

2012 AASRI Conference on Computational Intelligence and Bioinformatics

Bioinformatics Analysis and Characteristics of UL21 Protein from Duck Virus Enteritis

Yong Shi^{a,b} Anchun Cheng^{a,b,c,*}, Mingshu Wang^{a,b,c}

^a*Institute of Preventive Veterinary Medicine, Sichuan Agricultural University, Wenjiang, Chengdu city, Sichuan, 611130, P.R.China 2.*

^b*Avian Disease Research Center, College of Veterinary Medicine of Sichuan Agricultural University, 46 Xinkang Road, Ya'an, Sichuan 625014, P.R. China*

^c*Key Laboratory of Animal Disease and Human Health of Sichuan Province, Sichuan Agricultural University, Wenjiang, Chengdu city, Sichuan, 611130, P.R.China)*

Abstract

The UL21 protein of duck virus enteritis is analyzed by means of some software and online tools. The physicochemical properties results showed that the DEV UL21 product is a stable protein which consists of 561 amino acids and contain 27 potential phosphorylation sites, 3 potential glycosylation sites at aa residues 2, 172, 522. Both the signal peptide and the transmembrane region are not found. The secondary structure results revealed that UL21 protein is composed of 44.56% Alpha helix (h), 13.55% extended strand (e) and 41.89% random coil (c), respectively. In addition, the protein subcellular localization mainly locates at cytoplasmic with 65.2%, nuclear with 17.4 %. The phylogenetic tree and amino acid sequence comparison analysis revealed DEV UL21 protein is conserved and most closely related to Varicellovirus and Mardivirus. These results provided rational data to elucidate biological function and physiological features of the UL21 gene.

© 2012 Published by Elsevier B.V. Open access under [CC BY-NC-ND license](https://creativecommons.org/licenses/by-nc-nd/4.0/).

Selection and/or peer review under responsibility of American Applied Science Research Institute

Keywords: Duck virus enteritis(DVE); Duck plague(DP); UL21 Gene; UL21 protein.

* Anchun Cheng; Tel.: +86 835 2885774; fax: +86 835 2885774
E-mail address: chenganchun@vip.163.com.

1. Introduction

Duck Virus Enteritis(DVE), also called Duck Plague(DP), is a serious, contagious viral disease, and highly lethal in all ages of Anseriformes. The mortality and reduction in egg production in commercial caused significant economic losses[1-2]. Duck enteritis virus (DEV) is a member of the Alphaherpesvirinae subfamily, but it has not been divided into any genus according to the Eighth International Committee on Taxonomy of Viruses (ICTV) [3]. Previous researches focused on the epidemiology, diagnosis, prevention, the structure and morphogenesis of DEV[4-7], but now, more about the DEV gene expression, protein purification, protein function, detective studies have reported with molecular biology technology[8-10].

Different herpesvirus have similar structure. The herpes simplex virion has four components: core, capsid, tegument and the envelope. The tegument which links the capsid and the envelope, composed of a multitude of about 20 proteins, including VP1/2 (UL36), VP11/12(UL46), VP13/14 (UL47), VP16 (UL48), VP22 (UL49), ICP0, ICP4, US2, US3, US10, US11, UL11, UL13, UL14, UL16, UL17, UL21, UL37, UL41, UL51 and UL56[11].

The HSV-1 UL21 gene product, a capsid-associated tegument protein, promotes the outgrowth of long cellular processes when it is over-expressed in non-neural cells. It is presumed that UL21 protein physically associates with microtubules[13]. The early reports showed that both HSV and PRV UL21 gene products are not essential for viral replication in cultured cells and its deletion resulted in only marginally reduced titers but clearly decreased plaque sizes[14-15]. The PrV UL21 gene is a major determinant of PrV virulence, and its point mutations affecting the UL21 gene of live vaccine strain Bartha contribute to its attenuated phenotype[16]. In addition, PRV UL21 mutants which lacks UL21 gene has apparently reduced virulence for mice[17-18]. In short, the PRV UL21 protein associates with virulence. But as far, studies about DEV UL21 gene product are limited.

The DEV CHv strain genome was identified and sequenced in our laboratory. In this paper, the product encoded by DEV-UL21 gene, which is presumed as a capsid-associated tegument protein, is analyzed by means of bioinformatics methods. These works may provide some information for further studies on DEV UL21gene.

2. Materials and methods

2.1. DEV-UL21 gene and the deduced amino acid sequence

The DEV CHv strain, which is a highly virulent field strain, was obtained from the Key Laboratory of Animal Disease and Human Health of Sichuan Province. The DEV UL21 gene (GenBank Accession No. EU 195090) corresponding amino acid is deduced on line <http://mobyli.pasteur.fr/data/jobs/transeq>.

2.2. Analysis of the physico-chemical properties of DEV-UL21 gene product

The component of the DEV-UL21 protein sequence was analyzed by DNASTar7.0 and on-line tool. All of the websites of online predicted tools are shown in Table1.

