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Pre-processing and analysis of genomic data using data mining tools

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Résumé

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Abstract

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1 Introduction

Tuberculosis (TB) is a highly infectious disease caused by the bacterium *Mycobacterium Tuberculosis*. It primarily affects the lungs but can also impact other parts of the body. TB is a major global health concern, with millions of new cases and deaths reported annually. The disease is transmitted through airborne droplets when an infected person coughs or sneezes.

The emergence of multidrug-resistant (MDR) and extensively drug-resistant (XDR) strains of *M. tuberculosis* poses a significant challenge to TB control efforts. These strains are resistant to the most effective anti-TB drugs, making treatment more difficult and increasing the risk of transmission. The World Health Organization (WHO) has identified the need for new diagnostic tools, treatments, and vaccines to combat TB, particularly in the context of drug resistance.

This project aims to find a way to identify and analyze the strain of *M. tuberculosis* responsible for a TB infection using genomic data. By analyzing the genetic sequences of *M. tuberculosis* strains, we hope to develop a method that can accurately determine the strain type, which is crucial for effective treatment and control of the disease. We explore different approaches to achieve this goal.

We first use multiple clustering techniques to group similar strains based on the presence or absence of certain proteins. Then, we converted the genomic data into images of various resolutions and used a convolutional neural network (CNN) to classify the images.

2 Related Work

Genomics is a field that has been growing rapidly in the past few years. The advent of high-throughput sequencing technologies has made it possible to sequence the entire genome of an organism in a matter of days. This has led to an explosion of data, with the number of sequenced genomes increasing exponentially. This has created a need for new tools and algorithms to analyze this data. In this chapter, we review some of the existing tools and algorithms for analyzing genomic data. We also discuss some of the techniques we used in our project.

2.1 Genomic data analysis

Random forests

Another method that has been used for genomic data analysis is random forests (RF) [2]. This method is based on the idea of ensemble learning, where multiple decision trees are trained on different subsets of the data and then combined to make a final prediction.

AI applications in genomic analysis

Many researchers have used AI techniques to analyze genomic data. For example, [1] reviews different AI techniques that have been used for genomic data analysis, including CNNs, autoencoders, etc.

More recently, the authors of [9] designed a tool based on multiple LLM backends for multi-omics analysis with minimal human intervention.

2.2 DBSCAN and HDBSCAN

DBSCAN

DBSCAN works by defining a neighborhood around each point based on a distance metric (usually Euclidean distance) and a minimum number of points required to form a dense region. The algorithm proceeds as follows:

1. For each point in the dataset, find its neighbors within a specified radius ϵ .
2. If the number of neighbors is greater than or equal to the minimum number of points, a new cluster is formed.
3. The algorithm recursively expands the cluster by including all points that are reachable from the initial point.
4. Points that do not belong to any cluster are classified as noise.

HDBSCAN

HDBSCAN builds on the principles of DBSCAN by introducing a hierarchical clustering approach. It is more robust to varying densities and can identify clusters of different shapes and sizes. The algorithm works as follows:

1. Compute the mutual reachability distance between points, which takes into account the distance to the nearest neighbor.
2. Build a minimum spanning tree (MST) based on the mutual reachability distances.
3. Extract clusters from the MST by varying the density threshold, resulting in a hierarchy of clusters.
4. Assign points to clusters based on their membership in the hierarchy, allowing for flexible clustering.

2.3 Introduction to CNNs

The fundamental idea behind **Convolutional Neural Networks (CNNs)** was introduced by Kunihiko Fukushima^[4] in 1980 and later popularized by Yann LeCun in the 1990s with the LeNet architecture^[5]. CNNs are a class of deep learning models specifically designed for processing structured grid data, such as images. They are particularly effective for tasks like image classification, object detection, and segmentation.

CNNs work by applying convolutional filters to the input data, which allows them to learn spatial hierarchies of features. It shares some similarities with the more conventional neural networks, but there is at least two different types of layers that are specific to CNNs:

- **Convolutional layers:** These layers apply convolutional filters to the input data, extracting local features that are invariant to translation. The filters slide over the input data, computing dot products and producing feature maps.
- **Pooling layers:** These layers downsample the feature maps, reducing their spatial dimensions while retaining important information. Common pooling operations include max pooling and average pooling.

The architecture of a CNN typically consists of multiple convolutional and pooling layers followed by fully connected layers. The convolutional layers learn to extract features from the input data, while the fully connected layers perform the final classification or regression task. The training process involves optimizing the weights of the filters and fully connected layers using backpropagation and a loss function, such as cross-entropy for classification tasks.

2.4 K-fold cross validation

K-fold cross validation is a specific type of cross validation where the dataset is divided into k equally sized folds. The model is trained on $k - 1$ folds and tested on the remaining fold, and this process is repeated for each fold. The final performance is averaged over all folds to obtain a more reliable estimate of the model's performance. The graphical representation of K-fold cross validation is shown in Figure 1.

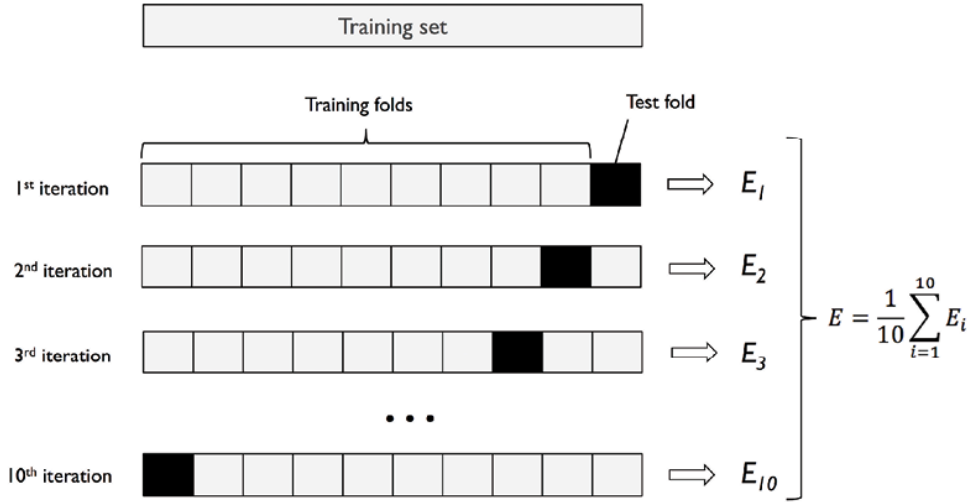


Figure 1: How K-fold cross validation works.^[6]

2.4.0.1 Stratified K-fold cross validation

Stratified K-fold cross validation is a variation of K-fold cross validation that ensures that each fold has the same proportion of samples from each class. This is particularly useful for imbalanced datasets, as it ensures that each fold has a representative sample of each class. Even if this method is recommended for imbalanced datasets, we did not have enough time to implement it, so we used the standard K-fold cross validation.

2.5 SMOTE

Data augmentation is a technique used to artificially increase the size of a dataset by applying various transformations to the existing data. This can help improve the model's performance by providing more diverse training samples and reducing overfitting. In our project, we used SMOTE (Synthetic Minority Over-sampling Technique) to generate synthetic samples for the minority classes in our dataset.

SMOTE works by creating new samples by interpolating between existing samples in the feature space. It generates synthetic samples by selecting a minority class sample, finding its nearest neighbors, and creating new samples by interpolating between the selected sample and its neighbors. This helps to balance the dataset by increasing the number of samples in the minority classes, which can improve the model's performance on those classes.

New samples are generated by taking a random sample from the minority class and finding its nearest neighbors in the feature space. The new sample is then created by interpolating between the selected sample and its neighbors, as shown in Figure 2. It uses the following formula:

$$x_{new} = x_i + \lambda(x_j - x_i) \quad (1)$$

Where x_{new} is the new sample, x_i is the selected sample, x_j is one of its nearest neighbors, and λ is a random number between 0 and 1. This process is repeated until the desired number of samples is generated for the minority class.

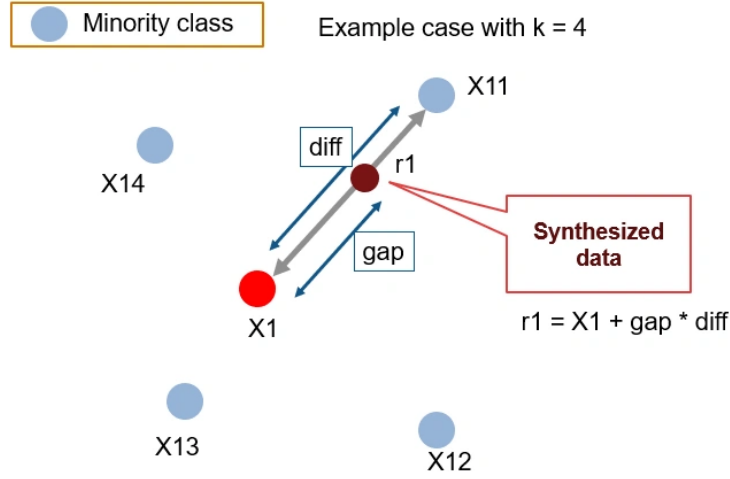


Figure 2: Illustration of the SMOTE algorithm.^[7]

3 Context

3.1 Work Environment

This project was done within the framework of the Master 2 Computer Science program at the **Laboratoire de Mathématiques Informatique et Applications (LAMIA)** of the **Université des Antilles**. The following tools and technologies were used to develop the algorithms and document the project:

- **Python**: a programming language widely used in scientific computing, data analysis, and machine learning.
- **GitHub**: a version control platform for managing the source code.
- **GitHub Copilot**: for code completion.
- **Jupyter Notebook**: an interactive environment for writing and executing Python code, ideal for data analysis and visualization.
- **Sklearn**: a Python library for machine learning that provides various algorithms and tools for data preprocessing, model selection, and evaluation.
- **Pandas**: a Python library for data manipulation and analysis, providing data structures and functions for working with structured data.
- **Matplotlib**: a Python library for graphics and data visualization, including 3D plotting capabilities.
- **L^AT_EX**: a typesetting system used for writing technical and scientific documents.
- A personal computer running Windows 11 with a **Ryzen 7 6800H** processor and **32 GB** of RAM.

3.2 Objectives

The main objective of this project is to explore and analyze genomic data, specifically the genomes of Tuberculosis (TB) strains, to identify patterns and relationships between different strains. As well as to apply various data mining and data analysis techniques, but also develop new methods and compare their performance to existing methods.

