ELSEVIER

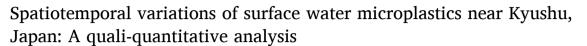
Contents lists available at ScienceDirect

Marine Pollution Bulletin

journal homepage: www.elsevier.com/locate/marpolbul



Baseline



Tsunefumi Kobayashi ^{a,1}, Mitsuharu Yagi ^{a,b,*,1}, Toshiya Kawaguchi ^a, Toshiro Hata ^c, Kenichi Shimizu ^{a,b}

- a Department of Fisheries Science, Graduate School of Fisheries and Environmental Sciences, Nagasaki University, 8528521 Bunkyo, Nagasaki, Japan
- ^b Institute of Integrated Science and Technology, Nagasaki University, 8528521 Bunkyo, Nagasaki, Japan
- ^c Civil and Environmental Engineering Program, Graduate School of Advanced Science and Engineering, Hiroshima University, 7398527 Higashi-Hiroshima, Hiroshima, Japan

ARTICLE INFO

Keywords: Microplastics East China Sea Neuston net FTIR-ATR Seasonal variability

ABSTRACT

Microplastics in the ocean are threatening marine ecosystems. Although plastic contaminants are ubiquitous, their distribution is thought to be heterogeneous. Here, we elucidate the spatial and temporal variations in the quanti-qualitative characteristics of microplastics near Kyushu, Japan in the East China Sea. Six surveys across nine stations were conducted over a 14-month period, and a total of 6131 plastic items were identified. The average microplastic abundance and size were 0.49 ± 0.92 (items·m⁻³ \pm S.D.), and 1.71 ± 0.93 (mm \pm S.D.), respectively. Differences between the highest and lowest abundances were 50-fold among monthly means, and 550-fold across all net tows. With respect to colour, polymer type, and shape, white and transparent polyethylene fragments were the dominant composition. There were significant differences for each of the analytical microplastic parameters among the survey months. Our results provide baseline data and lead to a more comprehensive understanding of the spatiotemporal characteristics of microplastic pollution.

Oceanic plastic pollution is a global-scale issue. Plastic products have been an indispensable part of modern society since the 1940s and 50s due to their favourable properties, such as low cost, light weight, high durability, and ease of design (Cole et al., 2011; Hammer et al., 2012). Approximately 80% of all manufactured, mass-produced plastics to date, however, have accumulated in landfills or natural environments (Geyer et al., 2017), and inappropriate management of waste plastics can lead to their transfer to marine environments (Gregory, 2009; Ryan et al., 2009; Kuroda et al., 2020). Approximately 10% of the produced plastics accumulate and persist in the marine ecosystems (Thompson, 2006), with an estimated 1.15-2.41 million tons introduced into the ocean via the river system every year (Jambeck et al., 2015; Lebreton et al., 2017). If current waste management trajectories are maintained, an additional ~12,000 million metric tons of plastic waste will be deposited into landfills or the natural environment by 2050 (Geyer et al., 2017), and the amount of plastic in the ocean will surpass that of fish by weight (World Economic Forum, 2016).

Plastic debris in the ocean is deteriorated by ultraviolet rays and

waves, and when it degrades to a size of \leq 5 mm, it is commonly referred to as microplastic (MP; Andrady, 2011). MPs are ubiquitous in the global oceans, and readily accumulate in marine ecosystems due to their small size and ease of ingestion by various marine organisms (De Witte et al., 2014; Auta et al., 2017). Once ingested, toxic substances amass inside an organism and lead to intestinal blockage or physical damage (Jovanović, 2017); therefore, negative impacts on marine organisms can be precluded by concentrating on MP management.

Drifting MPs on the sea surface are heterogeneously distributed throughout the globe, and positively correlate with epicentres of dumping in the Southeast Asian countries, including China (Jambeck et al., 2015). Accordingly, Isobe et al. (2015) reported total MP particle concentrations in the East Asian seas around Japan that were 16 and 27 times greater than those of the North Pacific Ocean and the world's oceans, respectively. Furthermore, marine MPs can vary with the seasons or currents (Moore, 2008; Martinez et al., 2009; Doyle et al., 2011); hence, studies with inadequate transect numbers and/or over short time scales fail to capture the spatiotemporal patterns in abundance and

^{*} Corresponding author at: Department of Fisheries Science, Graduate School of Fisheries and Environmental Sciences, Nagasaki University, 8528521 Bunkyo, Nagasaki, Japan.

E-mail address: yagi-m@nagasaki-u.ac.jp (M. Yagi).

 $^{^{1}\,}$ These authors contributed equally to this work.

characteristics (Ryan et al., 2009; Cole et al., 2011). Information on the long-term quantitative (abundance and size) and qualitative (shape, colour, and polymer) characteristics of MPs is limited, even though such studies would provide valuable pattern and baseline data for all future research (Cole et al., 2011). Additionally, data on MPs in the waters off Kyushu, Japan, which is a purported highly polluted area, are lacking. It was hypothesised in this study here that the significant spatiotemporal changes in abundance and type of sea surface MPs could be detected by long-term surveys; thus, repeated sea surface surveys over 14 months in the East China Sea were conducted to determine abundance, size, and type of MPs, and obtain the first regional assessment of spatiotemporal variations for this contaminant.

