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## ORIGINAL ARTICLE

# Suite of decision tree-based classification algorithms on cancer gene expression data

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## KEYWORDS

DNA microarray; Cancer; Classification; Decision trees; Ensample decision tree; Attribute selection Abstract One of the major challenges in microarray analysis, especially in cancer gene expression profiles, is to determine genes or groups of genes that are highly expressed in cancer cells but not in normal cells. Supervised machine learning techniques are used with microarray datasets to build classification models that improve the diagnostic of different diseases. In this study, we compare the classification accuracy among nine decision tree methods; which are divided into two main categories; the first is single decision tree C4.5, CART, Decision Stump, Random Tree and REPTree. The second category is ensample decision tree such Bagging (C4.5 and REPTree), AdaBoost (C4.5 and REPTree), ADTree, and Random Forests. In addition to the previous comparative analyses, we evaluate the behaviors of these methods with/without applying attribute selection (A.S.) techniques such as Chi-square attribute selection and Gain Ratio attribute selection. Usually, the ensembles learning methods: bagging, boosting, and Random Forest; enhanced classification accuracy of single decision tree due to the natures of its mechanism which generate several classifiers from one dataset and vote for their classification decision. The values of enhancement fluctuate

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between (4.99–6.19%). In majority of datasets and classification methods, Gain ratio attribute selection slightly enhanced the classification accuracy (~1.05%) due to the concentration on the most promising genes having the effective information gain that discriminate the dataset. Also, Chi-square attributes evaluation for ensemble classifiers slightly decreased the classification accuracy due to the elimination of some informative genes.

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#### 1. Introduction

The genome ribonucleic acid (RNA) expression studies allow systematic approaches to understand the relationship between gene expression profiles and disease states or different developmental stages of a cell. Microarray analysis provides quantitative information about the whole transcription profile of cells that make possible drug and therapeutics improvement, disease diagnosis, and comprehensible basic cell biology.

A DNA microarray technique allows to simultaneously observing the expression levels of thousands of genes during significant biological processes and across collections of related samples [1].

The datasets from microarray analysis, that enables the measurement of molecular signatures of diverse cells, becomes an important application of data mining, artificial intelligence and machine learning techniques to provide bioinformatics knowledge. In practical, supervised machine learning techniques used with microarray datasets to build classification models that improve the diagnostic of different diseases which easy to interpreted [2,3].

#### 1.1. Biological background

Cells are the fundamental working units of every living system. All the instructions needed to direct their actions are contained within the chemical deoxyribonucleic acid or shortly DNA. A DNA molecule is a double-stranded polymer composed of four basic molecular units namely nucleotides. The nitrogen bases include adenine (A), guanine (G), cytosine (C) and thymine (T). The genome provides a template for the synthesis of a variety of RNA molecules. The process of transcribing a gene's DNA sequence into RNA is called gene expression. A gene's expression level indicates the approximate number of copies that gene's RNA produced in a cell and it is correlated with the amount of the corresponding proteins made. This mechanism controls which genes are expressed in a cell and acts as a "volume control" that increases or decreases the level of expression of particular genes as necessary [4].

## 1.2. Microarray data format

A gene expression data set from a microarray experiment can be represented by a real-valued.

Expression matrix =  $\{G(i,j)|1 \le i \le n, 1 \le j \le m\}$ 

where the columns  $G = \{\vec{g_1}, \vec{g_2}, \dots, \vec{g_m}\}$  form the expression patterns of genes, the rows  $S = \{\vec{S_1}, \vec{S_2}, \dots \vec{S_n}\}$ .

