**Supplementary Protocol 2: eCLIP-seq Processing Pipeline**

**Requires:**

* Sequencing data (in fastq format).
* Genome STAR index directory (fasta files may be downloaded from UCSC; hg19)
* Repeat element STAR index directory (fasta files may be downloaded from RepBase)
* FASTA file containing barcodes for demultiplexing reads (described below)
* chrom.sizes file (tabbed file containing chromosome name and length, can be downloaded from UCSC; hg19)

**Installation (Programs Used & Version Information):**

This pipeline can be run with the following software installed on your environment. We recommend installation through the [Anaconda](https://anaconda.org/) package manager. Complete documentation can be found here: <https://github.com/yeolab/eclip>

**Yeo Lab Custom Script Versions:**

makebigwigfiles.py: <https://github.com/YeoLab/makebigwigfiles/>

clipper: <https://github.com/YeoLab/clipper>

clip\_analysis: <https://github.com/YeoLab/clipper/releases/tag/1.1>

demux.py: <https://github.com/YeoLab/eclipdemux>

Input normalization and IDR workflow (described below): <https://github.com/YeoLab/merge_peaks/releases/tag/v0.1.0>

**Other programs used:**

FastQC: v. 0.11.8

Cutadapt: v. 1.14

STAR: v. STAR\_2.7.6a

Samtools: v. 1.6

bedToBigBed: v. 2.7

Bedtools: v. 2.27.1

R: v. 3.3.2

fastq-sort: v. 0.8.0

umi\_tools: v. 1.0.0

**Python 2.7.17 :: Anaconda 2.1.0 (64-bit)**

Pysam 0.12.0.1

Bx 0.5.0

HTSeq 0.11.2

Numpy 1.10.2

Pandas 0.23.1

Pybedtools 0.8.1

Sklearn 0.17.1

Scipy 0.17.1

Matplotlib 2.2.2

Gffutils 0.8.7.1

Statsmodels 0.6.1

**perl=5.10.1 (changes to sorting in 5.22 may work but cause slightly different peak output)**

Statistics::Basic 1.6611

Statistics::Distributions 1.02

Statistics::R 0.34

**Outline of workflow:**

* (paired-end only) Demultiplexes paired-end reads using inline barcodes and extracts unique molecular identifiers (UMI) with eclipdemux. (single-end) extract unique molecular identifiers with umi\_tools
* Trims adapters & adapter-dimers with cutadapt
* Maps to repeat elements with STAR and filter
* Maps filtered reads to genome with STAR
* Removes PCR-duplicates with umi\_tools (single-end) or with a custom python script (barcodecollapsepe.py)
* (paired-end only) Merges multiple inline barcodes and filters R1 (uses only R2 for peak calling)
* Calls enriched peak regions (peak clusters) with CLIPPER
* Uses size-matched input sample to normalize and calculate fold-change enrichment within enriched peak regions with custom perl scripts (overlap\_peakfi\_with\_bam\_PE.pl, peakscompress.pl)

# Usage:

**Script Details**

Our processing pipeline is performed using Common Workflow Language ([CWL](https://www.commonwl.org/)) definition files that take sequencing reads and returns per-replicate peaks that can be used as inputs into an IDR-based workflow to merge replicates. Complete documentation can be found here: <https://github.com/yeolab/eclip>

**Human Readable Description of Steps**

**Note**: For paired-end data, until the merging step each script is run twice, once for each barcode used

**Identify unique molecular identifiers (UMIs) (SE)**: Use umi\_tools to extract unique molecular barcodes.

umi\_tools extract \

--random-seed 1 \

--bc-pattern NNNNNNNNNN \

--log EXAMPLE\_SE.rep1\_clip.---.--.metrics \

--stdin file\_R1.fastq.gz \

--stdout EXAMPLE\_SE.rep1.umi.r1.fq

**Demultiplexing inline barcodes and identify UMIs (PE):** Perform demultiplexing using a supplied barcodes FASTA file. Depending on protocol, --length may be longer than 5.

