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## Diphyllin: An effective anticandidal agent isolated from *Cleistanthus* collinus leaf extract



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#### ABSTRACT

In this study, diphyllin [9-(1,3-benzodioxol-5-yl)-4-hydroxy-6,7-dimethoxynaphtho[2,3-c]furan-1(3H)-one] was isolated from *Cleistanthus collinus* leaf extract. The isolated compound and leaf extract were evaluated for their *in vitro* anticandidal activity against Candida strains such as *Candida albicans*, *C. tropicalis*, and *C. glabrata*. Diphyllin was found to possess higher anticandidal activity against various *Candida* species with the Minimal Fungicidal Concentration (MFC) of 85–145  $\mu$ g and inhibition zone of 9.5  $\pm$  0.5–13.5  $\pm$  0.5 mm at 200  $\mu$ g concentration against the yeast pathogens studied. Thus, diphyllin was twice more active than miconazole against *C. glabrata*.

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#### 1. Introduction

In recent years, candidiasis is a major fungal infection caused by *Candida* species in humans and veterinary animals. Among them, 90% of nosocomial candidemia cases were due to *C. albicans* as causative agent associated with other candidal species such as *C. glabrata*, *C. tropicalis*, *C. parapsilosis*, *C. dubliniensis*, and *C. krusei* in the subcontinent [1–3]. In recent years, although a number of synthetic and natural derivative antifungal drugs developed in pharmacological industries were effective in controlling *Candida* infections, the toxicity, high cost, side effects, and development of drug-resistant strains due to frequent use of the drugs have led to several problems in candidiasis management [4–6]. Henceforth, a plant-derived novel agent with low toxicity and side effects has been examined to overcome and enhance the efficiency of treatment of fungal infections [7,8].

Anticandidal activities of plant extracts, oils, toxicants, metals, synthetic drugs, and natural products have been reported by many researchers and the frequency of discovery of new antifungal agents from plant sources emphasizes the increasing interest in the broad spectrum of activity against *Candida* species [9,10].

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Bioactive compounds are extracted from aromatic, toxic, and medicinal plants in pure or crude forms were considered recently as effective agents in controlling bacterial and fungal pathogens. Review of literature and the examination of botanicals against *Candida* species were significantly increased in the last decade [11,12]. The population of the Indian subcontinent has been traditionally using many plants as medicine for treatment of several microbial infections.

Cleistanthus collinus (Euphorbiaceae) is distributed in Asian countries with many potential pharmacological properties [13–16]. In this work, anticandidal activity of *C. collinus* leaf extract and its fraction against *C. albicans, C. tropicalis*, and *C. glabrata* have been examined. To the best of our knowledge, no study has been investigated the inhibitory effects of *C. collinus* extract and its fraction against different *Candida* species till date.

#### 2. Materials and methods

#### 2.1. Preparation of extracts

*C. collinus* samples were collected from Viralimalai, Tamil Nadu, India, in August 2011. The plant leaves were carefully separated and washed with running tap water and subsequently with distilled water to remove pollutants. The samples were shade-dried and minced to precede extraction. About 2 kg dried plant material

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was subjected to crude extract preparation using Soxhlet apparatus (Sigma Soxhlet Mantle, Tamil Nadu, India). Distilled water and ethyl acetate (Merck, Darmstadt, Germany) were used as solvents. Crude extracts were concentrated under reduced vacuum and stored for further analysis.

#### 2.2. Isolation and characterization of diphyllin

About 87 g of ethyl acetate extract was obtained and exactly 7 g was mixed with activated silica and filled at the top of the column. It was then subjected to first elution with 50 ml toluene. Thereafter, toluene was mixed with ethyl acetate in different ratios (9:1–1:9, and 0:10). Eighty-one fractions of 5 ml were collected in test tubes. These fractions were concentrated by evaporation and subjected to thin-layer chromatography (TLC). After TLC, comparable fractions (14–21) were obtained as a single compound. The isolated compound was further subjected to column chromatography and TLC for verifying the purity of the compound. This isolated compound was named as compound TE (toluene/ethyl acetate fractions). Fractionated compound TE was characterized using TLC, ultraviolet–visible (UV–Vis) spectral analysis, Fourier transform infrared (FTIR) spectral, nuclear magnetic resonance spectroscopy, mass spectrometry, and elemental analyses.

