in the soil metaphenome.

### Opportunities for the future Untangling the intricate web of metabolic interdependencies

Understanding the complexity of all metabolic interactions that result in measured phenotypes within a single organism is not yet achievable. However, recent advances in genome-enabled predictions, fluxomics (determining

rates of metabolic reactions), and modeling have made great strides towards deciphering specific metabolic pathways in single bacteria [32<sup>•</sup>]. In particular, flux-based analysis (FBA) is a promising approach that uses metabolic models to predict phenotypic responses of microorganisms to different environmental conditions [33]. A genome-enabled approach for prediction of metabolic interdependencies between soil community members is theoretically possible in the near future. For example, soil metagenomes can be explored for genes encoding specific bioactive compounds [34], or specific functional genes [35<sup>•</sup>] based on their distinct sequence signatures. Eventually reconstruction of biochemical reaction networks should be possible from annotated metagenomes, combined with stable isotope-based fluxomics, and

metabolic modeling [36]. When predicted metabolic pathways are incomplete — due to missing or missannotated genome sequence data — gap-filling can be employed to add the missing steps [33]. Gap filling can be aided by knowledge of the metabolite composition of soil, using sensitive mass spectrometry platforms to predict the identities of the metabolites. This combined with new computational approaches have great promise to increase the number of soil metabolite assignments in databases, such as recently demonstrated for human metabolites [37].

An attractive means to achieve increased resolution of metabolic dependencies is through dissection of complex soil microbiomes into discrete functional units, or guilds, responsible for a specific phenotype (cellulose decomposition, methanogenesis, sulfate reduction, and so on). This could be accomplished by combining soil isolates with known metabolic capabilities into ‘synthetic’ communities [38], or by selective enrichment in liquid media containing specific resource combinations. In both cases, the simplified communities should facilitate determination of metabolic exchange between species having specific metabolic capabilities [39,40<sup>\*</sup>]. However, a synthetic community that is built up from isolates loses the naturally adapted interactions between community members. Also, enrichment cultures in liquid medium do not include the spatial constraints inherent to soil microbes that predominantly reside in soil microaggregates [38]. Therefore, a future opportunity for identifying community phenotypes would be to construct naturally evolved and tractable model soil consortia in a soil environment. This would allow the study of metabolic and spatial interactions, and chemical signaling between microorganisms in their natural habitat.

Microfluidics has great potential to enable experimental manipulation of the soil microbiome to determine mechanisms underpinning specific microbial metabolic interactions and to understand and predict the influence of environmental gradients on specific microbial functions and with precise spatiotemporal control [41]. Microfluidics also lends itself to imaging in conjunction with concurrent developments in high resolution imaging platforms. For soil, it is particularly interesting to incorporate spatial heterogeneity into microfluidics, to approximate the complexity of the soil environment. We envision a future where specific interactions between soil microorganisms are visualized in a heterogeneous spatial context and at a micro-relevant scale using microfluidics and imaging. Combined with new modeling approaches, it should be possible to predict self-assembly of multi-species microbial communities on different rough surfaces mimicking a soil environment [42].

#### Interpreting the soil metapenome

Functions carried out by members of the soil microbiome that result in the soil metapenome, are now possible to

assess using techniques such as stable isotope probing (SIP) and multi-omics approaches. For example, stable isotopes can be used to track how specific nutrients are metabolized by interacting members of soil microbiomes and across trophic levels. In one study <sup>13</sup>C-SIP was used to determine succession over time during the assimilation of <sup>13</sup>C-labeled xylose, suggesting that labile C traveled through different trophic levels [43<sup>\*\*</sup>]. By contrast there were fewer changes in the phylogenetic composition of cellulose degraders over time, and they corresponded mainly to abundant, but poorly characterized members of the soil microbiome.