An example of a gene expression microarray dataset for Leukemia is shown in Table 1. the table organizes data into m columns (genes) and n rows (samples) where m mostly varies from thousand to hundred thousand according to the accuracy

Table 1	ble 1 Microarray data decision table.						
Samples	Attributes (genes)				Category		
	Gene 1	Gene 2		Gene m	<u> </u>		
1	G(1,1)	G(1,2)		G(1,m)	ALL		
2	G(2,1)	G(2,2)		G(2,m)	ALL		
					ALL		
					AML		
n	G( <i>n</i> ,1)	G( <i>n</i> ,2)		G(n,m)	AML		

of microarray image processing technique, while *n* is always less than 200 samples according to the previously collected datasets [5]. Category column presents the actual class of the sample. For the shown example AML stands for acute myeloid leukemia disease and ALL represents acute lymphoblastic.

Our study provides a performance comparison of nine decision tree methods. The rest of this paper is organized as the follows. In Section 2, we present brief challenges that faced in cancer classification area. In Section 3, we provide problem definitions. In Section 4, we exploit decision tree and microarray classification. In Section 5, we discuss related works in this domain. In Section 6, we explore the methodologies used in this work. In Section 7, we describe experimental setup. In Section 8, we present results and analysis. In Section 9, we conclude the paper.

## 2. Cancer classification challenges

Gene classification as domain of research poses a new challenges due to its unique problem nature. First, challenge comes from the unique nature of the available gene expression dataset; where most of these datasets has sample size below 200, vs. thousands to hundred thousands of genes presented in each tuples. Second, only a few numbers of these (genes) presents relevant attributes to the investigated disease. Third, comes from the presence of noise (biological and technical) inherent in the dataset. Fourth challenge arises from the application area, for instance accuracy is an important criterion in cancer classification task, but it is not the only goal, in cancer domain we want to achieve, biological relevancy as well as classification accuracy.

#### 3. Problem definition

There is no single classifier superior over the rest, for instance the classification accuracy is depend on the classification method, gene selection method, and dataset [7,8].

In this study we will use the notation provided by Ying Lu et al. [9].

Let  $X_1, X_2, ..., X_m$  be random variables for genes  $G_1$ ,  $G_2, ..., G_m$  respectively, where  $X_i$  has domain  $dom(X_i)$  which is the range of expression values for  $gene\ G_i$ .

Table 2 Confusion	n matrix.		
	Predicted	class	
Actual class		C1	C2
	C1	TP	FN
	C2	FP	TN

Let C be the random variable for the class labels, and  $dom(C) = \{1, ..., K\}$ , where K is denotes a total number of classes.

Let  $t = \{t.X_1, t.X_2, ..., t.X_m\}$  denotes a size m tuple of expression values for m genes. Let  $T = \{(t_1, c_1), (t_2, c_2)... (t_n, c_n)\}$  Denoting a training set of n tuples, where  $i = \{1, 2, ..., n\}, c_i \in dom(C)$  is the class label of tuple  $t_i$ .

Let the test set be  $S = \{t_1, t_2, \dots, t_l\}$  where *l* is the size of the test set.

Find a classification function Class, which gave maximal classification accuracy on *S*, where the classification accuracy calculated by divide number of correct classified instances on total number of instances.

$$Accuracy = \frac{TP + TN}{TP + FN + FP + TN}$$
 (1)

- True positive (TP) = the number of predicted positive cases that are actually positive.
- True negative (TN) = the number of predicted negative cases that are actually negative.
- False positive (FP) = the number of predicted positive cases that are actually negative.
- False negative (FN) = the number of predicted negative cases that are actually positive. Table 2 illustrates the confusion matrix for positive and negative tuples.

## 4. Decision tree and microarray classification

One of the main advantages of decision trees is the ability to generate understandable knowledge structures, i.e., hierarchical trees or sets of rules, a low computational cost when the model is being applied to predict or classify new cases, the ability to handle symbolic and numeric input variables, provision of a clear indication of which attributes are most important for prediction or classification [10].

There are two disadvantages that also represented a major weakness in using decision tree for microarray analysis problems in the past. The first one is their instability that is tightly connected with the second disadvantage - i.e., difficulties to branching the trees when the number of samples is too low. Instability of decision trees was successfully solved by ensembles methods where multiple trees built from different subsets of the initial dataset were built to improve the robustness of the final classifier. Unfortunately, ensembles of classifiers posses very low level of their knowledge understand-ability and are not appropriate for interpretation of the acquired knowledge. There were some studies that approached the problem of knowledge extraction from ensembles of classifiers [11], but all of them are too limited to be helpful for practical use. Quality of branching in decision tree is of fundamental importance to the final success of classifier.

