fMRI_Analysis

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Import data & processing

```
# Change file extension
# old_names <- list.files(path)</pre>
# new_names <- gsub(".1D", "1D.csv", old_names)</pre>
# file.rename(pasteO(path, old_names), pasteO(path, new_names))
# Import data
pheno <- read_csv("./data/phenotypic_CMU.csv") %>% janitor::clean_names()
df_file_names <-</pre>
  inner_join(data.frame(name = all_files,
                        sub id = as.numeric(substring(all files, 7, 13))),
             pheno, by = "sub id")
# Files for each group
asd_files <- df_file_names %>% filter(dx_group == 1) %>%
  select(1) %>% mutate(name = paste(path, name, sep = ""))
tc files <- df file names %>% filter(dx group == 2) %>%
  select(1) %>% mutate(name = paste(path, name, sep = ""))
roi <- read_csv("./data/dos160_labels.csv")</pre>
colnames(roi) <- c("number", "label")</pre>
# from paper: prediction of individual brain maturity using fmri
roi_network <- data.frame(</pre>
  network = c(rep("default", 34), rep("fronto_parietal", 21), rep("cingulo_opercular", 32),
              rep("sensorimotor", 33), rep("occipital", 22), rep("cerebellum", 18)),
  number = c(1, 4, 5, 6, 7, 11, 13, 14, 15, 17, 20, 63, 72, 73, 84, 85,
             90, 91, 92, 93, 94, 105, 108, 111, 112, 115, 117, 124, 132,
             134, 136, 137, 141, 146, # 1
             2, 3, 9, 10, 12, 16, 21, 22, 23, 24, 29, 34, 36, 88, 96, 99,
             101, 104, 107, 114, 116, # 2
             8, 18, 19, 25, 26, 27, 28, 30, 31, 33, 38, 39, 40, 44, 47, 57,
             58, 59, 61, 71, 76, 78, 80, 81, 87, 89, 95, 97, 100, 102, 103, 125, # 3
             32, 35, 37, 41, 42, 43, 45, 46, 48, 49, 50, 51, 52, 53, 54, 55, 56, 60,
             62, 64, 65, 66, 67, 68, 69, 70, 74, 75, 77, 79, 82, 83, 86, # 4
             106, 118, 119, 123, 126, 129, 133, 135, 139, 142, 145, 147, 148, 149,
             152, 153, 154, 156, 157, 158, 159, 160, # 5
             98, 109, 110, 113, 120, 121, 122, 127, 128, 130, 131, 138, 140,
             143, 144, 150, 151, 155 # 6
             ))
```

```
roi_comb <- inner_join(roi, roi_network, by = "number")</pre>
```

EDA

Table 1: Demographics for People with ASD and TC

| Characteristic | $\mathbf{ASD},N=14$ | Typical Controls, $N = 13$ | p-value |
|--|---------------------|----------------------------|---------|
| Age, yrs | 25.5 (21.2, 30.8) | 27.0 (21.0, 30.0) | 0.8 |
| Sex | | | > 0.9 |
| Male | 11 (79%) | 10 (77%) | |
| Female | 3 (21%) | 3 (23%) | |
| Handedness | , , | , | 0.6 |
| Ambi | 1(7.1%) | 1 (7.7%) | |
| L | 1(7.1%) | 0(0%) | |
| R | 12 (86%) | 12(92%) | |
| Full-Scale Intelligence Quotient (FIQ) | 117 (105, 123) | 110 (109, 124) | > 0.9 |
| Verbal Intelligence Quotient (VIQ) | 111 (99, 120) | 111 (108, 122) | 0.7 |
| Performance Intelligence Quotient | 115 (108, 121) | 109 (108, 123) | 0.7 |
| (PIQ) | , , , | | |

