# Prediction of survival rate for breast cancer patients

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#### Abstract

Breast cancer is a leading cause of cancer-related mortality among women worldwide. This study develops a predictive model for survival outcomes in breast cancer patients using logistic regression, leveraging demographic, clinical, and pathological factors from a prospective cohort dataset. The analysis focuses on identifying key predictors of mortality, evaluating model performance across racial groups, and addressing fairness in prediction accuracy. Tumor stage, grade, and hormone receptor status emerged as significant predictors. The initial logistic regression model achieved a moderate performance, with an area under the receiver operating characteristic curve (ROC-AUC) of 0.74. However, disparities in model performance were observed between racial groups, prompting the implementation of reweighting strategies to enhance fairness. These findings highlight the importance of equitable modeling approaches to improve prognostic accuracy and clinical outcomes in breast cancer care.

### Introduction

Breast cancer is highly prevalent, with X new cases annually worldwide. About 13.1% of women are diagnosed during their lifetime, accounting for 23% of cancer cases and 14% of cancer deaths<sup>1</sup>. Advances in diagnosis and tailored treatments have reduced mortality rates<sup>2</sup>. Survival depends on factors like tumor size, grade<sup>3</sup>, stage, lymph node involvement, socioeconomic status, and race<sup>4</sup>. Younger patients often have better outcomes, while disparities exist for lower-income and African American populations due to late diagnoses<sup>5</sup>. This report analyzes a cohort of breast cancer patients to predict mortality, identify key predictors, and evaluate model fairness across racial groups, aiming to improve outcomes and address disparities.

### Methods

Data Description The dataset includes 10 categorical variables: race (Black, White, Other), marital status (Divorced, Married, Separated, Single, Widowed), tumor stage (T1, T2, T3, T4), lymph node stage (N1, N2, N3), adjusted AJCC 6th stage (IIA, IIB, IIIA, IIIB, IIIC), tumor differentiation (Well, Moderately, Poorly, Undifferentiated), grade (1–4), tumor spread stage (Regional, Distant), estrogen receptor status (Positive, Negative), and progesterone receptor status (Positive, Negative). Additionally, there are 4 continuous variables: age, tumor size, regional nodes examined, and regional nodes positive.

Data Cleaning Column headers were renamed, categorical variables were converted to factors. tumor grade levels were recoded to ensure interpretability. Missing values were assessed, and log transformations were applied to highly skewed continuous variables (tumor size, regional nodes examined, and regional nodes positive) to normalize their distributions.

EDA Methods skim() function is applied to the dataset (model\_data) to compute detailed summary statistics for all variables. (table.1 & 2), Group-wise key statistics are calculated based on survival status(table.3) Cramér's V was used to quantify the strength of the association between categorical variables and the binary outcome (Alive/Dead), with values ranging from 0 (no association) to 1 (perfect association). Distributional plots, including proportional bar plots for race groups and histograms for continuous variables, were created to visualize the data. Boxplots stratified by survival status were used to explore relationships between continuous variables and the binary outcome. A correlation matrix for continuous variables was generated to assess pairwise relationship between variables.

Modeling Assumptions and Transformations Logistic regression was chosen as the primary method due to the binary nature of the outcome. Assumptions checked are: 1. The response variable (status) was confirmed to be binary by code.

2. The alias() function identified collinearity between models: grade2, grade3, and grade4 with other predictors, x6th\_stageIIIC with n\_stage, and differentiate with grade. For simplification, some variables were removed: x6th\_stage captures n\_stage's information, n\_stage was dropped. t\_stage (linked to tumor size), differentiate (overlapping with grade), and regional\_node\_positive (redundant with tumor size and regional\_node\_examined) were removed.

VIF were calculated, to ensure that all values were below 5 (table.4), indicating no multicollinearity. (dataset updated with dropped variable)

- 3. Continuous predictors were log-transformed, and their relationships with the log odds were examined (fig.12). This confirmed linearity. (dataset updated with transformed variable)
- 4. Independence of Errors: Since there were no group-level structures, the independence assumption was satisfied.
- 5. Outliers: Cook's Distance identified potential outliers exceeding 4/n, which flagged numerous points as influential, likely reflecting population variability rather than errors (fig.13). Models with and without these points were compared. Removing these points destabilized coefficients like grade, making them unreliable. Robust logistic regression (model\_robust) mitigated outliers impact, providing stable estimates for predictors (table.5). About 12% of data had reduced influence, while most observations are unaffected. Despite its advantages, we chose the original logistic regression for simplicity and familiarity.

Model Construction and Selection Models were constructed using predictors identified during EDA and assumption checks. Forward, backward, and stepwise selection were applied based on the AIC to select the final model. Interaction effects were tested by examining pairwise interactions between predictors.

Model Validation and Fairness The model was validated using 10-fold cross-validation, evaluating ROC-AUC, sensitivity, and specificity. Fairness was assessed by evaluating model performance across racial subgroups (White, Black, Other) based on subgroup-specific ROC-AUC values. To address disparities, fairness reweighting was applied by adjusting model weights to prioritize underrepresented groups. The reweighted model's performance was compared with the original final model. Predictor coefficients were interpreted as odds ratios, quantifying their impact on survival outcomes.

#### Results

EDA Cramér's V analysis identified x6th\_stage, n\_stage, and hormone receptor statuses as the strongest predictors of survival, while marital status, race, and a\_stage showed weaker associations (fig.1). Bar plots highlight racial disparities, with higher mortality among Non-White, particularly Black patients, and combining groups simplifies comparisons (figs.2-3). Histograms revealed significant right skewness for most continuous variables (except age), improved by log transformations for the other continuous vars (figs.4-5). Boxplots confirm these patterns (figs.6-10). A correlation matrix showed weak overall relationships and no multicollinearity, except regional\_node\_positive is moderately associated with both tumor size (0.24) and regional nodes examined (0.41), guiding modeling

choices (fig.11).

