

Palmer amaranth (*Amaranthus palmeri*) adaptation to agroecosystems

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2 ABSTRACT

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INTRODUCTION

Palmer amaranth (*Amaranthus palmeri* S. Watson) is currently considered one of the most economically damaged weed species to cropping systems in the United States. The species has showed a remarkable capacity to evolve resistance to herbicides. To date, Palmer amaranth has evolved resistance to eight herbicide sites of action (Heap, 2021), increasing the weed management complexity (Lindsay et al., 2017). Unmanaged Palmer amaranth in competition for water, light and nutrients can drastically impact on crop yields (Berger et al., 2015). For example, Palmer amaranth is documented with potential to reduce 91%, 68%, and 54% of corn (Massinga et al., 2001), soybean (Klingaman and Oliver, 1994), and cotton (Morgan et al., 2001) yields, respectively. Thus, Palmer amaranth poses an economical risk to sustainable agriculture.

Palmer amaranth is a fast growing summer annual forb indigenous to Sonoran Desert (Sauer, 1957). The species would eventually emerge as a threat to US agriculture in the 1990s. Palmer amaranth weediness is likely a result of human-assisted selection in combination with species biology. Farm mechanization, conservation agriculture (e.g., no-till), and reliance on herbicides for weed management are the main human-mediated selection of Palmer amaranth into cropping systems (Ward et al., 2013). On the other hand, Palmer amaranth is a prolific seed producer with a C4 photosynthetic apparatus (Wang et al., 1992). With a dioecy nature, Palmer amaranth male and female plants are obligate outcrosser species, increasing the chances of exchanging adaptive traits among plants (Oliveira et al., 2018). Also, Palmer amaranth small seed size (e.g, 1 mm) tend to thrive in no-tillage systems (Price et al., 2011), and spread across locations through farm equipment (Sauer, 1972), manure (Hartzler and Anderson, 2016), animals (Farmer et al.,

25 2017), and plant propagules (Yu et al., 2021). The dispersal capacity of Palmer amaranth make the species
26 one of the most successful cases of weed adaption to cropping systems.

27 Light and temperature are likely the main environment requirements for Palmer amaranth successful
28 adaptation. Palmer amaranth is reported with an extended germination period (Jha et al., 2010). Germination
29 of Palmer amaranth is triggered by 18 C soil temperature (Keeley et al., 1987), and optimal germination
30 and biomass production occur at 35/30 C day and night temperatures (Guo and Al-Khatib, 2003). Water
31 has not shown to limit Palmer amaranth fitness. Under continuous water stress, Palmer amaranth survived
32 and produced at least 14000 seeds plant-1 (Chahal et al., 2018). Also, seeds from Palmer amaranth growing
33 under water stress conditions were heavier, less dormant, and prompt for germination (Matzrafi et al.,
34 2021). The continuous global temperature warming can impact agriculture and promote niches for Palmer
35 amaranth invasion/adaptation into new environments. Currently, it is estimated that the greatest climatic
36 risk of Palmer amaranth establishment are agronomic crops in Australia and Sub-Saharan Africa (Kistner
37 and Hatfield, 2018). Temperature is a key factor limiting Palmer amaranth expansion to cooler geographies
38 (Briscoe Runquist et al., 2019); however, under future climate change Palmer amaranth is likely to expand
39 northward into Canada and Northern Europe (Kistner and Hatfield, 2018; Briscoe Runquist et al., 2019).

40 Palmer amaranth is already documented in agronomic crops of South America (Larran et al., 2017;
41 Küpper et al., 2017) and Southern Europe (Milani et al., 2021). In the US, Palmer amaranth is well
42 established at crop (Garetson et al., 2019) and non-crop land (Bagavathiannan and Norsworthy, 2016) in
43 the warm southern United States but its range is expanding to cool temperatures northward. For example,
44 herbicide resistant Palmer amaranth is widespread in Nebraska (Oliveira et al., 2021), Michigan (Kohrt
45 et al., 2017), and Connecticut (Aulakh et al., 2021). Successful cases of Palmer amaranth invasion and
46 near to eradication is reported in Minnesota (Yu et al., 2021). No Palmer amaranth actively growing
47 was found in Canada; however, Palmer amaranth seeds was detected in sweet potato slips (Page et al.,
48 2021). Nonetheless, it seems fated the need to manage Palmer amaranth in agronomic crops throughout
49 multiple environments in the near future. Therefore, strategies on Palmer amaranth management should
50 encompass the agroecosystem level but not only attempts to eradicate the weed. Most tactics to manage
51 Palmer amaranth are based on technology fixes (Scott, 2011), which are short-term (e.g., herbicide and/or
52 tillage) rather than long-term weed management. Strategies to manage Palmer amaranth should be built on
53 minimizing the species ability to adapt, grow and develop into agroecosystems.

