Training in Genomics and Plant Breeding- GBios

RNA Seq Data Analysis

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1 | A little bit of context











Sesame world trade

- 2014 USD 2500 per tonnes
- 2017 USD 2300 million Import
- 2017 USD 2100 million Export
- 2020 USD 373.3 million (Lignans)

Where ? Origin | Native? | Introduced?







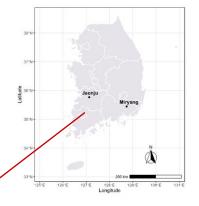
What

Wild **Variety**

Pedigree Landrace Characteristic Trait **Cultivar** Core collection

Mapping population (RILS, MAGIC, ...)





Federer Augmented Block Design

- Checks replicated 8 times

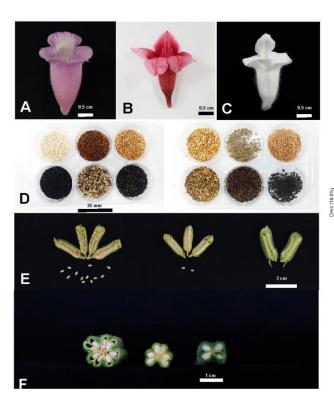




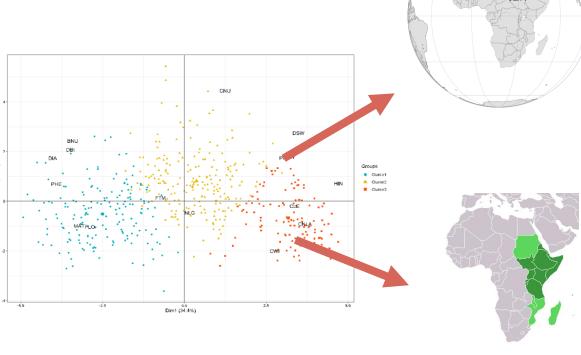
Total: 506 ccessions

Total: 24 traits

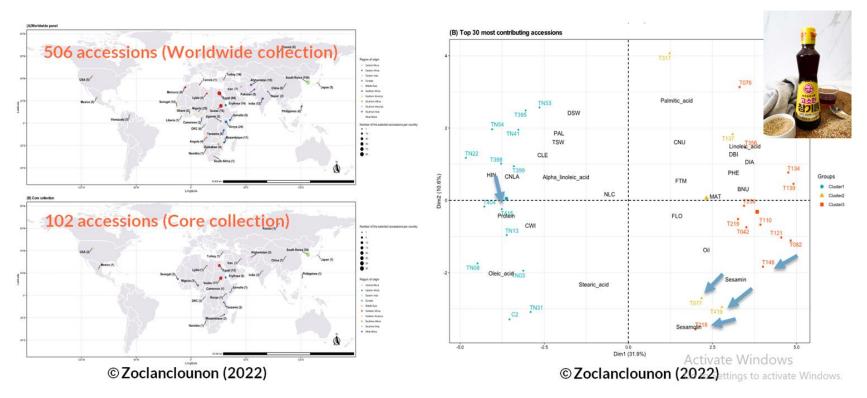
- Agronomic (18)
- Seed quality (06)
 - oil
 - fatty acids
 - sesamin
 - sesamolin



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Eastern and Northern Africa also contribute to the high yield accessions in the cluster 3



Crude Oil - Fatty acids [Palmitic, Stearic, Oleic, Linoleic, alpha linoleic] - TN03 & T415 Lignan [sesamin, sesamolin] - T218, T077, T419, & T148

Which omics concept have we covered so far?

Their utility

□ **Array-based:** Affymetrix axiom – Affimetrix GeneChip – Illumina Infinium Beadchip