#### Model Selection and Interretation

AIC values differed slightly (Full: 3039.8; Forward: 3039.8; Backward: 3037.5; Stepwise: 3037.5), but all methods yielded the same model. The stepwise model was chosen for its lower AIC (full results attached):

$$Logit(P) = \beta_0 + \beta_1 \times raceBlack + \beta_2 \times raceOther + \ldots + \beta_i \times X_i$$

Where  $\beta_i$  values are taken from Table 10,  $X_i$  represents other significant predictors, and  $Logit(P) = log(\frac{P}{1-P})$ , P is the probability of the status being 1.

**Interaction Effects** Pairwise interactions between predictors were tested, but none of the interaction terms had p-values below 0.05, consistent with earlier analyses showing weak or absent interaction effects (results attached)

Key Findings on Coefficients Table 6 highlights the coefficients for key predictors. Race was significant, with raceBlack (0.47961) associated with higher odds of death and raceOther (-0.42884) linked to lower odds compared to White patients. x6th\_stage was the strongest predictor, with higher stages (e.g., IIIC) significantly increasing mortality risk. Poorly differentiated tumors (Grades 3: 0.91053, Grade 4: 1.87333) also raised mortality odds, while positive hormone receptor status reduced the likelihood of death. Log-transformed predictors showed rn\_examined\_log (-0.29822) improved survival odds, while age\_log (1.09581) indicated higher mortality risk for older patients. Marital status had no strong associations.

Model Performance Cross-validation (10-fold) revealed an overall ROC-AUC of 0.7400, indicating moderately good performance with an acceptable ability to distinguish between "Alive" and "Dead" outcomes. The model demonstrated high sensitivity (0.985), correctly identifying most "Dead" cases, but low specificity (0.122), reflecting difficulty in identifying "Alive" cases (full results attached).

Performance by race groups revealed disparities, the White group had the highest ROC-AUC (0.7504), while the Black group (0.7021) and Other group (0.6584) showed lower performance. The combined minority group (ROC-AUC: 0.7313) improved but remained below the White group, highlighting a fairness gap. Reweighting narrowed this gap, increasing the minority group's ROC-AUC to 0.7358, closer to the White group's 0.7486, improving fairness while maintaining accuracy (table.8). coefficients for reweighted model are in table.7

### Conclusion and Limitation

This study developed a logistic regression model to predict breast cancer survival, highlighting key predictors and addressing racial disparities. Tumor stage and grade were the strongest predictors, with higher stages (e.g., IIIA, IIIB, IIIC) and poorly differentiated tumors (Grades 3 and 4) significantly increasing mortality risk. Positive hormone receptor status improved survival odds, while older age and inadequate lymph node evaluation increased mortality risk. Racial disparities were evident: Black patients faced higher odds of death (OR = 1.5) compared to White patients, while the "Other" race group had lower odds (OR = 0.6).

Reweighting improved fairness for underrepresented groups, increasing the minority group's ROC-AUC from 0.7313 to 0.7358 and narrowing the gap with the White group (0.7486). However, it may reduce generalizability to majority-dominant populations and alter predictor importance, requiring careful interpretation. Stepwise selection treated factor levels as independent variables (e.g., x6th\_stageIIIA as a dummy), potentially excluding some levels and raising interpretability concerns. Marital status was retained in the final model despite weak significance due to its potential theoretical relevance, confounding effects, or fairness considerations. All three aspects—reweighting, factor level treatment, and marital status inclusion—require further research to ensure robustness and interpretability.

### Contribution

Ada Guo wrote the abstract, introduction, and results-eda. Khue Nguyen analyzed results, wrote the conclusion and limitations, refined the results-section structure, and ensured clarity and completeness by adding references. Yixin Zheng wrote the methods and appendix section and constructed the Rmd file(cleaning, correlation matrix, Cramér's V, assumption checks, feature selection, model, cross-validation, fairness reweight).

#### Reference

- 1. Cao SS, Lu CT. Recent perspectives of breast cancer prognosis and predictive factors. Oncol Lett. 2016;12(5):3674-3678. doi:10.3892/ol.2016.5149
- 2. Cancer of the Breast (Female) Cancer Stat Facts. SEER. Accessed December 19, 2024. https://seer.cancer.gov/statfacts/html/breast.html

- 3. Bundred NJ. Prognostic and predictive factors in breast cancer. Cancer Treat Rev.  $2001;27(3):137-142.\ doi:10.1053/ctrv.2000.0207$
- Soerjomataram I, Louwman MWJ, Ribot JG, Roukema JA, Coebergh JWW. An overview of prognostic factors for long-term survivors of breast cancer. Breast Cancer Res Treat. 2008;107(3):309-330. doi:10.1007/s10549-007-9556-1
- 5. Phung MT, Tin Tin S, Elwood JM. Prognostic models for breast cancer: a systematic review. BMC Cancer. 2019;19(1):230. doi:10.1186/s12885-019-5442-6