Table1 The Websites For Analysis And Prediction of DEV UL21 Protein Bioinformatics

<i>Functions</i>	<i>Websites</i>
Protparam	http://www.expasy.org/tools/protparam.html

Signal peptide	http://www.cbs.dtu.dk/services/SignalP
Phosphorylation sites	http://www.cbs.dtu.dk/services/NetPhos/
Transmembrane region	http://genome.cbs.dtu.dk/services/TMHMM/
Hydrophobicity	http://www.expasy.org/tools/protscale.html
Glycosylation sites	http://www.cbs.dtu.dk/services/NetNGlyc/
Epitope analysis	http://www.cbs.dtu.dk/services/BepiPre
Subcellular localization	http://psort.nibb.ac.jp/form2.html
Secondary structure	http://npsa-pbil.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_mlrc.html
Tertiary structure	http://www.cbs.dtu.dk/services/CPHmodels/

2.3. Amino acid sequence comparison

Amino acid sequence comparison between the putative proteins encoded by DEV UL21 and other DEV strains(DEV UL21-like strain and DEV VAC strain) were aligned with the DNASTar 7.1 software in order to investigate differences in different DEV strains. Meanwhile, multiple sequences alignment of UL21 protein sequence of DEV, GaHV-2, GaHV-3, MeHV-1, HSV-1, EHV-1 and SuHV-1 were performed to validate the conservatism of herpesvirion UL21 product.

2.4. Phylogenetic analysis of the DEV-UL21 protein according to Compare with 25 Reference herpesviruses UL21 Protein Sequences

A phylogenetic tree was performed according to the amino acid sequences of the UL21 product in DEV and 25 Reference herpesviruses by using the MegAlign of DNASTar 7.1. The 25 reference herpesviruses UL21 protein sequences were employed from the NCBI GenBank nucleotide database (shown in Table 2).

Table 2 The Information About UL21 Protein Sequence of 25 Reference Herpesviruses

Genus	Virus name (Abbreviation)	GeneBank accession NO.	L [aa]
Simplexvirus	Cercopithecine herpesvirus 1(CeHV-1)	AF 533768	526
Simplexvirus	Cercopithecine herpesvirus 2(CeHV-2)	NC 006560	526
Simplexvirus	Bovine herpesvirus 2(BoHV-2)	AF 387490	522
Simplexvirus	Human herpesvirus 1(HSV-1)	NC 001806	535
Simplexvirus	Human herpesvirus 2(HSV-2)	NC 001798	532
Simplexvirus	Papiine herpesvirus 2(PaHV-2)	NC 007653	528
Simplexvirus	Saimiriine herpesvirus 1(SaHV-1)	HM 625781	537
Varicellovirus	Bovine herpesvirus 1(BoHV-1)	NC 001847	574
Varicellovirus	Bovine herpesvirus 5(BoHV-5)	NC 005261	603
Varicellovirus	Equine herpesvirus 1(EHV-1)	AY 464052	530
Varicellovirus	Equine herpesvirus 4(EHV-4)	AF 030027	529
Varicellovirus	Equid herpesvirus 9(EHV-9)	AP 010838	530
Varicellovirus	Felid herpesvirus 1 (FeHV-1)	FJ 478159	527

Varicellovirus	Human herpesvirus 3(HSV-3)	DQ 674250	541
Varicellovirus	Suid herpesvirus 1 (SuHV-1)	AY 363172	525
Varicellovirus	Canine Herpesvirus(CHV)	AY 768815	522
Iltovirus	Psittacid herpesvirus 1 (PsHV-1)	NC 005264	569
Iltovirus	Gallid herpesvirus 1(GaHV-1)	NC 006623	532
Mardivirus	Gallid herpesvirus 2(GaHV-2)	AF 439271	546
Mardivirus	Gallid herpesvirus 3(GaHV-3)	HQ 840738	532
Mardivirus	Meleagrid herpesvirus 1 (MeHV-1)	AF 282130	581
Roseolovirus	Human herpesvirus 7 (HSV-7)	AF 037218	430
Lymphocryptovirus	Human herpesvirus 4(HSV-4)	NC 007605	404
Rhadinovirus	Murid herpesvirus 4 (MHV-4)	AF 105037	644
Macavirus	Ovine herpesvirus 2 (OvHV-2)	AY 839756	400

3. Results

3.1. The character of DEV UL21 gene

The complete open reading frame (ORF) of the DEV-UL21 gene was predicted to encode a polypeptide containing 561 amino acids, and the corresponding amino acid sequence was showed below:

MEFHYWETINHNGVTIFYITRDGMRAYFACGGCILSVPRPPENDSDTQAEIAKFGIALRGITSGDLVLS
 NYVRSELGRRGLKWIIGDGEVFIDSLDLLGHTSGSSERDLCGTNSGDGSTERDLCGALEVEVRDQCIA
 EYMSVLEISSGLILSTGHIFSNYQVIKLYDVPIITNASSGFIYEPNRNAFALMQARLTSLPQSLAAMVDG
 LFDRIAVRRRGVREETKQTDVIITGKRSFGTVLVKHHGGERHRGSGEGTTLNTDDCDITTTLSRKHS
 RRGARKTTVSSSFVQVKYIPAVLNIWEYGAGNFKPTRSLGALWTVFCRIGDVVSQDISTWFGLEPEFN
 DARARIGDAIEASFGNIGELFVGYSMGRSVSSAQKFALVQYLCKGGYPNCYPPIEHLCVSLSADSESF
 PEPPRDIHLLVDTTNRLFRESIIWASSVAILSTRVKQLRVATDEDDSVMDDAETLFEMATDLLDTAQ
 EHQSIQLQRIARLASIIAEIYTTNDLMKTAIRTDRCFGNSYILNATIDAMCSSIFDEKCDIQKGVLTGLA
 LIDRRLKNAGLLG*

3.2. The physic chemical properties of DEV UL21 gene product

The open reading frame (ORF) of the DEV UL21 gene is expected to encode a protein with formula being $C_{2723}H_{4133}N_{765}O_{841}S_{24}$ containing 561 amino acids with a molecular mass of 61993.2 and an isoelectric point (pI) of 5.49. The total number of negatively charged residues (Asp + Glu) is 70, and the positively charged residues (Arg + Lys) is 57. The instability index (II) is computed to be 36.98, indicating that this classifies the protein as stable.