eclipdemux \

--metrics EXAMPLE\_PE.rep1\_clip.---.--.metrics \

--expectedbarcodeida C01 \

--expectedbarcodeidb D8f \

--fastq\_1 file\_R1.fastq.gz \

--fastq\_2 file\_R2.fastq.gz \

--newname rep2\_clip \

--dataset EXAMPLE\_PE \

--barcodesfile yeolabbarcodes\_20170101.fasta \

--length 5

* Use the FASTA ID for --expectedbarcodeida and --expectedbarcodeidb corresponding to each expected barcode of your IP sample. Use "NIL" for barcode-less size-matched input
* --length describes the length of the UMI to save.
* --metrics, --newname, --dataset are all labels that can be used to customize output file names
* --barcodesfile contents are described as follows:

**Inline barcode description:**

Each inline barcode is ligated to the 5’ end of Read1 and its id and sequence are listed below:

A01 ATTGCTTAGATCGGAAGAGCGTCGTGT

B06 ACAAGCCAGATCGGAAGAGCGTCGTGT

C01 AACTTGTAGATCGGAAGAGCGTCGTGT

D8f AGGACCAAGATCGGAAGAGCGTCGTGT

A03 ANNNNGGTCATAGATCGGAAGAGCGTCGTGT

G07 ANNNNACAGGAAGATCGGAAGAGCGTCGTGT

A04 ANNNNAAGCTGAGATCGGAAGAGCGTCGTGT

F05 ANNNNGTATCCAGATCGGAAGAGCGTCGTGT

RiL19/NIL AGATCGGAAGAGCGTCGTGT

(see eCLIP protocol document for full description of these oligos)

We have observed occasional double ligation events on the 5’ end of Read1, and we have found that to fix this requires we run cutadapt twice. Additionally, because two adapters are used for each library (to ensure proper balancing on the Illumina sequencer), two separate barcodes may be ligated to the same Read1 5’ end (often with 5’ truncations). To fix this we split the barcodes up into 15bp chunks so that cutadapt is able to deconvolute barcode adapters properly (as by default it will not find adapters missing the first N bases of the adapter sequence)

The barcodes file is made by appending one of the barcodes below (these are the same barcode sequences used to demultiplex). See example file ([yeolabbarcodes\_20170101.fasta](https://github.com/YeoLab/eclip/blob/master/example/inputs/yeolabbarcodes_20170101.fasta)):

AAGCAAT A01

GGCTTGT B06

ACAAGTT C01

TGGTCCT D8f

ATGACCNNNNT A03

TCCTGTNNNNT G07

CAGCTTNNNNT A04

GGATACNNNNT F05

To the 5’ adapter

CTTCCGATCT

**Cutadapt round 1**: Takes output from demultiplexed files. Run to trim off both 5’ and 3’ adapters on both reads

cutadapt \

-f fastq \

--match-read-wildcards \

--times 1 \

-e 0.1 \

-O 1 \

--quality-cutoff 6 \

-m 18 \

-a NNNNNAGATCGGAAGAGCACACGTCTGAACTCCAGTCAC \

-g CTTCCGATCTACAAGTT \

-g CTTCCGATCTTGGTCCT \

-A AACTTGTAGATCGGA \

-A AGGACCAAGATCGGA \

-A ACTTGTAGATCGGAA \

-A GGACCAAGATCGGAA \

-A CTTGTAGATCGGAAG \

-A GACCAAGATCGGAAG \

-A TTGTAGATCGGAAGA \

-A ACCAAGATCGGAAGA \

-A TGTAGATCGGAAGAG \

-A CCAAGATCGGAAGAG \

-A GTAGATCGGAAGAGC \

-A CAAGATCGGAAGAGC \

-A TAGATCGGAAGAGCG \

-A AAGATCGGAAGAGCG \

-A AGATCGGAAGAGCGT \

-A GATCGGAAGAGCGTC \

-A ATCGGAAGAGCGTCG \

-A TCGGAAGAGCGTCGT \

-A CGGAAGAGCGTCGTG \

-A GGAAGAGCGTCGTGT \

-o EXAMPLE\_PE.rep2\_clip.C01.r1.fqTr.fq \

-p EXAMPLE\_PE.rep2\_clip.C01.r2.fqTr.fq \

EXAMPLE\_PE.rep2\_clip.C01.r1.fq.gz \

EXAMPLE\_PE.rep2\_clip.C01.r2.fq.gz

**Fastqc round 1**: Run and examined by eye to make sure libraries look alright

fastqc -t 2 --extract -k 7 EXAMPLE\_PE.rep2\_clip.C01.r1.fqTr.fq -o .

fastqc -t 2 --extract -k 7 EXAMPLE\_PE.rep2\_clip.C01.r2.fqTr.fq –o .