54 The continuous Palmer amaranth dispersal and potential establishment into northern United States warrant
55 investigations on species morphology in such environments. Understanding Palmer amaranth biology and
56 growing strategies under different agroecosystems can enhance our knowledge on species adaptation. It
57 can also aid on designing proactive and ecological tactics to limit the species range expansion, reduce its
58 negative impact, and design resilient and sustainable farming systems (MacLaren et al., 2020). Therefore,
59 the objective of this study was to investigate the flowering pattern, biomass production, and height of
60 Palmer amaranth growing under corn, soybean and fallow at two timings across five locations in the United
61 States Midwest.

MATERIAL AND METHODS

62 Plant material and growing conditions

63 The study was performed with a *A. palmeri* accession (Per1) from Perkins County, Nebraska. Per1
64 accession collection is documented with no reported herbicide resistance (Oliveira et al., 2021). Three
65 weeks prior to the field experiment, seeds were planted in plastic trays containing potting-mix. Emerged
66 seedlings (1 cm) were transplanted into 200 cm⁻³ plastic pots (a plant pot-1). Palmer amaranth seedlings

were supplied with adequate water and kept under greenhouse conditions at Arlington, Clay Center, Lincoln, and Macomb; and kept outdoors in Grant. Palmer amaranth seedlings were kept under greenhouse/outdoors until the onset of the experiment (2-3 leaf stage/5 to 8 cm height).

Field study

The experiment was conducted in 2018 and 2019 under field conditions at five locations: Arlington (Washington County, Wisconsin), Clay Center (Clay County, Nebraska), Grant (Perkins County, Nebraska), Lincoln (Lancaster County, Nebraska), and Macomb (McDonough County, Illinois).

A glyphosate-resistant soybean cultivar (DSR-1950 R2Y at 296,400 seeds ha⁻¹), and a corn hybrid were planted at

Monthly mean air temperature and sum precipitation were obtained using Daymet weather data from June through September across the five locations in 2018 and 2019 (Correndo et al., 2021) (Figure 1)

The field experimental unit were six adjacent 9.1 m wide (12 rows at 72.2 cm row spacing) by 10.7 m long. Each experimental unit was planted with corn or soybean (DSR-1950 R2Y at 296,400 seeds ha⁻¹), or under fallow condition. Palmer amaranth seedlings (potting mix + two seedlings) were and gently transferring to the ground (6 cm deep and 8 cm wide). Twenty-four plants were equidistantly placed (0.76 m apart) between rows within each agroecosystems. After a week, one was eliminated and one was kept. There were two transplant timing: first (June 1) and second (July 1). There were 24 Palmer amaranth plants in each experimental unit, with a total of 144 plants for each location. The study was repeated twice.

After transplanting, Palmer amaranth flowering was monitored until the end of the study. When a plant started flowering, the day was recorded, plant sex was identified as male or female, and plant height was measured from soil surface to the plant top. Then, aboveground plant biomass was harvest near soil surface and oven dried at 65 C until reaching constant weight before the weight of biomass (g plant⁻¹) was recorded.

Statistical analyses

The statistical analyses were performed using R statistical software version 4.0.1. Data across locations and year were combined.

The cumulative Palmer amaranth flowering estimation was determined using a asymmetrical three parameter log logistic Weibull model of the drc package (Ritz et al., 2015).

$$Y(x) = 0 + (d - 0)\exp(-\exp(b(\log(x) - e)))$$

In this model, Y is the Palmer amaranth cumulative flowering, d is the upper limit (set to 100), and e is the XXX, and x day of year (doy).

The doy for 10, 50, and 90% Palmer amaranth cumulative flowering were determined using the *ED* function of drc package. Also, the 10, 50, and 90% Palmer amaranth cumulative flowering were compared among agroecosystems and timings using the *EDcomp* function of drc package. The *EDcomp* function compares the ratio of cumulative flowering using t-statistics, where P-value < 0.05 indicates that we fail to reject the null hypothesis.

Palmer amaranth height and biomass were performed with a linear mixed model using *lmer* function from "lme4" package (Bates et al., 2015). Plant height and biomass were transformed to meet model assumption of normality. In the model, agroecosystem (crop, soybean, fallow) was the fixed effect and year nested with location the random effects. Analysis of variance was performed with *anova* function from "car" package

106 (Fox and Weisberg, 2018). Marginal means and compact letter display were estimated with *emmeans* and
107 *cld* from packages “*emmeans*” and *multcomp* (Hothorn et al., 2008).