Some information about the DEV UL21 protein were obtained by some online tools (shown in table I): neither signal peptide nor transmembrane region was found in DEV UL21 protein the through online analysis; Phosphorylation sites analysis showed that there are 27 potential phosphorylation sites (showed in Fig. 1a) when the threshold is above 0.5, including serine 13, threonine 9, tyrosine 5; the DEV UL21 protein hydrophobic amine acid district centered in aa 50-80, 120-170, 180-210, 280-300, 350-410, 420-440 and 480-500 (shown in Fig.1b); NetNGlyc1.0 analysis shows DEV UL21 protein contains 3 potential N-linked glycosylation sites at aa residues 2, 172, 522 when the threshold of prediction score is above 0.5 (shown in Fig.1c); Epitope analysis by Bepipred 1.0 server shows that DEV UL21 protein epitope was centred in aa residue 36-49, 100-121, 178-183, 217-224, 244-263, 273-281, 303-310, 338-346, 405-416, 452-462 and 475-

480; the futhur analysis by using Protean program of DNASTar 7.1 (showed in Fig.1d); the analysis of the UL21 protein subcellular localization indicates that it locates in cytoplasmic with 65.2%, nuclear with 17.4 % , mitochondrial with 8.7%, vacuolar with 4.3% and vesicles of secretory system with 4.3%.

The prediction for DEV UL21 protein secondary structure is shown in Fig.2 and Fig3, the results suggest that of DEV UL21 protein consists of 44.56% Alpha helix (h), 13.55% extended strand (e) and 41.89% random coil (c) respectively. The Alpha helix of DEV UL21 protein is mainly located at aa 65-77, 119-143, 184-214, 341-353, 371-380, 422-451, 458-509, 521-534 and 544-556, Extended strand is mainly at aa 2-9, 14-19, 24-28, 295-300, 381-385, 393-403 and 417-421, the random coil are mainly situated in aa 36-44, 58-64, 97-107, 109-118, 215-254, 301-313, 404-415 and 534-543. However, there is no suitable template for modeling of tertiary structure of DEV UL21 protein could be found by CPHmodels-3.0 Server.

