Comparison of chromatographic stationary phases using Bayesian-based multilevel modeling

Agnieszka Kamedulska, Łukasz Kubik, Julia Jacyna, Wiktoria Struck-Lewicka, Michał J. Markuszewski, Paweł Wiczling\*

Department of Biopharmaceutics and Pharmacodynamics, Medical University of Gdańsk, Al. Gen. Hallera 107, 80-416 Gdańsk, Poland

ABSTRACT: We used a Bayesian multilevel model of chromatographic retention to compare five RP-HPLC stationary phases: XBridge Shield RP18, XTerra MS C18, XBridge Phenyl, XBridge C8 and Xterra MS C8. For that purpose we used a large dataset of retention times collected using chromatographic techniques coupled with mass spectrometry detection. The experiments were conducted in a gradient mode for a mixture of 300 small analytes for a wide range of pH values, in MeOH and ACN, at two temperatures, and for three gradient durations. Our analysis is based on a mechanistic model derived from principles and fundamentals of liquid chromatography and utilizes the literature prior knowledge about model parameters. The data and model allowed us to characterizes the between column differences in retention of neutral, acidic, and basic analytes. The proposed approach provides interpretable summary of stationary phase properties that can be readily used in decision-making, i.e. to transfer a method to other column based on a limited set of experiments.

The selection of stationary phases is an important aspect of analytical work. There are numerous methods and approaches used to select appropriate column for a given problem. All the methods has been recently reviewed alsewere1. The chromatographic stationary phases are usually characterized by a small set of experiments using few probe analytes. This method provide a rough estimates of “bulkiness”, “acidity”, “polarity” on retention. Also a very detailed physical (and thermodynamic) description of the chromatographic systems are possible, nevertheless they require an extensive sets of experiments and as such are available for few analytes2.

It also seems possible to characterize stationary phase using retention time data collected for a relatively large and heterogeneous group of compounds. Such datasets are rather easy to collect using chromatographic techniques coupled with mass spectrometry detection. There are, however, more difficult to analyze. Since such datasets carry much information about chromatographic retention they allow for a detailed multidimensional characterization of chromatographic stationary phases and ability to predict retention (along with uncertainty) based on various number of preliminary experiments (e.g. to predict retention time for a set of analytes given no, or several measurements collected using a different stationary phase). The accurate predictions of the retention time in liquid chromatography is required for rapid column screening, computer-assisted method development and method transfer3

In this work we applied the previously developed Bayesian multilevel framework4–7 to characterize chromatographic gradient retention time datasets collected using a multicomponent mixtures of analytes, two stationary phases, and a wide range of chromatographic conditions (pH, organic modifier, temperature, gradient program). The multilevel model

We decided to compared five RP-HPLC stationary phases XBridge Shield RP18, XTerra MS C18, XBridge Phenyl, XBridge C8 and Xterra MS C8. At the end we illustrate the usefulness of the model for decision making given access to different types of preliminary data.

# EXPERIMENTAL SECTION

## Data

The data were collected by performing 84 different liquid chromatography experiments using an initial mixture of 300 analytes. The experiments differed with respect to gradient duration (30, 90, and 270 min), pH of the mobile phase (from 2.5 to 10.5), type of organic modifier (methanol (MeOH) or acetonitrile (ACN)) and column temperature (25 °C and 35 °C).

Briefly the liquid chromatography experiments were carried out using an Agilent Technologies 1260 Infinity system (Agilent Technologies, Waldbronn, Germany) and a 6224 time of flight (TOF) mass spectrometer with a dual electrospray ionization source (Dual ESI) in positive polarity, using an XBridge Shield RP18, XTerra MS C18, XBridge Phenyl, XBridge C8 and Xterra MS C8 Waters Ltd., Milford, MA, USA, 3 mm × 50 mm, 2.5 μm). The extra column volume and system dwell volume (*Vd*) equaled 0.020 mL and 1.05 mL, respectively. The column hold-up volume (*V0*) was 0.266 mL 0.271, 0.271, 0.276, 0.284 mL and the flow rate (*F*) was 0.5 mL/min.