Due to high cost per experiment in microarray studies it is nowadays still acceptable for studies with 100 or yet less samples to represent benchmarking datasets for evaluation of the most complex classifiers [12]. Most of the microarray data to-day is collected in centralized repositories containing large numbers of samples like Gene Expression Omnibus (GEO) by National Center for Biotechnology Information (NCBI) or ArrayExpress by European Bioinformatics Institute (EBI) [13]. Unfortunately such repositories are too large and contain data coming from various sources using different protocols to serve as a benchmarking collection of datasets. Our study takes advantage of one of the largest publicly available repositories of gene expression measurements that were collected by EBI. Which is currently one of the most appropriate collections of gene expression samples for evaluation of classification methods [11].

#### 5. Related works

There is a numerous algorithms produced to construct classification models from the old nearest neighbor analysis and decision tree to the new SVM support vector machines [6]. For instance, Xiaosheng Wang et al. [5] compare the performance of NB (Naive Bayes), DT (Decision Tree), SVM Support Vector Machine) and k-NN (k-nearest neighbor) algorithm with several attribute selection (chi-square, information gain, Relief-F and symmetric uncertainty) the average accuracy in their study was between 69.33% and 90.01%. Peter et al. [14] uses Partial Least-Squares (PLS) regression as a feature selection method, and compare performance of several ensemble models the predictive accuracy in their study was between 61.2 and 99.4. Hong Hu et al. [7] provide new ensample method and compare it with several famous ensample method, the accuracy was between 60% for Prostate dataset and 98.9% for Lung Cancer dataset. Aik Choon et al. [15] comparisons, between single decision tree algorithm and ensemble based decision tree (Bagging, AdaBoost) Algorithm. The accuracy was between 52.38% and 93.29%. Table 3 shows the relevant works on cancer classification.

## 6. Methodologies

## 6.1. Classification methods

In this experimental study we focus on nine public decision tree methods, some of these methods build single decision tree such as C4.5, CART, REPTree, RandomTree, and Decision-Stump. The other are ensample decision tree such as ADTree, Random Forests, Bagging, and AdaBoost. These methods are described briefly as the following:

C4.5 algorithm top-down decision tree base proposed by Quinlan [16]. The algorithm is a successor of ID3, which determines at each step the most predictive attribute, and splits a node based on this attribute. Every node represents a decision point over the value of some attribute.

The split criterion calculated as the following:

 Calculate the expected information needed to classify a tuple in D

$$Info(D) = \sum_{i=1}^{m} p_i \log_2(p_i)$$
 (2)

where,  $p_i$  is the probability that an tuple in D belong to class  $C_i$ .