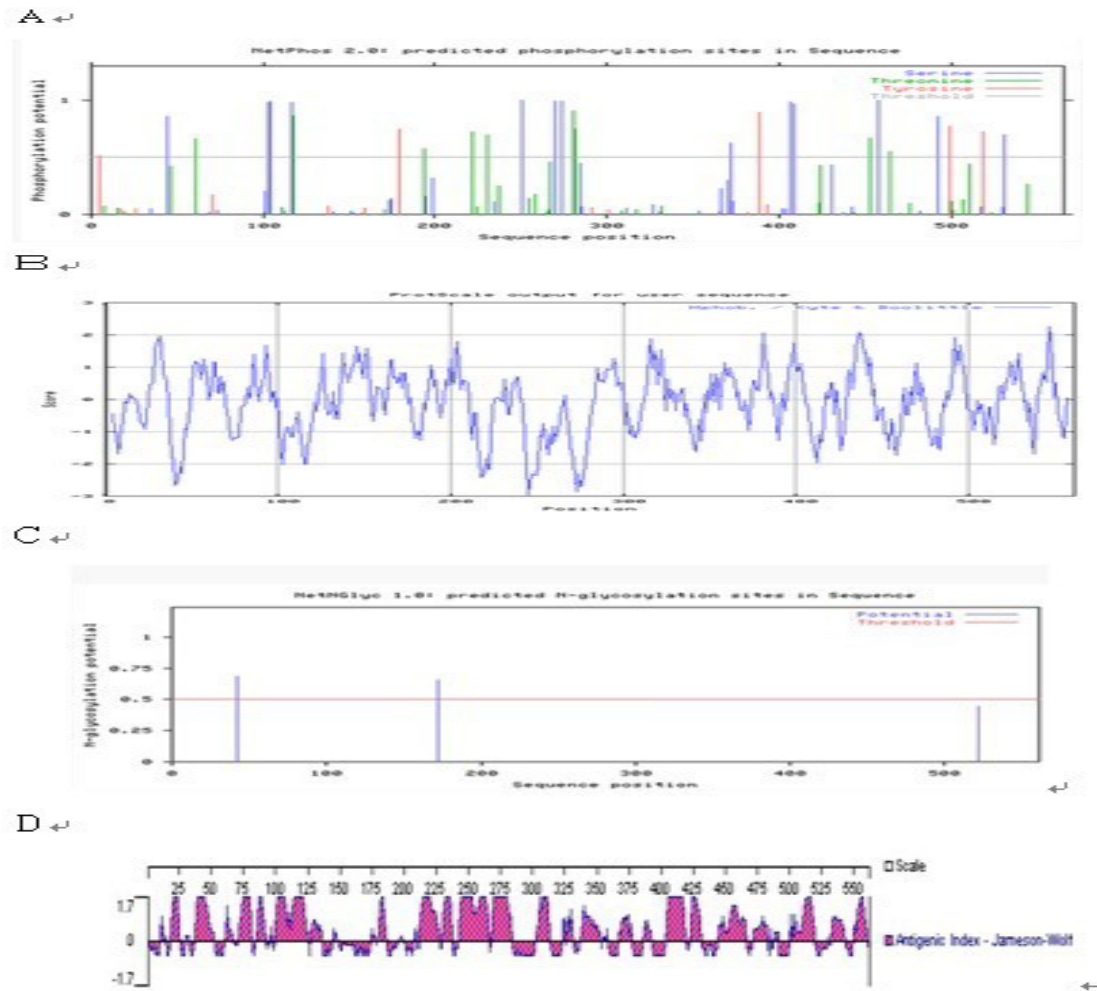


Fig. 1 A : The prediction result for potential phosphorylation sites of DEV UL21 protein. B : The prediction result for hydrophilicity domain of DEV UL21 protein, the hydrophilicity domain is in beneath with the score smaller than zero. C : Glycosylation sites of DEV UL21 protein by NetNGlyc1.0 D : the epitope analysis of DEV UL21 protein by Protean program of DNASTar 7.1

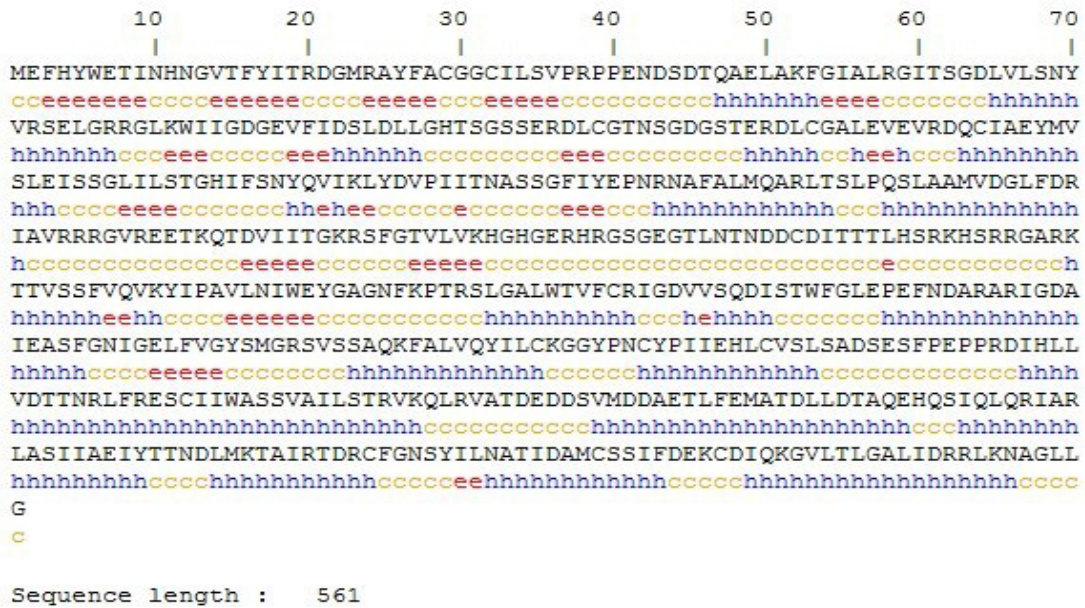


Fig. 2 The prediction of secondary structure of DEV UL21 protein by online tool. “h”= alpha helix; “e” = extended strand; “c”= random coil.

