

tssports : integrates tsRNA feature extraction, differential analysis, and extensive visualization in one

platforms all

<https://github.com/Xia-Youmei/tssports-master>

tssports can perform small RNA classification, differential gene analysis, principal component analysis (PCA), and visualize the related functions and mechanisms of tsRNA through pie chart, volcano chart, heat map, MA chart etc.

Installation

You can install the development version of *tssports* like so:

```
if (!requireNamespace("devtools", quietly = TRUE)) {  
  install.packages("devtools")  
}  
  
devtools::install_github('Xia-Youmei/tssports-master')
```

Alternatively, the latest version can be installed locally from Git-hub: <https://github.com/Xia-Youmei/tssports-master>, and use the R package for your own data.

```
if (!requireNamespace("remotes", quietly = TRUE)) {  
  install.packages("remotes")  
}  
  
remotes::install_local("file_address/tssports-master-main.zip", upgrade =  
F, dependencies = T)
```

Note: For most use cases it is not necessary to install the *tssports* package locally, If you have a bad Internet connection, you can choose this method, and because of the sample data is large, please be patient for a few minutes.

To start working with the package *tssports*, you can load it in the R environment with the following command.

```
library('tssports')
```

Example

There is an example downloaded from Gene Expression Omnibus under the accession code GSE144666, it has 6 data sets that are SRR11004011-13 (RNAs were sequenced with standard protocols), SRR11004020-22 (RNAs were treated with T4PNK, then AlkB before cDNA library construction), organism is *Mus musculus*, tissue is brain.

First, you need to determine the working directory located in the dataset address:

```
setwd("./examples")
```

sportsV1.1 processed and generate the output files into one folder, you can determine the working directory located in the dataset address, it will automatically processing those contain "-miR-|-mir-|-let-" keywords annotation commented out, and automatically add reads of same genes at the end of the file, the filename ended in "_output_collapse_miRNA.txt" is obtained.

```
collapse_mature_miRNA_reads()
```

This function will result in 3 unified files: 1.

sports_combined_sample_fragments_counts_matrix_all.txt; 2.

sports_combined_sample_fragments_counts_matrix_0.5.txt; 3.

sports_combined_sample_fragments_annotation.txt. It can also read the entire folder, automatically identify and process the files ending in "_miRNA.txt" in the folder, first it will collect and sort out all genes to form the first file contains gene sequence, gene sequence length, whether to match to the genome, annotated genes; After that, all the same gene sequences in the input file were sorted into one file, and the screening condition was set as the probability of each fragment not being zero in the sample was greater than 0.5 to form the second file. The third file is the original file that collates all the same gene sequences in the input file into one file without setting screening conditions and gives users sufficient follow-up custom analysis.

```
combine_read_counts()
```

1.sports_combined_sample_fragments_counts_matrix_all

Fragment	Length	Match_Genome	Annotation
----------	--------	--------------	------------

TCGCTGCGATCTATTGAAAGTCAGCCCTCGACACAAGGGTTTGT	44	Yes	28S-rRNA
--	----	-----	----------

TCACAGTGAACCGGTCTCTTTAA	23	NO	piRNA
-------------------------	----	----	-------

CGCGACCTCAGATCAGACGT	20	NO	28S-rRNA
----------------------	----	----	----------

TCGGATCCGTCTGAGCTTGGCTTT	24	NO	piRNA
--------------------------	----	----	-------

TCACAGTGAACCGGTCTCTTAA	22	NO	piRNA
------------------------	----	----	-------

TCTTTGGTTATCTAGCTGTATGTT	24	NO	piRNA
--------------------------	----	----	-------

GGCTGGTCCGAAGGTAGTGAGTTATCTCAATT	32	Yes	RYN1-YRNA
----------------------------------	----	-----	-----------

TCAGTCGGTCCTGAG	15	Yes	28S-rRNA
-----------------	----	-----	----------

TGGGCTGTAGTGCGCTATGC	20	Yes	misc_RNA
----------------------	----	-----	----------

