



# Assessing the relationship between the abundance of microplastics in sediments, surface waters, and fish in the Iran southern shores

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Received: 11 May 2021 / Accepted: 16 October 2021 / Published online: 24 October 2021  
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## Abstract

The purpose of this study was to investigate the microplastic (MP) pollution in sediments, surface waters, and four fish species in the northern coast of the Persian Gulf. Sampling was conducted in seven important regions during December 2019. The abundance of MPs was respectively  $190 \pm 35.5$  items/kg dry weight for sediments,  $9.28 \pm 2.1$  items/km<sup>2</sup> for surface waters, and  $0.33 \pm 0.05$  items/individual for fish. There was no correlation between MP abundance in surface water, sediment, and fish samples. Except for *Cynoglossus arel*, abundance of isolated MPs did not show significant relationships with body weight, body length, and gastrointestinal tract weight ( $P > 0.05$ ). MPs were ranged from 0.3 to 5 mm in size and were prevailed by fiber in shape; black, red, and blue in color; and polypropylene and polyethylene in polymer. This study may help in increasing our knowledge regarding MP pollution in marine water systems and biota.

**Keywords** Microplastics · Sediments · Surface waters · Fish · Polymer · Persian Gulf

## Introduction

Microplastic (MP) pollution exists in all aquatic ecosystems of the world. MPs are considered a permanent threat since they are found in different marine animal species including zooplankton (Cole et al. 2011), birds (Van Franeker et al. 2011), sea turtles (Tourinho et al. 2010), marine mammals (Besseling et al. 2015), and fish (e.g., Romeo et al. 2015; Güven et al. 2017; Arias et al. 2019; Rasta et al. 2021). These small particles can enter the environment directly

(primary MPs) or following the decomposition of larger plastics (secondary MPs) (Carr et al. 2016; Auta et al. 2017). Due to the high stability, it is difficult to remove MPs from the environment, especially in marine ecosystems, and it increases the worldwide concerns about these particles.

In the last few decades, a large number of MPs have been detected in the sediments, water columns, and water surface of marine environments (Cole et al. 2011; Eriksen et al. 2013; do Sul and Costa 2014; Auta et al. 2017; Rasta et al. 2020). Moreover, MP particles were reported in the gastrointestinal (GI) tract of many species of marine fish (e.g., Rochman et al. 2015; Neves et al. 2015; Ferreira et al. 2016), which are among the largest and most diverse groups of animals of high biological and economic importance. In fact, particles can be taken up from the sediments and water column by marine organisms. Besides, when the particles are consumed by various organisms, they can penetrate the food web and may result in bioaccumulation followed by possible negative effects on marine creatures (Kashiwada 2006; Akdogan and Guven 2019).

The risks associated with plastic particles are included: physical effects of materials (von Moos et al. 2012; Rochman et al. 2013), chemical compounds of plastics and chemicals adsorbed from the environment (Hirai et al. 2011) (e.g., stable bioaccumulation and toxic substances PBTs) (Holmes et al. 2012), and metals (Hosseini et al. 2020). Intestinal

Responsible Editor: V.V.S.S. Sarma

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obstruction, inhibition of gastric secretion, reduction of nutritional stimuli, reduction of steroid hormone levels, delayed ovulation, and lack of successful reproduction are the physiological effects of MPs on marine creatures (Auta et al. 2017; Hermabessiere et al. 2017). On the other hand, Wang et al. (2019) examined the effects of MP exposure on parental fish of marine medaka and reported MP accumulation in their liver, intestine, and gill, along with causing oxidative stress and histological changes. They also reported parental exposure to MP postponed incubation time and decreased the hatching rate. Moreover, consumption of polystyrene also changes the behavior of fish and disrupts the fat metabolism of freshwater fish (Pedà et al. 2016). Other studies also reported that ingestion of MP particles such as PE, PP, and PS by fish larva leads to death, decreasing head/body ratios, DNA breaking, and changing to swimming behavior (Pannetier et al. 2020), and exposure to PVC MPs caused inhibition of weight increase and growth, changing antioxidant-related gene expression in the livers (Xia et al. 2020).

The Persian Gulf is a waterway along the Oman Sea and the third largest gulf in the world. It has access to open sea from the east through the Strait of Hormuz and the Oman Sea and ends in the Arvand Rud delta (Arvand River) from the west (Taghizade et al. 2012; Bibak et al. 2021). This waterway is considered as an important and strategic region at the international level because of its abundant oil and

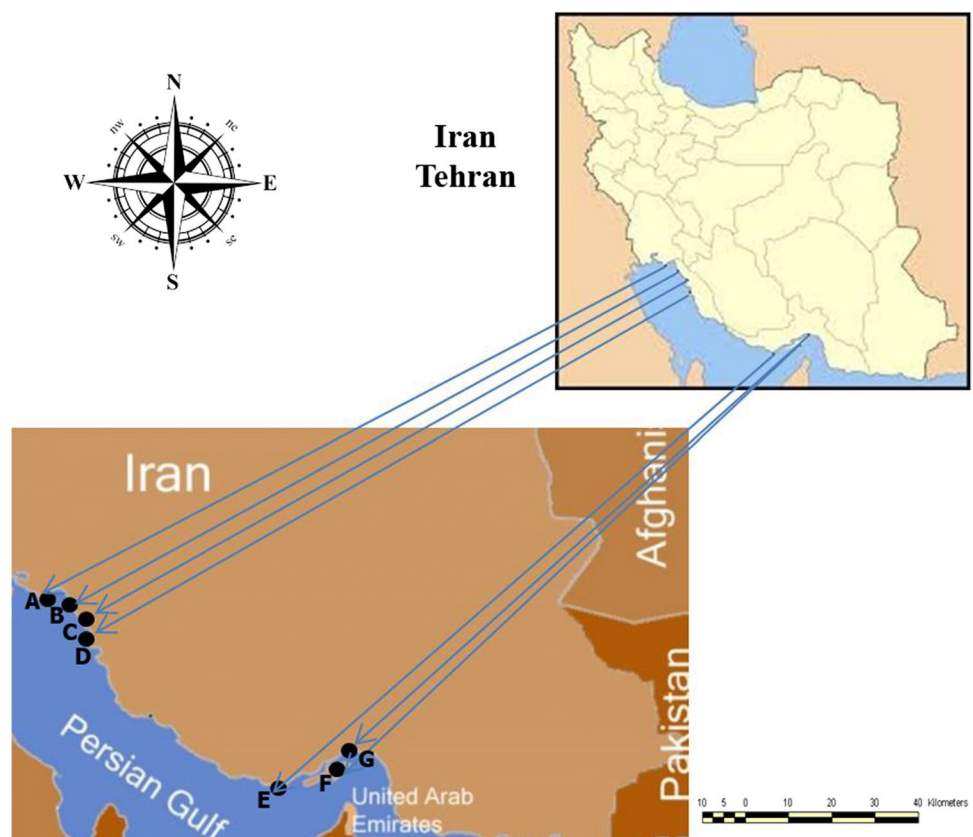
gas resources. whereas oil-related activities, untreated urban discharges, and industrial sewages impose high pollution pressure on the fauna and flora of the Gulf (Hosseini et al. 2015). Recently, different studies have been reported the MP pollution in the sediments, surface waters, and biota of the Persian Gulf (Naji et al. 2017; Akhbarizadeh et al. 2018; Naji et al. 2019; Nabizadeh et al. 2019), although there is no study on the relationship between MP pollution in the Gulf aquatic environments and fauna in a single study to date, reflecting the importance of a comprehensive study on MP pollution in the Persian Gulf. Consequently, the aim of this comprehensive study was to investigate the MP pollution in surface waters, sediments, and fish in different areas of the northern coast of the Persian Gulf (coast of Iran) as well their relationship with each other in the sampling areas for the first time.

## Materials and methods

### Sampling region

Sediments, surface waters, and fish species were sampled in seven sampling stations in December 2019 from the northern shores of the Persian Gulf. The stations are located in three provinces of Khuzestan, Bushehr, and Hormozgan (Fig. 1)

**Fig. 1** Sampling region. (A) Hendijan-Khozestan province, (B) Emam Hassan-Bushehr province, (C) Genaveh-Bushehr province, (D) Bushehr-Bushehr province, (E) Lengeh-Hormozgan province, (F) Qeshm-Hormozgan province, and (G) Bandar Abbas-Hormozgan province



and are selected on the basis of environmental characteristics and sources of pollutants. For instance, Hendijan (in Khuzestan province) is affected by fishing ports and residential houses. In Bushehr province, Emam Hassan, Genaveh and Bushehr stations are important commercial areas which are located near fishing and tourist areas and residential buildings. In Hormozgan province, Lengeh port is one of the important fishing and commercial area. Bandar Abbas and Qeshm are not only considered as important fishing and commercial ports, but also essential tourist areas in the southern region of Iran.

