

# Applying Urinary Biomarkers of 11-dehydrothromboxane B2 and 8-isoprostane to Understand the Health Effects of NO<sub>2</sub> Exposure

[https://github.com/Yang190809/data\\_repository\\_yang](https://github.com/Yang190809/data_repository_yang)

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## **Abstract**

Mounting evidence shows that exposure to NO<sub>2</sub> generates oxidative stress. Oxidative stress can cause lipid damage, which plays an important role in various respiratory and cardiovascular diseases. Urinary 8-isoprostane is a product of lipid peroxidation which can reflect systemic lipid damage. I built linear mix models to analyze the relationship between the level of urinary 8-isoprostane with the level of NO<sub>2</sub> exposure. I found that short term NO<sub>2</sub> exposure was associated with lipid peroxidation, reflected by increased concentrations of urinary 8-isoprostane associated with increasing exposure. One IQR (7.41 µg/m<sup>3</sup>) incremental change of 12-h NO<sub>2</sub> exposure was associated with an increase of 8-isoprostane level by 19.45% (95% CI: 14.37%, 24.55%, p-value <0.001). One IQR (8.64 µg/m<sup>3</sup>) incremental change of 24-h NO<sub>2</sub> exposure was associated an increase in 8-isoprostane level by 27.69% (95% CI: 20.50%, 34.95% , p-value <0.001). One IQR (4.47 µg/m<sup>3</sup>) incremental change of 1-week NO<sub>2</sub> exposure was associated with an increase in 8-isoprostane level by 25.15% (95% CI: 14.14%,36.31%, p-value <0.05). One IQR (2.74 µg/m<sup>3</sup>) incremental change of 2-week NO<sub>2</sub> exposure was associated with an increase in 8-isoprostane level by 15.68% (95% CI: 8.87%,22.65%, p-value <0.05).

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# 1 Rationale and Research Questions

Exposure to NO<sub>2</sub> associated with cardiovascular disease, lung function impairment and asthma (Mölter, A., et. al.,2013, Collart, P., et. al.,2018, Takenoue, Y., et. al., 2012 ). Mounting evidence shows that exposure to NO<sub>2</sub> generates oxidative stress (Hashemzadeh, B., et. al.,2019). Oxidative stress can cause lipid damage (Black, C. N., et. al., 2017), which plays an important role in various respiratory and cardiovascular diseases. (Zanolin, M. E., et. a.,2015, Lakshmi, S. V., et. al., 2009). I applied 8-isoprostane to investigate the health effects of short term NO<sub>2</sub> exposure.

8-isoprostane is the product of the oxidized cell membrane after being attacked by reactive oxygen species such as peroxides. Therefore, 8-isoprostane can reflect lipid damage (Danielsen et al., 2009). 8-isoprostane in urine does not have a diurnal variation and has been proven to be a stable biomarker showing systemic lipid damage. It has been applied in studying diseases such as type 2 diabetes mellitus, obesity, coronary heart disease, asthma, and acute respiratory distress syndrome (Nuernberg, A. M., et al., 2008). Nevertheless, the use of this biomarker in the research of air pollution and its health effects is scarce. One study in Iran discovered a significant positive relationship between short-term NO<sub>2</sub> exposure and 8-isoprostane in exhaled breath condensate (EBC) in healthy children aged 12-13 years old (Hashemzadeh, B., et. al., 2019). One study in New York City found that the increases in 1- to 5-day averages of nitrogen dioxide were significantly associated with increases in 8-isoprostane in EBC among 18-year old healthy and asthma affected individuals (Patel, M. M., et. al., 2013). My research question is: If the NO<sub>2</sub> exposure is positively associated with urinary 8-isoprostane? Among periods of 12-hour,24-hour,1-week and 2-weeks, which period has the most significant association with urinary 8-isoprostane?

## 2 Dataset Information

All the data are from Jim Zhang’s lab. Jim Zhang is a professor at the Nicholas School of Environment at Duke University. The file 8-iso is the data set of urinary biomarker 8-isoprostane. The file urine\_info is the data set that has information about the subjects who provided the urine samples. The urine list is a data set with lists of information of sample ID, subject ID and visits. The meaning of the columns as well as units and class of the data in each folder is listed below. Column names without descriptors are irrelevant to this study.

### 2.1 Urine list raw data set

The urine list has the information about the sample ID, subject ID and visits.

Column Name	Meaning	units	class of the data
Sample ID	the identity number of the samples	NA	integer
Subject ID	the identity number of the subjects	NA	integer
visit	the number of the 4 visits (1,2,3 or 4)	NA	integer

**Table 1**

## 2.2 8-isoprostane data set

The 8-isoprostane data set has information about 8-isoprostane concentration tested in the mass spectrum and the concentration of 8-isoprostane in original urine samples. The limit of detection for 8-isoprostane was 0.016ng/ml. Any value which is below 0.016 in the column of calculated Conc in 8-is data set should be excluded as an error.

Column Name	Meaning	Units	Class of the data
Sample ID	the identity number of the samples	NA	integer
Calculated Conc	the concentration tested by the machine	ng/ml	numeric
Sample Conc	the concentration in the original urine	ng/ml	numeric

**Table 2**

## 2.3 Urine Info

The urine info is the data set that has information about subjects characteristics and average pollutants exposure (NO<sub>2</sub>, SO<sub>2</sub>, O<sub>3</sub> and PM<sub>2.5</sub>) over the periods of 12 hours, 24 hours, one week and two weeks. It also includes average temperature and humidity over the periods of 12 hours, 24 hours, one week and two weeks.

Column name	Meaning	Units	Class of the data
ample_ID	the identity number of the samples	NA	integer
SubjectID	the identity number of the subjects	NA	integer
COLD	cold ( represent respiratory infection)	NA	category
MNST	menstration during visit	NA	category
last.ate	the hours to the last meal	hours	integer
wkday.start	the day that the subject start their work	NA	category
dt_smoke	second-hand smoke exposure in hours	hours	numeric
USG urine	specific gravity	g/ml	numeric
o3exp.12h	the exposure of ozone in 12h	ug/m3	numeric
pmexp.12h	the exposure of PM <sub>2.5</sub> in 12h	ug/m3	numeric
no2exp.12h	the exposure of NO <sub>2</sub> in 12h	ug/m3	numeric
so2exp.12h	the exposure of SO <sub>2</sub> in 12h	ug/m3	numeric
Temp.12h	temperature in 12h	ug/m3	numeric
RHx.12h	humidity in 12h	ug/m3	numeric

**Table 3**

## 2.4 Data Wrangling

My data wrangling started with loading the data set of 8-isoprostane and changing the column names. Then I merged this data set with the urine list to match the sample ID with subjects ID and visit. Then this merged data was merged with the urine info data set using subject ID and visit as the matching keys. After the second merging, the rows with NAs were removed. Then I calculated the normalized 8-isoprostane using the specific urine gravity. This normlization can adjust the dilution in urine. I obtained my final data by selecting columns that is revelant to my rearch question. Finally, the processed data was saved into a processed data folder.