Authors	Dataset	Att. Sel.	Classifier accuracy [%	]		
Xiaosheng Wang et al. [5]	Colon		NB	C4.5	SVM	k-nn
		Chi	88.71	90.32	87.1	87.1
		Inf.	85.48	85.48	87.1	87.1
		RF	87.1	85.48	87.1	87.1
		SU	87.1	91.94	87.1	88.71
			Highest accuracy		Average accuracy	
	Colon		91.94		83.1074	
	Cns		90		72.3362	
	DLBCL		87.93		70.7054	
	Leukemia1		97.22		92.013	
	Lung		100		97.9547	
	Prostate		96.08		90.9766	
	Breast		88.46		69.3911	
	Leukemia2		98.25		87.593	
Peter J. Tan et al. [14]	Dataset		single C4.5	RF	Ad C5.0	MML Oblique Forest
	Leukemia	PLS dimensionality	94.3	96.2	95.7	96.7
	Breast	·	65.2	71.2	67.9	69.2
	Central nervous		61.2	64.5	63.2	65.9
	Colon		80.9	84.7	82.7	88.8
	Lung		98	96.2	98.2	99.4
	Prostate		83	90.6	88.1	91.3
	Prostate		65	69.3	51.5	53.2
Hong Hu. et al. [7]	Dataset		C4.5	RF	Ad C4.5	Ba C4.5
	Breast		62.9	61.9	61.9	66
	Lung		95	98.3	96.1	97.2
	Lymphoma		78.7	80.9	85.1	85.1
	Leukemia		79.2	86.1	87.5	86.1
	Colon		82.3	75.8	77.4	82.3
	Ovarian		95.7	94.1	95.7	97.6
	Prostate		33.3	52.4	33.3	42.9
Aik Choon Tan et al. [15]	Dataset		C4.5	Ba C4.5	Ad C4.5	
	Leukemia		91.18	91.18	91.18	
	Breast		63.16	89.47	89.47	
	tumors outcome		85	88.33	88.33	
	Colon		95.16	93.55	90.32	
	Lung		92.62	93.29	92.62	
	Prostate		67.65	73.53	67.65	
	Prostate outcome		52.38	85.71	76.19	

- Calculate the expected information required to classify a tuple from *D* based partitioning by *A*.

$$Info_A(D) = \sum_{i=1}^{m} \frac{|D_i|}{|D|} \times Info(D_i)$$
(3)

The term  $\frac{|D_j|}{|D|}$  acts as the weight of the *j*th partition.

- Calculate information gain of attribute A.

$$Gain(A) = Info(D) - Info_A(D)$$
 (4)

- Calculate split information of attribute A

$$SplitInfo_{A}(D) = \sum_{i=1}^{\nu} \frac{\mid D_{j} \mid}{\mid D \mid} \times log_{2}\left(\frac{\mid D_{j} \mid}{\mid D \mid}\right)$$
 (5)

- Calculate gain ratio:

$$GainRation(A) = \frac{Gain(A)}{SplitInfo_{A}(D)}$$
 (6)

The attribute with the maximum gain ratio is selected as best splitting attribute.

CART (Classification and Regression Tree), it is analysis which is based on the paper by Breiman et al. [17]. Is binary decision trees, which split a single variable at each node. CART approach can also produce classification trees, which depends on the type of the dependent variable (categorical or numerical) CART Similar to C4.5 but use Gini index as split criteria that calculated as the following

$$Gini(D) = 1 - \sum_{i=1}^{m} p_i^2$$
 (7)

The Gini index of attribute A for a binary spilt is calculated as the following:

$$Gini_A(D) = \frac{|D_2|}{|D|}Gini(D_1) + \frac{|D_2|}{|D|}Gini(D_2)$$
 (8)

RandomTree: is constructing a tree that randomly selected attributes at each node. It performs no pruning.

DecisionStump: is an algorithm builds simple binary decision 'stumps' (1 level decision tress) for both nominal and numeric classification task. It deal with mission values by extending a third branch from the stump or treating 'missing' as a separate attribute value. It does regression (based on mean-squared error) or classification (based on entropy) [18].

REPTree: algorithm is a fast decision tree learner it is also based on C4.5 algorithm and can produce classification (discrete outcome) or regression trees (continuous outcome). It builds a regression/decision tree using information gain/variance and prunes it using reduced-error pruning (with backfitting).

ADTree: Applied Alternating decision trees, it is a generalization of decision trees, voted decision trees and voted decision stumps. The algorithm boosting procedures to decision tree algorithms to produce accurate classifiers. The classifiers are in the form of a majority vote over a number of decision trees but having a smaller and easier to understand classification rules [19].

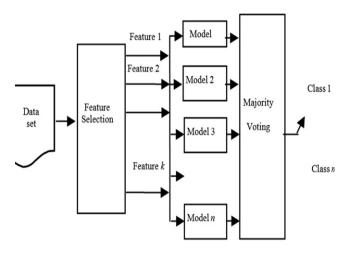


Figure 1 Overview of the ensemble classifier.

