## Data Analysis in R

## Analysis of Variance (ANOVA)

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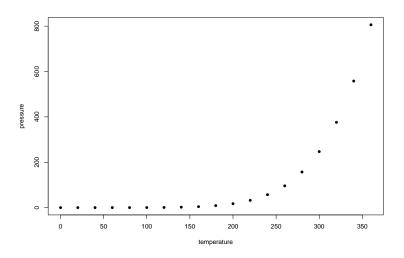
## Check your data

 Quantitative data is information about quantities; that is, information that can be measured and written down with numbers. Example:

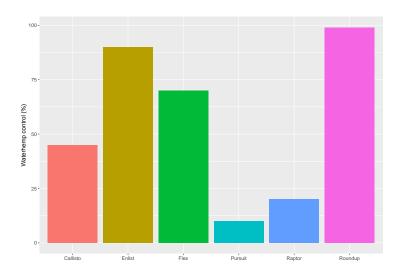
x-axis = 1.5, 5.6, 12.8, 26.7. 39.4, 45.1

- Proceed with regression
- Numeric or integer
- **Qualitative** data is information about qualities; information that can't actually be measured. *Example*: x-axis = Pursuit, Roundup, Callisto
  - Proceed with ANOVA
  - Factor

## Quantitative data



## Qualitative data



#### The ANOVA Test

- A way to find if experiment results are significant. It helps you to figure out if you need to reject the null hypothesis or accept the alternate hypothesis
- Test groups to see if there's a difference between them
  - A group of weed scientists are trying different herbicides for Palmer amaranth control. You want to see if one herbicide is better than others
  - A company has 20 industrial hemp varieties from outside United States.
     They want to know which ones performed better in Wisconsin
  - Students from different colleges take the same exam. You want to see if one college outperforms the other

## Assumptions of ANOVA

- Independence
- Normality
- Homogeneity of variances (aka, Homoscedasticity)

#### Parametric and Non-Parametric Tests

- Parametric Tests: Relies on theoretical distributions of the test statistic under the null hypothesis and assumptions about the distribution of the sample data (i.e., normality)
- Non-Parametric Tests: Referred to as "Distribution Free" as they do not assume that data are drawn from any particular distribution

## Case of study

library(lattice)

• Using the data barley from package lattice

```
## yield variety year site
## 1 41.33333 Velvet 1931 Crookston
## 2 25.23333 Peatland 1932 Crookston
## 3 36.56666 Trebi 1931 University Farm
## 4 26.16667 Glabron 1932 Crookston
## 5 48.86667 Manchuria 1931 Waseca
```

- A data frame with 120 observations on the following 4 variables
  - Yield (averaged across three blocks) in bushels/acre.
  - Variety (factor) with 10 levels "Svansota", "No. 462", "Manchuria", "No. 475", "Velvet", "Peatland", "Glabron", "No. 457", "Wisconsin No. 38", "Trebi".
  - Year (factor) with 2 levels 1931 and 1932
  - Site (factor) with 6 levels: "Grand Rapids", "Duluth", "University Farm", "Morris", "Crookston", "Waseca"

## Homogeneity of variances

Bartlett test

```
##
## Bartlett test of homogeneity of variances
##
## data: yield by variety
## Bartlett's K-squared = 7.9816, df = 9, p-value = 0.536

• Levene test
leveneTest(barley$yield, barley$variety)

## Levene's Test for Homogeneity of Variance (center = median)
## Df F value Pr(>F)
```

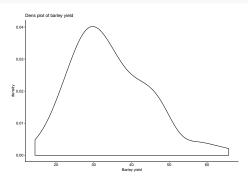
110

##

## group 9 0.747 0.6652

## Normality

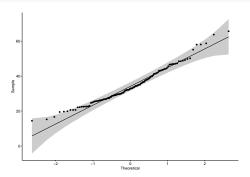
 Density plot: the density plot provides a visual judgment about whether the distribution is bell shaped.



## Q-Q plot

 Q-Q plot: Q-Q plot (or quantile-quantile plot) draws the correlation between a given sample and the normal distribution. A 45-degree reference line is also plotted.