RESULTS

108 Cumulative flowering

109 Palmer amaranth growing in corn resulted in a longer flowering pattern compared to fallow and soybean
110 (Figure 2A). Palmer amaranth reached 10% flowering in soybean at doy 180, which was slightly different
111 from fallow (doy 180.9; $P = 0.01$) and corn (doy 181.7; $P = 0.00$). Nonetheless, the 10% cumulative
112 Palmer amaranth flowering in soybean, fallow and corn occurred at the end of June. The 50% Palmer
113 amaranth cumulative flowering occurred in July. For example, Palmer amaranth reached 50% in fallow at
114 doy 193.4, followed by soybean (doy 194.8), corn (doy 206.6). Similar trend was observed at 90% Palmer
115 amaranth cumulative flowering. Palmer amaranth growing in corn reached 90% flowering at doy 252.6
116 (early September), which was 37.8 and 32.2 days after Palmer amaranth 90% flowering in fallow and
117 soybean, respectively.

118 Palmer amaranth cumulative flowering from second transplanting timing ranged from mid July to mid
119 September (Figure 2). All cumulative flowering comparisons among agroecosystems were significant (P
120 < 0.00) Palmer amaranth growing in fallow resulted in earlier flowering time compared to soybean and
121 corn. Palmer amaranth growing in fallow reached 10%, 50%, and 90% flowering time at day 203.8, 214.4,
122 and 232.2, respectively. Palmer amaranth growing in soybean reached 10% flowering at doy 210.9, which
123 was 6 days prior to corn (P -value = 0.00). Similar trend was observed at 50% flowering, whereas Palmer
124 amaranth reached 50% flowering in corn (doy 233.0) 4 days after soybeans (doy 228.9; $P = 0.00$). The 90%
125 Palmer amaranth cumulative flowering occurred at same day in corn (260.9) and soybean (260.5; $P = 0.66$).

126 Height and biomass

127 Palmer amaranth accumulated more biomass when growing in fallow compared to Palmer amaranth
128 growing in soybean and corn (Figure 3). At first transplanting time, Palmer amaranth biomass was 78.3
129 g plant-1, 26.4 g plant-1 and 14.9 g plant-1 in fallow, soybean and corn, respectively. Palmer amaranth
130 produce less biomass at second transplanting time. For example, Palmer amaranth growing in fallow
131 resulted in 49.2 g plant-1, 3.4 g plant-1 in soybean and 1.4 g plant when growing in corn.

132 For example, Palmer amaranth growing at first cohort accumulated 90%, 85% and 31% more biomass
133 when compared to second cohort at corn, soybean and fallow, respectively

134 In addition, Palmer amaranth growing in corn and soybean at second cohort had a similar end-season
135 flowering pattern.

DISCUSSION

136 Our study showed the Palmer amaranth flowering pattern, grow and development varied within
137 agroecosystems and timing. In general, Palmer amaranth produced more biomass and taller plants
138 when growing at first cohort rather than second cohort. At first cohort, resources (e.g., soil nutrients) and
139 conditions (e.g., light) were more timely available for both species, crop and weed. Thus, Palmer amaranth
140 growing strategy was to mimic the crop species (Figure 4). In contrast, at second cohort, Palmer amaranth
141 grow and development was limited due to the crop competitive ability at advanced development stages.

142 Taller Palmer amaranth plants is a strategy to compete for light in between crop rows in absence of canopy.
143 For example, at first cohort, corn and Palmer amaranth had abundant light to grow and develop, which
144 likely aid its extended the flowering pattern. However, at second cohort, Palmer amaranth was transplanted

145 when corn canopy was nearly closed, which reduced Palmer amaranth competition. As a result, Palmer
146 amaranth height and biomass was nearly 50% and 90% lower compared to its first cohort, respectively.
147 Similar trend was observed to Palmer amaranth growing in soybeans. Palmer amaranth growing without
148 crop competition produced highest amount of biomass. The Palmer amaranth strategy in fallow was to
149 invest biomass in growing plant width and height. Nonetheless, Palmer amaranth produced 31% less
150 biomass in second cohort compared to first cohort timing. Therefore, crop competition was not the only
151 factor limiting Palmer amaranth growing in the second cohort.

152 It was not evaluate seed production due to Palmer amaranth harvest at early flowering. Nonetheless, it is
153 reported a strong positive correlation between Palmer amaranth biomass and seed production (Schwartz
154 et al., 2016; Spaunhorst et al., 2018). In our study, Palmer amaranth growing at first cohort accumulated
155 nearly 50% more overall biomass when compared to second cohort. Therefore, Palmer amaranth plants
156 growing in the second cohort is likely to produce less seeds regardless the agroecosystem. Seed production
157 and deposition in the seedbank is a key factor for species perpetuation (Menges, 1987). Palmer amaranth
158 germination is influenced by soil depth and seed viable seeds were fount after 36 months buried in the
159 seedbank (Sosnoskie et al., 2013). Therefore, tactics that promote crop advantage against Palmer amaranth,
160 including early crop planting, preemergence applied herbicide, and crop residue (e.g. cover crops) can
161 minimize the negative impact of Palmer amaranth in agroecosystems.

