# Manual of VirusRecom

# Detecting recombination of viral lineages using information theory

Home page: <a href="https://github.com/ZhijianZhou01/VirusRecom/Zhijian

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#### 1. Introduction

#### 1.1. Background

Recombination is common in viruses. In general, recombination can be divided into two categories: homologous recombination including normal homologous recombination and abnormal homologous recombination, and non-homologous recombination. Homologous recombination occurs between homologous or similar sequences and plays a major role in recombination. The normal homologous recombination was defined as that recombination region is homologous or similar. However, aberrant homologous generally results in sequence deletion or duplication due to the recombination region is not in the homologous region.

Identifying homologous recombinations from highly similar sequences is a challenge, due to the uncertainty of an emerging genomic variance originating from recombination or *in-situ* mutation. Herein, we present VirusRecom, an efficient tool for recombination analysis of viral genome with high similarity based on information theory. The new method evaluates the likelihood of recombination by quantifing recombination contribution using weighted information content (WIC).

#### 1.2. Functions

VirusRecom was written by Python 3, the releases were created by Pyinstaller. The functions are:

- (I) Calculate the recombination contribution of each polymorphic sites.
- (II) Identify the recombination events and potential recombination region with p-value.
  - (III) Scan the potential recombination breakpoint.

#### 2. Download and install

VirusRecom and all the updated versions is freely available for non-commercial user at <a href="https://github.com/ZhijianZhou01/VirusRecom/releases">https://github.com/ZhijianZhou01/VirusRecom/releases</a>. After obtaining the program, users could directly run the program in Windows, MacOS or Linux (Ubuntu 16.04 or more) systems without installation.

In general, the executable file of VirusRecom is located at the folder of Main. Then, just double click the VirusRecom.exe (windows system) or VirusRecom (Linux or MacOS system) to start. If you could not get permission to run VirusRecom on linux system, you could change permissions by chmod -R 777 VirusRecom.

## 3. Description of parameters

#### 3.1. Getting help

VirusRecom is a command line interface program, users can get help documentation of the software by entering VirusRecom -h or VirusRecom --help.

```
usage:
VirusRecom [-h] [-a ALIGNMENT] [-q QUERY] [-l LINEAGE] [-g GAP]
[-m METHOD] [-w WINDOW] [-s STEP] [-mr MAX_REGION]
[-p PERCENTAGE] [-b BREAKPOINT] [-bw BREAKWIN]
[-t THREAD] [-y Y_START]
```

#### **Example of usage:**

(i) If the input-sequence data was not aligned:

```
VirusRecom -q XE.fasta -l Lineage_Dir -g n -m p -w 100 -s 20 -t 2
```

(ii) If the input-sequence data has been aligned:

```
VirusRecom -a alignment.fasta -q XE_ -l lineage_name_list.txt -g n -m p -w 100 -s 20
```

# 3.2. Parameters

Parameters	Description
-h,help	show this help message and exit.
<b>-</b> a	FilePath of an aligned sequence set (*.fasta format) containing all
	sequences used for analysis, then the alignment will be skipped.
	Default is null. If using, name of each sequence in aligned sequence
	set requires containing the mark (a unique string) of the lineage.
<b>-</b> q	FilePath of query lineage (potential recombinant, *.fasta format).
	Note, if the '-a alignment.fasta' has been used, please enter the mark
	(a unique string) of queried recombinant here, such as '- $q$ XE_', not a
	FilePath.
-l	DirPath of reference lineages. One sequence file (*.fasta format) per
	lineage, and each lineage could contain multiple sequences. Note, if
	the '-a alignment.fasta' has been used, please enter a text file
	containing the marks (a unique string) of lineages here, not a DirPath.
-g	Gaps (-) in the alignment were used in analysis? '-g y' means to
	reserve gaps, and '-g n' means to delete gaps.
-m	Scanning method of recombination analysis. '-m p': using
	polymorphic sites only, '-m a': using all the monomorphic sites and
	polymorphic sites.
-w	Number of nt sites per sliding window. Note: if the '-m p' has been
	used, -w refers to the number of polymorphic sites per windows.
-S	Step size for scanning these sites. Note: if the '-m p' has been used,
	-w refers to the number of polymorphic sites per jump.
-mr	The maximum allowed recombination region. Note: if the '-m p'
	method has been used, it refers the maximum number of polymorphic
	sites contained in a recombinant region.

Parameters	Description
-cp	The cutoff threshold of proportion (cp, default was 0.9) for searching
	recombination regions when mWIC/EIC $\geq cp$ , the maximum value
	of <i>cp</i> is 1. For detection in genus level, about 0.5 is recommended.
<i>-b</i>	Whether to run the breakpoint scan of recombination. '-b y': yes, '-b
	n': no. Note: this option only takes effect when '-m p' has been
	specified!
-bw	The window size (polymorphic sites, default is 200) used
	forbreakpoint scan. The step size is fixed at 1. Note: this option only
	takes effect when '-m p -b y' has been specified!
-t	Number of threads used for the multiple sequence alignments (MSA),
	default is 1.
<b>-</b> <i>y</i>	Specify the starting value of the Y axis in the picture, the default is 0.