Random forests: ensemble decision tree methods by resampling attributes, produced by Leo Breiman [19]. This early random decision trees method combines bagging and random feature selection methods to generate multiple classifiers. Random Forests based on CART method.

Bagging: produced by Leo Breiman [20,21] it uses a bootstrap technique to resample the training data sets D. To form a resampled data set  $D_i$ . Each sample in D has a probability of 1/n of being drawn in any trial. The most often predicted class label will be the final classification result.

AdaBoost: The Boosting method was first developed by Freund [22]. The initial classifier is constructed from the original data set where every sample has an equal distribution ratio of 1. In the Boosting method training data set  $D_i$ , the distribution ratios are made different among samples depending on their prediction accuracy in the previous data set  $D_{i-1}$ . If a sample has a lower prediction accuracy rate in  $D_{i-1}$ , it will be given a higher weight in  $D_i$  and therefore get a higher possibility to be selected in  $D_i$ . Fig. 1 illustrates an overview of the ensemble classifier.

## 6.2. Data preprocessing

Among thousands of genes whose expression levels are measured, not all are wanted for classification. We need to select a few numbers of genes highly related with classes for classification, which is called informative genes.

Chi-square  $(\chi^2)$  attributes evaluate. The chi-square  $(\chi^2)$  method evaluates features individually by measuring their chi-squared statistic with respect to the classes. The  $\chi^2$  value

$$\chi^{2}(a) = \sum_{\nu=\nu} \sum_{i=1}^{n} \frac{[A_{i}(a=\nu) - E(a=\nu)]2}{E_{i}(a=\nu)}$$
(9)

where V is the set of possible values for a, n the number of classes,  $A_i(a = V)$  the number of samples in the ith class with a = v, and  $E_i(a = v)$  the expected value of  $A_i(a = v)$ ;

 $E_i(a = v) = P(a = v)P(c_i)N$ , where P(a = v) is the probability of a = v,  $P(c_i)$  the probability of one sample labeled with the *i*th class, and N the total number of samples [23].

**Table 4** Description of microarray dataset where AML: acute myeloid leukemia, ALL: acute lymphoblastic leukemia, AD: adenocarcinomas, SQ: squamous cell carcinomas, COID carcinoids, and NL: normal lung. BR: Breast, PR: prostate, LN: lung, CO: colon, PA: patient, CO: control.

Dataset	Sample no.	Gene no.	Category				
			Tumor		Normal		
Breast	62	16383	43		19		
Breast1	86	16382	43		43		
Colon	36	7458	18		18		
Lung2	88	16382	69		19		
Prostate	146	12626	65		81		
Prostate2	108	12554	92		16		
Prostate3	188	16383	92		96		
Lung1	197	10937	AD	NL	SQ	COID	
-			139	17	21	20	
Multi tissues	103	16383	BR	PR	LN	CO	
			26	26	28	23	
Leukemia	72	7130	ALL		AML		
			46		26		
Lymphoma	60	16381	PA		CO		
-			40		20		

Gain ratio features evaluation. Expected information (entropy) is used to sort the attribute in D, according to higher informative gene and eliminate the attribute with low gain ratio see Eqs. (7) and (8).

#### 6.3. Datasets

In this study, we select eleven microarray datasets these dataset were breast cancer, breast1 cancer, lung2 cancer, prostate cancer (normal prostate tissue adjacent to tumor vs. tumor tissue), prostate2 cancer, Prostate3 cancer, and lymphoma, these dataset were collected from EBI,

http://www.ebi.ac.uk/array-express and multi tissues [24], lung1 cancer [24], colon tumor [25], Leukemia [24], were collected from GEO repository. The latest four dataset had been extensively tested in many previous studies [25–27]. Table 4 provides briefly description of microarray dataset.

#### 7. Experimental setup

Eleven dataset was collected from EBI and GEO, used to evaluate classification performance. These datasets were described briefly in Table 3.