4 Genomic data

Genomic data refers to the information contained in the DNA of an organism, which includes the sequences of nucleotides (Adenine, Thymine, Cytosine, and Guanine). These are gathered through various methods, such as DNA sequencing, and can be used to study the genetic makeup of organisms, identify genetic variations, and understand the relationships between different species. The Institute Pasteur has a large collection of genomic data, including the genomes of various strains of Tuberculosis (TB), which is a major public health concern worldwide.

4.1 The FASTA format

The FASTA format is a text-based format for representing nucleotide or protein sequences [3, 8]. It is widely used in bioinformatics to store and exchange sequence data. A FASTA file consists of one or more sequences, each represented by a header line starting with a greater-than symbol (>), followed by the sequence itself on the next line. The sequence codes for a protein who then codes for a gene. The header line can contain additional information about the sequence, such as its identifier, description, or source. An example of a FASTA file is shown in Figure 3.

```
>sequence_id
ATCGATCGATCGATCGATCGATCG
>sequence_id_2
GCTAGCTAGCTAGCTAGCTAGCTA
>sequence_id_3
TAGCTAGCTAGCTAGCTAGCTAGC
>sequence_id_4
AGCTAGCTAGCTAGCTAGCTAGCT
>sequence_id_5
CATCGATCGATCGATCGATCGATC
```

Figure 3: Example of a FASTA file containing two sequences.

The FASTA format is simple and easy to read, making it suitable for storing large amounts of sequence data. It is also compatible with many bioinformatics tools and software, allowing for easy analysis and manipulation of sequence data.

5 Clustering genomic data

5.1 Preprocessing Genomic Data

Our data is a collection of Tuberculosis (TB) genomes, where each file contains the proteins and their respective nucleotide sequences in FASTA format, for a total of 7057 files. These genomes were obtained from the **Institut Pasteur** and are part of the **Mycobacterium tuberculosis complex** (MTBC), which is a group of closely related bacteria that cause TB. The goal is to cluster these genomes based on their protein sequences and find ways to distinguish between different TB strains.

We started by creating a dataset containing the number of times each protein appears in each genome. This was done by parsing the FASTA files and counting the occurrences of each protein sequence, we then removed the outliers, namely "hypothetical proteins" and "putative proteins", which are not well characterized and do not provide useful information for clustering. The resulting dataset is a CSV file of 7057 columns plus 1 column for the protein names and 2732 rows plus 1 row for the header.

Considering the size of the dataset, we used Principal Component Analysis (PCA) set to 95% variance to reduce the dimensionality of the data while preserving as much variance as possible. PCA is a technique that transforms the data into a new coordinate system, where the first principal component captures the maximum variance, the second captures the second maximum variance, and so on.

We then calculated the silhouette score for different values of k , ranging from 2 to 10, to determine the optimal number of clusters. The silhouette score was calculated using the 'silhouette_score' function from the 'sklearn.metrics' module, which takes the data and the cluster labels as input. The optimal number of clusters is the one that maximizes the silhouette score.

5.1.1 First Results

The initial results showed that the silhouette score was highest for $k = 9$, indicating that this was the optimal number of clusters. However, the clusters were not well separated, or some points are alone in their cluster, as seen in Figure 4. Which suggests that the data may not be well suited for clustering with K-means, or that the features used for clustering are not informative enough. We also tried without applying PCA but achieved similar results, we also observed fewer clusters, as seen in Figure 6.

After reviewing the data, we found that there is significant overlap between the points, so we switched to 3D visualization to better understand the clusters. With Figure 5 and Figure 7, we can see that the clusters are still not well separated, but we can conclude that a 2D space was not appropriate for visualizing the clusters.

These results suggest that K-means clustering may not be the best approach for this dataset, or that the data may need further or a different preprocessing step to improve the clustering results. We will explore alternative clustering algorithms in the next section.

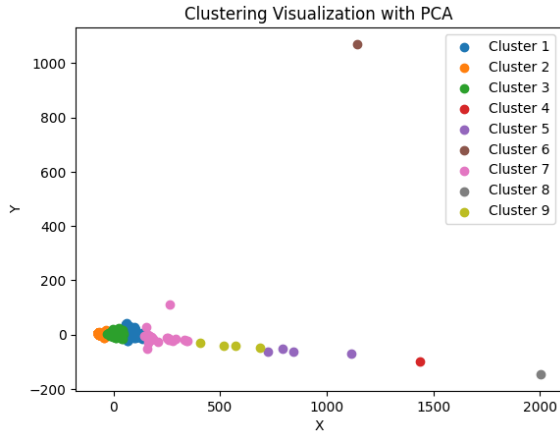


Figure 4: Clusters obtained with K-means clustering after applying PCA.

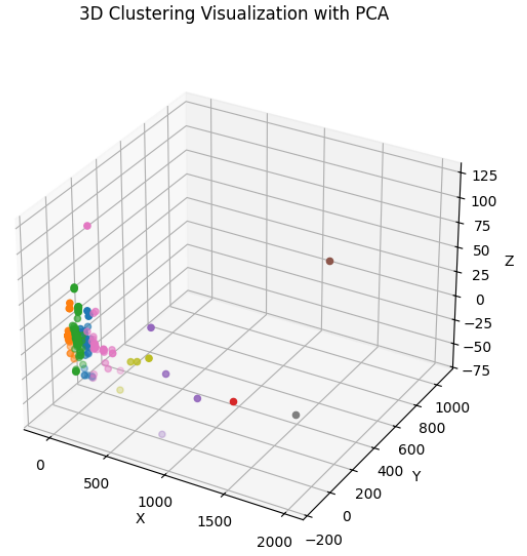


Figure 5: Clusters obtained with K-means clustering after applying PCA in 3D.

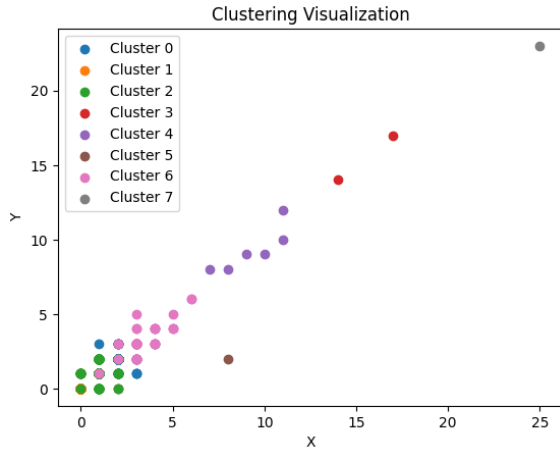


Figure 6: Clusters obtained with K-means clustering without applying PCA.

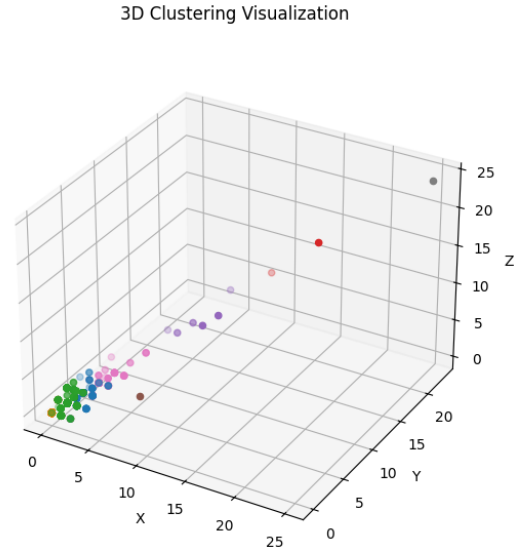


Figure 7: Clusters obtained with K-means clustering without applying PCA in 3D.

5.2 New Approach using DBSCAN and HDBSCAN

To address the poor clustering results obtained with K-means, we explored alternative clustering algorithms, specifically **DBSCAN** and **HDBSCAN**.

DBSCAN (Density-Based Spatial Clustering of Applications with Noise) is a density-based clustering algorithm that groups points that are close together based on a distance measurement and a minimum number of points. It is particularly effective for datasets with varying densities and can identify noise points that do not belong to any cluster.

HDBSCAN (Hierarchical Density-Based Spatial Clustering of Applications with Noise) is an extension of DBSCAN that builds a hierarchy of clusters and allows for more flexible clustering by varying the density threshold. It can also handle varying cluster shapes and sizes.

5.2.1 Results

During our testing, we applied both DBSCAN and HDBSCAN with an epsilon value of 1 and a minimum number of samples of 5. These techniques were both applied to the dataset with and without PCA. We can see from Figure 20 to Figure 11 that the clusters obtained with DBSCAN and HDBSCAN are more just a bit more discernible than those obtained with K-means clustering. But the clusters are still closely packed together, we can conclude that the problem lies with the way the dataset was created. A similar observation can be made when not applying PCA, as shown in Figure 20 to Figure 23 in the appendix.

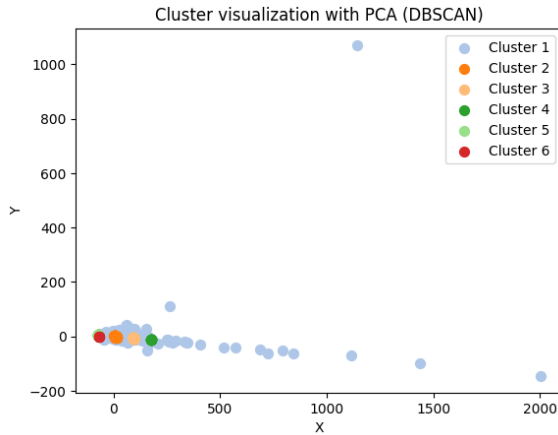


Figure 8: Clusters obtained with DBSCAN after applying PCA.

3D DBSCAN Clustering Visualization with PCA

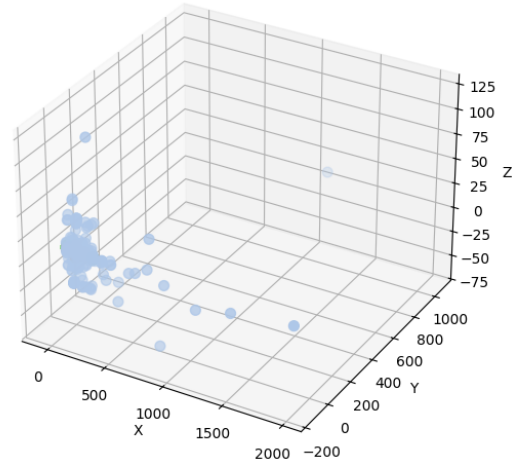


Figure 9: Clusters obtained with DBSCAN after applying PCA in 3D.