Six sea surface surveys were carried out from April 2019 to June 2020 using the training vessel T/V Kakuyo-maru (155 gross tonnage: Faculty of Fisheries, Nagasaki University), with each survey including nine sampling stations (Fig. 1; Supplementary Table S1). To avoid the possible effects of tides on MPs abundance by tide, i.e., the spring and neap tides, sampling was carried out around the spring tide, except for April 2019 and June 2020. Although each sampling was carried out in one day, we could not collect each sample just at the same tide times, i. e., flood and low tide, due to technical difficulties. A total of 54 samples were collected using a neuston net (JMA, RIGO Co., Ltd., Tokyo, Japan; rectangular mouth opening, 0.75 m × 0.75 m; length, 3 m; mesh size, $350 \mu m$) originally designed for sampling neustonic organisms such as zooplankton, fish larvae, and eggs near the sea surface (Isobe et al., 2015). According to the net mesh size, MPs ranging from 350 μm to 5 mm were targeted. To measure the volume of water that passed through during sampling, a flow meter (5571A, RIGO Co., Ltd., Tokyo, Japan) was attached to the net mouth. The net was towed from a boom installed at the bow-port side of the vessel to ensure that it was kept 5 m from the ship's side and prevent debris disturbance by the bow wave. The net was towed around each station for \sim 10 min at a vessel log speed of 2.0 knots (Doyle et al., 2011; GESAMP, 2019). Significant wave height, wind speed, water temperature, and salinity during sampling were recorded by the onboard monitoring system (Supplementary Fig. S1). After sample retrieval, nets were rinsed with filtered sea water to ensure that all debris and organisms were attached to the cod end. Contents of the cod end were then carefully put into a sample bottle (2000 ml), and when debris larger than 100 mm was collected, the surface of the debris was rinsed using filtered water to collect any present MPs in a 350 μm strainer net prior to removal. The samples were preserved with a 5% formalin solution buffered with sodium borate, and stored at room temperature until sorting.

The volume of filtered water $V(m^3)$ was calculated from the flow meter readings according to Eq. (1):

$$V = D \times R \times A_N \times 2/3 \tag{1}$$

where D is a constant in the towing distance per revolution of the flow meter (0.107 m); R is the number of revolutions of the flow meter; A_N is the area of the net mouth (0.5625 m^2) ; and 2/3 is the proportion of the neuston net frame that was submerged during towing, where the net was equipped with floats (buoys) that were placed to collect MPs from the upper \sim 0.5 m surface layer. D represents the filtration efficiency that was calculated by towing the same distance with and without the net in a water tunnel at Tokyo University of Marine Science and Technology (Michida et al., 2019). Although areal units (e.g., items per km²) are frequently reported in similar research, when comparing wider areas such as the Mediterranean Sea and Atlantic Ocean (Collignon et al., 2012; Eriksen et al., 2013; Silvestrova and Stepanova, 2021), we adopted a volumetric approach (number of items per cubic meter of seawater) as the standardised measurement of debris for each sampling station; however we can still compare areas A (m²) by converting the recorded values according to Eq. (2):

$$A = D \times R \times A_N \times 3/4 \tag{2}$$

Furthermore, considering that the number of small plastic particles decreases exponentially with depth, it has been recommended that the concentration of MPs should be integrated vertically (Kukulka et al., 2012; Reisser et al., 2015). The theoretical concentration per square meter (items· m^{-2}) of MP particles on the surface can also be calculated by accounting for wind speed and significant wave height during the survey (Supplementary Fig. S1).

Sorting and identification of MPs were carried out at the Fish and Ships Laboratory of the Faculty of Fisheries, Nagasaki University, using the methods described in Michida et al. (2019). To reduce sample contamination before and during analysis, the laboratory was closed to minimise airflow by closing the windows and door. All equipment and working counter tops were disinfected with a 90% alcohol solution prior to analysis, and the experimental equipment was rinsed with deionised water prior to analysis. The equipment was covered with aluminium foil while not in operation. Additionally, during sorting, petri dishes with distilled water were placed within the work area and subsequently observed under a microscope to check for airborne contamination (Lam et al., 2020). Samples were visually examined by dissecting stereo microscopy (SMZ745T, Nikon Corporation, Tokyo, Japan), and debris was initially categorized as plastic and non-plastic by differentiating their colour, shape, and physical response properties (e.g., softness and texture) (Doyle et al., 2011; Hidalgo-Ruz et al., 2012). Non-plastic debris consisted of zooplankton, fish larvae and juveniles, crustaceans, wood fragments, and algae. In instances where many non-plastic particles

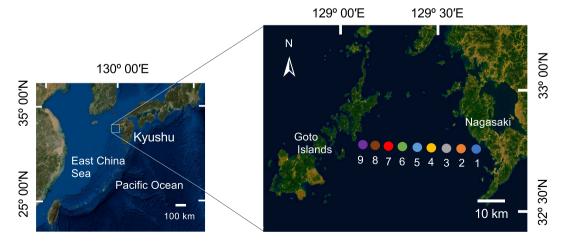


Fig. 1. Sampling sites for sea surface microplastic collection in the waters off the west coast of Kyushu, Japan. See Table S1 for details on the sampling locations, including coordinates and water depth.

were identified and difficult to sort, samples were treated with 30% $\rm H_2O_2$ at room temperature for one week to dissolve interfering organic matter (Michida et al., 2019), sorted, and individually stored in pill cases

All particles were photographed by a digital camera (NOA630, WRAYCAM Corporation, Tokyo, Japan) attached to a microscope, and the longest side of the major axis was measured using the MicroStudio software. The size range of MPs analysed was from 350 µm to 5 mm. MPs were categorized by four shape types (fragment, form, film, fibre) and eight colours (white, transparent, black, yellow, green, blue, red) based on the methods of Cheung et al. (2016) and Lam et al. (2020). Finally, all plastic polymer compositions were identified using an attenuated total reflection Fourier transform infrared spectrometer (ATR-FTIR; FT-IR-4600, JASCO Corporation, Tokyo, Japan) that exhibited a spectrum range from 4000 to 400 cm⁻¹ at a resolution of 4 cm⁻¹ and performed eight scans per sample. Background blank scans per every 50 samples were conducted for comparison, and the top-plate and diamond prism were cleaned with 75% ethanol prior to each sample analysis. The polymer type was identified by comparing a standard reference from KnowItAll Spectroscopy Library (Wiley Science Solutions, New Jersey, USA), and PEAKPICK function was adopted to confirm three or more plastic polymer characteristic peaks in the spectrum. The Hit Quality Index (HQI) was also recorded.