All experiments described in this paper were performed using libraries from Weka 3.7.1 machine learning environment [28]. A lot of studies used Weka in classification task, for examples [26,29]. Nine selected decision tree classifiers are used to build the classification models, these classifier was briefly described above (Section 6.1), as well as two type of attribute selection (Chi-square, Gain Ratio) were used to reduce the initial set of available genes, and evaluate the performance of these methods, on elected subset of attribute.

Each classification method was used "as it is" in Weka environment which means that no additional parameter tuning

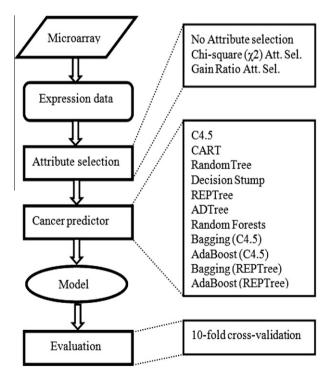


Figure 2 Cancer classification system.

was performed before or during classification performance comparison. Each feature selection method was used in combination with all nine classification models included in comparison. As well as we evaluate AUC of the classification method using, each test we used 10-fold cross-validation. For bagging and AdaBoost method we used (C4.5) and REPTree as a classifiers and applying 10 iterations each experiment.

We summarize our machine learning work for DNA microarray in three main stages. First stage is attribute selection, second is choosing appropriate predictor, and third is produced model evaluation as shown in Fig. 2.

## 8. Results and analysis

We report the results from nine methods that used to study the usefulness of the decision tree to predict disease status. For each method we evaluated classification accuracy on the original dataset (without attribute selection) and also by using Chisquare and Gain ratio as attribute selection.

First, we applied the methods on breast cancer Fig. 3 shows the accuracy of the methods. We notes that Random Forest, AdaBoost (C4.5), and AdaBoost (REPTree) gave higher accuracy on original dataset (98.39%), AdaBoost (C4.5) perform higher accuracy with Chi-square attribute selection (96.77) and Gain ratio attribute selection (98.39%).

Second, we applied the methods on colon cancer Fig. 4 shows accuracy of the methods. We notice that AdaBoost (C4.5) perform higher accuracy on original dataset (100%), and RandomForest, CART, AdaBoost(C4.5), AdaBoost(REPTree) and Bagging(REPTree) gave the same and higher accuracy with Gain ratio attribute selection (97.22%), while DecisionStump gave higher accuracy with Chi-square attribute selection (97.22%).

Third, we applied the methods on leukemia dataset; Fig. 5 shows the accuracy of the method. We notice that Bagging(REPTree) perform higher accuracy on original dataset (93.06%), AdaBoost(REPTree) perform higher accuracy with Chi-square Attribute selection (95.83%) and gain ratio attributes selection (97.22%).

Fourth, we applied the method on lung1 dataset Fig. 6 shows the accuracy of the methods, we notice that Ada-Boost(REPTree) gave higher accuracy on the original dataset (95.43%) and with Chi-square Attribute selection (94.42%), while Bagging(C4.5) gave higher accuracy with Gain ratio attribute selection (93.91%).

Fifth, we applied the methods on lung2 dataset Fig. 7 shows the accuracy of the methods, we notice that CART, Ada-Boost(C4.5) gave higher accuracy on the original dataset (97.73%), while CART gave higher accuracy with Gain ratio attribute selection (98.86%), and AdaBoost(C4.5) gave higher accuracy with Chi-square Attribute selection (97.73%).

Sixth, we applied the methods on lymphoma dataset Fig. 8 shows the accuracy of the method, we notice that Random Forest, Bagging(REPTree) and AdaBoost(C4.5) gave high performance on original dataset(100%), while ADTree, Decision-Stump, CART AdaBoost(C4.5), Bagging(REPTree), and REPTree gave accuracy with Gain ratio attribute selection (100%) while Random Forest AdaBoost(C4.5) gave higher accuracy with Chi-square attribute selection (100%).

