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BioScape: A Modeling and Simulation Language for Bacteria-Materials Interactions

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**Abstract**

We design BioScape, a concurrent language for the stochastic simulation of biological and bio-materials processes in a reactive environment in 3D space. BioScape is based on the Stochastic Pi-Calculus, and it is motivated by the need for individual-based, continuous motion, and continuous space simulation in modeling complex bacteria-materials interactions. Our driving example is a bio-triggered drug delivery system for infection-resistant medical implants. Our models in BioScape will help in identifying biological targets and materials strategies to treat biomaterials associated bacterial infections.

The novel aspects of BioScape include syntactic primitives to declare the scope in space where species can move, diffusion rate, shape, and reaction distance, and an operational semantics that deals with the specifics of 3D locations, verifying reaction distance, and featuring random movement. We define a translation from BioScape to 3*π* and prove its soundness with respect to the operational semantics.

*Keywords:* Stochastic simulation, stochastic Pi-calculus, bacteria-materials interaction

# Introduction

In contrast to the now deep and multidimensional understanding of how tissue cells interact with the surface of biomaterials, comparatively little is known about the influence of surface properties on interactions with bacteria. These interactions are clearly very important, however. Biomaterials-associated infection (BAI) is a major clinical problem [[1](#_bookmark19),[13](#_bookmark31),[35](#_bookmark53)]. Current strategies to mitigate BAI concentrate on engineering antimicrobial [[16](#_bookmark34),[18](#_bookmark36),[20](#_bookmark38),[22](#_bookmark40)] or antifouling [[9](#_bookmark27),[14](#_bookmark32),[19](#_bookmark37),[33](#_bookmark50)] coatings for specific biomedical devices.

While valuable clinically in the short-term, this approach ignores the fact that many devices, particularly those involving regenerative strategies, require surfaces that must controllably interact with both tissue cells and bacteria. Often, surfaces

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optimized to promote tissue-cell interactions also support undesirable bacterial col- onization (Fig. [1](#_bookmark4)).

## Concurrent Modeling of Bio- material Interactions

As an alternative to models built around sets of ordinary differential equations (ODEs), process algebras are formal languages where multiple objects with different behavioral at- tributes can interact with each other and dynamically influence overall sys- tem development. Process algebras are being used to model biological systems [[30](#_bookmark44),[6](#_bookmark22),[2](#_bookmark20)], where they are particularly at-

tractive, because of their ability to ac- commodate new objects and new be- havioral attributes as the complex bi- ological system becomes better under- stood.

Currently, however, modeling lan- guages based on concurrent synchro- nization either lack spatial attributes (SPiM [[27](#_bookmark45)], Kappa [[12](#_bookmark29)], Petri Nets [[26](#_bookmark46)]) or stochasticity (SpacePi [[15](#_bookmark33)], Shape Calculus [[3](#_bookmark21)]), or they offer only a lim- ited notion of space (BioAmbients [[23](#_bookmark41)], BioPepa [[10](#_bookmark28)]). As findings from biolog- ical experiments reveal, inter and intra- cellular dynamics and signaling path- ways depend on the location and move- ment of particles [[17](#_bookmark35)]. Recently, new spatial modeling languages allowing ex- plicit description of temporal spatial dynamics of biochemical processes have been proposed (SpacePi [[15](#_bookmark33)], DCA [[36](#_bookmark54)], *L*Π [[34](#_bookmark52)], Stochsim [[24](#_bookmark42)]). Other agent- based platforms [[21](#_bookmark39)] include C-Immsim

Fig. 1Grit-blasted Ti6Al4V used in orthopedic im-

plants promotes osteoblast adhesion/spreading (fluo- rescence image) but also enhances staphylococcal col- onization (SEM inset). After Wu, Libera et al. [[37](#_bookmark55)]

Released AmA

* bind
* diffuse
* flow
* kill
* hydrolyze



Planktonic bacterium

* adsorb (A p)
* diffuse
* flow
* killed by AmA (Kp >> Ka)
* reproduce (Rp ≠ Ra)
* metabolize (Mp ≠ Ma)

Medium Flow

Bound AmA

* stay bound
* pH -release
* contact-release

Adsorbed bacterium

* desorb
* killed by AmA (Ka << Kp)
* reproduce (Ra ≠ Rp)
* metabolize (Ma ≠ Mp)
* produce ECM

Fig. 2 Concurrency models flexibly account for mul- tiple object types and copies. Each type has char- acteristic attributes. Expression of one attribute dy- namically influences other objects and what attribute each might next express.

[[32](#_bookmark51),[8](#_bookmark23)] and PathSim visualizer [[28](#_bookmark47)]. However, few of them support individual based,

continuous motion, and continuous space stochastic simulation [[4](#_bookmark24)], which are im- portant features for modeling temporal spatial dynamics of biochemical processes accurately. To address this problem we design BioScape, a new language incorpo- rating both stochasticity and 3D spatial attributes.