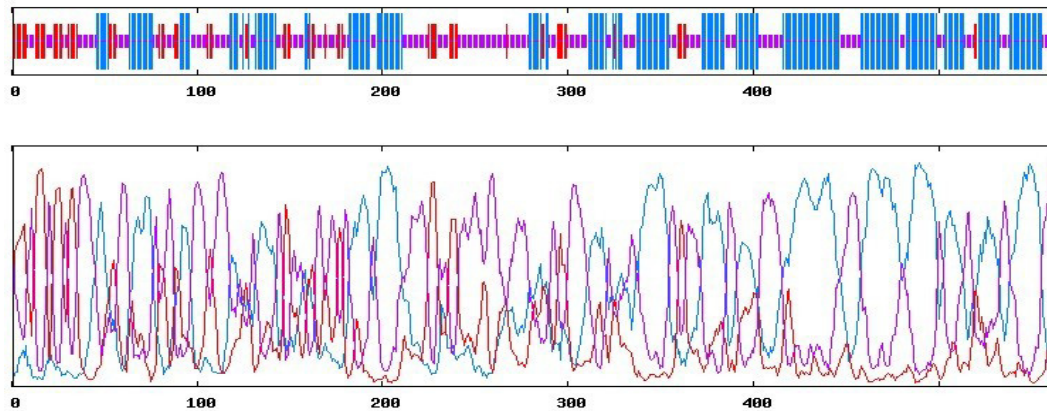
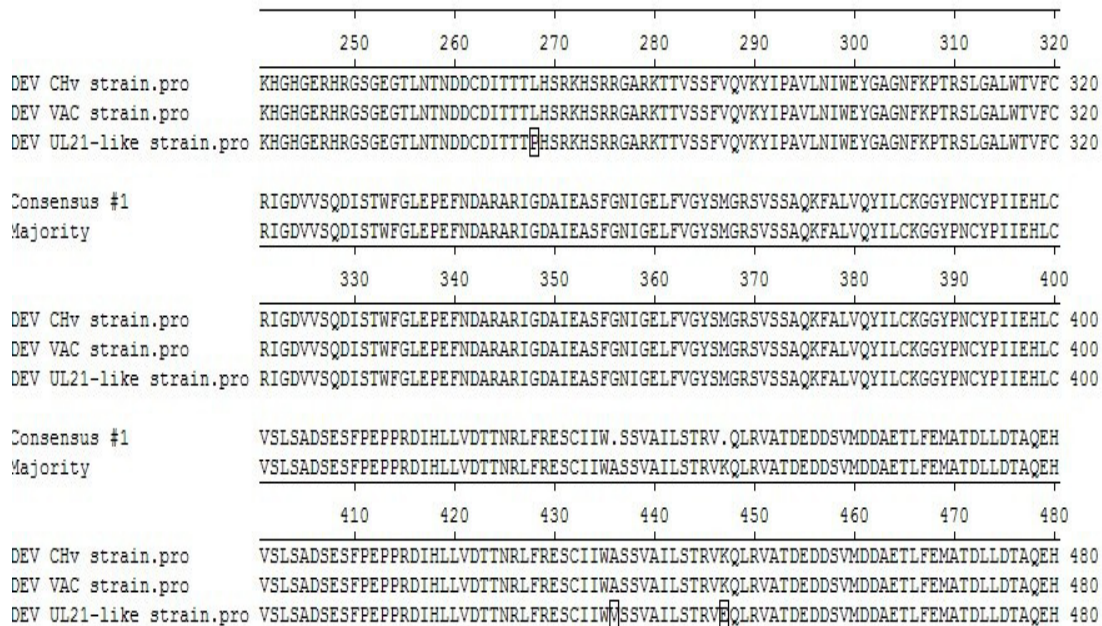


Fig. 3 The prediction of distribution of secondary structure of DEV UL21 protein.



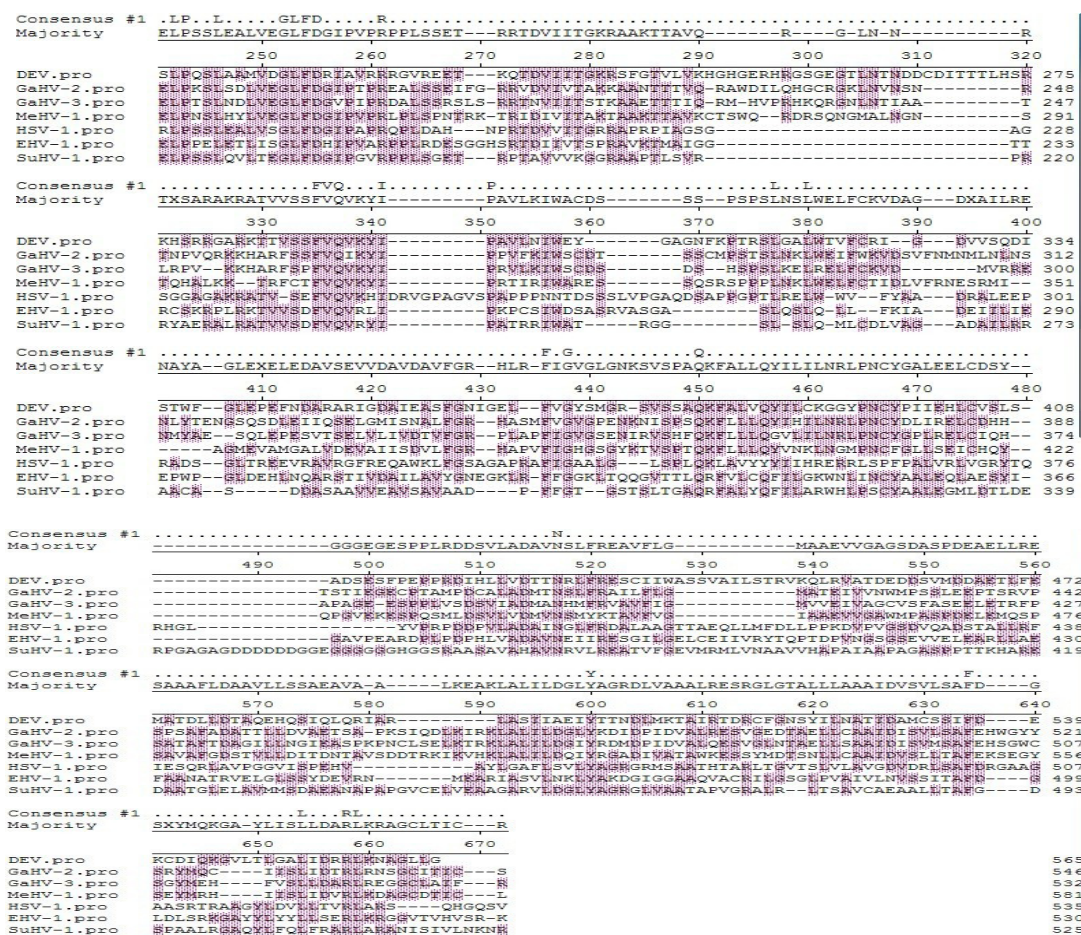


Fig. 5 Multiple sequences alignment of UL21 protein sequence of DEV, GaHV-2, GaHV-3, MeHV-1, HSV-1, EHV-1 and SuHV-1. The conserved structural motifs that are characteristic of the protein are shadowed.