**Cutadapt round 2**: Takes output from cutadapt round 1. Run to trim off the 3’ adapters on read 2, to control for double ligation events.

cutadapt \

-f fastq \

--match-read-wildcards \

--times 1 \

-e 0.1 \

-O 5 \

--quality-cutoff 6 \

-m 18 \

-A AACTTGTAGATCGGA \

-A AGGACCAAGATCGGA \

-A ACTTGTAGATCGGAA \

-A GGACCAAGATCGGAA \

-A CTTGTAGATCGGAAG \

-A GACCAAGATCGGAAG \

-A TTGTAGATCGGAAGA \

-A ACCAAGATCGGAAGA \

-A TGTAGATCGGAAGAG \

-A CCAAGATCGGAAGAG \

-A GTAGATCGGAAGAGC \

-A CAAGATCGGAAGAGC \

-A TAGATCGGAAGAGCG \

-A AAGATCGGAAGAGCG \

-A AGATCGGAAGAGCGT \

-A GATCGGAAGAGCGTC \

-A ATCGGAAGAGCGTCG \

-A TCGGAAGAGCGTCGT \

-A CGGAAGAGCGTCGTG \

-A GGAAGAGCGTCGTGT \

-o EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.fq \

-p EXAMPLE\_PE.rep2\_clip.C01.r2.fqTrTr.fq \

EXAMPLE\_PE.rep2\_clip.C01.r1.fqTr.fq \

EXAMPLE\_PE.rep2\_clip.C01.r2.fqTr.fq

**Fastqc round 2:** Takes output from STAR rmRep. Runs a second round of fastqc to verify that after read grooming the data still is usable.

fastqc -t 2 --extract -k 7 EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.fq -o .

fastqc -t 2 --extract -k 7 EXAMPLE\_PE.rep2\_clip.C01.r2.fqTrTr.fq –o .

**Fastq-sort**: Takes cutadapt round 2 output and sorts to reduce randomness

fastq-sort --id EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.fq > EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.sorted.fq

fastq-sort --id EXAMPLE\_PE.rep2\_clip.C01.r2.fqTrTr.fq > EXAMPLE\_PE.rep2\_clip.C01.r2.fqTrTr.sorted.fq

**STAR rmRep**: Takes sorted output from cutadapt round 2. Maps to human specific version of RepBase used to remove repetitive elements, helps control for spurious artifacts from rRNA (& other) repetitive reads.

STAR \

--runMode alignReads \

--runThreadN 8 \

--genomeDir homo\_sapiens\_repbase\_v2 \

--genomeLoad NoSharedMemory \

--alignEndsType EndToEnd \

--outSAMunmapped Within \

--outFilterMultimapNmax 30 \

--outFilterMultimapScoreRange 1 \

--outFileNamePrefix EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.sorted.STAR \

--outSAMtype BAM Unsorted \

--outFilterType BySJout \

--outBAMcompression 10 \

--outReadsUnmapped Fastx \

--outFilterScoreMin 10 \

--outSAMattrRGline ID:foo \

--outSAMattributes All \

--outSAMmode Full \

--outStd Log \

--readFilesIn EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.sorted.fq EXAMPLE\_PE.rep2\_clip.C01.r2.fqTrTr.sorted.fq

**Re-name files**: re-name repeat-mapped outputs

mv EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.sorted.STARAligned.out.bam EXAMPLE\_PE.rep2\_clip.C01.r1.fq.repeat-mapped.bam

mv EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.sorted.STARUnmapped.out.mate1 EXAMPLE\_PE.rep2\_clip.C01.r1.fq.repeat-unmapped.fq

mv EXAMPLE\_PE.rep2\_clip.C01.r1.fqTrTr.sorted.STARUnmapped.out.mate2 EXAMPLE\_PE.rep2\_clip.C01.r2.fq.repeat-unmapped.fq

