

Stat 115/215 Lab

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Outline

- Introduction to MAESTRO (Part I of HW).
- Integrating scRNA-seq and scATAC-seq data (Part IV of HW).

MAESTRO

Wang et al. *Genome Biology* (2020) 21:198
<https://doi.org/10.1186/s13059-020-02116-x>

Genome Biology

SOFTWARE

Open Access

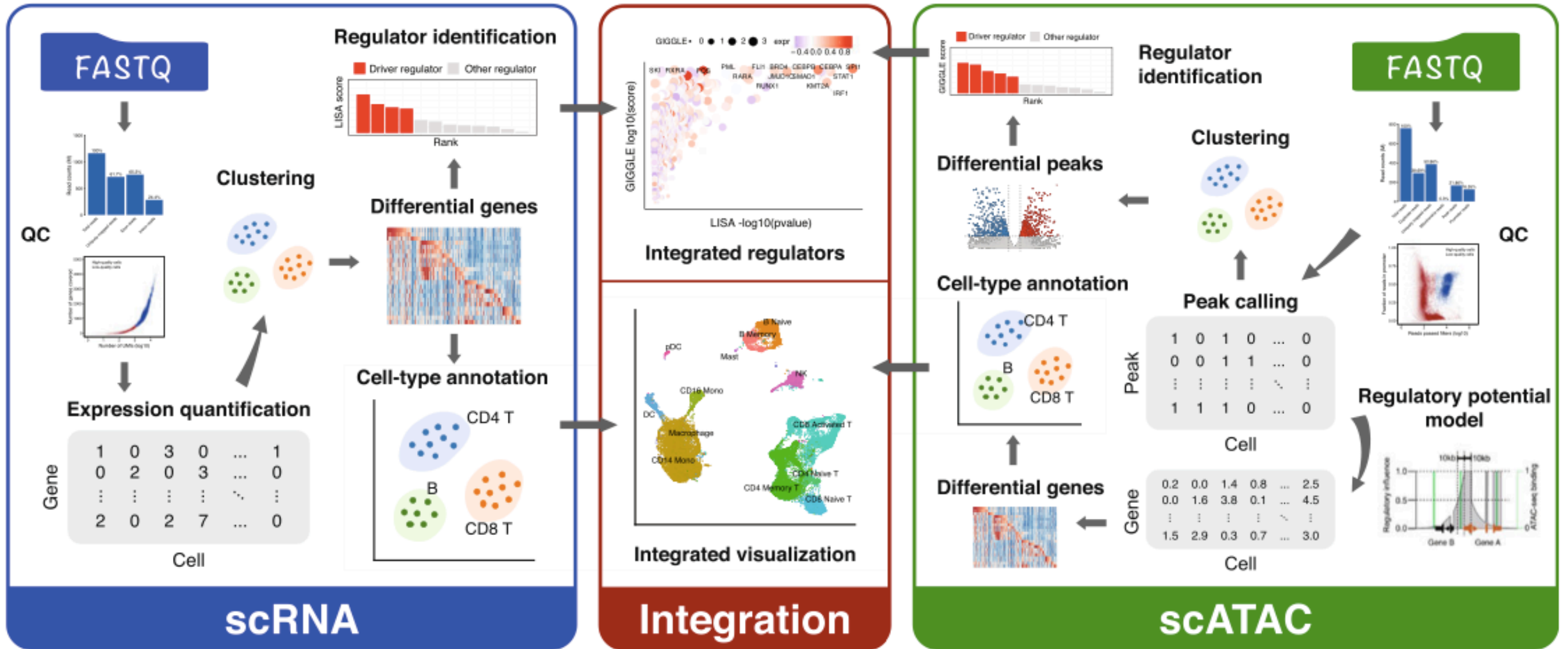
Integrative analyses of single-cell transcriptome and regulome using MAESTRO



Chenfei Wang^{1,2†}, Dongqing Sun^{3†}, Xin Huang⁴, Changxin Wan³, Ziyi Li³, Ya Han³, Qian Qin³, Jingyu Fan³, Xintao Qiu^{2,5}, Yingtian Xie^{2,5}, Clifford A. Meyer^{1,2}, Myles Brown^{2,5}, Ming Tang^{1,2}, Henry Long^{2,5}, Tao Liu^{6*} and X. Shirley Liu^{1,2*} 

What is MAESTRO?