Fig. [2](#_bookmark5) illustrates the elements of a simple instance of the model. It includes four objects: planktonic bacteria; adsorbed bacteria; gel-bound antibacterial agent

(AmA); and released AmA. Each has attributes, many of which are inter-related. Both bacteria, for example, can metabolize, albeit at different rates. Metabolism is particularly significant for bound AmA, since acidic products will lower the local pH, reduce the strength of electrostatic AmA-gel binding, and enable the pH-release of bound AmAs. Released AmA can then diffuse and/or flow, and potentially kill bound or planktonic bacteria, again with different efficiencies, because of the en- hanced microbial resistance of biofilm bacteria. The complexity of such a model can rapidly explode as more attributes are assigned to each object. Note that spatial coordinates and stochasticity are critical to understanding the interactions between different objects and with the substrate, because they all rely on proximity. So far, existing process-algebra modeling languages do not support this. Furthermore, while there are now a number of models being developed to explain the biofilm for- mation [[31](#_bookmark49),[38](#_bookmark56)], none takes into account the controllable properties of the substrate and how this can influence bacterial adhesion, proliferation, and phenotypic change.

**1.4**

**1.2**

**1.0**

Optical density

**0.8**

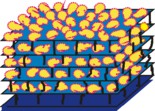
**0.6**

**0.4**

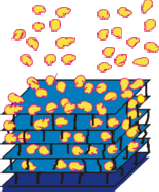
**0.2**

**0.0**

**0 5 10 15 20 25**



bacteria



bacteria

**Time, hours**

**8.0**

**7.5**

**7.0**

pH in the medium

**6.5**

**6.0**

**5.5**

**5.0**

**4.5**

**4.0**

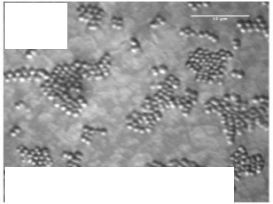
pH change

Fig. 3 pH variation during S. epidermidis growth

Fig. 4 Bio-triggered release of antibacterial agents from pH-responsive layer-by-layer hydrogel thin film.

A number of experimentally measurable quantities can be used to calibrate ele- ments of the model. Among these are the bacterial proliferation rates, the biofilm morphology, the AmA minimum inhibitory (MIC) and bactericidal (MBC) concen- trations, gel capacities for AmA binding and pH-dependent properties, and com- ponent diffusivities, among others. For example, measurements by Sukhishvili’s group of the pH-dependent release of L5 antimicrobial peptide and its effect on S. epidermidis [[25](#_bookmark43)] have been used by Compagnoni and her group to develop a pro- totype implementation in BioScape (Fig. [3](#_bookmark6)). This data corresponds to the schema described in Fig. [4](#_bookmark7). This model includes computational processes for planktonic bacterial motion, adhesion, and proliferation on an L5-loaded hydrogel surface as well as for the local metabolic pH decrease, triggered AmA release, and killing of bacteria (Figs. [5](#_bookmark8) C/D). Experimental images (Figs. [5](#_bookmark8) A/B) are in good agreement with our computational results.

## Language Design



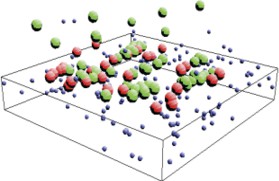
A

10 m

**as-synthesized**

The 3D aspect of BioScape is inspired by 3*π* [[7](#_bookmark25)] and SpacePi [[15](#_bookmark33)]. In BioScape every process has an implicit affine map and a shape (Fig. [7](#_bookmark10)). The shape is defined by a set of point coordinates that is local to a process’ system of coordinates. While the syntax of BioScape describes processes within its implicit local frame, its seman- tics places processes in a global 3D space (Fig. [8](#_bookmark11)), by assign- ing an affine map to each pro- cess – a located process. The application of an affine map to the shape returns the shape lo- cated in the global frame. The

Fig. 5 Optical micrographs of NJ 9709 S. epidermidis cul- tured on (A) as-synthesized and (B) L5-loaded (PMAA)10 gels. BioScape simulations with 40 bacteria and (C) an as-synthesized gel film and (D) an L5-loaded gel film. Live bacteria are green, dead bacteria are red, and L5 molecules are small blue dots.

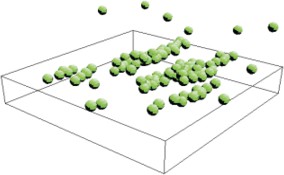


B

10 m

**peptide-loaded**

D



C

affine map is what characterizes the position of the process. Movement is then mod-

eled by updating a process’ affine map. On the contrary, 3*π* is a low level language that gives absolute control of spatial attributes to the programmer. For example, the programmer can guard an interaction by checking whether two processes are close enough, and it can also assign affine maps to processes. For example, collision checks in 3*π* would have to be implemented by the programmer checking corre- sponding distances with every other entity in the system, and taking into account the shape of every process. In contrast, in BioScape, the programmer specifies species declaring a reaction radius, as in SpacePi, and the operational semantics enforces the proximity requirement. Therefore, affine maps are not available to the programmer in BioScape, unlike 3*π*. While 3*π* is a general calculus for processes in space, BioScape has been designed to program biological and biomaterial processes and their interactions.