Concentration values were expressed in terms of the number of plastic items per cubic meter of seawater (items·m $^{-3}$), and all statistical data were expressed as mean items ± 1 standard deviation (S.D.). Since the quantitative data posed a non-nominal distribution (Shapiro-Wilk test: abundance, $p=1.1\times 10^{-11}$; size, $p=1.1\times 10^{-4}$) and the variance exhibited hetero-homogeneity (Bartlett test: abundance, $p=2.2\times 10^{-16}$, size, $p=6.8\times 10^{-6}$), a non-parametric analysis (Kruskal-Wallis test) was used. If the test indicated a significant difference, multiple comparisons were then performed using a Steel-Dwass post-hoc test. To test the differences among qualitative data (i.e., colour, shape, and polymer composition), Chi-square (χ^2) tests were applied. All statistical analyses were performed in R (v.1.2.5033).

A total of 6131 plastic items were identified in this survey. The mean and median abundance were 0.49 ± 0.92 items·m⁻³ ($n=54,\pm S.D.$), and 0.12 ± 0.56 items·m⁻³ ($n=54,\pm$ median absolute deviation—M.A.D.), respectively (Table 1). The Kruskal-Wallis test showed a significant difference in the mean abundance between months (n=6,p<0.0001; Fig. 2a), while that observed among the sampling stations was insignificant (n=9,p=0.95; Fig. 2b). The highest observed abundance between surveys was 1.97 ± 1.49 items·m⁻³ (October 2019), which was ~50-fold higher than the lowest value: 0.04 ± 0.03 items·m⁻³ (April

Table 1 Microplastic abundance: number of plastic items per cubic meter of seawater (items·m $^{-3}$), and longest size of major axis (mm) collected from the surface waters off the west coast of Kyushu, Japan. S.D.: standard deviation; M.A.D.: median absolute deviation.

Sampling date	Abundance (items·m ⁻³)			Size (mm)		
	N	Mean \pm S.D.	Median \pm M.A.D.	N	Mean \pm S.D.	$\begin{array}{l} \text{Median} \ \pm \\ \text{M.A.D.} \end{array}$
11th Apr. 2019	9	0.04 ± 0.03	0.02 ± 0.03	40	$\begin{array}{c} \textbf{2.32} \pm \\ \textbf{1.05} \end{array}$	2.00 ± 0.80
3rd Jun. 2019	9	$\begin{array}{c} 0.09 \pm \\ 0.08 \end{array}$	$\textbf{0.07} \pm \textbf{0.06}$	85	$\begin{array}{c} 1.96 \pm \\ 0.63 \end{array}$	1.96 ± 0.48
3rd Jul. 2019	9	$\begin{array}{c} \textbf{0.27} \pm \\ \textbf{0.31} \end{array}$	$\textbf{0.18} \pm \textbf{0.23}$	563	$\begin{array}{c} 2.10 \; \pm \\ 0.32 \end{array}$	$\textbf{2.22} \pm \textbf{0.23}$
4th Oct. 2019	9	$\begin{array}{c} \textbf{1.97} \pm \\ \textbf{1.49} \end{array}$	1.65 ± 1.00	4343	$\begin{array}{c} 1.61 \pm \\ 0.19 \end{array}$	$\textbf{1.66} \pm \textbf{0.17}$
12th Nov. 2019	9	$\begin{array}{c} 0.29 \pm \\ 0.24 \end{array}$	$\textbf{0.17} \pm \textbf{0.20}$	608	$\begin{array}{c} 2.13 \pm \\ 0.25 \end{array}$	2.21 ± 0.21
1st Jun. 2020	9	$\begin{array}{c} \textbf{0.25} \pm \\ \textbf{0.45} \end{array}$	$\textbf{0.06} \pm \textbf{0.28}$	492	$\begin{array}{c} 1.81\ \pm \\ 0.36\end{array}$	1.84 ± 0.21
Total	54	$\begin{array}{c} 0.49 \pm \\ 0.92 \end{array}$	$\textbf{0.12} \pm \textbf{0.56}$	6131	$\begin{array}{c} 1.99 \pm \\ 0.57 \end{array}$	1.93 ± 0.39

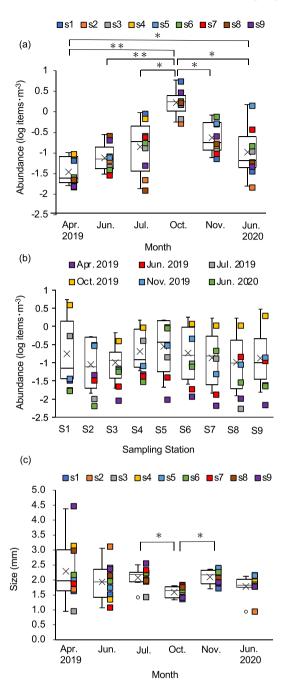


Fig. 2. Box-and-whisker plots of the spatiotemporal patterns of the (a) abundance and (b) longest size of the major axis of microplastics in the waters off the west coast of Kyushu, Japan (*p < 0.05 and **p < 0.01 were determined by the Steel-Dwass post-hoc test). The y-axes in (a) and (b) are in \log_{10} scale. Lower and upper box boundaries indicate the 25th and 75th percentiles, cross marks represent the mean, interior box lines are the median, and exterior whiskers are the 10th and 90th percentiles. Plot abundance at each sampling station was n = 6.

2019; Fig. 2). Furthermore, the highest abundance among the sampling stations was 5.50 items·m $^{-3}$ (October 2019), which was 550-fold higher than the lowest value; 0.01 items·m $^{-3}$ (April 2019).

The mean and median size of the MPs were $1.71\pm0.93~\text{mm}$ (n=6131) and $1.45\pm0.73~\text{mm}$ (n=6131; Table 1), respectively. The Kruskal-Wallis test confirmed a significant difference in the mean sizes among the months (n=6,p<0.05; Fig. 2c). Combining all months and stations, there was a trend of increasing abundance of MPs with decreasing mean size when x ranged from 1 to 5 mm or when it was <1

mm (Fig. 3a). These trends were also confirmed by the size frequency in each month (Fig. 3b–g).