Statistic	N	Mean	St. Dev.	Min	Pctl(25)	Pctl(75)	Max
8-isoprostane	302	3.806	5.780	0.186	0.747	4.005	48.299
11dhTxB2	302	1.510	1.259	0.141	0.812	1.861	14.255

Table 4

## 1. Data wrangling

```
# Set working directory
getwd()

## [1] "C:/Users/26059/OneDrive/Desktop/ENV 872 R/Yang_ENV872/directory_yang"

# Load packages
library(tidyverse)
library(lmerTest)
library(MuMIn)
library(car)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
# Set ggplot theme
mytheme <- theme_classic(base_size = 25) +
  theme(axis.text = element_text(color = "black"),
        legend.position = "top")
theme_set(mytheme)

# Load dataset 1
is <- read.csv("raw_data/8iso.csv")
is <- select(is, Sample.ID, Calculated.Conc, Sample.Conc)
colnames(is)[colnames(is) == "Calculated.Conc"] <- "is.origin"
colnames(is)[colnames(is) == "Sample.Conc"] <- "is.conc"

# Load dataset 2
list <- read.csv("raw_data/urine_list.csv")

#merge
merge1 <- merge(x=is,y=list,by="Sample.ID",all=TRUE)

urine.info <- read.csv("raw_data/urine_info.csv")
merge3 <- merge(x=merge1,y=urine.info,by=c("Subject.ID","visit"),all=TRUE)
dim(merge3)

## [1] 344 69

full.data <- na.omit(merge3)
dim(full.data)

## [1] 307 69
```

```
mean.cre<- mean(full.data$USG)
mean.cre
```

```
## [1] 1.016117
```

```
#calculate normalized biomarker concentration
full.data$is.ad <- (full.data$is.conc/(1-full.data$USG))*(1-mean.cre)

#select useful columns
final.dat<- select(full.data, Sample.ID, Subject.ID, is.ad, is.conc, is.origin, group,

#save processed data set
write.csv(final.dat, row.names = FALSE, file = "processed_data/final.dat.csv")
```

## 2.Explore the data

```
library(ggplot2)
#load processed data set
dat <- read.csv("processed_data/final.dat.csv")
#remove outliers
summary(dat$is.origin)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##  1.508   6.220  13.421  45.703  38.281 605.614
```

```
dat1<- dat[-c(4,7,89,147,167),]
```

```
#test normality
shapiro.test(dat1$is.ad)
```

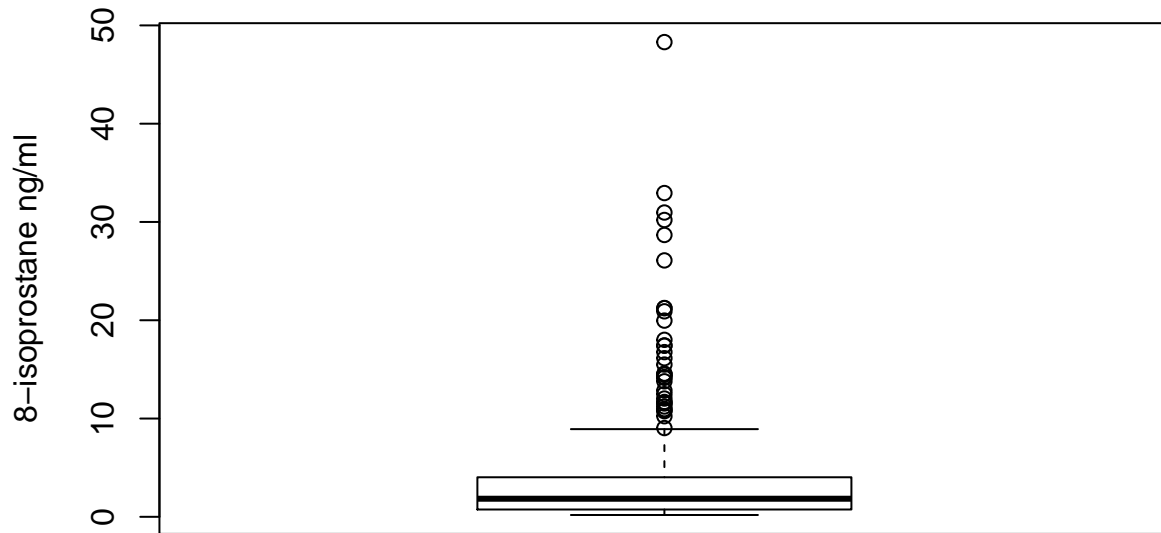
```
##
##  Shapiro-Wilk normality test
##
## data:  dat1$is.ad
## W = 0.58916, p-value < 2.2e-16
```

```
shapiro.test(log(dat1$is.ad))
```

```
##
##  Shapiro-Wilk normality test
##
## data:  log(dat1$is.ad)
## W = 0.98127, p-value = 0.0005511
```

```
#explore the data
```

```
is.plot1 <- boxplot(dat1$is.ad,colors="darkblue",ylab="8-isoprostane ng/ml")
```

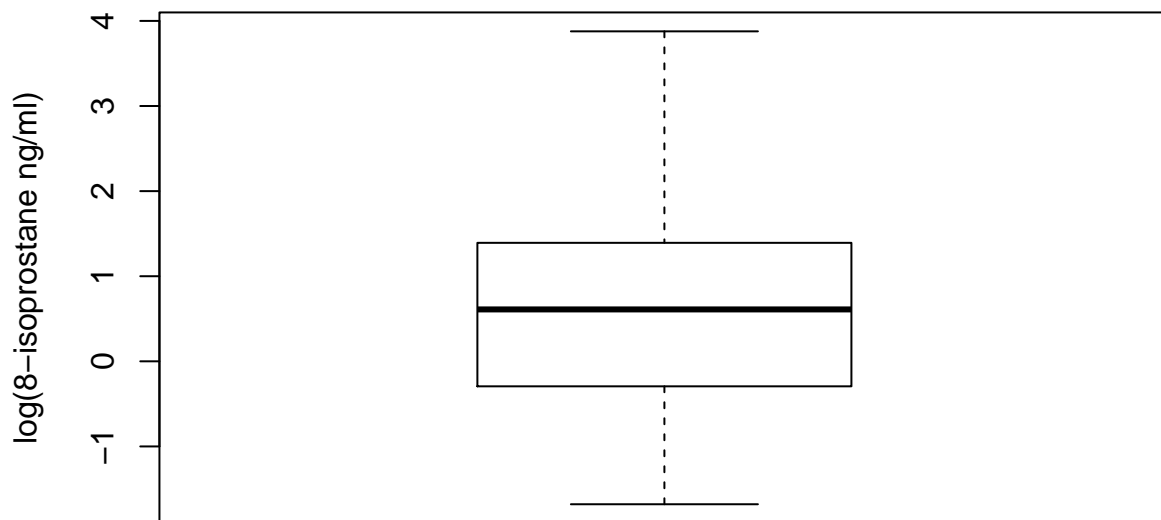


**Figure 1: Plot of 8-isoprostane**

Figure 1 shows that the range of concentration of 8-isoprostane is huge and it is not normalized

```
is.plot1 <- boxplot(log(dat1$is.ad),color="darkblue",ylab="log(8-isoprostane ng/ml)")
```





**Figure 2: Plot of log(8-isoprostane)**

Figure 2 shows that the log(8-isoprostane) looks more normalized than original scale of 8-isoprostane.

```
#calculate IQR for each period
sum3<-summary(dat$no2exp.12h)
no2.12 <-sum3[5]-sum3[2]
no2.12
```

```
## 3rd Qu.
## 7.409404
```

```
sum4<-summary(dat$no2exp.24h)
no2.24 <-sum4[5]-sum4[2]
no2.24
```

```
## 3rd Qu.
## 8.638027
```

```
sum5<-summary(dat$no2exp.1w)
no2.1w <-sum5[5]-sum5[2]
no2.1w
```

```
## 3rd Qu.
## 8.617326
```

```
sum6<-summary(dat$no2exp.2w)
no2.2w <-sum6[5]-sum6[2]
no2.2w
```

```
## 3rd Qu.
## 2.736119
```