#### 2.3. Anticandidal activity

C. albicans (NCIM 3471), C. tropicalis (NCIM 3118), and C. glabrata (NCIM 3236) were obtained from National Collection of Industrial Microorganisms (NCIM), National Chemical Laboratory, Maharashtra, India, and used for anticandidal analysis. A primarily anticandidal test was carried out for aqueous, ethyl acetate extracts, and fractioned compound by well-diffusion method as described by Magaldi et al. [17]. Yeast was inocula prepared from 18-h-old mother cultures. Yeast inocula were spread on petri dishes containing Sabouraud dextrose agar and wells were made using a sterile cork borer. Extracts and fractioned compound were dissolved in sterile 4% dimethyl sulfoxide (DMSO). Thereafter, 100, 200, 400, and 800 µg extracts and fractions were loaded on the

wells. Standard antifungal agent miconazole 50  $\mu g$  and 4% DMSO were used as positive and negative controls. The plates were incubated at 37 °C for 24–48 h and the zone of inhibition (ZOI) was measured.

#### 2.4. Minimal fungicidal concentration

Minimal fungicidal concentration of the fractioned compound was evaluated by the broth macro dilution method, to determine the minimum inhibitory concentration (MIC) of the fractioned compound that inhibited visible growth of test pathogens. Mid exponential culture ( $10\,\mu$ l) was seeded with the fractioned compound at concentrations of 2–200 µg in 1 ml total volume of Sabouraud dextrose broth incubated at 37 °C for 24 h with mild agitation at 100 rpm. After the incubation, the culture pellets were obtained by centrifugation (REMI, Maharashtra, India), resuspended in  $100\,\mu$ l sterile broth, and the total suspension swabbed onto the Sabouraud dextrose agar plates and allowed to incubate for a further 24–48 h at 37 °C [18].

#### 3. Results

#### 3.1. Characterization of isolated compound TE

#### 3.1.1. Physical properties of compound TE

About 179 mg dry weight of the residue was obtained from identified fractions. Fractionated compound was crystal in nature and green in color, soluble in all organic solvents. The  $R_{\rm f}$  value of this compound was 0.37 in toluene/ethyl acetate (4:1) in mobile phase.

#### 3.1.2. Ultraviolet-visible spectral analysis

The UV–vis spectra exhibited an absorption bond at 278 nm, which can be assigned to  $\pi$ – $\pi$ \* transition of carboxyl and aromatic groups. This gives an idea about the structured compound containing hetero atom having nonbonding electrons (Fig. 1).

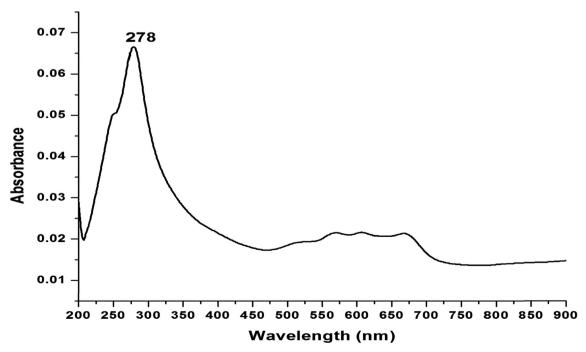


Fig. 1. UV-Visible spectra of fractioned compound TE (Diphyllin).

#### 3.1.3. FTIR spectral analysis of compound TE

Infrared spectrum of TE recorded in KBr medium (4000–450 cm $^{-1}$ ) showed a number of bands (Fig. 2). The tentative assignments of various stretching and bending frequencies for fractioned compound TE are listed in Table 1. A broad band observed at 3437 cm $^{-1}$  can be assigned to the OH stretching vibration. The medium bands at 3037 and 3031 cm $^{-1}$  are attributed to aromatic C=H stretching vibration. The ketonic bond observed due to stretching vibration, that is C = O, appears at 1764 cm $^{-1}$ . In in-plane bending, bands appear in the region 1378 cm $^{-1}$ . The median band observed at 1086 cm $^{-1}$  can be attributed to C=O=C bending vibration. The presence of absorption bands in the region 846–727 cm $^{-1}$  is due to out-of-plane bending vibrations of C=H bands at 727 cm $^{-1}$ . The aromatic substituted vibration appears as a strong absorption band at 626 cm $^{-1}$ .