Amino acid sequence comparison among different DEV strain shows that the sequences of DEV CHV and VAC strain are identical, the UL21-like strain are similar with them except aa position 268, 436, 447 respectively (shown in Figure 4). Multiple sequences alignment of UL21 protein sequences we selected showed the conserved region of UL21 protein centred in aa 69-84, 93-103, 112-124, 174-189, 230-260, 274-283 and 335-341.

3.4. . Phylogenetic analysis about the UL21 protein sequence of DEV and 25 reference herpesviruses

A phylogenetic tree was established based on UL21 protein sequence of DEV and those of 25 reference herpesviruses (display in Fig.6). The result shows there are 4 mainly branches: Varicellovirus, Mardivirus and DEV in a large branches; Simplexvirus, and Iltovirus in a same branches; Betaherpesvirinae and Gammaherpesvirinae in other two branches respectively. The DEV are with MeHV-1, GaHV-2 and GaHV-3 in a monophyletic clade. Protein sequence comparison (showed in Fig.7) by Clustal multiple revealed that DEV UL21 protein shares 26.4%, 27.3%, 26.2% similarity with GaHV-2, GaHV-3, MeHV-1 and 33.0%,

Fig. 6 Evolutionary relationships of the putative DEV CHv UL21 protein with its 25 reference herpesviruses(Table 2). Phylogenetic tree of these proteins was generated by using the MegAlign program with Clustal V multiple alignment of DNASTar 7.1.

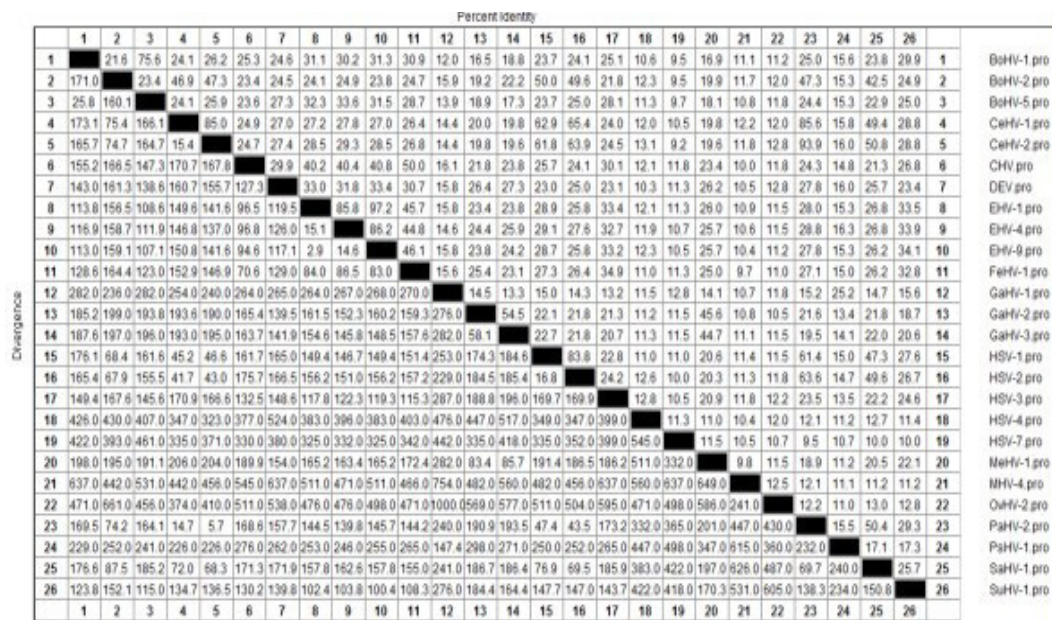


Fig.7 Identify analysis of the UL21 protein sequences of DEV and other 25 alphaherpesviruses by using DNASTar 7.1 multiple alignments.

4. Discussion

Previous reports about UL21 protein are mainly focused on the PRV and HSV-1. The UL21 protein is a tegument protein, and weakly associated with the capsid [13]. The deletion mutants which lacked the whole or partly UL21 gene sequences, were proved that UL21 protein is dispensable for growth both in cultured cells or in vivo [14-15,19]. There is a complex interaction between the UL16 and UL21 tegument proteins in PRV and HSV-1 [15,20-23]. In addition, the PRV UL21 gene product associates with virulence, and package pUL46, pUL49, and pUS3 efficiently [24]; the HSV-1 UL21 gene product associates with microtubules, indicating that the UL21 protein may have function of transport. All above revealed UL21 product of HSV and PRV is a regulating protein. The UL21 gene is conserved among herpes virions [25], and we presumed the product of DEV UL21 gene may have similar function.

There are few reports about UL21 protein of DEV, so we study the DEV UL21 protein predicted information base on bioinformatics software and online tools. DEV UL21 protein doesn't contain the signal peptide and the transmembrane region, suggesting the protein is not secreted protein or membrane glycoprotein. Protein phosphorylation on serine, threonine, and tyrosine (Ser/Thr/Tyr) is generally considered the major regulatory posttranslational modification functional in prokaryotes [26]. There are 27 potential phosphorylation sites were found in DEV UL21 protein, which may associates with regulating function, and the apparent molecular mass may be more than predicted value 61993.2. The analysis of the UL21 protein subcellular localization indicates that the UL21 gene product of DEV could both local in cytoplasmic and nuclear, which is similar with PRV and HSV-1 protein [13-14,17].