- MAESTRO (Model-based Analyses of Single-cell Transcriptome and RegulOme) is a pipeline for the analysis and integration of scRNA-seq and scATAC-seq data.
- MAESTRO can start from a FASTQ file and return a processed Seurat object (thereby performing alignment, demultiplexing of barcodes, cell-type annotation, etc).



Running MAESTRO

- Maestro is built using Snakemake (Mölder et al., 2021). Snakemake is a tool for workflow management.
- Four steps for running on Cannon:
 - Source the conda environment.
 - Configure the pipeline using MAESTRO scrna-init or MAESTRO scatac-init. You do not need to run this on a compute node.
 - Run the analysis as a batch job. I specified 60 GB of RAM, 5 hours of run-time, and 16 CPUs.
 - Analyze the results (copy and paste text files and transfer images to your local computer).

Initialization for scRNA-seq

```
usage: MAESTRO scrna-init [-h] [--platform {10x-genomics,Dropseq,Smartseq2}]
--fastq-dir FASTQ_DIR [--fastq-prefix FASTQ_PREFIX]
[--fastq-barcode FASTQ_BARCODE]
[--fastq-transcript FASTQ_TRANSCRIPT]
[--species {GRCh38,GRCm38}] [--cores CORES]
[--rseqc] [--directory DIRECTORY]
[--outprefix OUTPREFIX]
[--count-cutoff COUNT_CUTOFF]
[--gene-cutoff GENE_CUTOFF]
[--cell-cutoff CELL_CUTOFF] --mapindex MAPINDEX
[--rsem RSEM] [--whitelist WHITELIST]
[--barcode-start BARCODE_START]
[--barcode-length BARCODE_LENGTH]
[--umi-start UMI_START] [--umi-length UMI_LENGTH]
[--lisadir] [--signature SIGNATURE]
```