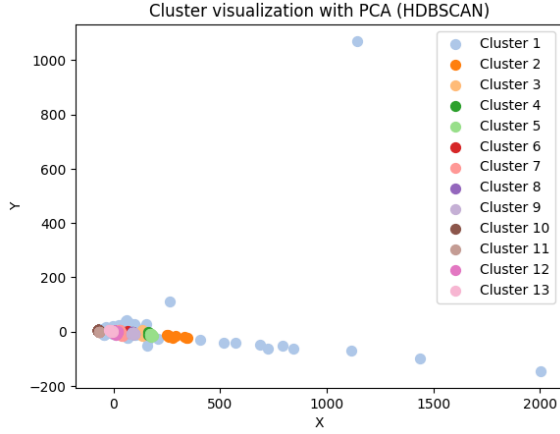


Figure 10: Clusters obtained with DBSCAN after applying PCA.

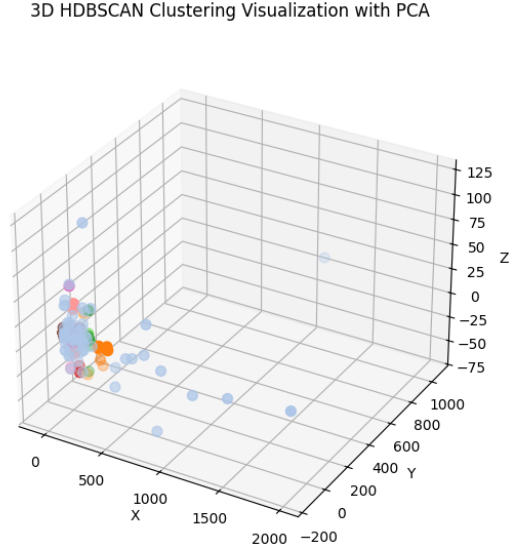


Figure 11: Clusters obtained with DBSCAN after applying PCA in 3D.

5.3 Trying an autoencoder

To further explore the clustering of the genomic data, we decided to try an autoencoder. After getting the embeddings from the autoencoder, we applied the K-means clustering algorithm to the encoded data. We can see in Figure 12 and Figure 13 that the clusters obtained with K-means clustering on the encoded data are more discernible than those obtained with K-means clustering on the original data. However, we observe a great intra-cluster distance, which suggests that the autoencoder may not have learned a good representation of the data, that the data is not well suited, or that the features used for clustering are not informative enough.

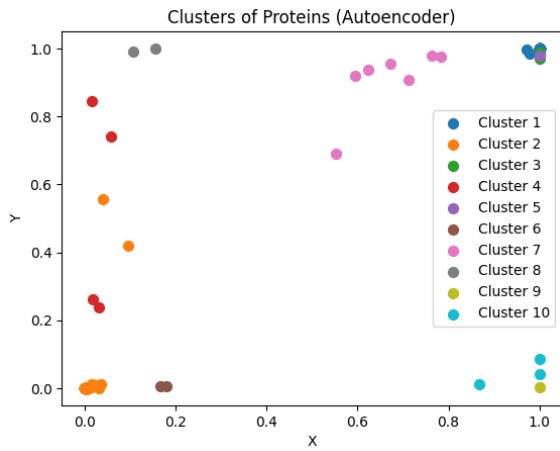


Figure 12: Clusters obtained with K-means clustering on the encoded data.

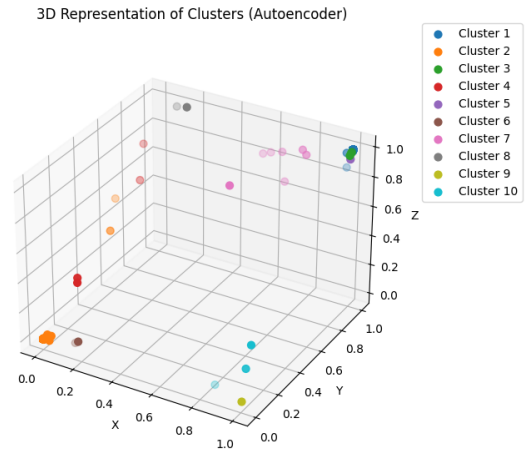


Figure 13: Clusters obtained with K-means clustering on the encoded data in 3D.

6 Using CNNs to identify TB Strains

The clustering approach did not yield satisfactory results, so we decided to explore a different approach by converting the genomic data into images. This approach allows us to leverage the power of Convolutional Neural Networks (CNNs) to classify the images and identify the TB strains. These images are generated from the genomic data by applying various metrics to the nucleotide sequences, which are then converted into pixel values. The resulting images can be used to train a CNN to classify the TB strains based on their genomic data. This approach has the potential to improve the classification accuracy and provide a more robust method for identifying TB strains.

6.1 Building the dataset

For this experiment, each genome was converted into an image where each pixel is determined by a combination of three metrics of following metrics:

- **Chargaff** index: provides the fraction of nucleic bases G and C of the sequence. The formula is:

$$100 \times \frac{Gcount + Ccount}{Sequence_size} \quad (2)$$

- **Component** index: evaluates the distribution of the bases in a sequence by comparing the number of each base A and T to the expected frequency of those bases in a balanced distribution. The formula is:

$$\frac{(Acount + Tcount) + \frac{Sequence_size}{2}}{\frac{Sequence_size}{2}} \quad (3)$$

- **Diversity** index: measures the diversity of bases in a sequence by comparing the number of different subsequences of a given size to the expected frequency of those subsequences in a balanced distribution. The formula is:

$$\frac{Number_{unique\ subsequences}}{Expected_{subsequences\ frequency}} \quad (4)$$

- **Skew** index: measures the asymmetry of the distribution of nucleobases in a sequence. The formula is:

$$\frac{(Gcount - Ccount)}{Sequence_size} \quad (5)$$

- **Nuclescore** index: empirical measure that combines multiple metrics from a genome sequence to synthesize the nucleotide information of a genome in order to differentiate species based only on their sequences. The formula is:

$$\log_2\left(\frac{Variance \times GC_{ratio} \times AT_{count}/GC_{ratio}}{\sqrt{Sequence_size}}\right) \quad (6)$$

We get ten combinations of these metrics, each represented by a different color channel in the image after being normalized to the range $[0, 255]$. This conversion is applied at different resolutions, from 4px (2x2) to 100px (10x10), but for the training, all images were resized to a 32x32px resolution. The resulting images put into their corresponding class based on the strain they belong to, as shown in Figure 14. We have the following classes:

- **M** with 185 samples.
- **East Asian** with 1651 samples.
- **East-African Indian** with 267 samples.
- **Euro-American** with 4409 samples.
- **Indo-Oceanic** with 259 samples.

We also made mosaic images by combining the corresponding images from each combination of metrics resulting in a 5x2 grid rectangular image as shown in Figure 15. We chose to a 5x2 grid because it creates complete images without any empty pixels, which is important for the training of the CNN. The images are also resized to a 20x50px resolution for training.

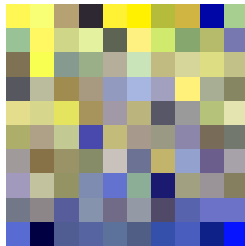


Figure 14: Example of an image generated from the genome of a TB strain using the Chargaff, Component and Diversity metrics at 100px resolution.

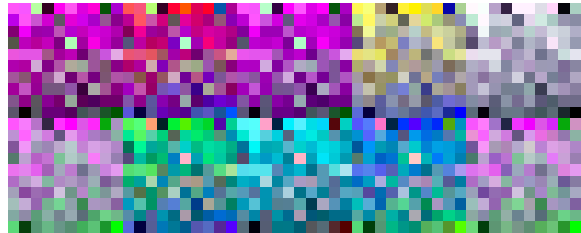


Figure 15: Example of a mosaic image obtained from the individual images at 100px resolution.

6.1.1 Initial testing

The resulting dataset is highly imbalanced, with some classes having significantly more samples than others. This can lead to biased models that perform poorly on minority classes. We decided to train our CNN on the dataset as-is, without any preprocessing or balancing techniques, to see how it performs on the imbalanced dataset. For the sake of brevity, we will only present the results for the combination of Chargaff, Composition and Diversity metrics at 100px resolution, for the mosaic images we will present the results for the 100px resolution as well.

6.1.1.1 Results

As shown in Figure 16, the 4px images yielded the worst performance, with a validation accuracy between 66% and 76%. We then observe a significant improvement in performance as the resolution increases with the accuracy starting to stabilize around 92% and 94% starting from the 36px resolution. Figure 17 and Table 1 show that the overall performances of the model are good, except for the East-African Indian and Indo-Oceanic classes with more moderate performances.

East-African Indian shows a moderate recall of 0.779 and a precision of 0.904, indicating that the model is able to correctly classify most of the samples in this class, but it also misclassifies some samples as other classes. The Indo-Oceanic class has a recall of 0.988, indicating that the model is able to correctly classify most of the samples in this class, but it has a lower precision of 0.718, indicating that the model misclassifies some samples as this class.

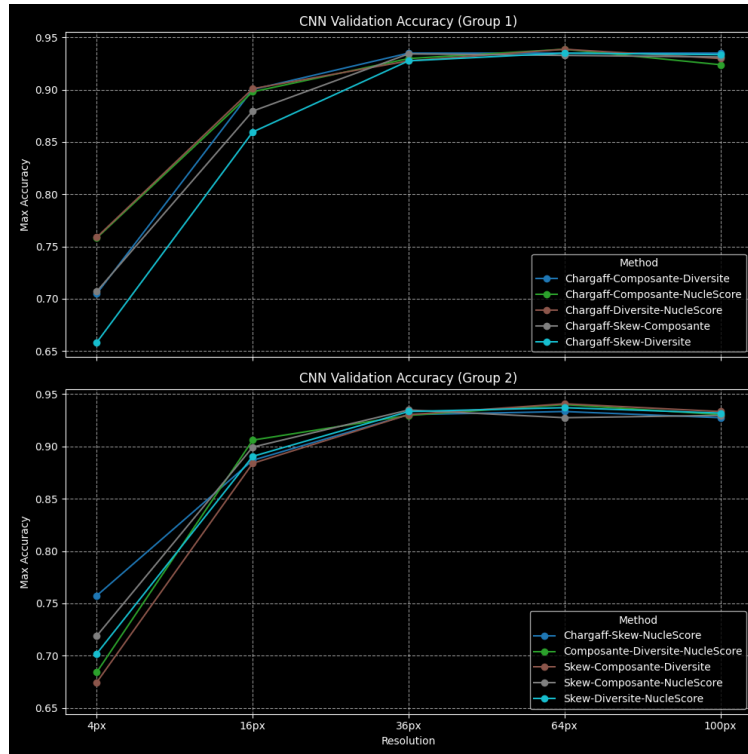


Figure 16: Graph showing the validation accuracy of the model on the different methods at 100px resolution without any preprocessing or balancing techniques applied.