### 3.Build models

#### 3.1 Build models for 12-hour NO2 exposure

```
mo1<- lmer(log(is.ad) ~no2exp.12h+o3exp.12h+ pmexp.12h+so2exp.12h+Temp.12h+RHx.12h + last.ate, data=dat)
#find the best model with lowest AIC
step(mo1)
```

```
## Backward reduced random-effect table:
```

```
##
##               Eliminated npar  logLik      AIC      LRT Df Pr(>Chisq)
## <none>                16 -474.63   981.26
## (1 | Subject.ID)      0   15 -493.94 1017.88 38.616  1  5.159e-10 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
##
```

```
## Backward reduced fixed-effect table:
```

```
## Degrees of freedom method: Satterthwaite
```

```
##
```

```
##               Eliminated  Sum Sq Mean Sq NumDF  DenDF F value  Pr(>F)
## Temp.12h              1  0.0261  0.0261     1 276.301  0.0314 0.859411
## COLD                  2  0.0677  0.0677     1 266.912  0.0819 0.774976
## pmexp.12h             3  0.0956  0.0956     1 271.141  0.1159 0.733779
## go.home               4  0.6585  0.3293     2  83.839  0.4012 0.670812
## wkday.start           5  0.3986  0.3986     1 291.709  0.4848 0.486792
## so2exp.12h            6  0.4401  0.4401     1 293.944  0.5343 0.465386
## MNST                  7  0.8942  0.8942     1 272.520  1.0861 0.298261
## last.ate              8  0.8829  0.8829     1 276.178  1.0773 0.300216
## dt_smoke              9  0.9658  0.9658     1 293.061  1.1845 0.277332
```

```

## RHx.12h          10  2.0108  2.0108      1 251.786  2.4885 0.115939
## no2exp.12h       0 12.2057 12.2057      1 228.742 14.9910 0.000141 ***
## o3exp.12h        0  4.0334  4.0334      1 225.577  4.9538 0.027023 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Model found:
## log(is.ad) ~ no2exp.12h + o3exp.12h + (1 | Subject.ID)

#best model for 12-hour NO2 exposure
mo1.1<- lmer(log(is.ad) ~ no2exp.12h +o3exp.12h + (1 | Subject.ID),data=dat1)
summary(mo1.1)

## Linear mixed model fit by REML. t-tests use Satterthwaite's method [
## lmerModLmerTest]
## Formula: log(is.ad) ~ no2exp.12h + o3exp.12h + (1 | Subject.ID)
## Data: dat1
##
## REML criterion at convergence: 910.8
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -1.99963 -0.62729 -0.03595  0.60429  2.58701
##
## Random effects:
##  Groups      Name      Variance Std.Dev.
## Subject.ID (Intercept) 0.5187   0.7202
## Residual              0.8142   0.9023
## Number of obs: 302, groups: Subject.ID, 89
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept) -0.187878   0.212372 292.646414  -0.885 0.377064
## no2exp.12h   0.025912   0.006693 228.741989   3.872 0.000141 ***
## o3exp.12h    0.031066   0.013958 225.576673   2.226 0.027023 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) n2x.12
## no2exp.12h -0.812
## o3exp.12h  -0.489  0.132

```

```
#check F value
```

```
anova(mo1.1)
```

```
## Type III Analysis of Variance Table with Satterthwaite's method
```

```
##           Sum Sq Mean Sq NumDF  DenDF F value    Pr(>F)
## no2exp.12h 12.2057 12.2057     1 228.74 14.9910 0.000141 ***
## o3exp.12h   4.0334  4.0334     1 225.58  4.9538 0.027023 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
#check collinearity
```

```
vif(mo1.1)
```

```
## no2exp.12h  o3exp.12h
```

```
##    1.017673    1.017673
```

```
#get R2
```

```
r.squaredGLMM (mo1.1)
```

```
##           R2m          R2c
```

```
## [1,] 0.03761012 0.412126
```

```
#identify outliers
```

```
res1.1 <- resid(mo1, type = "pearson")
```

```
dat[which(abs(res1.1) > 2.5),]
```

```
##  [1] Sample.ID    Subject.ID    is.ad         is.conc       is.origin     group
##  [7] COLD          MNST          last.ate      wkday.start   go.home       Smoker
## [13] dt_smoke      o3exp.12h     pmexp.12h     no2exp.12h    so2exp.12h    Temp.12h
## [19] RHx.12h       o3exp.24h     pmexp.24h     no2exp.24h    so2exp.24h    Temp.24h
## [25] RHx.24h       o3exp.1w      pmexp.1w      no2exp.1w     so2exp.1w     Temp.1w
## [31] RHx.1w        o3exp.2w      pmexp.2w      no2exp.2w     so2exp.2w     Temp.2w
## [37] RHx.2w
## <0 rows> (or 0-length row.names)
```

```
#calculate IQR change
```

```
(exp(0.025912)-1)*no2.12
```

```
##    3rd Qu.
```

```
## 0.1945016
```

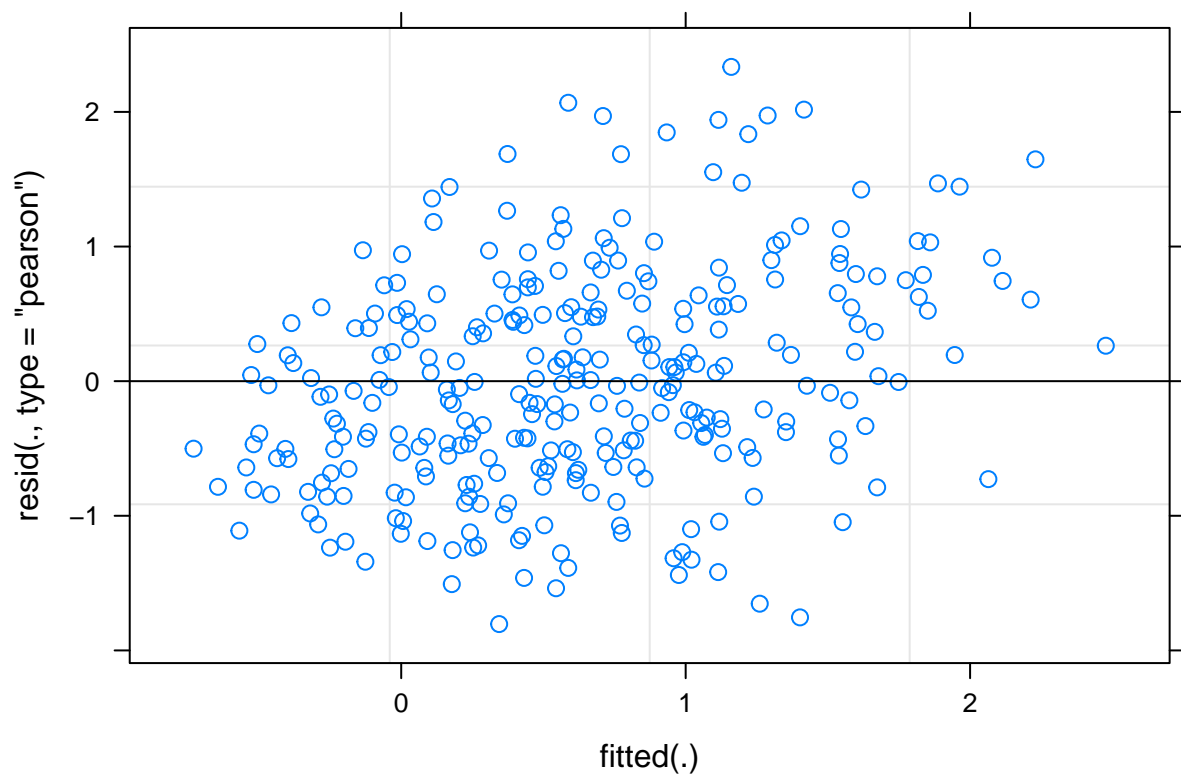
```
(exp(0.025912 +0.006693)-1)*no2.12
```

```
## 3rd Qu.  
## 0.2455652
```

```
(exp(0.025912 -0.006693)-1)*no2.12
```

```
## 3rd Qu.  
## 0.1437785
```

```
#plot residues  
plot(mo1.1)
```



**Figure 3: diagnostic plot of model 1.1**

Figure 3 shows that the higher the fitted value, the higher the residue. However, in general it looks fine.