Ammonium bicarbonate, ammonium acetate and ammonium formate were selected as buffers to control the pH of the mobile phase during chromatographic separation. The pH of the buffers (nominal aqueous pH) was adjusted to the desired pH (ammonium formate: 2.5, 3.3, 4.1, 8.9, and 9.7; ammonium acetate: 4.9 and 5.8; and ammonium bicarbonate: 6.8 and 10.5) by an appropriate addition of formic acid, acetic acid and ammonia, respectively. The pH was measured at 25 °C and 35 °C using an S220 pH meter (Mettler Toledo, Greifensee, Switzerland) with an InLab® Routine Pro ISM electrode after mixing an organic modifier with the buffer solution.

The MassHunter Profinder B.08.00 (Agilent Technologies, Waldbronn, Germany) was selected to find all the matches per formula using “Batch Targeted Feature Extraction” (containing 300 predefined mases for each analyte included in the mixture). The data for analysis were restricted to analytes that had “IdentificationScores” higher than 95%, that were present on at least 42 chromatograms, and that had less than 2 dissociation steps in a pH range from 2 to 11.

The functional groups and structural elements were determined using Checkmol (version 0.5b N. Haider, University of Vienna, 2003-2018).8 The lipophilicity (*log P*), dissociation constant (*pKalit*) were calculated using the ACD/Labs program9 based on the structures of analytes generated from SMILES strings.

More details about data extraction can be found in our previous work.4 Raw data for selected analytes is shown in Figure S1.

## Structural Model

The details of the model are provided in supplementary material. Briefly, a standard chromatographic model was employed in this work.10,11 For each analyte the effect of pH was accounted for by the following function describing the relationship between the isocratic retention factor and *pH* for an analyte with *R* dissociation steps and *R+1* forms12:

(1)

where *r* represents thedissociation step, denotes the *rth* dissociation constant, andrepresents the retention factor of a particular form of the analyte in a given chromatographic condition. Furthermore, it was assumed that depends on the organic modifier content, pH and temperature according to the following equation:

(2)

where represents the logarithm of retention factors extrapolated to 0% of organic modifier content at 25 °C for mobile phase pH 7 for the neutral and dissociated forms of the analyte; and denote slopes in the Neue equation; denotes the change in due to the increase in temperature by 10 °C, *apHr* denotes pH effects for cations and anions. In this parametrization of the Neue equation, the *S1* parameter reflects the difference between the logarithm of retention factors between the 0% organic modifier content and 100% organic modifier content.

Furthermore, a linear relationship between *pKa* values and organic modifier content was assumed:

(3)

where denotes dissociation constants of an analyte in given chromatographic conditions, denotes aqueous *pKa*, and denotes the slope due to changes in the organic modifier.

## Measurement-Error Model

The observed retention factors (*tRobs,z*) were modeled using the following model:

(4)

where *z* denotes the *zth* measurement and *student\_t* denotes the student’s t-distribution with the mean given by the predicted retention time *tR,z*, scale *σ* (analyte and column specific) and normality parameter . The retention time *tR,z* under an organic modifier gradient was calculated utilizing the well-known integral equation:

(5)

where *t0*denotes column hold-up (dead) time, *te* denotes extra column time, and denotes the instantaneous isocratic retention factor corresponding to the mobile phase composition at time *t* at the column inlet for a particular observation. The numerical solution of this integral equation was carried out using the method of steps with 4 and 10 steps for methanol and acetonitrile gradients using the method proposed by Nikitas et al.10

## Analyte-Level Model

Parameter were assumed to be different for each analyte to account for between analyte variability (BAV). The relationship was of the form:

(5)

where *Ri* denotes an individual value of a parameter *R*, is the mean value that correspond to a typical analyte with *logP=*2.2, is a regression coefficient, and denotes standard deviation. For correlated parameter a multivariate normal distribution was used.The BAV was assigned for *logkw* and *S1m* (*S1* in MeOH) for neural forms of analytes (these parameters were correlated (ρ) and depended on *logP* (*β*)), the effects of ACN on *S1* (*dS1*), the effect of temperature on logkw (*dlogkT*), the effects of dissociation on *logkw*, *S1m* and *dS1* (*dlogkw*, *dS1m*, *ddS1* for acids and bases), and the effect of column on these parameters (*clogkw*, *cS1m*, *cβ*, *cdS1*, *cdlogkT*, *cdlogkw*, *cdS1m*, *cddS1*). The pH effects for cations and anions (*apH*) were assumed to be the same across analytes but different across the columns (*capH*). S2 parameter was assumed to be the same across analytes and columns but different in MeOH and ACN (*logS2m*, and *dlogS2* for ACN effect). Also the BAV was assigned for dissociation related parameters: *pKaw* (dependent on literature *pKa* values), the parameters for acids and bases in MeOH (, and the effect of ACN on (. The dissociation related parameters were assumed to be independent of the column. The standard deviation of BAV was denoted as ω and *cω* for the parameters of the neutral form of analyte on XBridge Shield RP18 (ω) and for the difference between the other columns and XBridge Shield RP18(ω), *ωT*and *cωT* for temperature effects, κ and cκ for the parameter related to dissociated forms and τ for the parameters related to dissociation constant.

## Priors

The Bayesian model requires specification of priors that allow to incorporate domain expertise into inferences. In this work priors were selected based on literature knowledge as discussed previously.4 The means for the priors describing column effects were assumed 0. The standard deviation were 0.5 of the standard deviations/scales used for XBridge Shield RP18 parameters.

## Bayesian Inference

**Technical.** Multilevel modeling was performed in Stan/cmdstanr33 software linked with Rstudio34. For the inference we used 8 Markov chains with 500 iterations after 1000 warm up 1000 iterations. The *reduce\_sum* function was used to accelerate the calculations by parallelizing the execution of a single Stan chain across multiple cores. Convergence diagnostics were checked using Gelman-Rubin statistics and trace plots. No divergence was reported in the model. The R code, data and Stan code used to analyze the data are publicly available from GitHub (https://github.com/wiczling/bmm-fivecolumns). The raw data are also available through a repository.36 The calculations were run on the Tryton computing cluster in Centre of Informatics Tricity Academic Supercomputer.

**Predictions Using a Limited Set of Experiments.** The model applicability was illustrated using selected 6 analytes with different acidic/basic properties: acridine (monoprotic acid), baclofen (zwitterion: acidic and basic group), hydrocortisone (neutral), pioglitazone (zwitterion: basic and acidic group), quinine (diprotic: 2 basic groups), and tolbutamide (monoprotic base). In this work we present the limited data predictions that correspond to a future predictions given access to the experimental data collected for XBridge Shield RP18 column. For this problem all the population parameters were fixed and the other were re-estimated using the additional data. It answers a problem of how accurate can we be in predicting analyte retention on other columns in a situation of having a very good understating of retention in one reference column. The predictions were summarized as uncertainty chromatograms (posterior distribution of retention times expected for a given set of chromatographed analytes under given conditions).12 The uncertainty chromatogram visualizes the uncertainty for the locations of the maximum of each peak on a given chromatogram. Any area under the uncertainty chromatogram for a particular analyte can be probabilistically interpreted as a fraction of analytes (similar with respect to predictors and gathered data) that are expected to have a retention time within the range that the area was calculated.