**Fastq-sort**: Takes unmapped output from STAR rmRep and sorts it to account for issues with STAR not outputting first and second mate pairs in order

fastq-sort --id EXAMPLE\_PE.rep2\_clip.C01.r1.fq.repeat-unmapped.fq > EXAMPLE\_PE.rep2\_clip.C01.r1.fq.repeat-unmapped.sorted.fq

fastq-sort --id EXAMPLE\_PE.rep2\_clip.C01.r2.fq.repeat-unmapped.fq > EXAMPLE\_PE.rep2\_clip.C01.r2.fq.repeat-unmapped.sorted.fq

**STAR genome mapping**: Takes output from STAR rmRep. Maps unique reads to the human genome

STAR \

--runMode alignReads \

--runThreadN 8 \

--genomeDir /stage/hg19\_star\_sjdb \

--genomeLoad NoSharedMemory \

--readFilesIn \

EXAMPLE\_PE.rep2\_clip.C01.r1.fq.repeat-unmapped.sorted.fq \

EXAMPLE\_PE.rep2\_clip.C01.r2.fq.repeat-unmapped.sorted.fq \

--outSAMunmapped Within \

--outFilterMultimapNmax 1 \

--outFilterMultimapScoreRange 1 \

--outFileNamePrefix EXAMPLE\_PE.rep2\_clip.C01.r1.fq.repeat-unmapped.sorted.STAR \

--outSAMattributes All \

--outSAMtype BAM Unsorted \

--outFilterType BySJout \

--outReadsUnmapped Fastx \

--outFilterScoreMin 10 \

--outSAMattrRGline ID:foo \

--outStd Log \

--alignEndsType EndToEnd \

--outBAMcompression 10 \

--outSAMmode Full

**Re-name BAM:** rename genome-mapped outputs

mv EXAMPLE\_PE.rep2\_clip.C01.r1.fq.repeat-unmapped.sorted.STARAligned.out.bam EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mapped.bam

**Name sort BAM:** sort output from STAR by name to ensure read pairs are adjacent.

samtools sort -n -o EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.bam EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mapped.bam

**Barcode\_collapse\_pe (PE)**: takes output from STAR genome mapping. Custom random-mer-aware script for PCR duplicate removal.

barcodecollapsepe.py \

-o EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDup.bam \

-m EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDup.metrics \

-b EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.bam

**Position sort BAM**: Takes output from barcode collapse PE (or from SE namesort bam). Sorts resulting bam file for use downstream.

samtools sort -o EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.bam EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDup.bam

**Barcode\_collapse\_se (SE):** takes output from STAR genome mapping. Use umi\_tools dedup to identify the extracted random-mer from the previous step and perform PCR duplicate removal.

umi\_tools dedup \

--random-seed 1 \

---I EXAMPLE\_SE.rep1\_clip.umi.r1.fq.genome-mappedSoSo.bam \

--method unique \

--output-stats EXAMPLE\_SE.rep1\_clip.umi.r1.fq.genome-mappedSoSo.txt \

-S EXAMPLE\_SE.rep1\_clip.umi.r1.fq.genome-mappedSoSo.rmDup.bam

**Samtools index**: Takes output from sortSam, makes bam index for use downstream.

samtools index EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.bam

**Samtools merge (PE only)**: Takes inputs from multiple final bam files. Merges the two technical replicates for further downstream analysis.

samtools merge EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.bam EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.bam EXAMPLE\_PE.rep2\_clip.D8f.r1.fq.genome-mappedSo.rmDupSo.bam

**Samtools index**: Takes output from sortSam, makes bam index for use downstream.

samtools index EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.bam

**Samtools view (PE only)**: Takes output from samtools merge. Only outputs the second read in each pair for use with a single stranded peak caller. This is the final bam file to perform analysis on.

samtools view -f 128 -b -o EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.bam

**Make normalized read density bigwig files**: Takes input from samtools view. Makes bw files to be uploaded to the genome browser or for other visualization. Use --direction f for SE clip as reads are not reversed.