Combining all months and stations, fragments were the primary shape type (76.0%), followed by foams (14.8%), fibres (5.6%), and films (3.5%); meanwhile, no notable primary MP such as microbeads and resin pellets were found (Fig. 4a). Furthermore, white was the most abundant colour (48.3%), followed by transparent (20.2%) and blue (11.4%; Fig. 4b). With regards to polymer composition, polyethylene (PE) was the predominant type (80.0%), followed by polystyrene (PS; 9.0%) and polypropylene (PP; 5.9%; Fig. 4c). Other polymer types identified included polyvinyl chloride (PVC; 0.9%) and polyethylene terephthalate (PET; 0.4%). Although overall trends of qualitative characterisation were roughly upheld in each individual month, Chi-square (χ^2) tests indicated significant differences in shape, colour, and polymer type between them (p < 0.0001 for each qualitative variable).

The work presented here represents the first report of a stationary, long-term survey of sea surface MPs from the marine waters surrounding Japan. Results from this study revealed that sea surface MPs displayed statistically significant spatiotemporal variability of quantitative (abundance and size) and qualitative (shape, colour and polymer type) characteristics in the waters off the west coast of Kyushu, Japan. These results strongly supported the hypothesis of heterogeneous MP

distribution in space and time (Doyle et al., 2011; Gajšt et al., 2016; Lam et al., 2020). Indeed, differences between the highest and lowest average monthly abundances were 50-fold and 550-fold between all net tows, respectively.

In the study area, MP items were found in each of the 54 tows used for sampling over the 14-month analysis period (six surveys x nine stations). Although spatiotemporal heterogeneity of MP abundance was observed in the study area, there was no significant difference among the sampling stations during the survey period, thereby suggesting that surface MPs do not locally aggregate at a particular station within the survey area. Sea surface MPs can be found globally from semi-closed bays (Lattin et al., 2004; Chen et al., 2018; Kashiwabara et al., 2021; Nakano et al., 2021) to the Arctic Ocean (Peeken et al., 2018; Kanhai et al., 2020). Mean abundances of MPs are observed to be ten orders of magnitude, ranging from 10^{-5} items·m⁻³ in the equatorial Pacific (Spear et al., 1995) to 10^5 items·m⁻³ off the southern coast of Korea (Song et al., 2014), depending on sea area and mesh size of the net. The mesh size of the neuston or manta nets primarily affects the absolute amount of plastic particles (Kang et al., 2015; Barrows et al., 2017; Lindeque et al., 2020; Tokai et al., 2021). Globally, the mean abundance in the surface waters collected by a net with 290-350 µm mesh size was 0.96 ± 2.05 items·m⁻³ (Shim et al., 2018). In the present study, the

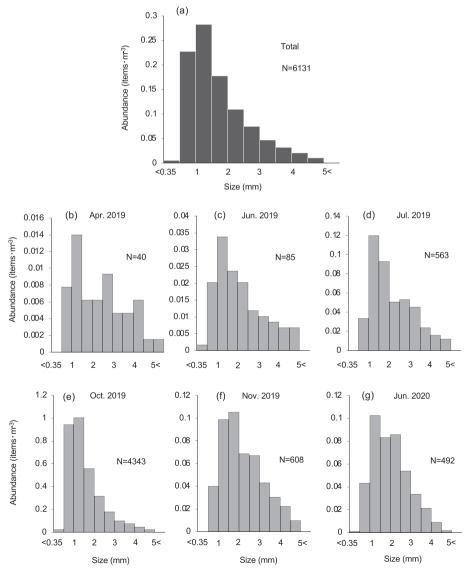


Fig. 3. Size distribution of microplastic items in (a) total and (b)-(g) of each sampling date, showing mean abundances (items·m⁻³).

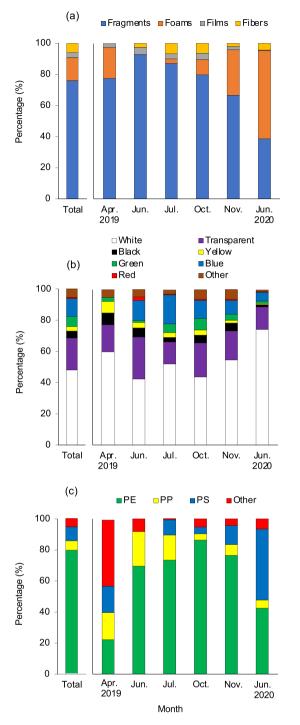


Fig. 4. Total and monthly percentage compositions for (a) shape (fibres, films, foams, and fragments), (b) colour (white, transparent, black, yellow, green, blue, red, and other), and (c) polymer type (polyethylene, PE; polypropylene, PP; polystyrene, PS) and other microplastic abundances collected from the surface waters off the west coast of Kyushu, Japan. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

average mean abundance was 0.49 ± 0.92 items·m⁻³, as evaluated by a neuston net with a 350 µm mesh size; this value is approximately half of the global average mean abundance. This value was similar to that found in the Tokyo Bay in May 2019 using the same neuston net (0.15–0.90 items·m⁻³, mean = 0.53, n=4), except for one outliner (Nakano et al., 2021). Contrarily, Isobe et al. (2015) reported an abundance in the East Asian Seas of 3.74 items·m⁻³, and deemed the area a hotspot of sea

surface MPs. The reasons for this discrepancy are unknown, but it may be related to the spatial and/or temporal variability of MP abundance seen in the present study. In fact, the mean abundance recorded in Tokyo Bay shot up to $3.98~{\rm items\cdot m^{-3}}$ when including the outliner in May 2019 (17.75 items·m $^{-3}$), leading the authors to conclude that the bay is one of the most polluted areas in the world with respect to MPs (Nakano et al., 2021). Our study also found considerable variability in abundance, ranging from 0.01 to 5.50 items·m $^{-3}$; thus, further multiple samplings with repeated surveys are needed to better assess and characterise variations in abundance and distribution patterns.