Genotyping

□**NGS-based:** GBS – DArT-seq – RAD-seq – ddRAD – REST-Seq

Whole genome sequencing – Pangenomes – Structural Variations

Trait mapping: GWAS – QTL detection





Step 2: Generate genomic resources

Genome assembly
Genes SSRs QTL
Database SNPs
Annoted genome

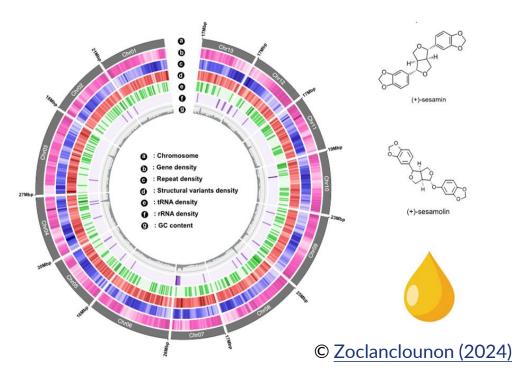
Molecular markers

Step 2: Generate genomic resources

1.16 ton per hectare | high oil content of 50.2% | sesamin : 3.96 mg/g sesamolin 2.57mg/g | Linoleic acid: 44.5%



Sesamum indicum cv Goenbaek



Circos plot of Goenbaek genome

Data: https://www.ncbi.nlm.nih.gov/bioproject/810203

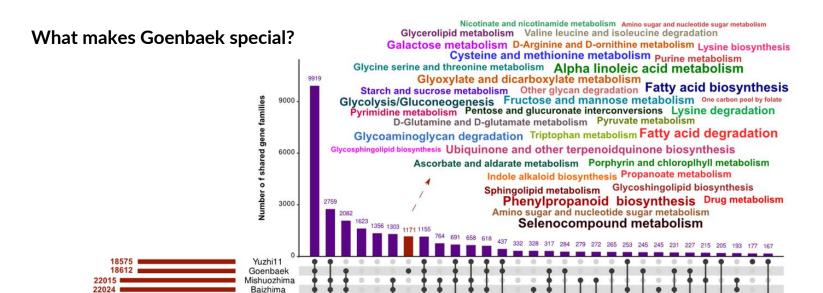
Step 2: Generate genomic resources

Swetha Zhongzhi13

20000

10000

Gene families number per species



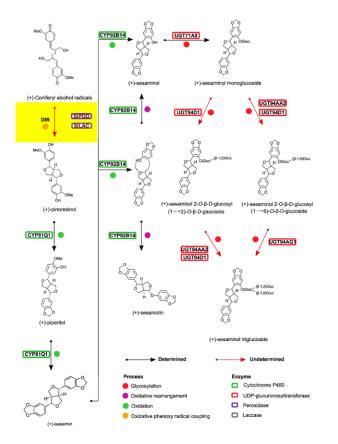




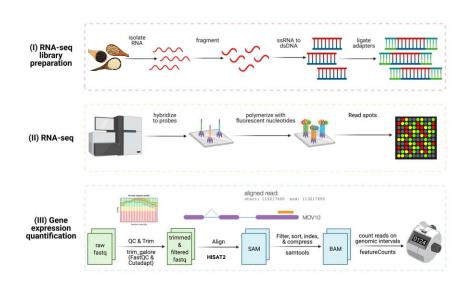
Step 3: Investigate key genes of interest



Step 3: Investigate key genes of interest



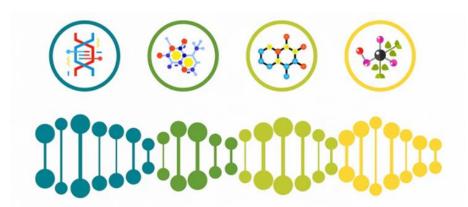
Transcriptomics in action



White vs Black | Rich oil vs Low oil

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2 | Data Acquisition



Data Acquisition | Metadata matters !!!

Variety name	Key characteristic	NCBI Project	SRA ID
ZZM4728	High oil content (59g/100g seed)	PRJNA400575	SRR6010085,SRR6010086,SRR6010087
ZZM2161	Low oil content (48g/100g seed)	PRJNA400575	SRR6010088,SRR6010089,SRR6010090