Recent application of metatranscriptomics [44], metaproteomics [22] and metabolomics [45] are also helping to fill gaps in our knowledge about genes that are expressed and/or translated into proteins, and metabolic interactions that are possible under a given soil resource regime. Although there have been some recent advances in use of a multi-omics approach to decipher soil microbial community functions [22], there remain significant challenges to overcome, including functional gene annotation, and extraction and identification of macromolecules (metabolites and proteins). Future advances in mass spectrometry technologies that will facilitate higher throughput, yield and depth of coverage of proteins have enormous potential to open the current bottleneck in soil proteomics [12,14]. Also, as the depth of metagenome sequencing continues to increase, we are seeing much better metagenome assemblies; in particular when combined with long-read sequencing technologies [12]. Along with better assembled metagenomes come a higher number of complete- to near-complete genome bins from soil [12]. A future opportunity thus lies in the use of genome bins as databases for searching soil metaproteomes [46]. The advantages of using genome bins is similar to that of using isolate genomes; that is, entire operons with genes encoding complete pathways and genes encoding regulatory mechanisms are intact and phylogeny can be coupled to function because the entire 16S rRNA gene is on the genome together with the functional genes. By application of comparative genomics approaches it should be possible to predict phenotypes directly from the genome bins [47<sup>\*</sup>] without the necessity to cultivate the represented species. Expression data (transcripts and/or proteins) can then be mapped to the binned genomes to determine their phenotypes [46]. The collective binned genomes, with expression data, has potential to illuminate details of the soil metapenome.

#### Conclusions

Individual microbial phenotypes, including the combined metabolic outputs of the community members, together generate the higher scale outcomes of the soil metapenome. Interpreting bulk dynamics of microbial genes and gene-products involved in the soil metapenome amidst the high diversity and complexity of soil microbiomes

requires linking mechanistic details to emergent properties. Advances in genome-enabled predictions, fluxomics, and modeling, combined with metabolomics, SIP and imaging technologies, have great promise to identify and track the exchange of signaling molecules and metabolites among soil organisms, which will enable transitioning from the metagenome to the metaphenome. This knowledge is important for prediction of the impacts of environmental perturbations on key functions carried out by the soil microbiome and will enable development of new approaches for optimizing soil carbon cycling, managing nutrient transport, and sustaining crop production.

## Acknowledgements

This research was supported by the Department of Energy Office of Biological and Environmental Research (BER) and is a contribution of the Scientific Focus Area 'Phenotypic response of the soil microbiome to environmental perturbations'. PNNL is operated for the DOE by Battelle Memorial Institute under Contract DE-AC05-76RLO1830. Ted Tanasse and Hans Bernstein (both at PNNL) are acknowledged for their help with the figure illustrations.