makebigwigfiles \

--bw\_pos EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.norm.pos.bw \

--bw\_neg EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.norm.neg.bw \

--bam EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam \

--genome hg19.chrom.sizes \

--direction r

**Clipper**: Takes results from samtools view. Calls peaks on those files.

clipper \

--species hg19 \

--bam EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam \

--save-pickle \

--outfile EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.peakClusters.bed

**Input normalization:** Compares the number of reads within the IP sample to the number of reads within the size-matched INPUT sample across Clipper-called peak clusters. This step is performed both within this pipeline as well as within the merge\_peaks pipeline using the same perl scripts.

samtools view -cF 4 EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam > ip\_mapped\_readnum.txt

samtools view -cF 4 EXAMPLE\_PE.rep2\_input.NIL.r1.fq.genome-mappedSo.rmDupSo.r2.bam > input\_mapped\_readnum.txt

overlap\_peakfi\_with\_bam\_PE.pl \

EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam \

EXAMPLE\_PE.rep2\_input.NIL.r1.fq.genome-mappedSo.rmDupSo.r2.bam \

EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.peakClusters.bed \

ip\_mapped\_readnum.txt \

input\_mapped\_readnum.txt \

EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.peakClusters.normed.bed

perl compress\_l2foldenrpeakfi\_for\_replicate\_overlapping\_bedformat.pl \

EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.peakClusters.normed.bed \

EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.peakClusters.normed.compressed.bed

**Peak normalization vs SMInput and**

**reproducible peak / IDR analysis**

Peak normalization vs paired SMInput datasets is run as a second processing pipeline (merge\_peaks) available with additional documentation on github (<https://github.com/YeoLab/merge_peaks>). Input files for normalization pipeline include .bam and .peak.bed files (generated through the pipeline above), as well as a manifest file pairing eCLIP datasets with their paired SMInput datasets as follows.

# Requires:

* Read 2 BAM file outputs from the core pipeline for each replicate IP and INPUT (ie. EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam, EXAMPLE\_PE.rep2\_input.NIL.r1.fq.genome-mappedSo.rmDupSo.r2.bam). Or Read1 BAM file outputs if processing SE reads (ie. EXAMPLE\_SE.rep1\_clip.umi.r1.fq.genome-mappedSoSo.rmDup.bam)
* CLIPPER peak files from the core pipeline for each replicate (ie. EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.peakClusters.bed)

# Installation:

This pipeline can be run with the following software installed on your environment. We recommend installation through the [Anaconda](https://anaconda.org/) package manager:

* **perl=5.10.1 (changes to sorting in 5.22 may work but cause slightly different peak output)**
  + Statistics::Basic 1.6611
  + Statistics::Distributions 1.02
  + Statistics::R 0.34
* **IDR=2.0.2 (base environment):**
  + python=3.4.5
  + numpy=1.11.3
  + scipy=0.19.1
  + setuptools=32.3.1.post20170108
  + matplotlib=2.0.0b4
* cwl=1.0

# Outline of workflow:

* Normalize CLIP BAM over INPUT for each replicate (overlap\_peakfi\_with\_bam.cwl)
* Peak compression/merging on input-normalized peaks for each replicate (compress\_l2foldenrpeakfi\_for\_replicate\_overlapping\_bedformat\_outputfull.cwl)
* Entropy calculation on CLIP and INPUT read probabilities within each peak for each replicate (make\_informationcontent\_from\_peaks.cwl)
* Reformat \*.full files into \*.bed files for each replicate (full\_to\_bed.cwl)
* Run IDR on peaks ranked by entropy (idr.cwl)
* Calculates summary statistics at different IDR cutoffs (parse\_idr\_peaks.cwl)
* Normalize CLIP BAM over INPUT using new IDR peak positions (overlap\_peakfi\_with\_bam.cwl)
* Identifies reproducible peaks within IDR regions (get\_reproducing\_peaks.cwl)

# Usage:

Below is a description of all fields required to be filled out in the manifest file. See https://github.com/YeoLab/merge\_peaks/blob/master/example/204\_RBFOX2.yaml for a full example for the ENCODE RBFOX2 HepG2 eCLIP experiment.