https://www.ebi.ac.uk/ena/browser/view/SRR6010085

https://www.ncbi.nlm.nih.gov/sra/?term=SRR6010085





Data Acquisition | srahunter

```
srahunter download -i accession_list.txt
```

Data Acquisition | srahunter

```
(srahunter_env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ vi accession_list.txt
(srahunter env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ srahunter download -i accession list.txt
Downloading with list: accession list.txt
Number of t: 6
Download path: /home/angeomics/data/tmp_srahunter
Max size: 50G
Output directory: /home/angeomics/data
Currently downloading: SRR6010085
The command used was: prefetch -p -X 50G SRR6010085 --output-file /home/angeomics/data/tmp srahunter/SRR6010085.sra
2024-10-30T18:30:23 prefetch.3.1.1: 1) Resolving 'SRR6010085'...
2024-10-30T18:30:25 prefetch.3.1.1: Current preference is set to retrieve SRA Normalized Format files with full base quality scores
2024-10-30T18:30:25 prefetch.3.1.1: 1) Downloading 'SRR6010085'...
2024-10-30T18:30:25 prefetch.3.1.1: SRA Normalized Format file is being retrieved
2024-10-30T18:30:25 prefetch.3.1.1: Downloading via HTTPS...
2024-10-30T18:42:10 prefetch.3.1.1: HTTPS download succeed
2024-10-30T18:42:18 prefetch.3.1.1: 'SRR6010085' is valid: 1750385155 bytes were streamed from 1750378499
2024-10-30T18:42:18 prefetch.3.1.1: 1) 'SRR6010085' was downloaded successfully
Generating fastg for: SRR6010085
The command used was: fasterg-dump --skip-technical -p -e 6 /home/angeomics/data/tmp_srahunter/SRR6010085.sra --outdir /home/angeomics/data
join : ------ 100% concat : ------ 100%
            : 13,305,758
spots read
reads read : 26,611,516
reads written : 26,611,516
Processing SRR6010085 completed successfully.
(srahunter env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ ls -rltha
total 7.6G
drwxr-x--- 7 angeomics angeomics 4.0K Oct 30 18:29 ...
-rw-r--r-- 1 angeomics angeomics   11 Oct 30 18:29 accession_list.txt
drwxr-xr-x 2 angeomics angeomics 4.0K Oct 30 18:42 tmp_srahunter
-rw-r--r-- 1 angeomics angeomics 3.8G Oct 30 18:45 SRR6010085_2.fastq
-rw-r--r-- 1 angeomics angeomics 3.8G Oct 30 18:46 SRR6010085_1.fastq
drwxr-xr-x 3 angeomics angeomics 4.0K Oct 30 18:46
(srahunter env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$
```

Line 1 (@SRR6010085.1 FCC0CF3ACXX:8:1101:1600:2212 length=90):

- Starts with @, marking it as the header line.
- SRR6010085.1: The read identifier, which includes the SRA accession number (SRR6010085) and a unique read number (.1).
- FCC0CF3ACXX:8:1101:1600:2212: Details about the sequencing run and position of the read in the Illumina flow cell:
 - FCC0CF3ACXX: Flow cell ID.
 - 8: Lane number.
 - 1101:1600:2212: X and Y coordinates of the cluster within the flow cell.
- length=90: Specifies the read length in base pairs.

Line 2 (GTTTGAT...TGAATT):

Contains the DNA sequence for this read.

- •Line 3 (+SRR6010085.1 FCC0CF3ACXX:8:1101:1600:2212 length=90):
- •Begins with a + symbol, marking it as the separator line.
- •Optionally, this line repeats the read identifier, but it can be left blank.

Line 4 (CCCFFFF...DECD):

•Shows the quality scores for each base in the sequence. Each character corresponds to a base in Line 2 and represents its quality score (often in ASCII, with F and higher generally indicating high quality).

Click https://www.drive5.com/usearch/manual/quality_score.html) to learn about quality scores

Task: How many reads we have in each fastq file?

```
| angeomics@DESKTOP-UB182! × + | v | |
| (base) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ grep "@SRR" SRR6010085_1.fastq | wc -l |
| 13305758 | (base) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ grep "@SRR" SRR6010085_2.fastq | wc -l |
| 13305758 | (base) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ |
```