162 Palmer amaranth has shown to be responsive to thermal units (e.g, growing degree days). A study
163 in North Carolina demonstrated that Palmer amaranth emergence in response to soil temperature, with
164 seedlings emerging before July prior to crop canopy closure (Piskackova et al., 2021). In addition, in a
165 bareground study, Palmer emergence pattern in Nebraska was influenced by soil management.
166 Early tillage resulted in 90% Palmer amaranth emergence in June

167 can cause significant soybean yield loss (Korres et al., 2019)

DISCLOSURE/CONFLICT-OF-INTEREST STATEMENT

168 The authors declare that the research was conducted in the absence of any commercial or financial
169 relationships that could be construed as a potential conflict of interest.

AUTHOR CONTRIBUTIONS

170 RW: designed the experiments; AJ, CP, MB, MO, and SS: conducted the experiments; MO: analyzed the
171 data and wrote the manuscript; AJ, CP, MB, MO, SS, and RW: conceptualized the research. All authors
172 reviewed the manuscript.

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1 SUPPLEMENTAL DATA

175 Supplementary Material should be uploaded separately on submission, if there are Supplementary Figures,
176 please include the caption in the same file as the figure. LaTeX Supplementary Material templates can be
177 found in the Frontiers LaTeX folder

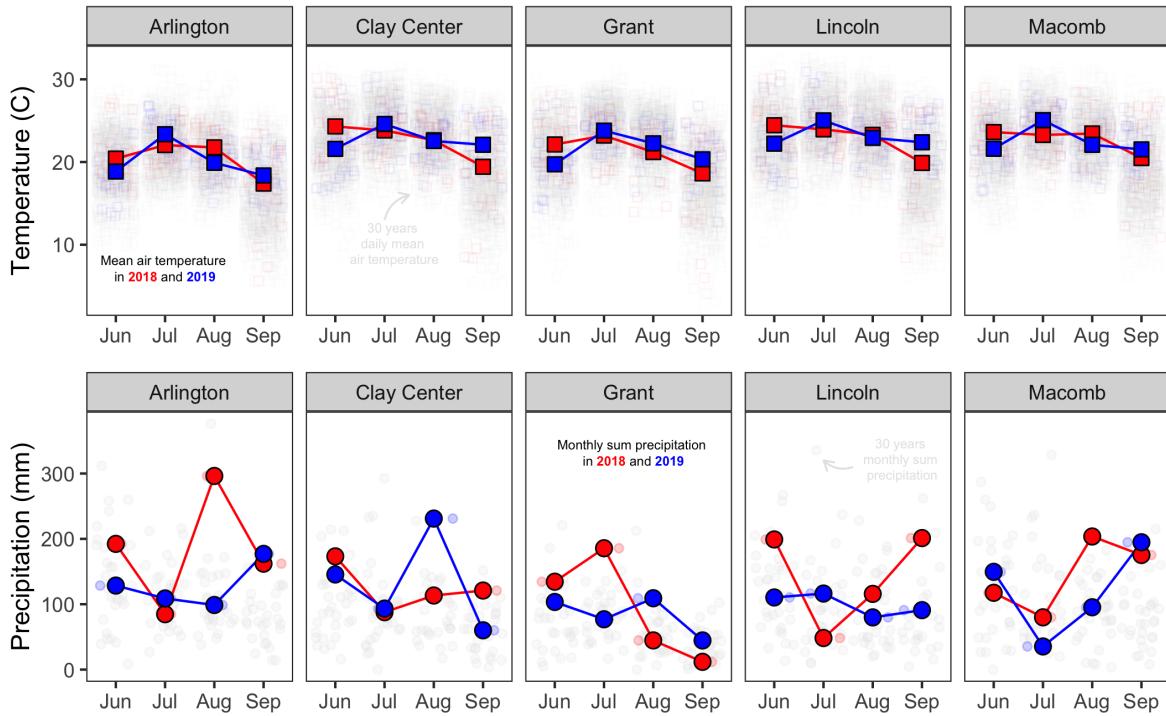


Figure 1. Mean average temperature (C) and monthly sum precipitation (mm) at Arlington, WI, Clay Center, NE, Grant, NE, Lincoln, NE and Macomb, IL

2 REFERENCES

178 A reference list should be automatically created here. However it won't. Pandoc will place the list of
 179 references at the end of the document instead. There are no convenient solution for now to force Pandoc to
 180 do otherwise. The easiest way to get around this problem is to edit the LaTeX file created by Pandoc before
 181 compiling it again using the traditional LaTeX commands.