The observed variability in mean abundance may not be caused by physical disturbances such as wave height, wind velocity, or close-range transport. Buoyant MP particles are mixed vertically and distributed within the upper water column by wind and turbulent transport; thus, surface abundance decreases with increasing wind velocity (Kukulka et al., 2012; Reisser et al., 2015). Although wind-driven mixing was likely a factor throughout the present study, this physical turbulence could not explain the observed variability alone as wind velocity during sampling did not correlate with either abundance or a significant wave height (Supplementary Fig. S1a-b). Furthermore, nearby transport arising from adjacent rivers and landfills also could not explain the observed variability in abundance. The high unidirectional flow of rivers drives the movement of plastic particles into the ocean (Browne et al., 2010; Moore et al., 2002). During rainy seasons, surface runoff increases and releases MPs from inland areas to streams and rivers (Williams and Simmons, 1999; Cunningham and Wilson, 2003; Nakano et al., 2021), resulting in increased MP concentrations of the adjacent coastal sea areas (Cheung et al., 2016); however, the abundance of MP items in the survey area of the present study did not increase during the rainy season. Therefore, the observed variability in abundance here may have arisen from long-range transport, rather than short-range transport. The hydrodynamic environment of this study area is strongly influenced by the Kuroshio current (Katoh et al., 1996). Isobe et al. (2015) showed that small plastic fragments near Japan (upstream from the East and Southeast Asian countries) were responsible for discharging large amounts of plastic waste into the ocean (Jambeck et al., 2015) and occurred due to the Kuroshio and Tsushima currents. A northward branched channel from the Kuroshio currents was observed by a general ocean circulation model; however, no clear relationship was found between geostrophic mean flow (Supplementary Fig. S2) and the dramatic increase in MP abundance in October 2019 (Fig. 2a). This suggests that the strength of northward branched currents in the study area could not explain the observed increase in abundance, and perhaps this explosion originated from massive clusters generated by extreme flash flooding and/or Changjiang River discharge. In fact, lower salinity in October 2019 was observed (Supplementary Fig. S1d). One storm resulted in litter being deposited at even greater distances from the river mouth and coastal area (Lattin et al., 2004). Particle tracing model analyses are needed to pinpoint the emission sources, and fixed-point observations are required to ascertain the future ecological consequences of these anthropogenic particles.

In this study, the heterogeneity of MP characteristics was accompanied by measurements of mean size (Fig. 2b). Although many recent studies have reported MP sizes (e.g., Cózar et al., 2014; Reisser et al., 2015; Gajšt et al., 2016; Lam et al., 2020), information on the seasonal variability of sizes are still lacking. The mean sizes in October 2019 were significantly smaller than those of the preceding months (July and November 2019; Fig. 2b). Plastic debris is gradually degraded as it moves through the ocean currents and gets repeatedly washed ashore on beaches, before returning to the ocean (Andrady, 2011; Hidalgo-Ruz et al., 2012; Isobe et al., 2015). Thus, the observed difference in mean size could be due to more intense physical processes, longer drifting and/or residence times (i.e., older debris), or secondary plastics originating from larger debris. Furthermore, the size distribution of MP items should be noted. Overall, the mode of item size was ~1–1.5 mm and the abundance decreased rapidly at sizes <1 mm (Fig. 3a); these trends were

also observed in every monthly size distribution (Fig. 3b–g). Mesh selectively was a major reason for the observed skewed distribution, as most MPs < 1 mm could not be collected by the mesh size used (350 $\mu m;$ Tokai et al., 2021). Other factors, such as coastal deposition (Hinata et al., 2017), sessile organisms (Zettler et al., 2013; Long et al., 2015), and bioaccumulation by zooplankton (Cole et al., 2015; Desforges et al., 2015) also could have contributed to the patterns observed, as they play potentially significant roles in the removal of MPs from the surface waters.

Significant differences in qualitative characteristics (shape, colour, and polymer type) were also observed in the present study (Fig. 4). All MPs identified here were classified by secondary plastics. Fragments were the most frequent shape type observed (Fig. 4a); meanwhile, white and transparent colour types were predominant and later identified as PE by FT-IR analysis (Fig. 4b and c). These compositions are in agreement with previous studies carried out in the Pearl River Estuary (Lam et al., 2020), Tokyo Bay (Nakano et al., 2021), and the western North Pacific Ocean (Yamashita et al., 2007); however, the composition of all three characteristics varied significantly among the survey months. For example, foam shapes were relatively high in June 2020 and accompanied by an increase in low density PS (polystyrene). The density of plastics varies with the polymer type (Browne et al., 2010). The density, along with the MP size and geographic emission variability, seem to largely affect transportation routes and the resulting spatiotemporal heterogeneity in MPs. More importantly, the qualitative and quantitative characteristics of MPs affect ingestive behaviour by marine biota. Ory et al. (2017) reported that fish selectively ingested specific colours of MPs more closely resembling zooplankton, a significant factor in the growing global concern of bioaccumulated MP particles found in the gastrointestinal tracts of fish (e.g., Collard et al., 2017; Lusher et al., 2016; Rummel et al., 2016). Once plastic waste is released into the ocean, complete retrieval is impossible, and there is an urgent need for global cooperation before the amount of marine plastic will outweigh the fish.

CRediT authorship contribution statement

Tsunefumi Kobayashi: Formal analysis, Investigation, Writing – original draft, Visualization. Mitsuharu Yagi: Conceptualization, Methodology, Investigation, Writing – original draft, Writing – review & editing, Supervision, Funding acquisition. Toshiya Kawaguchi: Investigation, Writing – review & editing. Toshiro Hata: Investigation, Writing – review & editing. Kenichi Shimizu: Investigation, Writing – review & editing.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This work was supported by JSPS KAKENHI (Grant Number JP18K14790). The authors are grateful to the captain, officers, and crews of the training ship *Kakuyo-maru*, and also acknowledge the "Fish and Ships Laboratory" staff from the Faculty of Fisheries, Nagasaki University, who directly and indirectly helped throughout the entirety of the study. Finally, thanks to the editor and two anonymous reviewers for their valuable comments and suggestions that greatly improved the quality of this manuscript.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.marpolbul.2021.112563.