# Table, Plots, and Code Results

# Tables

Table 1: Skim Summary for Categorical Variables

skim_variable	n_missingcom	plete_ra	ttactor.orde	eredfactor.n_u	niq	ukactor.top_counts
race	0	1	FALSE		3	Whi: 3413, Oth: 320, Bla: 291
marital_status	0	1	FALSE		5	Mar: 2643, Sin: 615, Div: 486,
						Wid: 235
t_stage	0	1	FALSE		4	T2: 1786, T1: 1603, T3: 533,
						T4: 102
n_stage	0	1	FALSE		3	N1: 2732, N2: 820, N3: 472
x6th_stage	0	1	FALSE		5	IIA: 1305, IIB: 1130, III: 1050,
						III: 472
differentiate	0	1	FALSE		4	Mod: 2351, Poo: 1111, Wel:
						543, Und: 19
grade	0	1	FALSE		4	2: 2351, 3: 1111, 1: 543, 4: 19
a_stage	0	1	FALSE		2	Reg: 3932, Dis: 92
estrogen_status	0	1	FALSE		2	Pos: 3755, Neg: 269
progesterone_stat	tus 0	1	FALSE		2	Pos: 3326, Neg: 698
status	0	1	FALSE		2	Ali: 3408, Dea: 616

Table 2: Skim Summary for Numeric Variables

skim_variable n	_missing complet	e_rate	mean	$\operatorname{sd}$	p0	p25	p50	p75	p100	hist
age	0	1 5	53.972167	8.963134	30	47	54	61	69	_
tumor_size	0	1 3	30.473658	21.119696	1	16	25	38	140	<b>-</b>
regional_node_examir	ned 0	1 1	14.357107	8.099675	1	9	14	19	61	
reginol_node_positive	0	1 4	1.158052	5.109331	1	1	2	5	46	

Table 3: Summary Statistics Grouped by Survival Status

status	mean_age sd_a	ge mean_tumor_siz <b>s</b> o	d_tumor_size p	prop_white prop_	_black_other	n_obs
Alive	53.75910 8.8084	29.26878	20.30317	0.8518192	0.1481808	3408
Dead	55.15097 9.6982	91 37.13961	24.11611	0.8279221	0.1720779	616

Table 4: Variance Inflation Factors for Predictors

Variable	GVIF	Df	GVIF_Ratio
age	1.106908	1	1.052097
race	1.058083	2	1.014215
marital_status	1.127489	4	1.015112
x6th_stage	1.967732	4	1.088293
grade	1.118473	3	1.018836
a_stage	1.210950	1	1.100432
tumor_size	1.365304	1	1.168462
estrogen_status	1.484914	1	1.218570
progesterone_status	1.434692	1	1.197786
regional_node_examined	1.225729	1	1.107127

Table 5: Unstable Coefficients

Variable	Full_Coef	Full_SE	No_Outliers_Coef	No_Outliers_SE	Robust_Coef	Robust_SE
(Intercept)	-5.8924387	1.3069930	-30.590617	438.7266805	-5.0273109	1.3799307
raceOther	-0.4272352	0.2010365	-2.877142	0.7253748	-0.5745467	0.2281736
grade2	0.5309961	0.1831397	16.658587	438.7223333	0.4079614	0.1935342
grade3	0.9060030	0.1917058	17.208447	438.7223392	0.7801358	0.2013946
grade4	1.8504135	0.5421565	17.639998	438.7259010	1.6966710	0.5472605

Table 6: Stepwise Model Coefficients

Predictor	Estimate	Std. Error	Z-Value	P-Value
(Intercept)	-5.5402471	1.2360055	-4.4823805	0.0000074
raceBlack	0.4796129	0.1604322	2.9895047	0.0027943
raceOther	-0.4288428	0.2008699	-2.1349280	0.0327669
marital_statusMarried	-0.2257394	0.1398532	-1.6141167	0.1065021
$marital\_statusSeparated$	0.6844028	0.3807017	1.7977402	0.0722182
marital_statusSingle	-0.0355339	0.1726281	-0.2058410	0.8369152
$marital\_statusWidowed$	0.0321840	0.2181767	0.1475133	0.8827269
x6th_stageIIB	0.5232540	0.1439353	3.6353418	0.0002776
x6th_stageIIIA	0.9870602	0.1411669	6.9921488	0.0000000
x6th_stageIIIB	1.5728667	0.3020784	5.2068165	0.0000002
x6th_stageIIIC	2.0388975	0.1606928	12.6881737	0.0000000
grade2	0.5325726	0.1829927	2.9103483	0.0036103
grade3	0.9105293	0.1915207	4.7542082	0.0000020
grade4	1.8733288	0.5418545	3.4572545	0.0005457
estrogen_statusPositive	-0.7232933	0.1754783	-4.1218401	0.0000376
progesterone_statusPositive	-0.5680958	0.1266501	-4.4855520	0.0000073
rn_examined_log	-0.2982186	0.0800080	-3.7273614	0.0001935
age_log	1.0958051	0.2938018	3.7297427	0.0001917

Table 7: Reweighted Logistic Regression Model Results

Predictor	Estimate	Std_Error	Odds_RatioX9	95CILower. X95.	.CIUpper	. P.Value
(Intercept)	-5.126	1.161	0.006	0.001	0.058	1.02e-
						05
raceBlack	0.455	0.135	1.577	1.211	2.052	0.000713
raceOther	-0.426	0.147	0.653	0.490	0.871	0.003683
$marital\_statusMarried$	-0.248	0.134	0.781	0.600	1.015	0.064877
$marital\_statusSeparated$	0.696	0.350	2.006	1.011	3.981	0.046555