#### 3.1.4. <sup>1</sup>H NMR spectral studies of compound TE

The proton magnetic resonance spectra of the fractioned compound TE was recorded (Table 2) in CDCl<sub>3</sub> solvent (Fig. 3) and the resonance signals were given in. The integration of the spectra indicates the number of proton to be 16. Resonance signal at  $\delta$  3.98 ppm is due to O—CH<sub>3</sub> protons. The aromatic proton appears as multiples in the range of  $\delta$  6.83–6.56 ppm and the substitute benzene ring appears in the range of  $\delta$  6.40–6.37 ppm. The OH proton appears at  $\delta$  7.76 ppm signal. The signal due to methoxy proton appears, that is O—CH<sub>2</sub>—O, at  $\delta$  5.995.95 ppm and the —CH<sub>2</sub>—O proton appears at  $\delta$  5.27 ppm. Thus, the <sup>1</sup>H NMR spectra reveal the presence of aromatic, methoxy, O—CH<sub>2</sub>—O, and —CH<sub>2</sub>—O groups in the compound. The intensity ratio obtained for signals correlates well with the total number of protons under chemically equivalent and magnetically active nuclei.

#### 3.1.5. <sup>13</sup>C NMR spectral studies of compound TE

The spectra of the fractioned compound TE were recorded in the CDCl<sub>3</sub> solvent, as shown in Fig. 4, and the data are presented in Table 3. A spectrum shows absorption of carboxyl carbon at 169.55 and 169.44 ppm. The chemical shift of aromatic carbons appear at 146.71, 129.8–118.35 ppm. The substitute's aromatic carbon can be distinguished from other carbons by its decreased peak height. Its lacks a proton and hence suffers from longer relaxation time with a diminished nuclear Overhauser effect. The peak at 149.52 ppm may be assigned to the substitute's carbon in the

**Table 1** FT-IR spectrum of fractioned compound TE.

Absorption (cm <sup>-1</sup> )	Assignment
3437	OH(b)
3037-3031	C—H aromatic
2931	C—H aliphatic
1764	C=0
1378	In plan bending bass of aromatic ring
1056	C-O-C
846-727	Out of plan bending of aromatic
727	Substitutes aromatic ring

**Table 2** <sup>1</sup>H NMR spectra of fractioned compound TE.

Resonance signals	Assignment
3.98	O-CH <sub>3</sub>
6.83-6.56	Aromatic proton
6.40-6.37	Substitutes benzene ring
7.76	OH
5.99-5.95	O-CH <sub>2</sub> -O
5.27	-CH <sub>2</sub> -O

ring. The peaks at 151.09 and 149.52 are due to aromatic carbon with O atom. The sharp signal at 149.71 is due to aromatic *ortho*-carbon bond with OH group. Peaks at 134.76 and 129.82 ppm are due to aromatic ring attached with another aromatic ring as a single O bond carbon. The peaks at 11.08–100.59 are due to aromatic carbons. The peak at 69.02 ppm is due to Ar—C—O carbon. The chemical shifts of <sup>13</sup>C atoms of the fractioned compound have been assigned relative to the assignments available for individuals of the compound. The <sup>13</sup>C NMR signals of the compound and various assignments to different carbon atoms are in good agreement with the <sup>1</sup>H NMR.

#### 3.1.6. Mass spectrum analysis of compound TE

The mass spectrum of the fractionated compound was obtained on element ionization mode. The molecular mass was observed at 378 m/z, which is close to the expected value of  $380 \, m/z$  (Fig. 5). The mass spectral fragment studies show that the molecular ions peak at m/z 378, that is  $M^{-2}$  peak ( $C_{21}H_{16}O_7$ ), which confirms the molecular mass of the compound. The peak at m/z 366, 345, 319,

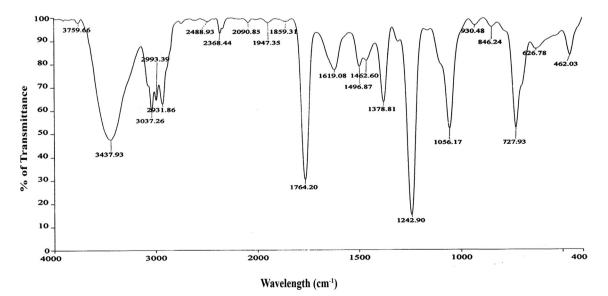


Fig. 2. FT-IR spectrum of fractioned compound TE (Diphyllin).