BAM file containing the merged-barcode (read 2 only) post PCR duplicate removal CLIP reads mapping to the genome for Replicate 1:

rep1\_clip\_bam\_file:

class: File

path: EXAMPLE\_PE.rep1\_clip.A01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam

BAM file containing the merged-barcode (read 2 only) post PCR duplicate removal INPUT reads mapping to the genome for Replicate 1:

rep1\_input\_bam\_file:

class: File

path: EXAMPLE\_PE.rep1\_input.A01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam

BED file containing the called peak clusters for Replicate 1 **Output from CLIPPER**. This pipeline performs input normalization:

rep1\_peaks\_bed\_file:

class: File

path: EXAMPLE\_PE.rep1\_clip.A01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.peakClusters.bed

BAM file containing the merged-barcode (read 2 only) post PCR duplicate removal CLIP reads mapping to the genome for Replicate 2:

rep2\_clip\_bam\_file:

class: File

path: EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam

BAM file containing the merged-barcode (read 2 only) post PCR duplicate removal INPUT reads mapping to the genome for Replicate 2:

rep2\_input\_bam\_file:

class: File

path: EXAMPLE\_PE.rep2\_input.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.bam

BED file containing the called peak clusters for Replicate 2 **Output from CLIPPER**. This pipeline performs input normalization:

rep2\_peaks\_bed\_file:

class: File

path: EXAMPLE\_PE.rep2\_clip.C01.r1.fq.genome-mappedSo.rmDupSo.merged.r2.peakClusters.bed

Final output files:

### FINAL OUTPUTS

merged\_peaks\_custombed: reproducible\_peaks.bed

merged\_peaks\_bed: reproducible\_peaks.custombed

# To run the workflow:

* Ensure that the yaml file is accessible and that wf\_get\_reproducible\_eclip\_peaks.cwl is in your $PATH.
* Type: ./204\_RBFOX2.yaml

# Outputs

* \*.merged\_peaks\_bed: this is the BED6 file containing reproducible peaks as determined by entropy-ordered peaks between two replicates.
  + chromosome
  + start
  + end
  + geomean of the log2 fold changes
  + minimum of the -log10 p-value between two replicates
  + strand This is probably what will be useful.
* \*.full files: these tabbed outputs have the following columns (in order):
  + chromosome
  + start
  + end
  + name (colon separated region)
  + reads in CLIP
  + reads in INPUT
  + p-value
  + chi value or (F)isher
  + (F)isher or (C)hi square test
  + enriched or depleted
  + negative log10p value (set to 400 if Perl Statistics::Distributions reports ‘p = 0’)
  + log2 fold change
  + entropy
* \*.idr.out: direct output from IDR, used only as an intermediate for QC checks
  + IDR chromosome
  + IDR start
  + IDR end
  + IDR name
  + IDR score (min(int(log2(-125*IDR*), 1000))
  + IDR strand
  + localIDR (-log10(*Local IDR*))
  + globalIDR (-log10(*Global IDR*))
  + Rep1 merged peak start
  + Rep1 merged peak end
  + Rep1 entropy
  + Rep2 merged peak start
  + Rep2 merged peak end
  + Rep2 entropy
* \*.idr.out.normed.bed: output from IDR as a bed file
  + chromosome
  + start
  + end
  + minimum of the -log10(p-value) of the region in rep1 and rep2
  + geometric mean of the l2fc
  + strand
* \*.custombed: contains individual replicate information. The headers are:
  + IDR region (entire IDR identified reproducible region)
  + peak (reproducible peak region)
  + geomean of the l2fc
  + rep1 log2 fold change
  + rep2 log2 fold change
  + rep1 -log10 p value
  + rep2 -log10 p value

**Changelog**:

1.P.2 20160426 – Clarified section regarding inline demultiplexing to specify which steps occur to generate fastq files which are submitted to the ENCODE DCC.

2.0 20180724 – Replaced input normalization section with new CWL pipeline, which now includes both input normalization as well as identification of reproducible peaks using the IDR framework

2.2 20200409 - Fixed steps to include steps for SE processing and renamed example files to mirror actual outputs from the latest pipeline version.