Seventh, we applied the methods on prostate dataset Fig. 9 shows the accuracy of the methods, we notice that

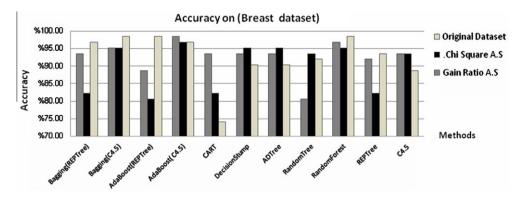


Figure 3 Accuracy of the methods on breast dataset.

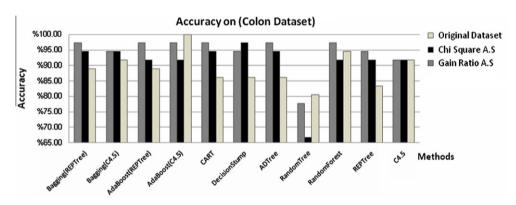


Figure 4 Accuracy of the methods on colon dataset.

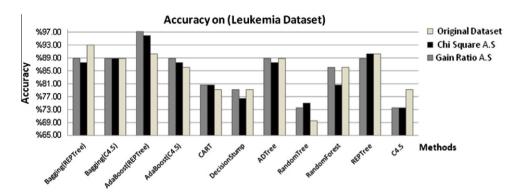


Figure 5 Accuracy of the methods on leukemia dataset.

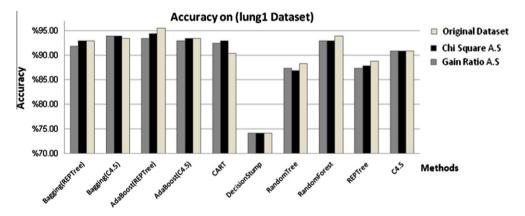


Figure 6 Accuracy of the methods on lung1 dataset.

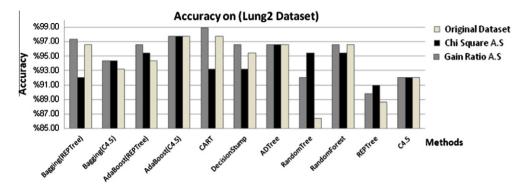


Figure 7 Accuracy of the methods on lung2 dataset.

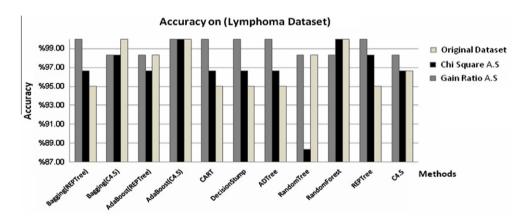


Figure 8 Accuracy of the methods on lymphoma dataset.

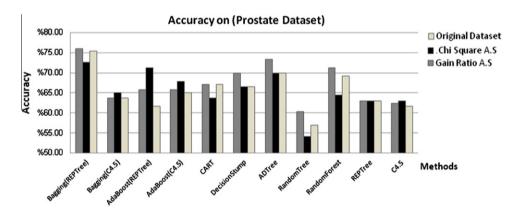


Figure 9 Accuracy of the methods on prostate dataset.

Bagging(REPTree) gave higher accuracy on the original dataset (75.34%), with Chi-square Attribute selection (72.60%) and Gain ratio attribute selection (75.92%).

Eigth, we applied the methods on prostate2 dataset Fig. 10 shows the accuracy of the methods, we notice that Ada-Boost(C4.5), AdaBoost(REPTree) and Bagging(C4.5) gave higher accuracy on the original dataset (91.67%), Bagging(C4.5) gave higher accuracy with Chi-square Attribute selection (90.74%) and Gain ratio attribute selection (91.67%), also we applied the method on the rest datasets. Table 5 summarized the minimum and maximum accuracy of these methods.