### Sample collection

Triplicate surface water samples were collected from each site (A to G) at the depth of 3–4 m and about 150–200 m distance of shoreline by a neuston net with an opening diameter of 50 cm, a length of 180 cm, and a mesh size of 50  $\mu\text{m}$  (Thompson et al. 2004). The net was towed for 10 min at a speed of 3 knots on the water surface (depth of  $\approx 30$  cm) and in the direction of the wind (Free et al. 2014). A flow meter was used to obtain the volume of filtered water through the net. The collected samples in the cod end were washed in glass containers. Next, they were stabilized with 70% isopropyl alcohol to prevent water from putrefaction (Martins and Sobral 2011; Toumi et al. 2019) and transferred to the laboratory at 4 °C.

Sediment samples were collected from a depth of 5 cm at the intertidal zone and also at the same time as water sampling. This sampling was performed using a stainless-steel core with a diameter of 15 cm (Claessens et al. 2011; Löder and Gerdtz 2015). Approximately 1-kg sediment of each site was collected and placed in glass containers and packaged with aluminum foil to avoid air contamination. Following the transfer of sediments to the lab, the sediment was kept at 60 °C to dry completely (Nor and Obbard 2014). The temperature is less than the melting point and deformation of polymers, so it is not expected that the shape of the polymers will change.

A total of 280 fish specimens were bought from local fishermen in the Persian Gulf from 4 commercial species (*Otolithes ruber*, *Liza abu*, *Sphyræna forsteri*, and *Cynoglossus arel*) (40 fish per region or in other words, 10 individuals for each species at each sampling station). The target

fish are commonly caught by cast net (for *Liza abu*), gill net (for *Otolithes ruber* and *Sphyræna forsteri*), and fish pot (for *Cynoglossus arel*) by local fishermen in the limited location of each sampling station. The fishes were wrapped in aluminum foil and transferred to the laboratory via a cool box (Lusher et al. 2013; Su et al. 2019). In the laboratory, the total length (TL) (cm), total weight (TW) (g), and weight of gastrointestinal tract (W of GI tract) were measured (Table 1).

### Microplastic extraction

The surface waters and sediment from each station were initially filtered through a metal sieve with a mesh size of 5 mm in order to separate large plastic pieces. Afterward, the filtrates were subjected to 30% hydrogen peroxide oxidation (20 mL) in the presence of aqueous 0.05 M Fe (II) solution (20 mL) as a catalyst to digest unstable organic matter at 60 °C (Free et al. 2014). Hydrogen peroxide was added until no more natural organic material can be seen. For extraction of sediments, 100 g of dry sediment was added to a glass flask. Then, 800 mL of saline solution (NaCl; 1.2 g cm<sup>-3</sup>) was added to a glass flask and shaken for 5 min by hand (Graca et al. 2017; Toumi et al. 2019). After about 1-h settlement, the supernatant was filtered on a 5- $\mu\text{m}$  Whatman cellulose nitrate filter to preserve MPs. The filter papers were washed with ionized water for removing the salt and finally were dried at 60 °C. The procedure was repeated thrice for every sediment sample. For surface water MPs were also separated by NaCl 1.2 g cm<sup>-3</sup> and analyzed. It is considered that NaCl solution cannot separate MPs with high density (Van Cauwenberghe et al. 2015; Quinn et al. 2017), although recent studies have been indicated that density separation with NaCl solution probably allows separation of fibers regardless of their density (Graca et al. 2017; Rasta et al. 2020).

For fish species, the GI tract of each individual was transferred to a 250-mL glass jar and then 10% potassium hydroxide solution (KOH) was added into the glass jar (Rochman et al. 2015). Recent studies have recognized 10% KOH as the most suitable solution for the digestion of intestinal contents of fish (Karami et al. 2017). The added solution was at least three times the volume of biological material. The samples were incubated within 5 days to ensure the entire

**Table 1** The biometric of all of the species fish (mean  $\pm$  SD), total length (TL), total weight (TW), and W of GI tract (the weight of gastrointestinal tract)

Species	Number	TL (cm), mean $\pm$ SD	TW (g), mean $\pm$ SD	W of GI tract (g), mean $\pm$ SD
<i>O. ruber</i>	70	29.5 $\pm$ 1.3	255.8 $\pm$ 24.5	24.7 $\pm$ 0.7
<i>L. abu</i>	70	21.6 $\pm$ 1.3	123.6 $\pm$ 19.5	14.5 $\pm$ 4.1
<i>S. forsteri</i>	70	52.8 $\pm$ 5.8	589.5 $\pm$ 151.7	38.7 $\pm$ 13.9
<i>C. arel</i>	70	29.4 $\pm$ 2.8	139.2 $\pm$ 64.5	3.2 $\pm$ 2.0

digestion via the digestive tract at 60 °C (Savoca et al. 2019). Afterward, the residual substance was filtered through 5- $\mu$ m Whatman paper. The filters were dried in a Petri dish at 60 °C for 24 h (Li et al. 2020).

### Identification of microplastics

At the end, the filters were observed under a stereomicroscope ZTX-E with  $\times 40$  magnification, and MPs were removed from other components. Several criteria including similar thickness in the length of fibers, identical of colors all over the MPs, and lack of cellular structures on particles were applied for visual observation of MP particles (Hidalgo-Ruz et al. 2012). Moreover, we discarded organic matter like wood through testing for hardness via pressure applied by forceps. Particles were counted and classified in different shapes (film, fiber, and fragment) and colors (red, blue, black, yellow, white, green, and purple). In terms of size, MP particles were measured by a calibrated scaled lens (Abidli et al. 2017) and categorized into six classes: 300–500, 500–1000, 1000–2000, 2000–3000, 3000–4000, and 4000–5000  $\mu$ m. The maximum length of particles was considered for measuring the MPs.

The chemical composition and morphological properties of each MPs were analyzed by scanning electron microscope (SEM; Hitachi SU 3500) and an energy-dispersive X-ray microanalyzer (EDS; Amptek, USA). Besides, in terms of polymer identification, 120 particles (33.61% of all isolated particles) were selected in different shapes, colors, and sizes and validated using ATR-FTIR by a Nicolette Nexus 470 (Thermo Nicolet, USA) connected to the OMNIC Software. The spectral range of each spectrum was in 650–4000  $\text{cm}^{-1}$  region, 4  $\text{cm}^{-1}$  resolution, and average over 50 scans. Before each test, a background air spectrum was recorded (Qiu et al. 2016). Hummel polymer sample library was used to identify MP polymer in the sample. Spectra matching with  $\geq 70\%$  were accepted among the reference substance and sample.

### Quality control and quality assurance

To prevent air-borne contamination, nitrile gloves and cotton lab coats were worn in the whole steps of experiments. All apparatuses and work surfaces were rinsed with alcohol 70% before and after using. All solutions were filtered before using to decrease the possibility of contamination. Aluminum foils were used to cover the samples during the experiments. Nine control blanks (three for surface waters, three for sediments, and three for fish) containing pure water were run during the analysis. A total of 8 fibers were detected in the control blanks which final data were corrected by subtracting the blank contamination from particles counted in samples.

### Data analysis

One-way ANOVA was used to compare the mean abundance of micro plastics (total of films, fragments, and fibers) of water, sediment, and fish between the stations. The same test was also applied for each fish species. Before ANOVA, normality of the data was examined with the Kolmogorov–Smirnov test. The significance of Pearson's correlation coefficient was tested for the relationships between microplastic abundance for water–sediment, water–fish, and fish–sediment combinations and between microplastic abundance in GI tract and fish size (body length, body weight, and GI tract weight). Excel 2019 and MiniTab 17 were performed to draw graphs, and SPSS 22 was used for the statistical analysis of the data.

## Results

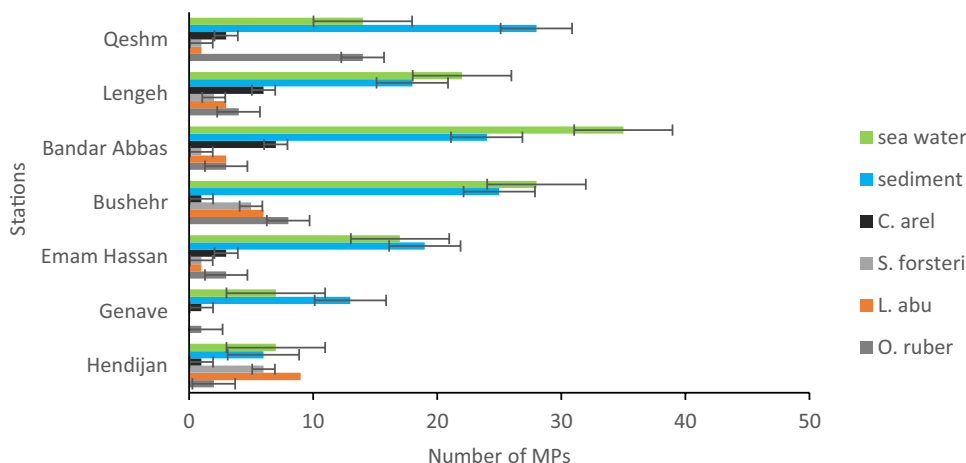
### Microplastic abundance in surface waters, sediments, and fish

Generally, MPs were found in all surface water and sediment stations as well as fish species. A total number of 133, 130, and 94 particles were detected in the sediments (mean  $\pm$  SD =  $190 \pm 35.5$  items/kg dry weight), surface waters ( $9.28 \pm 2.1$  items/ $\text{km}^2$ ), and fish ( $0.33 \pm 0.05$  items/individual), respectively. The highest abundance of MPs in the surface waters were detected in Bandar Abbas, Bushehr, and Lengeh stations, respectively. There were no significant differences between the different sampling stations, while they were highest in Qeshm, Bushehr, and Bandar Abbas stations in sediment samples. The results of the present study illustrated that 28.5% of fish had MPs in their GI tract, and *Otolithes ruber* (mean  $\pm$  SD =  $0.43 \pm 0.12$  items/individual), *Liza abu* ( $0.34 \pm 0.09$  items/individual), *Cynoglossus arel* ( $0.33 \pm 0.14$  items/individual), and *Sphyræna forsteri* ( $0.24 \pm 0.08$  items/individual) were the most polluted fish species, respectively. Bushehr, Qeshm, and Hendijan were the most polluted region based on the numbers of MPs in investigated species, respectively (Fig. 2).