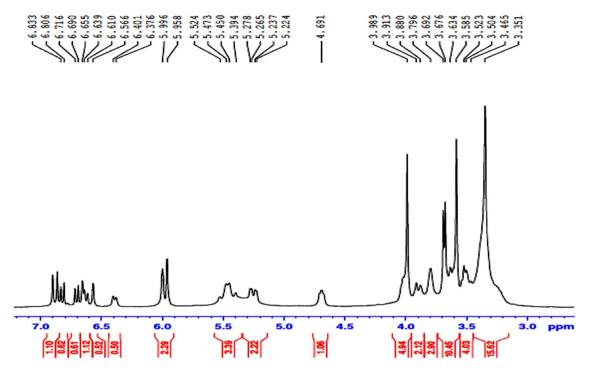


Fig. 3. <sup>1</sup>H NMR spectra of fractioned compound TE (Diphyllin).

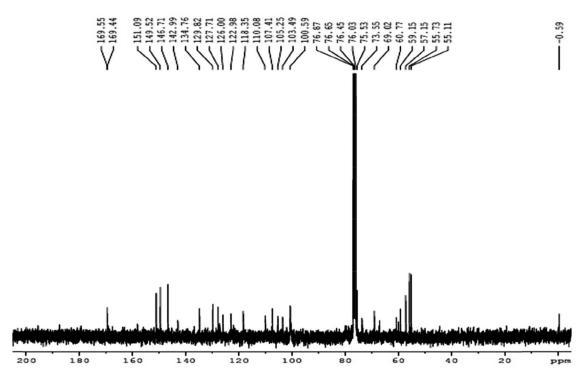


Fig. 4. <sup>13</sup>C NMR spectra of fractioned compound TE (Diphyllin).

and 285 are due to  $C_{21}H_{15}O_6^+$ ,  $C_{20}H_{14}O_5^+$ ,  $C_{19}H_{11}O_5^+$ , and  $C_{18}H_8O_4^+$ , respectively.

#### 3.1.7. Elemental analysis of compound TE

The compound TE was analyzed for carbon, hydrogen, and nitrogen. The results of elemental analyses are given below

	C%	Н%	0%
Calculated	66.31	4.24	29.45
Observed	66.29	4.24	29.27

**Table 3** <sup>13</sup>C NMR spectra of fractioned compound TE.

Resonance signals	Assignment
169.55 & 169.47	C=0
146.71 & 129.82, 129.82-118.35	Aromatic carbon
118.85	Substitutes aromatic carbon
149.52	Ar-O-CH <sub>2</sub>
55.73, 55.11	O-CH <sub>3</sub>
151.09 & 149.52	Aromatic carbon bonded with O-CH <sub>3</sub>
142.99	Ar—OH
134.76 & 129.82	Ar—Ar carbon
60.77 & 59.15	Ar—CH <sub>2</sub> —O
69.02	Ar—C—O

The above data indicate that the molecular formula of the fractioned compound TE is  $C_{21}H_{16}O_7$  and the molecular weight of the compound is 380.

#### 3.1.8. 2D structure elucidation and name of the compound TE

On the basis of the spectral studies, the fractioned compound TE was successfully drawn in the ChemDraw® Standard 14.0 software and the name was identified as 9-(1,3-benzodioxol-5-yl)-4-hydroxy-6,7dimethoxynaphtho[2,3-c]furan-1(3H)-one (diphyllin) (Fig. 6). The name and synonyms of the diphyllin and the source of the plant and properties of the compound are given in Table 4.

#### 3.2. Anticandidal activity

The anticandidal activity was determined against *C. albicans*, *C. tropicalis*, and *C. glabrata* from the aqueous and ethyl acetate solvent-based *C. collinus* leaf extracts. The ethyl acetate extract only showed activity against *C. albicans* at 800  $\mu g$  and other pathogens were resistant (Table 5). Further, the ethyl acetate extract was fractionated with toluene/ethyl acetate (3:2) solvents to get one single compound. Thereafter, the fraction, identified as a compound diphyllin, was tested against selected yeast pathogens (Fig. 7). All selected *Candida* species were found to be highly sensitive to the fractionated compound at 200  $\mu g$ . *C. glabrata* and *C. albicans* were found to be highly susceptible (11–13.5 ± 0.5 mm ZOI at 200  $\mu g$ ) to the isolated botanicals. The MFC values of the compound

**Fig. 6.** Structure of Diphyllin (9-(1,3-benzodioxol-5-yl)-4-hydroxy-6,7-dimethoxy-naphtho[2,3-*c*]furan-1(3*H*)-one).

were observed at  $85-145 \mu g/ml$  (Table 5) against all tested *Candida* species. For the standard antifungal drug miconazole used in the test, zones of inhibition in the range of  $17.5 \pm 0.5-20.5 \pm 0.5$  mm were observed against the tested pathogens.