### 3.2 Build models for 24-hour NO<sub>2</sub> exposure

```
mo2<- lmer(log(is.ad) ~ no2exp.24h+o3exp.24h+ pmexp.24h+so2exp.24h+Temp.24h+RHx.24h + last.ate, data=dat1)
step(mo2)
```

```
## Backward reduced random-effect table:
```

```
##
##               Eliminated npar  logLik      AIC      LRT Df Pr(>Chisq)
## <none>                16 -475.04   982.07
## (1 | Subject.ID)      0   15 -493.92 1017.84 37.768   1 7.969e-10 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
## Backward reduced fixed-effect table:
```

```
## Degrees of freedom method: Satterthwaite
```

```
##
##               Eliminated  Sum Sq Mean Sq NumDF   DenDF F value    Pr(>F)
## so2exp.24h           1  0.0016  0.0016     1 280.151  0.0019 0.9655516
## Temp.24h             2  0.0161  0.0161     1 277.135  0.0193 0.8895889
## wkday.start          3  0.0126  0.0126     1 279.050  0.0152 0.9021154
## COLD                 4  0.1555  0.1555     1 268.565  0.1873 0.6655198
## go.home              5  0.9235  0.4617     2  84.443  0.5575 0.5747522
## RHx.24h              6  0.4390  0.4390     1 258.415  0.5298 0.4673562
## dt_smoke             7  0.5832  0.5832     1 291.289  0.7037 0.4022322
## pmexp.24h            8  0.8868  0.8868     1 273.549  1.0799 0.2996466
## MNST                 9  0.8850  0.8850     1 274.024  1.0795 0.2997231
## last.ate            10  1.0377  1.0377     1 281.175  1.2708 0.2605860
## no2exp.24h           0 12.3002 12.3002     1 231.652 15.1608 0.0001292 ***
## o3exp.24h            0  5.0095  5.0095     1 225.520  6.1746 0.0136869 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
## Model found:
```

```
## log(is.ad) ~ no2exp.24h + o3exp.24h + (1 | Subject.ID)
```

```
mo2.1<- lmer(log(is.ad) ~ no2exp.24h + o3exp.24h + (1 | Subject.ID),data=dat1)
summary(mo2.1)
```

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method [
```

```
## lmerModLmerTest]
```

```
## Formula: log(is.ad) ~ no2exp.24h + o3exp.24h + (1 | Subject.ID)
```

```
## Data: dat1
```

```
##
```

```
## REML criterion at convergence: 910.4
```

```
##
```

```
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -1.88882 -0.65529 -0.03642  0.58790  2.66141
##
## Random effects:
##   Groups      Name      Variance Std.Dev.
##   Subject.ID (Intercept) 0.5243   0.7241
##   Residual                0.8113   0.9007
## Number of obs: 302, groups: Subject.ID, 89
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept) -0.339832   0.243838 284.149326  -1.394 0.164503
## no2exp.24h   0.031555   0.008104 231.652052   3.894 0.000129 ***
## o3exp.24h    0.030798   0.012394 225.520144   2.485 0.013687 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) n2x.24
## no2exp.24h -0.857
## o3exp.24h  -0.505  0.196
```

```
anova(mo2.1)
```

```
## Type III Analysis of Variance Table with Satterthwaite's method
##              Sum Sq Mean Sq NumDF  DenDF F value    Pr(>F)
## no2exp.24h 12.3002 12.3002     1 231.65 15.1608 0.0001292 ***
## o3exp.24h   5.0095  5.0095     1 225.52  6.1746 0.0136869 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
vif(mo2.1)
```

```
## no2exp.24h  o3exp.24h
##   1.039908   1.039908
```

```
r.squaredGLMM (mo2.1)
```

```
##              R2m      R2c
## [1,] 0.03854593 0.4159847
```

```
res2.1 <- resid(mo2.1, type = "pearson")
dat[which(abs(res2.1) > 2.5),]
```

```
## [1] Sample.ID Subject.ID is.ad is.conc is.origin group
## [7] COLD MNST last.ate wkday.start go.home Smoker
## [13] dt_smoke o3exp.12h pmexp.12h no2exp.12h so2exp.12h Temp.12h
## [19] RHx.12h o3exp.24h pmexp.24h no2exp.24h so2exp.24h Temp.24h
## [25] RHx.24h o3exp.1w pmexp.1w no2exp.1w so2exp.1w Temp.1w
## [31] RHx.1w o3exp.2w pmexp.2w no2exp.2w so2exp.2w Temp.2w
## [37] RHx.2w
## <0 rows> (or 0-length row.names)
```

```
#calculate IQR change
(exp(0.031555)-1)*no2.24
```

```
## 3rd Qu.
## 0.276919
```

```
(exp(0.031555 +0.008104)-1)*no2.24
```

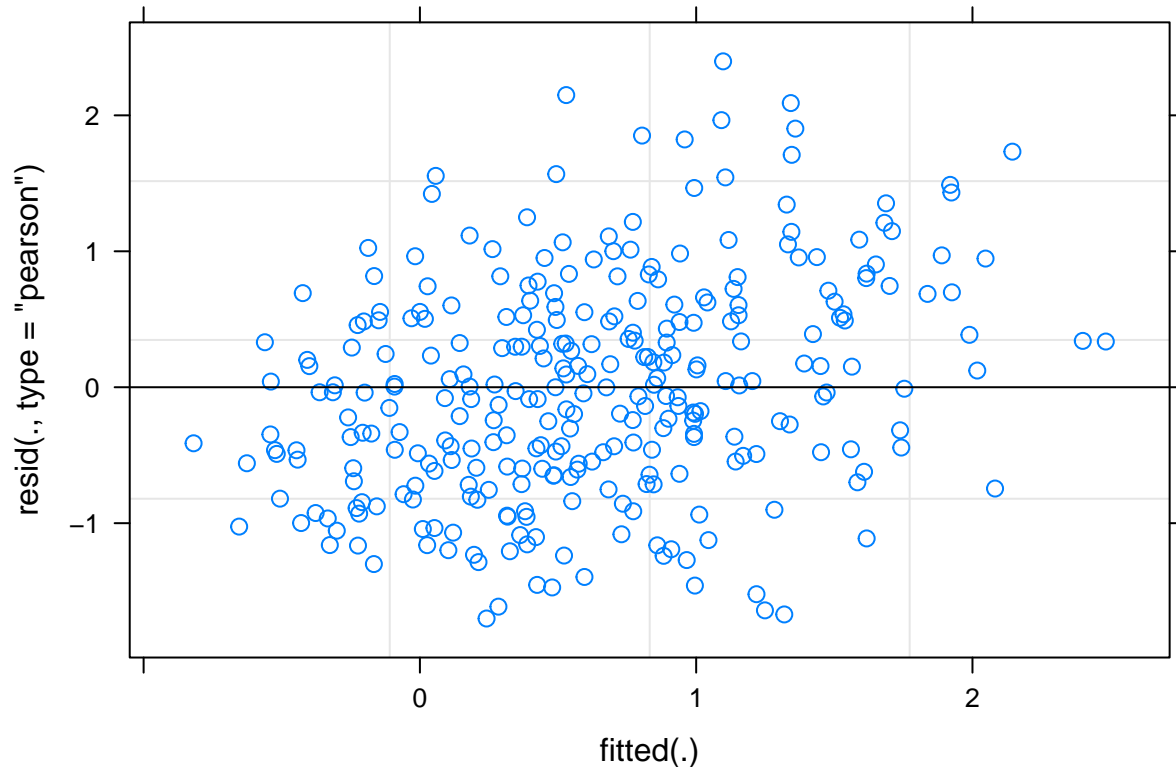
```
## 3rd Qu.
## 0.3494593
```

```
(exp(0.031555 -0.008104)-1)*no2.24
```

```
## 3rd Qu.
## 0.2049643
```

```
plot(mo2.1)
```





**Figure 4: diagnostic plot of model 2.1**

Figure 4 shows that the higher the fitted value, the higher the residue. However, in general it looks fine. 3.3 Build models for 1-week NO<sub>2</sub> exposure

```
mo3<- lmer(log(is.ad) ~ no2exp.1w+o3exp.1w+ pmexp.1w+so2exp.1w+Temp.1w+RHx.1w + last.ate
step(mo3)
```