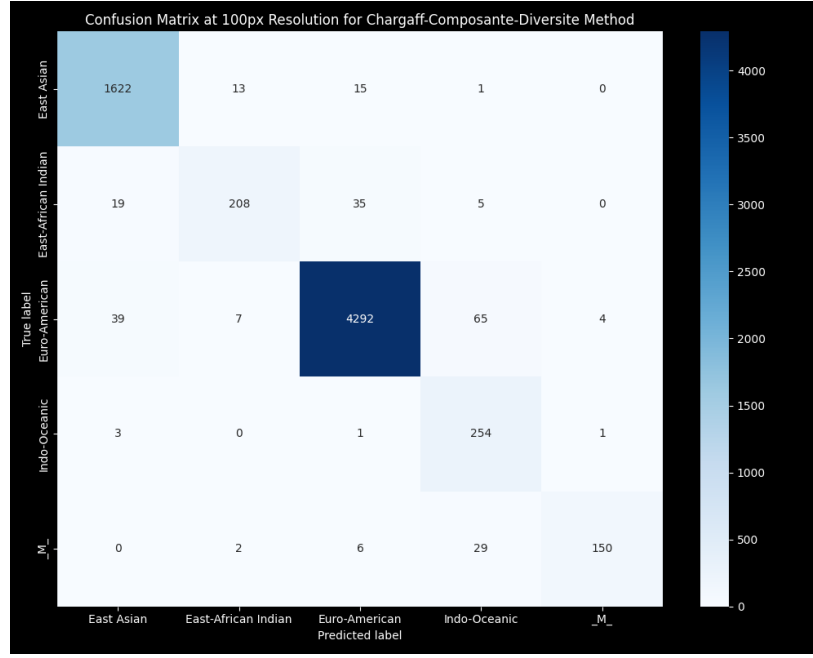


Figure 17: Confusion matrix of the model on the images obtained from the Chargaff-Component-Diversity combination at 100px resolution without any preprocessing or balancing techniques applied.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.965	0.982	0.988	0.987
East-African Indian	0.904	0.779	0.997	0.988
Euro-American	0.987	0.974	0.976	0.975
Indo-Oceanic	0.718	0.988	0.985	0.985
M	0.968	0.802	0.999	0.994

Table 1: Performance metrics of the model on the images obtained from the Chargaff-Component-Diversity combination at 100px resolution without any preprocessing or balancing techniques applied.

For the mosaic images, we observed similar results, with the model achieving a validation accuracy of around 95% at 100px resolution. While most of the metrics are similar to the individual images, the East-African Indian class improved its recall, the precision dropped to 0.662. This indicates that, while the model is able to correctly classify most of the samples in this class, other classes are confused with this class.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.976	0.970	0.992	0.987
East-African Indian	0.662	0.858	0.982	0.977
Euro-American	0.991	0.963	0.984	0.970
Indo-Oceanic	0.771	0.973	0.988	0.988
M	0.966	0.908	0.999	0.997

Table 2: Performance metrics of the model on the mosaic images at 100px resolution without any preprocessing or balancing techniques applied.

Overall, this model shows good performance on the imbalanced dataset, but there is still room for improvement. But we will explore different techniques to address the class imbalance issue in the following sections.

6.2 Mitigating class imbalance

We have multiple ways of addressing the class imbalance issue in our dataset, including:

- Cross validation
- Under-sampling
- Data augmentation
- Combining under-sampling and data augmentation

We will explore each of these techniques in the following sections and compare the results to see which one yields the best performance.

6.2.1 Cross validation

Cross validation is a technique used to evaluate the performance of a model by splitting the dataset into multiple subsets, or folds. The model is trained on a subset of the data and tested on the remaining data, and this process is repeated for each fold. This allows for a more robust evaluation of the model's performance, as it reduces the risk of overfitting to a specific subset of the data.

Remaking the dataset

During our testing, we realized that we were testing our model with images the model was trained on, which is not a good practice. We decided to split our dataset into a training set and a test set, with 90% of the samples in the training set and 10% in the test set. The files are separated into two folders to avoid any overlap between the training and test sets.

6.2.1.1 Results

During our testing, we used the K-fold cross validation technique with 5 folds with 15 epochs and a batch size of 32. We observed that the model was able to achieve a validation accuracy of around 96% during validation for all of 10 combinations of the metrics at 100px resolution, as shown in Figure 18. The confusion matrix in Figure 19 shows that the model was able to correctly classify most of the samples, with only a few misclassifications.

With Table 3 we can see that the model has good performance across most classes. Surprisingly enough, the M class, which has the least number of samples, has better performance than the East-African Indian class, which has more samples. This suggests that the model is able to generalize well to the minority classes, even with the class imbalance issue.

For the mosaic images, we applied the same k-fold cross validation technique with 5 folds and 15 epochs, and we observed similar results compared to the individual images. The East-African Indian class has a slightly lower precision but a slightly higher recall, indicating that the model is able to correctly classify most of the samples in this class, but it also misclassifies some samples as other classes. The Indo-Oceanic class has a recall of 1, indicating that the model was able to correctly classify all the samples in this class, but it has a lower precision of 0.684, indicating that the model misclassifies some samples as this class. Specificity and accuracy are nearly the same as the individual images, as shown in Table 4.

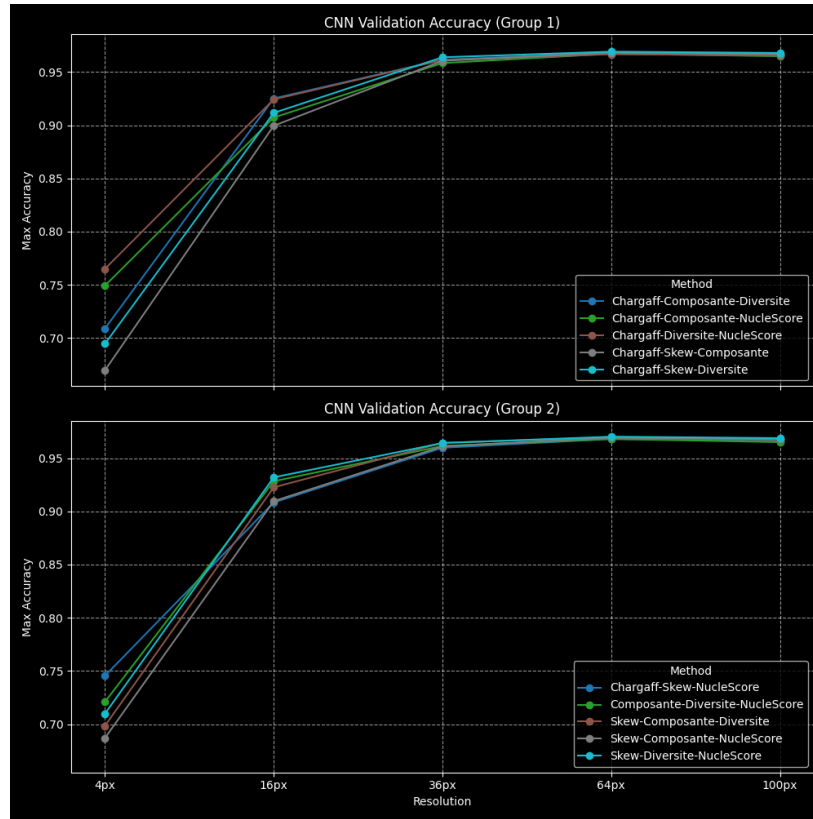


Figure 18: Graph showing the validation accuracy of the CNN model with k-fold cross validation applied on the different methods.

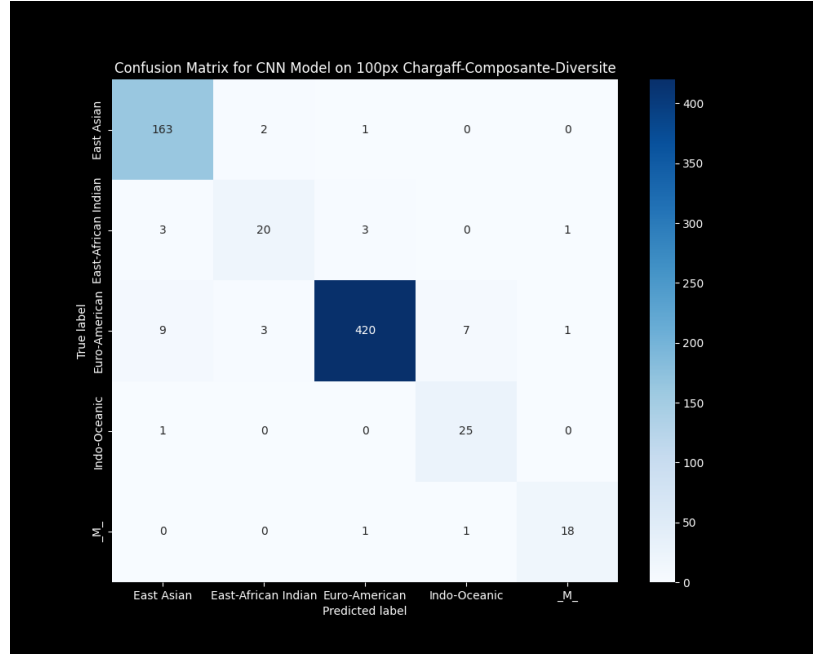


Figure 19: Confusion matrix of the CNN model with k-fold cross validation applied on the images obtained from the Chargaff-Component-Diversity combination at 100px resolution.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.926	0.982	0.975	0.976
East-African Indian	0.800	0.741	0.992	0.982
Euro-American	0.988	0.955	0.979	0.963
Indo-Oceanic	0.758	0.962	0.988	0.987
M	0.900	0.900	0.997	0.994

Table 3: Performance metrics with k-fold cross validation applied on the images obtained from the Chargaff-Component-Diversity combination at 100px resolution.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.932	0.988	0.977	0.979
East-African Indian	0.724	0.778	0.988	0.979
Euro-American	0.995	0.945	0.992	0.962
Indo-Oceanic	0.684	1.000	0.982	0.982
M	0.944	0.850	0.998	0.994

Table 4: Performance metrics with k-fold cross validation applied on the mosaic images at 100px resolution.

The performances of the model with cross validation are close to the ones obtained without cross validation. It seems to perform slightly better, but the difference is not significant enough to conclude that cross validation improves the performance of the model. However, it is important to note that cross validation is a good practice to evaluate the performance of a model, as it reduces the risk of overfitting to a specific subset of the data and provides a more robust evaluation of the model's performance.

6.2.2 Under-sampling

Under-sampling is a technique used to address the class imbalance issue by reducing the number of samples in the majority classes. In our application, we decided to reduce the number of samples in the Euro-American and East Asian classes to a total of 1000 samples each, while keeping the other classes unchanged. This was done to balance the dataset and ensure that the model is not biased towards the majority classes.