#### 4. Discussion

In recent years, the number of researches focused on drug development from plant sources to treat infectious has notably increased. This plant material has been used in various biological studies [19,20]. Among all the properties, antifungal activity has received the most attention. *Candida* species are normal flora, harmless yeast-like fungi in healthy humans, but they can cause infections in skin and mucosal membranes under immunecompromised situations [21]. In this study, we evaluated the anticandidal activity of *C. collinus* leaf extracts and its fractions.

Hot aqueous extract of *C. collinus* did not show any activity against different species at least concentration but its displayed moderate anticandidal activity against only *C. albicans* at 800 µg. But earlier it was reported that cold aqueous extract if *C. collinus* exhibited good anticandidal activity (>11 mm as maximum ZOI)

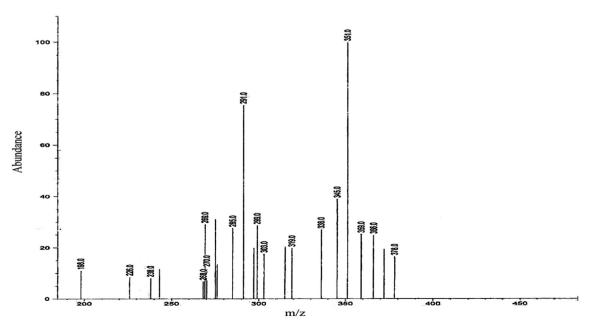


Fig. 5. The mass spectrum of fractioned compound TE (Diphyllin).

**Table 4** Isolated compound name and synonyms, compound source and bio activity.

S. no	Name and synonyms	Compound source	Bio activity
1	4-Hydroxy-6,7-dimethoxy-9-[3,4-(methylenedioxy) phenyl]-naphtho[2,3-c]furan-1(3H)-one	Lignan from roots of <i>Diphylleia grayi</i> , leaves of <i>Cleistanthus collinus</i> , <i>Justicia procumbens</i> and <i>Haplophyllum hispanicum Zerenex Molecular</i> [ZBioX-0173]	Cytotoxin; Zerenex Molecular [ZBioX-
2	9-(1,3-Benzodioxol-5-yl)-4-hydroxy-6,7- dimethoxynaphtho[2,3-c]furan-1(3H) on [German] [ACD/IUPAC Name]		0173]
3	9-(1,3-Benzodioxol-5-yl)-4-hydroxy-6,7-dimethoxynaphtho[2,3-c]furan-1(3H)-one [ACD/IUPAC Name]		
4	9-(1,3-Benzodioxol-5-yl)-4-hydroxy-6,7- diméthoxynaphto[2,3-c]furan-1(3H)-one [French] [ACD/IUPAC Name]		
5	Naphtho(2,3-c)furan-1(3H)-one, 4-hydroxy-6,7-dimethoxy-9-(3,4-(methylenedioxy)phenyl)-		
6	Naphtho(2,3-c)furan-1(3H)-one, 9-(1,3-benzodioxol-5-yl)-4-hydroxy-6,7-dimethoxy-		
7	Naphtho[2,3-c]furan-1(3H)-one, 9-(1,3-benzodioxol-5-yl)-4-hydroxy-6,7-dimethoxy- [ACD/Index Name] 22055-22-7 [RN]		
8	9-(13-Benzodioxol-5-yl)-4-hydroxy-6,7- dimethoxynaphtho(2,3-c)furan-1(3H)-one		
9	9-(Benzo[d][1,3]dioxol-5-yl)-4-hydroxy-6,7- dimethoxynaphtho[2,3-c]furan-1(3H)-one		
10	9-Benzo[1,3]dioxol-5-yl-4-hydroxy-6,7-dimethoxy- 3H-naphtho[2,3-c]furan-1-one		
11	Diphyllin		

Source of information: http://www.chemspider.com/Chemical-Structure.90798.html?rid=86c435ce-27e1-4d77-9f1d-1d1410b5091b.