FIGURES

- 182 Aulakh, J. S., Chahal, P. S., Kumar, V., Price, A. J., and Guillard, K. (2021). Multiple herbicide-resistant
 183 Palmer amaranth (*Amaranthus palmeri*) in Connecticut: Confirmation and response to POST herbicides.
 184 *Weed Technology* 35, 457–463. doi:10.1017/wet.2021.6.
- 185 Bagavathiannan, M. V., and Norsworthy, J. K. (2016). Multiple-Herbicide Resistance Is Widespread in
 186 Roadside Palmer Amaranth Populations. *PLOS ONE* 11, e0148748. doi:10.1371/journal.pone.0148748.
- 187 Bates, D., Mächler, M., Bolker, B., and Walker, S. (2015). Fitting Linear Mixed-Effects Models Using
 188 *Lme4*. *Journal of Statistical Software* 67, 1–48. doi:10.18637/jss.v067.i01.
- 189 Berger, S. T., Ferrell, J. A., Rowland, D. L., and Webster, T. M. (2015). Palmer Amaranth (*Amaranthus*
 190 *palmeri*) Competition for Water in Cotton. *Weed Science* 63, 928–935. doi:10.1614/WS-D-15-00062.1.
- 191 Briscoe Runquist, R. D., Lake, T., Tiffin, P., and Moeller, D. A. (2019). Species distribution models
 192 throughout the invasion history of Palmer amaranth predict regions at risk of future invasion and reveal
 193 challenges with modeling rapidly shifting geographic ranges. *Sci Rep* 9, 2426. doi:10.1038/s41598-
 194 018-38054-9.

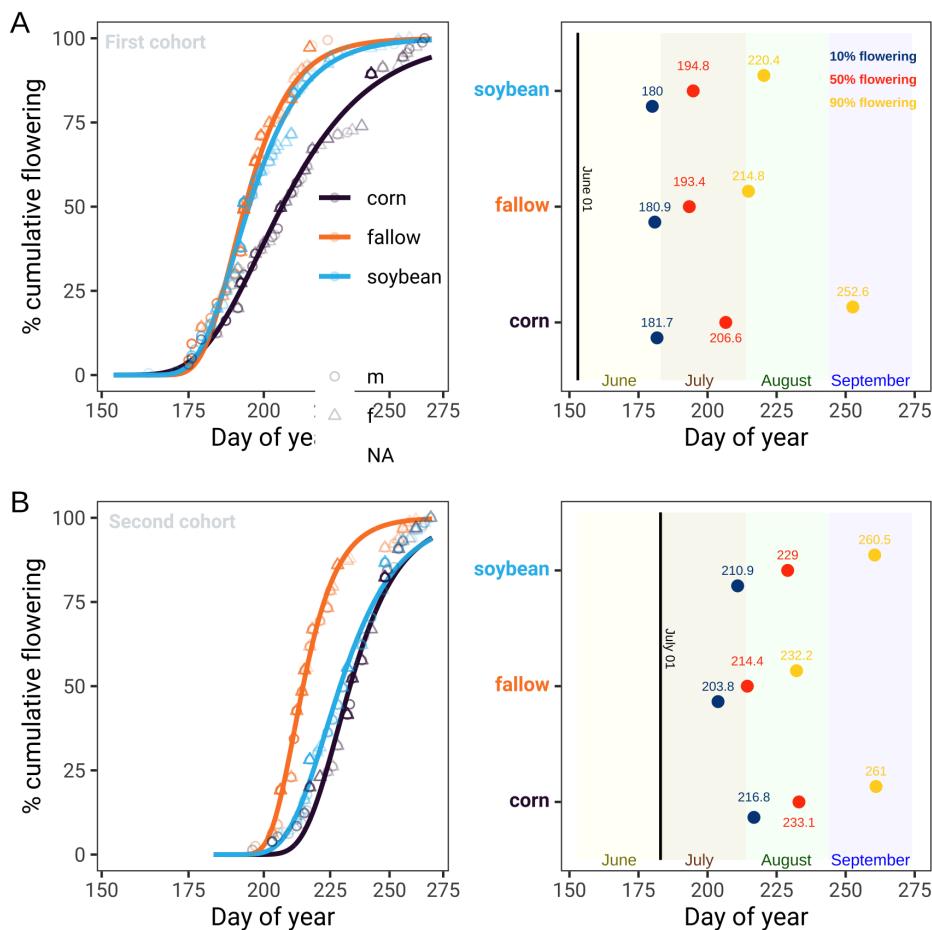


Figure 2. Cumulative flowering of Palmer amaranth at first and second transplant timing (A) and day of year of 10, 50, and 90 cumulative flowering at first and second transplant timing (B)