Results

This time we observe a drop in the validation accuracy of the model (Figure 28) compared to the previous results, with the accuracy being between 89% and 93% for the 100px resolution. There is also a significant drop in the performance of the model on the East-African Indian class, while the recall slightly improves, the precision drops significantly, indicating that the model identify other classes as East-African Indian. This is likely due to the reduced number of samples in the Euro-American and East Asian classes, which may have led to a less representative training set for these classes. It can be seen in Figure 29 and Table 5.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.964	0.982	0.988	0.987
East-African Indian	0.571	0.889	0.972	0.969
Euro-American	0.998	0.936	0.996	0.957
Indo-Oceanic	0.765	1.000	0.988	0.988
M	0.905	0.950	0.997	0.996

Table 5: Performance metrics with under-sampling applied on the images obtained from the Chargaff-Component-Diversity combinations at 100px resolution.

For the mosaic images, we observe an even more significant drop in the precision of the East-African Indian class, which drops to 0.361 from 0.724 with cross validation on these images (Figure 6). The model misclassifies a lot of Euro-American samples as East-African Indian, more than there are actual East-African Indian samples in the dataset (Figure 31).

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.952	0.952	0.984	0.976
East-African Indian	0.361	0.815	0.940	0.935
Euro-American	0.992	0.893	0.987	0.926
Indo-Oceanic	0.765	1.000	0.988	0.988
M	0.864	0.950	0.995	0.994

Table 6: Performance metrics with under-sampling applied on the mosaic images at 100px resolution.

Instead of improving the performance of the model like we hoped, under-sampling led to a drop in the performance of the model, especially on the East-African Indian class. Still, even with the lowest number of samples, the M class keeps stable metrics. We're still unsure why this is the case, but it could be due to the fact that the M class has a very distinct pattern that the model can pick up on, even with a reduced number of samples.

6.2.3 Data augmentation

For this test, we use the SMOTE-generated images to increase the number of samples in the minority classes, East-African Indian, Indo-Oceanic and M, the other classes are kept unchanged. The aforementioned classes are augmented so that the resulting number of samples contains 25% of generated images for each class.

Results

With data augmentation, we once again observe similar behavior for the East-African Indian and Indo-Oceanic classes between the individual and mosaic images. The individual images seem to yield a better precision metric for the Indo-Oceanic class (Table 7), while the mosaic images favors the East-African Indian class (Table 8) in that regard. But this time, those two classes are roughly on par with each other, with metrics that are close, if not slightly better than with cross validation. This technique seems to be promising, as it allows us to increase the number of samples in the minority classes while keeping the majority classes unchanged.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.976	0.976	0.992	0.988
East-African Indian	0.759	0.815	0.989	0.982
Euro-American	0.986	0.968	0.975	0.970
Indo-Oceanic	0.812	1.000	0.991	0.991
M	0.950	0.950	0.998	0.997

Table 7: Performance metrics the images obtained from the Chargaff-Component-Diversity combinations at 100px resolution with data augmentation applied on the training set.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.970	0.976	0.990	0.987
East-African Indian	0.815	0.815	0.992	0.985
Euro-American	0.993	0.970	0.987	0.976
Indo-Oceanic	0.722	1.000	0.985	0.985
M	0.947	0.900	0.998	0.996

Table 8: Performance metrics the mosaic images at 100px resolution with data augmentation applied on the training set.

6.2.4 Combining techniques

Results

This time, we combined the under-sampling and data augmentation techniques hoping to get the best results. But, while the results for the individual images are similar to the ones obtained from the previous techniques (Table 9), the mosaic images show a significant drop in the performance of the model (Table 10). The East-African Indian class has a precision of 0.307, which is the worst performance we have seen so far. Once again, the model misclassified a lot of Euro-American samples as East-African Indian, which may be a side effect of the combination.

Earlier, we mentioned that we would only present the results for images at 100px resolution for the Chargaff-Component-Diversity combination, but it's worth mentioning that, for the individual images, we observed a disparity in validation accuracy between the different combinations. The accuracy ranges from 75% to 82% at 100px resolution, they are also the lowest accuracy observed for the individual images as shown in Figure 36.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.953	0.970	0.984	0.981
East-African Indian	0.706	0.889	0.985	0.981
Euro-American	0.995	0.952	0.992	0.966
Indo-Oceanic	0.743	1.000	0.986	0.987
M	0.950	0.950	0.998	0.997

Table 9: Performance metrics with combined under-sampling and data augmentation techniques applied on the images obtained from the Chargaff-Component-Diversity combinations at 100px resolution.

Class	Precision	Recall	Specificity	Accuracy
East Asian	0.968	0.922	0.990	0.973
East-African Indian	0.307	1.000	0.906	0.910
Euro-American	0.995	0.864	0.992	0.909
Indo-Oceanic	0.839	1.000	0.992	0.993
M	0.950	0.950	0.998	0.997

Table 10: Performance metrics with combined under-sampling and data augmentation techniques applied on the mosaic images at 100px resolution.

6.3 Comparing the results

We first ran the CNN model on the dataset without any preprocessing or balancing techniques applied, and we observed that the model was able to achieve a validation accuracy of between 92% and 94% at 100px resolution. However, this model was trained and tested on the same dataset, which means that the model may be biased. The dataset was also highly imbalanced, with some classes having significantly more samples than others.

After splitting the dataset into a training set and a test set, we applied k-fold cross validation which yielded decent results. But the East-African Indian and Indo-Oceanic classes had lower precision and recall metrics compared to the other classes, except for the Indo-Oceanic class which had a recall of 1 with the mosaic images. Then we tried under-sampling the dataset, which led to a drop in the performance for both individual and mosaic images. Data augmentation was then applied to the dataset, which allowed us to increase the number of samples in the minority classes. This led to better performance for the Indo-Oceanic class with the individual images, while the East-African Indian class had a better performance with the mosaic images. Finally, we combined under-sampling and data augmentation techniques, which led to a drop in the performance of the model for the mosaic images, while the individual images remained stable.

Overall, the best results were obtained with SMOTE data augmentation, which allowed us to increase the number of samples in the minority classes while keeping the majority classes unchanged.

7 Conclusion

A Presenting the Institut Pasteur and LAMIA

A.1 The Institut Pasteur

The Institut Pasteur is a renowned research institute located in Paris, France, dedicated to the study of biology, microorganisms, diseases, and vaccines. Founded in 1887 by Louis Pasteur, the institute has played a pivotal role in the development of modern microbiology and immunology. It is known for its contributions to the understanding of infectious diseases and the development of vaccines, including those for rabies and tuberculosis. Founded by Louis Pasteur in 1887, the Institut Pasteur has been at the forefront of scientific research and innovation for over a century.

A.2 Laboratoire de Mathématiques Informatique et Applications (LAMIA)

Laboratoire de Mathématiques Informatique et Applications (LAMIA) is a research laboratory part of Université des Antilles and is located in Guadeloupe. It focuses on the intersection of mathematics, computer science, and their applications in various fields, including data science, machine learning, and artificial intelligence. LAMIA conducts research that aims to advance theoretical understanding and practical applications of mathematical and computational methods.

B Additional Figures

B.1 DBSCAN and HDBSCAN Clustering

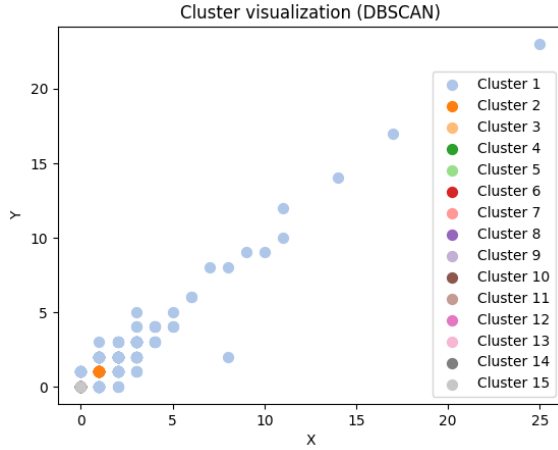


Figure 20: Clusters obtained with DBSCAN after applying PCA.

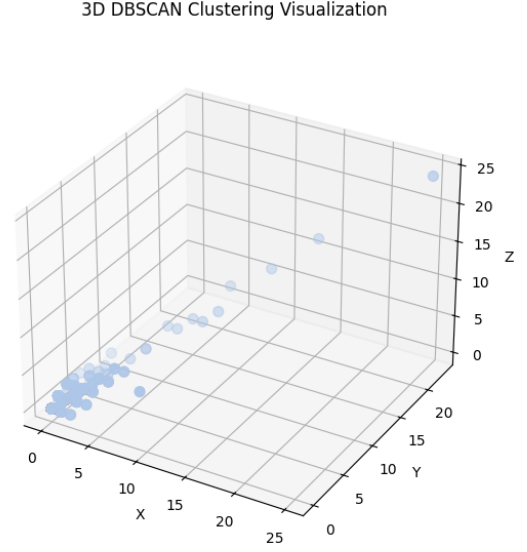


Figure 21: Clusters obtained with DBSCAN after applying PCA in 3D.

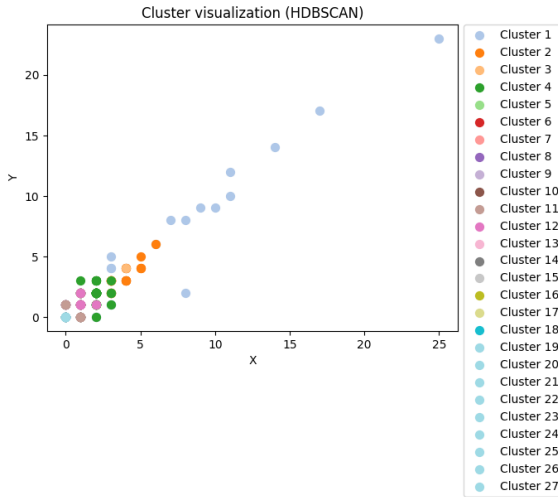


Figure 22: Clusters obtained with DBSCAN after applying PCA.

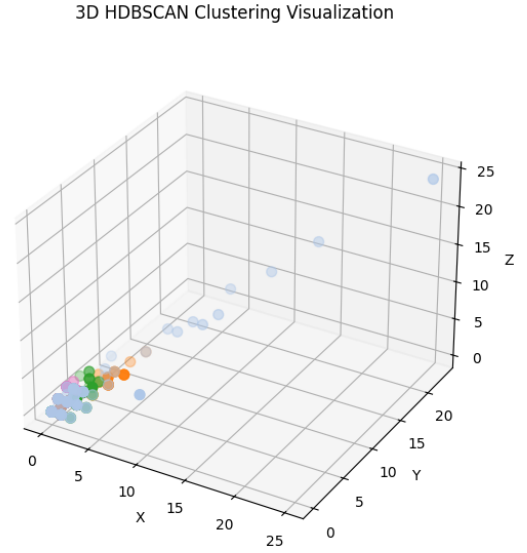


Figure 23: Clusters obtained with DBSCAN after applying PCA in 3D.