References

- Andrady, A.L., 2011. Microplastics in the marine environment. Mar. Pollut. Bull. 62, 1596–1605. https://doi.org/10.1016/j.marpolbul.2011.05.030.
- Auta, H.S., Emenike, C., Fauziah, S., 2017. Distribution and importance of microplastics in the marine environment: a review of the sources, fate, effects, and potential solutions. Environ. Int. 102, 165–176. https://doi.org/10.1016/j. envirt.2017.02.013.
- Barrows, A.P.W., Neumann, C.A., Berger, M.L., Shaw, S.D., 2017. Grab vs. neuston tow net: a microplastic sampling performance comparison and possible advances in the field. Anal. Methods 9, 1446–1453. https://doi.org/10.1039/C6AY02387H.
- Browne, M.A., Galloway, T.S., Thompson, R.C., 2010. Spatial patterns of plastic debris along estuarine shorelines. Environ. Sci. Technol. 44, 3404–3409. https://doi.org/10.1021/es903784e.
- Chen, M., Jin, M., Tao, P., Wang, Z., Xie, W., Yu, X., Wang, K., 2018. Assessment of microplastics derived from mariculture in Xiangshan Bay, China. Environ. Pollut. 242. 1146–1156. https://doi.org/10.1016/j.envpol.2018.07.133.
- Cheung, P.K., Cheung, L.T.O., Fok, L., 2016. Seasonal variation in the abundance of marine plastic debris in the estuary of a subtropical macro-scale drainage basin in South China. Sci. Total Environ. 562, 658–665. https://doi.org/10.1016/j. scitotenv.2016.04.048.
- Cole, M., Lindeque, P., Halsband, C., Galloway, T.S., 2011. Microplastics as contaminants in the marine environment: a review. Mar. Pollut. Bull. 62, 2588–2597. https://doi. org/10.1016/j.marpolbul.2011.09.025.
- Cole, M., Webb, H., Lindeque, P.K., Fileman, E.S., Halsband, C., Galloway, T.S., 2015. Isolation of microplastics in biota-rich seawater samples and marine organisms. Sci. Rep. 4, 4528. https://doi.org/10.1038/srep.04528.
- Collard, F., Gilbert, B., Compère, P., Eppe, G., Das, K., Jauniaux, T., Parmentier, E., 2017.
 Microplastics in livers of European anchovies (*Engraulis encrasicolus*, L.). Environ.
 Pollut. 229, 1000–1005. https://doi.org/10.1016/j.envpol.2017.07.089.
- Collignon, A., Hecq, J.-H., Glagani, F., Voisin, P., Collard, F., Goffart, A., 2012. Neustonic microplastic and zooplankton in the North Western Mediterranean Sea. Mar. Pollut. Bull. 64, 861–864. https://doi.org/10.1016/j.marpolbul.2012.01.011.
- Cózar, A., Echevarria, F., Gonzalez-Gordillo, J.I., Irigoien, X., Ubeda, B., Hernandez-Leon, S., Palma, A.T., Navarro, S., Garcia-de-Lomas, J., Ruiz, A., Fernandez-de-Puelles, M.L., Duarte, C.M., 2014. Plastic debris in the open ocean. Proc. Natl. Acad. Sci. 111, 10239–10244. https://doi.org/10.1073/pnas.1314705111.
- Cunningham, D.J., Wilson, S.P., 2003. Marine debris on beaches of the greater Sydney region. J. Coast. Res. 19, 421–430.
- De Witte, B., Devriese, L., Bekaert, K., Hoffman, S., Vandermeersch, G., Cooreman, K., Robbens, J., 2014. Quality assessment of the blue mussel (Mytilus edulis): comparison between commercial and wild types. Mar. Pollut. Bull. 85, 146–155. https://doi.org/10.1016/j.marpolbul.2014.06.006.
- Desforges, J.-P.W., Galbraith, M., Ross, P.S., 2015. Ingestion of microplastics by zooplankton in the Northeast Pacific Ocean. Arch. Environ. Contam. Toxicol. 69, 320–330. https://doi.org/10.1007/s00244-015-0172-5.
- Doyle, M.J., Watson, W., Bowlin, N.M., Sheavly, S.B., 2011. Plastic particles in coastal pelagic ecosystems of the Northeast Pacific ocean. Mar. Environ. Res. 71, 41–52. https://doi.org/10.1016/j.marenvres.2010.10.001.
- Eriksen, M., Mason, S., Wilson, S., Box, C., Zellers, A., Edwards, W., Farley, H., Amato, S., 2013. Microplastic pollution in the surface waters of the Laurentian Great Lakes. Mar. Pollut. Bull. 77, 177–182. https://doi.org/10.1016/j.marpolbul.2013.10.007.
- Gajšt, T., Bizjak, T., Palatinus, A., Liubartseva, S., Kržan, A., 2016. Sea surface microplastics in Slovenian part of the Northern Adriatic. Mar. Pollut. Bull. 113, 392–399. https://doi.org/10.1016/j.marpolbul.2016.10.031.
- GESAMP, 2019. Guidelines of the monitoring and assessment of plastic litter and microplastics in the ocean. In: Kershaw, P.J., Turra, A., Galgani, F. (Eds.), (IMO/FAO/UNESCO-IOC/UNIDO/WMO/IAEA/UN/UNEP/UNDP/ISA Joint Group of Experts on the Scientific Aspects of Marine Environmental Protection). Rep. Stud. GESAMP No. 99, pp. 130.
- Geyer, R., Jambeck, J.R., Law, K.L., 2017. Production, use, and fate of all plastics ever made. Sci. Adv. 3, 25–29. https://doi.org/10.1126/sciadv.1700782.
- Gregory, M.R., 2009. Environmental implications of plastic debris in marine settings—entanglement, ingestion, smothering, hangers-on, hitch-hiking and alien invasions. Philos. Trans. R. Soc. B Biol. Sci. 364, 2013–2025. https://doi.org/10.1098/rstb.2008.0265.
- Hammer, J., Kraak, M.H.S., Parsons, J.R., 2012. Plastics in the Marine Environment: The Dark Side of a Modern Gift. pp. 1–44. https://doi.org/10.1007/978-1-4614-3414
- Hidalgo-Ruz, V., Gutow, L., Thompson, R.C., Thiel, M., 2012. Microplastics in the marine environment: a review of the methods used for identification and quantification. Environ. Sci. Technol. 46, 3060–3075. https://doi.org/10.1021/es2031505.
- Hinata, H., Mori, K., Ohno, K., Miyao, Y., Kataoka, T., 2017. An estimation of the average residence times and onshore-offshore diffusivities of beached microplastics based on the population decay of tagged meso- and macrolitter. Mar. Pollut. Bull. 122, 17–26. https://doi.org/10.1016/j.marpolbul.2017.05.012.
- Isobe, A., Uchida, K., Tokai, T., Iwasaki, S., 2015. East Asian seas: a hot spot of pelagic microplastics. Mar. Pollut. Bull. 101, 618–623. https://doi.org/10.1016/j. marrolbul.2015.10.042
- Jambeck, J.R., Geyer, R., Wilcox, C., Siegler, T.R., Perryman, M., Andrady, A., Narayan, R., Law, K.L., 2015. Plastic waste inputs from land into the ocean. Science 347, 768–771. https://doi.org/10.1126/science.1260352.
- Jovanović, B., 2017. Ingestion of microplastics by fish and its potential consequences from a physical perspective. Integr. Environ. Assess. Manag. 13, 510–515. https:// doi.org/10.1002/jeam.1913.