#### ggqqplot(barley\$yield)



## Normality test

```
library(nortest)
pearson.test(barley$yield)

##

## Pearson chi-square normality test
##

## data: barley$yield
## P = 16.5, p-value = 0.1236
```

- ullet From the output, the p-value > 0.05 implying that the distribution of the data are not significantly different from normal distribution. In other words, we can assume the normality.
- Other tests of normality (Shapiro-Wilk)

#### Data transformation

- Assuming data (barley yield) is non-gaussean (non-normal)
- Load the package bestNormalize

```
#install.packages("bestNormalize") # if needed
library(bestNormalize)
```

#### Package 'bestNormalize'

August 20, 2019

Type Package

Title Normalizing Transformation Functions

Version 142

Date 2019-08-20

Description Estimate a suite of normalizing transformations, including a new adaptation of a technique based on ranks which can guarantee normally distributed transformed data if there are no ties: ordered quantile normalization (ORQ). ORQ normalization combines a rank-mapping approach with a shifted logit approximation that allows the transformation to work on data outside the original domain. It is also able to handle new data within the original domain via linear interpolation. The package is built to estimate the best normalizing transformation for a vector consistently and accurately. It implements the Box-Cox transformation, the Yeo-Johnson transformation, there types of Lambert WxF transformations, and the ordered quantile normalization transformation. It also estimates the normalization efficacy of other commonly used transformations.

URL https://github.com/petersonR/bestNormalize

### Transforming the data

•

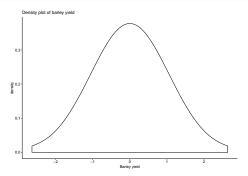
```
data <- bestNormalize(barley$yield, loo = TRUE)</pre>
```

data

```
Best Normalizing transformation with 120 Observations
##
   Estimated Normality Statistics (Pearson P / df, lower => r
   - No transform: 1.5
##
## - Box-Cox: 0.4818
## - Log b(x+a): 0.4818
## - sqrt(x+a): 0.9909
## - \exp(x): 136.9621
## - arcsinh(x): 0.4818
## - Yeo-Johnson: 0.4818
## - orderNorm: 0.1212
## Estimation method: Out-of-sample via leave-one-out CV
```

## Visual plots of transformed yield date

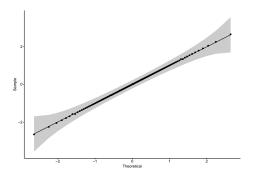
 Density plot: the density plot provides a visual judgment about whether the distribution is bell shaped (transformed data)



## Q-Q plot of transformed yield date

 Q-Q plot: Q-Q plot (or quantile-quantile plot) draws the correlation between a given sample and the normal distribution. A 45-degree reference line is also plotted

#### ggqqplot(data\$x.t)



## Normality test after data transformation

```
##
## Pearson chi-square normality test
##
## data: data$x.t
```

ullet From the output, the p-value > 0.05 implying that the distribution of the data are not significantly different from normal distribution. In other words, we can assume the normality.

pearson.test(data\$x.t)

## P = 0.4, p-value = 1

#### Post-Hoc ANOVA

Use package: Ime4

#### Package 'lme4'

March 5, 2019

Version 1.1-21

Title Linear Mixed-Effects Models using 'Eigen' and S4

Contact LME4 Authors <lme4-authors@lists.r-forge.r-project.org>

**Description** Fit linear and generalized linear mixed-effects models.

The models and their components are represented using S4 classes and methods. The core computational algorithms are implemented using the 'Eigen' C++ library for numerical linear algebra and 'RcppEigen' ``glue''.

Depends R (>= 3.2.0), Matrix (>= 1.2-1), methods, stats

LinkingTo Rcpp (>= 0.10.5), RcppEigen

**Imports** graphics, grid, splines, utils, parallel, MASS, lattice, boot, nlme (>= 3.1-123), minqa (>= 1.1.15), nloptr (>= 1.0.4)

Suggests knitr, rmarkdown, PKPDmodels, MEMSS, testthat (>= 0.8.1), ggplot2, mlmRev, optimx (>= 2013.8.6), gamm4, pbkrtest, HSAUR2, numDeriv, car, dfoptim

VignetteBuilder knitr

LazyData yes

License GPL (>= 2)

