

Genetic Diversity of *Salmonella* Pathogenicity Islands SPI-5 and SPI-6 in *Salmonella* Newport

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Abstract

Salmonella enterica subspecies *enterica* serotype Newport is one of the common serotypes causing foodborne salmonellosis outbreaks in the United States. *Salmonella* Newport consists of three lineages exhibiting extensive genetic diversity. Due to the importance of *Salmonella* pathogenicity islands 5 and 6 (SPI-5 and SPI-6) in virulence of pathogenic *Salmonella*, the genetic diversity of these two SPIs may relate to different potentials of *Salmonella* Newport pathogenicity. Most *Salmonella* Newport strains from North America belong to *Salmonella* Newport lineages II and III. A total 28 *Salmonella* Newport strains of lineages II and III from diverse sources and geographic locations were analyzed, and 11 additional *Salmonella* genomes were used as outgroup in phylogenetic analyses. SPI-5 was identified in all *Salmonella* Newport strains and 146 single nucleotide polymorphisms (SNPs) were detected. Thirty-nine lineage-defining SNPs were identified, including 18 non-synonymous SNPs. Two 40-kb genomic islands (SPI5-GI1 and SPI5-GI2) encoding bacteriophage genes were found between tRNA-ser and pipA. SPI5-GI1 was only present in *Salmonella* Newport multidrug-resistant strains of lineage II. SPI-6 was found in all strains but three Asian strains in *Salmonella* Newport lineage II, whereas the three Asian strains carried genomic island SPI6-GI1 at the same locus as SPI-6 in other *Salmonella*. SPI-6 exhibited 937 SNPs, and phylogenetic analysis demonstrated that clustering of *Salmonella* Newport isolates was a reflection of their geographic origins. The sequence diversity within SPI-5 and SPI-6 suggests possible recombination events and different virulence potentials of *Salmonella* Newport. The SNPs could be used as biomarkers during epidemiological investigations.

Introduction

NONTYPHOIDAL *SALMONELLA* spp. cause an estimated 1.4 million foodborne illnesses annually in the United States, accounting for 11% of all foodborne infections (Scallan *et al.*, 2011). *Salmonella enterica* subspecies *enterica* serotype Newport (*Salmonella* Newport) causes over 100,000 infections annually. *Salmonella* Newport has ranked as the third serotype causing illnesses and has been responsible for multistate foodborne outbreaks in the United States (CDC, 2006; Greene *et al.*, 2008), including outbreaks associated with ground beef and tomatoes since 2002 (Bell *et al.*, 2012; Greene *et al.*, 2008; Schneider *et al.*, 2011).

Salmonella Newport consists of three lineages with extensive genetic diversity (Sangal *et al.*, 2010). Most *Salmonella* Newport strains from Europe belong to lineage I,

whereas most North American strains belong to lineages II and III (Sangal *et al.*, 2010). Whole genome sequence analysis of 28 *Salmonella* Newport strains from diverse sources and locations grouped the strains into lineages II and III with clustering explained by geographic origins (Cao *et al.*, 2013). The Asian strains were clustered separately from American strains. To arrive at a comprehensive evolutionary picture of *Salmonella* Newport, it would be necessary to include all three lineages; however, no *Salmonella* Newport lineage I strain has been sequenced to date.

Genomic island (GI) is a gene cluster that has been acquired via horizontal gene transfer (Langille *et al.*, 2010). Pathogenicity islands are gene clusters encoding virulence determinants that are usually absent in nonpathogenic strains of the same or closely related species (Sabbagh *et al.*, 2010). A total of 22 *Salmonella* pathogenicity islands (SPIs) have

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been identified to date (Sabbagh *et al.*, 2010). *Salmonella* Newport genomes contained SPI-1 through SPI-4 sequences (unpublished data) and showed extensive diversities at the region around *mutS* downstream of SPI-1 (Cao *et al.*, 2013).

SPI-5 was first identified in the *Salmonella* Dublin genome between tRNA-*serT* and *copR* and was found to consist of five genes (*pipA*, *pipB*, *pipC*, *sopB*, and *pipD*) (Wood *et al.*, 1998). These five genes displayed high similarity with genes from bacteriophages Gifsy-1 and Gifsy-2 (Figueroa-Bossi *et al.*, 2001). SPI-5 plays a vital role in pathogenicity and encodes effectors of SPI-1 and SPI-2 (Sabbagh *et al.*, 2010). For example, *sopB* encodes a translocated effector protein of type III secretion systems (T3SS) in SPI-1 under control of *hilA*, whereas *pipB* encodes a translocated effector of T3SS in SPI-2 under control of *ssrAB* (Knodler *et al.*, 2002; Hensel, 2004). SPI-5 contributes to the colonization of the spleen in chickens (Rychlik *et al.*, 2009). Mutations in SPI-5 genes significantly reduced the enteropathogenicity of *Salmonella* (Wood *et al.*, 1998).

SPI-6 is located between tRNA-*aspV* and *sinR* at centisome 7 in *Salmonella* encoding a type six secretion system (T6SS) and a *Salmonella* atypical fimbriae (*saf*) cluster (Sabbagh *et al.*, 2010). SPI-6 has different gene contents in various serotypes. For example, it is a 47-kb island in *Salmonella* Typhimurium (Folkesson *et al.*, 1999) and a 59-kb island in *Salmonella* Typhi (Parkhill *et al.*, 2001). T6SS is widespread in bacteria (Schwarz *et al.*, 2010), and its gene products perform diverse functions (Blondel *et al.*, 2009;

Jani and Cotter, 2010), one of which is to mediate antagonistic interactions between bacteria (Hood *et al.*, 2010). Folkesson *et al.* (Folkesson *et al.*, 2002) reported that the deletion of SPI-6 reduced the invasion activity of *Salmonella* Typhimurium into Hep2 cells. The *saf* genes are located downstream of T6SS in SPI-6 and are present in most clinical isolates of *Salmonella* (Folkesson *et al.*, 1999; Humphries *et al.*, 2003). However, the *saf* operon encoding nonfimbrial adhesion elements does not contribute to virulence in mice (Folkesson *et al.*, 1999).