```
## Backward reduced random-effect table:
```

```
##
```

```
##           Eliminated npar  logLik      AIC    LRT Df Pr(>Chisq)
```

```
## <none>                16 -475.45  982.89
```

```
## (1 | Subject.ID)      0   15 -493.45 1016.90 36.001  1  1.972e-09 ***
```

```
## ---
```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
##
```

```
## Backward reduced fixed-effect table:
```

```
## Degrees of freedom method: Satterthwaite
```

```
##
```

```
##           Eliminated Sum Sq Mean Sq NumDF  DenDF F value    Pr(>F)
```

```
## Temp.1w           1 0.0000  0.0000     1 256.639  0.0000 0.9986105
```

```
## RHx.1w          2 0.0271  0.0271      1 288.772  0.0317 0.8588147
## COLD            3 0.0560  0.0560      1 271.043  0.0657 0.7979384
## wkday.start     4 0.1240  0.1240      1 258.909  0.1459 0.7027697
## go.home         5 0.8717  0.4359      2  84.614  0.5134 0.6002845
## pmexp.1w        6 0.2371  0.2371      1 293.316  0.2793 0.5975635
## dt_smoke        7 0.4455  0.4455      1 291.942  0.5270 0.4684347
## last.ate        8 0.6026  0.6026      1 277.597  0.7195 0.3970452
## MNST            9 0.7572  0.7572      1 273.657  0.9102 0.3409085
## o3exp.1w       10 1.4655  1.4655      1 229.000  1.7684 0.1849006
## no2exp.1w        0 4.4044  4.4044      1 229.706  5.2961 0.0222690 *
## so2exp.1w        0 9.8969  9.8969      1 230.979 11.9006 0.0006669 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Model found:
## log(is.ad) ~ no2exp.1w + so2exp.1w + (1 | Subject.ID)
```

```
mo3.1<- lmer(log(is.ad) ~ no2exp.1w + so2exp.1w + (1 | Subject.ID),data=dat1)
summary(mo3.1)
```

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method [
## lmerModLmerTest]
## Formula: log(is.ad) ~ no2exp.1w + so2exp.1w + (1 | Subject.ID)
## Data: dat1
##
## REML criterion at convergence: 913.2
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -1.95288 -0.61932 -0.07413  0.62697  2.63863
##
## Random effects:
## Groups      Name      Variance Std.Dev.
## Subject.ID (Intercept) 0.5199   0.7211
## Residual              0.8316   0.9119
## Number of obs: 302, groups: Subject.ID, 89
##
## Fixed effects:
##              Estimate Std. Error    df t value Pr(>|t|)
## (Intercept)   0.76276    0.29669 261.12832   2.571 0.010698 *
## no2exp.1w      0.02877    0.01250 229.70566   2.301 0.022269 *
## so2exp.1w     -0.12342    0.03578 230.97927  -3.450 0.000667 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
##
## Correlation of Fixed Effects:
##          (Intr) n2xp.1
## no2exp.1w -0.624
## so2exp.1w -0.331 -0.476
```

```
anova(mo3.1)
```

```
## Type III Analysis of Variance Table with Satterthwaite's method
##          Sum Sq Mean Sq NumDF  DenDF F value    Pr(>F)
## no2exp.1w 4.4044  4.4044     1 229.71  5.2961 0.0222690 *
## so2exp.1w 9.8969  9.8969     1 230.98 11.9006 0.0006669 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
vif(mo3.1)
```

```
## no2exp.1w so2exp.1w
##  1.293508  1.293508
```

```
r.squaredGLMM (mo3.1)
```

```
##          R2m          R2c
## [1,] 0.02719595 0.4014215
```

```
#calculate IQR change
(exp( 0.02877)-1)*no2.1w
```

```
## 3rd Qu.
## 0.2515213
```

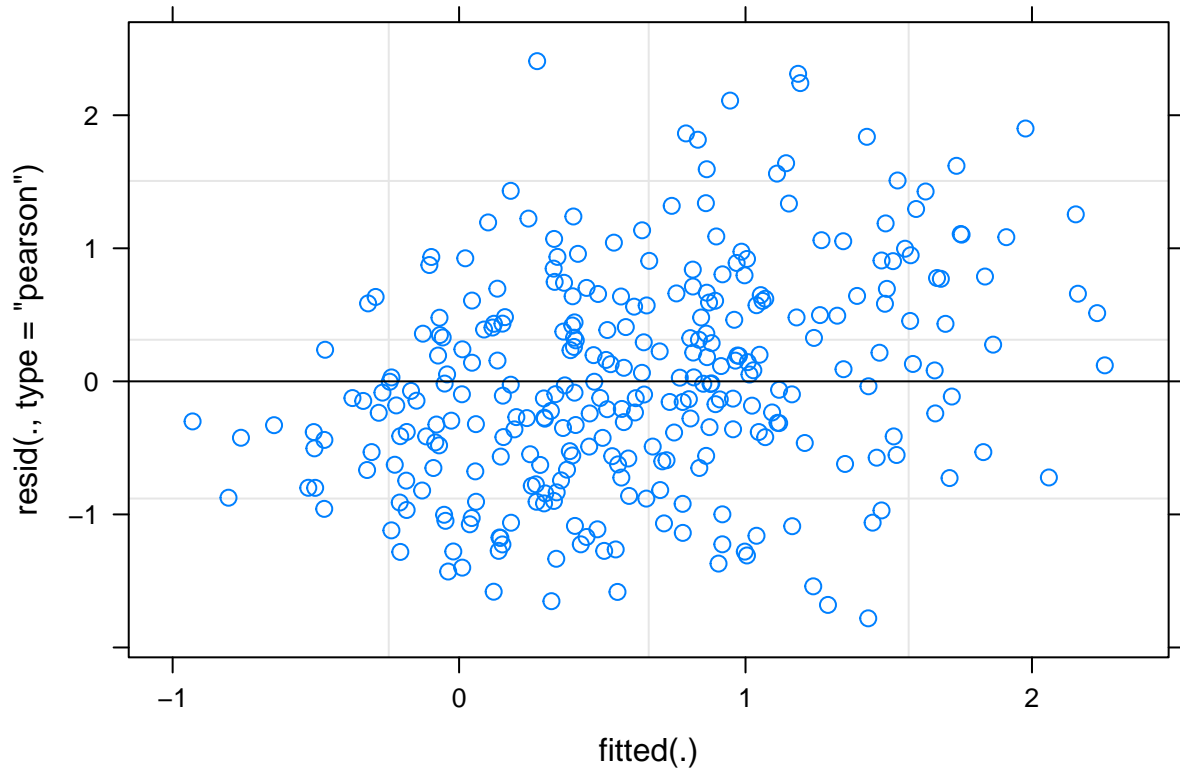
```
(exp( 0.02877 +0.01250)-1)*no2.1w
```

```
## 3rd Qu.
## 0.3630776
```

```
(exp( 0.02877 -0.01250)-1)*no2.1w
```

```
## 3rd Qu.
## 0.1413507
```

```
plot(mo3.1)
```



**Figure 5: diagnostic plot of model 3.1**

Figure 5 shows that the higher the fitted value, the higher the residue. However, in general it looks fine. 3.4 Build models for 2-week NO<sub>2</sub> exposure

```
mo4<- lmer(log(is.ad) ~ no2exp.2w+o3exp.2w+ pmexp.2w+so2exp.2w+Temp.2w+RHx.2w + last.ate +  
summary(mo4)
```

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method [  
## lmerModLmerTest]  
## Formula: log(is.ad) ~ no2exp.2w + o3exp.2w + pmexp.2w + so2exp.2w + Temp.2w +  
##      RHx.2w + last.ate + wkday.start + go.home + COLD + MNST +  
##      dt_smoke + (1 | Subject.ID)  
##      Data: dat1  
##  
## REML criterion at convergence: 946.3  
##  
## Scaled residuals:
```

```
##      Min      1Q   Median      3Q      Max
## -2.03734 -0.65274 -0.04204  0.61324  2.64864
##
## Random effects:
##   Groups      Name      Variance Std.Dev.
## Subject.ID (Intercept) 0.4961   0.7043
## Residual                0.8617   0.9283
## Number of obs: 302, groups: Subject.ID, 89
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)   -0.293745   2.831683 283.544175  -0.104    0.917
## no2exp.2w      0.066813   0.056690 283.012735   1.179    0.240
## o3exp.2w       0.040363   0.074186 278.563880   0.544    0.587
## pmexp.2w      -0.002124   0.005735 284.434919  -0.370    0.711
## so2exp.2w     -0.085834   0.104764 286.681042  -0.819    0.413
## Temp.2w       0.023248   0.120432 279.830238   0.193    0.847
## RHx.2w        -0.009879   0.036280 285.735687  -0.272    0.786
## last.ate      0.010671   0.013349 280.291066   0.799    0.425
## wkday.start   0.042621   0.084409 190.300335   0.505    0.614
## go.homewed    -0.252430   0.302338  79.849719  -0.835    0.406
## go.homeweekend -0.130340   0.343381  80.299526  -0.380    0.705
## COLDY         0.058713   0.204809 270.397059   0.287    0.775
## MNSTY        -0.482831   0.490378 266.959455  -0.985    0.326
## dt_smoke     -0.022624   0.032845 284.361134  -0.689    0.492
```