Predictor	Estimate	Std_Error (	Odds_RatioX9	95CILower.	X95CIUpper.	P.Value
marital_statusSingle	0.034	0.162	1.035	0.753	1.422	0.833041
$marital\_statusWidowed$	0.123	0.204	1.131	0.759	1.687	0.544852
$x6th\_stageIIB$	0.533	0.136	1.704	1.305	2.225	9.02e-
						05
$x6th\_stageIIIA$	0.978	0.134	2.660	2.047	3.457	2.52e-
						13
$x6th\_stageIIIB$	1.565	0.287	4.781	2.726	8.386	4.82e-
						08
$x6th\_stageIIIC$	2.006	0.152	7.433	5.519	10.012	< 2e-16
grade2	0.506	0.172	1.659	1.184	2.324	0.003266
grade3	0.862	0.180	2.368	1.664	3.370	1.67e-
						06
grade4	1.825	0.521	6.203	2.233	17.232	0.000463
$estrogen\_statusPositive$	-0.714	0.166	0.490	0.354	0.678	1.70e-
						05
progesterone_statusPositive	e -0.503	0.121	0.605	0.477	0.767	3.37e-
						05
$rn\_examined\_log$	-0.277	0.076	0.758	0.653	0.880	0.000270
age_log	0.974	0.276	2.648	1.543	4.545	0.000410

 $\label{thm:comparison} \mbox{Table 8: Comparison of ROC-AUC Values Across Models} \\ \mbox{and Racial Groups}$ 

Model Type	White	Black	Other	Minority
Original	0.7504	0.7021	0.6584	0.7313
Reweighted	0.7486	0.7052	0.6657	0.7358