Based on Pearson correlation, there was not any correlation between shape, size, color, and number of MPs in surface water–fish, fish–sediment, and sediment–surface water. According to one-way ANOVA analysis, there was no significant relationship between the number of extracted MPs from fish GI tract with MP particles taken from surface water and sediment ( $P > 0.05$ ). The results of the correlation analysis did not show significant relationships in the abundance of MPs isolated from fish GI tract with body length, body weight, and GI tract weight in *S. forsteri*, *L. abu*, and *O. ruber*, while their abundance in *C. arel* displayed



**Fig. 2** The number of MPs per sampling items in different regions (items/kg for sediments, items/km<sup>2</sup> for surface waters, and items/individual for fish GI tract)



significant relationships with body weight, body length, and GI tract weight ( $P < 0.05$ ).

### Characterization of microplastics

Fiber was the most prevalent shape in all sampling stations, and those samples were taken from the surface waters (84.71% frequency), sediments (88%), and fish (86.67%). Fragments and films were the next dominant shape in the samples, respectively (Fig. 3A). In terms of color, black, red, and blue were the most common particles in both sediments and surface waters respectively, while blue, black, and red were the most frequent MPs in the fish GI tract, respectively. Other colors such as white, purple, yellow, and green were detected in extracted particles as well (Fig. 3B). MP particles were ranged from 300 to 5000  $\mu\text{m}$  in size. The category 1000–2000  $\mu\text{m}$  was the most common size in the fish (30.85% frequently) and sediment (44.36%) samples, while the class 500–1000  $\mu\text{m}$  with 27.69% was the predominant size in the surface waters.

### Surficial textural characteristics and chemical composition of the MPs

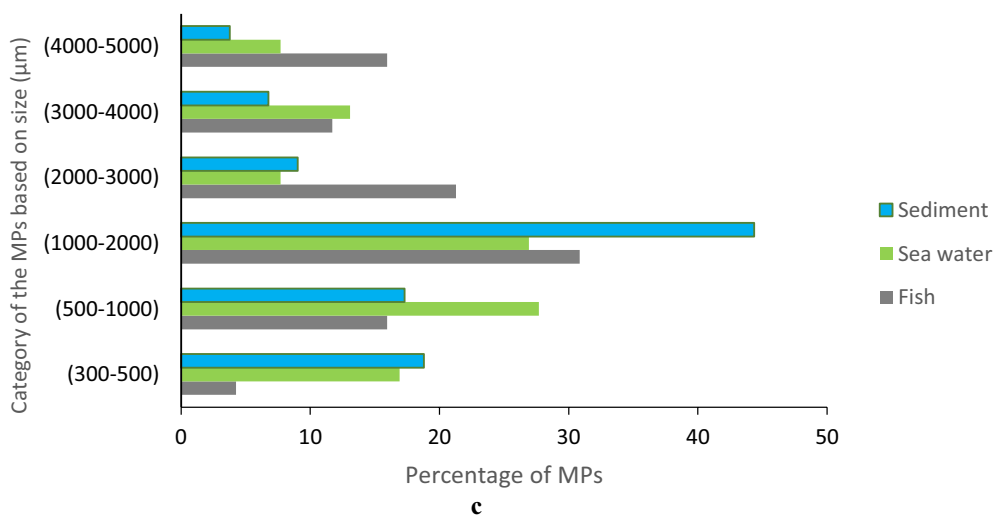
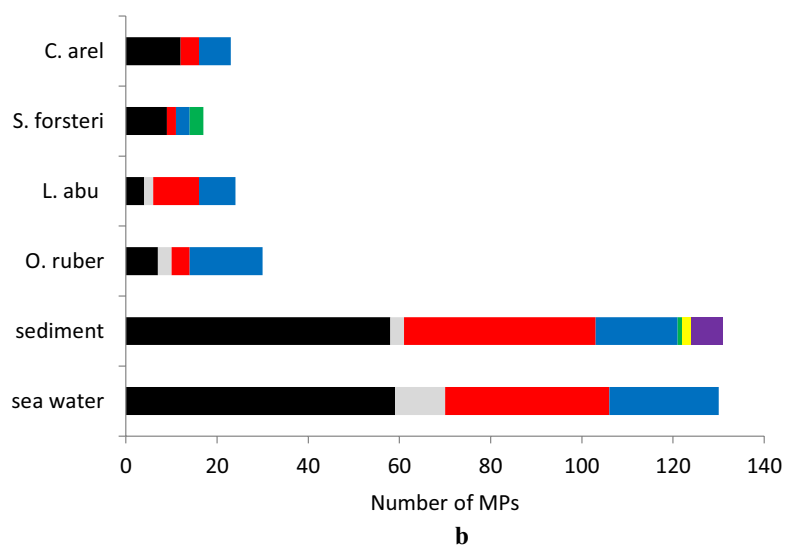
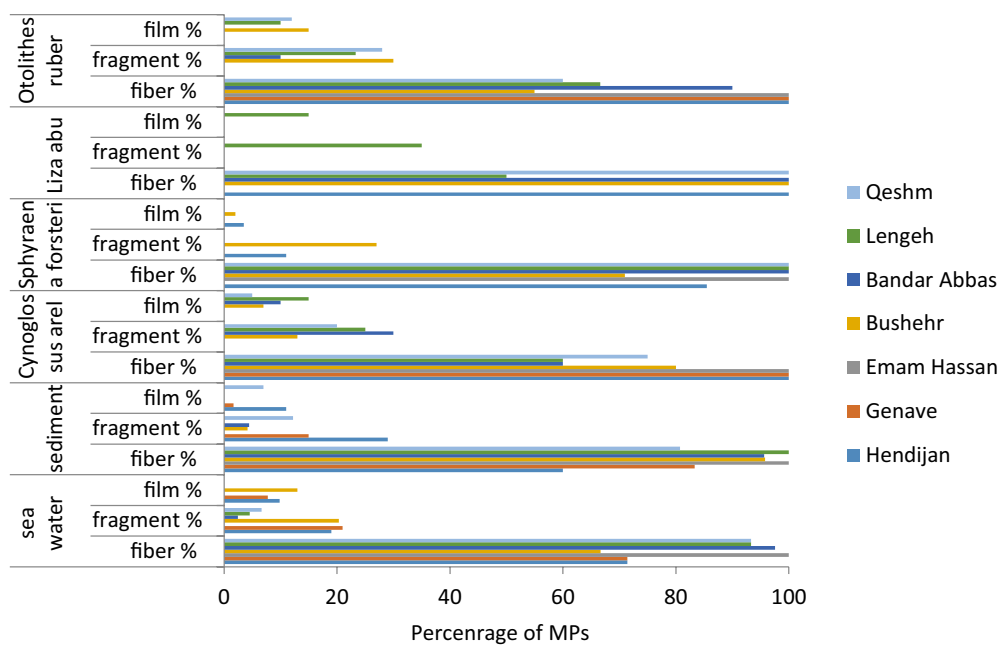
Figure 4 shows the results obtained from the analysis of the SEM images. Generally, MP particles appeared with different surfaces of non-smooth, smooth, and edged shapes. The fiber MPs were observed as narrow and smooth shapes (Fig. 4B, H). Fragment MPs appeared with non-smooth structures (Fig. 4A, E, F) and irregularly edged shapes (Fig. 4D, G). Film MPs showed an irregularly edged shape with a soft texture (Fig. 4C). The analysis results of X-ray diffraction (EDS) spectroscopy revealed some elements such as carbon (C), oxygen (O), iron (Fe), aluminum (Al), silicon (Si), calcium (Ca), magnesium (Mg), and potassium (K) in the structures of MPs. Carbon and oxygen were the main elements of MPs (Fig. 4). Of the 120 selected particles for

validation, 76 particles (63.3%) were confirmed as MPs. As a whole, five different polymers including polypropylene (PP), polystyrene (PS), polyethylene (PE), polyvinyl acetate (PVAC), and polyvinyl propionate (PVPR) were identified using ATR-FTIR spectrometry (Fig. 5). PE was the dominant polymer (43.42%) in the collected MPs. Other identified polymers were included: PP (25%), PS (18.42%), PVAC (6.58%), and PVPR (6.58%) in the identified MPs.