`step(mo4)`

```
## Backward reduced random-effect table:
##
##              Eliminated npar  logLik      AIC      LRT Df Pr(>Chisq)
## <none>                      16 -473.16  978.33
## (1 | Subject.ID)           0   15 -490.95 1011.90 35.572  1  2.458e-09 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Backward reduced fixed-effect table:
## Degrees of freedom method: Satterthwaite
##
##              Eliminated Sum Sq Mean Sq NumDF DenDF F value Pr(>F)
## Temp.2w         1  0.0321  0.0321     1 279.830  0.0373 0.8470709
## COLD            2  0.0678  0.0678     1 271.212  0.0790 0.7788488
## pmexp.2w        3  0.0718  0.0718     1 280.831  0.0839 0.7722626
## RHx.2w          4  0.1283  0.1283     1 259.795  0.1506 0.6982381
```

```
## go.home          5  0.5742  0.2871      2  83.645  0.3385 0.7138446
## o3exp.2w         6  0.1074  0.1074      1 270.074  0.1266 0.7222560
## dt_smoke         7  0.4478  0.4478      1 292.437  0.5298 0.4672622
## wkday.start      8  0.5832  0.5832      1 263.734  0.6961 0.4048516
## last.ate         9  0.7859  0.7859      1 278.303  0.9379 0.3336707
## MNST            10  0.8190  0.8190      1 274.894  0.9847 0.3219216
## no2exp.2w        0  4.5348  4.5348      1 221.626  5.4726 0.0202064 *
## so2exp.2w        0 10.7883 10.7883      1 222.380 13.0193 0.0003808 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Model found:
## log(is.ad) ~ no2exp.2w + so2exp.2w + (1 | Subject.ID)
```

```
mo4.1<- lmer(log(is.ad) ~ no2exp.2w + so2exp.2w + (1 | Subject.ID),data=dat1)
summary(mo4.1)
```

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method [
## lmerModLmerTest]
## Formula: log(is.ad) ~ no2exp.2w + so2exp.2w + (1 | Subject.ID)
##   Data: dat1
##
## REML criterion at convergence: 910.7
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -2.07871 -0.63093 -0.03594  0.59131  2.66937
##
## Random effects:
##   Groups      Name      Variance Std.Dev.
##   Subject.ID (Intercept) 0.5184   0.7200
##   Residual                0.8286   0.9103
## Number of obs: 302, groups: Subject.ID, 89
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)   0.07802    0.54464 232.36291   0.143 0.886220
## no2exp.2w     0.05572    0.02382 221.62605   2.339 0.020206 *
## so2exp.2w    -0.13645    0.03782 222.37980  -3.608 0.000381 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) n2xp.2
```

```
## no2exp.2w -0.904
## so2exp.2w 0.079 -0.471
```

```
anova(mo4.1)
```

```
## Type III Analysis of Variance Table with Satterthwaite's method
##           Sum Sq Mean Sq NumDF  DenDF F value    Pr(>F)
## no2exp.2w  4.5348   4.5348     1 221.63   5.4726 0.0202064 *
## so2exp.2w 10.7883  10.7883     1 222.38  13.0193 0.0003808 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
vif(mo4.1)
```

```
## no2exp.2w so2exp.2w
##  1.284372  1.284372
```

```
r.squaredGLMM (mo4.1)
```

```
##           R2m           R2c
## [1,] 0.02811885 0.4021322
```

```
#calculate IQR change
(exp(0.05572)-1)*no2.2w
```

```
## 3rd Qu.
## 0.156784
```

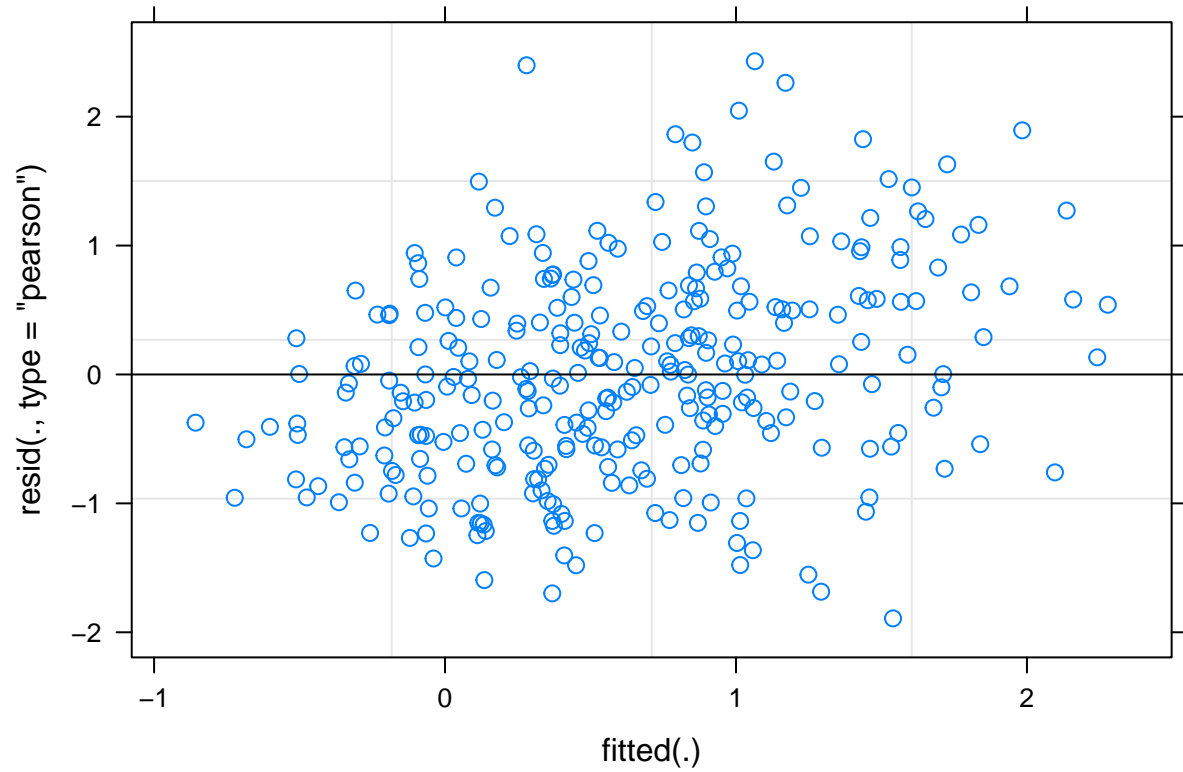
```
(exp(0.05572 +0.02382)-1)*no2.2w
```