Parameter tuning

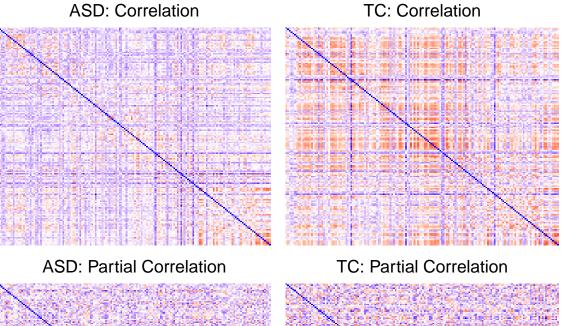
```
#
                  p = ncol(file_sel), verbose = FALSE)
#
#
   pc_final \leftarrow ifelse(pc_res@pMax < x, 1, 0)
   g <- graph.adjacency(pc_final, mode = "undirected")
#
   len \leftarrow length(E(q))
#
    data.frame(alpha = x, num\_edge = len)
# }
# saveRDS(pc_alpha, file = "./data/pc_alpha.rds")
pc_alpha <- readRDS("./data/pc_alpha.rds")</pre>
pc_alpha_res <- do.call(rbind, pc_alpha)</pre>
fig_1 <- pc_alpha_res %>%
  ggplot(aes(x = log(alpha), y = num_edge)) +
  geom_point() + geom_line() + theme_bw() +
  labs(x = "log(alpha)", y = "Number",
       title = "Number of Undirected Edges by alpha in PC")
sel_alpha_pc <- pc_alpha_res$alpha[9]</pre>
# --- MMHC ---
\# mmhc_alpha_grid <- exp(seq(-3, -6, length = 20)) \# Try similar number of eqdes
# mmhc alpha <- foreach(x = mmhc alpha grid) %do% {
   mmhc_res <- pchc::mmhc(as.matrix(file_sel), alpha = x)</pre>
#
#
# mmhc_final <- ifelse(1 - exp(mmhc_res$ini$pvalue) < x, 1, 0)</pre>
# g <- graph.adjacency(mmhc_final, mode = "undirected")</pre>
# len \leftarrow length(E(g))
#
#
   data.frame(alpha = x, num_edge = len)
# saveRDS(mmhc_alpha, file = "./data/mmhc_alpha.rds")
mmhc_alpha <- readRDS("./data/mmhc_alpha.rds")</pre>
mmhc_alpha_res <- do.call(rbind, mmhc_alpha)</pre>
fig_2 <- mmhc_alpha_res %>%
  ggplot(aes(x = log(alpha), y = num_edge)) +
  geom_point() + geom_line() + theme_bw() +
  labs(x = "log(alpha)", y = "Number",
       title = "Number of Undirected Edges by alpha in MMHC")
sel_alpha_mmhc <- mmhc_alpha_res$alpha[12]</pre>
```

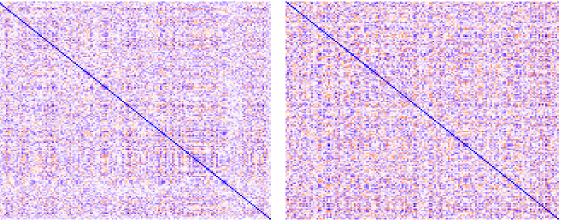
Individual-level analysis

```
# Normalize for 0 to 1
normalize <- function(x){
  t_x <- (x - min(x))/(max(x) - min(x))
  return(t_x)}

indepTest <- gaussCItest # conditional independence test
# --- ASD ---</pre>
```

```
# Randomly select one ASD sample
set.seed(123)
asd dat <- fread(sample(as.matrix(asd files), 1), select = c(1:160))
suffStat_asd <- list(C = cor(asd_dat), n = nrow(asd_dat))</pre>
# Correlation
asd_corr <- cor(asd_dat)</pre>
t_asd_corr <- normalize(asd_corr)</pre>
diag(t_asd_corr) <- 0</pre>
# Partial correlation
asd_pcorr <- ppcor::pcor(asd_dat)</pre>
t_asd_pcorr <- normalize(asd_pcorr$estimate)</pre>
diag(t_asd_pcorr) <- 0</pre>
# --- TC ---
# Randomly select one TC sample
set.seed(123)
tc_sel <- sample(as.matrix(tc_files), 1)</pre>
tc_dat <- fread(tc_sel, select = c(1:160))</pre>
suffStat_tc <- list(C = cor(tc_dat), n = nrow(tc_dat))</pre>
# Correlation
tc_corr <- cor(tc_dat)</pre>
t_tc_corr <- normalize(tc_corr)</pre>
diag(t_tc_corr) <- 0</pre>
# Partial Correlation
tc_pcorr <- ppcor::pcor(tc_dat)</pre>
t_tc_pcorr <- normalize(tc_pcorr$estimate)</pre>
diag(t_tc_pcorr) <- 0</pre>
# --- Make plots ---
col_fun <- colorRamp2(c(0, 0.5, 1), c("blue", "white", "red"))</pre>
grid.newpage()
pushViewport(viewport(layout = grid.layout(nr = 3, nc = 2)))
# ASD
pushViewport(viewport(layout.pos.row = 1, layout.pos.col = 1))
draw(Heatmap(t_asd_corr, col = col_fun, cluster_rows = F, cluster_columns = F,
             show_row_names = F, show_column_names = F,
              show_heatmap_legend = F, column_title = "ASD: Correlation"), newpage = F)
upViewport()
pushViewport(viewport(layout.pos.row = 2, layout.pos.col = 1))
draw(Heatmap(t_asd_pcorr, col = col_fun, cluster_rows = F, cluster_columns = F,
             show_row_names = F, show_column_names = F,
             show_heatmap_legend = F, column_title = "ASD: Partial Correlation"), newpage = F)
upViewport()
# TC
pushViewport(viewport(layout.pos.row = 1, layout.pos.col = 2))
draw(Heatmap(t_tc_corr, col = col_fun, cluster_rows = F, cluster_columns = F,
```