### Discussion

This study investigated the occurrence and distribution of microplastics in the sediments, surface waters, and fish of the Persian Gulf, coastal environment of Iran. MPs were detected in all sample stations and fish species. The average number of MPs in the present study was  $190 \pm 35.5$  items/kg dry weight for sediments and  $9.28 \pm 2.1$  items/km<sup>2</sup> for surface waters. There are several studies addressing MP pollution in the sediments and surface waters of the Persian Gulf. Naji et al. (2017) found 307 MP particles with a range of 0 to 125 and mean  $\pm SD$   $61 \pm 49$  items/kg dry weight of littoral sediments of the Persian Gulf which was lower than the present study, and they considered that fishing activity is a possible source of pollutant. Kor and Mehdiinia (2020) reported that all 15 examined stations in the surface waters of the Persian Gulf were contaminated by MPs with a range of  $1.5 \times 10^3$ – $4.6 \times 10^4$  and a mean of  $1.8 \times 10^4$  items/km<sup>2</sup> which was much higher than the present study. Merchant ships, fishery, and tourist activities likely are the different sources of MP pollution in the examined stations.

In this study, MPs were detected in all four fish species with a total number of 94 particles. Other studies in the Persian Gulf also reported the MP pollution in the fish tissues and other biotas. For instance, Akhbarizadeh et al. (2018) examined the MPs and toxic element pollution in fish muscles of the Persian Gulf and reported that the



**Fig. 3** Comparison of microplastics in terms of shape (A), color (B), and size (C) in surface water, sediment, and fish species from Persian Gulf

average abundance of MPs in the *Platycephalus indicus*, *Alepes djedaba*, *Epinephelus coioides*, and *Sphyrna jello* was  $18.50 \pm 4.55$ ,  $8.00 \pm 1.22$ ,  $7.75 \pm 2.16$ , and  $5.66 \pm 1.69$  items/10 g fish muscle, respectively. Abbasi et al. (2018) investigated the GI tract, gills, skin, and muscle of four fish species and also the muscle and exoskeleton of a tiger prawn of the Musa Estuary in the Persian Gulf and extracted 828 MP particles from all caught species. The mean abundance of MPs ranged from 7.8 items/individual for tiger prawn (*Penaeus semisulcatus*) to 21.8 items/individual for a fish species (*Platycephalus indicus*). The results of the present study showed 28.5% of fish with an average of  $0.33 \pm 0.05$  items/individual contaminated with MPs. These values are lower than those previously reported for fish in the northern Persian Gulf. Generally, the abovementioned studies reported MP pollution in different internal organs of fish, and it can be the reason for the low abundance of MPs in the GI tract of fish in this study. Moreover, recently, some studies have demonstrated that MPs found in some fish tissues such as the muscle and liver may be affected by background or external contamination during experimental analysis (Su et al. 2019; Akoueson et al. 2020). In fact, Akoueson et al. (2020) suggested that it is not feasible to decrease lab environment contamination to zero, even with high-quality assurance approaches. Consequently, it can be likely that MP content in the fish muscle may be due to laboratory environment contamination in the previous studies in the Persian Gulf which led to reporting a higher value of MPs than the present study. Moreover, differences in the protocol such as the kind of chemicals used to digest tissues, MP size classes, and identification methods have a great influence on resulting MP abundances. On the other hand, the intake of MPs by fish can be various depending on species, location, sampling time, habitat, and feeding habit, although Baalkhuyur et al. (2020) reported a low level of MPs in the commercial fish from the western part of the Persian Gulf, with an average of  $0.057 \pm 0.019$  items/individual. There are numerous studies in other parts of the world which reported low levels of MP ingestion in the fish GI tract similar to the present study as well. For example, Neves et al. (2015) investigated 26 species of commercial fish from the Portuguese coast in different habitats and detected a mean of  $0.27 \pm 0.63$  items/individual of MPs in their GI tract. The MP abundance range in the wild fish of Pearl River Estuary from South China was 0.17 to 1.33 items/individual (Lin et al. 2020). Consequently, the low prevalence of MPs in the studied fish suggests that these species is not as extremely affected by MPs as other fish in other regions of the world (e.g., Vendel et al. 2017; Ory et al. 2018). Another aspect

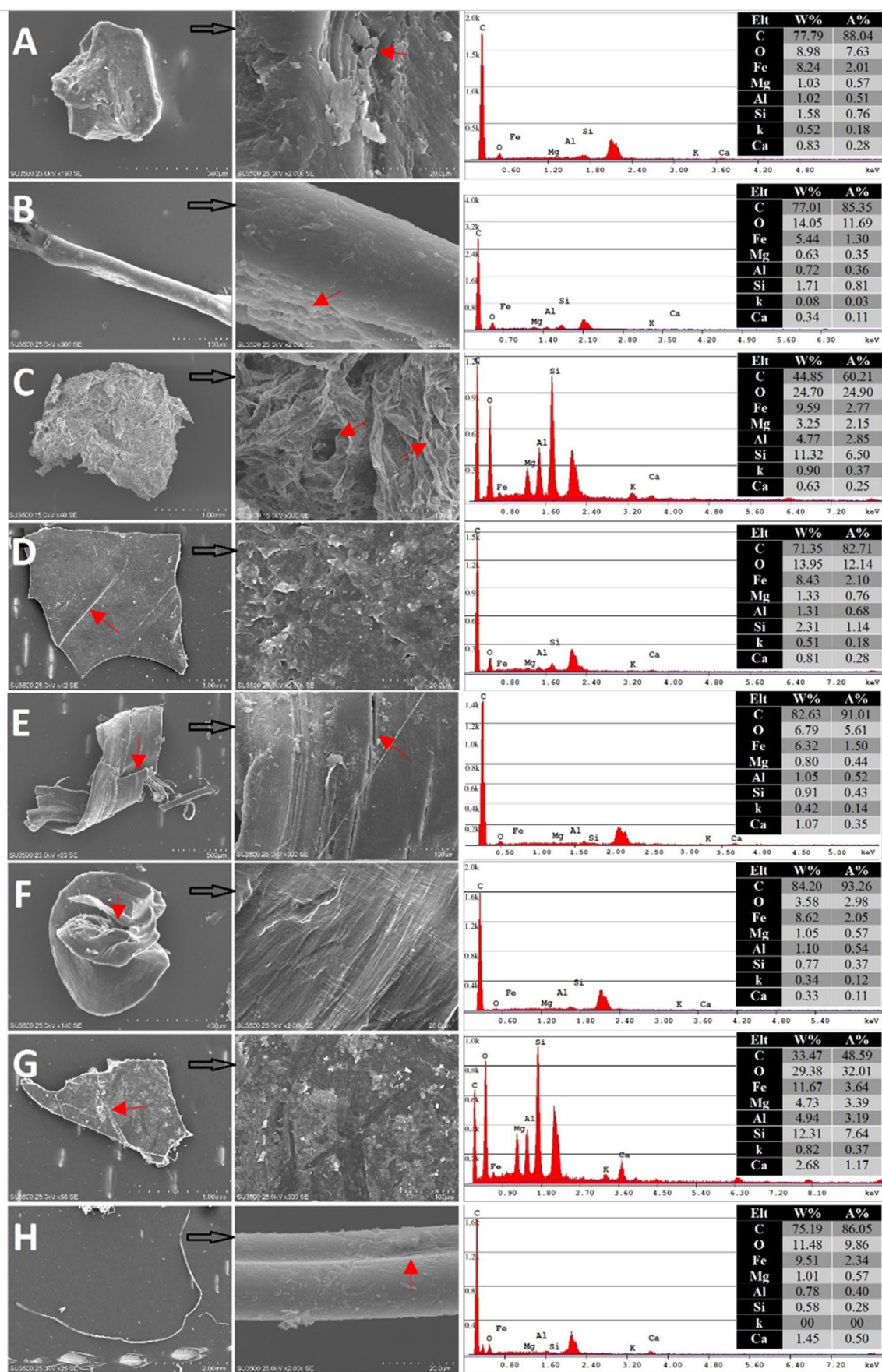
to consider is that regardless of some studies that reported KOH as a suitable solution for the digestion of fish tissues (e.g., Dehaut et al. 2016; Karami et al. 2017), other studies reported that KOH can degrade some polymers such as PET and PVC (Pfeiffer and Fischer 2020). Therefore, it can affect the abundance of MPs found in fish in this study.

There was no correlation in MP abundance between surface water with sediment and also sampling sites in sediments and surface waters with fish species. It can be due to several complicated factors such as water quality, water circulation, and the MP characteristics. Similarly, Yuana et al. (2019) did not found a significant correlation between MP abundance in surface waters and sediments in Poyang Lake, China. On the contrary, Zhang et al. (2020) detected a positive correlation between the MP abundance in fish and sediment based on the different shapes and colors, while no correlation was found between fish and water samples according to any classification in Shengsi, China.