**Table 5**Anticandidal activity of *C. collinus* extracts and Diphyllin.

Yeast pathogens		Candida albicans		Candida tropicalis		Candida glabrata	
Samples	Concentrations (μg)	ZI <sup>*</sup>	MFC <sup>**</sup> (μg/mL)	ZI <sup>*</sup>	MFC** (μg/mL)	ZI°	MFC** (μg/mL)
Aqueous	100	_	_	_	_	_	-
extracts	200	_		_		_	
	400	-		-		-	
	800	M		_		_	
Ethyl acetate extract	100	=	=	=	=	_	_
	200	_		_		_	
	400	M		_		_	
	800	$9.5 \pm 0.5$		M		_	
Diphyllin	100	M	≥85	M	≥110	M	≥145
	200	11 ± 0		$9.5 \pm 0.5$		13.5 ± 0.5	
	400	$13.25 \pm 0.25$		11.25 ± 0.25		$14.5 \pm 0.5$	
	800	15 ± 0.5		12.5 ± 0.5		$17.5 \pm 0.5$	
4% DMSO		-	=	-	_	_	-
Micoconazole (50 μg) (Positive control)		19.5 ± 0.5		17.5 ± 0.5		$22.5 \pm 0.5$	

<sup>\*</sup> Results are expressed as mean ± standard deviation of values from triplicate experiments.

and exhibited MICs  $600 \,\mu g/ml$  and MFC  $750 \,\mu g/ml$  against *C. albicans* [13]. Significant ZOI was observed in ethyl acetate extract at  $800 \,\mu g$  compared to the aqueous extract. From this study, it was found that some important phytocompounds might be present in ethyl acetate extract. Earlier, we investigated the preliminary phytochemicals, and aqueous and ethyl acetate extracts were qualitatively screened using gas chromatography–mass spectrometry (GC–MS). Fifteen major phytocompounds were found to be present in ethyl acetate extract, major compounds among them being silane, trimethyl[5-methyl-2-(1-methyl ethyl)phenoxy]-anthracene (7.06%). Tannins, terpenoids, flavonoids, saponins, glycosides, steroids, and alkaloids were also found in ethyl acetate

extract. Tannins, terpenoids, glycosides, flavonoids, and saponins were observed in the aqueous extract [22]. The major compounds were observed in the ethyl acetate extract other than aqueous extract by TLC and GC–MS analyses [23,24]. The isolated phytocompound diphyllin showed higher level of inhibition against all tested *Candida* species at 200 µg. Before that many researcher reported good anticandidal activity of medicinal plants. They reported only the anticandidal activity of crude extracts and did not find any promising fractionated compounds [25,26].

Diphyllin is a major glycoside compound present in *C. collinus* plant. Anjaneyulu et al. [27] reported a new diphyllin diglycoside from *C. collinus* heartwood. From the methanolic extract, the CHCl<sub>3</sub>

<sup>\*\*</sup> Average (MFC) minimal fungal inhibition concentration; M – Moderate activity; DMSO – Dimethyl sulphoxide

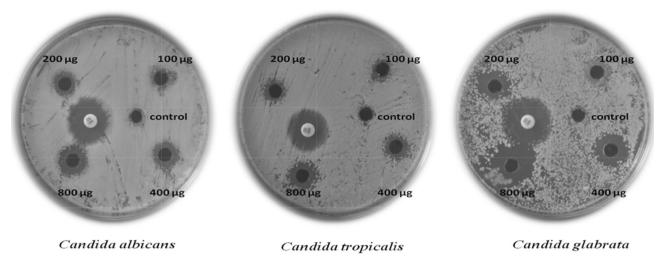


Fig. 7. Anticandidal activity of Diphyllin.

rig. 7. Anticandidal activity of Diphyllin.

Investigations of extracts from *C. collinus* plant leaves revealed a complex group of compounds [29,30]. The toxic active principles of *C. collinus* in the leaves are arylnaphthalene lignin lactones—diphyllin and its glycoside derivatives Cleistanthin A and B, and Collinusin [31,32]. Diphyllin and Cleistanthin A and B were commonly known as "Oduvin" in the past. Also, the lignans Cleistanone, Cleistanthin C, and Cleistanthin D are present in *C. collinus*. The toxicity of the *C. collinus* leaves have been primarily due to Cleistanthin A and B [33]. Diphyllin was isolated free and also as 3,4-di-*O*methylxylopyranoside from *C. collinus* leaves and as its fi-Dglucopyranoside from its bark [34]. The fruits of *C. collinus* have been shown to contain sitosterol and lupeol [35].