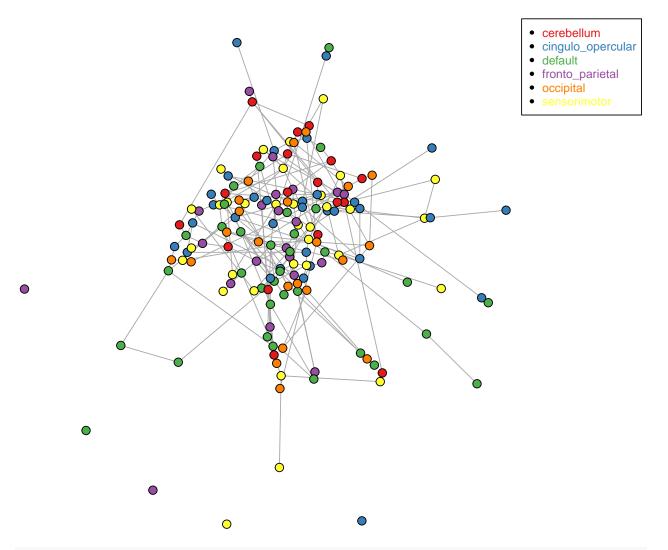




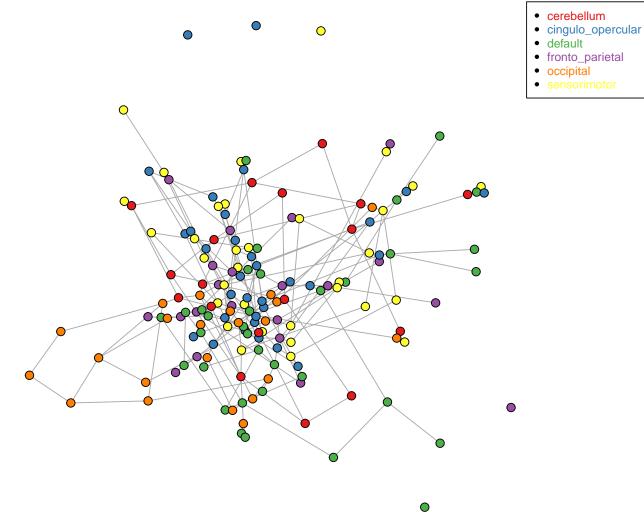
```
# Change weights of layout
weight_community <-</pre>
  function(row, membership, weight_within = 0.5, weight_between = 100) {
    if (membership$network[which(membership$number == row[1])] ==
      membership$network[which(membership$number == row[2])])
    {
      weight = weight_within
    else {
      weight = weight_between
    return(weight)
 }
# Colors for 6 networks
my_color <- RColorBrewer::brewer.pal(6, "Set1")</pre>
# PC plot for ASD vs. TD
# Groups
df_grp <- roi_comb %>%
 select(1, 3)
# --- ASD ---
# PC
asd_pc <- pc(suffStat_asd, indepTest, alpha = sel_alpha_pc,</pre>
             p = ncol(asd_dat), verbose = FALSE)
asd_res <- ifelse(asd_pc@pMax < sel_alpha_pc, 1, 0)</pre>
g_asd <- graph.adjacency(asd_res, mode = "undirected") # convert to igraph</pre>
el_asd <- get.edgelist(g_asd) # get edgelist</pre>
layout_wgt <- apply(el_asd, 1, weight_community, df_grp)</pre>
g_asd$layout <- layout_with_fr(g_asd, weights = layout_wgt)</pre>
# make plot
plot(g_asd, vertex.size = 3.5, vertex.label = NA,
     vertex.color = my_color[as.factor(df_grp$network)],
     main = "Topological Structure of ASD from PC")
legend("topright", legend = levels(as.factor(df_grp$network)),
      pch = 20, text.col = my_color, cex = 0.8)
```