https://www.ncbi.nlm.nih.gov/sra/?term=SRR6010085

Data Acquisition | fastqc

```
fastqc -t 8 -o fastqc_dir/ SRR6010085_*.fastq
```

Data Acquisition | fastqc

```
(fastqc_env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ fastqc -t 8 -o fastqc_dir/ SRR6010085_*.fastq
null
null
Started analysis of SRR6010085_1.fastq
Started analysis of SRR6010085_2.fastq
Approx 5% complete for SRR6010085_1.fasto
Approx 5% complete for SRR6010085_2.fastq
Approx 10% complete for SRR6010085_1.fastq
 Approx 10% complete for SRR6010085_2.fastq
 Approx 15% complete for SRR6010085_1.fastq
 Approx 15% complete for SRR6010085_2.fastq
 Approx 20% complete for SRR6010085_1.fastq
 Approx 20% complete for SRR6010085_2.fastq
 Approx 25% complete for SRR6010085_2.fastq
 Approx 25% complete for SRR6010085_1.fastq
 Approx 30% complete for SRR6010085_2.fastg
 Approx 30% complete for SRR6010085_1.fastq
 Approx 35% complete for SRR6010085_2.fastg
 Approx 35% complete for SRR6010085_1.fastq
 Approx 40% complete for SRR6010085_2.fastq
 Approx 40% complete for SRR6010085_1.fastq
 Approx 45% complete for SRR6010085_2.fastq
 Approx 45% complete for SRR6010085 1.fasto
 Approx 50% complete for SRR6010085_2.fastq
 Approx 50% complete for SRR6010085_1.fastq
 Approx 55% complete for SRR6010085 2.fasto
 Approx 55% complete for SRR6010085_1.fastq
 Approx 60% complete for SRR6010085_2.fastq
 Approx 60% complete for SRR6010085_1.fastq
 Approx 65% complete for SRR6010085_2.fastq
Approx 65% complete for SRR6010085_1.fastq
 Approx 70% complete for SRR6010085_2.fastq
 Approx 70% complete for SRR6010085_1.fastq
Approx 75% complete for SRR6010085_2.fastq
Approx 75% complete for SRR6010085_1.fastq
 Approx 80% complete for SRR6010085_2.fastq
Approx 80% complete for SRR6010085_1.fastq
Approx 85% complete for SRR6010085_2.fastq
Approx 85% complete for SRR6010085_1.fastq
 Approx 90% complete for SRR6010085_2.fastq
 Approx 90% complete for SRR6010085_1.fastq
 Approx 95% complete for SRR6010085_2.fastq
Approx 95% complete for SRR6010085_1.fastq
Analysis complete for SRR6010085_2.fastq
Analysis complete for SRR6010085_1.fastq
(fastgc_env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$
```

Please check the report

Data Cleaning | fastp

```
fastp

fastp --detect_adapter_for_pe \
    --overrepresentation_analysis \
    --correction --cut_right --thread 2 \
    --html trimmed_dir/SRR6010085.fastp.html \
    --json trimmed_dir/SRR6010085.fastp.json \
    -i SRR6010085_1.fastq -I SRR6010085_2.fastq \
    -o trimmed_dir/SRR6010085_1.fastq -0 trimmed_dir/SRR6010085_2.fastq
```

Data Cleaning | fastp

```
angeomics@DESKTOP-UB182: X + V
 (fastp_env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ fastp --detect_adapter_for_pe \
        --overrepresentation_analysis \
        --correction --cut_right --thread 2 \
        --html trimmed_dir/SRR6010085.fastp.html --json trimmed_dir/SRR6010085.fastp.json \
        -i SRR6010085_1.fastq -I SRR6010085_2.fastq \
        -o trimmed_dir/SRR6010085_1.fastq -O trimmed_dir/SRR6010085_2.fastq
Detecting adapter sequence for read1...
AAAGGCTTACGGTGGATACCTAGGCACCCAGAGACGAGAAGGGCGTAGTAATCGACGAA
Detecting adapter sequence for read2...
No adapter detected for read2
Read1 before filtering:
total reads: 13305758
total bases: 1197518220
Q20 bases: 1157527932(96.6606%)
030 bases: 1073681705(89.6589%)
Read2 before filtering:
total reads: 13305758
total bases: 1197518220
Q20 bases: 1102927865(92.1011%)
030 bases: 996543643(83.2174%)
Read1 after filtering:
total reads: 12402218
total bases: 1059211267
Q20 bases: 1050942053(99.2193%)
030 bases: 997961572(94.2174%)
Read2 after filtering:
total reads: 12402218
total bases: 1008532481
020 bases: 995490379(98.7068%)
Q30 bases: 928508411(92.0653%)
Filtering result:
reads passed filter: 24804436
reads failed due to low quality: 1742
reads failed due to too many N: 0
reads failed due to too short: 1805338
reads with adapter trimmed: 192723
bases trimmed due to adapters: 6579734
reads corrected by overlap analysis: 4204
bases corrected by overlap analysis: 4314
Duplication rate: 28.224%
Insert size peak (evaluated by paired-end reads): 90
JSON report: trimmed_dir/SRR6010085.fastp.json
HTML report: trimmed_dir/SRR6010085.fastp.html
fastp --detect_adapter_for_pe --overrepresentation_analysis --correction --cut_right --thread 2 --html trimmed_dir/SRR6010085.fastp.html --json trimmed_dir/SRR6010085.fastp.json -i SRR6010085_1.fastq -I SRR6010085_2.fastq -o trimmed_dir/SRR6010085_1.
/SRR6010085_1.fastg -0 trimmed_dir/SRR6010085_2.fastg
fastp v0.23.4, time used: 278 seconds
 (fastp_env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$
```