URL https://github.com/lme4/lme4/

#### Post-Hoc ANOVA

#### **Packages**

```
library(lme4) # model
## Loading required package: Matrix
##
## Attaching package: 'Matrix'
## The following objects are masked from 'package:tidyr':
##
##
       expand, pack, unpack
## Registered S3 methods overwritten by 'lme4':
##
     method
                                      from
    cooks.distance.influence.merMod.car
##
    influence.merMod
##
                                      car
##
    dfbeta.influence.merMod
                                      car
     dfbetas.influence.merMod
##
                                      car
library(emmeans) # anova
library(lmerTest) # lsmeans
```

##

#### Model

• Mixed model:

Fixed: Variety

Random: Year

fit <- lmer(yield ~ variety \* site + (1|year), data=barley)</pre>

## Summary

summary(fit)

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method [
## lmerModLmerTest]
## Formula: yield ~ variety * site + (1 | year)
##
     Data: barlev
##
## REML criterion at convergence: 449.8
##
## Scaled residuals:
##
     Min 10 Median 30 Max
## -1.877 -0.491 0.000 0.491 1.877
##
## Random effects:
   Groups Name
##
                    Variance Std.Dev.
## year (Intercept) 13.29 3.645
   Residual
                       50.35 7.096
## Number of obs: 120, groups: year, 2
##
## Fixed effects:
                                            Estimate Std. Error df
##
## (Intercept)
                                             23.1500 5.6406 16.7881
## varietyNo. 462
                                             -0.7333 7.0955 59.0002
```

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#### **ANOVA**

#### anova(fit)

```
## Type III Analysis of Variance Table with Satterthwaite's method
## Sum Sq Mean Sq NumDF DenDF F value Pr(>F)

## variety 1052.6 116.95 9 59 2.3230 0.02592 *

## site 6633.9 1326.77 5 59 26.3529 6.95e-14 ***

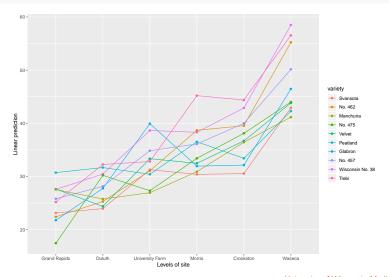
## variety:site 1205.8 26.79 45 59 0.5322 0.98544

## ---

## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

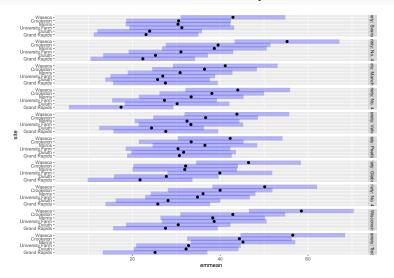
#### Visual interaction

emmip(fit, variety ~ site | site) # include CIs = TRUE



Data Analysis in R

#### plot(emmeans(fit, ~ site | site\*variety))



#### LS Means

#### Looking the site level (Factor)

```
lssite<-emmeans(fit, ~site, contr="pairwise", adjust="none", type="response")
## NOTE: Results may be misleading due to involvement in interactions
lssite$emmeans</pre>
```

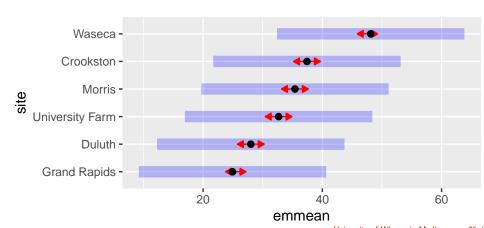
```
site
                         SE
##
                 emmean
                             df lower.CL upper.CL
##
   Grand Rapids 24.9 3.03 1.68
                                   9.21
                                           40.6
   Dulluth
                                           43.7
##
                 28.0 3.03 1.68 12.28
   University Farm 32.7 3.03 1.68 16.95 48.4
##
   Morris
               35.4 3.03 1.68 19.68 51.1
##
   Crookston
                37.4 3.03 1.68 21.70
                                           53.1
##
##
   Waseca
                  48.1 3.03 1.68 32.39
                                           63.8
##
```

- ## Results are averaged over the levels of: variety
- ## Degrees-of-freedom method: kenward-roger
- ## Confidence level used: 0.95

#### Plot

• Plotting the site *level* (Factor)

```
plot(lssite, comparisons =TRUE, adjust="none")
```