Significant correlations were found between fish body weight, body length, and GI tract weight with MP abundance isolated from the fish GI tract in *C. arel* ( $P < 0.05$ ), in line with the results of Huang et al. (2020) who reported the same relationships in the fish caught from Zhanjiang mangrove wetland, China. As a result, it seems that by increasing fish size (length and weight) and GI tract weight, the number of MPs in the *C. arel* will be raised, while it is not true for *S. forsteri*, *L. abu*, and *O. ruber* since the abundance of MPs did not show significant relationships with body weight, body length, and GI tract weight ( $P > 0.05$ ).

The result of this study showed that fiber was the most typical shape of MPs found in the surface waters, sediments, and fish. Domestic sewage, degradation of fishing net, and textile are considered as the most common origin of fibers in the marine environments (Browne et al. 2011; Cole et al. 2011). A high amount of fibers in the present study can be concluded from fishing gears such as rope and net, since our sampling sites were located in the areas with high fishing activity and also most polymer type of fiber found in this study was PP and PE which are used for fishery industry. This is consistent with several previous studies in the Persian Gulf which reported fibers as the main shape (Naji et al. 2017, 2019 and Kor and Mehdinia 2020). Fragment and films were also found in this study that can be originated from the fragmentation of larger plastics such as plastic bottles and bags (Rocha-Santos and Duarte 2017). Consequently, it can be stated that secondary MPs (taken from segregation of larger plastics) were the main types of shape in the Gulf.

Regarding the color, black, blue, and red were the most dominant in all sampling sites and fish GI tract. A reason for the higher number of colored particles may be that these particles are more easily identified by visual detection compared with white or transparent particles. Although, the





**Fig. 4** SEM/EDS analysis of found MPs in the sediment MPs (A fragment, B fiber), surface water (D, E fragment, C film), and fish GI tract (F, G fragment, H fiber). Left: SEM picture with low magnification; center: SEM picture with high magnification; right: EDS results showing the different elements found with respective percentages. Red arrows illustrate the cracks and different structures of MPs

most likely reason can be due to their high frequency in the environment. These colored MPs likely originated from anthropogenic sources such as synthetic textiles (clothing and carpets from beach dwellers), breaking down of consumer products, and releasing colored fibers through the fishing industry into the Persian Gulf. Other studies found that white and blue (Kor and Mehdinia 2020), black (Naji et al. 2019), blue and white (Castillo et al. 2016), and black or gray (Abbasi et al. 2018) are the most prevalent colors in the sediments, surface waters, and biota of the Persian Gulf. In fact, MP colors can be applied to initially assess the origins and types of chemical compounds and also the source of MPs (Rocha-Santos and Duarte 2017). However, colors can change by weathering and the formation of biofilms (Eriksen et al. 2013; Stolte et al. 2015). Therefore, this results in a nearly controversial discussion about colors. Furthermore, colored MPs may attract predators since they can resemble the prey color (Kühn et al. 2015; Abayomi et al. 2017) reflecting the increasing concerns about colored MP particles, although, in this study, a significant correlation was not found between MP color in GI tract contents compared to the water and sediment.

In terms of size, the most predominant MPs were smaller than 2000  $\mu\text{m}$  in the present study. Similarly, other studies in the English Channel (Lusher et al. 2013), Lake Hovsgol, Mongolia (Free et al. 2014), and Haikou Bay, China (Qi et al. 2020) reported particles > 2000  $\mu\text{m}$  as the predominant size in the sediments, surface waters, and fish. Previous studies in the Persian Gulf showed that particles 1000–3000  $\mu\text{m}$  (Kor and Mehdinia 2020), > 2000– $\leq$  5000  $\mu\text{m}$  (Dobaradaran et al. 2018), and 100–500  $\mu\text{m}$  (Abbasi et al. 2018) were the most common MP size in the surface waters, sediments, and biota, respectively, although sampling methods can influence the MP size composition of this study with other studies. Kor and Mehdinia (2020) reported that MP size distribution in the Persian Gulf likely is in conformity with MP size in global ocean surface water. In fact, particle size is an important factor affecting MP bioavailability and bioaccumulation since their size can be similar to the food size of organisms (Dantas et al. 2012; Ory et al. 2018). Therefore, the abundance of MPs, especially smaller sizes (< 2000  $\mu\text{m}$ ), detected in the present study raises the ingestion potential through a range of biota.

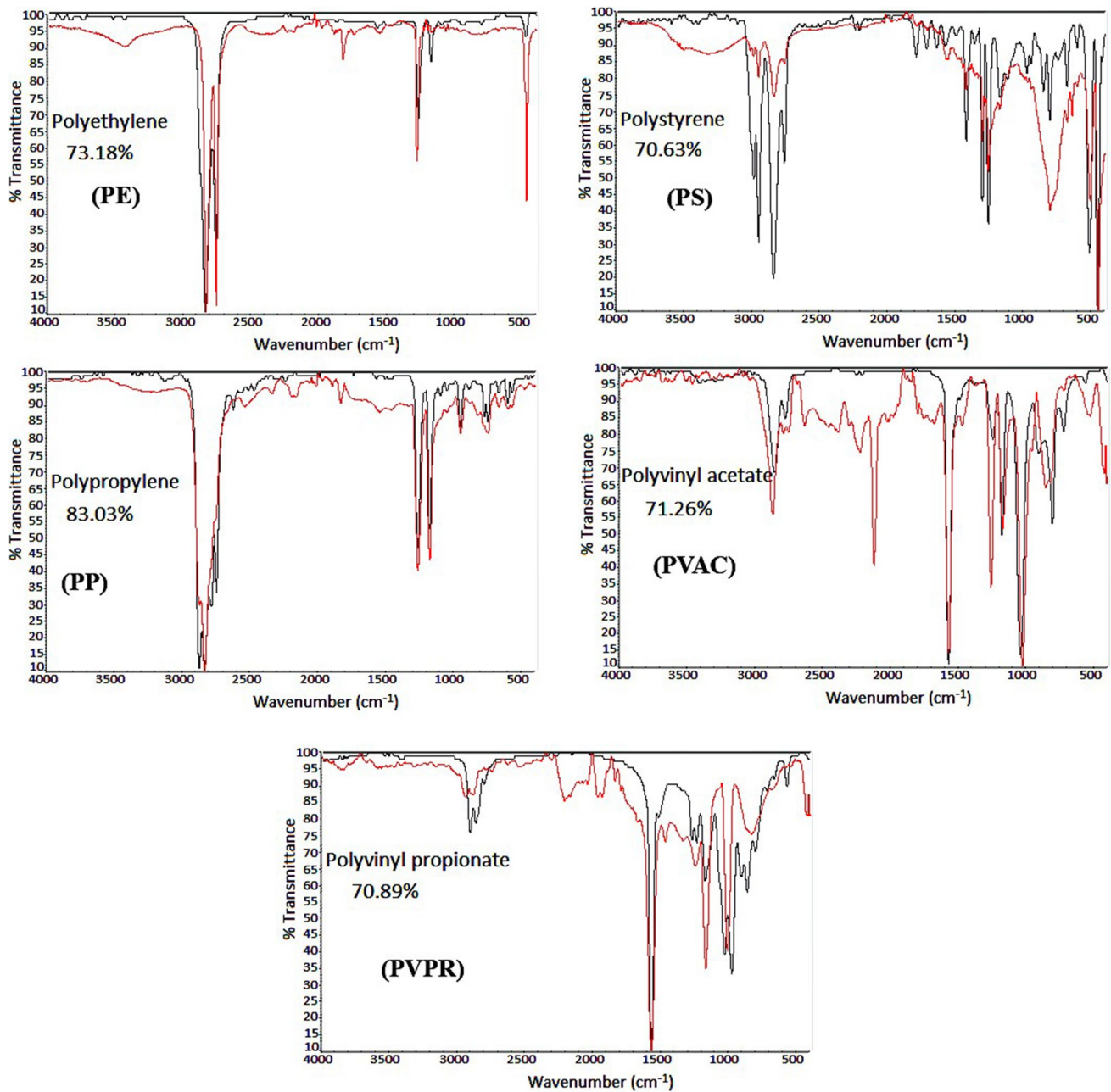
Based on EDS analyses, carbon and oxygen were the main elements of particles in this study which are considered as the main components of plastics (do Sul and Costa

2014). Also, silicon is applied in polymeric materials (Mehdinia et al. 2020). Consequently, it can be concluded that the analyzed particles in this study were microplastic, although other elements such as iron, aluminum, calcium, potassium, and magnesium detected on particles can be absorbed by MPs from the surrounding area (Liu et al. 2019; Mataji et al. 2020) or they can be used as chemical additives in the production process of plastic materials (do Sul and Costa 2014). The cracks and abrasions observed on the surface of MPs can indicate the period of MP exposure to the environment, weathering by physical or chemical agents, or mechanical degradation that occurred in larger pieces of plastics (Abbasi et al. 2018; Liu et al. 2019). These developed structures on the surfaces of MPs have already intensified their capacity to absorb various environmental pollutants, including heavy metals and organic pollutants (such as stable organic pollutants and polycyclic aromatic hydrocarbons) (Wardrop et al. 2016; Wen et al. 2018).