## New Features of BioScape

* We introduce three new parameters for each process: movement space, movement step, and shape to be specified by the programmer when defining a process. We take the definition of bacteria as an example. The movement space restricts the space within which bacteria can move. In Fig. [5](#_bookmark8), bacteria can never penetrate the gel film. Thus we define the volume above the film as the movement space for bacteria. We assume that the scheduler will randomly place the initial con- centration of bacteria in that space. The movement step indicates the distance that bacteria can move in a time interval. The shape of bacteria represents the volume that each bacterium occupies in space. The shape is instrumental in avoiding collisions and overlaps.
* Besides standard reaction operations send, receive and delay, BioScape has a geometric operation move, that can be generalized to any affine transformation. The difference is that while reaction operations have stochastic reaction rates, the operation move is always enabled. In order to avoid an unfair competition between reactions and geometric operations, we split the semantics into two reduction relations, and we consider a heterogeneous choice operator with probabilistic and non-deterministic branches. Intuitively, it is like having two choice operators.
* The design decisions of having the scheduler assign initial affine maps to each process and of having the operational semantics enforcing proximity requirements significantly simplifies the models’ code.

BioScape is a collaboration tool that has helped us develop a long term multidis- ciplinary research program to study how surface properties affect interactions with bacteria.

# Syntax

BioScape is based on the stochastic *π*-calculus [[29](#_bookmark48)] with primitives for processes in 3D space. We assume a set of names N ranged over by *x, y,.. .*, and a global three-di-

mensional Euclidean space. The syntax of BioScape

is defined in Fig. [6](#_bookmark9). (*νx*@r*,*rad)*.P* defines

channel name *x* with two parameters r and rad*∈* R*≥*0 in process *P* ; r is the stochastic rate for communications through channel *x* and rad is the communication radius. The radius is the maximum distance between pro- cesses in order to communicate through chan-

*P, Q* ::= (*νx*@r*,*rad)*.P | P | Q | X*(*x*)

*M* ::= **0** *| π.P* + *M*

*π* ::= delay@r *|* !*x*(*x*) *|* ?*x*(*y*) *|* mov

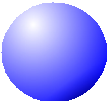
*D* ::= *∅ | D, X*(*x*)@*ξ, ω, σ* = *M*

*E* ::= *∅ | E, x*@r*,* rad

Fig. 6 BioScape Syntax

nel *x*, and the reaction rate determines whether two processes that are close enough to react actually do. *P | Q* is parallel composition of processes. *M* is the *hetero- geneous* choice, where + is associative and commutative. *M* may have reaction branches and movement branches. The reaction branches are probabilistic, while the movement branches are non-deterministic, since reactions are subject to reac- tion rates, while movement is always enabled. **0** is the empty process. The prefix *π* denotes the action that the process *π.P* can perform. delay@r is a spontaneous and unilateral reaction of a single process, and r is the stochastic rate. !*x* is the output prefix and ?*x* is the input prefix. We add a new geometry prefix mov to translate a process. We use standard syntactic abbreviations such as *π.P* for *π.P* + 0 and *π* for *π.*0. *X*(*x*) is a definition call.

*D* is a global list of definitions. *X*(*x*)@*ξ, ω, σ* = *M* defines process *X* with argu- ment *x*, movement space *ξ*, step *ω* and shape *σ*. The movement space *ξ* is a set of point coordinates in the global coordinate system defining a volume. Intuitively, *X* can move within *ξ*. The step *ω ∈* R*≥*0, is the distance *X* can move in a single step, and it corresponds to the diffusion rate of *X*; the shape of *X* is *σ*, an arbitrary vol- ume in space defined as a set of coordinates in the local coordinate system (Fig. [7](#_bookmark10)). The movement space for the empty process **0** is everywhere, the global space, and



i

o j

k k



i

j

o

Fig. 7. Shapes (*σ*) in their local coordinate system (i,j,k) with origin o.



i

distance o j k

i

o j

o j

k

k

o

i

o j

k

k

z

o j

i

i

x

y

Fig. 8. Shapes in the global coordinate system (x,y,z) – (*μ*(*σ*)).

NR.Par

NR.Move

*μ′* = translate(*ω, μ, r*) *μ′*(*σ*) *⊆ ξ X*(*x*)@*ξ, ω, σ* = mov*.P* + *M ∈ D*

*E ▶ {X*(*y*)*}μ → {P* [*y/x*]*}μ'*

SR.Delay

*E ▶ A → B* trans(*B*) *∩* trans(*C*)= *∅*

*E ▶ A | C → B | C*

SR.Com

*X*(*x*)@*ξ, ω, σ* = [*delay*@r*.P*](mailto:delay@r.P)+ *M ∈ D*

*E ▶ {X*(*y*)*}μ →*r *{P* [*y/x*]*}μ*

*X*(*x*)@*ξ, ω, σ* =!*z*(*u*)*.P* + *M ∈ D Y* (*y*)@*ξ, ω, σ* =?*z*(*v*)*.Q* + *N ∈ D* dis(*μ, μ′*) *≤* rad

*E, z*@r*,* rad *▶ {X*(*x′*)*}μ | {Y* (*y′*)*}μ' →*r *{P* [*x′/x*]*}μ | {Q*[*y′/y*][*u/v*]*}μ'*

Fig. 9. Stochastic (SR) and Non-Stochastic (NR) Reduction Relations (Sample rules)

its movement step is 0 by default. *X* can be defined by at most one equation in *D*. *E* is an environment of channel name declarations. *x*@r*,* rad declares channel name *x* with reaction rate r and reaction radius rad. A channel name *x* appears at

most once in *E*.