The secondary structure is related with protein function. The alpha helix of the protein have higher chemical bonding energy, can firmly maintain proteinic higher structure. The alpha helix of the protein plays a important role in DNA binding motifs, but it seldom become B cell epitopes because it is difficult to be recognized by antibody better, and usually locates at protein interior. Extended strand and random coil are more noncohesive flexibility structures and always include B cell dominant epitopes, because they are more loosen texture, which are easy to generate retortion and stretch out of the proteinic surface and be recognized by antibody [27-29]. DEV UL21 protein consists of 44.56% Alpha helix (h), 13.55% extended strand (e) and 41.89% random coil (c) respectively, indicating more extended strand and random coil structure may contain certain B cell epitopes. Previous results suggest that herpes simplex tegument proteins are processed for antigen presentation in vivo and are possible candidate compounds for herpes simplex vaccines [30], about 20 main antigenic determinants in epitope analysis further suggests that UL21 protein is possible candidate compounds for DEV vaccines to the prevention and diagnosis of the duck virus enteritis.

Amino acid sequence comparison among different DEV strain shows that there may be light difference of UL21 protein among different DEV strains. We presumed the difference of identical gene may cause virulence diversity. The result of multiple sequences alignment of UL21 protein sequence of DEV, GaHV-2, GaHV-3, MeHV-1, HSV-1, EHV-1 and SuHV-1 supports that UL21 gene is conserved among Alphaherpesvirinae at least.

From Fig.6, the established phylogenetic tree based on DEV CHv UL21 protein with its 25 reference herpesviruses and cluster analysis results show DEV has a close evolutionary relationship with GaHV-2, GaHV-3, MeHV-1, which belong to Mardivirus, but protein sequence comparison showed in Fig.7 revealed similarity between DEV UL21 protein and Varicellovirus (including EHV-1, EHV-4, EHV-9 and FeHV-1) is higher than those between Mardivirus. It has reported DEV dUTPase gene product, gI, UL15, UL27, UL35 and UL55 proteins also have a close relationship with Mardivirus [10,31-35], we presumed DEV may be one

member of *Mardivirus* or belong to an individual genus within the *Alphaherpesvirinae* subfamily based on these results. More researches are required to define which genus of herpesvirus the DEV belongs to.

In short, bioinformatics analysis of DEV UL21 protein by some software and online tools provides some important information about the molecular characteristics. Our work provided some basic information for the further DEV UL21 protein research.

Acknowledgment

The research was supported by China 973 program (2011CB111606, China Agricultural Research System(CARS-43-8), Changjiang Scholars and Innovative Research Team in University(PCSIRT0848) and National science and technology support program for agriculture (2011BAD34B03) An-chun Cheng and Ming-shu Wang are the corresponding authors. Institute of Preventive Veterinary Medicine, Sichuan Agricultural University, Wenjiang, Chengdu city, Sichuan, 611130, P.R.China. Avian Disease Research Center, College of Veterinary Medicine of Sichuan Agricultural University, 46# Xinkang Road, Ya'an, Sichuan 625014, P.R. China. Key Laboratory of Animal Disease and Human Health of Sichuan Province, Sichuan Agricultural University, Wenjiang, Chengdu city, Sichuan, 611130, P.R.China.. Tel.: +86 835 2885774; fax: +86 835 2885774; E-mail address: chenganchun@vip.163.com (A. Cheng); mshwang@163.com (M. Wang).

References

- [1] Thomas N. Tully, Gerry M. Dorrestein, Saunders Ltd. .Waterfowl. Handbook of avian medicine (Second Edition), USA: Saunders Ltd,2009.
- [2] Sandhu TS, Metwally SA. Diseases of poultry.Blackwell Publishing, 2008
- [3] Fauquet CM, Mayo MA, Maniloff J . Virus Taxonomy: Eighth report of the international committee on taxonomy of viruses, Elsevier Academic Press, California, 2005
- [4] Yang F, Jia WX, Yue H, et al. Development of quantitative real-time polymerase chain reaction for duck enteritis virus DNA. Avian Dis,AAAP.San Jose Blvd 2005;49:397-400,
- [5] Jia RY, Cheng AC, Wang MS, et al. Studies on ultrastructure of duck enteritis virus chv virulent strain”,Chinese J.Virol 2007;23:202-6,.
- [6] Yuan GP, Cheng AC, Wang MS, et al. Electron microscopic studies of the morphogenesis of duck enteritis virus”,Avian Dis, AAAP, San Jose Blvd 2005;49:50-5
- [7] Pan H, Cao R, Liu L, et al. Prokaryotic expression of N-terminal antigenic domain of duck plague virus gB protein and the establishment of putative indirect ELISA assay. Wei Sheng Wu Xue Bao. 2008;48:98-102.
- [8] Xie W, Cheng AC, Wang MS. Expression and characterization of the UL31 protein from duck enteritis virus. Virology Journal 2009;6, doi:10.1186/1743-422X-6-19
- [9] Xiang J, Cheng AC, Wang MS.Molecular Cloning and sequence analysis of the Duck Enteritis Virus Nucleocapsid Gene UL38. Biomedical Engineering and Informatics, 2009. BMEI '09. 2nd International Conference on, 17-19 Oct. 2009
- [10] Li LJ, Cheng AC, Wang MS.Molecular Cloning and sequence analysis of the Duck Plague Virus gI Gene, Bioinformatics and Biomedical Engineering (iCBBE), 2010 4th International Conference on, 18-20 June 2010
- [11] Valerio V, Eve D. Determination of Interactions between tegument proteins of herpes simplex virus type 1. Journal of virology 2005;79:9566–71
- [12] Li YF , Huang B.Molecular characterization of the genome of duck enteritis virus. Virology 2009;391:151–61
- [13] Hiroki Takakuwa, Fumi Goshima .Herpes simplex virus encodes a virion-associated protein which promotes long cellular processes in overexpressing cells.Genes to Cells 2001: 6 : 6955-66