# RESULTS AND DISCUSSION

In this work, we applied a mechanistic model to describe the retention data of small molecules obtained for a wide range of chromatographic conditions and for five chromatographic columns. The model was built based on known fundamentals of gradient chromatography and prior knowledge available in the literature. The methodology allow to characterize the stationary phases properties using the parameters of the Neue model. This allows for an easy interpretation and application of this parameters in decision making. The model parameters are summarized in Table S1 and Figure S2. The key parameters are provided in Figures 1-3. These parameters provide a concise summary of influence of various factors affecting retention of analytes on the tested columns for a typical (having *logP* of 2.2) neutral, acidic, and basic analyte.

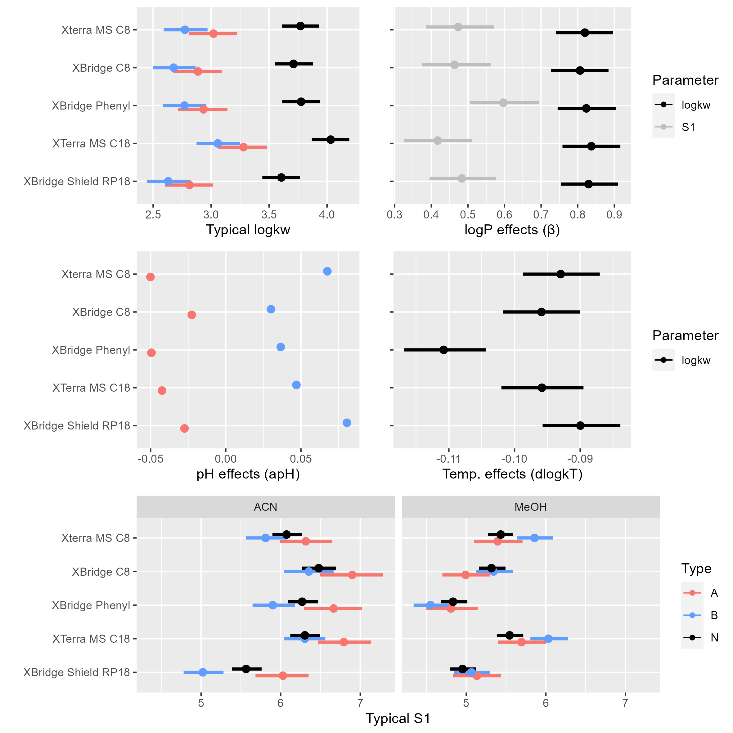


Figure 1. Summary of marginal posterior distributions of the key population-level parameters characterizing retention of analytes in the tested columns.

The interpretation of parameters and all the effects is straightforward. Each parameter isolates a certain effect of a design factor (pH, fi, temperature, etc) on retention and provides quantitative summary. As an example, the typical logkw of a neutral form of an analyte (a measure of hydrophobicity) is 3.6 for XBridge Shield RP18, and is 0.42, 0.17, 0.10, and 0.17 higher for XTerra MS C18, XBridge Phenyl, XBridge C8 and Xterra MS C8 columns. The typical slope S1 (in MeOH) is 4.96 and the difference between XBridge Shield RP18 and the other column is 0.59, -0.12, 0.36, 0.48.

The logkw for acids and bases is lower by -0.79 and -0.97 on XBridge Shield RP18. The between column differences for that parameter are small.

The pH effects on logkw for acids and bases (apH) are small and negative for acids and positive for bases. This effects reflects the changes in stationary phase properties due to pH of the mobile phases.

The log P effects logkw and S1 with a slope of 0.83 and 0.48 for XBridge Shield RP 18. The effect of other column on the parameter are small. The largest difference is between XBridge C8 and Xterra MS C18 (about 0.18) for cS1.