Data Cleaning | FastQC on trimmed data

```
FastQC on trimmed data

fastqc -t 8 -o fastqc_trimmed_dir/ trimmed_dir/SRR6010085_*.fastq
```

Data Cleaning | FastQC on trimmed data

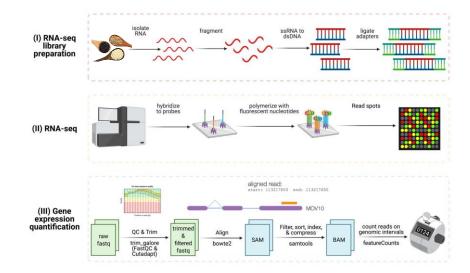
```
angeomics@DESKTOP-UB182! X
(fastqc_env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$ fastqc -t 8 -o fastqc_trimmed_dir/ trimmed_dir/SRR6010085_*.fastq
null
Started analysis of SRR6010085_1.fastq
Started analysis of SRR6010085_2.fastq
Approx 5% complete for SRR6010085_1.fastq
Approx 5% complete for SRR6010085_2.fastq
Approx 10% complete for SRR6010085_2.fastq
Approx 10% complete for SRR6010085 1.fastq
Approx 15% complete for SRR6010085_2.fastq
Approx 15% complete for SRR6010085 1.fastq
Approx 20% complete for SRR6010085_2.fastq
Approx 20% complete for SRR6010085_1.fastq
Approx 25% complete for SRR6010085_2.fastq
Approx 25% complete for SRR6010085 1.fastq
Approx 30% complete for SRR6010085_2.fastq
Approx 30% complete for SRR6010085_1.fastq
Approx 35% complete for SRR6010085_2.fastq
Approx 35% complete for SRR6010085_1.fastq
Approx 40% complete for SRR6010085_2.fastq
Approx 40% complete for SRR6010085 1.fastq
Approx 45% complete for SRR6010085_2.fastq
Approx 45% complete for SRR6010085_1.fastq
Approx 50% complete for SRR6010085_2.fastq
Approx 50% complete for SRR6010085_1.fastq
Approx 55% complete for SRR6010085_2.fastq
Approx 55% complete for SRR6010085_1.fastq
Approx 60% complete for SRR6010085_2.fastq
Approx 60% complete for SRR6010085_1.fastq
Approx 65% complete for SRR6010085_2.fastq
Approx 65% complete for SRR6010085 1.fastq
Approx 70% complete for SRR6010085_2.fastq
Approx 70% complete for SRR6010085 1.fastq
Approx 75% complete for SRR6010085_2.fastq
Approx 75% complete for SRR6010085_1.fastq
Approx 80% complete for SRR6010085_2.fastq
Approx 80% complete for SRR6010085_1.fastq
Approx 85% complete for SRR6010085_2.fastq
Approx 85% complete for SRR6010085_1.fastq
Approx 90% complete for SRR6010085_2.fastq
Approx 90% complete for SRR6010085_1.fastq
Approx 95% complete for SRR6010085_2.fastq
Approx 95% complete for SRR6010085_1.fastq
Analysis complete for SRR6010085 2.fastg
Analysis complete for SRR6010085_1.fastq
(fastqc_env) angeomics@DESKTOP-UB1829G:/home/angeomics/data$
```