The objectives of the current study were to investigate the different virulence potential via identifying genetic diversity in SPI-5 and SPI-6 of *Salmonella* Newport lineages II and III and to identify markers in SPI-5 and SPI-6 for *Salmonella* Newport subtyping.

Materials and Methods

Genomes

Twenty-eight *Salmonella* Newport genomes from diverse sources and locations from our previous work (Table 1) and 11 outgroup genomes were analyzed in the current study (Cao *et al.*, 2013; Lienau *et al.*, 2013; Timme *et al.*, 2013), including *Salmonella* Tennessee CDC07_0191 (ACBF00000000), *Salmonella* Kentucky CVM29188 (ABAK000000000), *Salmonella* Kentucky CDC191 (ABEI000000000), *Salmonella* Gallinarum 287/91 (AM933173.1), *Salmonella* Dublin CT02021853 (CP001144.1), *Salmonella* Hadar RI_05P066 (ABFG000000000), *Salmonella* Typhimurium

TABLE 1. GENOME INFORMATION OF *SALMONELLA* NEWPORT STRAINS USED IN THE STUDY^a

Strain	Tree label	Accession number	Genome size (Mbp)	Number of contigs
CVM35185	Bison_TN_2004	AHTJ000000000	4.71	95
CVM35199	Caprine_TN_2004	AHTK000000000	4.75	72
CVM21539	Chicken_MO	AHTL000000000	4.71	71
CVM33953	Ground_turkey_MD_2003	AHTM000000000	4.80	88
CVM35188	Equine_TN_2004_1	AHTN000000000	4.71	66
CVM21559	Turkey_CO	AHTO000000000	4.74	64
CVM19447	Frog_Vietnam	AHTP000000000	4.67	59
CVM19449	Fish_Hong_Kong	AHTQ000000000	4.70	76
CVM19567	Fish_Vietnam	AHTR000000000	4.67	53
CVM35202	Equine_TN_2004_2	AHTS000000000	4.96	72
CVM21550	Swine_TX	AHTT000000000	4.92	73
CVM22513	Cattle_NC_2003	AHTU000000000	4.90	72
CVM21538	Chicken_GA	AHTV000000000	4.93	70
CVM22425	Cattle_AZ_2003	AHTW000000000	4.93	69
CVM22462	Canine_AZ_2003	AHTX000000000	5.02	384
CVMN18486	Ground_turkey_NM_2008	AHTY000000000	4.93	85
CVMN1543	Ground_beef_GA_2004	AHTZ000000000	4.89	77
CVM21554	Swine_IL_2001	AHUA000000000	4.69	44
CVM19443	Shrimp_India	AHUB000000000	4.81	70
CVM37978	Spinach_CO_2008	AHUC000000000	4.80	49
CVM19593	Cheese_Mexico	AHUD000000000	4.65	74
CVM19470	Squid_Vietnam	AHUE000000000	4.73	84
CVM19536	Pepper_Vietnam	AHUF000000000	4.65	70
CVM4176	Pig_ear_CA	AHUG000000000	4.73	62
Levine 1	Farm_1_VA_2007	AJMN000000000	4.81	91
Levine 15	Farm_15_VA_2007	AJMO000000000	4.81	75
SL254	<i>S. Newport</i> SL254	ABEN010000000	4.83	0
SL317	<i>S. Newport</i> SL317	ABEW000000000	4.95	63

^aThese genomes were selected from our published study.

LT2 (NC_003197.1), *Salmonella* Typhimurium SL1344 (NC_016810.1), *Salmonella* Typhimurium D23580 (NC_016854.1), *Salmonella* Typhimurium 14028S (CP001363.1) and *Salmonella* 4,5,12:i:- SL474 (ABAO00000000).

Phylogenetic analysis

A whole genome parsimony tree was reconstructed based on 131,855 informative single nucleotide polymorphisms (SNPs) with the Tree analysis using New Technology (TNT) program (Goloboff *et al.*, 2008). The phylogenetic analysis found a minimum tree length with 20 reiterations using Section Search, Ratchet, Drift, and Tree fusing methods, and it calculated 100,000 bootstrapping replicates. Multiple sequence alignment using MULCLE with default parameter (Edgar, 2004) in SEAVIEW (Galtier *et al.*, 1996) identified 146 SNPs in SPI-5, 937 SNPs in SPI-6 (excluding *saf* genes), and 355 SNPs in *saf* genes. Parsimony trees of SPI-5, SPI-6, and *saf* genes were reconstructed using TNT and the same parameters as above. Certain strains were not included in analyses of SPI-5, SPI-6, or the *saf* genes because of the poor data quality of the draft genomes, such as canine_AZ_2003, bison_TN_2004, and equine_TN_2004_1.