- Kang, J.-H., Kwon, O.Y., Lee, K.-W., Song, Y.K., Shim, W.J., 2015. Marine neustonic microplastics around the southeastern coast of Korea. Mar. Pollut. Bull. 96, 304–312. https://doi.org/10.1016/j.marpolbul.2015.04.054.
- Kanhai, L.D.K., Gardfeldt, K., Krumpen, T., Thompson, R.C., O'Connor, I., 2020. Microplastics in sea ice and seawater beneath ice floes from the Arctic Ocean. Sci. Rep. 10, 5004. https://doi.org/10.1038/s41598-020-61948-6.
- Kashiwabara, L.M., Kahane-Rapport, S.R., King, C., DeVogelaere, M., Goldbogen, J.A., Savoca, M.S., 2021. Microplastics and microfibers in surface waters of Monterey Bay National Marine Sanctuary, California. Mar. Pollut. Bull. 165, 112148 https://doi. org/10.1016/j.marpolbul.2021.112148.
- Katoh, O., Teshima, K., Abe, O., Fujita, H., Miyaji, K., Morinaga, K., Nakagawa, N., 1996. Process of the Tsushima Current formation revealed by ADCP measurements in summer. J. Oceanogr. 52, 491–507. https://doi.org/10.1007/BF02239051.
- Kukulka, T., Proskurowski, G., Morét-Ferguson, S., Meyer, D.W., Law, K.L., 2012. The effect of wind mixing on the vertical distribution of buoyant plastic debris. Geophys. Res. Lett. 39, n/a-n/a. https://doi.org/10.1029/2012GL051116.
- Kuroda, M., Uchida, K., Tokai, T., Miyamoto, Y., Mukai, T., Imai, K., Shimizu, K., Yagi, M., Yamanaka, Y., Mituhashi, T., 2020. The current state of marine debris on the seafloor in offshore area around Japan. Mar. Pollut. Bull. 161, 111670. https://doi.org/10.1016/j.marpolbul.2020.111670.
- Lam, T.W.L., Fok, L., Lin, L., Xie, Q., Li, H.-X., Xu, X.-R., Yeung, L.C., 2020. Spatial variation of floatable plastic debris and microplastics in the Pearl River Estuary, South China. Mar. Pollut. Bull. 158, 111383 https://doi.org/10.1016/j.marpolbul.2020.111383.
- Lattin, G.L., Moore, C.J., Zellers, A.F., Moore, S.L., Weisberg, S.B., 2004. A comparison of neustonic plastic and zooplankton at different depths near the southern California shore. Mar. Pollut. Bull. 49, 291–294. https://doi.org/10.1016/j. marpolbul.2004.01.020.
- Lebreton, L.C.M., van der Zwet, J., Damsteeg, J.-W., Slat, B., Andrady, A., Reisser, J., 2017. River plastic emissions to the world's oceans. Nat. Commun. 8, 15611. https://doi.org/10.1038/ncomms15611.
- Lindeque, P.K., Cole, M., Coppock, R.L., Lewis, C.N., Miller, R.Z., Watts, A.J.R., Wilson-McNeal, A., Wright, S.L., Galloway, T.S., 2020. Are we underestimating microplastic abundance in the marine environment? A comparison of microplastic capture with nets of different mesh-size. Environ. Pollut. 265, 114721. https://doi.org/10.1016/j.envpol.2020.114721.
- Long, M., Moriceau, B., Gallinari, M., Lambert, C., Huvet, A., Raffray, J., Soudant, P., 2015. Interactions between microplastics and phytoplankton aggregates: impact on their respective fates. Mar. Chem. 175, 39–46. https://doi.org/10.1016/j. marchem.2015.04.003.
- Lusher, A.L., O'Donnell, C., Officer, R., O'Connor, I., 2016. Microplastic interactions with North Atlantic mesopelagic fish. ICES J. Mar. Sci. 73, 1214–1225. https://doi.org/ 10.1093/icesjms/fsv241.
- Martinez, E., Maamaatuaiahutapu, K., Taillandier, V., 2009. Floating marine debris surface drift: convergence and accumulation toward the South Pacific subtropical gyre. Mar. Pollut. Bull. 58, 1347–1355. https://doi.org/10.1016/j. marpolbul.2009.04.022.
- Michida, Y. et al., 2019. Guidelines for Harmonizing Ocean Surface Microplastic Monitoring Mesthods. 71 pp. Ministry of the Environment, Japan. 10.25607/OBP-51 3.
- Moore, C.J., 2008. Synthetic polymers in the marine environment: a rapidly increasing, long-term threat. Environ. Res. 108, 131–139. https://doi.org/10.1016/j.envres 2008 07 025
- Moore, C.J., Moore, S.L., Weisberg, S.B., Lattin, G.L., Zellers, A.F., 2002. A comparison of neustonic plastic and zooplankton abundance in southern California's coastal waters. Mar. Pollut. Bull. 44, 1035–1038. https://doi.org/10.1016/S0025-326X(02) 00150-9.