```
## 3rd Qu.
## 0.2265202
```

```
(exp(0.05572 -0.02382)-1)*no2.2w
```

```
## 3rd Qu.
## 0.08868928
```

```
plot(mo4.1)
```



**Figure 6: diagnostic plot of model 4.1**

Figure 3 shows that the higher the fitted value, the higher the residue. However, in general it looks fine.

#### **4.Ploting Results 4.1 Relationship between log(8-isoprostane) and 12-hour NO2 exposure**



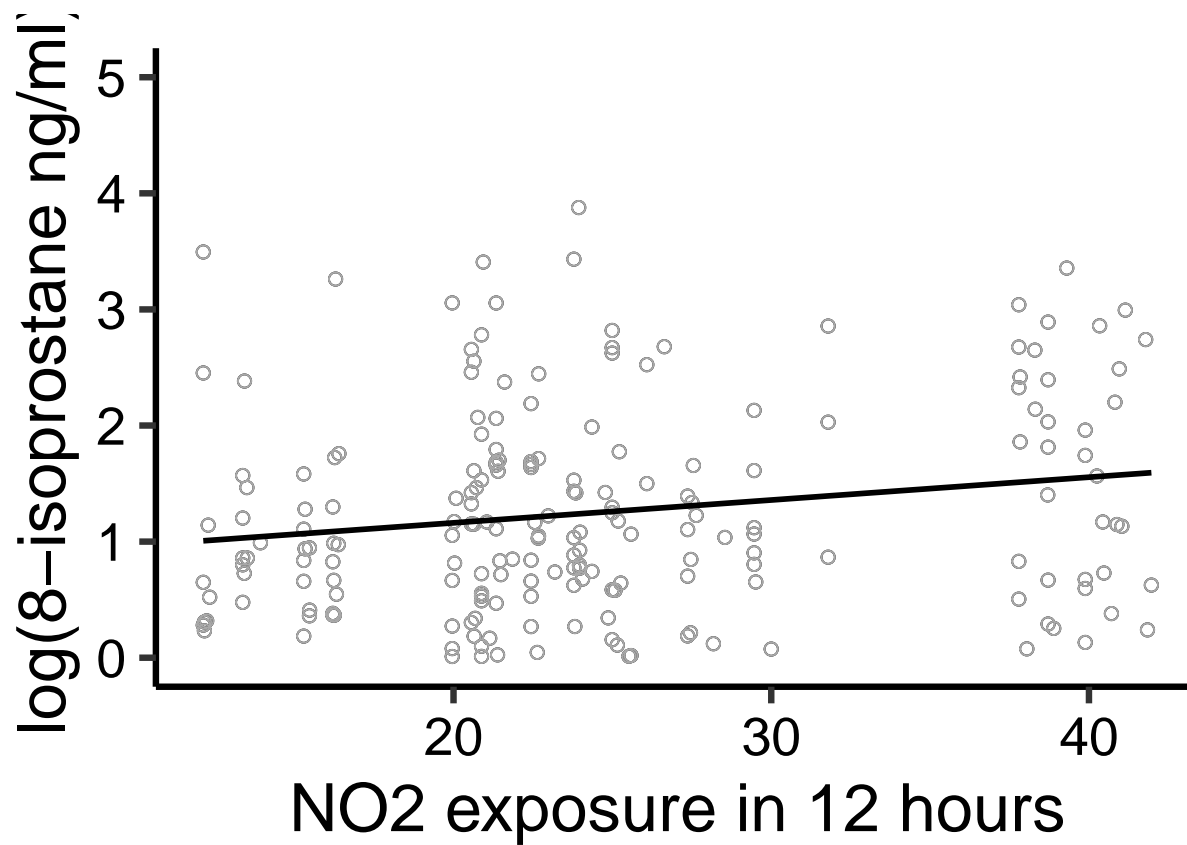
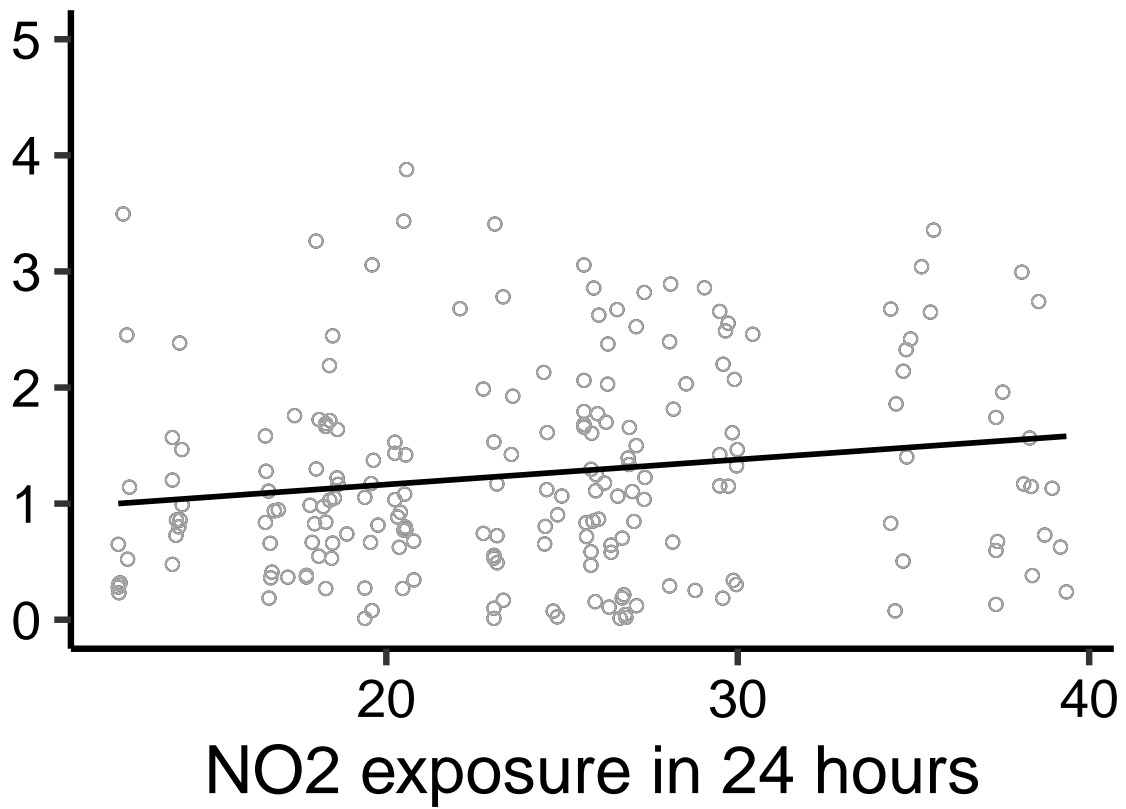


Figure 7: Relationship between log(8-isoprostane) and 12-hour NO2 exposure

Figure 7 shows that there is a positive relationship between 12-hour NO2 exposure and log(8-isoprostane).



**Figure 8:**Relationship between  $\log(8\text{-isoprostane})$  and 24-hour  $\text{NO}_2$  exposure

Figure 8 shows that there is a positive relationship between 24-hour  $\text{NO}_2$  exposure and  $\log(8\text{-isoprostane})$ .

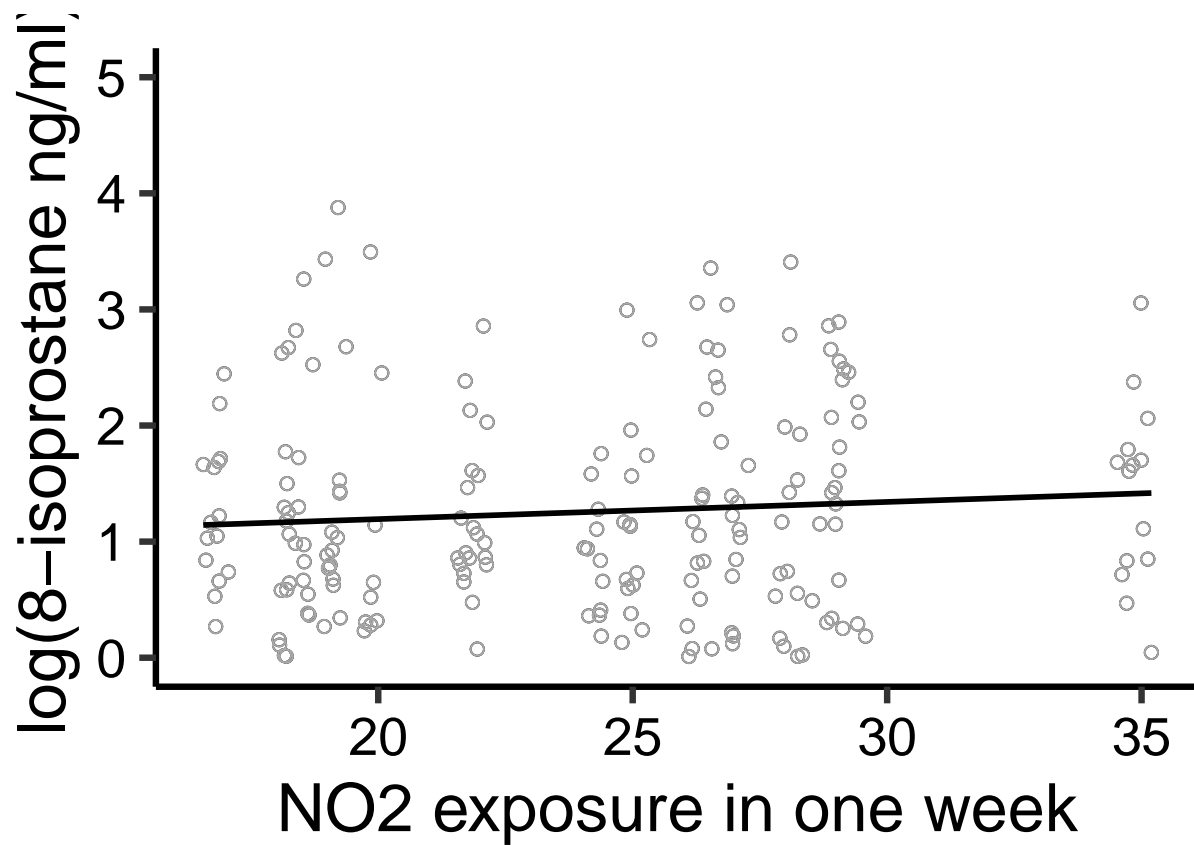
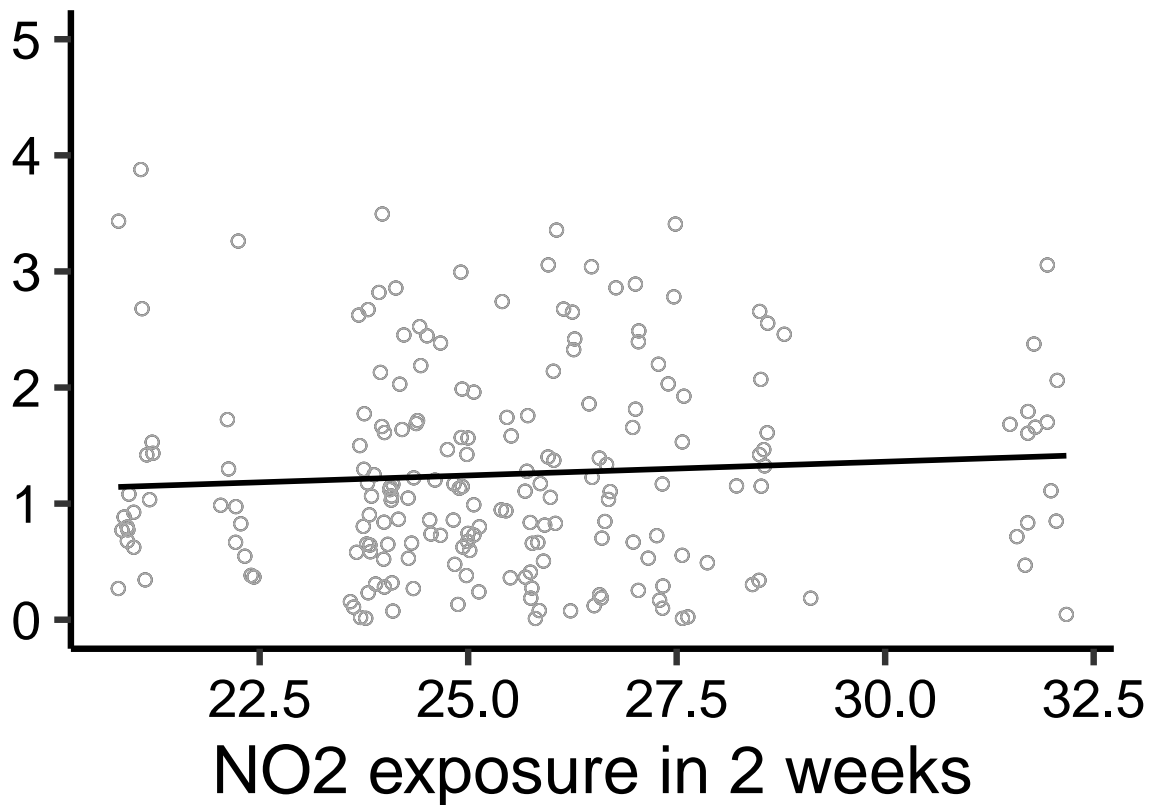


Figure 9: Relationship between log(8-isoprostane) and 1-week NO<sub>2</sub> exposure

Figure 9 shows that there is a positive relationship between 1-week NO<sub>2</sub> exposure and log(8-isoprostane).



**Figure 10: Relationship between log(8-isoprostane) and 2-week NO2 exposure**

Figure 10 shows that there is a positive relationship between 2-week NO2 exposure and log(8-isoprostane).

### 3 Exploratory Analysis

The Shapiro tests showed that concentrations of urinary 8-isoprostane is not normalized (p-value<0.001). I log-transformed it to put it into my models as the respondent variables.

### 4 Analysis

I used linear mixed models with participant-specific intercepts. The inclusion of participants as a variable in mixed-effects models account for the correlation of repeated measurements from the same individuals and precludes the need to control for participant characteristics (e.g., age, gender, BMI, the identity of smoker or non-smoker) that do not change across the four longitudinal measurements.