Consider the following simple example of a bacterium Bac, that can either move or divide into two daughter cells. A more complex example can be found in Section

[5](#_bookmark14). Bac is defined with movement space movB, movement step stepB, and shape shapeB. Intuitively, bacteria can move within movB, with random steps of length stepB, and the shape shapeB is at all times contained within movB. The prefix mov represents a random movement of length stepB. delay@1.0.(Bac() | Bac()) represents mitosis, the division of a bacterium into two daughter cells: Bac() | Bac(), and the [delay@1.0](mailto:delay@1.0) prefix is used to model the fact that division is not an instantaneous reaction.

Bac()@movB, stepB, shapeB = mov.Bac() + delay@1.0.(Bac() | Bac())

# Operational Semantics

The operational semantics of BioScape is based on two reduction relations: a non- deterministic relation, *E ▶D A → B*, for geometric transformations, in our case move, and a stochastic relation, *E ▶D A →*r *B*, for reactions such as synchronization and delay. We often omit *D* to simplify the notation.

We use *μ* to represent an affine map; *μ*(*s*)= *M × s* + *V* , where *M* is a matrix

and *V* is a vector [[7](#_bookmark25)] (see Fig. [7](#_bookmark10)). *μ*(*σ*) computes the location and orientation of a process in the global coordinate system. When a process is initialized, it is assigned a random *μ* within its movement space *ξ* in the global frame (*μ*(*σ*) *⊆ ξ*).

We define *conﬁgurations A, B, . . .* as parallel compositions of *located processes*. A located process *{P}μ,σ* is a process *P* annotated with affine map *μ* and shape *σ*. *A, B, ...* ::= *{P}μ,σ | A | B |* (*νx*@r*,*rad)*.A*. For clarity, we often omit the shape *σ*.

Evaluation contexts are given as follows. We write **C**[*A*] for the context **C** in which the hole *·* has been substituted with *A*: **C** ::= *· |* (*νx*@r*,*rad)*.***C** *| A |* **C** *|* **C** *| A*

We define trans, a function on configurations that returns the shapes of its processes located in the global frame (Fig. [8](#_bookmark11)), such that trans(*{P}μ,σ*) = *μ*(*σ*), trans((*νx*@r*,*rad)*.A*)= trans(*A*) and trans(*A | B*)= trans(*A*) *∪* trans(*B*). We write dis(*μ, μj*) for the distance between the origin of *μ* and the origin of *μj* in the global frame (Fig. [8](#_bookmark11)). We denote translate(*ω*,*μ*,*r*) the function that generates a new local affine map *μj*, using the movement step *ω*, the old map *μ*, and a random number *r*.

As usual, fn is a function that returns the set of free names of a process or a configu- ration, bn is a function

(*S.Loc*) *P ≡ Q* implies *{P}μ ≡ {Q}μ*

(*S.Loc.N u*) (*νx*@r*,*rad)*.{P}μ ≡ {*(*νx*@r*,*rad)*.P}μ*

(*S.Loc.P ar*) *μ*1(*shape*(*P* )) d *μ*2(*shape*(*Q*)) = *μ*(*shape*(*P | Q*))

*{P | Q}μ ≡ {P}μ*1 *| {Q}μ*2

Fig. 10 Structural Equivalence (Sample Rules)

that returns the set of bound names of a process or a configuration, and =*α* equates two processes or configurations that differ only in their bound names.

We define the shape of processes inductively as follows:

*shape*(**0**)= *∅ shape*(*X*(*a*)) = *σ* if *X*(*x*)@*ξ, ω, σ* = *M ∈ D shape*((*νx*@r*,*rad)*.P* )= *shape*(*P* ) *shape*(*P | Q*)= *shape*(*P* ) d *shape*(*Q*)

where d gives a shape obtained by composing two shapes through juxtaposition.

For different applications we can choose suitable functions to realise d, we only require d to be a commutative and associative operator, i.e. *σ*1 d *σ*2 = *σ*2 d *σ*1 and (*σ*1 d *σ*2) d *σ*3 = *σ*1 d (*σ*2 d *σ*3). *μ*(*shape*(*P* )) computes the space occupied by a process *P* in the global coordinate system.

The structural equivalence, *≡*, is the smallest equivalence relation that contains the rules in Fig. [10](#_bookmark13), and such that parallel composition is commutative, associative,

and has neutral element *{***0***}μ* for any *μ*. Rule

S.Loc

uses the standard struc-

tural equivalence on processes, Rule S.Loc.Nu allows a channel definition to move through located processes, and Rule S.Loc.Par permits to move from configura- tions to a single located process (and viceversa) through the creation of a new affine map derived from the old ones and the shape of the processes.