The result of ATR-FTIR spectrometry indicated that 63.3% of particles were MPs, while PP (density 0.90  $\text{g}/\text{cm}^3$ ), PE (0.91–0.96  $\text{g}/\text{cm}^3$ ), PS (1.05  $\text{g}/\text{cm}^3$ ), PVAC (1.19  $\text{g}/\text{cm}^3$ ), and PVPR (0.91  $\text{g}/\text{cm}^3$ ) were the dominant polymers. The low ratio of validation particles as plastics in this study can be due to inadequate spectrum intensity of fibers by ATR-FTIR spectrometry; nevertheless, none of the test particles (36.7%) were identified as natural organic materials. Besides, the density of all found polymers in the current study was lower than 1.2  $\text{g}/\text{cm}^3$  which can indicate the limits of suspending high-density MPs by NaCl. Fishing nets, supermarket bags, packaging, and plastic bottles are the most important sources of PP, PE, and PS in aquatic ecosystems (Claessens et al. 2011; Elvers 2016; Fu et al. 2020). In the present study, sampling sites (e.g., Bushehr, Bandar Abbas, and Qeshm) were situated in the regions with high fishing activity, leading to the generation of fragmented ropes and nets (which are usually made of PP and PE) into the Gulf. Moreover, the tourist industry, recreational and commercial boating, and releasing of untreated sewage by dwelling buildings near beaches (De Sales-Ribeiro et al. 2020; Rasta et al., 2020) can transport a variety of types of polymers into the Persian Gulf.

## Conclusion

In this study, microplastic pollution was investigated in the surface waters, sediments, and four fish species in 7 sampling stations located on the northern shores of the Persian Gulf. MPs were found in all sampling stations and fish species. There was no correlation between MP abundance in surface water, sediment, and fish samples. Except for *C. arel*, the abundance of MPs did not show significant relationships with body weight, body length,



**Fig. 5** ATR-FTIR of representative samples identified in the north of Persian Gulf (black line is polymer reference; red line is sample polymer)

and GI tract weight ( $P > 0.05$ ). Although the abundance of MPs in fish and sampling stations in the present study was lower than in other regions of the world, we suggest to continue monitoring MP prevalence in sediments, seawaters, and coastal fishes from the Persian Gulf to recognize the changes in time. We also recommend that MP pollution must be investigated in the fish food chain of these areas.

**Acknowledgements** This study was financially supported by the Iran National Science Foundation (INSF) (Grant No: 96016853).

**Authors' contributions** AA: formal analysis. MST: methodology and data curation. MB: methodology and writing—original draft. MR: data curation and writing—original draft. HTJ: formal analysis. BRA: conceptualization and methodology.

**Data availability** All data in this article will be available upon request.

## Declarations

**Ethics approval** The authors pledged that no data from this study have been published elsewhere.

**Consent to participate** To avoid damage to the fish stock, the lowest number of fish sample was collected.

**Consent for publication** All the authors agreed to publish the data in this journal.

**Conflict of interest** The authors declare no competing interests.

## References

- Abayomi OA, Range P, Al-Ghouti MA, Obbard JP, Almeer SH, Ben-Hamadou R (2017) Microplastics in coastal environments of the Arabian Gulf. *Mar Pollut Bull* 124(1):181–188. <https://doi.org/10.1016/j.marpolbul.2017.07.011>
- Abbasi S, Soltani N, Keshavarzi B, Moore F, Turner A, Hassanaghaei M (2018) Microplastics in different tissues of fish and prawn from the Musa Estuary, Persian Gulf. *Chemosphere* 205:80–87. <https://doi.org/10.1016/j.chemosphere.2018.04.076>
- Abidli S, Toumi H, Lahbib Y, El Menif NT (2017) The first evaluation of microplastics in sediments from the complex lagoon-channel of Bizerte (Northern Tunisia). *Water Air Soil Pollut* 228(7):262. <https://doi.org/10.1007/s11270-017-3439-9>
- Akdogan Z, Guven B (2019) Microplastics in the environment: a critical review of current understanding and identification of future research needs. *Environ Pollut* 113011. <https://doi.org/10.1016/j.envpol.2019.113011>
- Akhbarizadeh R, Moore F, Keshavarzi B (2018) Investigating a probable relationship between microplastics and potentially toxic elements in fish muscles from northeast of Persian Gulf. *Environ Pollut* 232:154–163. <https://doi.org/10.1016/j.envpol.2017.09.028>
- Akoueson F, Sheldon LM, Danopoulos E, Morris S, Hotten J, Chapman E, Li J, Rotshell, J.M., 2020. A preliminary analysis of microplastics in edible versus non-edible tissues from seafood samples. *Environmental pollution*, p.114452. <https://doi.org/10.1016/j.envpol.2020.114452>
- Arias AH, Ronda AC, Oliva AL, Marcovecchio JE (2019) Evidence of microplastic ingestion by fish from the Bahía Blanca estuary in Argentina. *South America Bull Environ Contam Toxicol* 102(6):750–756. <https://doi.org/10.1007/s00128-019-02604-2>
- Auta HS, Emenike CU, Fauziah SH (2017) Distribution and importance of microplastics in the marine environment: a review of the sources, fate, effects, and potential solutions. *Environ Int* 102:165–176. <https://doi.org/10.1016/j.envint.2017.02.013>
- Baalkhuyur FM, Qurban MA, Panickan P, Duarte CM (2020) Microplastics in fishes of commercial and ecological importance from the Western Arabian Gulf. *Mar Pollut Bull* 152:110920. <https://doi.org/10.1016/j.marpolbul.2020.110920>
- Bakir A, Rowland SJ, Thompson RC (2014) Transport of persistent organic pollutants by microplastics in estuarine condition. *Estuar Coast Shelf Sci* 140:14–21. <https://doi.org/10.1016/j.ecss.2014.01.004>
- Besseling E, Foekema EM, Van Franeker JA, Leopold MF, Kühn S, Bravo Rebolledo EL, Heße E, Mielke L, IJzer J, Kamminga P, Koelmans AA (2015) Microplastic in a macro filter feeder: Humpback whale *Megaptera novaeangliae*. *Mar Pollut Bull*. <https://doi.org/10.1016/j.marpolbul.2015.04.007>
- Bibak M, Tahmasebi S, Sattari M, Kafaei R, Ramavandi B (2021) Empirical cumulative entropy as a new trace elements indicator to determine the relationship between algae-sediment pollution in the Persian Gulf, Southern Iran. *Environ Sci Pollut Res*. <https://doi.org/10.1007/s11356-020-10838-5>
- Browne MA, Crump P, Niven SJ, Teuten E, Tonkin A, Galloway T, Thompson R (2011) Accumulation of microplastic on shorelines worldwide: sources and sinks. *Environ Sci Technol* 45(21):9175–9179. <https://doi.org/10.1021/es201811s>
- Carr SA, Liu J, Tesoro AG (2016) Transport and fate of microplastic particles in wastewater treatment plants. *Water Res* 91:174–82. <https://doi.org/10.1016/j.watres.2016.01.002>
- Castillo AB, Al-Maslamani I, Obbard JP (2016) Prevalence of microplastics in the marine waters of Qatar. *Mar Pollut Bull* 111(1–2):260–267
- Claessens M, De Meester S, Van Landuyt L, De Clerck K, Janssen CR (2011) Occurrence and distribution of microplastics in marine sediments along the Belgian coast. *Mar Pollut Bull* 62(10):2199–2204. <https://doi.org/10.1016/j.marpolbul.2011.06.030>
- Cole M, Lindeque P, Halsband C, Galloway TS (2011) Microplastics as contaminants in the marine environment: a review. *Mar Pollut Bull* 62(12). <https://doi.org/10.1016/j.marpolbul.2011.09.025>
- Dantas DV, Barletta M, Costa MF (2012) The seasonal and spatial patterns of ingestion of polyfilament nylon fragments by estuarine drums (*Sciaenidae*). *Environ Sci Pollut Res* 19:600–606. <https://doi.org/10.1007/s11356-011-0579-0>
- Dehaut A, Cassone AL, Frère L, Hermabessiere L, Himber C, Rinnert E, Rivière G, Lambert C, Soudant P, Huvet A, Duflos G, Paul-Pont I (2016) Microplastics in seafood: benchmark protocol for their extraction and characterization. *Environ Pollut* 215:223–233. <https://doi.org/10.1016/j.envpol.2016.05.018>
- De Sales-Ribeiro C., Brito-Casilla Y, Fernandez A, Caballero MJ (2020) An end to the controversy over the microscopic detection and effects of pristine microplastics in fish organs. *Sci. Rep.* 24;10(1):12434. <https://doi.org/10.1038/s41598-020-69062-3>
- Dobaradaran S, Schmidt TC, Nabipour I, Khajehmadi N, Tajbakhsh S, Saeedi R, Mohammadi MJ, Keshkar M, Khorsand M, Ghasemi FF (2018) Characterization of plastic debris and association of metals with microplastics in coastline sediment along the Persian Gulf. *Waste Manage* 78:649–658. <https://doi.org/10.1016/j.wasman.2018.06.037>
- do Sul JAI, Costa MF, (2014) The present and future of microplastic pollution in the marine environment. *Environ Pollut* 185:352–364. <https://doi.org/10.1016/j.envpol.2013.10.036>
- Elvers B (ed) (2016) Ullmann's polymers and plastics: products and processes. Wiley-VCH, Germany
- Eriksen M, Maximenko N, Thiel M, Cummins A, Lattin G, Wilson S, Hafner J, Zellers A, Rifman S (2013) Plastic pollution in the South Pacific subtropical gyre. *Mar Pollut Bull* 68(1–2):71–76. <https://doi.org/10.1016/j.marpolbul.2012.12.021>
- Ferreira P, Fonte E, Soares ME, Carvalho F, Guilhermino L (2016) Effects of multi-stressors on juveniles of the marine fish *Pomatoschistus microps*: gold nanoparticles, microplastics and temperature. *Aquat Toxicol*. 2016 Jan;170:89–103. <https://doi.org/10.1016/j.aquatox.2015.11.011>
- Free CM, Jensen OP, Mason SA, Eriksen M, Williamson NJ, Boldgiv B (2014) High-levels of microplastic pollution in a large, remote, mountain lake. *Mar Pollut Bull* 85:156–163. <https://doi.org/10.1016/j.marpolbul.2014.06.001>
- Fu Z, Chen G, Wang W, Wang J (2020) Microplastic pollution research methodologies, abundance, characteristics and risk assessments for aquatic biota in China. *Environ Pollut*. Nov; 266 (Pt 3):115098. <https://doi.org/10.1016/j.envpol.2020.115098>
- Güven O, Gökdağ K, Jovanović B, Kideys AE, (2017) Microplastic litter composition of the Turkish territorial waters of the Mediterranean Sea, and its occurrence in the gastrointestinal tract of fish. *Environ Pollut* 223:286–294. <https://doi.org/10.1016/j.envpol.2017.01.025>