#### LS Means

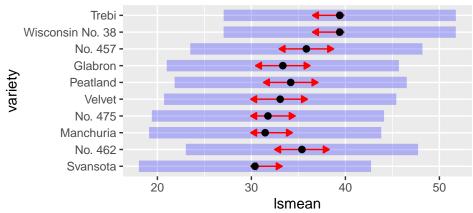
Looking the variety level (Factor)

```
lsvar<-lsmeans(fit, ~variety, contr="pairwise", adjust="none", type="response")</pre>
## NOTE: Results may be misleading due to involvement in interactions
lsvar$lsmeans
##
   variety
                  lsmean
                          SE df lower.CL upper.CL
##
   Svansota
                    30.4 3.29 2.34
                                     18.0 42.7
##
   No. 462
                    35.4 3.29 2.34 23.0 47.7
##
   Manchuria
                   31.5 3.29 2.34 19.1 43.8
   No. 475
                   31.8 3.29 2.34 19.4 44.1
##
##
   Velvet
                   33.1 3.29 2.34 20.7 45.4
   Peatland
                   34.2 3.29 2.34 21.8 46.5
##
            33.3 3.29 2.34 21.0 45.7
##
   Glabron
                   35.8 3.29 2.34 23.5 48.2
## No. 457
   Wisconsin No. 38
                   39.4 3.29 2.34 27.0 51.7
##
##
                    39.4 3.29 2.34 27.0 51.8
   Trebi
##
## Results are averaged over the levels of: site
## Degrees-of-freedom method: kenward-roger
## Confidence level used: 0.95
```

#### LS Means

• Plotting the **variety** level (factor)

```
plot(lsvar, comparisons =TRUE, adjust="none")
```



#### Non-Parametric Tests

- Case of study: Percentage data (0 to 100%)
  - Weed Control
  - Disease
  - Insect damage

#### Non-Parametric Tests

Cover crop management (Kolby's study)

```
sample_n(Data, size=5)
```

```
## # A tibble: 5 x 5
                                                                  control
     trt
                                              trtn
                                                      rep crop
     <chr>>
                                              <fct> <dbl> <chr>
                                                                    <dbl>
## 1 No-till + POST
                                              3
                                                        4 Soybean 0.9
                                                        3 Soybean 0.99
   2 Cereal Rye terminated at plant + POST
  3 \text{ No-till + POST}
                                                        2 Soybean 0.7
## 4 Cerealy Rye forage harvest + PRE fb POST 11
                                                        3 Soybean
                                                                    0.99
## 5 Cereal Rye terminated at plant + POST
                                                        1 Soybean
                                                                     0.9
```

Values must be between 0 and 1

# Generalized Linear Mixed Models using Template Model Builder

- Fit linear and generalized linear mixed models with various extensions, including zero-inflation. The models are fitted using maximum likelihood estimation via 'TMB' (Template Model Builder)
- Random effects are assumed to be Gaussian on the scale of the linear predictor and are integrated out using the Laplace approximation.
   Gradients are calculated using automatic differentiation

## Homogeneity of variances

Bartlett test

```
bartlett.test(control ~ trtn, data=Data)
##
##
    Bartlett test of homogeneity of variances
##
## data: control by trtn
## Bartlett's K-squared = Inf, df = 10, p-value < 2.2e-16

    levene test

leveneTest(Data$control, Data$trtn)
## Levene's Test for Homogeneity of Variance (center = median)
       Df F value Pr(>F)
##
## group 10 1.8176 0.09158 .
```

## Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.05 '.' 0.1 ' ' 1

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#### Model

```
library(glmmTMB)
model <- glmmTMB(control ~ trtn + (1|rep), data=Data, beta_family(link = "logit"))</pre>
```