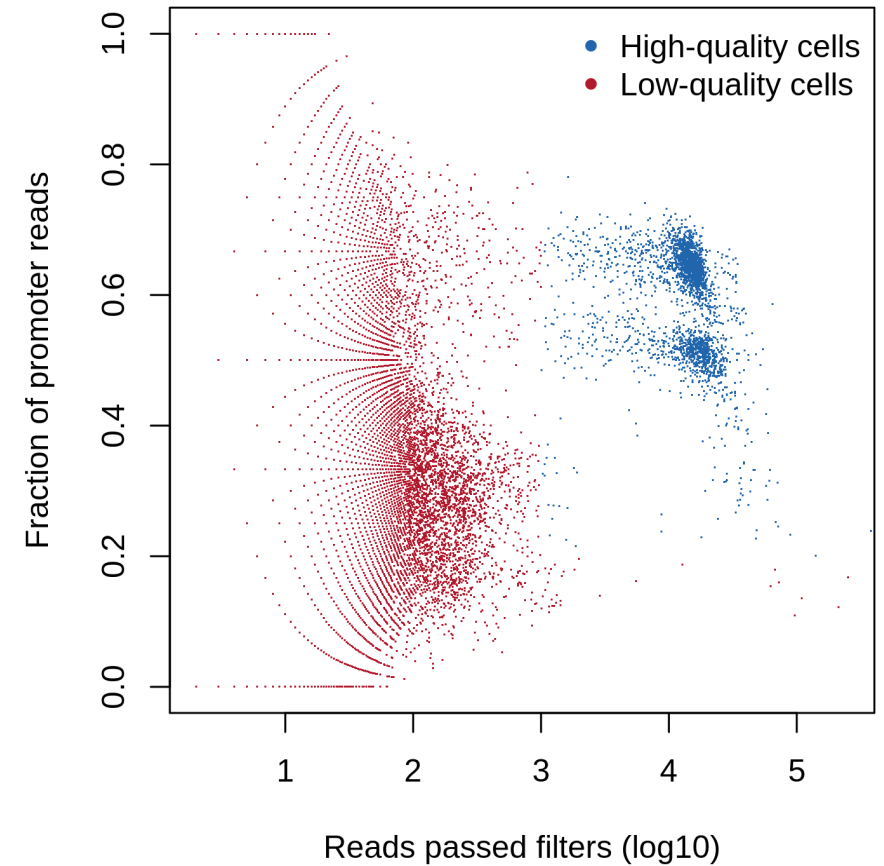
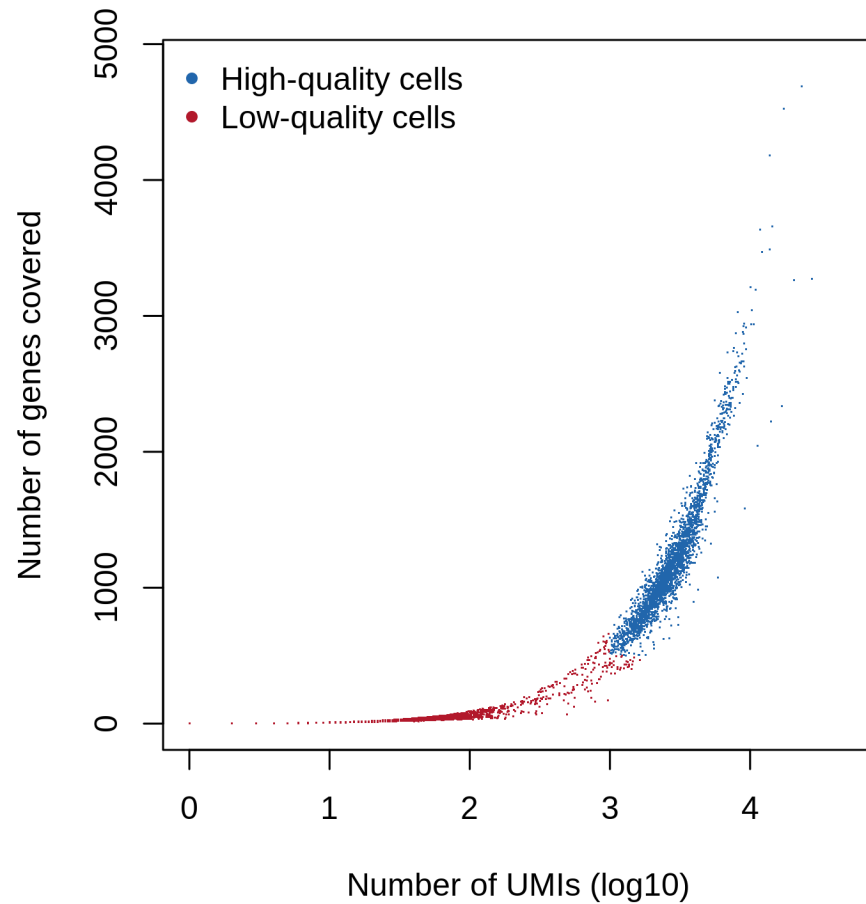
Initialization for scATAC-seq

```
usage: usage: MAESTRO scatac-init [-h]
      [--platform {10x-genomics,sci-ATAC-seq,microfluidic}]
      [--format {fastq,bam,fragments}]
      [--fastq-dir FASTQ_DIR]
      [--fastq-prefix FASTQ_PREFIX] [--bam BAM]
      [--frag FRAG] [--species {GRCh38,GRCm38}]
      [--cores CORES] [--directory DIRECTORY]
      [--outprefix OUTPREFIX] [--peak-cutoff PEAK_CUTOFF]
      [--count-cutoff COUNT_CUTOFF]
      [--frip-cutoff FRIP_CUTOFF]
      [--cell-cutoff CELL_CUTOFF] --giggleannotation
      GIGGLEANNOTATION [--fasta FASTA]
      [--whitelist WHITELIST] [--custompeak]
      [--custompeak-file CUSTOMPEAK_FILE] [--shortpeak]
      [--clusterpeak] [--rpmodel {Simple,Enhanced}]
      [--genedistance GENEDISTANCE] [--annotation]
      [--method {RP-based,peak-based,both}]
      [--signature SIGNATURE]
```