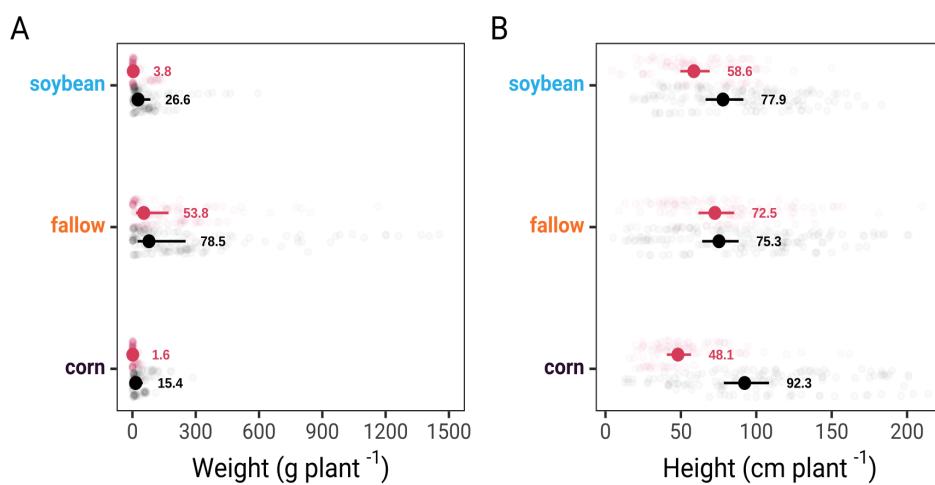


Figure 3. Palmer amaranth biomass (A) and height (B) growing in corn, fallow, and soybean across Arlington, Clay Center, Grant, Lincoln, and Macomb



Figure 4. Harvest Palmer amaranth plants at 40 days after first transplant timing. From left to right, Palmer amaranth growing in fallow, soybean and corn in Arlington, Wisconsin

- 195 Chahal, P. S., Irmak, S., Jugulam, M., and Jhala, A. J. (2018). Evaluating Effect of Degree of Water Stress
196 on Growth and Fecundity of Palmer amaranth (*Amaranthus palmeri*) Using Soil Moisture Sensors.
197 *Weed Science* 66, 738–745. doi:10.1017/wsc.2018.47.
- 198 Correndo, A. A., Moro Rosso, L. H., and Ciampitti, I. A. (2021). Retrieving and processing agro-
199 meteorological data from API-client sources using R software. *BMC Research Notes* 14, 205.
200 doi:10.1186/s13104-021-05622-8.
- 201 Farmer, J. A., Webb, E. B., Pierce, R. A., and Bradley, K. W. (2017). Evaluating the potential for weed seed
202 dispersal based on waterfowl consumption and seed viability. *Pest Management Science* 73, 2592–2603.
203 doi:10.1002/ps.4710.
- 204 Fox, J., and Weisberg, S. (2018). *An R Companion to Applied Regression*. SAGE Publications Available at:
205 <http://books.google.com?id=uPNrDwAAQBAJ>.
- 206 Garetson, R., Singh, V., Singh, S., Dotray, P., and Bagavathiannan, M. (2019). Distribution of herbicide-
207 resistant Palmer amaranth (*Amaranthus palmeri*) in row crop production systems in Texas. *Weed
Technology* 33, 355–365. doi:10.1017/wet.2019.14.
- 209 Guo, P., and Al-Khatib, K. (2003). Temperature effects on germination and growth of redroot pigweed
210 (*Amaranthus retroflexus*), Palmer amaranth (*A. Palmeri*), and common waterhemp (*A. rudis*). *Weed
Science* 51, 869–875. doi:10.1614/P2002-127.
- 212 Hartzler, B., and Anderson, M. (2016). Palmer amaranth: It's here, now what? 10.
- 213 Heap, I. (2021). Internation Herbicide-Resistant Weed Database. Available at: <http://www.weedscience.org/Home.aspx> [Accessed July 26, 2021].
- 215 Hothorn, T., Bretz, F., and Westfall, P. (2008). Simultaneous Inference in General Parametric Models.
216 *Biometrical Journal* 50, 346–363. doi:10.1002/bimj.200810425.
- 217 Jha, P., Norsworthy, J. K., Riley, M. B., and Bridges, W. (2010). Annual Changes in Temperature and Light
218 Requirements for Germination of Palmer Amaranth (*Amaranthus palmeri*) Seeds Retrieved from Soil.
219 *Weed Science* 58, 426–432. doi:10.1614/WS-D-09-00038.1.