Genetic characterizations of SPI-5 genomic islands 1 and 2 (SPI5-GI1 and SPI5-GI2), and SPI-6 genomic island 1 (SPI6-GI1)

Genetic organizations of SPI5-GIs and SPI6-GI1 were displayed using Mauve (Darling *et al.*, 2004). The best match of genes in SPI5-GI1, SPI5-GI2, and SPI6-GI1 was determined using blastp (Altschul *et al.*, 1990), followed by verification using tblastn (Altschul *et al.*, 1990).

Distance matrix

MEGA 6.05 (Tamura *et al.*, 2011) was used to calculate evolutionary distances (number of differences) over sequence pairs with 10,000 bootstrap iterations for SPI-5, SPI-6, and *saf* genes.

Results

Phylogenetic tree based on whole genome data

A whole genome phylogenetic tree was constructed using more than 131,855 SNPs (Fig. 1). To better display the evolutionary relationship between *Salmonella* Newport strains, we selected 11 genomes as outgroups. There were six

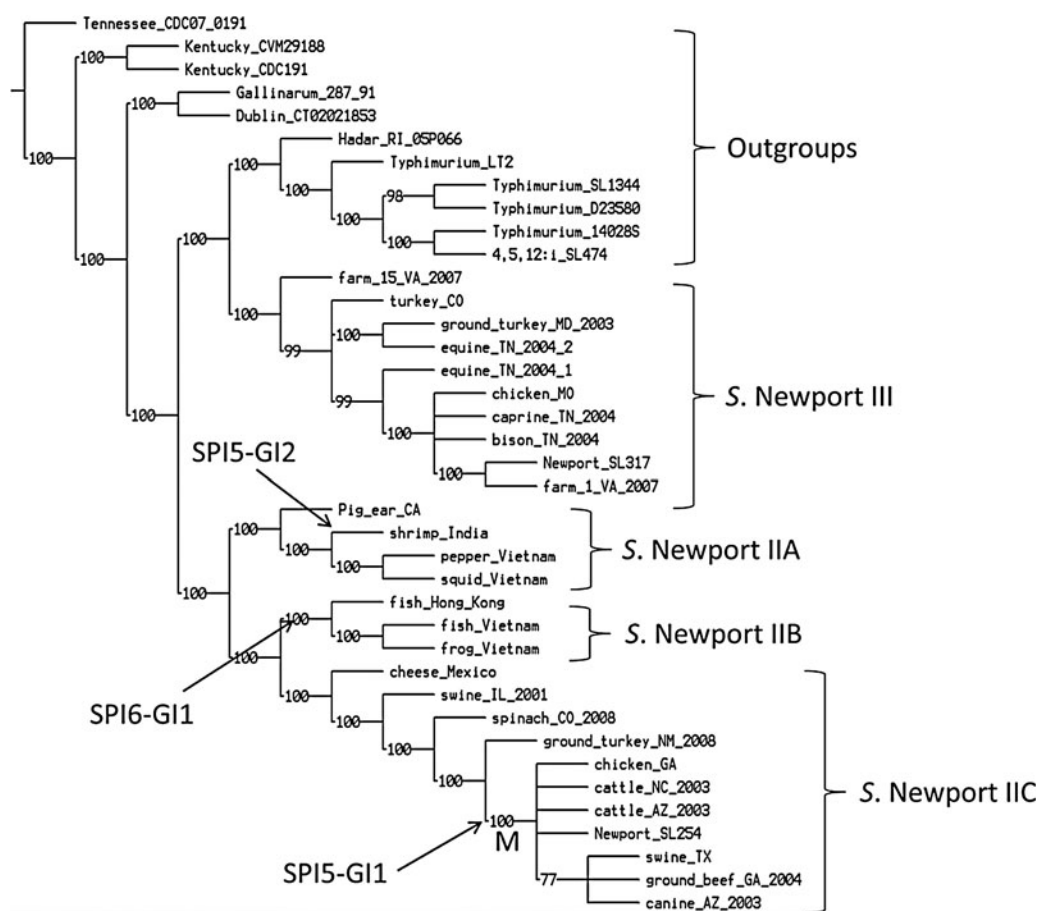


FIG. 1. Whole-genome parsimony tree of *Salmonella* Newport and 11 outgroup genomes. *Salmonella* Newport strains showed phylogenies identical to those of a previous study (Cao *et al.*, 2013). There are six equally most parsimonious trees identified with a length of 209,114 single nucleotide polymorphisms, consistency index of 0.616, and retention index of 0.888. Two gene clusters, SPI5-GI1 and SPI5-GI2, encoding bacteriophage genes are displayed. The rest of the genomes do not contain SPI5-1 or SPI5-2.

equally most parsimonious trees with the same branch order at the subgroup level, meaning that the *Salmonella* Newport strains in each subgroup were the same in all the resulting trees. *Salmonella* Newport strains were divided into lineages II and III. Lineage II was further grouped into subgroups IIA, IIB, and IIC. All multidrug-resistant (MDR) strains were in node M of subgroup IIC (Cao *et al.*, 2013).

Genetic diversity of *Salmonella* pathogenicity island 5

SPI-5 was present in 28 *Salmonella* Newport and 11 outgroup genomes. SPI-5 variations included insertions and SNPs. Two genomic islands encoding prophage genes were found between tRNA-*ser* and *pipA* in certain genomes and designated as SPI-5 genomic islands 1 and 2 (SPI5-GI1 and SPI5-GI2) (Supplementary Fig. S1; Supplementary Data are available online at www.liebertpub.com/fpd). SPI5-GI1 was only present in node M, which contained MDR strains (Cao *et al.*, 2013). SPI5-GI2 was only found in strain shrimp_India. The rest of the *Salmonella* Newport genomes included in the current work do not contain any insertion in SPI-5. The SPI5-GI1 and SPI5-GI2 were approximately 41 and 44 kb in length, respectively, encoding prophage genes (Supplementary Tables S1 and S2).