Topological structures from PC

Topological Structure of ASD from PC



Topological Structure of TC from PC



Group-level prediction

```
pc_res <- pc(suffStat, indepTest, alpha = sel_alpha_pc,</pre>
                p = ncol(dat), verbose = FALSE)
  pc_final <- ifelse(pc_res@pMax < sel_alpha_pc, 1, 0)</pre>
  g <- graph.adjacency(pc_final, mode = "undirected")</pre>
  # measurements
  # distance class
  ecc <- mean(eccentricity(g)) # eccentricity</pre>
  # rad <- radius(g) # radius</pre>
  # gir <- girth(g, circle = FALSE) # girth
  # connection class
  ass <- assortativity_degree(g, directed = FALSE)</pre>
  tra <- transitivity(g, type = "global")</pre>
  # community class
  mod <- modularity(g, as.factor(df_grp$network))</pre>
  m_dist <- mean_distance(g, directed = FALSE) # mean distance</pre>
  # assor <- assortativity_degree(g, directed = FALSE) # assortivity degree
  # den <- edge_density(g, loops = FALSE) # density
  # dia <- diameter(g, directed = FALSE) # diameter
  data.frame(
    id = df file names$sub id[x],
    eccentricity = ecc, assortativity = ass,
    transitivity = tra, modularity = mod, mean_distance = m_dist,
    asd = ifelse(df_file_names$dx_group[x] == 1, 1, 0))}
pc_res_all_2 <- do.call(rbind, pc_res_all) %>%
 mutate(asd = as.factor(ifelse(asd == 1, "asd", "no_asd")))
# Split train vs. Test
set.seed(1234)
tr_id <- createDataPartition(y = pc_res_all_2$asd, p = 0.75, list = FALSE)</pre>
tr_dat_pc <- pc_res_all_2[tr_id, -1]</pre>
ts_dat_pc <- pc_res_all_2[-tr_id, -1]
# qlm
set.seed(1234)
glm_fit_pc <- train(x = tr_dat_pc[, 1:5], y = tr_dat_pc$asd,</pre>
                    method = "glm", metric = "ROC",
                     trControl = ctrl)
pc_test_prob <- predict(glm_fit_pc, newdata = ts_dat_pc, type = "prob")[, 1]</pre>
pc_test_pred <- ifelse(pc_test_prob > 0.5, "asd", "no_asd")
confusionMatrix(data = as.factor(pc_test_pred),
                reference = ts_dat_pc$asd,
                 positive = "no_asd")
```

```
## Confusion Matrix and Statistics
##
## Reference
```

```
## Prediction asd no asd
##
       asd
                2
##
       no asd
##
##
                   Accuracy: 0.8333
##
                     95% CI: (0.3588, 0.9958)
##
       No Information Rate: 0.5
       P-Value [Acc > NIR] : 0.1094
##
##
##
                      Kappa: 0.6667
##
##
    Mcnemar's Test P-Value: 1.0000
##
               Sensitivity: 1.0000
##
##
               Specificity: 0.6667
##
            Pos Pred Value: 0.7500
##
            Neg Pred Value: 1.0000
##
                Prevalence: 0.5000
##
            Detection Rate: 0.5000
##
      Detection Prevalence: 0.6667
##
         Balanced Accuracy: 0.8333
##
##
          'Positive' Class : no_asd