## Plots

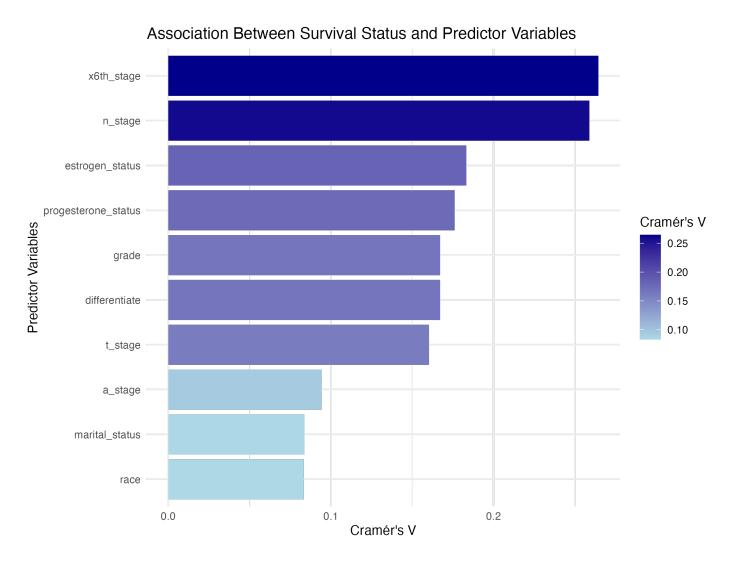


Figure 1: Cramér's V Associations

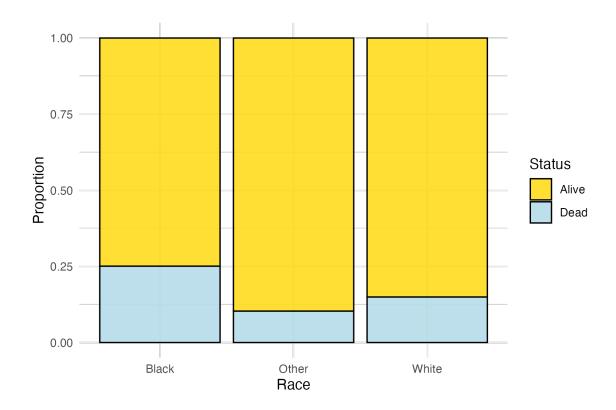


Figure 2: Survival Status by Race

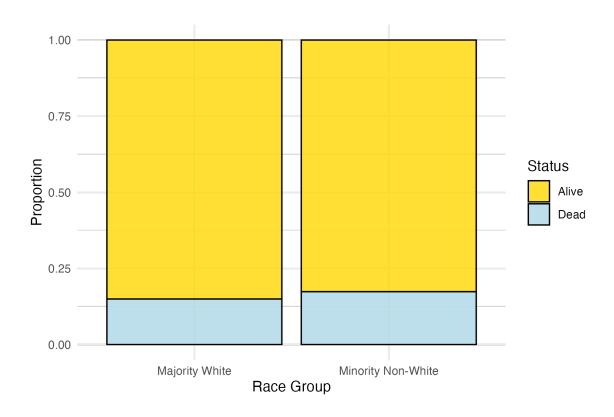


Figure 3: Survival Status by Combined Race Groups

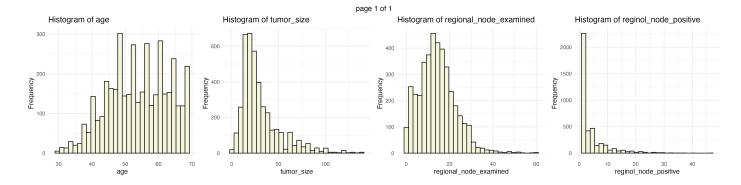


Figure 4: Histograms for Original Continuous Variables

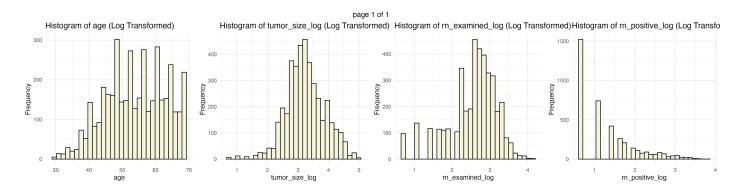


Figure 5: Histograms for Log-Transformed Continuous Variables

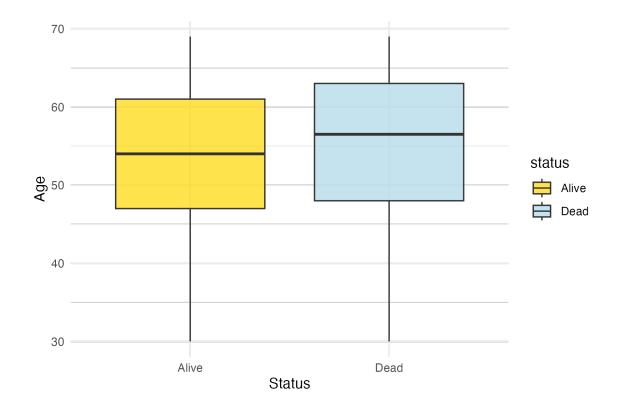


Figure 6: Age by Survival Status

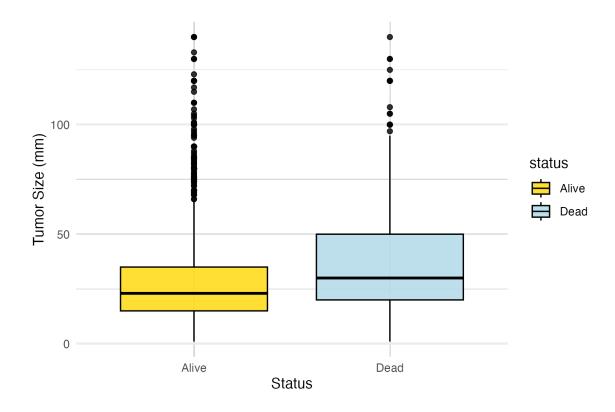


Figure 7: Tumor Size by Survival Status

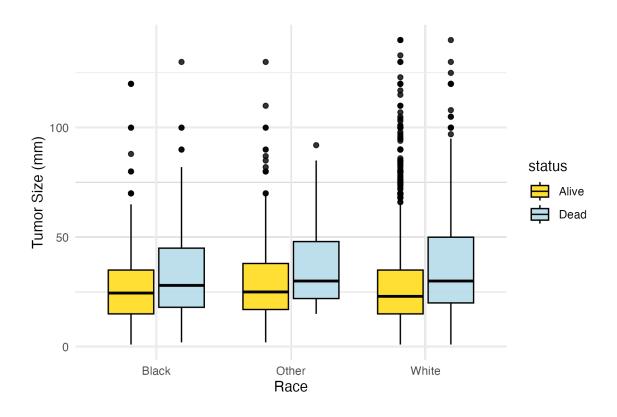