- [14] Baines JD, Koyama AH, Huang T, et al. The UL21 gene products of herpes simplex virus1 are dispensable for growth in cultured cells. *Virology*,1994;68: 2929-36
- [15] Klupp BG, Bottcher S, Granzowet H ,et al. Complex formation between the UL16 and UL21 Tegument Proteins of Pseudorabies Virus. *Journal of virology* 2005;79 : 1510–22
- [16] Klupp BG, Lomniczi B, Visser N, et al. Mutations affecting the UL21 gene contribute to avirulence of pseudorabies virus vaccine strain Bartha. *Virology* 1995: 212:466-73
- [17] Wind DN., Wagenaar F, Pol J, et al. The pseudorabies virus homology of the herpes simplex virus UL21 gene product is acapsid protein which is involved in capsid maturation. *Virol* 1992;66 : 7096-103
- [18] Klop RF, Klupp BG, Fuchs W, et al. Influence of Pseudorabies Virus Proteins on Neuroinvasion and Neurovirulence in Mice. *Journal of virology* 2006;80 : 5571–6
- [19] Frans W, Jan MA. Deletion of the UL21 gene in Pseudorabies virus results in the formation of DNA-deprived capsids:an electron microscopy study. *Vet. Res*,2001: 32:47–54
- [20] Amy LH, David G. Meckes Jr, et al. Interaction domains of the UL16 and UL21 tegument proteins of herpes simplex virus. *Journal of virology*, 2010;84 :2963–71
- [21] Pei CY, David G. Meckes, Jr, et al. Analysis of the interaction between the UL11 and UL16 tegument proteins of herpes simplex virus. *Journal of virology* 2008;82: 10693–700
- [22] David G, Meckes Jr., Jacob AM, et al .Complex mechanisms for the packaging of the UL16 tegument protein into herpes simplex virus. *Virology* 2010;398:208–13
- [23] David G, Meckes, Jr., John WW. Dynamic Interactions of the UL16 Tegument Protein with the Capsid of Herpes Simplex Virus. *Journal of virology* 2007;81: 13028-36
- [24] Michael K, Klupp BG, Karger A, et al. Efficient Incorporation of Tegument Proteins pUL46, pUL49, and pUS3 into Pseudorabies Virus Particles Depends on the Presence of pUL21. *Journal of virology* 2007: 81 :1048-51
- [25] Barbara J. Kellya, Cornel Fraefelb, Anthony L. Functional roles of the tegument proteins of herpes simplex virus type 1. *Virus Research* 2009: 145: 173-86
- [26] Macek B, Gnad F, Soufi B. Phosphoproteome analysis of E. coli reveals evolutionary conservation of bacterial Ser/Thr/Tyr Phosphorylation. *Biochemistry and Molecular Biology*,2007;7:299-307
- [27] Xu C, Li XR, Xin HY, et al. Cloning and molecular characterization of gC gene of duck plague virus. *Chinese Veterinary Science* 2008;38:1038-44.
- [28] Sun T, Cheng AC, Wang MS, et al. Prediction of epitopes on B cell of UL6 gene of duck enteritis virus and prokaryotic expression of major antigen determinant sequence. *Veterinary Science in China* .2008;.38:939-45
- [29] Barlow DJ, Edwards MS ,Thornton JM. Continuous and discontinuous protein antigenic determinants. *Nature* 1986;322:747-48
- [30] David MK..Recognition of Herpes Simplex Virus Type 2 Tegument Proteins by CD4 T Cells Infiltrating Human Genital Herpes Lesions. *Journal of virology*1998;72:7476-83
- [31] Zhao LC, Cheng AC, Wang MS ,et al.Characterization of codon usage bias in the dUTPase gene of duck enteritis virus,” *Progress in Natural Science* 18 (2008) 1069–1076.
- [32] Cai MS, Cheng AC, Wang MS, et al. Characterization of synonymous codon usage bias in the Duck Plague virus UL35 Gene,*Intervirology* 2009;52:266–278.
- [33] Jiang L, Lin D, Cheng AC, et al. Bioinformatic analysis of UL27 gene of Duck Plague Virus CHv Strain. *Bioinformatics and Biomedical Engineering (iCBBE)*, 2010 4th International Conference on, 2010
- [34] Zhu HG, Li HX, Han ZX. Identification of a spliced gene from duck enteritis virus encoding a protein homologous to UL15 of herpes simplex virus 1. *Virology Journal* 2011, 8:156
- [35] Wu Y, Cheng AC, Wang MS. Molecular characterization analysis of newly identified Duck Enteritis Virus UL55 Gene. *Bioinformatics and Biomedical Engineering (iCBBE)*, 2010 4th International Conference on, 18-20 June 2010