Sample rules for both the non-stochastic (*E ▶ A → B*) and stochastic (*E ▶ A →*r *B*) reduction relation of BioScape are given in Fig. [9](#_bookmark12). The condition *μj*(*σ*) *⊆ ξ* of NR.Move ensures the new located process *{P* [*y/x*]*}μ′* is within its movement space *ξ*. NR.Move can be easily generalized to any affine map application. The NR.Par condition trans(*B*) *∩* trans(*C*)= *∅* means that reduction does not cause collisions

or overlaps. Regarding the stochastic reduction relation, r in *E ▶ A →*r *B* is the rate for synchronization or delay. The condition dis(*μ, μj*) *≤* rad in SR.Com ensures that located processes *{X*(*xj*)*}μ* and *{Y* (*yj*)*}μ′* are close enough to communicate through channel *x*. The remaining rules are standard.

# Simulation

The simulation algorithm has two phases: reaction and movement. The reaction phase is based on Gillespie’s algorithm as implemented in SPiM, and instead of keeping only concentrations for each agent species, it also keeps 3D information – (x,y,z) – for each instance. Gillespie’s algorithm produces two outputs in each iter- ation: a reaction to be executed next, and a time interval to update the simulation time. If the selected reaction is an interaction between two agents (send/receive) then the algorithm uses 3D location information to identify two individual agents close enough to interact, and proceeds as described in the operational semantics (SR-Com). If there are no two such agents it proceeds to the movement phase. If the selected reaction is a first order reaction (delay), the algorithm propagates 3D information as described in the operational semantics (SR-Delay). The move- ment phase uses the time interval generated by Gillespie’s algorithm and moves each agent in a random direction a distance proportional to the diffusion rate (*ω*) in that period of time. The movement phase takes into account collision detection as described in the operational semantics (NR-Par).

Overcrowding is beyond the scope of this paper. However there could be several approaches to address it. Cells have external regulators that allow them to react to molecules on the outside of neighboring cells, and those external regulators emit a signal to inhibit mitosis in overcrowded conditions. Therefore, if it were relevant to the system being modeled, the most natural way to address overcrowding would be to program it into the model. Alternatively, the modeling language could address overcrowding in different ways. For instance, if enough movement steps fail within a given movement area in one iteration, reactions exacerbating overcrowding could be disabled. This choice would have the effect of modeling dormancy in cells such as bacteria in Maturation I or Maturation II stage of biofilm formation.

# Example

In this section, we present the BioScape model for the bio-triggered drug delivery system from Figs. [4](#_bookmark7) and [5](#_bookmark8). We first define the communication channels release, kill and bind with reaction rate and reaction radius.

Channel release is for the communication between hydronium ions and em- bedded drug molecules, channel kill is for the communication between released drug molecules and bacteria, and channel bind is for the communication between bacteria in solution and binding sites on the hydrogel film substrate. BacF() rep- resents a free bacterium in solution. Free bacteria can move, bind to the hydrogel film, grow, acidify the environment by producing hydronium ions, and get killed by

drug molecules. BacB() represents a bacterium bound to the hydrogel film. Bound bacteria cannot move, but they can grow, acidify the environment by producing hydronium ions, and get killed by drug molecules. BindSite() represents a binding site on the hydrogel film. DeadBac() represents a dead bacterium. Dead bacteria can move and degrade. MolB() represents a drug molecule embedded in the hydrogel film. Drug molecules can be released as hydrogel ions are produced. MolF() repre- sents released drug molecules. They can move and kill bacteria. HIon() represents a hydronium ion, which determines the pH value of the environment. Hydrogen ions can move and release embedded drug molecules. resX, stepX and shapeX represent *ξ*, *ω* and *σ* for species X. The following is the BioScape code.

[release@0.004,](mailto:release@0.004) 2.0

[kill@0.001,](mailto:kill@0.001) 0.5 HIon()@resIon, stepIon, shapeIon [bind@3.0,](mailto:bind@3.0) 0.2 = mov.HIon() + !release.HIon()

BacF()@resBF, stepBF, shapeBF = BindSite()@resBS, stepBS, shapeBS mov.BacF() + = ?bind.BindSite()

!bind.BacB() +

delay@0.2.(BacF() | BacF()) + DeadBac()@stepDB, stepDB, shapeDB delay@0.005.(BacF() | HIon()) + = [delay@0.1](mailto:delay@0.1)

?kill.DeadBac()

MolB()@resMB, stepMB, shapeMB

BacB()@resBB, stepBB, shapeBB = = ?release.MolF() delay@0.1.(BacB() | BacF()) +

delay@0.005.(BacB() | HIon()) + MolF()@resMF, stepMF, shapeMF

?kill.DeadBac() = mov.MolF() + !kill

To simulate this model, the programmer decides on an initial concentration of free bacteria (BacF()) and bound molecules of AmA (MolB()). The simulation results of Fig. [5](#_bookmark8) C, show the controlled experiment of assuming only an initial concentration of free bacteria and no antibacterial agent. The results in Fig. [5](#_bookmark8) D, in contrast, show the effect of AmA molecules on the size and number of bacteria clusters. Figs. [5](#_bookmark8) A and B show the experimental images consistent with our results. We now consider an example illustrating the interleaving of stochastic and non- stochastic rules starting from one free bacterium, one hydronium ion and one bound molecule of AmA: *{*BacF()*}μ*1 *| {*HIon()*}μ*2 *| {*MolB()*}μ*3 *,* to showcase the inter- leaving of movement steps with reaction steps (communication or delay). To make

our example easier to follow, we unfold all three definitions as follows:

*{*mov.BacF() + !bind.BacB() + delay@1.5.(BacF() | BacF())

+delay@0.5.(BacF() | HIon()) + ?kill.DeadBac()*}μ*1

*| {*mov.HIon() + !release.HIon()*}μ*2

*| {*?release.MolF()*}μ*3

The process has movement steps and reactions available. We first reduce all move

steps. We start with mov.BacF(). Assume an affine map *μj* such that: 1) *μj* =

1

1

translate(stepBF*, μ*1*, r*), for some random number *r*; 2) the translated shape of the free bacterium shapeBF is within the movement space for free bacteria resBF:

*μj* (shapeBF) *⊆* resBF, and 3) the new shape of the free bacterium does not over- lap with the other shapes in space: trans(*{*BacF()*}μ′* ) *∩* trans(*{*HIon()*}μ*2 *|*

1

*{*MolB()*}*

*μ*3

1

) = *∅.* If there is no such *μj* , the configuration remains unchanged,

1

and the next movement step can be reduced.

Using the non-stochastic reduction rules, NR.Move and NR.Par, we have:

*{*mov.BacF() + !bind.BacB() + delay@1.5.(BacF()|BacF()) + delay@0.5.(BacF()|HIon())

+ ?kill.DeadBac()*}μ*1 *| {*mov.HIon() + !release.HIon()*}μ*2 *| {*?release.MolF()*}μ*3

*→ {*BacF()*}μ' | {*mov.HIon() + !release.HIon()*}μ*2 *| {*?release.MolF()*}μ*3

1

We next reduce mov.HIon() using NR.Move and NR.Par:

*{*BacF()*}μ' | {*mov.HIon() + !release.HIon()*}μ*2 *| {*?release.MolF()*}μ*3

1

*→ {*BacF()*}μ' | {*HIon()*}μ' | {*?release.MolF()*}μ*3

1 2

Unfolding *{*BacF()*} ′*

*μ*

1

and *{*HIon()*} ′*

2

*μ*

for the sake of clarity:

*{*mov.BacF() + !bind.BacB() + delay@1.5.(BacF()|BacF()) + delay@0.5.(BacF()|HIon())

+?kill.DeadBac()*}μ' | {*mov.HIon() + !release.HIon()*}μ' | {*?release.MolF()*}μ*3

1 2

There are three available reactions, one communication on channel release and two delays. The next reaction will be determined using the corresponding reaction rates. Assume that the next reaction is the communication between HIon() and MolB() through channel release. Using the stochastic rules SR.Com and SR.Par, if the distance between the processes is within release’s reaction radius (dis(*μj , μ*3) *≤* 2*.*0) then the last process reduces to:

2

*{*mov.BacF() + !bind.BacB() + delay@1.5.(BacF()|BacF()) + delay@0.5.(BacF()|HIon())

+?kill.DeadBac()*}μ' | {*HIon()*}μ' | {*MolF()*}μ*3

1 2

Otherwise, the configuration remains unchanged. The next reduction is a movement step, where we reduce all available mov prefixes. This includes reducing mov.BacF(), mov.HIon(), and mov.MolF(), and subsequently interleaving stochastic (reaction) and non-stochastic (movement) reductions.

# Translation from BioScape to 3*π*

We define a translation from BioScape into a variant of Cardelli-Gardner’s 3*π* with definitions instead of replication, and prove that it is sound with respect to the oper- ational semantics (Thm. [6.1](#_bookmark16)). Since 3*π* does not include stochasticity, we translate only the spatial aspect of BioScape into 3*π*.

In an attempt to make this paper

self contained, we include in this sec- tion material from *Processes in Space* [[7](#_bookmark25)]. “3*π* is a proper extension of *π*- calculus with by-value communication of geometric data Δ, data comparisons Δ =*σ* Δ*.P* , and frame shifting *μ*[*P* ].

Δ ::= *xc | ... | μ*[Δ]

*π* ::= ?*σx*(*x′*) *|* !*σx*(Δ) *|* Δ =*σ* Δ

*P* ::= 0 *| π.P | P* + *P′ | P|P′ |* (*νx*)*P | μ*[*P* ]

*D* ::= *∅ | D, X*(*x*)= *P*

(Red Comm) Δ*A '→ є* implies !*σx*(Δ)*.P* + *P′ |*

?*σx*(*y*)*.Q* + *Q′A → P|Q{y/є}*

*≺*

The syntax of 3*π* is shown in Table [1](#_bookmark15).