Figure 8: Tumor Size by Race and Survival Status

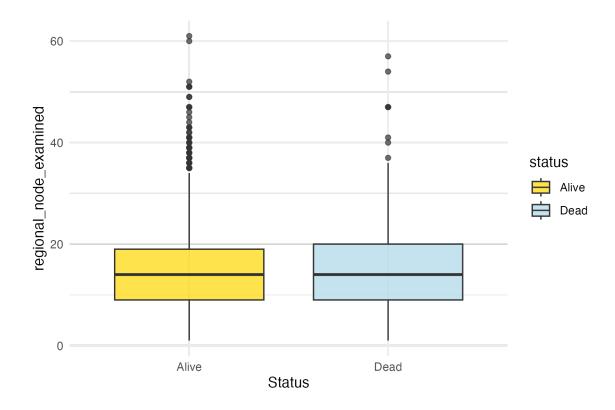


Figure 9: Regional Node Examined by Survival Status

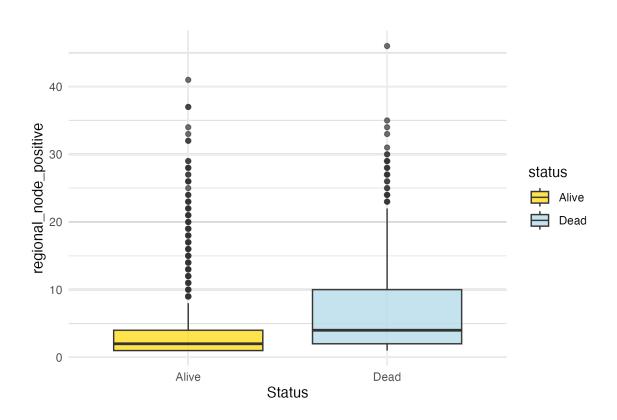


Figure 10: Regional Node Positive by Survival Status

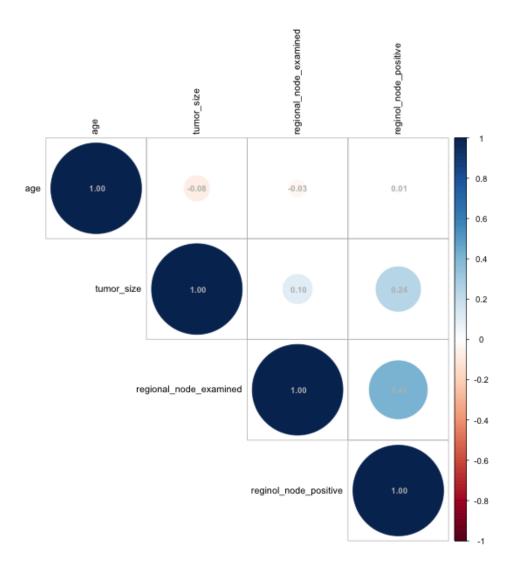


Figure 11: Correlation Matrix for Continuous Variables

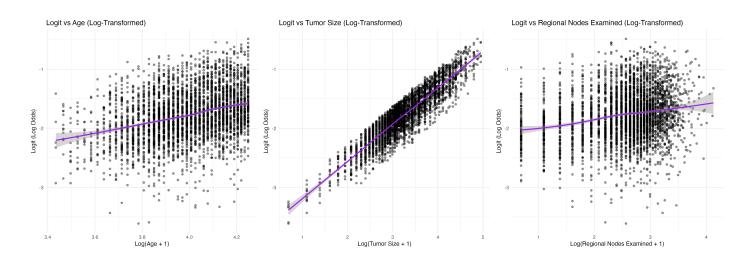


Figure 12: Log Odds Relationship with Predictors

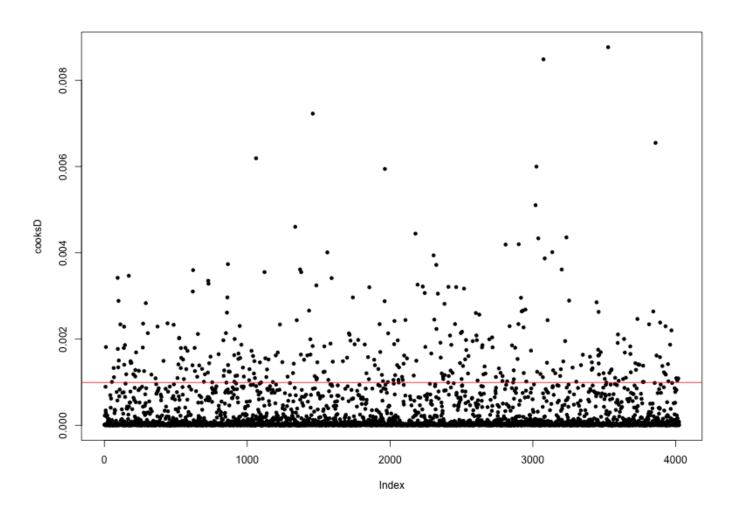


Figure 13: Cook's Distance for Outlier Detection

#### Code Results

For full code results, please refer to the .txt files available in the results folder of the GitHub repository: Yixin-Zheng/p8130\_finalproject.