## Summary

```
Sum <- summarv(model)</pre>
Sum$coefficients
## $cond
##
                Estimate Std. Error
                                      z value
                                                   Pr(>|z|)
## (Intercept) -0.8771015  0.3208830 -2.733399  6.268431e-03
## trtn3
                2.5672054 0.4223463 6.078437 1.213594e-09
## trtn4
                3.4124799 0.5599591 6.094159 1.100145e-09
## trtn5
               4.0050302
                          0.5967980 6.710864 1.934760e-11
               4.0050179
                          0.5967956 6.710870 1.934668e-11
## trtn6
## trtn7
               3.3517779 0.5556180 6.032522 1.614203e-09
## trtn8
                1.9697344
                           0.4639821
                                      4.245281 2.183199e-05
               3.6705235
                          0.5774255 6.356704 2.061281e-10
## t.rt.n9
## trtn10
               3.4741377 0.5642906 6.156646 7.430164e-10
                          0.5879081 6.535864 6.324327e-11
## trtn11
               3.8424874
## trtn12
               4.0050179 0.5967956 6.710870 1.934668e-11
##
## $zi
## NULL
##
## $disp
## NULL
```

#### **ANOVA**

We should use Anova.glmmTBM

```
Anova(model, test.statistic = "Chisq", type = "II")

## Analysis of Deviance Table (Type II Wald chisquare tests)

##

## Response: control

## Chisq Df Pr(>Chisq)

## trtn 97.126 10 < 2.2e-16 ***

## ---

## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1</pre>
```

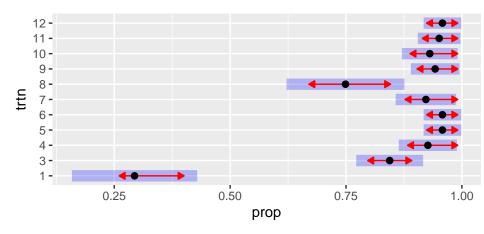
#### LS Means

```
lsm=emmeans(model , ~ trtn, contr="pairwise", adjust="none", type = "response")
lsm
## $emmeans
```

```
trtn prop SE df lower.CL upper.CL
##
##
       0.294 0.0666 35
                        0.178
                                 0.444
     0.844 0.0355 35 0.758 0.904
##
##
     0.927 0.0309 35 0.834 0.969
##
     0.958 0.0200 35 0.893 0.984
## 6
     0.958 0.0200 35 0.893 0.984
##
     0.922 0.0321 35
                        0.827
                                0.967
##
     0.749 0.0627 35
                        0.603
                                0.854
                        0.862 0.977
##
     0.942 0.0258 35
##
   10
       0.931 0.0296 35
                      0.841 0.971
##
   11
       0.951 0.0227 35 0.878
                                0.981
##
  12 0.958 0.0200 35 0.893
                                0.984
##
## Confidence level used: 0.95
## Intervals are back-transformed from the logit scale
##
## $contrasts
##
   contrast odds.ratio
                        SE df t.ratio p.value
##
   1 / 3
               0.0767 0.0324 35 -6.078 <.0001
              0 0330 0 0185 35 -6 094
                                     < 0001
```

#### Plot

```
plot(lsm, comparisons =TRUE, adjust="none")
```



```
CLD(lsm, alpha=0.05, Letters=letters, adjust="none", reversed = TRUE)

## Warning: 'CLD' will be deprecated. Its use is discouraged.

## See '? CLD' for an explanation. Use 'pwpp' or 'multcomp::cld' instead.

## Warning in CLD.emm_list(lsm, alpha = 0.05, Letters = letters, adjust =

## "none", : `CLD()` called with a list of 2 objects. Only the first one was

## used.

## Warning: 'CLD' will be deprecated. Its use is discouraged.

## See '? CLD' for an explanation. Use 'pwpp' or 'multcomp::cld' instead.

## trtn prop SE df lower.CL upper.CL .group
```

```
##
  5 0.958 0.0200 35 0.893 0.984 a
## 12 0.958 0.0200 35 0.893 0.984 a
## 6 0.958 0.0200 35 0.893 0.984 a
  11 0.951 0.0227 35 0.878 0.981 a
##
  9 0.942 0.0258 35 0.862 0.977 a
##
## 10 0.931 0.0296 35 0.841 0.971 ab
## 4 0.927 0.0309 35 0.834 0.969 ab
## 7 0.922 0.0321 35 0.827 0.967 ab
## 3 0.844 0.0355 35 0.758 0.904 bc
## 8 0.749 0.0627 35 0.603 0.854 c
## 1 0.294 0.0666 35 0.178 0.444
                                  d
##
## Confidence level used: 0.95
```

## **Plotting**