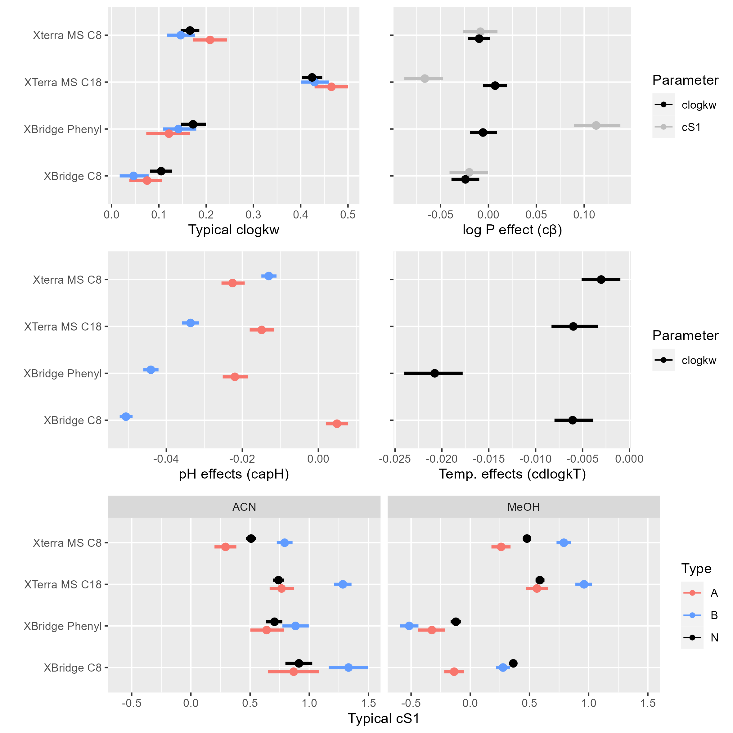


Figure 2. Summary of marginal posterior distributions of the key population-level parameters characterizing the difference in retention of analytes between the indicated column and XBridge Shield RP18 column.

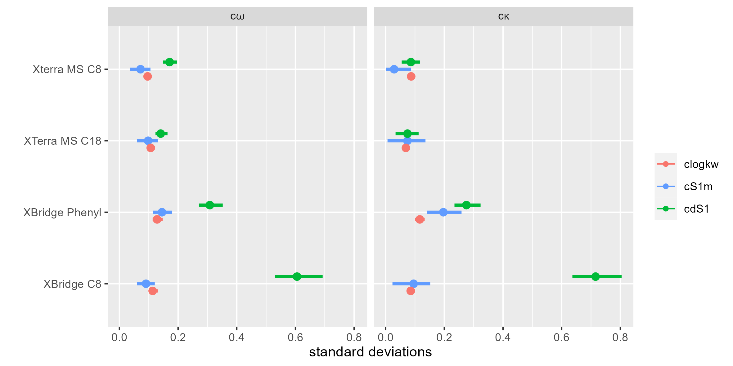


Figure 3. Summary of marginal posterior distributions of the BAV parameters characterizing the difference in retention of analytes between the indicated column and XBridge Shield RP18 column.

The model predictions are well calibrated with the data, as shown in Figure S10. The individual and population predictions versus observed retention times are relatively symmetrically distributed around the line of identity indicating model applicability for predictions. The individual and population predictions for several analytes are shown in Figure S6. The individual prediction are very precise and close to observed data. The population predictions are also well calibrated however, but are less precise.

All model parameters jointly affect analyte retention. To better illustrate this joined effect of parameters, we simulated the average retention factors for the typical acidic, basic and neutral analyte with log P of 0, 3 and 6. The results are present in Figure 4. We also provide various isocratic prediction for selected analytes to illustrate the impact of parameters on retention (Figure S3, S4, S8 and S9)