## Code Appendix

```
knitr::opts_chunk$set(echo = TRUE)
library(tidyverse)
library(janitor)
library(skimr)
library(dplyr)
library(ggplot2)
library(caret)
library(corrplot)
library(lsr)
library(vcd)
library(car)
library(gridExtra)
library(robustbase)
library(leaps)
library(pROC)
library(knitr)
# Import data and clean column names
data <- read.csv("./data/Project_2_data.csv") %>%
 clean_names()
# Select relevant covariates (variables 1-14) and outcome variable
model_data <- data %>%
 dplyr::select(-survival_months)
# Convert categorical variables to factors and relabel `grade`
```

```
model data <- model data %>%
 mutate(
   race = factor(race),
   marital_status = factor(marital_status),
   t_stage = factor(t_stage),
   n_stage = factor(n_stage),
   x6th stage = factor(x6th stage),
   differentiate = factor(differentiate),
   a_stage = factor(a_stage),
    estrogen_status = factor(estrogen_status),
   progesterone_status = factor(progesterone_status),
    status = factor(status, levels = c("Alive", "Dead")),
   grade = case_when(
     grade == "1" ~ "1",
      grade == "2" ~ "2",
      grade == "3" ~ "3",
      grade == " anaplastic; Grade IV" ~ "4",
     TRUE ~ NA character
    ) %>% factor(levels = c("1", "2", "3", "4"))
 )
# Summarize structure of the cleaned dataset
summary(model_data)
# Summary statistics for continuous and categorical variables
skimmed_data <- skim(model_data)</pre>
skim_categorical <- skimmed_data %>%
 filter(skim type == "factor") %>%
 select(-starts_with("numeric"), -skim type) %>%
 na.omit()
write.csv(skim_categorical, "tables/skim_categorical_summary.csv", row.names = FALSE)
```

```
skim numeric <- skimmed data %>%
  filter(skim type == "numeric") %>%
  select(-starts_with("factor"), -skim_type) %>%
 na.omit()
colnames(skim_numeric) <- gsub("^numeric\\.", "", colnames(skim_numeric))</pre>
write.csv(skim_numeric, "tables/skim_numeric_summary.csv", row.names = FALSE)
# Key statistics grouped by survival status
summary_by_status <- model_data %>%
  group_by(status) %>%
  summarise(
    mean_age = mean(age, na.rm = TRUE),
    sd_age = sd(age, na.rm = TRUE),
    mean_tumor_size = mean(tumor_size, na.rm = TRUE),
    sd_tumor_size = sd(tumor_size, na.rm = TRUE),
    prop_white = mean(race == "White", na.rm = TRUE),
    prop black other = mean(race != "White", na.rm = TRUE),
    n obs = n()
  )
write.csv(summary_by_status, "tables/summary_by_status.csv", row.names = FALSE)
# Define categorical variables to analyze
variables <- c("race", "marital_status", "t_stage", "n_stage", "x6th_stage",</pre>
               "differentiate", "grade", "a_stage", "estrogen_status", "progesterone_status
# Initialize a vector to store Cramér's V results
results <- numeric(length(variables))</pre>
# Calculate Cramér's V for each variable
for (i in seq_along(variables)) {
  var <- variables[i]</pre>
```

```
# Select outcome and predictor variable, omitting missing values
  df_temp <- model_data %>%
    dplyr::select(status, all_of(var)) %>%
    na.omit()
  # Convert both columns to factors
  x <- droplevels(as.factor(df temp$status))</pre>
  y <- droplevels(as.factor(df temp[[var]]))</pre>
  # Create contingency table and calculate Cramér's V
  table_var <- table(x, y)</pre>
  results[i] <- cramersV(table var)</pre>
}
# Create a dataframe with results
association_df <- data.frame(Variable = variables, CramersV = results)</pre>
# Plot Cramér's V values
cramerV_association <- ggplot(association_df, aes(x = reorder(Variable, CramersV), y = CramersV), y = CramersV
  geom_bar(stat = "identity") +
  coord_flip() +
  scale_fill_gradient(low = "lightblue", high = "darkblue") +
  labs(
    title = "Association Between Survival Status and Predictor Variables",
    x = "Predictor Variables",
    y = "Cramér's V",
    fill = "Cramér's V"
  ) +
  theme_minimal()
ggsave("plots/cramerV_association.png", plot = cramerV_association, width = 8, height = 6)
# Proportional Bar Plot for Survival Status by Race
```

```
race barplot <- ggplot(model data, aes(x = race, fill = status)) +</pre>
 geom_bar(position = "fill", alpha = 0.8, color = "black") +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Race",
   y = "Proportion",
   fill = "Status"
 )
ggsave("plots/race_proportional_barplot.png", plot = race_barplot, width = 6, height = 4)
# Combine "Black" and "Other" into a single group "Minority Non-White"
model_data_race_combined <- model_data %>%
 mutate(
   race combined = case_when(
     race == "White" ~ "Majority White",
     race %in% c("Black", "Other") ~ "Minority Non-White"
   ),
   race_combined = factor(race_combined, levels = c("Majority White", "Minority Non-White"
 )
# Proportional Bar Plot for Combined Race Groups
race_combined_barplot <- ggplot(model_data_race_combined, aes(x = race_combined, fill = sta
 geom_bar(position = "fill", alpha = 0.8, color = "black") +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme minimal() +
 labs(
   x = "Race Group",
   y = "Proportion",
   fill = "Status"
```

```
ggsave("plots/race_combined_proportional_barplot.png", plot = race_combined_barplot, width
continuous_vars <- model_data %>%
  dplyr::select(age, tumor_size, regional_node_examined, reginol_node_positive) %>%
  na.omit()
df1 <- as.data.frame(continuous vars)</pre>
hist_list <- lapply(names(continuous_vars), function(col) {</pre>
  ggplot(continuous_vars, aes_string(x = col)) +
    geom_histogram(fill = "beige", color = "black", bins = 30) +
    labs(
      title = paste("Histogram of", col),
      x = col,
      y = "Frequency"
    ) +
    theme_minimal()
})
# Arrange plots in a grid
hist_grid <- marrangeGrob(hist_list, nrow = 1, ncol = 4)</pre>
ggsave("plots/original_histograms_grid.png", hist_grid, width = 16, height = 4)
df log <- df1 %>%
  dplyr::select(-tumor_size, -regional_node_examined, -reginol_node_positive) %>%
  mutate(
    tumor_size_log = log(df1$tumor_size + 1),
    rn examined \log = \log(df1\$regional node examined + 1),
    rn positive log = log(df1$reginol node positive + 1)
  )
log_hist_list <- lapply(names(df_log), function(col) {</pre>
```

```
ggplot(df_log, aes_string(x = col)) +
    geom_histogram(fill = "beige", color = "black", bins = 30) +
    labs(
      title = paste("Histogram of", col, "(Log Transformed)"),
     x = col,
     y = "Frequency"
    ) +
    theme_minimal()
})
# Arrange plots in a grid
log_hist_grid <- marrangeGrob(log_hist_list, nrow = 1, ncol = 4)</pre>
ggsave("plots/log_transformed_histograms_grid.png", log_hist_grid, width = 16, height = 4)
# Age by survival status
age_boxplot <- ggplot(model_data, aes(x = status, y = age, fill = status)) +</pre>