Data Cleaning | MultiQC

```
MultiQC
multiqc fastqc_trimmed_dir/ trimmed_dir/ --outdir multiqc_dir
```

Data Cleaning | MultiQC

3 | Alignment



Alignment with HISAT2 | Genome Data

```
Assembly and annotation data: <a href="https://zenodo.org/records/6350881">https://zenodo.org/records/6350881</a>
```

```
Source: https://www.researchsquare.com/article/rs-4887813/v1
```

- Assembly: Sesamum_indicum_goenbaek.fasta
- Annotation: Sesamum_indicum_goenbaek.gff3

Alignment with HISAT2 | Build index

hisat2-build Sesamum_indicum_goenbaek.fasta Sesamum_indicum_goenbaek

```
angeomics@DESKTOP-UB182! X
(hisat2_env) angeomics@DESKTOP-UB1829G:/home/angeomics/analyses$ ls -rlth
total 669M
-rw-r--r-- 1 angeomics angeomics 255M Feb 22 2022 Sesamum_indicum_goenbaek.fasta
-rw-r--r- 1 angeomics angeomics 28M Mar 14 2022 Sesamum_indicum_goenbaek.gff3
-rw-r--r-- 1 angeomics angeomics 63M Oct 30 21:44 Sesamum_indicum_goenbaek.4.ht2
-rw-r--r 1 angeomics angeomics 1.8K Oct 30 21:44 Sesamum_indicum_goenbaek.3.ht2
-rw-r--r-- 1 angeomics angeomics
                                   8 Oct 30 21:44 Sesamum_indicum_goenbaek.8.ht2
-rw-r--r-- 1 angeomics angeomics 12 Oct 30 21:44 Sesamum_indicum_goenbaek.7.ht2
-rw-r--r- 1 angeomics angeomics 63M Oct 30 21:47 Sesamum_indicum_goenbaek.2.ht2
-rw-r--r-- 1 angeomics angeomics 88M Oct 30 21:47 Sesamum_indicum_goenbaek.1.ht2
-rw-r--r-- 1 angeomics angeomics 64M Oct 30 21:47 Sesamum_indicum_goenbaek.6.ht2
-rw-r--r- 1 angeomics angeomics 110M Oct 30 21:47 Sesamum_indicum_goenbaek.5.ht2
(hisat2_env) angeomics@DESKTOP-UB1829G:/home/angeomics/analyses$
```

Alignment with HISAT2 | Mapping

```
hisat2 \
-p 8 \
-x Sesamum_indicum_goenbaek \
-1 /home/angeomics/data/01.SRA/trimmed_dir/SRR6010085_1.fastq \
-2 /home/angeomics/data/01.SRA/trimmed_dir/SRR6010085_2.fastq \
-S SRR6010085.bam
```

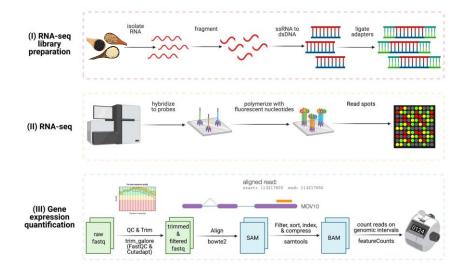
```
angeomics@DESKTOP-UB182! X
(hisat2_env) angeomics@DESKTOP-UB1829G:/home/angeomics/data/01.SRA/trimmed_dir/SRR6010085_1.fastq -2 /home/angeomics/data/01.SRA/trimmed_dir/SRR6010085_2.fastq -5 SRR6010085.bam
12402218 reads; of these:
 12402218 (100.00%) were paired; of these:
   2715669 (21.90%) aligned concordantly 0 times
   9387390 (75.69%) aligned concordantly exactly 1 time
   299159 (2.41%) aligned concordantly >1 times
   2715669 pairs aligned concordantly 0 times; of these:
     25232 (0.93%) aligned discordantly 1 time
   2690437 pairs aligned 0 times concordantly or discordantly; of these:
     5380874 mates make up the pairs: of these:
       5167736 (96.04%) aligned 0 times
       201644 (3.75%) aligned exactly 1 time
       11494 (0.21%) aligned >1 times
79.17% overall alignment rate
(hisat2_env) angeomics@DESKTOP-UB1829G:/home/angeomics/analyses$
```