(Red Cmp) Δ*A*

Δ*′* implies Δ =*σ* Δ*′.PA → P*

Each data term and value has a sort *σ ∈ {***c***,* **a***,* **p***,* **v***,* **m***}*, denoting channels, scalars, points, vectors, and maps re- spectively. A geometric data can be a value or a variable, or a function on val-

(Red Par) *PA → Q* implies *P | RA → Q | R*

(Red Res) *PA → Q* implies (*νx*)*PA →* (*νx*)*Q* (Red *≡*) *P′ ≡ P, PA → Q, Q ≡ Q′*

imply *P′A → Q′*

Table 1 3*π* syntax and reduction

ues and variables, or a frame shift. An action term *π* can be an input ?*σx*(*xj*), an output !*σx*(Δ), or a data comparison Δ =*σ* Δ. The input and output actions are

analogous to *π*-calculus actions, while the data comparison evaluates to *P* if Δ and Δ*j* evaluate to the same value. Actions are restricted by sorting constraints: chan- nels must have sort **c**; *xj* in input must have sort *σ*; Δ in output must have sort *σ*; Δ and Δ*j* in data comparison must have sort *σ*.

Process terms are the standard *π*-calculus terms, plus the frame shift process *μ*[*P* ]: it means running the process *P* in the global frame *A* shifted by the affine map obtained by evaluating the map *μ*.

The reduction relation *A '→*, which relates two processes relative to the global frame

*A*, appears in Table [1](#_bookmark15).

Reduction rules are the rule of a by-value *π*-calculus with data terms Δ, but Red Comm and Red Cmp rules depend on an evaluation relation *A '→*, that evaluates a data Δ to value *ϵ* in a global frame *A*. Data comparison requires the data evaluation

Δ*A* Δ*j*, meaning there is a data value *ϵ* such that Δ*A '→ ϵ* and Δ*jA '→ ϵ*.”

*≺*

We define three functions *−*SPA, *−*PRO and *−*DEF to translate spatial con-

figurations, processes and definitions, respectively. In *A* SPA

*X*;*E*;*D*;r

, *A* is a spatial

configuration, *X* is a triple containing the movement space, shape and step of the current definition, *E* is a set of channel declarations, *D* is a set of definitions, and *r* is a real number. We will abuse the notation and consider *E* and *D* sets or lists.

Similarly for *P* PRO and *D* DEF *′* .

*X*;*E*;*D*;r *X*;*E*;*D* ;r

**[Process Translation]**

(*νx*@r*,*rad)*.P* PRO

*X*;*E*;*D*;r

PRO

= *P*

*X*;*<x*@*r,rad>,E*;*D*;r

P.Res

PRO

*P | Q*

*X*;*E*;*D*;r

PRO

*X*;*E*;*D*;r

= *P*

*| Q*

PRO

*X*;*E*;*D*;r

P.Par

PRO

*X*(*y*)

*X*;*E*;*<X*(*x*)@*ξ,σ,ω*=*M>,D*;r

PRO

**0**

*X*;*E*;*D*;r

=*X*(*y*) P.Def

=**0** P.Nil

PRO

*π.P* + *M*

=*{ π.P*

*X*;*E*;*D*;r

PRO

*X*;*E*;*D*;r

PRO

*X*;*E*;*D*;r

*}∪ M*

P.Cho

[delay@r*.P *PRO PRO](mailto:delay@r.P)