Some genomes did not contain the entire SPI5-GI1 and SPI5-GI2 between tRNA-*ser* and *pipA*; however, partial sequences of SPI5-GIs were identified. For example, a gene cluster in SPI5-GI1 (SNSL254_A1155 to SNSL254_A1177, 5' to 3') was present in *Salmonella* Typhi CT18, *Salmonella*

Paratyphi B SPB7, *Salmonella* Paratyphi C RKS4594, and *S. Choleraesuis* SC-B67. Similarly, part of the SPI5-GI2 sequence (SEEN443_12678 to SEEN443_12753, 5' to 3') was identified in *Salmonella* Weltevreden HI_N05-537, *Salmonella* Newport SNSL317, *Salmonella* Typhimurium DT104, and *Salmonella* Saintpaul SARA29. The blast matches indicated that 74% and 52% of the genes in SPI5-GI1 and SPI5-GI2, respectively, encoded hypothetical or bacteriophage proteins. Based on current annotation, no gene relating to virulence or antimicrobial resistance was present. Both SPI5-GIs contained genes encoding a methylase (Supplementary Tables S1 and S2).

The five genes in SPI-5 possessed 146 SNPs (Supplementary Table S3). The phylogenetic tree of SPI-5 showed that *Salmonella* Newport lineages II and III were separated by outgroup genomes (Fig. 2). TNT program identified 227 equally most parsimonious trees with the same taxa at lineage level, meaning that *Salmonella* Newport isolates in each lineage were clustered together and separated by outgroup in the resulting trees. SNPs in SPI-5 could not distinguish *Salmonella* Newport at the subgroup level in lineage II. Pairwise distance matrix showed SNPs differences between *Salmonella* Newport and other serotypes (Table 2). The average differences between lineages II and III were 40 SNPs but only 18 SNPs between *Salmonella* Typhimurium and lineage II.

A total of 39 SNPs in SPI-5 defined lineages II and III, meaning that all strains in each lineage shared the same

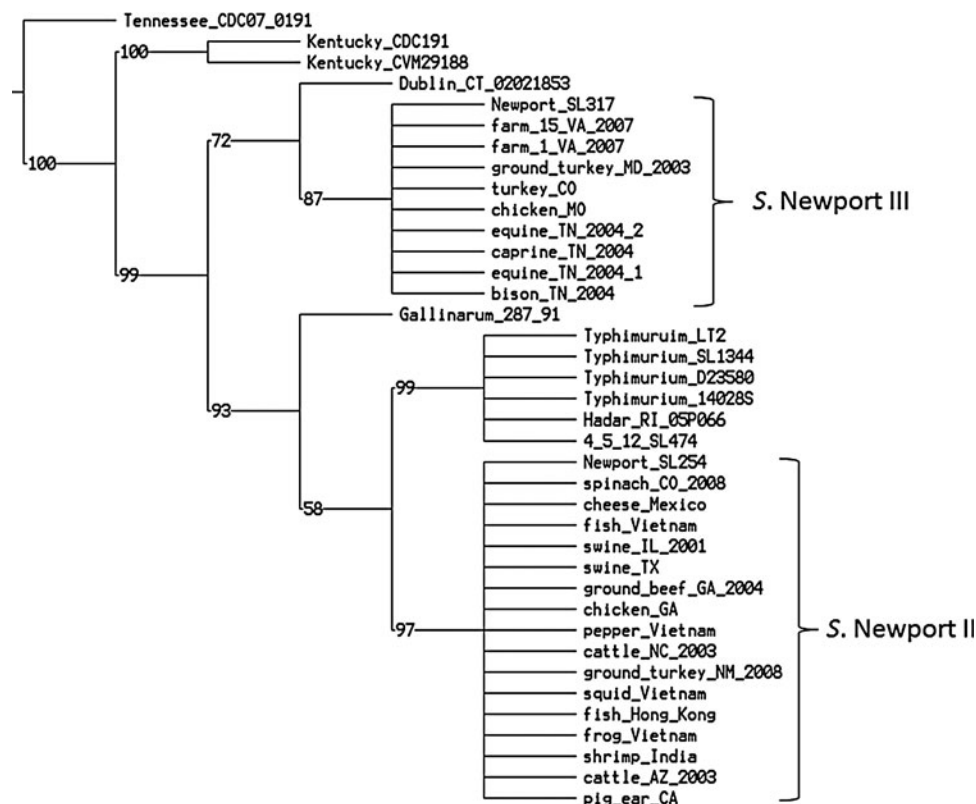


FIG. 2. Parsimony phylogenetic tree of SPI-5 genes. There are 227 equally most parsimonious trees identified with a length of 187 single nucleotide polymorphisms, and consistency index of 0.797, and retention index (RI) of 0.942. Lineages II and III were separated by outgroup genomes. Lineage II displays close relationship with *Salmonella* Typhimurium group, *Salmonella* 4,[5],12:i:- SL474, *Salmonella* Hadar RI_05P066, and *Salmonella* Gallinarum 287/91; lineage III shows a close relationship with *Salmonella* Dublin CT_02021853.