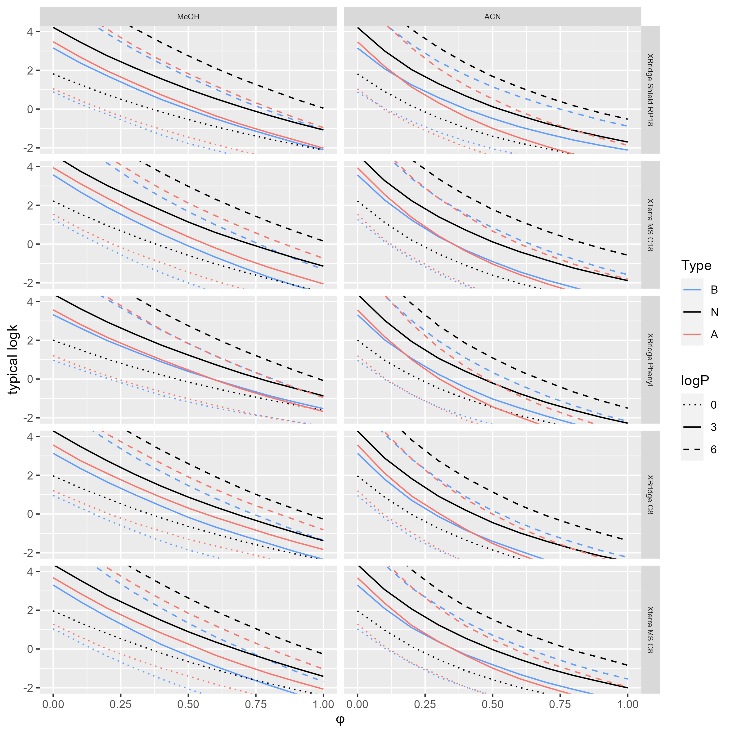


Figure 4. Graphical display of the typical retention profiles (log k vs fi ) for basic, neutral and acidic form of an analyte with log P of 0, 3, and 6 at 25oC. The uncertainty is not present to improve readability.

The model provides quantitative results allowing to predict likely chromatogram given various number of experimental data (even lack of any data). Such prediction can be used for decision making. In this work we show the comparison of population (future observations when no experimental data are available) and limited data predictions (future predictions given access to the limited experimental data, in this work to all experiments collected for XBridge Shield RP 18). Simulations are shown in Figures 5 and 6. The population predictions (Figure 4) are of limited usefulness as they lead to a very large uncertainties. Basically, the uncertainties for population predictions are mostly driven by unexplained variability, *ω*, which is large. However, by adding information one can decrease this variability. For example, the added predicted value of XBridge Shield RP18 data is large. For this limited data predictions the uncertainty is almost entirely reduced for XBridge Shield RP18 column (as there is a lot of data). For other columns there is still some proportion of uncertainty left (*cω*). Since cωis small (about 0.1)one can expect a fairly precise predictions. The between analyte variabilities are on a log (with base 10) scale and so these standard deviations has to be interpreted accordingly. For example the between analyte variability of 0.1 for clogkw implies that the predicted effects of the column are on the order of ± 0.1, which corresponds to a multiplicative factors from exp(-2.3\*0.1)=0.79 to exp(2.3\*0.1)=1.26. For simple problems involving few analyte it indicates that one is able to predict retention with an uncertainty of few minutes that might be of practical usefulness.