*X*;*E*;*D*;r

!*x*(*y*)*.P*

*X*;*<x*@*r,rad>,E*;*D*;r

=(*νm*)(!**c***m*() *|*?**c***m*()*. * *P *PRO

=?**c***cx*(*ok*)*.*?**c***ok*(*pos*)*.*?**p***pos*(*p*)*.*

*X*;*E*;*D*;r

) *m /∈ fn*(*P* ) P.Del

PRO

?*x*(*z*)*.Q*

*X*;*<x*@*r,rad>,E*;*D*;r

*abs*(*p −* H) *≤ rad* =**a** 1*.*!**c***ok*(*y*)*. * *P *PRO

*cx, ok, pos,p ∈/* fn(*P* ) P.Out

*X*;*<x*@*r,rad>,E*;*D*;r

=(*νok, pos*)(!**c***cx*(*ok*)*.*!**c***ok*(*pos*)*.*

PRO

mov*.P*

*<ξ,σ,ω>*;*E*;*D*;r

!**p***pos*(H)*.*?**c***ok*(*z*)*. * *Q * PRO

=*into*(T(*↑* (r*, ω*))[H]*, σ, ξ*) =**a** 1*.*

*X*;*<x*@*r,rad>,E*;*D*;r

) *cx ∈/* fn(*Q*) P.In

PRO

T(*↑* (r*, ω*))[ *P*

*<ξ,σ,ω>*;*E*;*D*;r

] P.Mov

**[Spatial Configuration Translation]**

SPA

*{P}μ*

*X*;*E*;*D*;r

PRO

*X*;*E*;*D*;r

= *μ*[ *P*

] S.Loc

SPA

*A | B*

*X*;*E*;*D*;r

(*νx*@r*,*rad)*.A* SPA

SPA

*X*;*E*;*D*;r

= *A*

*| B*

= *A* SPA

SPA

*X*;*E*;*D*;r

S.Par

S.Res

*X*;*E*;*D*;r *X*;*<x*@*r,rad>,E*;*D*;r

**[Definition Translation]**

*X*(*x*)@*ξ, σ, ω* = *M, D′* DEF

DEF

*X*;*E*;*D*;r

*∅*

= *∅* D.Nil

=

*X*;*E*;*<X*(*x*)@*ξ,σ,ω*=*M>,D*;r

PRO

(*X*(*x*)= *M*

*<ξ,σ,ω>*;*E*;*<X*(*x*)@*ξ,σ,ω*=*M>,D*;r

*′* DEF

*X*;*E*;*D*;r

)*, D*

D.Def

The translation of a BioScape choice process is the set of translations of its branches, because 3*π* does not allow reduction under its choice operator. Otherwise the reductions obtained from translating the branches would be blocked. Conse- quently, we extend naturally 3*π*’s reduction and congruence to sets (see Table [2](#_bookmark17)), and we also define homomorphic extensions of parallel composition, frame shift, prefix and restriction over sets. The trickiest rules are the ones for translating communication, P.In for input and P.Out for output. First, the input sends to the output two private channels, *ok* and *pos*, and subsequently its position (H) via channel *pos*. The output calculates the distance between itself and the position sent by the input with the function *abs*(*p −* H), i.e. the absolute value of the difference between the positions of the processes. If this distance is less than the reaction radius (rad), then the output sends a communication on the channel of success and performs the process *P* : when the input receives a communication on the *ok* channel, it performs process *Q*.

For the translation of movement in Rule P.Mov, we assume a function *into*(*p, sh, sp*) that checks whether a shape *s*, centered at point *p*, is contained within space *sp*. T(*v*) is a translation map with vector *v*, and *↑* (r*, ω*) is a vector of length *ω* and direction r. The translation checks whether *into*(T(*↑* (r*, ω*))[H]*, σ, ξ*) corresponding to condition *μj*(*σ*) *⊆ ξ* in NR.Move. Finally, since delay is not a prefix in 3*π*, it is translated as a communication over a channel.

**Theorem 6.1 (Soundness)** *If E ▶D A → B or E ▶D A →r B then there exists*

*∅*;*E*;*D*;*r*

*S such that A SPA*

*∅*;*E*;*D*;*r*

*A →∗*

*S and S ≡ B SPA*

*for some number r.*

In our translation, the global frame *A* is the identity. Although sound, this translation is not complete in the sense that not all reductions in the trans- lated code arise from corresponding re- ductions in the source code. Achiev- ing completeness is more complex and would require collision detection and a more refined mechanism for checking proximity.

# Conclusions

Let *S* and *T* be sets

* 1. *S | T* = *{*(*P | Q*) *| P ∈ S,Q ∈ T}*
  2. *μ*[*{∅}*]= *∅* and

*μ*[*{P}∪ S*]= *{μ*[*P* ]*}∪ μ*[*S*]

* 1. *π.S* = *{π.P | P ∈ S}*
  2. (*νm*)*.S* = *{*(*νm*)*.P | P ∈ S}*

(Set.Par) *SA → S’* implies *S | TA → S’ | T* (Set.Red) *PA → P′* imp. *{P}∪ QA → {P′}∪ Q* (S.Set.Par) *S ≡ S’* imp. *S | T ≡ S’ | T* (S.Set.Red) *P ≡ P′* imp. *{P}∪ Q ≡ {P′}∪ Q*

Table 2 Operations, Reduction and Congruence on Sets

We define BioScape for the modeling and simulation of complex bacteria-materials interactions. BioScape builds on 3*π* [[7](#_bookmark25)] and SpacePi [[15](#_bookmark33)] merging an affine space geometry, reaction radius [1](#_bookmark18) and reaction boundary. This combination of features is strictly motivated by the nature of the models we are capturing, as described in the introduction: stochasticity, movement, individual process location (in con- trast with homogeneously mixed reactants in a volume), interaction in proximity, and movement confinement. We formulate a reduction semantics for BioScape and

1 Although this concept may be derived in 3*π*, it is convenient in practice to have as primitive.

demonstrate it in an example of pH-triggered drug release in the presence of bacte- rial infection. Furthermore, we validate BioScape with a translation into Cardelli- Gardner’s 3*π*, and prove its soundness with respect to the operational semantics. Complete technical details can be found in the companion technical report [[11](#_bookmark30)].

The process calculus that most closely resembles BioScape is the Shape Calculus [[3](#_bookmark21)], a CCS-like timed calculus, with simulating tool BioShape [[5](#_bookmark26)]. These are some of the differences. BioScape is stochastic, but the Shape Calculus is not. The Shape Calculus does not allow dynamic creation of channels, but BioScape does. Agents in BioScape can be modified by affine transformations useful in modeling phenomena such as cell growth, but the Shape Calculus only allows movement specified with a velocity vector. The Shape Calculus has a time primitive for describing a delay, while BioScape has a stochastic delay. The specification of an agent in BioScape de- scribes an area where it is allowed to be, but the Shape Calculus does not. This area is instrumental in describing biomaterials such as antibacterial surfaces and prevent- ing bacteria from penetrating the surface while allowing antibacterial molecules to do so. On the other hand, such behavior would have to be programmed in the description of the agent in the Shape Calculus.

We develop an implementation of BioScape based on SPiM, and show prelim- inary simulation results in agreement with wet-lab experiments. We are currently working on a parallel simulation algorithm implemented in CUDA, and our proto- type implementation can handle one million agents in 3D space.

Our long-term modeling objective is to identify combinations of substrate vari- ables that most significantly inhibit bacterial colonization and promote tissue inte- gration.

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