TABLE 2. AVERAGE PAIRWISE DISTANCE (NO. OF NUCLEOTIDE DIFFERENCE) OF SPI-5 AND *saf* FIMBRIAL OPERON IN GENOMES OF *SALMONELLA* NEWPORT AND OUTGROUPS

	<i>Tennessee</i>	<i>Kentucky</i>	<i>Dublin</i>	<i>Newport III</i>	<i>Gallinarum</i>	<i>Typhimurium</i>
SPI-5						
Tennessee						
Kentucky	62 (6)					
Dublin	76 (6)	73 (6)				
Newport III	70 (6)	71 (6)	18 (4)			
Gallinarum	88 (6)	83 (6)	50 (6)	50 (6)		
Typhimurium	83 (6)	82 (6)	41 (5)	31 (5)	41 (5)	
Newport II	81 (6)	74 (6)	49 (6)	40 (5)	35 (5)	18 (4)
<hr/>						
	<i>Newport IIA&B</i>	<i>Shrimp_India^a</i>	<i>Tennessee</i>	<i>Pig_ear_CA^a</i>	<i>Newport III</i>	<i>Typhimurium</i>
<hr/>						
<i>saf</i> operon						
Newport IIA&B						
Shrimp_India ^a	79 (7)					
Tennessee	71 (7)	21 (4)				
Pig_ear_CA ^a	210 (9)	212 (9)	217 (9)			
Newport III	210 (9)	210 (9)	215 (9)	11 (3)		
Typhimurium	137 (9)	141 (9)	144 (9)	197 (9)	197 (9)	
Newport IIC	223 (9)	221 (9)	222 (9)	108 (8)	109 (8)	194 (9)
Gallinarum	233 (8)	236 (6)	241 (9)	105 (8)	104 (8)	193 (9)
						67 (7)

Distances were calculated using the concatenated alignment of single nucleotide polymorphisms in SPI-5 and *saf* fimbrial operon that estimate the diversity between two major lineages and outgroup genomes observed. Standard deviation is listed in parentheses.

^aShrimp_India and Pig_ear_CA are *Salmonella* Newport strains and not included in the group Newport IIA&B and Newport IIC.

nucleotide sequence (4 SNPs in *pipA*, 9 in *pipB*, 7 in *pipC*, 5 in *sopB*, and 14 in *pipD*) (Table 3). Among the lineage-defining SNPs, 18 SNPs led to nonsynonymous substitutions, including 4, 7, 2, 2, and 3 nonsynonymous substitutions in *pipA*, *pipB*, *pipC*, *sopB*, and *pipD*, respectively.

Genetic diversity of *Salmonella* pathogenicity island 6

An intact SPI-6 (T6SS part and *saf* operon) was present in all *Salmonella* Newport genomes, except the Asian strains in subgroup IIA including shrimp_India, squid_Vietnam, and pepper_Vietnam (Fig. 1). These three Asian strains contained one common gene cluster, named SPI-6 genomic island 1 (SPI6-GI1), and the *saf* genes. Thus, the “T6SS part” and the *saf* genes were analyzed separately. The complete genome *Salmonella* Virchow SL491 had gene contents identical to those of these Asian strains. Thus, *Salmonella* Virchow SL491 was used as an example to show the genetic characterization of SPI6-GI1 (Supplementary Table S4). According to the annotation, SPI6-GI1 did not carry any gene known to be related to virulence or antimicrobial resistance.

The phylogenetic tree based on T6SS was constructed using 937 SNPs (Fig. 3). TNT program identified 208 equally parsimonious trees with the same taxa at the subgroup level. The tree reflected the geographic origin of the isolates at the lineage level, meaning that the Asian strains in subgroup IIB were clustered separately from all American strains. Among the American strains, lineage III and subgroup IIC were separated. There were 672 SNP differences between IIB and IIC, but only 222 between IIB and *Salmonella* Hadar RI_05P066.

All 28 *Salmonella* Newport strains contained the *safABCD* operon. Similar to the T6SS tree, the six Asian strains were grouped together and clustered separately from the American

strains (Fig. 4). Subgroup IIB strains clustered together. Strain shrimp_India displayed a distant relationship with the other five Asian strains. *Salmonella* Tennessee contained 71 SNP differences with IIA&B and only 21 SNP differences with shrimp_India (Fig. 4, Table 2). In the American group, lineage III and subgroup IIC were separated by *Salmonella* Gallinarum 287/91. Strain pig_ear_CA in IIA seemed to be an exception, showing a close relationship with lineage III (Table 2). Additionally, a gene cluster consisting of the *tcfABCD* fimbrial operon, *tinR*, and *tioA*, was only present in strains squid_Vietnam and pepper_Vietnam in subgroup IIA (Supplementary Fig. S2).

Discussion

SPIs play significant roles in causing human illness (Sabbagh *et al.*, 2010). Due to the similarities in nucleotide sequence between bacteriophages and pathogenicity islands (PAIs), PAIs likely originated from phage via horizontal gene transfer (HGT). Examples include SPIs (Sabbagh *et al.*, 2010), *Vibrio* pathogenicity island, and *Staphylococcus aureus* pathogenicity island 1 (SaPI1) (Boyd *et al.*, 2001). Knodler *et al.* (2002) reported that SPI-5 genes might have been acquired through HGT from lambdoid phages, including Gifsy-1 and Gifsy-2. Therefore, bacteriophages may play vital roles in virulence activities of *Salmonella* and facilitate survival of the bacteria in different environments. For example, bacteriophages have been important for the genomic evolution of *Salmonella* Montevideo and *Salmonella* Enteritidis (Allard *et al.*, 2012, 2013).