CTGGGCTGTAGTGCGCTATGC	21	Yes	misc_RNA
-----------------------	----	-----	----------

CGCTGCGATCTATTGAAAGTCAGCCCTCGACACAAGGGTTTGT	43	Yes	28S-rRNA
---	----	-----	----------

CGCGACCTCAGATCAGAC	18	NO	28S-rRNA
--------------------	----	----	----------

CATTGATCATCGACACTTCGAACGCACTTGCGGCCCGGGT	41	NO	5.8S-rRNA
--	----	----	-----------

2.sports_combined_sample_fragments_counts_matrix_0.5.txt

Sequence	SRR11004011	SRR11004012	SRR11004013	SRR11004020	SRR11004021
----------	-------------	-------------	-------------	-------------	-------------

SRR11004022					
-------------	--	--	--	--	--

TCGCTGCGATCTATTGAAAGTCAGCCCTCGACACAAGGGTTTGT	269161	143901	82697	49357	
--	--------	--------	-------	-------	--

96480	132035				
-------	--------	--	--	--	--

TCACAGTGAACCGGTCTCTTTAA	204627	136474	72396	2679	5694	38834
-------------------------	--------	--------	-------	------	------	-------

CGCGACCTCAGATCAGACGT	93017	119623	66666	30488	33792	143835
----------------------	-------	--------	-------	-------	-------	--------

TCGGATCCGTCTGAGCTTGGCTTT	66940	50898	49442	25380	38967	73429
--------------------------	-------	-------	-------	-------	-------	-------

TCACAGTGAACCGGTCTCTTAA	45249	32876	14415	503	997	8267
------------------------	-------	-------	-------	-----	-----	------

TCTTTGGTTATCTAGCTGTATGTT	35142	28972	27240	1199	3243	9601
--------------------------	-------	-------	-------	------	------	------

GGCTGGTCCGAAGGTAGTGAGTTATCTCAATT	32900	20960	14730	52832	34279	
----------------------------------	-------	-------	-------	-------	-------	--

44606						
-------	--	--	--	--	--	--

mmu-miR-3960	0	0	0	12	8	2
--------------	---	---	---	----	---	---

mmu-miR-3473g	0	0	0	2	1	1
---------------	---	---	---	---	---	---

mmu-miR-6987-5p	0	0	0	2	3	1
-----------------	---	---	---	---	---	---

```
mmu-miR-1951    0    0    0    1    1    3

3.sports_combined_sample_fragments_annotation.txt
Sequence      SRR11004011 SRR11004012 SRR11004013 SRR11004020 SRR11004021
SRR11004022
TCGCTGCGATCTATTGAAAGTCAGCCCTCGACACAAGGGTTTGT      269161  143901  82697  49357
96480  132035
TCACAGTGAACCGGTCTCTTTAA 204627  136474  72396  2679  5694  38834
CGCGACCTCAGATCAGACGT   93017  119623  66666  30488  33792  143835
TCGGATCCGTCTGAGCTTGGCTTT   66940  50898  49442  25380  38967  73429
TCACAGTGAACCGGTCTCTTAA 45249  32876  14415  503 997 8267
TCTTTGGTTATCTAGCTGTATGTT   35142  28972  27240  1199  3243  9601
GGCTGGTCCGAAGGTAGTGAGTTATCTCAATT   32900  20960  14730  52832  34279
44606
mmu-miR-511-5p  0    0    0    0    0    1
mmu-miR-505-3p  0    0    0    0    0    1
mmu-miR-7074-5p 0    0    0    0    0    1
mmu-miR-7229-5p 0    0    0    0    0    1
mmu-miR-6984-3p 0    0    0    0    0    1
```