- Graca B, Szewc K, Zakrzewska D, Dołęga A, Szczerbowska-Boruchowska M (2017) Sources and fate of microplastics in marine and beach sediments of the Southern Baltic Sea—a preliminary study. *Environ Sci Pollut Res* 24(8):7650–7661. <https://doi.org/10.1007/s11356-017-8419-5>
- Hermabessiere L, Dehaut A, Paul-Pont I, Lacroix C, Jezequel R, Soudant P, Duflos G (2017) Occurrence and effects of plastic additives on marine environments and organisms: a review. *Chemosphere* 182:781–793. <https://doi.org/10.1016/j.chemosphere.2017.05.096>
- Hidalgo-Ruz V, Gutow L, Thompson RC, Thiel M (2012) Microplastics in the marine environment: a review of the methods used for identification and quantification. *Environ Sci Technol* 46(6):3060–3075. <https://doi.org/10.1021/es2031505>
- Hirai H, Takada H, Ogata Y, Yamashita R, Mizukawa K, Saha M, Kwan C, Moore C, Gray H, Laursen D, Zettler ER, Farrington JW, Reddy CM, Peacock EE, Ward MW (2011) Marine Pollution Bulletin, 29 Jun 2011, 62(8):1683–1692. <https://doi.org/10.1016/j.marpolbul.2011.06.004>
- Holmes LA, Turner A, Thompson RC (2012) Adsorption of trace metals to plastic resin pellets in the marine environment. *Environ Pollut* 160:42–48. <https://doi.org/10.1016/j.envpol.2011.08.052>
- Hosseini M, Nabavi SM, Nabavi SN, Pour NA (2015) Heavy metals (Cd, Co, Cu, Ni, Pb, Fe, and Hg) content in four fish commonly consumed in Iran: risk assessment for the consumers. *Environ Monit Assess* 187:237. <https://doi.org/10.1007/s10661-015-4464-z>
- Huang JS, Koongolla JB, Li HX, Lin L, Pan YF, Liu S, He WH, Maharana D, Xu XR (2020) Microplastic accumulation in fish from Zhanjiang mangrove wetland. *South China Sci Total Environ* 708:134839. <https://doi.org/10.1016/j.scitotenv.2019.134839>
- Hosseini R, Hossein Sayadia M, Aazami J, Savabieasfehanic M (2020) Accumulation and distribution of microplastics in the sediment and coastal water samples of Chabahar Bay in the Oman Sea, Iran. *Mar Pollut Bull* Volume 160, November 2020, 111682. <https://doi.org/10.1016/j.marpolbul.2020.111682>
- Karami A, Golieskardi A, Choo CK, Romano N, Ho YB, Salamatinia B (2017) A high-performance protocol for extraction of microplastics in fish. *Sci Total Environ* 578:485–494. <https://doi.org/10.1016/j.scitotenv.2016.10.213>
- Kashiwada S (2006) Distribution of nanoparticles in the sea-through medaka (*Oryzias latipes*). *Environ Health Perspect* 114(11):1697–1702. <https://doi.org/10.1289/ehp.9209>
- Kor K, Mehdiinia A (2020) Neustonic microplastic pollution in the Persian Gulf. *Mar Pollut Bull* 150, January 2020, 110665. <https://doi.org/10.1016/j.marpolbul.2019.110665>
- Kühn S, Van Oyen A, Booth AM, Meijboom A, Van Franeker JA (2018) Marine microplastic: preparation of relevant test materials for laboratory assessment of ecosystem impacts. *Chemosphere* 213:103–113. <https://doi.org/10.1016/j.chemosphere.2018.09.032>
- Li B, Su L, Zhang H, Deng H, Chen Q, Shi H (2020) Microplastics in fishes and their living environments surrounding a plastic production area. *Sci. Total Environ.* p.138662. <https://doi.org/10.1016/j.scitotenv.2020.138662>
- Lin L, Ma LS, Li HX, Pan YF, Liu S, Zhang L, Peng JP, Fok L, Xu XR, He WH (2020) Low level of microplastic contamination in wild fish from an urban estuary. *Mar. Pollut. Bull.* 160, p.111650.
- Liu S, Jian M, Zhou L, Li W (2019) Distribution and characteristics of microplastics in the sediments of Poyang Lake. *China Water Sci Technol* 79(10):1868–1877. <https://doi.org/10.2166/wst.2019.185>
- Löder MGJ, Gerdt G (2015) Methodology used for the detection and identification of microplastics—a critical appraisal. In: Bergmann, M., Gutow, L., Klages, M. (Eds.), *Marine Anthropogenic Litter*. Springer, Cham, pp. 201–227. [https://doi.org/10.1007/978-3-319-16510-3\\_8](https://doi.org/10.1007/978-3-319-16510-3_8)
- Lusher AL, Mchugh M, Thompson RC (2013) Occurrence of microplastics in the gastrointestinal tract of pelagic and demersal fish from the English Channel. *Mar Pollut Bull* 67(1–2):94–99. <https://doi.org/10.1016/j.marpolbul.2012.11.028>
- Martin J, Lusher A, Thompson RC (2017) The deposition and accumulation of microplastics in marine sediments and bottom water from the Irish Continental Shelf. *Sci Rep.* 7, 10772. <https://doi.org/10.1038/s41598-017-11079-2>
- Martins J, Sobral P (2011) Plastic marine debris on the Portuguese coastline: a matter of size? *Mar Pollut Bull* 62(12):2649–2653. <https://doi.org/10.1016/j.marpolbul.2011.09.028>
- Mataji A, Taleshi MS, Balimoghaddas E (2020) Distribution and characterization of microplastics in surface waters and the southern Caspian Sea coasts sediments. *Arch Environ Contam Toxicol* 78(1):86–93. <https://doi.org/10.1007/s00244-019-00700-2>
- Mehdiinia A, Dehbandi R, Hamzehpour A, Rahnama R (2020) Identification of microplastics in the sediments of southern coasts of the Caspian Sea, north of Iran. *Environ Pollut* 258:113738. <https://doi.org/10.1016/j.envpol.2019.113738>
- Nabizadeh R, Sajadi M, Rastkari N, Yaghmaei K (2019) Microplastic pollution on the Persian Gulf shoreline: a case study of Bandar Abbas city, Hormozgan Province. *Iran Mar Pollut Bull* 145:536–546. <https://doi.org/10.1016/j.marpolbul.2019.06.048>
- Naji A, Esmaili Z, Mason SA, Vethaak AD (2017) The occurrence of microplastic contamination in littoral sediments of the Persian Gulf. *Iran Environ Sci Pollut Res* 24(25):20459–20468. <https://doi.org/10.1007/s11356-017-9587-z>
- Naji A, Nuri M, Amiri P, Niyogi S (2019) Small microplastic particles (S-MPPs) in sediments of mangrove ecosystem on the northern coast of the Persian Gulf. *Mar. Pollut Bull.* 146, pp.305–311. <https://doi.org/10.1016/j.marpolbul.2019.06.033>
- Neves D, Sobral P, Lia Ferreira J, Pereira T (2015) Ingestion of microplastics by commercial fish off the Portuguese coast. *Mar Pollut Bull* 101(1):119–126. <https://doi.org/10.1016/j.marpolbul.2015.11.008>
- Nor NHM, Obbard JP (2014) Microplastics in Singapore’s coastal mangrove ecosystems. *Mar Pollut Bull* 79(1–2):278–283. <https://doi.org/10.1016/j.marpolbul.2013.11.025>
- Ory N, Chagnon C, Felix F, Fernández C, Ferreira JL, Gallardo C, Ordóñez OG, Henostroza A, Laaz E, Mizraji R, Mojica H (2018) Low prevalence of microplastic contamination in planktivorous fish species from the southeast Pacific Ocean. *Mar Pollut Bull* 127:211–216. <https://doi.org/10.1016/j.marpolbul.2017.12.016>
- Pannetier P, Morin B, Le Bihanic F, Dubreil L, Clérancieu C, Chouvet F, Van Arkel K, Danion M, Cacho J (2020) Environmental samples of microplastics induce significant toxic effects in fish larvae. *Environ Int* 134. <https://doi.org/10.1016/j.envint.2019.105047>
- Pedà C, Caccamo L, Fossi MC, Gai F, Andaloro F, Genovese L, Perdicchi A, Romeo T, Maricchiolo G (2016) Intestinal alterations in European sea bass *Dicentrarchus labrax* (Linnaeus, 1758) exposed to microplastics: preliminary results. *Environ Pollut* 212:251–256. <https://doi.org/10.1016/j.envpol.2016.01.083>
- Pfeiffer F, Fischer EK (2020) Various digestion protocols within microplastic sample processing—evaluating the resistance of different synthetic polymers and the efficiency of biogenic organic matter destruction. *Front Environ Sci* 8:572424. <https://doi.org/10.3389/fenvs.2020.572424>
- Qi H, Fu D, Wang Z, Gao M, Peng L (2020) Microplastics occurrence and spatial distribution in seawater and sediment of Haikou Bay in the northern South China Sea. *Estuar Coast Shelf Sci* 239:106757. <https://doi.org/10.1016/j.ecss.2020.106757>
- Qiu Q, Tan Z, Wang J, Peng J, Li M, Zhan Z (2016) Extraction, enumeration and identification methods for monitoring microplastics in the environment. *Estuar Coast Shelf Sci* 176:102–109. <https://doi.org/10.1016/j.ecss.2016.04.012>
- Rasta M, Sattari M, Taleshi MS, Namin JI (2020) Identification and distribution of microplastics in the sediments and surface waters of