By calculating the average accuracy for all methods on each dataset, from Fig. 11 we notice AdaBoost (C4.5) gave higher average accuracy on original dataset (91.23%) and with using Chi-square Attribute selection (90.83%) and Bagging (REP-Tree) gave higher average accuracy Gain ratio attribute selection (92.64%).

Also, we notice the ensamples decision tree is significantly improving the accuracy of single decision tree classifier such as (C4.5 and REPTree).

In this study we intended to use several dataset from the same cancer types to focus on the following facts: it is important to classify the samples produced from the same

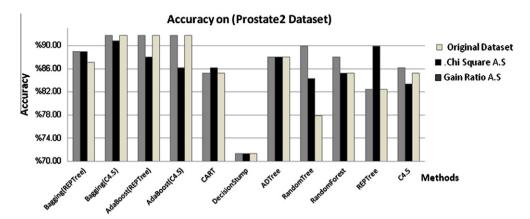


Figure 10 Accuracy of the methods on prostate2 dataset.

		Gain ratio A. S.		Chi-square A. S.		Original data	
		Method	%	Method	%	Method	%
Breast1	Max. Acc.	Ad(C4.5)	98.39	Ad(C4.5)	86.05	Ba(REPTree)	81.40
	Min. Acc.	RT	67.44	ADTree	75.58	RT	77.91
Multi tissues	Max. Acc.	Ad(C4.5)	97.09	Ad(REPTree)	95.15	CART	94.17
	Min. Acc.	DS	44.66	DS	47.57	DS	46.60
Prostate3	Max. Acc.	RF	98.94	RF	100	Ad(C4.5)	99.47
	Min. Acc.	ADTree	86.70	DS	86.7	DS	86.70

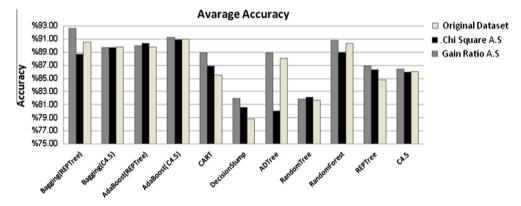


Figure 11 Average accuracy of the methods for all dataset.

laboratory, as well as the same preprocessing algorithm, and same microarray type.

#### 9. Conclusions

This experimental study compares classification performance of different nine decision tree algorithms via using of eleven cancerous microarray datasets. These algorithms include five single decision tree (C4.5, REPTree, CART, DecisionStump, and RandomTree) as well as four ensample decision tree methods AdaBoost (C4.5), AdaBoost (REPTree), Bagging (REPTree), Bagging (C4.5), ADTree. In addition, the effect of attribute selection on building decision tree based models is investigated.

From the obtained results we can highlights some interesting conclusions: The ensample method (AdaBoost, Bagging and Random Forest) significantly improve the classification accuracy of single decision tree, Due to building several classifier and voting techniques. Accuracy of AdaBoost (C4.5) outperforms other ensample methods on original dataset without using of attribute selection. AdaBoost (REPTree), AdaBoost (C4.5) outperform other method with Chi-square attribute evaluation, Bagging (REPTree) outperform other method with Gain Ratio attribute.

Gain ratio attribute selection significantly improves classifier accuracy on majority of the dataset and classification method, but chi-square attribute selection didn't significantly improve the classification accuracy, as well as it decreases

classifier accuracy such as C4.5, due to the elimination of some informative genes. By analyzing the results obtained with multiclass microarray datasets (lung1. Multi-tissues dataset) we noticed that CART and AdaBoost (C4.5) outperform other methods on the original dataset. While AdaBoost (REPTree) outperforms with Chi-square attribute selection. Usually, Bagging (C4.5) and AdaBoost (C4.5) outperform with Gain ratio attribute selection.

Lastly, decision trees are particularly attractive for biologists due to their interpretability, being able to highlight which genes are actually influencing the classification task as well as results show that decision tree classifiers might play an important role in microarray analysis in the future.

But small numbers of genes shared to constrict tree from microarray dataset is still critical criteria because missing such genes mean missing classification result.

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