Alignment with HISAT2 | Mapping

samtools view --threads 8 -bS -o SRR6010085.bam SRR6010085.sam

```
angeomics@DESKTOP-UB182! X
(samtools_env) angeomics@DESKTOP-UB1829G:/home/angeomics/analyses$ samtools view --threads 8 -bS -o SRR6010085.bam SRR6010085.sam
(samtools_env) angeomics@DESKTOP-UB1829G:/home/angeomics/analyses$ ls -rlth
total 9.9G
-rw-r--r-- 1 angeomics angeomics 255M Feb 22 2022 Sesamum_indicum_goenbaek.fasta
-rw-r--r- 1 angeomics angeomics 28M Mar 14 2022 Sesamum_indicum_goenbaek.qff3
-rw-r--r-- 1 angeomics angeomics 63M Oct 30 21:44 Sesamum_indicum_goenbaek,4.ht2
-rw-r--- 1 angeomics angeomics 1.8K Oct 30 21:44 Sesamum_indicum_goenbaek.3.ht2
-rw-r-- 1 angeomics angeomics 8 Oct 30 21:44 Sesamum_indicum_goenbaek.8.ht2
-rw-r--r-- 1 angeomics angeomics 12 Oct 30 21:44 Sesamum_indicum_goenbaek.7.ht2
-rw-r--- 1 angeomics angeomics 63M Oct 30 21:47 Sesamum_indicum_goenbaek.2.ht2
-rw-r--r-- 1 angeomics angeomics 88M Oct 30 21:47 Sesamum_indicum_goenbaek.1.ht2
-rw-r--r-- 1 angeomics angeomics 64M Oct 30 21:47 Sesamum_indicum_goenbaek.6.ht2
-rw-r--r-- 1 angeomics angeomics 110M Oct 30 21:47 Sesamum_indicum_goenbaek.5.ht2
-rw-r--r-- 1 angeomics angeomics 0 Oct 30 22:10 Sesamum_indicum_goenbaek
-rw-r--r 1 angeomics angeomics 7.1G Oct 30 22:23 SRR6010085.sam
-rw-r--r-- 1 angeomics angeomics 2.2G Oct 31 10:57 SRR6010085.bam
(samtools_env) angeomics@DESKTOP-UB1829G:/home/angeomics/analyses$
```

4: Abundance estimation



Abundance count | featuresCounts

```
featureCounts \
-T 4 -s 2 -p -t gene \
-g ID -a Sesamum_indicum_goenbaek.gff3 \
-o SRR6010085.gene.txt SRR6010085.bam
```

```
angeomics@DESKTOP-UB182: X + V
(subread_env) angeomics@DESKTOP-UB1829G:/home/angeomics/analyses$ featureCounts -T 4 -s 2 -p -t gene -q ID -a Sesamum_indicum_goenback.gff3 -o SRR6010085.gene.txt SRR6010085.ban
             Input files : 1 BAM file
             Output file : SRR6010085.gene.txt
                 Sunnary : SRR6010085.gene.txt.summary
              Paired-end : yes
        Count read pairs : no
      Annotation : Sesamum_indicum_goenbaek.gff3 (GTF)
Dir for temp files : ./
                  Threads : 4
      Multimapping reads : not counted
  Multi-overlapping reads : not counted
  Load annotation file Sesanum indicum goenback.gff3 ...
    Features: 23539
    Meta-features : 23539
    Chromosomes/contigs : 13
  Process BAM file SRR6010085.bam..
    Strand specific : reversely stranded
    Paired-end reads are included
    The reads are assigned on the single-end mode.
    Total alignments: 2635
    Successfully assigned alignments : 6873644 (26.1%)
    Running time : 0.30 minutes
  Write the final count table.
  Write the read assignment summary
   Summary of counting results can be found in file "SRR6010085.gene.txt.sum
(subread_env) angeomics@DESKTOP-UB1829G:/hone/angeomics/analyses$
```