- 220 Keeley, P. E., Carter, C. H., and Thullen, R. J. (1987). Influence of Planting Date on Growth of Palmer
221 Amaranth (*Amaranthus palmeri*). *Weed Science* 35, 199–204. doi:10.1017/S0043174500079054.
- 222 Kistner, E. J., and Hatfield, J. L. (2018). Potential Geographic Distribution of Palmer
223 Amaranth under Current and Future Climates. *Agricultural & Environmental Letters* 3, 170044.
224 doi:10.2134/ael2017.12.0044.
- 225 Klingaman, T. E., and Oliver, L. R. (1994). Palmer Amaranth (*Amaranthus palmeri*) Interference in
226 Soybeans (*Glycine max*). *Weed Science* 42, 523–527. doi:10.1017/S0043174500076888.
- 227 Kohrt, J. R., Sprague, C. L., Nadakuduti, S. S., and Douches, D. (2017). Confirmation of a Three-Way
228 (Glyphosate, ALS, and Atrazine) Herbicide-Resistant Population of Palmer Amaranth (*Amaranthus*
229 *palmeri*) in Michigan. *Weed Science* 65, 327–338. doi:10.1017/wsc.2017.2.
- 230 Korres, N. E., Norsworthy, J. K., and Mauromoustakos, A. (2019). Effects of Palmer Amaranth
231 (*Amaranthus palmeri*) Establishment Time and Distance from the Crop Row on Biological and
232 Phenological Characteristics of the Weed: Implications on Soybean Yield. *Weed Science* 67, 126–135.
233 doi:10.1017/wsc.2018.84.
- 234 Küpper, A., Borgato, E. A., Patterson, E. L., Netto, A. G., Nicolai, M., Carvalho, S. J. P. de, Nissen, S.
235 J., Gaines, T. A., and Christoffoleti, P. J. (2017). Multiple Resistance to Glyphosate and Acetolactate
236 Synthase Inhibitors in Palmer Amaranth (*Amaranthus palmeri*) Identified in Brazil. *Weed Science* 65,
237 317–326. doi:10.1017/wsc.2017.1.
- 238 Larran, A. S., Palmieri, V. E., Perotti, V. E., Lieber, L., Tuesca, D., and Permingeat, H. R. (2017). Target-site
239 resistance to acetolactate synthase (ALS)-inhibiting herbicides in *Amaranthus palmeri* from Argentina.
240 *Pest Management Science* 73, 2578–2584. doi:10.1002/ps.4662.
- 241 Lindsay, K., Popp, M., Norsworthy, J., Bagavathiannan, M., Powles, S., and Lacoste, M. (2017). PAM:
242 Decision Support for Long-Term Palmer Amaranth (*Amaranthus palmeri*) Control. *Weed Technology*
243 31, 915–927. doi:10.1017/wet.2017.69.
- 244 MacLaren, C., Storkey, J., Menegat, A., Metcalfe, H., and Dehnen-Schmutz, K. (2020). An ecological
245 future for weed science to sustain crop production and the environment. A review. *Agron. Sustain. Dev.*
246 40, 24. doi:10.1007/s13593-020-00631-6.
- 247 Massinga, R. A., Currie, R. S., Horak, M. J., and Boyer, J. (2001). Interference of Palmer amaranth in corn.
248 *Weed Science* 49, 202–208. doi:10.1614/0043-1745(2001)049[0202:IOPAIC]2.0.CO;2.
- 249 Matzrafi, M., Osipitan, O. A., Ohadi, S., and Mesgaran, M. B. (2021). Under pressure: Maternal effects
250 promote drought tolerance in progeny seed of Palmer amaranth (*Amaranthus palmeri*). *Weed Science*
251 69, 31–38. doi:10.1017/wsc.2020.75.
- 252 Menges, R. M. (1987). Weed Seed Population Dynamics during Six Years of Weed Management Systems
253 in Crop Rotations on Irrigated Soil. *Weed Science* 35, 328–332. Available at: <http://www.jstor.org/stable/4044593>.
- 255 Milani, A., Panizzo, S., Farinati, S., Iamonico, D., Sattin, M., Loddo, D., and Scarabel, L. (2021). Recent
256 Discovery of *Amaranthus palmeri* S. Watson in Italy: Characterization of ALS-Resistant Populations
257 and Sensitivity to Alternative Herbicides. *Sustainability* 13, 7003. doi:10.3390/su13137003.