Aligiannis et al. [36] proposed a classification on MIC of plant material extracts (strong inhibitors, MIC up to 500 μg/ml); moderate inhibitors (MIC between 600 and 1500 μg/ml); and weak inhibitors (MIC above 1600 µg/ml). On the basis of our MIC results, fractionated compound of C. collinus extract showed strong inhibition (85–145 μg/ml) against C. albicans followed by C. tropicalis and C. glabrata. Candida species is responsible for the majority of yeast infections in humans and veterinary animals at immunecompromised situations. Among them, in 90% of cases, C. albicans is the most causative agent associated with disease to serious fungal infection. Moreover, C. albicans form biofilm with C. tropicalis, C. glabrata, and other Candida species, which has also been associated with disease [37,38]. Previously, the toxicity property of fractioned compound was studied against mouse 3 T3-L1 preadipocytes cell proliferation. The ethyl acetate fraction (diphyllin) showed 23-59% anti-proliferative activity (concentration necessary to inhibit cell growth at 50% is  $\sim$ 180 µg/ml) [39]. From our study, we found an assured isolated compound with anticandidal activity against Candida species with less toxicity.

In conclusion, it can be said that the results of this study indicated that diphyllin, the fractionated compound of *C. collinus* ethyl acetate extract, exhibited strong inhibition against *C. albicans*, *C. tropicalis*, and *C. glabrata*. Also, to the best of our knowledge, this

is the first detailed study of *C. collinus* extract and its fractioned compound against *Candida* species. Further studies on the mode of action of diphyllin are required to understand its anticandidal effects.

#### **Conflict of interest**

The authors declared that no conflict of interest.

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#### References

- [1] Horn DL, Neofytos D, Anaissie EJ, Fishman JA, Steinbach WJ, Olyaei AJ. Epidemiology and outcomes of candidemia in 2019 patients: data from the Prospective Antifungal Therapy Alliance Registry. Clin Infect Dis 2009;48:1695–703.
- [2] Douglas LJ. Candida biofilms and their role in infection. Trends Microbiol 2003;11:30–6.
- [3] Edwards JE. Candida species. In: Mandell, Douglas, Bennett, editors. Principles and practice of infectious diseases. New York, USA: Churchill Livingstone; 1995. p. 2289–301.
- [4] Khan ZU, Chandy R, Metwali KE. Candida albicans strain carriage in patients and nursing staff of an intensive care unit: a study of morphotypes and resistotypes. Mycoses 2003;46:476–86.
- [5] Klepser ME. Antifungal resistance among Candida species. Pharmacotherapy 2001;21:124S-32S.
- [6] Runyoro DKB, Matee MIN, Ngassapa OD, Joseph CC, Mbwambo ZH. Screening of Tanzanian medicinal plants for anti-candida activity. BMC Complement Altern Med 2006;30:6–11.
- [7] Wagner H. Synergy research: approaching a new generation of phytopharmaceuticals. Fitoterapia 2011;82:34–7.
- [8] Hassan STS, Masarčíková R, Berchová K. Bioactive natural products with antiherpes simplex virus properties. J Pharm Pharmacol 2015;67:1325–36.
- [9] Ali NA, Julich WD, Kusnick C, Lindequist U. Screening of Yemeni medicinal plants for antibacterial and cytotoxic activities. J Ethnopharmacol 2001;74:173–9.
- [10] Sanjenbam P et al. Anticandidal activity of silver nanoparticles synthesized using Streptomyces sp. VITPK1. J Mycol Med 2014;24(3):211–9.
- [11] Duarte MCT, Figueira GM, Rehder VLG, Delarmelina C. Anti-Candida activity of Brazilian medicinal plants. J Ethnopharmacol 2005;97:305–11.
- [12] Feldmesser M. New and emerging antifungal agents: impact on respiratory infections. Am J Respir Med 2003;2:371–83.
- [13] Maji S, Dandapat P, Ojha D, Maity C, Halder SK, Das Mohapatra PK, et al. In vitro antimicrobial potentialities of different solvent extracts of ethnomedicinal plants against clinically isolated human pathogens. J Phytol 2010;2(4):57–64.
- [14] Govindachari TR, Sathe SS, Viswanathan N, Pai BR, Srinivasan M. Chemical constituents of *Cleistanthus collinus* (Roxb.). Tetrahedron 1969;25:2815–21.