Running the pipeline on Cannon

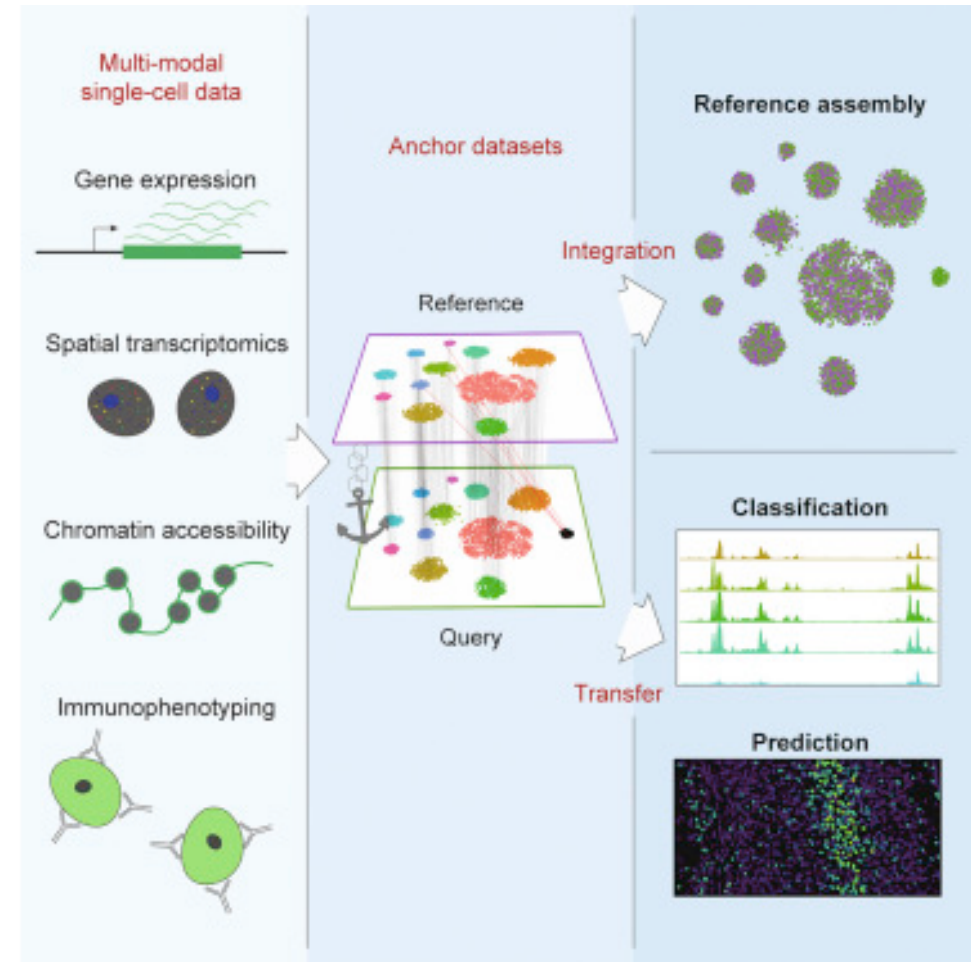
- Make sure that you have specified sufficient resources.
- Since you only have ~100GB of space in your home directory, you will likely need to delete large files from previous HWs before running MAESTRO.
- When you run `snakemake`, you should be in the `multiome_scrna` folder.