This function requires the user to input the set of the last two digits in the SRR filename of the experimental group to distinguish the experimental group from the control group. For example, 20:22 is required for difference analysis in the example data. This function will automatically recognize the sports_combined_sample_fragments_counts_matrix_0.5.txt file output from the previous function. After processing, you will get 4 files, 1. Match all fragment annotations to genes, and get the file sports_counts_all.txt; 2. Match all the annotations to genes, and processed with the DESeq2 R package to obtain the differentially expressed genes, and obtain the file sports_deg_fDR005_2fc_all.txt. 3. All fragments were annotated as genes, and the differentially expressed genes were obtained after processing with DESeq2, and the reads were normalized to cpm (Counts per million) value to obtain the file sports_cpm_fdr005_2fc_all.txt. 4. Save all the differential genes calculated by DESeq2, without distinguishing log2FoldChange and padj, and get the file sports_DEG_all.txt;

```
getdegs(20:22)

1.sports_counts_all.txt
      SRR11004011 SRR11004012 SRR11004013 SRR11004020 SRR11004021 SRR11004022
28S-rRNA   269161  143901  82697  49357  96480  132035
piRNA      204627  136474  72396  2679   5694   38834
RNY1-YRNA   32900  20960  14730  52832  34279  44606
misc_RNA    28044  35480  18645  13025  21257  15968
5.8S-rRNA   22734  13872  2363  21703  16378  11993
45S-rRNA    12263  12825  15348  12838  27474  53615
mature-tRNA-Gly-GCC_5_end;mature-tRNA-Gly-CCC_5_end 10286  8054  6349
15440  24174  38846
5S-rRNA     8157  19663  8814  4571  8266  13170

2.sports_deg_fDR005_2fc_all.txt
ID baseMean log2FoldChange lfcSE stat pvalue padj
mature-tRNA-His-GTG 30035.3716708793 9.18307904145374 0.580202283242111
15.8273748771543 2.01452256749788e-56 2.96940626449187e-53
pre-tRNA-Leu-CAA 21805.5636328603 9.85974407854832 0.658354292658678
14.9763496471346 1.04818154091913e-50 7.72509795657397e-48
mature-tRNA-Arg-CCT;mature-tRNA-Arg-CCG 468.48506156553 7.40567460160257
0.655365046544287 11.3000756458593 1.31105017035912e-29
6.44162650369779e-27
```

```

mature-tRNA-Arg-ACG_CCA_end 43397.7312599756 7.76515581481599
0.692681783635114 11.2102786564782 3.630486126182e-29 1.33783413749807e-26
pre-tRNA-Tyr-GTA 505.057106284187 9.2287311054882 0.828001518415102
11.1457900743384 7.50750007057211e-29 2.21321102080466e-26
mature-tRNA-Pro-TGG_5_end;mature-tRNA-Pro-CGG_5_end;mature-tRNA-Pro-AGG_5_end
1946.81081659445 9.928241334204 1.04537621880354 9.49729021535149
2.15423010298028e-21 5.29222528632156e-19
mature-tRNA-Arg-ACG 1850.07541775039 6.35805962609316 0.67452163520523
9.42602771245233 4.25913920030078e-21 8.96853025891908e-19

```

3.sports_cpm_fdr005_2fc_all.txt

```

ID SRR11004011 SRR11004012 SRR11004013 SRR11004020 SRR11004021 SRR11004022
28S-rRNA 14.3200235328116 13.5965848351574 13.0443714276675
14.6978424726186 14.9935127374893 14.6639604791408
piRNA 13.9245729094712 13.5201406814239 12.8524707211966
10.4952997919691 10.9115022019083 12.89856566266
RNY1-YRNA 11.2882200871046 10.8179018026627 10.5560816430719
14.7959962196167 13.5006881015406 13.0984586794382
misc_RNA 11.057924529273 11.5769419854079 10.8959117870999
12.776022553392 12.8113773297272 11.6166995041221
5.8S-rRNA 10.7552419878891 10.2228471945342 7.92102213175833
13.5125514135644 12.4352583087336 11.2038607265397
45S-rRNA 9.86541494133145 10.109728768677 10.6153362976346
12.7551626430638 13.1814608784966 13.36382994117
mature-tRNA-Gly-GCC_5_end;mature-tRNA-Gly-CCC_5_end 9.61208227726735
9.43932057807877 9.34318987544876 13.0213797611129 12.9968713182784
12.8990113391204
5S-rRNA 9.27799186112964 10.7257992535941 9.81583605097291
11.2657012714733 11.4490159975246 11.3388688232466