##
# === MMHC ===
# Obtain graph metrics from MMHC
mmhc_res_all <- foreach(x = seq(nrow(df_file_names))) %do% {</pre>
  dat <- fread(paste(path, df_file_names$name[x], sep = ""), select = c(1:160))</pre>
 mmhc_res <- pchc::mmhc(as.matrix(dat), alpha = sel_alpha_mmhc)
  mmhc_final <- ifelse(exp(mmhc_res$ini$pvalue) < sel_alpha_mmhc, 1, 0)</pre>
  g <- graph.adjacency(mmhc_final, mode = "undirected")</pre>
  # measurements
  # distance class
  ecc <- mean(eccentricity(g)) # eccentricity</pre>
  # rad <- radius(g) # radius</pre>
  # gir <- girth(g, circle = FALSE) # girth
  # connection class
  ass <- assortativity_degree(g, directed = FALSE)</pre>
  tra <- transitivity(g, type = "global")</pre>
  # community class
  mod <- modularity(g, as.factor(df_grp$network))</pre>
 m_dist <- mean_distance(g, directed = FALSE) # mean distance</pre>
  # assor <- assortativity_degree(g, directed = FALSE) # assortivity degree
  # den <- edge_density(g, loops = FALSE) # density</pre>
  # dia <- diameter(g, directed = FALSE) # diameter
 data.frame(
   id = df file names$sub id[x],
```

```
eccentricity = ecc,
    assortativity = ass, transitivity = tra, modularity = mod,
    asd = ifelse(df_file_names$dx_group[x] == 1, 1, 0))}
mmhc_res_all_2 <- do.call(rbind, mmhc_res_all) %>%
  mutate(asd = as.factor(ifelse(asd == 1, "asd", "no_asd")))
# Split train vs. Test
set.seed(1234)
tr_id <- createDataPartition(y = mmhc_res_all_2$asd, p = 0.75, list = FALSE)
tr_dat_mmhc <- mmhc_res_all_2[tr_id, -1]</pre>
ts_dat_mmhc <- mmhc_res_all_2[-tr_id, -1]
# qlm
set.seed(1234)
glm_fit_mmhc <- train(x = tr_dat_mmhc[, 1:5], y = tr_dat_mmhc$asd,</pre>
                      method = "glm", metric = "ROC",
                      trControl = ctrl)
mmhc_test_prob <- predict(glm_fit_mmhc, newdata = ts_dat_mmhc, type = "prob")[, 1]</pre>
mmhc_test_pred <- ifelse(mmhc_test_prob > 0.5, "asd", "no_asd")
confusionMatrix(data = as.factor(mmhc_test_pred),
                reference = ts_dat_mmhc$asd,
                positive = "no_asd")
## Confusion Matrix and Statistics
##
             Reference
##
## Prediction asd no_asd
##
       asd
                3
##
       no_asd
              0
                       3
##
##
                  Accuracy : 1
                    95% CI: (0.5407, 1)
##
##
       No Information Rate: 0.5
##
       P-Value [Acc > NIR] : 0.01563
##
##
                     Kappa: 1
##
  Mcnemar's Test P-Value : NA
##
               Sensitivity: 1.0
##
##
               Specificity: 1.0
##
            Pos Pred Value : 1.0
            Neg Pred Value: 1.0
##
##
                Prevalence: 0.5
            Detection Rate: 0.5
##
##
      Detection Prevalence: 0.5
##
         Balanced Accuracy: 1.0
##
##
          'Positive' Class : no_asd
```