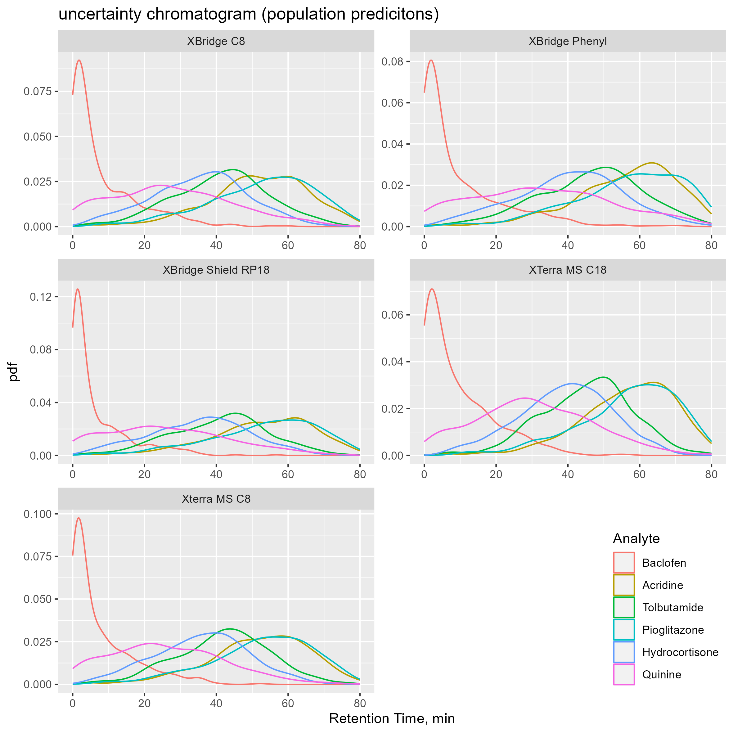


Figure 5. Uncertainty chromatograms displaying the predictions for 6 selected analytes using no preliminary information. Each peak represents the range of analyte retention factors compatible with prior and preliminary data.

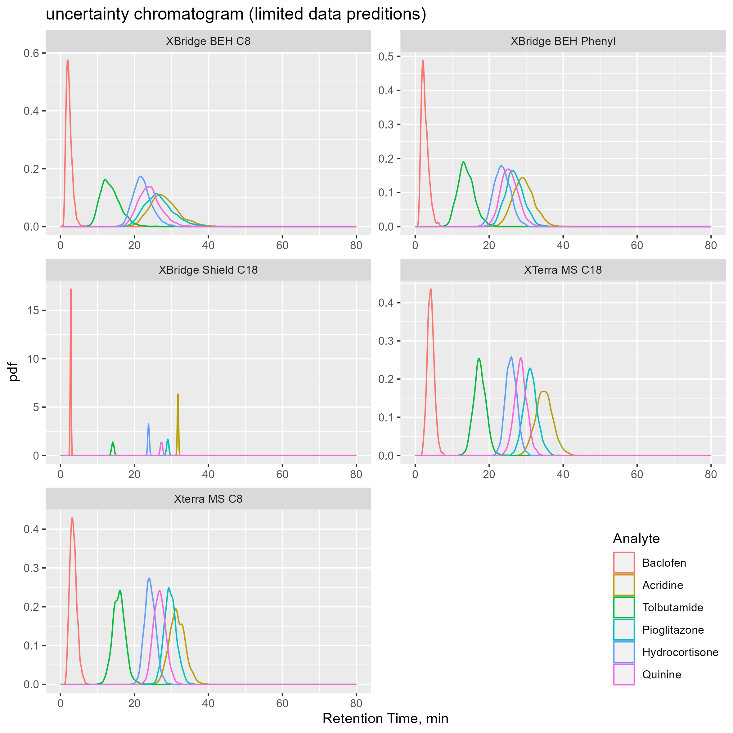


Figure 6. Uncertainty chromatograms displaying the predictions for 6 selected analytes based on XBridge Shield RP18 data. Colors correspond to different analytes that are identified in the legend.

<including column as fixed vs. random effects, possibility of adding predictors, like retention of a compound under some standard conditions).

# CONCLUSIONS

This work demonstrates the application of a Bayesian multilevel model to compare various stationary phases using large datasets collected for a wide range of chromatographic conditions. The analysis characterizes the difference in chromatographic retention of neutral, acidic, and basic analytes. It also provides a way to characterize the influence of pH, temperature, organic modifier type and content on analyte retention.

# ASSOCIATED CONTENT

## Supporting Information

The following Supporting Information is available free of charge at the ACS website:

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# AUTHOR INFORMATION

## Corresponding Author

\* wiczling@gumed.edu.pl

## Author Contributions

ŁK, JJ, WSL collected the experimental data; ŁK and AK prepared the data for analysis; AK and PW analyzed the data; PW, AK wrote the paper with input from all authors; PW conceived of the presented idea, designed the study and supervised the project; and MM helped supervise the project.