- Anzali Wetland in the Southwest Caspian Sea, Northern Iran. *Mar Pollut Bull* 160, p.111541. [10.1016/j.marpolbul.2020.111541](https://doi.org/10.1016/j.marpolbul.2020.111541).
- Rasta M, Sattari M, Taleshi MS, Namin JI (2021) Microplastics in different tissues of some commercially important fish species from Anzali Wetland in the Southwest Caspian Sea. *Northern Iran Mar Pollut Bull* 169:112479. <https://doi.org/10.1016/j.marpolbul.2021.112479>
- Rocha-Santos T, Duarte A (Eds.) (2017) Characterization and analysis of microplastics. In: *Comprehensive Analytical Chemistry*, 75. Elsevier.
- Rochman CM, Browne MA, Halpern BS, Hentschel BT, Hoh E, Karapanagioti HK, Rios-Mendoza LM, Takada H, Thompson RC (2013) Classify plastic waste as hazardous. *Nature* 494, 169–171. [14; 494\(7436\):169–71. https://doi.org/10.1038/494169a](https://doi.org/10.1038/494169a)
- Rochman CM, Tahir A, Williams SL, Baxa DV, Lam R, Miller JT, Werorilangi TFC, S, The SJ, (2015) Anthropogenic debris in seafood: plastic debris and fibers from textiles in fish and bivalves sold for human consumption. *Sci Rep* 5:14340. <https://doi.org/10.1038/srep14340>
- Romeo T, Pietro B, Ped`a C, Consoli P, Andaloro F, Fossi MC, (2015) First evidence of presence of plastic debris in stomach of large pelagic fish in the Mediterranean Sea. *Mar Pollut Bull* 95(1):358–361. <https://doi.org/10.1016/j.marpolbul.2015.04.048>
- Savoca S, Capillo G, Mancuso M, Bottari T, Crupi R, Branca C, Romano V, D'Angelo FC, GN, (2019) Spanò Microplastics occurrence in the tyrrhenian waters and in the gastrointestinal tract of two congener species of Seabreams *Environ. Toxicol Pharmacol* 67(2019):35–41. <https://doi.org/10.1016/j.etap.2019.01.011>
- Stolte A, Forster S, Gerdt S, Schubert H (2015) Microplastic concentrations in beach sediments along the German Baltic coast. *Mar Pollut Bull* 99(1–2):216–229. <https://doi.org/10.1016/j.marpolbul.2015.07.022>
- Su L, Deng H, Li B, Chen Q, Pettigrove V, Wu C, Shi H (2019) The occurrence of microplastic in specific organs in commercially caught fishes from coast and estuary area of East China. *J Hazard Mater* 365(2019):716–724. <https://doi.org/10.1016/j.jhazmat.2018.11.024>
- Taghizade M, Khosrotehrani K, Lak R, Aghanabati SA, Peyrowan H (2012) Geochemistry, paleoclimatology and paleogeography of the Northeast Region of the Persian Gulf (case study from Southern Hormuzgan, Iran). *Iranian Journal of Earth Sciences* 4:110–119
- Thompson RC, Olsen Y, Mitchell RP, Davis A, Rowland SJ, John AW, McGonigle D, Russell AE (2004) Lost at sea: where is all the plastic? *Science* 304(5672):838. <https://doi.org/10.1126/science.1094559>
- Toumi H, Abidli S, Bejaoui M (2019) Microplastics in freshwater environment: the first evaluation in sediments from seven water streams surrounding the lagoon of Bizerte (Northern Tunisia). *Environ Sci Pollut Res* 26(14):14673–14682. <https://doi.org/10.1007/s11356-019-04695-0>
- Tourinho PS, Do Sul JAI, Fillmann G (2010) Is marine debris ingestion still a problem for the coastal marine biota of southern Brazil? *Mar Pollut Bull. Mar*;60 (3):396–401. <https://doi.org/10.1016/j.marpolbul.2009.10.013>.
- Van Franeker JA, Blaize C, Danielsen J, Fairclough K, Gollan J, Guse N, Hansen PL, Heubeck M, Jensen JK, Le Guillou G, Olsen B (2011) Monitoring plastic ingestion by the northern fulmar *Fulmarus glacialis* in the North Sea. *Environ Pollut.* 159(10):2609–15. <https://doi.org/10.1016/j.envpol.2011.06.008>
- Von Moos N, Burkhardt-Holm P, Kohler A (2012) Uptake and effects of microplastics on cells and tissue of the blue mussel *Mytilus edulis* after an experimental exposure. *Environ. Sci. Technol.* 16; 46(20):11327–35. <https://doi.org/10.1021/es302332w>.
- Wang J, Li Y, Lu L, Zheng M, Zhang X, Tian H, Wang W, Ru S (2019) Polystyrene microplastics cause tissue damages, sex-specific reproductive disruption and transgenerational effects in marine medaka (*Oryzias melastigma*). *Environ Pollut.* 254, p.113024. <https://doi.org/10.1016/j.envpol.2019.113024>
- Wardrop P, Shimeta J, Nuggeoda D, Morrison PD, Miranda A, Tang M, Clarke BO (2016) Chemical pollutants sorbed to ingested microbeads from personal care products accumulate in fish. *Environ Sci Technol.* 50(7), pp.4037–4044. <https://doi.org/10.1021/acs.est.5b06280>
- Wen X, Du C, Xu P, Zeng G, Huang D, Yin L, Yin Q, Hu L, Wan J, Zhang J, Tan S (2018) Microplastic pollution in surface sediments of urban water areas in Changsha, China: abundance, composition, surface textures. *Mar Pollut Bull* 136:414–423. <https://doi.org/10.1016/j.marpolbul.2018.09.043>
- Xia W, Rao Q, Deng X, Chen J, Xie P (2020) Rainfall is a significant environmental factor of microplastic pollution in inland waters. *Sci Total Environ* 732(25)
- Yuan W, Liu X, Wang W, Di M, Wang J (2019) Microplastic abundance, distribution and composition in water, sediments, and wild fish from Poyang Lake, China. *Ecotoxicol Environ Saf* 170:180–187. <https://doi.org/10.1016/j.ecoenv.2018.11.126>
- Zhang D, Cui Y, Zhou H, Jin C, Yu X, Xu Y, Li Y, Zhang C (2020) Microplastic pollution in water, sediment, and fish from artificial reefs around the Ma'an Archipelago, Shengsi China. *Sci Total Environ* 703:134768. <https://doi.org/10.1016/j.scitotenv.2019.134768>

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