- [15] Arivoli S, Samuel T. Larvicidal efficacy of Cleistanthus collinus (Roxb.) (Euphorbiaceae) leaf extracts against vector mosquitoes (Diptera: Culicidae). Asian Pac J Trop Biomed 2011;1(2):281–3.
- [16] Maity S. Ethnobotany of lateritic West Bengal [Ph.D Thesis]. Midnapore: Vidyasagar University; 2002.
- [17] Magaldi S, Mata-Essayag S, Hartung de Capriles C, Perez C, Colella MT, Carolina O, et al. Well diffusion for antifungal susceptibility testing. In J Infect Dis 2004;8:39–45.
- [18] CLSI. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts, 3rd edn. Approved standard M27-A3. Wayne, PA: Clinical and Laboratory Standards Institute; 2008.
- [19] Pinho PM, Kijjoa A. Chemical constituents of the plants of the genus *Cleistanthus* and their biological activity. Phytochem Rev 2007;6:175–82.
- [20] Pradheep Kumar CP, Panneerselvam N, Rajesh S, Shanmugam G. Cytotoxic and genotoxic effects of cleistanthin B in normal and tumour cells. Mutagenesis 1996;11:543–7.
- [21] Staib P, Kretschmar M, Nichterlein T, Hof H, Morschhauser J. Differential activation of a *Candida albicans* virulence gene family during infection. Proc Natl Acad Sci USA 2000;97:6102–7.
- [22] Suman T, Elangomathavan R. Bio-prospecting of *Cleistanthus collinus* and its antibacterial activity. Asian J Pharm Clin Res 2013;6(4):206–9.
- [23] Parasuraman S, Raveendran R, Madhavravo C. GC–MS analysis of leaf extracts of Cleistanthus collinus Roxb. (Euphorbiaceae). Int J Ph Sci 2009;1(2):284–486.
- [24] Suman T, Chakkaravarthi K, Elangomathavan R. Phyto-chemical profiling of Cleistanthus collinus leaf extracts using GC-MS analysis. Res J Pharm Tech 2013;6(11):1173-7.
- [25] Naeini A, Jalayer Naderi N, Shokri H. Analysis and in vitro anti-Candida antifungal activity of *Cuminum cyminum* and *Salvadora persica* herbs extracts against pathogenic Candida strains. J Myco Méd 2014;24:13–8.
- [26] Naeini A, Khosravi AR, Chitsaz M, Shokri H, Kamlnejad M. Anti-Candida albicans activity of some Iranian plants used in traditional medicine. J Myco Méd 2009:19:168–72.

- [27] Anjaneyulu ASR, Atchljta R, Ramachandra R. A new diphyllin glycoside from Cleistanthus collinus. Phytochemistry 1975;14. 187-76.
- [28] Devi CJ, Vasantha KPK. Chemical examination of fruits of *Cleistanthus collinus*. Der Pharm Chem 2011;3(6):160–4.
- [29] Rajagopal Naidu S, Venkat Rao P, Subrahmanyam CA. The microscopy and chemistry of oduvin. J Proc Inst Chem India 1944;16:59–63.
- [30] Maiti PC, Das AK. Chemical examination of the fruits of *Cleistanthus collinus*. Curr Sci 1965:34:79–81.
- [31] Anjaneyulu AS, Ramaiah PA, Row LR, Venkateswarlu R, Pelter A, Ward RS. New lignans from the heartwood of *Cleistanthus collinus*. Tetrahedron 1981:37:3641–52.
- [32] Anjaneyulu AS, Ramaiah PA, Rao R. Crystalline constituents of Euphorbiaceae; Part XVI A new Diphyllin glycoside from Cleistanthus collinus. Indian J Chem 1977:15:10-1.
- [33] Ramesh C, Ravindranath N, Ram TS, Das B. Arylnaphthalide lignans from *Cleistanthus collinus*. Chem Pharm Bull (Tokyo) 2003;51:1299–300.
- [34] Govindachari TR, Sathe SS, Viswanathan N, Pai BR, Srinivasan M. Chemical constituents of *Cleistanthus collinus*. Tetrahedron 1969;25:2815–21.
- [35] Lakshmi TG, Srimannarayan G, Subbarao NV. A new glucoside from Cleistanthus collinus. Curr Sci 1970;39:395–6.
- [36] Aligiannis N, Kalpotzakis E, Mitaku S, Chinou IB. Composition and antimicrobial activity of the essential oils of two *Origanum* species. J Agric Food Chem 2001;40:4168–70.
- [37] Staib P, Kretschmar M, Nichterlein T, Hof H, Morschhauser J. Differential activation of a Candida albicans virulence gene family during infection. Proc Natl Acad Sci USA 2000;97:6102–7.
- [38] Heinig MJ, Francis J, Pappagiansis D. Mammary candidosis in lactating women. | Hum Lact 1999;15:281–8.
- [39] Suman T, Elangomathavan R, Ilavenil S, Ramesh S. In vitro cytotoxic effect of Cleistanthus collinus extracts and fractions on mouse cell line. J App Pharm Sci 2014;4(10):044–6.