Viewing the QC results



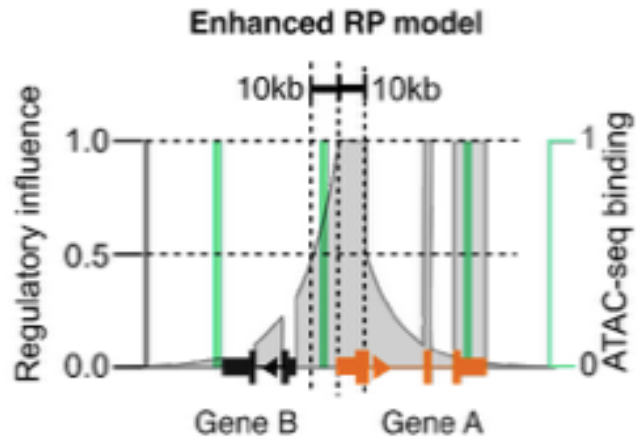
Integrating scRNA-seq and scATAC-seq

- A strength of scRNA-seq is that we have direct knowledge of the gene expression (this is better for things such as cell type annotation).
- In Part IV of the homework, we use “label transferring” to integrate scATAC-seq with scRNA-seq.
- (Big) Hint:
https://satijalab.org/seurat/articles/atacseq_integration_vignette.html
contains most of the code needed for this part of the HW.

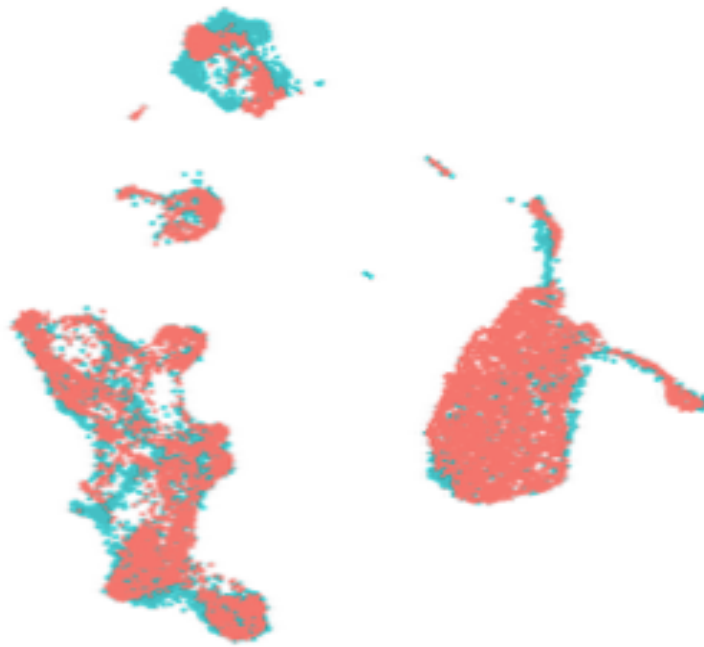


MAESTRO develops a regulatory potential model to predict gene activity in scATAC-seq data. From this, the two datasets can be treated as two different batch and a batch-correction method (CCA) can be applied.