```

4.sports_DEG_all.txt

```

ID baseMean log2FoldChange lfcSE stat pvalue padj
28S-rRNA 117369.725044431 0.382472823614208 0.644227936830634
0.593691769245269 0.552718289120833 0.673819151856151
piRNA 50767.1086731965 -2.40366212934164 0.765431138235381
-3.14027220643652 0.00168790918441722 0.00846251067289452
RNY1-YRNA 45719.3346323336 2.44151119683615 0.707386312647438
3.45145382824646 0.000557575147822217 0.00336997078222502
misc_RNA 22395.6619710466 0.67498992210017 0.6596268775384
1.02329050723206 0.306170510703025 0.448158225199861
5.8S-rRNA 19601.5728991033 1.97867701752085 0.882338784732432
2.24253659904668 0.0249267126520048 0.0724693776115484
45S-rRNA 24872.0434625502 2.26075136993462 0.569544671009807
3.96940132180727 7.20534381777695e-05 0.000689654336844366
mature-tRNA-Gly-GCC_5_end;mature-tRNA-Gly-CCC_5_end 21344.3984569
2.86332800867512 0.563247054211885 5.08360938111179 3.70329152216592e-07
9.00411658654179e-06
5S-rRNA 9981.49076365101 0.648247200530487 0.639794072975421
1.01321226299543 0.310958784289966 0.450329597790607

```

This function will automatically identify sports_DEG_fdr005_2fc_all.txt file in the folder, and categorizing the miRNAs, extract contains keywords "-miR-|-let-" for miRNA_diff. TXT file, The tsRNA_diff.txt file containing the keyword "tRNA", the rsRNA_diff.txt file containing the keyword "rRNA", and the ysRNA_diff.txt file containing the keyword "YRNA".

```
tsRNA_ann_classify()
```

1.mirRNA_diff.txt

ID	baseMean	log2FoldChange	lfcSE	stat	pvalue	padj
mmu-miR-2137	87.8349839348083		5.9563172036757	0.799484930810467		
7.45019321081833		9.3203633809477e-14	7.63234201306495e-12			
mmu-miR-153-3p	316.296855176786		-5.03376515234459	0.750489390856289		
-6.70731020807795		1.98244544430187e-11	1.12389407111575e-09			
mmu-miR-340-5p	1374.52373372071		-4.43397551622456	0.754895118332545		
-5.87363119530906		4.26351320188798e-09	1.39653743546286e-07			
mmu-miR-101a-3p	13763.2199746233		-4.86630273572994	0.87811726649203		
-5.5417458708792		2.9947070804607e-08	8.82839647319815e-07			
mmu-miR-690	1186.05912973371		3.84780460164948	0.713714629912904		
5.39123683385759		6.99743788822742e-08	1.983504509086e-06			
mmu-miR-9-3p	2296.27478279842		-4.20391814855742	0.810478351339246		
-5.18695920947278		2.13755526571688e-07	5.62635082440478e-06			
mmu-miR-136-3p	124.245832937843		-5.32869206253836	1.04845251393674		
-5.08243529554823		3.72626263079409e-07	9.00411658654179e-06			