SPI5-GIs containing bacteriophage genes also may play significant roles in virulence. We hypothesized that SPI5-GI1 was originally acquired by the most recent common ancestor of node M via HGT and transmitted it vertically to the

TABLE 3. SINGLE NUCLEOTIDE POLYMORPHISMS (SNPs) OF SPI-5 GENES DEFINING *SALMONELLA* NEWPORT LINEAGES II AND III

Gene	<i>S. Newport</i> SL254 (lineage II)	<i>S. Newport</i> SL317 (lineage III)	Nuc	AA	Position
<i>pipA</i>	SNSL254_A1184	SNSL317_A1439	G->A	D/N	70
<i>pipA</i>	SNSL254_A1184	SNSL317_A1439	A->C	R/P	328
<i>pipA</i>	SNSL254_A1184	SNSL317_A1439	G->C	R/P	329
<i>pipA</i>	SNSL254_A1184	SNSL317_A1439	T->C	V/A	485
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	A->G	N/D	217
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	A->C	D/A	308
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	T->C	S	369
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	A->C	K/Q	412
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	A->G	K/D	517
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	A->C	K/D	519
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	A->G	N/D	532
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	C->A	T/N	554
<i>pipB</i>	SNSL254_A1185	SNSL317_A1440	T->A	L	558
<i>sopB</i>	SNSL254_A1187	SNSL317_A1442	A->G	A	114
<i>sopB</i>	SNSL254_A1187	SNSL317_A1442	T->C	S/P	127
<i>sopB</i>	SNSL254_A1187	SNSL317_A1442	T->C	V/A	134
<i>sopB</i>	SNSL254_A1187	SNSL317_A1442	T->C	D	480
<i>sopB</i>	SNSL254_A1187	SNSL317_A1442	T->C	G	1473
<i>pipC</i>	SNSL254_A1186	SNSL317_A1441	A->C	D/A	29
<i>pipC</i>	SNSL254_A1186	SNSL317_A1441	G->A	A	48
<i>pipC</i>	SNSL254_A1186	SNSL317_A1441	G->T	L	66
<i>pipC</i>	SNSL254_A1186	SNSL317_A1441	T->C	L	69
<i>pipC</i>	SNSL254_A1186	SNSL317_A1441	G->T	L	186
<i>pipC</i>	SNSL254_A1186	SNSL317_A1441	C->T	Y	192
<i>pipC</i>	SNSL254_A1186	SNSL317_A1441	A->G	T/A	193
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	C->T	D	189
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	A->G	E	216
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	A->G	I/V	259
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	G->A	A	345
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	T->C	Y	435
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	C->T	A/V	509
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	C->T	F	822
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	C->T	A/V	920
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	A->G	T	966
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	A->G	K	969
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	T->C	I	972
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	A->C	R	1002
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	C->T	P	1068
<i>pipD</i>	SNSL254_A1189	SNSL317_A1444	G->A	S	1095

A total of 39 SNPs in five genes in SNP-5 were identified. They defined *Salmonella* Newport lineages II and III, and could be used as potential biomarkers to differentiate strains during outbreak trace-back investigations. There are a total 18 SNPs causing nonsynonymous substitutions.

offspring strains (Fig. 1). SPI5-GII may have become functionally compatible with the genomes in node M, which includes all MDR strains (Cao *et al.*, 2013). The presence of SPI5-GI2 indicates that the location between tRNA-*ser* and *pipA* may be a hot spot for independent acquisitions of foreign genetic elements. Functional studies of SPI-5 with and without these GIs might be important to examine the possible role of both SPI5-GIs. Both SPI5-GIs contained genes encoding a methylase, which could potentially regulate chromosome replication, cell cycle events, pathogenicity, and gene expression (Fang *et al.*, 2012; Davis *et al.*, 2013).

The SPI-5 genes could be considered targets for resequencing and biomarkers to rapidly differentiate lineages II and III. We performed positive selection tests for *pipA* and *pipB* using codon-based Z tests in MEGA6, indicating that these two genes were under positive selection. Positive se-

lection played critical roles in the evolution of bacterial pathogens in that it accounts for 1.2% of the *Salmonella* core genome including virulence genes (Soyer *et al.*, 2009). Soyer *et al.* reported that three genes showed evidence of positive selection in SPI-1 through SPI-6, including *pipB* (SPI-5) and *safC* (SPI-6) (Soyer *et al.*, 2009). Since *Salmonella* Newport and *pipA* were not included in Soyer's study, *pipA* may show serotype-specific positive selection in *Salmonella* Newport.

Nonsynonymous substitutions in SPI-5 may have influenced the pathogenicity of the corresponding isolates. Two nonsynonymous substitutions were identified in domain CHASE3 in *pipA*, which is associated with signal transduction pathways in bacteria (Zhulin *et al.*, 2003). *pipB* encodes a translocated effector of T3SS in SPI-2 (Knodler *et al.*, 2002; Hensel, 2004). Moreover, a *pipB* null mutant caused reduced virulence in bovine hosts (Wood *et al.*, 1998) and facilitates