## References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Amundson R, Berhe AA, Hopmans JW, Olson C, Sztein AE, Sparks DL: **Soil and human security in the 21st century**. *Science* 2015, **348**:1261071-1-1261071-6.
2. Calderon K, Spor A, Breuil M-C, Bru D, Bizouard F, Violle C, Barnard RL, Philippot L: **Effectiveness of ecological rescue for altered soil microbial communities and functions**. *ISME J* 2017, **11**:1-12.
3. Crowther TW, Todd-Brown KEO, Rowe CW, Wieder WR, Carey JC, Machmuller MB, Snoek BL, Fang S, Zhou G, Allison SD et al.: **Quantifying global soil carbon losses in response to warming**. *Nature* 2016, **540**:104-108.
4. Sollins P, Homann P, Caldwell BA: **Stabilization and destabilization of soil organic matter: mechanisms and controls**. *Geoderma* 1996, **74**:65-105.
5. Ling LL, Schneider T, Peoples AJ, Spoering AL, Engels I, Conlon BP, Mueller A, Schaberle TF, Hughes DE, Epstein S et al.: **A new antibiotic kills pathogens without detectable resistance**. *Nature* 2015, **22**:455-459.
6. Thompson LR, Sanders JG, McDonald D, Amir A, Ladau J, Locey KJ, Prill RJ, Tripathi A, Gibbons SM, Ackermann G et al.: **The Earth Microbiome Project Consortium: A communal catalogue reveals Earth's multiscale microbial diversity**. *Nature* 2017 <http://dx.doi.org/10.1038/nature24621>.  
Provides extensive microbiome comparison across diverse habitats on Earth, including a range of soil habitats.
7. Howe A, Williams RJ, Yang F, Meyer F, Hofmockel KS: **Characterization of core carbon cycling genes in fertilized prairie soils**. *PLoS One* 2016, **11**:e0166578 1-14.
8. Hartman WH, Ye R, Horwath WR, Tringe SG: **A genomic perspective on stoichiometric regulation of soil carbon cycling**. *ISME J* 2017, **11**:2652-2665.
9. Carini P, Marsden PJ, Leff JW, Morgan EE, Strickland MS, Fierer N: **Relic DNA is abundant in soil and obscures estimates of soil microbial diversity**. *Nat Microbiol* 2016, **19**:16242.
10. Emerson JB, Adams RI, Román CM, Brooks B, Coil DA, Dahlhausen K, Ganz HH, Hartmann EM, Hsu T, Justice NB, Paulino-Lima IG: **Schrödingers microbes: tools for distinguishing the living from the dead in microbial ecosystems**. *Microbiome* 2017, **5**:86.
11. Flemming HC, Wingender J: **The biofilm matrix**. *Nat Rev Microbiol* 2010, **8**:623-633.
12. White RA III, Bottos EM, Chowdhury TR, Zucke JD, Brislawn CJ, Nicora CD, Fansler SJ, Glaesemann KR, Glass K, Jansson JK: **Moleculo long-read sequencing facilitates assembly and genomic binning from complex soil metagenomes**. *mSystems* 2016, **1**:1-15.
13. Doolittle WF, Zhaxybayeva O: **Metagenomics and the units of biological organization**. *BioScience* 2010, **60**:102-112.
14. Jansson JK, Baker ES: **A multi-omic future for microbiome studies**. *Nat Microbiol* 2016, **1**:1-3.
15. Luo Z, Baldock J, Wang E: **Modelling the dynamic physical protection of soil organic carbon: Insights into carbon predictions and explanation of the priming effect**. *Global Change Biol* 2017, **23**:5273-5283.