# 4. Example of usage

# **4.1.** Unaligned input-sequences

VirusRecom owns a pipeline built in to handle unaligned sequences. In this case, multiple sequence alignment is performed by MAFFT (Katoh & Standley, 2013) with alignment strategy of "auto". All the unaligned sequences from the query lineage need to contained in a file (\*.fasta format), such as XE.fasta. For other lineages as reference lineages, the unaligned sequences from each lineage also need to contained in a file, and all the sequence files are placed in the same folder, for example (D:\Test\lineages).

AY.45.fasta

B.1.617.2.fasta

B.1.1.7.fasta

BA.1.fasta

BA.2.fasta

Then, execute the command to detect recombination events, for example.

VirusRecom -q D: \Test\XE.fasta -l D:\Test\ lineages -g n -m p -w 100 -s 20 -t 2

#### 4.2. Aligned input-sequences

Users can also provide an independent aligned sequences dataset which was performed from any other alignment program, including all the sequences from query lineage and reference lineages. Notably, each sequence in the aligned sequences dataset needs to contain a unique mark representing its lineage, for example (XE-others.fasta).

```
>XE hCoV-19/England/PHEC-YYR4GXD/2022
>XE_hCoV-19/England/LSPA-3C834E6/2022
>AY.45_hCoV-19/SouthAfrica/NICD-N18409/2021
-----taaacgaactttaaaatctntgnggctgtcactcggctgcatgcttagtgcactcacgcag
>AY.45 hCoV-19/SouthAfrica/NICD-N18483/2021
>AY.45_hCoV-19/Denmark/DCGC-173261/2021
gatct gtt ctctaaac gaactt taaaat ct gt gt gg ct gt cac te gg ct gcat gct tag t gcac te ac geag \\
>B.1.617.2 hCoV-19/India/KA-CRL-KIMS-245/2021
>B.1.617.2 hCoV-19/Turkey/HSGM-F14312/2022
>B.1.1.7_hCoV-19/Turkey/HSGM-FS7372/2021
>B.1.1.7_hCoV-19/USA/FL-BPHL-6009/2021
>BA.1_hCoV-19/USA/MN-CDC-IBX539664392073/2021
>BA.1_hCoV-19/India/KA-SEQ_8548_S89_R1_001/2021
>BA.1 hCoV-19/Mexico/CHH InDRE FB2199 E08113577401 S11007/2022
>BA.2_hCoV-19/Denmark/DCGC-294641/2021
>BA.2 hCoV-19/Malaysia/MGVI GS0822/2022
```

Then, prepare a text document that labels all the names of reference lineages, for example (lineages.txt).

```
AY.45_
B.1.617.2_
B.1.1.7_
BA.1_
BA.2_
```

It is recommended to include the character "-" in the mark, which ensures that the mark of each lineage is unique. Notably, the mark of query lineage cannot be placed in the lineages.txt.

Then, execute the command to detect recombination events, for example.

```
VirusRecom -a XE-others.fasta -q XE_ -l D:\Test\lineages.txt -g n -m p -w 100 -s 20
```

Note, the mark of query lineage was specified in the command, and it is recommended to include the character "-" in the mark.

## 4.3. Output result

As the Unaligned input-sequences above as an example. The output directory is automatically created and is located under the input directory. There are three sub-directories and the aggregated report (Possible recombination event in XE\_.txt) under the output directory, including run\_record, WICs of sites and WICs of slide\_window.

- (I) In the directory of run\_record, the alignment file created by MAFFT is reserved.

  If -g n is specified, and the Record of deleted gap sites\_\*.txt file containg all the gap sites will be created. Besides, If -m p is specified, and the Record of same sites in aligned sequence\_\*.txt file containg all the same sites will be created.
- (II) In the directory of WICs of sites, the \*\_WIC contribution from lineage in

sites.pdf and the \*\_WIC contribution from lineage in sites.xlsx are used to record the WIC value for each site.

- (III) In the directory of WICs of slide\_window, the \*\_WIC contribution from lineage in sliding window.pdf and the \*\_WIC contribution from lineage in sliding window.xlsx are used to record the mean WIC of each sliding window.
- (IV) The identified recombination events and region are aggregated in the \*\_Possible recombination event in XE\_.txt file.

Besides, if **-b** y is specified, for eaxmple, **-b** y **-bw** 200, then VirusRecom will perform the search of recombination breakpoint. The negative logarithm of *p*-value in each site is in the \*\_-lg(p-value) for potential breakpoint.pdf and \*\_-lg(p-value) for potential breakpoint.xlsx.

# 5. Bug report

You can tell us any problems which you encounter in usage, and so that we can further improve VirusRecom. Submit your question in <u>GitHub issuse</u> (https://github.com/ZhijianZhou01/VirusRecom/issues) of VirusRecom or send email to zjzhou@hnu.edu.cn.

#### References

Katoh, K., & Standley, D. M. (2013). MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol*, 30(4), 772-780. doi:10.1093/molbev/mst010