I used NO<sub>2</sub> exposure as predicting variables and log-transformed urinary biomarkers as dependent variables. For each biomarker, I built 4 models to the exposure over the periods of 12h, 24h, one week, and two weeks. I used a backward stepwise model selection method to select the co-variables for each model. The co-variables that I tested in the models were last meal, the start of the workday, respiratory infection status, menstruation, and the time of second-hand exposure. I also tested the SO<sub>2</sub> exposure, ozone exposure and PM<sub>2.5</sub> exposure, the average temperature and humidity during a corresponding period.

Both 12-hour, 24-hour, 1-week and 2-week NO<sub>2</sub> exposure showed significant correlations with the level of urinary 8-isoprostane, especially the 2-week NO<sub>2</sub> exposure. *One IQR (7.41 µg/m<sup>3</sup>) incremental change of 12-h NO<sub>2</sub> exposure was associated with an increase of 8-isoprostane level by 19.45% ( 95% CI: 14.37%, 24.55%, F value=14.99, p-value <0.001). The Adjusted R-squared =0.4121, which is the fraction of total variance explained by the model* One IQR (8.64 µg/m<sup>3</sup>) incremental change of 24-h NO<sub>2</sub> exposure was associated an increase in 8-isoprostane level by 27.69% ( 95% CI: 20.50%, 34.95% , F value=15.16, p-value <0.001 ). The Adjusted R-squared = 0.4160, which is the fraction of total variance explained by the model. *One IQR (4.47 µg/m<sup>3</sup>) incremental change of 1-week NO<sub>2</sub> exposure was associated with an increase in 8-isoprostane level by 25.15% ( 95% CI: 14.14%,36.31%, F value=5.30p-value <0.05 ). The Adjusted R-squared = 0.4014, which is the fraction of total variance explained by the model* One IQR (2.74 µg/m<sup>3</sup>) incremental change of 2-week NO<sub>2</sub> exposure was associated with an increase in 8-isoprostane level by 15.68% ( 95% CI: 8.87%,22.65%, F value=5.47p-value <0.05 ). The Adjusted R-squared = 0.4021, which is the fraction of total variance explained by the model.

## 5 Summary and Conclusions

Short term NO<sub>2</sub> exposure was associated with urinary 8-isoprostane. This finding meet my hypothesis. Urinary 8-isoprostane indicates the lipid peroxidation from the whole body, reflecting the systemic oxidative stress. It indicates short term NO<sub>2</sub> exposure can cause significant higher systemic oxidative stress.

## 6 References

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