16. Bach EM, Williams RJ, Hargreaves SK, Yang F, Hofmockel KS: **Greatest soil microbial diversity found in micro-habitats**. *Soil Biol Biochem* 2018, **218**:217-226.
17. O'Brien SL, Jastrow JD: **Physical and chemical protection in hierarchical soil aggregates regulates soil carbon and nitrogen recovery in restored perennial grasslands**. *Soil Biol Biochem* 2013, **61**:1-3.
18. Rillig MC, Muller LAH, Lehmann A: **Soil aggregates as massively concurrent evolutionary incubators**. *ISME J* 2017, **11**:1943-1948.  
Present a concept for evolution in soil aggregates; considering aggregates as a compartmentalized microbial incubator and present a hypothesis that this will lead to high levels of soil microbial diversity.
19. Cordero OX, Datta MS: **Microbial interactions and community assembly at microscales**. *Curr Opin Microbiol* 2016, **31**:227-234.  
Discuss the importance of microscale spatial organization for resulting microbial community interactions, stability and functional productivity.
20. Myrold DD, Zeglin LH, Jansson JK: **The potential of metagenomic approaches for understanding soil microbial processes**. *Soil Sci Soc Am J* 2013, **78**:3-10.
21. Mackelprang R, Saleska SR, Jacobsen CS, Jansson JK, Tas N: **Permafrost meta-omics and climate change**. *Ann Rev Earth Planet Sci* 2016, **44**:439-462.
22. Hultman J, Waldrop MP, Mackelprang R, David MM, McFarland J, Blazewicz SJ, Harden J, Turetsky MR, McGuire AD, Shah MB et al.: **Multi-omics of permafrost, active layer and thermokarst bog soil microbiomes**. *Nature* 2015, **521**:208-212.
23. DeAngelis KM, Pold G, Topcuoglu BD, van Diepen LTA, Varney RM, Blanchard JL, Melillo J, Frey SD: **Long-term forest soil warming alters microbial communities in temperate forest soils**. *Frontiers Microbiol* 2015, **6**:1-13.  
Found that long-term warming favored bacteria with low numbers of rRNA operons, corresponding to an oligotrophic lifestyle.
24. Lopez-Mondejar R, Zuhlke D, Becher D, Riedel K, Baldrian P: **Cellulose and hemicellulose decomposition by forest soil bacteria proceeds by the action of structurally variable enzymatic systems**. *Sci Rep* 2016, **6**:25279.  
Identified several genes and proteins from forest soil bacterial isolates that are involved in cellulose degradation, including catabolic enzymes and transporters.
25. Woo HL, Hazen TC, Simmons BA, DeAngelis KM: **Enzyme activities of aerobic lignocellulolytic bacteria isolated from wet tropical forest soils**. *Syst Appl Microbiol* 2014, **37**:60-67.
26. Coyte KZ, Schluter J, Foster KR: **The ecology of the microbiome: networks, competition and stability**. *Science* 2015, **350**:663-666.  
Used models to show that cooperation between community members in host-associated microbiomes reduces community stability, whereas competition stabilized communities.
27. Van der Heijden MGA, Hartmann M: **Networking in the plant microbiome**. *PLOS Biol* 2016, **14**:1-9.