Notes  
The authors declare no competing financial interest.

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# REFERENCES

(1) Žuvela, P.; Skoczylas, M.; Jay Liu, J.; Ba̧czek, T.; Kaliszan, R.; Wong, M. W.; Buszewski, B. Column Characterization and Selection Systems in Reversed-Phase High-Performance Liquid Chromatography. *Chem. Rev.* **2019**, *119* (6), 3674–3729. https://doi.org/10.1021/acs.chemrev.8b00246.

(2) Gritti, F.; Guiochon, G. Adsorption Mechanism in RPLC. Effect of the Nature of the Organic Modifier. *Anal Chem* **2005**, *77* (13), 4257–4272. https://doi.org/10.1021/ac0580058.

(3) Gritti, F. Perspective on the Future Approaches to Predict Retention in Liquid Chromatography. *Anal. Chem.* **2021**, *93* (14), 5653–5664. https://doi.org/10.1021/acs.analchem.0c05078.

(4) Kamedulska, A.; Kubik, Ł.; Jacyna, J.; Struck-Lewicka, W.; Markuszewski, M. J.; Wiczling, P. Toward the General Mechanistic Model of Liquid Chromatographic Retention. *Anal. Chem.* **2022**, *94* (31), 11070–11080. https://doi.org/10.1021/acs.analchem.2c02034.

(5) Kamedulska, A.; Kubik, Ł.; Wiczling, P. Statistical Analysis of Isocratic Chromatographic Data Using Bayesian Modeling. *Anal Bioanal Chem* **2022**, *414* (11), 3471–3481. https://doi.org/10.1007/s00216-022-03968-x.

(6) Wiczling, P.; Kamedulska, A.; Kubik, L. Application of Bayesian Multilevel Modeling in the Quantitative Structure-Retention Relationship Studies of Heterogeneous Compounds. *ANALYTICAL CHEMISTRY*, 2021, *93*, 6961–6971. https://doi.org/10.1021/acs.analchem.0c05227.

(7) Wiczling, P. Analyzing Chromatographic Data Using Multilevel Modeling. *ANALYTICAL AND BIOANALYTICAL CHEMISTRY*, 2018, *410*, 3905–3915. https://doi.org/10.1007/s00216-018-1061-3.

(8) Haider, N. Functionality Pattern Matching as an Efficient Complementary Structure/Reaction Search Tool: An Open-Source Approach. *Molecules* **2010**, *15* (8), 5079–5092. https://doi.org/10.3390/molecules15085079.

(9) ACD/Labs. *Release 12.0*; Advanced Chemistry Development Inc.: Toronto, ON, Canada, www.acdlabs.com, 2022., 2011.

(10) Nikitas, P.; Pappa-Louisi, A. New Equations Describing the Combined Effect of PH and Organic Modifier Concentration on the Retention in Reversed-Phase Liquid Chromatography. *J Chromatogr A* **2002**, *971* (1–2), 47–60. https://doi.org/10.1016/s0021-9673(02)00965-2.

(11) Nikitas, P.; Pappa-Louisi, A. Retention Models for Isocratic and Gradient Elution in Reversed-Phase Liquid Chromatography. *Journal of Chromatography A* **2009**, *1216* (10), 1737–1755. https://doi.org/10.1016/j.chroma.2008.09.051.

(12) Jano, I.; Hardcastle, J. E.; Zhao, K.; Vermillion-Salsbury, R. General Equation for Calculating the Dissociation Constants of Polyprotic Acids and Bases from Measured Retention Factors in High-Performance Liquid Chromatography. *J Chromatogr A* **1997**, *762* (1–2), 63–72. https://doi.org/10.1016/s0021-9673(96)00739-x.

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