Task: Create a loop for the mapping and the abundance estimation stages

Task: Create a loop for the mapping step

```
REFERENCE="Sesamum_indicum_goenbaek.fasta"
INDEX_NAME="Sesamum_indicum_goenbaek"
if [ ! -f "${INDEX_NAME}.1.ht2" ]; then
 echo "Building HISAT2 index..."
 hisat2-build "$REFERENCE" "$INDEX_NAME"
 echo "Index building completed."
 echo "Index files found, skipping index building."
FASTO DIR="/home/angeomics/data/01.SRA/trimmed dir"
for FILE in ${FASTQ_DIR}/*_1.fastq; do
  SAMPLE_NAME=$(basename "$FILE" _1.fastq)
  FASTQ1="${FASTQ_DIR}/${SAMPLE_NAME}_1.fastq"
  FASTO2="${FASTO DIR}/${SAMPLE NAME} 2.fastg"
  SAM_FILE="${SAMPLE_NAME}.sam"
  BAM_FILE="${SAMPLE_NAME}.bam"
  echo "Mapping reads for $SAMPLE_NAME..."
  hisat2 -p $THREADS -x "$INDEX_NAME" -1 "$FASTQ1" -2 "$FASTQ2" -S "$SAM_FILE"
  echo "Mapping completed for $SAMPLE_NAME."
  echo "Converting $SAM_FILE to BAM format..."
  samtools view --threads $THREADS -bS -o "$BAM_FILE" "$SAM_FILE"
  rm "$SAM_FILE"
 echo "BAM conversion completed for $SAMPLE_NAME."
```

Task: Create a loop for the abundance estimation stage

```
...
                                Abundance Estimation Loop
GFF3="Sesamum_indicum_goenbaek.gff3"
THREADS=4
for BAM_FILE in *.bam; do
 SAMPLE_NAME=$(basename "$BAM_FILE" .bam)
  # Run featureCounts for each BAM file
  featureCounts \
   -T $THREADS \
   -s 2 \
    -p \
    -t gene \
    -g ID \
    -a "$GFF3" \
    -o "${SAMPLE_NAME}.gene.txt" \
    "$BAM FILE"
 echo "Abundance estimation completed for $BAM_FILE"
```

Make a table of gene count

```
Make a gene count table
paste <(awk 'BEGIN {OFS="\t"} {print $1,$7}' SRR6010085.gene.txt) \</pre>
  <(awk 'BEGIN {OFS="\t"} {print $7}' SRR6010086.gene.txt) \</pre>
  <(awk 'BEGIN {OFS="\t"} {print $7}' SRR6010087.gene.txt) \</pre>
  <(awk 'BEGIN {OFS="\t"} {print $7}' SRR6010088.gene.txt) \</pre>
  <(awk 'BEGIN {OFS="\t"} {print $7}' SRR6010089.gene.txt) \</pre>
  <(awk 'BEGIN {OFS="\t"} {print $7}' SRR6010090.gene.txt) | \</pre>
  grep -v '^\#' > sesame_count.txt
```

5: DEG analysis with DIANE Package

https://diane.bpmp.inrae.fr/

5: DEG analysis with DIANE Package

DIANE Package enable post counting analyses covering differentially expressed gene, coexpression analyses and regulatory network inference.

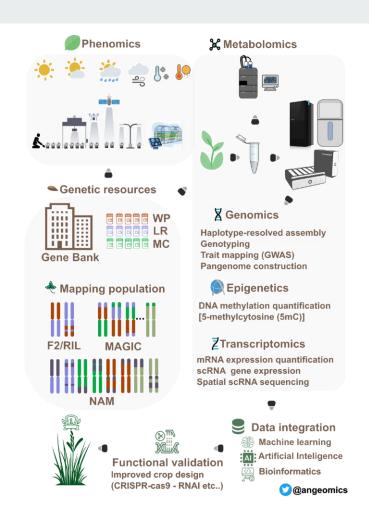
We will follow the documentation at: https://github.com/OceaneCsn/DIANE

Bonus

□Omics enables fast-forward breeding for a

food-secure world

- **□**Genetic diversity is a paramount
- □Big data Bioinformatics Machine learning
- ☐ Genetic engineering Gene editing



Thank you