B.2 Initial CNN Model Performance

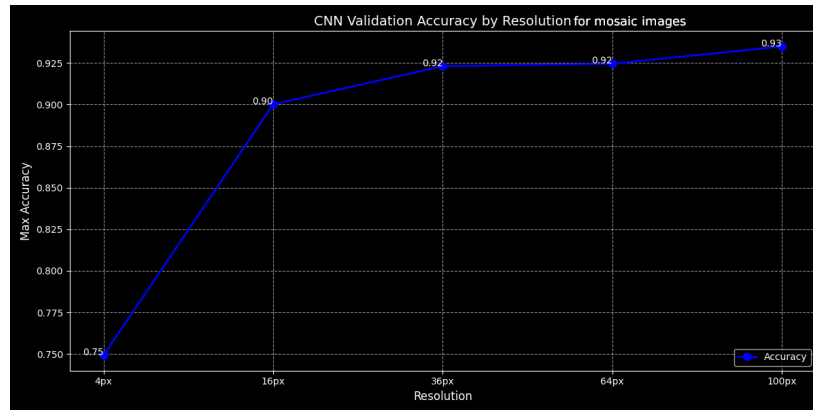


Figure 24: Graph showing the validation accuracy of the CNN model on the mosaic images at 100px resolution without any additional techniques applied.

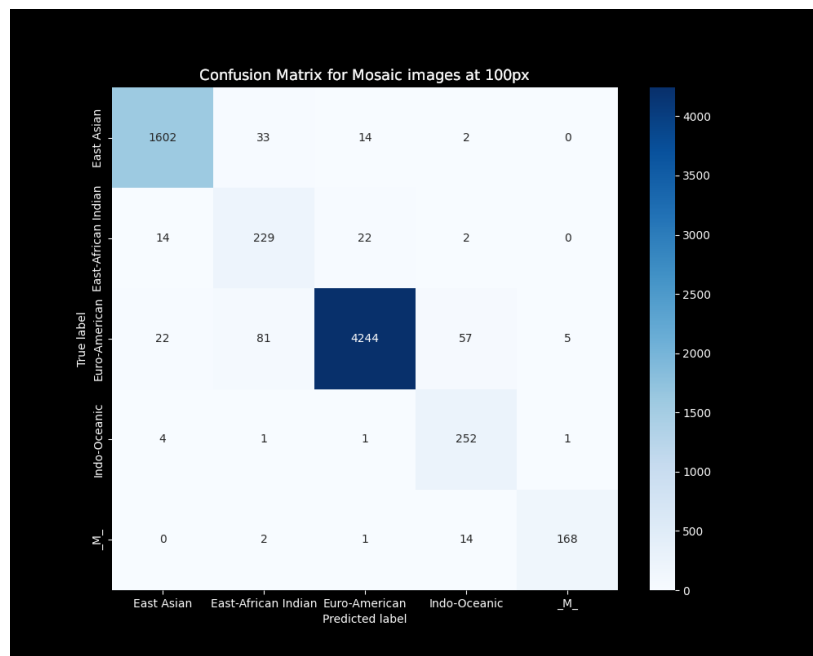


Figure 25: Confusion matrix on the mosaic images at 100px resolution without any additional techniques applied.

B.3 K-Fold Cross Validation

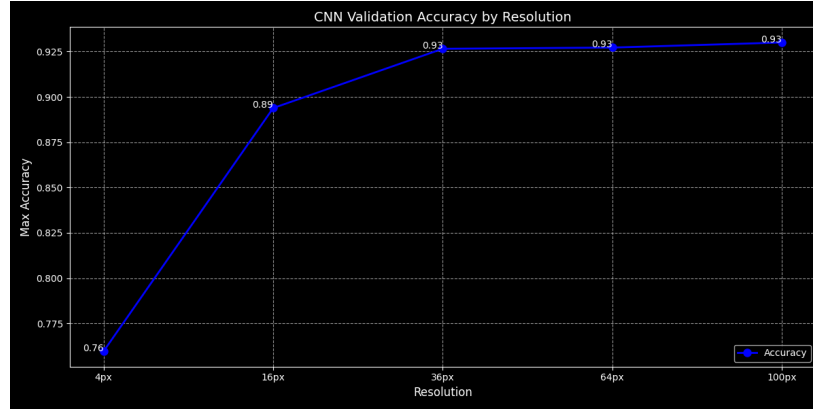


Figure 26: Graph showing the validation accuracy of the CNN model with k-fold cross validation applied on the mosaic images.

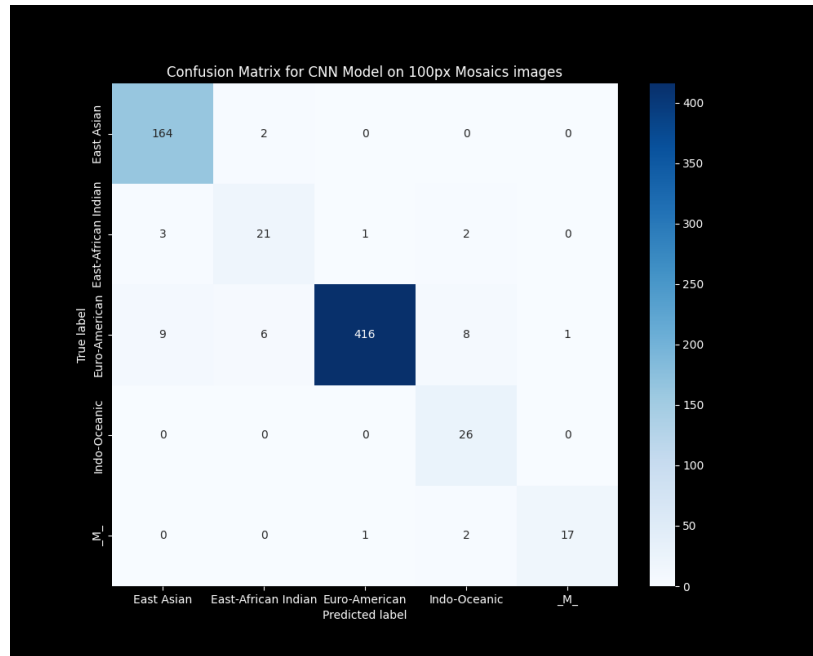


Figure 27: Confusion matrix with k-fold cross validation applied on the mosaic images at 100px resolution.

B.4 Under-Sampling techniques applied

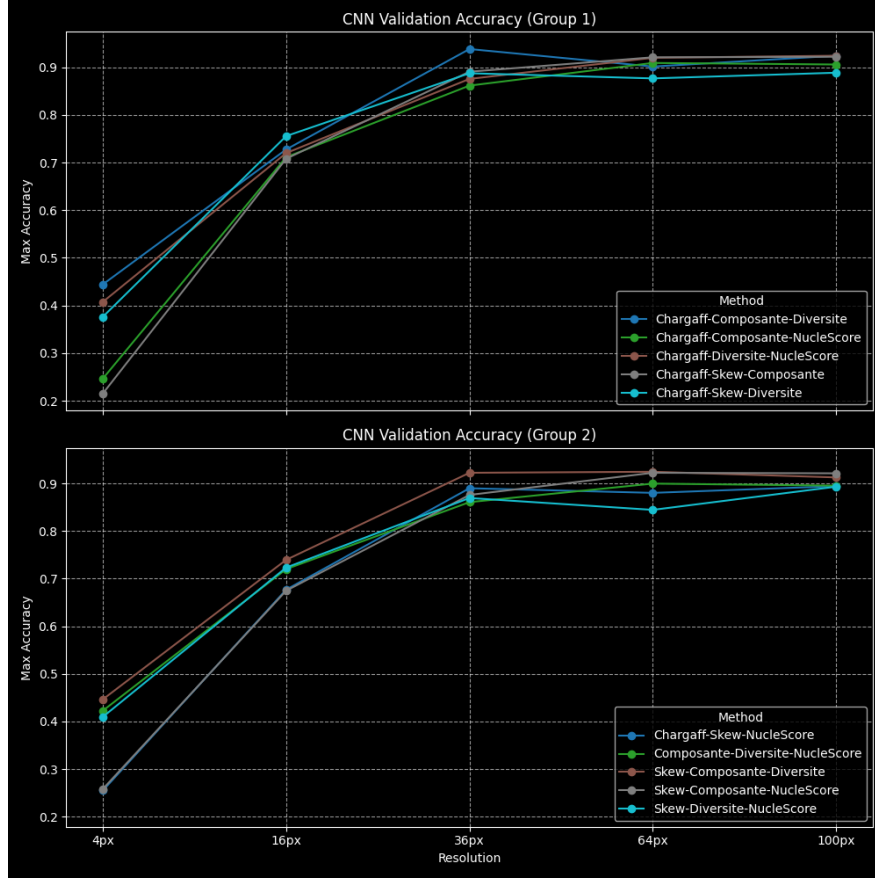


Figure 28: Graph showing the validation accuracy of the CNN model with under-sampling applied on the different methods.

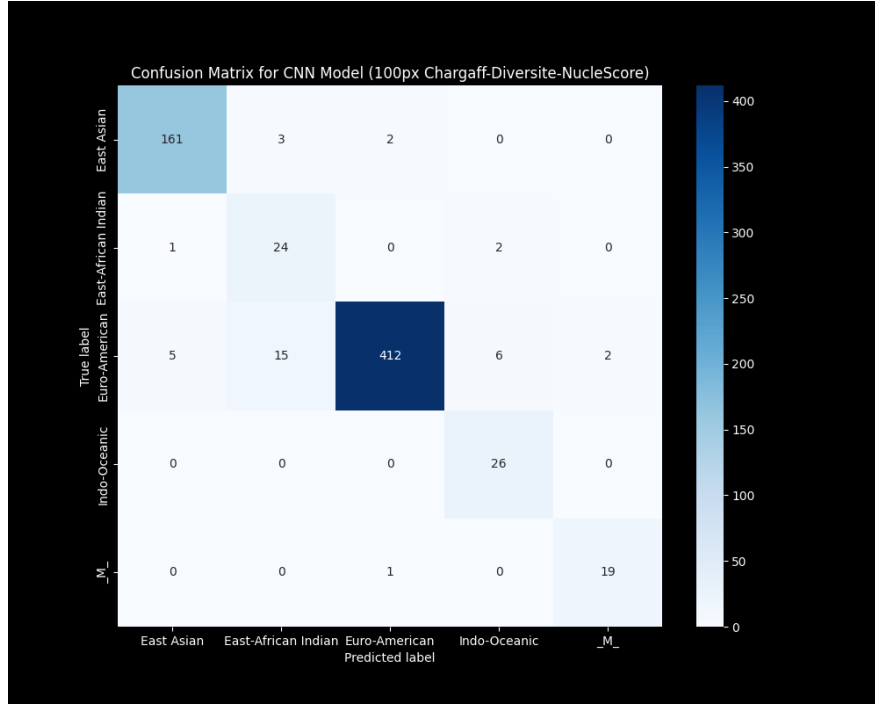


Figure 29: Confusion matrix with under-sampling applied on the images obtained from the Chargaff-Diversity-NucleScore combination at 100px resolution.