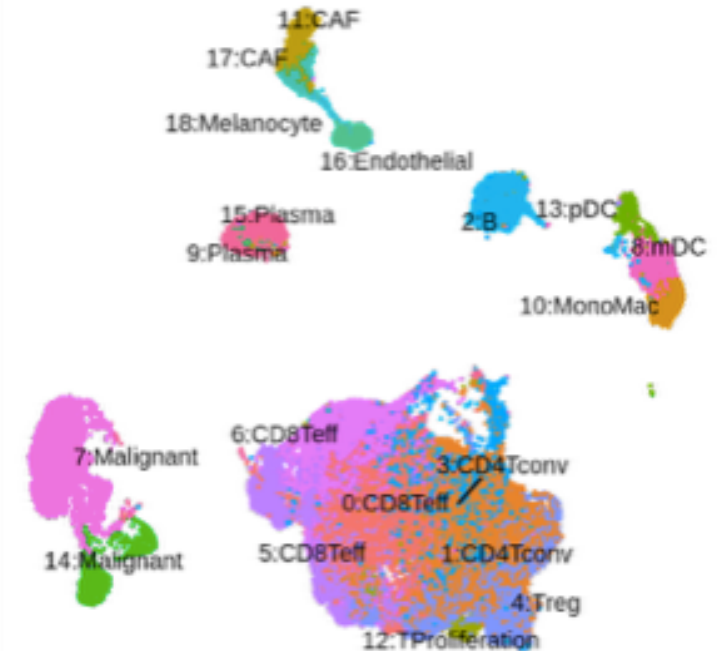
Gene activity modelling from scATAC-seq



Integration of PBMC scRNA-seq and scATAC-seq



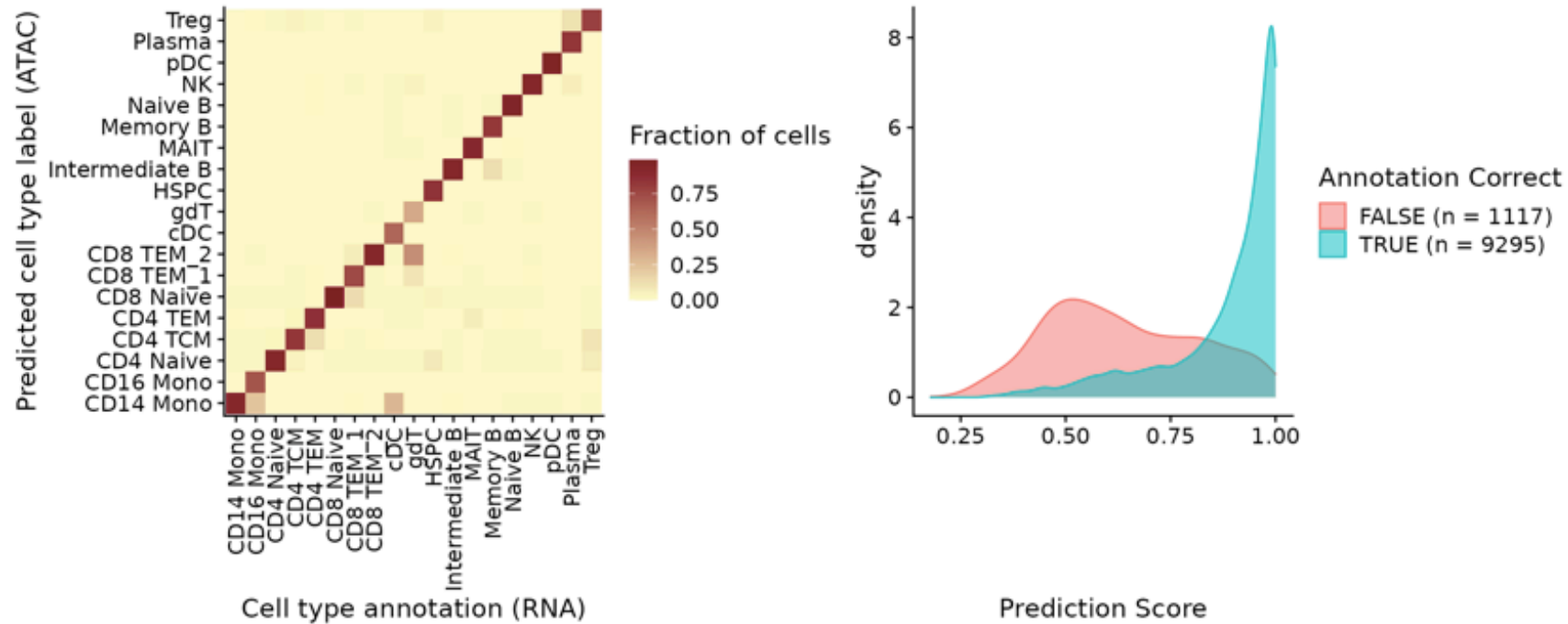
Integration of large-scale scRNA-seq and scATAC-seq



Key steps for Part IV

- Step 1: Match barcodes between scRNA-seq and scATAC-seq (most of the code for this part is already given).
- Step 2: Run `FindTransferAnchors()` to obtain a "mapping" between the scRNA-seq and the scATAC-seq data.
- Step 3: Run `TransferData()` to annotate the scATAC-seq data using the scRNA-seq data.
- Step 4: Compare the inferred labels to the true labels.

Sample plot for 3.3 and 3.4 (See Vignette for code)



Other hints and suggestions for homework

- Part I, Problem 2: use `scATAC_cell_filtering.png` instead of `scATAC_read_distr.png`.
- Part II, Problem 2: Consider the Seurat function `PercentageFeatureSet` and look at the provided example.
- Part II, Problem 5.1: The first answer provided by the link is wrong, make sure you read the follow-up to understand why.