- Nakano, H., Arakawa, H., Tokai, T., 2021. Microplastics on the sea surface of the semiclosed Tokyo Bay. Mar. Pollut. Bull. 162, 111887. https://doi.org/10.1016/j. marpolbul.2020.111887.
- Ory, N.C., Sobral, P., Ferreira, J.L., Thiel, M., 2017. Amberstripe scad Decapterus muroadsi (Carangidae) fish ingest blue microplastics resembling their copepod prey along the coast of Rapa Nui (Easter Island) in the South Pacific subtropical gyre. Sci. Total Environ. 586, 430–437. https://doi.org/10.1016/j.scitotenv.2017.01.175.
- Peeken, I., Primpke, S., Beyer, B., Gütermann, J., Katlein, C., Krumpen, T., Bergmann, M., Hehemann, L., Gerdts, G., 2018. Arctic sea ice is an important temporal sink and means of transport for microplastic. Nat. Commun. 9, 1505. https://doi.org/ 10.1038/s41467-018-03825-5.
- Reisser, J., Slat, B., Noble, K., du Plessis, K., Epp, M., Proietti, M., de Sonneville, J., Becker, T., Pattiaratchi, C., 2015. The vertical distribution of buoyant plastics at sea: an observational study in the North Atlantic Gyre. Biogeosciences 12, 1249–1256. https://doi.org/10.5194/bg-12-1249-2015.
- Rummel, C.D., Löder, M.G.J., Fricke, N.F., Lang, T., Griebeler, E.-M., Janke, M., Gerdts, G., 2016. Plastic ingestion by pelagic and demersal fish from the North Sea and Baltic Sea. Mar. Pollut. Bull. 102, 134–141. https://doi.org/10.1016/j. marrollul.2015.11.043
- Ryan, P.G., Moore, C.J., van Franeker, J.A., Moloney, C.L., 2009. Monitoring the abundance of plastic debris in the marine environment. Philos. Trans. R. Soc. B Biol. Sci. 364, 1999–2012. https://doi.org/10.1098/rstb.2008.0207.
- Shim, W.J., Hong, S.H., Eo, S., 2018. Marine microplastics: abundance, distribution, and composition, in: Microplastic Contamination in Aquatic Environments. Elsevier, pp. 1–26. https://doi.org/10.1016/B978-0-12-813747-5.00001-1.
- Silvestrova, K., Stepanova, N., 2021. The distribution of microplastics in the surface layer of the Atlantic Ocean from the subtropics to the equator according to visual analysis. Mar. Pollut. Bull. 162, 111836. https://doi.org/10.1016/j.marpolbul.2020.111836.
- Song, Y.K., Hong, S.H., Jang, M., Kang, J.-H., Kwon, O.Y., Han, G.M., Shim, W.J., 2014. Large accumulation of micro-sized synthetic polymer particles in the sea surface microlayer. Environ. Sci. Technol. 48, 9014–9021. https://doi.org/10.1021/ es501757s.
- Spear, L.B., Ainley, D.G., Ribic, C.A., 1995. Incidence of plastic in seabirds from the tropical pacific, 1984–1991: relation with distribution of species, sex, age, season, year and body weight. Mar. Environ. Res. 40, 123–146. https://doi.org/10.1016/0141-1136(94)00140-K.
- Thompson, R.C., 2006. Plastic debris in the marine environment: consequences and solutions. In: Krause, J.C., Nordheim, H., Bräger, S. (Comp.), Marine Nature Conservation in Europe 2006. Federal Agency for Nature Conservation, Bonn, pp. 107–115. http://www.habitatmarenatura2000.de.
- Tokai, T., Uchida, K., Kuroda, M., Isobe, A., 2021. Mesh selectivity of neuston nets for microplastics. Mar. Pollut. Bull. 165, 112111. https://doi.org/10.1016/j. marnolbul.2021.112111.
- Williams, A.T., Simmons, S.L., 1999. Sources of riverine litter: the River Taff, South Wales, UK. Water Air Soil Pollut. 112, 197–216. https://doi.org/10.1023/A: 1005000724803.
- World Economic Forum, Ellen MacArthur Foundation and McKinsey & Company, 2016.

 The New Plastics Economy Rethinking the Future of Plastics. http://www.ellenmacarthurfoundation.org/publications.
- Yamashita, R., Takada, H., Murakami, M., Fukuwaka, M., Watanuki, Y., 2007. Evaluation of noninvasive approach for monitoring PCB pollution of seabirds using preen gland oil. Environ. Sci. Technol. 41, 4901–4906. https://doi.org/10.1021/es0701863.
- oil. Environ. Sci. Technol. 41, 4901–4906. https://doi.org/10.1021/es0701863
 Zettler, E.R., Mincer, T.J., Amaral-Zettler, L.A., 2013. Life in the "plastisphere": microbial communities on plastic marine debris. Environ. Sci. Technol. 47, 7137–7146. https://doi.org/10.1021/es401288x.