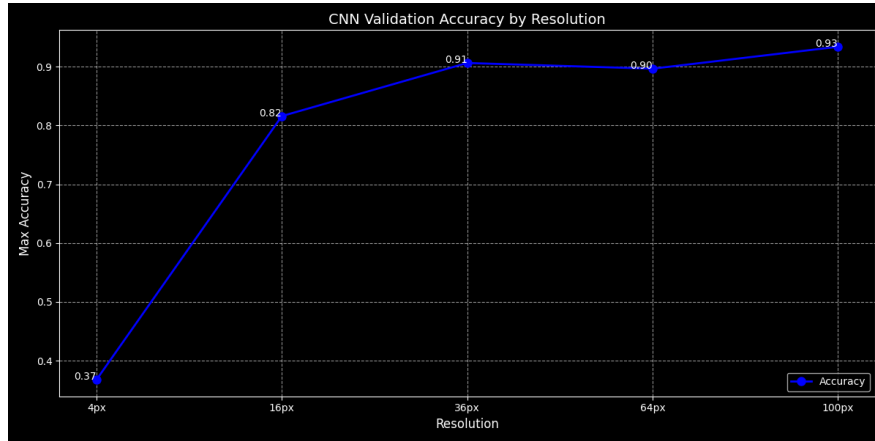


Figure 30: Graph showing the validation accuracy of the CNN model with under-sampling applied on the mosaic images.

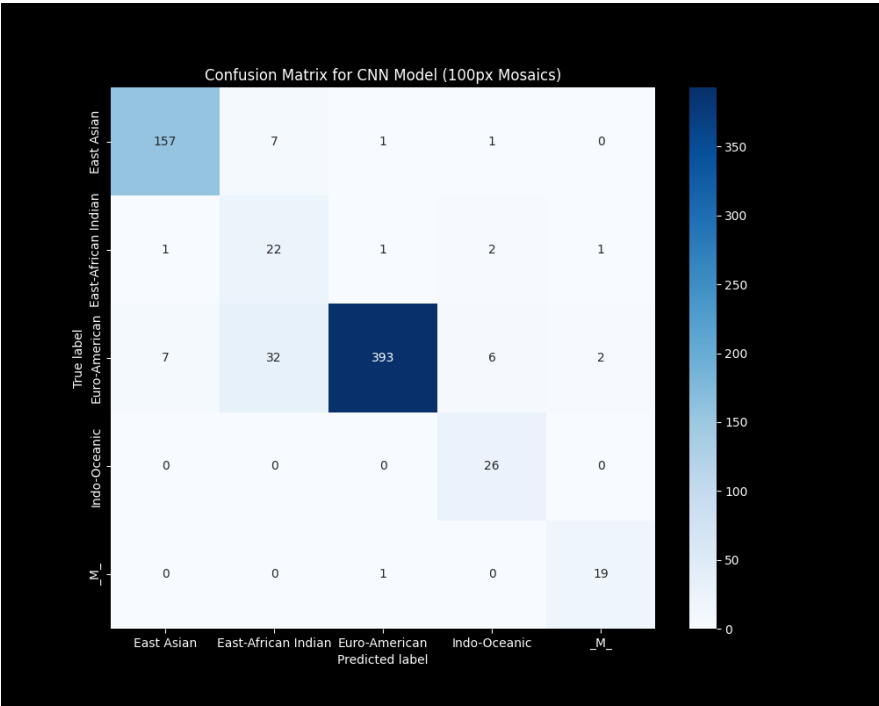


Figure 31: Confusion matrix with under-sampling applied on the mosaic images at 100px resolution.

B.5 Data Augmentation techniques applied

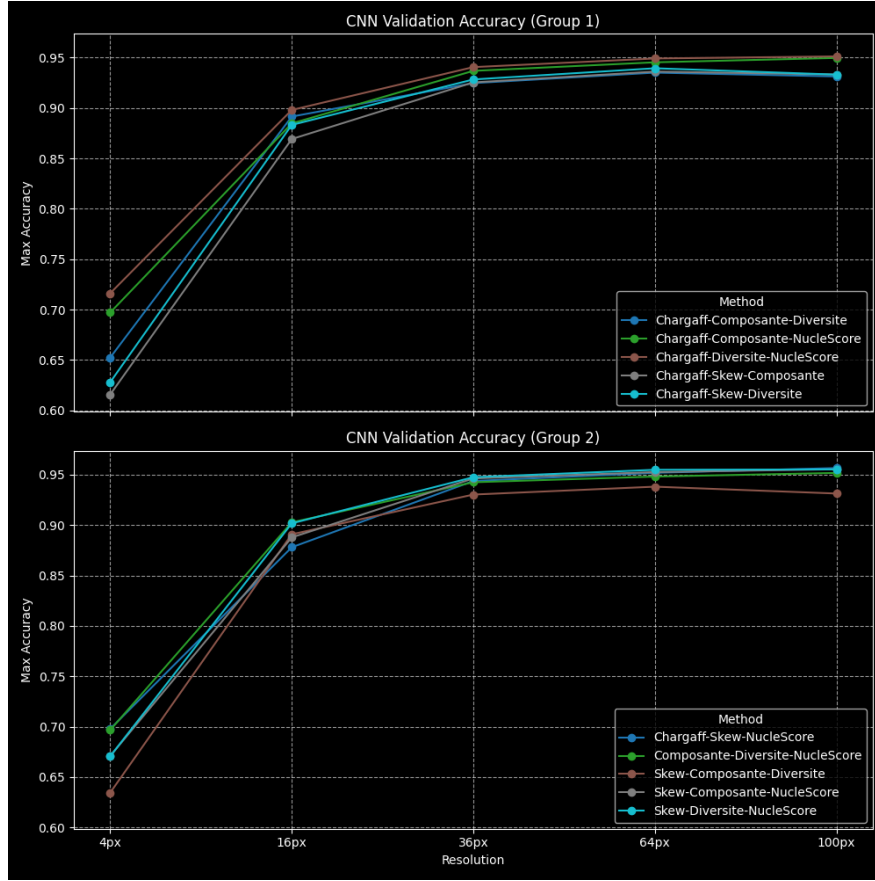


Figure 32: Graph showing the validation accuracy of the CNN model with data augmentation applied on the different methods.

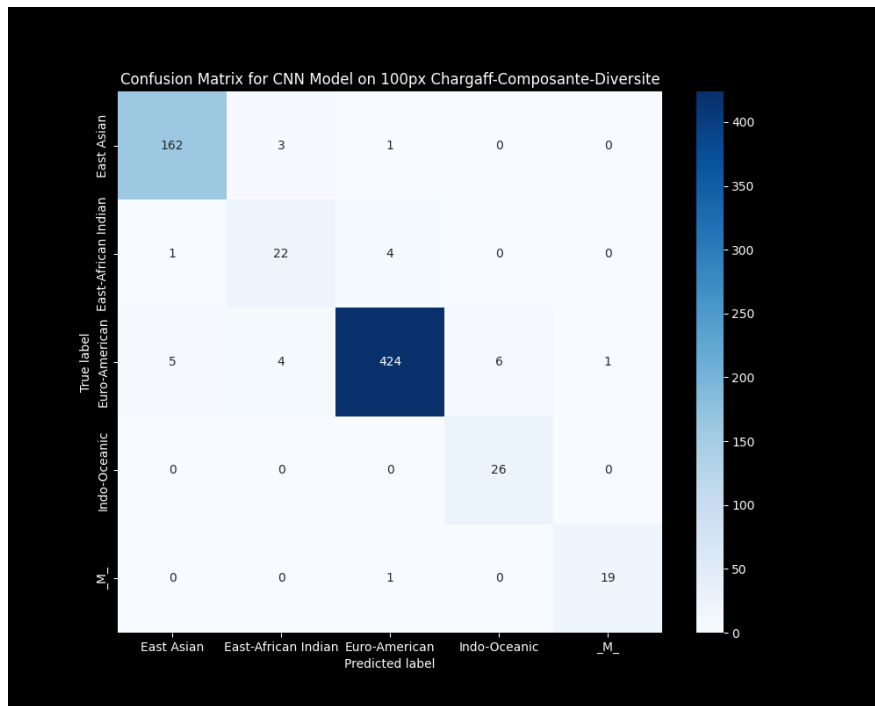


Figure 33: Confusion matrix with data augmentation applied on the images obtained from the Chargaff-Diversity-NucleScore combination at 100px resolution.

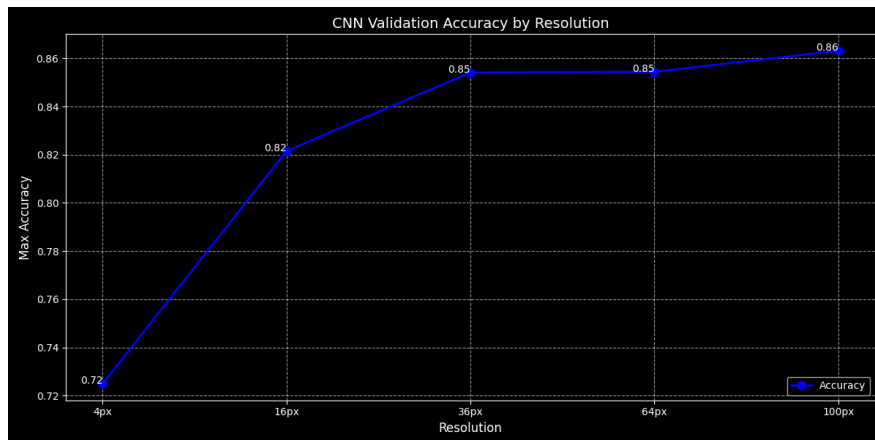


Figure 34: Graph showing the validation accuracy of the CNN model with data augmentation applied on the mosaic images.