Discuss microbial networks and microbial hubs as well as their potential importance for soil fertility and plant health.

28. DeAngelis KM: **Chemical communication connects soil food webs.** *Soil Biol Biochem* 2016, **102**:48-51.
  29. Baron R, Brodie EL, Mayberry-Lewis J, Hummel E, Da Rocha UN, Chakraborty R, Bowen BP, Karaoz U, Cadillo-Quiroz H, Garcia-Pichel F, Northen TR: **Exometabolite niche partitioning among sympatric soil bacteria.** *Nat Commun* 2015, **6**:8289.
  30. Paez-Espino D, Eloie-Fadrosch EA, Pavlopoulos GA, Thomas AD, Huntemann M, Mikhailova N, Rubin E, Ivanova NN, Kyrpides NC: **Uncovering Earth's virome.** *Nature* 2016, **536**:425-430.
- A large number of novel viral sequences were discovered by searching metagenomes for CRISPR sequences. This approach increased the number of known viral genes by approximately 16-fold, including many novel soil viruses. However, the functions of these viruses are largely unknown.
31. Jacoby R, Peukert M, Succurro A, Koprivova A, Kopriva S: **The role of soil microorganisms in plant mineral nutrition – current knowledge and future directions.** *Front Plant Sci* 2017 <http://dx.doi.org/10.3389/fpls.2017.01617>.
  32. Varman AM, He L, Follenfant R, Wu W, Wemmer S, Wrobel SA, Tang YJ, Singh S: **Decoding how a soil bacterium extracts building blocks and metabolic energy from ligninolysis provides road map for lignin valorization.** *PNAS* 2016, **113**:E5802-E5811.
- Determined details of central metabolism carried out by the soil bacterium *Sphingobium* sp. SYK-6 during metabolism of lignin-derived monomers using a combination of <sup>13</sup>C fingerprinting and <sup>13</sup>C-metabolic flux analysis.
33. Cuevas DA, Edirisinghe J, Henry CS, Overbeek R, O'Connell TG, Edwards RA: **From DNA to FBA: how to build your own genome-scale metabolic model.** *Front Microbiol* 2016, **7**:1-11.
  34. Charlop-Powers Z, Owen JG, Reddy BVB, Ternei MA, Brady SF: **Chemical-biogeographic survey of secondary metabolism in soil.** *PNAS* 2014, **111**:3757-3762.
  35. Nelson MB, Martiny AC, Martiny JHB: **Global biogeography of microbial nitrogen-cycling traits in soil.** *PNAS* 2016, **113**:8033-8040.
- Used a trait based approach to determine nitrogen-cycling pathways in soil. They found that the frequency of N-cycling genes varied across different habitats, but that taxa responsible for different steps in the nitrogen cycle were relatively consistent across different geographic locations. This example, illustrates the need for complementary approaches to decipher the functional roles of soil microbial community members that result in a measured phenotype.
36. Henry CS, Bernstein HC, Weisenborn P, Taylor RC, Lee J-Y, Zucker J, Song H-S: **Microbial community metabolic modeling: a community data-driven network reconstruction.** *J Cell Physiol* 2016, **231**:23390-23245.
  37. Metz TO, Baker ES, Schymanski EL, Renslow RS, Thomas DG, Causon TJ, Webb IK, Hann S, Smith RD, Teeguarden JG: **Integrating ion mobility spectrometry into mass spectrometry-based exposome measurements: what can it add and how far can it go?** *Bioanalysis* 2017, **9**:81-98.
  38. Ponomarova O, Patil KR: **Metabolic interactions in microbial communities: untangling the Gordian knot.** *Curr Opin Microbiol* 2015, **27**:37-44.
  39. Romine MF, Rodionov DA, Maezato Y, AOsterman AL, Nelson WC: **Underlying mechanisms for syntrophic metabolism of essential enzyme cofactors in microbial communities.** *ISME J* 2017, **11**:1434-1446.
  40. Romine MF, Rodionov DA, Maezato Y, Anderson LN, Nandhikonda P, Rodionova IA, Carre A, Li X, Xu C, Clauss TRW et al.: **Elucidation of roles for vitamin B12 in regulation of folate, ubiquinone and methionine metabolism.** *PNAS* 2017:E1205-E1214.
- Novel activity based probes were used to determine previously unknown roles of vitamin B12 in a model microbial community.
41. Stanley CE, van der Heiden MGA: **Microbiome-on-a-chip: new frontiers in plant-microbiota research.** *Trends Microbiol* 2017, **25**:610-613.
  42. Wang G, Or D: **Trophic interactions induce spatial self-organization of microbial consortia on rough surfaces.** *Sci Rep* 2014, **4**:6757.
  43. Pepe-Ranney C, Campbell AN, Koechli CN, Berthrong S, Buckley DH: **Unearthing the ecology of soil microorganisms using a high resolution DNA-SIP approach to explore cellulose and xylose metabolism in soil.** *Front Microbiol* 2016, **7**:1-17.
- Results suggest that life-history traits, are important in determining which soil microbes are capable of responding to different <sup>13</sup>C-labeled substrates. The substrates were tracked as they were metabolized through the soil food web. Several uncharacterized bacteria responded to cellulose, whereas most of the taxa that responded to xylose were labeled due to metabolite transfer across trophic levels.
44. Singer E, Wagner M, Woyke T: **Capturing the genetic makeup of the active microbiome in situ.** *ISME J* 2017, **11**:1949-1963.
  45. Baran R, Brodie EL, Mayberry-Lewis J, Hummel E, Nunes Da Rocha U, Chakraborty R, Bowen BP, Karaoz U, Cadillo-Quiroz H, Garcia-Pichel F, Northen TR: **Exometabolite niche partitioning among sympatric soil bacteria.** *Nat Commun* 2015, **6**:8289.
  46. White RA III, Callister SJ, Moore RJ, Baker ES, Jansson JK: **The past, present and future of microbiome analyses.** *Nat Protoc* 2016, **11**:2049-2053.
  47. Brbic M, Piskorec M, Vidulin V, Krisko A, Smuc T, Supek F: **The landscape of microbial phenotypic traits and associated genes.** *Nucleic Acids Res* 2016, **44**:10074-10090.
- Developed ProTraits that uses supervised machine learning algorithms for annotation of microbial phenotypes. ProTraits assigns phenotypes using comparative genomics and pattern associations and was employed to develop a network of microbial phenotypic traits.