2.tsRNA_diff.txt file

ID	baseMean	log2FoldChange	lfcSE	stat	pvalue	padj
mature-tRNA-His-GTG	30035.3716708793		9.18307904145374	0.580202283242111		
15.8273748771543		2.01452256749788e-56	2.96940626449187e-53			
pre-tRNA-Leu-CAA	21805.5636328603		9.85974407854832	0.658354292658678		
14.9763496471346		1.04818154091913e-50	7.72509795657397e-48			
mature-tRNA-Arg-CCT;mature-tRNA-Arg-CCG	468.48506156553		7.40567460160257			
0.655365046544287		11.3000756458593	1.31105017035912e-29			
6.44162650369779e-27						
mature-tRNA-Arg-ACG_CCA_end	43397.7312599756		7.76515581481599			
0.692681783635114		11.2102786564782	3.630486126182e-29	1.33783413749807e-26		
pre-tRNA-Tyr-GTA	505.057106284187		9.2287311054882	0.828001518415102		
11.1457900743384		7.50750007057211e-29	2.21321102080466e-26			
mature-tRNA-Pro-TGG_5_end;mature-tRNA-Pro-CGG_5_end;mature-tRNA-Pro-AGG_5_end						
1946.81081659445		9.928241334204	1.04537621880354	9.49729021535149		
2.15423010298028e-21		5.29222528632156e-19				
mature-tRNA-Arg-ACG	1850.07541775039		6.35805962609316	0.67452163520523		
9.42602771245233		4.25913920030078e-21	8.96853025891908e-19			

3.rsRNA_diff.txt

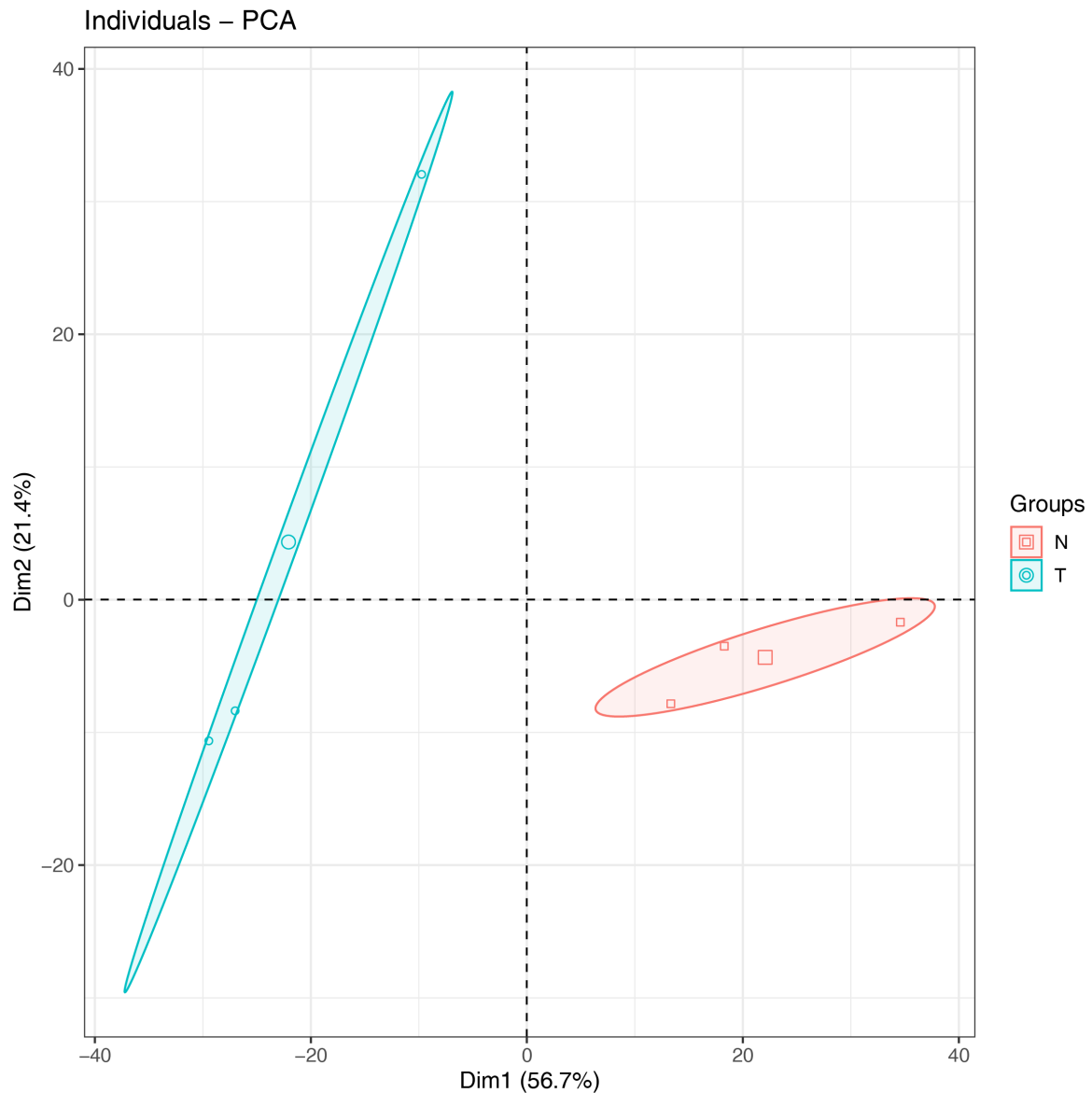
ID	baseMean	log2FoldChange	lfcSE	stat	pvalue	padj
45S-rRNA	24872.0434625502		2.26075136993462	0.569544671009807		
3.96940132180727		7.20534381777695e-05	0.000689654336844366			
rRNA	12.4284548829842		2.37423147013067	0.937010532600679		
2.53383648051529		0.0112821365345229	0.0402660272442777			