- 258 Morgan, G. D., Baumann, P. A., and Chandler, J. M. (2001). Competitive Impact of Palmer Amaranth
259 (Amaranthus palmeri) on Cotton (*Gossypium hirsutum*) Development and Yield. *Weed Technology* 15,
260 408–412. doi:10.1614/0890-037X(2001)015[0408:CIOPAA]2.0.CO;2.
- 261 Oliveira, M. C., Gaines, T. A., Patterson, E. L., Jhala, A. J., Irmak, S., Amundsen, K., and Knezevic, S.
262 Z. (2018). Interspecific and intraspecific transference of metabolism-based mesotrione resistance in
263 dioecious weedy Amaranthus. *The Plant Journal* 96, 1051–1063. doi:10.1111/tpj.14089.
- 264 Oliveira, M. C., Giacomini, D. A., Arsenijevic, N., Vieira, G., Tranel, P. J., and Werle, R. (2021).
265 Distribution and validation of genotypic and phenotypic glyphosate and PPO-inhibitor resistance in
266 Palmer amaranth (Amaranthus palmeri) from southwestern Nebraska. *Weed Technology* 35, 65–76.
267 doi:10.1017/wet.2020.74.
- 268 Page, E. R., Nurse, R. E., Meloche, S., Bosveld, K., Grainger, C., Obeid, K., Filotas, M., Simard, M.-J., and
269 Laforest, M. (2021). Import of Palmer amaranth (Amaranthus palmeri S. Wats.) Seed with sweet potato
270 (*Ipomea batatas* (L.) Lam) slips. *Can. J. Plant Sci.*, CJPS-2020-0321. doi:10.1139/CJPS-2020-0321.
- 271 Piskackova, T. A. R., Reberg-Horton, S. C., Richardson, R. J., Jennings, K. M., Franca, L., Young, B. G.,
272 and Leon, R. G. (2021). Windows of action for controlling palmer amaranth (Amaranthus palmeri)
273 using emergence and phenology models. *Weed Research* 61, 188–198. doi:10.1111/wre.12470.
- 274 Price, A. J., Balkcom, K. S., Culpepper, S. A., Kelton, J. A., Nichols, R. L., and Schomberg, H. (2011).
275 Glyphosate-resistant Palmer amaranth: A threat to conservation tillage. *Journal of Soil and Water
276 Conservation* 66, 265–275. doi:10.2489/jswc.66.4.265.
- 277 Ritz, C., Baty, F., Streibig, J. C., and Gerhard, D. (2015). Dose-Response Analysis Using R. *PLOS ONE*
278 10, e0146021. doi:10.1371/journal.pone.0146021.
- 279 Sauer, J. (1957). Recent Migration and Evolution of the Dioecious Amaranths. *Evolution* 11, 11–31.
280 doi:10.2307/2405808.
- 281 Sauer, J. D. (1972). The dioecious amaranths: A new species name and major range extensions. *Madroño*
282 21, 426–434. Available at: <http://www.jstor.org/stable/41423815>.
- 283 Schwartz, L. M., Norsworthy, J. K., Young, B. G., Bradley, K. W., Kruger, G. R., Davis, V. M., Steckel,
284 L. E., and Walsh, M. J. (2016). Tall Waterhemp (Amaranthus tuberculatus) and Palmer amaranth
285 (Amaranthus palmeri) Seed Production and Retention at Soybean Maturity. *Weed Technology* 30,
286 284–290. doi:10.1614/WT-D-15-00130.1.
- 287 Scott, D. (2011). The Technological Fix Criticisms and the Agricultural Biotechnology Debate. *J Agric
288 Environ Ethics* 24, 207–226. doi:10.1007/s10806-010-9253-7.
- 289 Sosnoskie, L. M., Webster, T. M., and Culpepper, A. S. (2013). Glyphosate Resistance Does Not
290 Affect Palmer Amaranth (Amaranthus palmeri) Seedbank Longevity. *Weed Science* 61, 283–288.
291 doi:10.1614/WS-D-12-00111.1.
- 292 Spaunhorst, D. J., Devkota, P., Johnson, W. G., Smeda, R. J., Meyer, C. J., and Norsworthy, J. K. (2018).
293 Phenology of Five Palmer amaranth (Amaranthus palmeri) Populations Grown in Northern Indiana and
294 Arkansas. *Weed Science* 66, 457–469. doi:10.1017/wsc.2018.12.
- 295 Wang, J. L., Klessig, D. F., and Berry, J. O. (1992). Regulation of C4 Gene Expression in Developing
296 Amaranth Leaves. *The Plant Cell* 4, 173–184. doi:10.1105/tpc.4.2.173.

- 297 Ward, S. M., Webster, T. M., and Steckel, L. E. (2013). Palmer Amaranth (*Amaranthus palmeri*): A Review.
298 *Weed Technology* 27, 12–27. doi:10.1614/WT-D-12-00113.1.
- 299 Yu, E., Blair, S., Hardel, M., Chandler, M., Thiede, D., Cortilet, A., Gunsolus, J., and Becker, R. (2021).
300 Timeline of Palmer amaranth (*Amaranthus palmeri*) invasion and eradication in Minnesota. *Weed
301 Technology*, 1–31. doi:10.1017/wet.2021.32.