##

Others

```
# === PC ===
# ASD
run <- function(dat_files = asd_files, net = "default") {</pre>
  vars <- roi network %>%
    filter(network == net) %>% pull(number)
  len <- length(vars)</pre>
  all_pc <- mclapply(1:nrow(dat_files), function(i){</pre>
  dat <- fread(dat_files[i, ], select = vars)</pre>
  suffStat \leftarrow list(C = cor(dat), n = nrow(dat))
  pc_res <- pc(suffStat, indepTest, alpha = 0.05,</pre>
                p = ncol(dat), verbose = FALSE)
  pc_final <- ifelse(pc_res@pMax < 0.05, 1, 0)</pre>
  adj_mat <- graph.adjacency(pc_final, mode = "max")</pre>
  # number of edges
  num <- nrow(get.edgelist(adj_mat))/((len * (len - 1))/2)</pre>
  }, mc.cores = num.cores)
  return(do.call(rbind, all pc))
}
asd_default_pc <- run(asd_files, "cerebellum")</pre>
tc_default_pc <- run(tc_files, "cerebellum")</pre>
t.test(asd_default_pc, tc_default_pc)
asd_all_pc <- mclapply(1:nrow(asd_files), function(i){</pre>
  dat <- fread(asd_files[i, ], select = c(1:160))</pre>
  suffStat \leftarrow list(C = cor(dat), n = nrow(dat))
  pc_res <- pc(suffStat, indepTest, alpha = 0.1,</pre>
                p = ncol(dat), verbose = FALSE)
  pc_final <- ifelse(pc_res@pMax < 0.1, 1, 0)</pre>
  adj_mat <- graph.adjacency(pc_final, mode = "max")</pre>
  # number of edges
  num <- nrow(get.edgelist(adj_mat))</pre>
}, mc.cores = num.cores)
asd_res <- do.call(rbind, asd_all_pc)</pre>
tc_all_pc <- mclapply(1:nrow(tc_files), function(i){</pre>
 dat <- fread(tc_files[i, ], select = c(1:160))</pre>
  suffStat \leftarrow list(C = cor(dat), n = nrow(dat))
 pc_res <- pc(suffStat, indepTest, alpha = 0.1,</pre>
```

```
p = ncol(dat), verbose = FALSE)
  pc_final <- ifelse(pc_res@pMax < 0.1, 1, 0)</pre>
  adj_mat <- graph.adjacency(pc_final, mode = "max")</pre>
  # number of edges
 num <- nrow(get.edgelist(adj_mat))/(160 * 159/2)</pre>
}, mc.cores = num.cores)
tc_res <- do.call(rbind, tc_all_pc)</pre>
t.test(asd_res, tc_res)
# --- Try alpha = 0.05 ---
# ASD
asd_all_res_pc <- common_conn(asd_files, alpha = 0.05, prct = 0.5)
tc_all_res_pc <- common_conn(tc_files, alpha = 0.05, prct = 0.5)
# --- Different alpha ---
alpha_ls \leftarrow seq(0.01, 0.5, length = 10)
# ASD
asd_all_res_pc_alpha <-
  mclapply(alpha_ls, function(alpha){
    # find the number of connections for each alpha
    pc_res <- common_conn_pc(dat_files = asd_files, alpha = alpha, prct = 0.5)</pre>
}, mc.cores = num.cores)
# saveRDS(asd_all_res_pc_alpha, "./data/asd_all_pc_alpha.RData")
# TC
tc_all_res_pc_alpha <-
  mclapply(seq(0.01, 1, by = 0.01), function(alpha){}
    # find the number of connections for each alpha
    pc_res <- common_conn_pc(dat_files = tc_files, alpha = alpha, prct = 0.5)</pre>
}, mc.cores = num.cores)
 \# \ saveRDS(tc\_all\_res\_pc\_alpha, \ "./data/tc\_all\_pc\_alpha.RData") \\
# Comparison
num_conn_pc <- data.frame(</pre>
 asd = lapply(asd_all_res_pc_alpha, function(x) nrow(x)) %>% unlist(),
 tc = lapply(tc_all_res_pc_alpha, function(x) nrow(x)) %>% unlist()
)
```