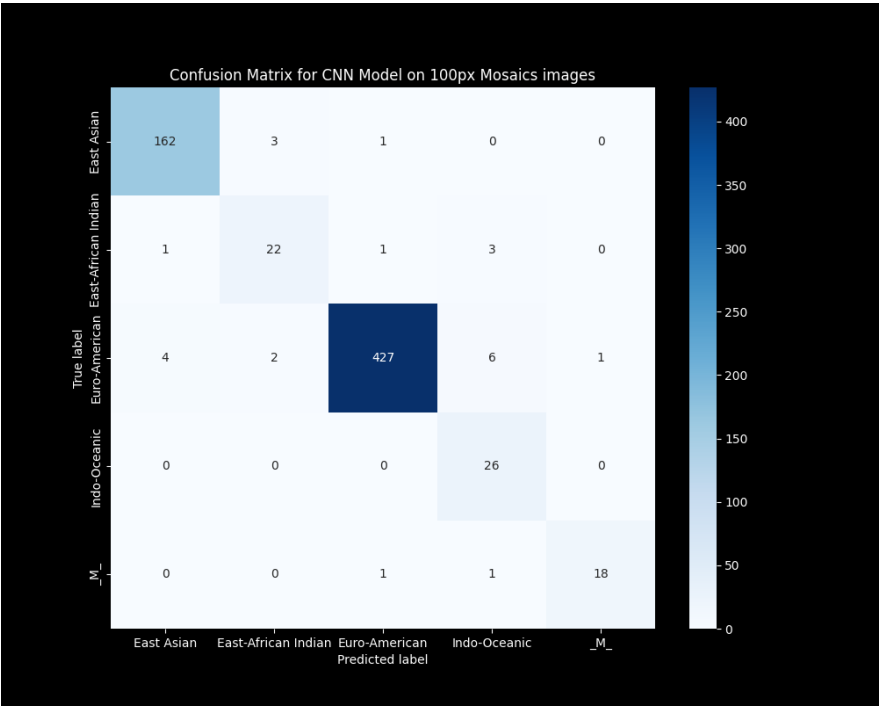


Figure 35: Confusion matrix with data augmentation applied on the mosaic images at 100px resolution.

B.6 Combined Techniques

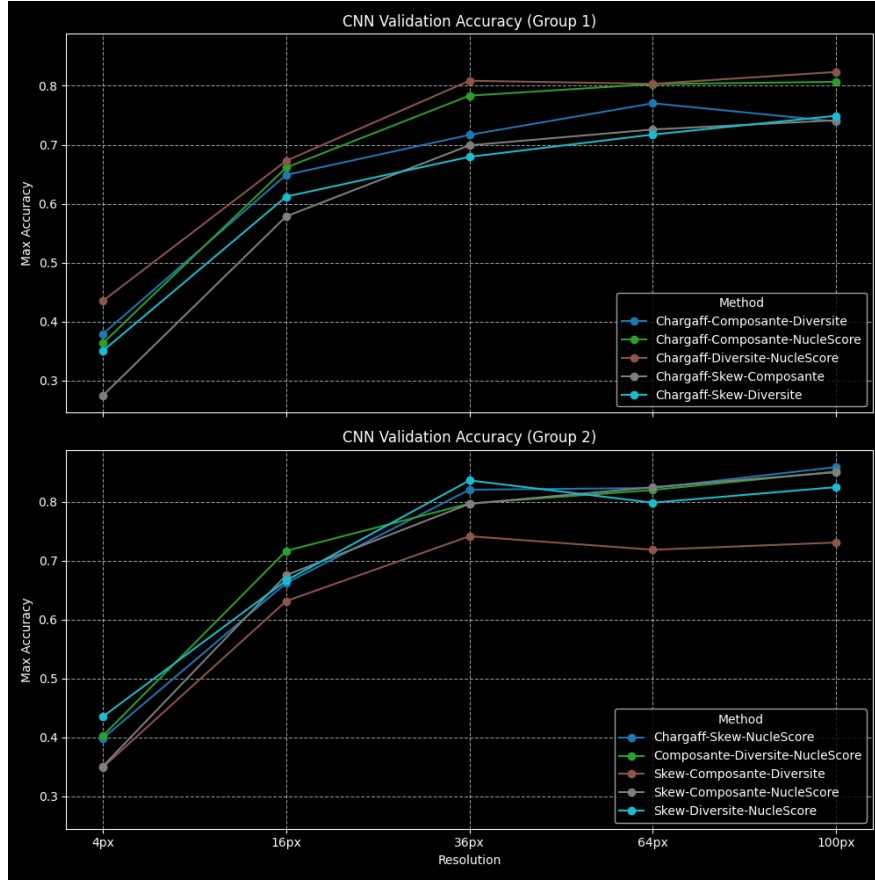


Figure 36: Graph showing the validation accuracy of the CNN model with combined under-sampling and data augmentation techniques applied on the different methods.

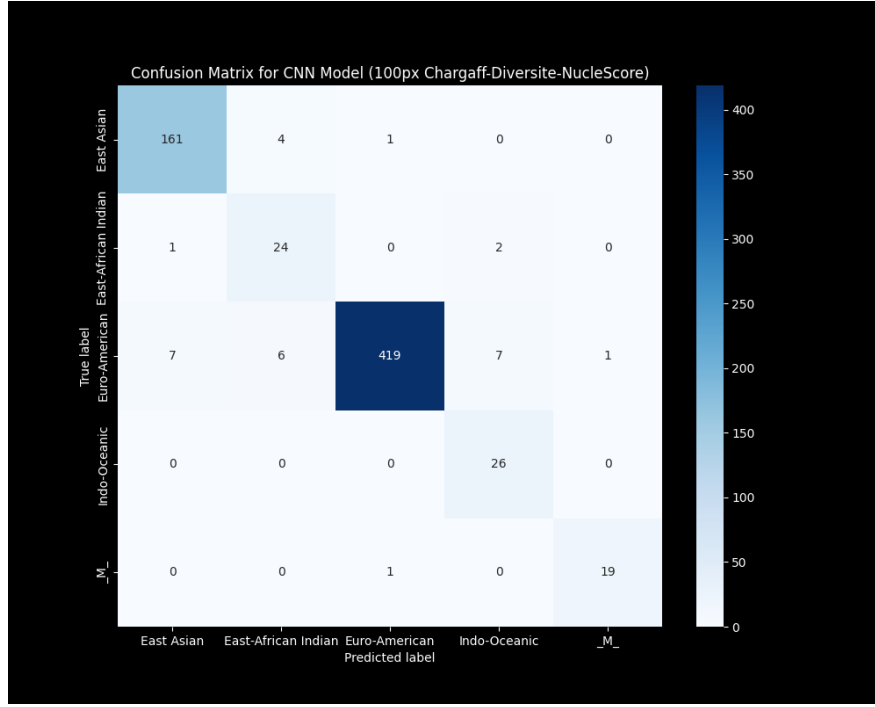


Figure 37: Confusion matrix with combined under-sampling and data augmentation techniques applied on the images obtained from the Chargaff-Diversity-NucleScore combination at 100px resolution.

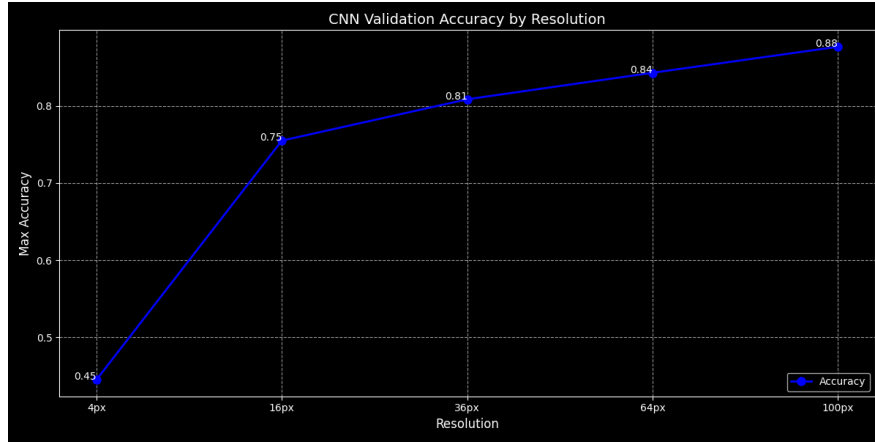


Figure 38: Graph showing the validation accuracy of the CNN model with combined under-sampling and data augmentation techniques applied on the mosaic images.

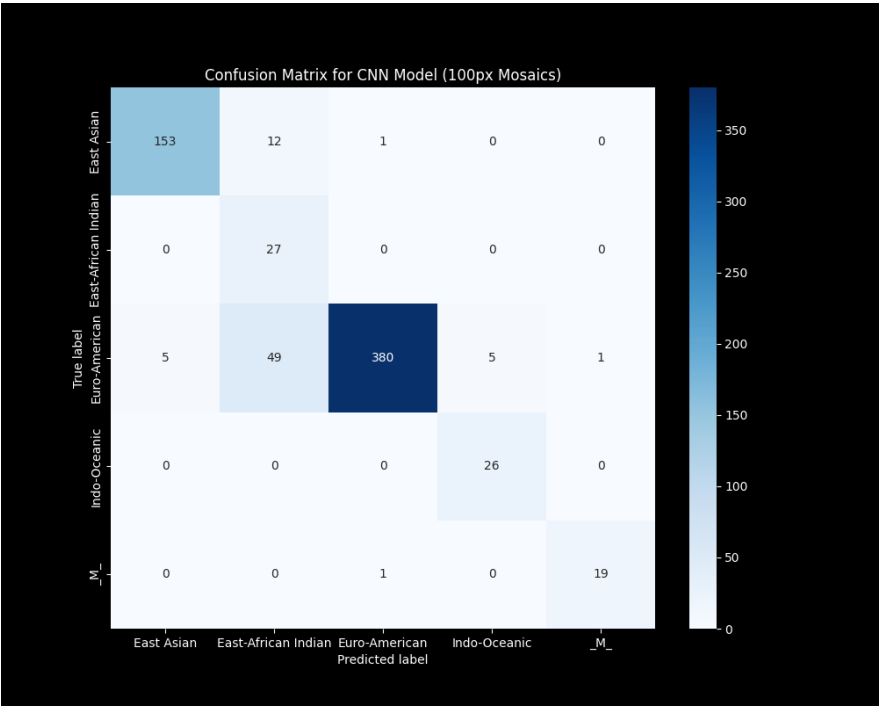


Figure 39: Confusion matrix with combined under-sampling and data augmentation techniques applied on the mosaic images at 100px resolution.

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