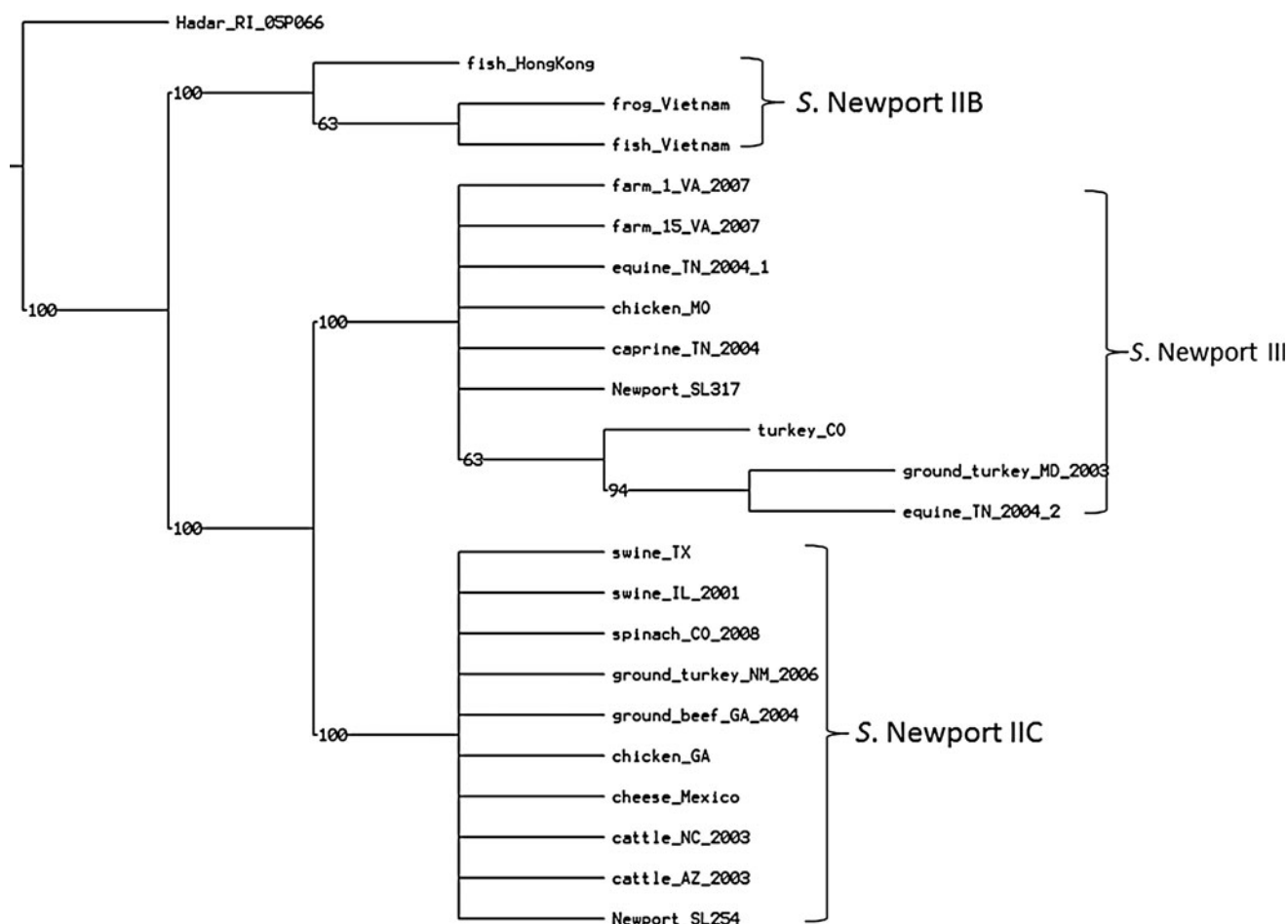


FIG. 3. Parsimony phylogenetic tree of SPI-6 genes. There were 208 equally most parsimonious trees determined with a length of 1029 single nucleotide polymorphisms, consistency index of 0.914, and retention index (RI) of 0.984. SPI-6 clustering reflects geographic origins. The Asian strains were clustered separately from the American strains.

colonization of the cecum in chickens (Soyer *et al.*, 2009). Nonsynonymous mutations were determined in the Chapterone_III domain in *pipC*, which is involved in T3SS and in delivering virulence effector proteins from *Salmonella* to host cells (Luo *et al.*, 2001). The genes under positive selection could be possible targets for mutational studies (Soyer *et al.*, 2009).

The three Asian strains in IIA may have different virulence attributes because they do not contain T6SS, which is a major component in SPI-6 (Jani and Cotter, 2010). *Salmonella* Gallinarum 287/91, *Salmonella* Virchow SL491, and *Salmonella* Paratyphi B SPB7 did not contain SPI-6 either (Blondel *et al.*, 2009). Thus, the gain or loss of SPI-6 has occurred independently in different serotypes. We could not determine whether SPI6-GII was introduced independently or if it replaced T6SS. Since SPI-6 was located next to *tRNA-aspartate* and contained Rhs family protein genes, both of which are associated with rearrangement or acquisition of new genetic elements (Hill, 1999; Pukatzki *et al.*, 2009), this location is likely to be a hot spot for recombination events. In the phylogenetic trees of SPI-6 and *saf*, the American strains in both lineages were clustered separately from the Asian strains, indicating that geographic location played an im-

portant role in the evolution and diversity of SPI-6. Based on the distribution of T6SS, *saf*, and *tcf* genes, the acquisitions of these clusters were independent events.

The findings in the current study distinguish *Salmonella* Newport lineages as well as MLST analyses (Cao *et al.*, 2013). Moreover, no *Salmonella* Newport lineage I strain has been sequenced to date. The lineage I strains may possess the lineage-specific SNPs in SPI-5 and SPI-6 because lineage I displayed a distant relationship with lineages II and III (Sangal *et al.*, 2010).