4.ysRNA_diff.txt

ID	baseMean	log2FoldChange	lfcSE	stat	pvalue	padj
RNY3-YRNA	1275.60735378423		2.23955706124081	0.603348544285474		
3.71187944754726		0.000205725970976646	0.00156309320216277			
RNY1-YRNA	45719.3346323336		2.44151119683615	0.707386312647438		
3.45145382824646		0.000557575147822217	0.00336997078222502			

This function will automatically identifies the sports_counts_all.txt file in the folder, selects the top 1000 genes by multiple change, and generates a PDF file of the principal component analysis using the ggplot2 R package.

```
pca(20:22)
```



function of visualization

pie_plot_tsRNA_aa.pdf: This function will automatically identifies the tsRNA_diff.txt file in the folder, and uses the ggplot2 R package to draw the pie chart of different amino acid classes of tsRNA, including Glu, Gly, Val, and Ser, generate the pie_plot_tsRNA_aa.pdf file.

pie_plot_tsRNA_end.pdf: This function will automatically identify the tsRNA_diff.txt file in the folder, and use the ggplot2 R package to draw the pie chart of different tsRNA end categories, including 5'end, 3'end, CCA end, generate the pie_plot_tsRNA_end.pdf file.

maplot.pdf: This function will automatically identifies the sports_deg_all.txt file in the folder and uses the ggplot2 R package to draw the MA map of sports output differentially expressed genes, generating the "maplot.pdf" file.

heatmap_plot.pdf: This function will automatically identifies the sports_cpm_fdr005_2fc_all.txt and sports_DEG_fdr005_2fc_all.txt files in the folder, selects and uses the ggplot2 R package to draw a heat map of the differentially expressed genes that sports outputs, Generate the heatmap_plot.pdf file.

volcano_plot: This function will will automatically identify the sports_DEG_all.txt file in the folder and use the ggplot2 R package to draw the volcano plot of sports output differentially expressed genes, $\log_2FC > 1$, $p \leq 0.05$, generate the "volcano_plot.pdf" file.

visualization()

Those pictures shown below are the sample data output, and the format is adjusted by Adobe Illustrator software.