In addition, the *tcf* fimbrial operon, *tinR*, and *tioA* were only identified downstream of *sinR* in IIA, pepper_Vietnam, and squid_Vietnam (Supplementary Fig. S2). These genes were found downstream of SPI-6 in *Salmonella* Typhi, but not in *Salmonella* Typhimurium (Sabbagh *et al.*, 2010). Porwollik (Porwollik, 2011) reported that *Salmonella* with a broad host range always possess higher numbers of fimbrial operons than those with host restriction. Diversification of the fimbrial operon in *Salmonella* may contribute to virulence activities (Yue *et al.*, 2012; Allard *et al.*, 2013). Moreover, the typhoid-associated gene *tcfA* has been more common in nontyphoidal *Salmonella* than known, and it is expressed during *Salmonella* invasion activities (Suez *et al.*, 2013).

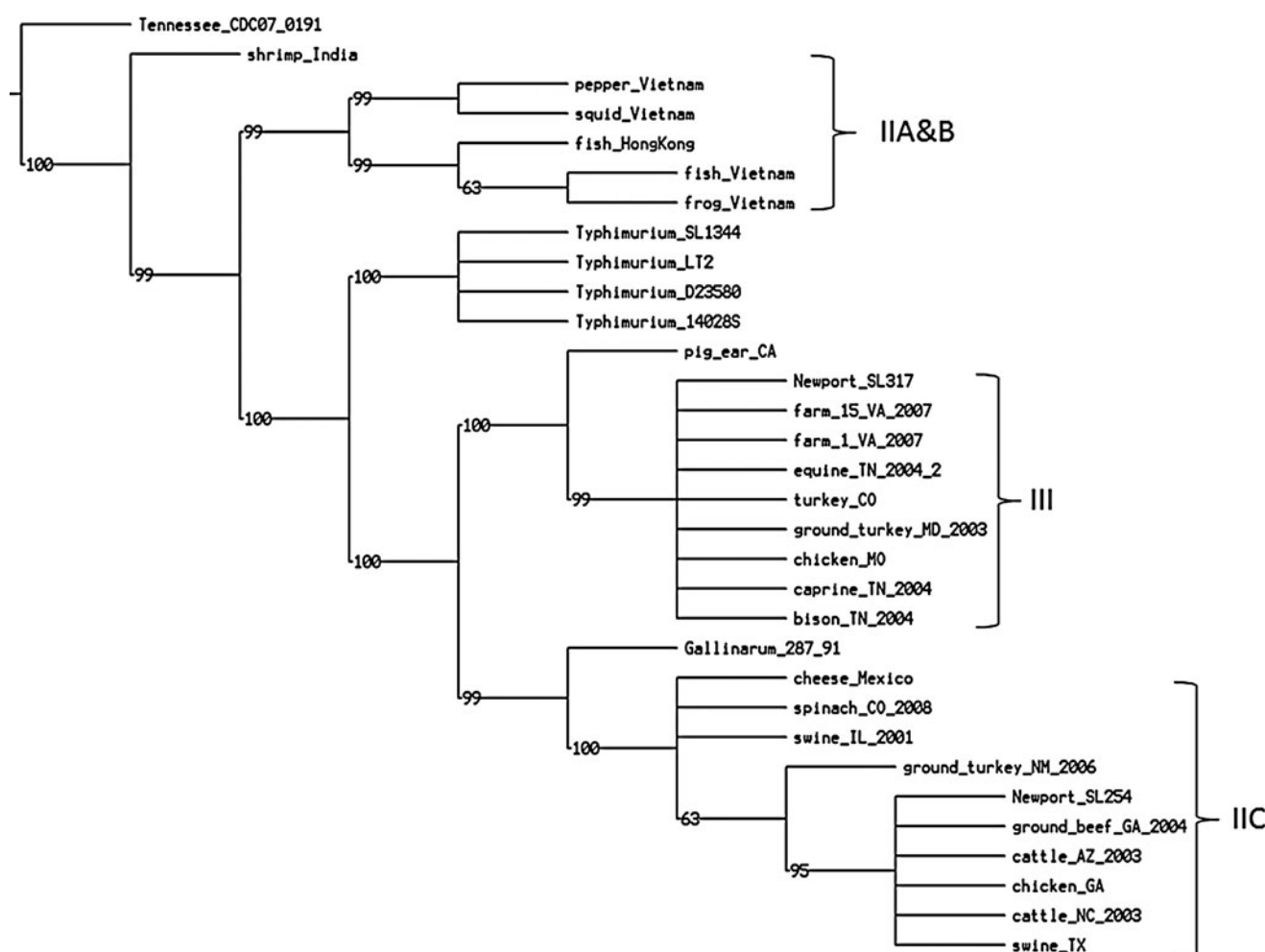


FIG. 4. Parsimony phylogenetic tree of *saf* gene cluster. There are 210 equally most parsimonious trees determined with a length of 493 SNP, consistency index of 0.840, and retention index of 0.970. The *saf* genes clustering reflect geographic origins. All Asian strains were clustered separately from the American strains.

Conclusions

SPI-5 and SPI-6 possess extensive differences in *Salmonella* Newport lineages II and III. SPI-5 contained both insertions and substitutions, including SPI5-GI1 and SPI5-GI2. SPI6-GI1 was present in the Asian strains of IIA. These genomic islands may contribute to virulence in their hosts. The SNPs in SPI-5 and SPI-6 could be used as biomarkers for rapid detection and epidemiological investigations to differentiate *Salmonella* Newport lineages II and III. The *tcp* genes may relate to host range and virulence activity in pepper_Vietnam and squid_Vietnam.

Acknowledgments

This work was supported in part by the Joint Institute for Food Safety and Applied Nutrition, University of Maryland.

Disclosure Statement

No competing financial interests exist.

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