  geom_boxplot(alpha = 0.7) +
  scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
  theme_minimal() +
  labs(
   x = "Status",
    y = "Age"
  )
ggsave("plots/age_by_status_boxplot.png", plot = age_boxplot, width = 6, height = 4)
# Tumor size by survival status
tumor_boxplot <- ggplot(model_data, aes(x = status, y = tumor_size, fill = status)) +</pre>
  geom_boxplot(alpha = 0.8, color = "black") +
  scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
  theme_minimal() +
  labs(
    x = "Status",
    y = "Tumor Size (mm)"
```

```
ggsave("plots/tumor_size_by_status_boxplot.png", plot = tumor_boxplot, width = 6, height =
# Tumor size by race and survival status
tumor_race_boxplot <- ggplot(model_data, aes(x = race, y = tumor_size, fill = status)) +</pre>
 geom_boxplot(alpha = 0.8, color = "black") +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Race",
   y = "Tumor Size (mm)"
 )
ggsave("plots/tumor_size_by_race_status_boxplot.png", plot = tumor_race_boxplot, width = 6,
# Regional Node Examined by survival status
rn_examined_boxplot <- ggplot(model_data, aes(x = status, y = regional_node_examined, fill</pre>
 geom_boxplot(alpha = 0.7) +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Status",
   y = "regional_node_examined"
 )
ggsave("plots/rn_examined_by_status_boxplot.png", plot = rn_examined_boxplot, width = 6, he
# Reginol Node Positive by survival status
rn_positive_boxplot <- ggplot(model_data, aes(x = status, y = reginol_node_positive, fill =</pre>
 geom_boxplot(alpha = 0.7) +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
```

```
labs(
    x = "Status",
    y = "regional_node_positive"
  )
ggsave("plots/rn_positive_by_status_boxplot.png", plot = rn_positive_boxplot, width = 6, he
correlation_matrix <- cor(continuous_vars, use = "pairwise.complete.obs")</pre>
# Pairwise relationships (correlation matrix for continuous variables)
correlation_plot <- function() {</pre>
  corrplot(
    correlation_matrix,
    method = "circle",
   type = "upper",
    tl.col = "black",
    addCoef.col = "grey",
    number.cex = 0.8,
    tl.cex = 0.9
  )
}
png("plots/correlation_matrix_plot.png", width = 800, height = 600)
correlation_plot()
dev.off()
# Binary or Dichotomous Response Variable
unique(model_data$status)
#The response variable (status) has exactly two categories: Alive and Dead.
# Fit an initial logistic regression model
model_data_1 <- model_data %>%
 mutate(
    race = relevel(race, ref = "White"), # Set "White" as reference
```

```
grade = relevel(grade, ref = "1"),  # Set "Grade 1" as reference
    x6th stage = relevel(x6th stage, ref = "IIA") # Set "IIA" as reference
  )
alias_results <- capture.output(alias(glm(status ~ ., data = model_data_1, family = binomia
# Save the results to a text file in the `results` folder
writeLines(alias_results, "results/alias_results_model1.txt")
model_data_2 <- model_data_1 %>%
  dplyr::select(-differentiate, -n_stage, -t_stage, -reginol_node_positive)
# Perform multicollinearity check with alias()
alias_results_2 <- capture.output(alias(glm(status ~ ., data = model_data_2, family = binor
writeLines(alias_results_2, "results/alias_results_model2.txt")
# Fit a logistic regression model
model vif <- glm(status ~ ., data = model data 2, family = binomial)</pre>
# Calculate VIF
vif_values <- vif(model_vif)</pre>
vif_df <- as.data.frame(vif_values)</pre>
vif_df <- tibble::rownames_to_column(vif_df, var = "Variable")</pre>
colnames(vif_df) <- c("Variable", "GVIF", "Df", "GVIF_Ratio")</pre>
write.csv(vif_df, "tables/vif.csv", row.names = FALSE)
continuous_vars_log_odds <- model_data_2 %>%
  dplyr::select(age, tumor_size, regional_node_examined, status) %>% # Include 'status'
  na.omit()
# Log-transform tumor size and regional nodes examined
df_log_odds <- continuous_vars_log_odds %>%
  mutate(
```

```
age_log = log(age + 1),
    tumor size log = log(tumor size + 1),
    rn_examined_log = log(regional_node_examined + 1)
  )
linearity_test <- glm(status ~ age_log + tumor_size_log + rn_examined_log,</pre>
                       data = df_log_odds,
                       family = binomial)
df_log_odds$logit <- predict(linearity_test, type = "link")</pre>
plot1 <- ggplot(df_log_odds, aes(x = age_log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Age (Log-Transformed)",
       x = \text{"Log(Age + 1)"}, y = \text{"Logit (Log Odds)"}) +
  theme_minimal()
plot2 <- ggplot(df_log_odds, aes(x = tumor_size_log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Tumor Size (Log-Transformed)", x = "Log(Tumor Size + 1)", y = "Log
  theme_minimal()
plot3 <- ggplot(df_log_odds, aes(x = rn_examined_log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Regional Nodes Examined (Log-Transformed)",
       x = "Log(Regional Nodes Examined + 1)", y = "Logit (Log Odds)") +
  theme_minimal()
```

```
log odds grid <- grid.arrange(plot1, plot2, plot3, ncol = 3)</pre>
ggsave("plots/logit_grid_plot.png", plot = log_odds_grid, width = 18, height = 6)
# Update
model_data_3 <- model_data_2 %>%
 mutate(
    tumor_size_log = log(tumor_size + 1),
    rn_examined_log = log(regional_node_examined + 1),
    age_log = log(age + 1)
  ) %>%
  dplyr::select(-tumor_size, -regional_node_examined, -age)
# Fit logistic regression model with log-transformed predictors
full_model <- glm(status ~ .,</pre>
                 data = model_data_3, family = binomial)
# Calculate Cook's Distance
cooksD <- cooks.distance(model vif)</pre>
# Plot Cook's Distance
png("plots/cooks_distance_plot.png", width = 800, height = 600)
plot(cooksD, pch = 20)
abline(h = 4 / nrow(model_data_3), col = "red")
dev.off()
# Identify influential observations
influential_obs <- which(cooksD > 4 / nrow(model_data_3))
# Build Model
model_no_outliers <- glm(status ~ .,</pre>
                          data = model_data_3[-influential_obs, ], family = binomial)
model_robust <- glmrob(status ~ .,</pre>
```

```
data = model data 3, family = binomial, method = "Mqle")
# Extract summaries
full_coefficients <- summary(full_model)$coefficients</pre>
no_outliers_coefficients <- summary(model_no_outliers)$coefficients</pre>
robust_coefficients <- summary(model_robust)$coefficients</pre>
# Identify unstable coefficients
unstable coeffs <- which(
  (abs(full_coefficients[, "Estimate"] - no_outliers_coefficients[, "Estimate"]) > 2) |
  (abs(full_coefficients[, "Std. Error"] - no_outliers_coefficients[, "Std. Error"]) > 2)
)
unstable_coeffs_df <- data.frame(
  Variable = rownames(full_coefficients)[unstable_coeffs],
  Full_Coef = full_coefficients[unstable_coeffs, "Estimate"],
  Full_SE = full_coefficients[unstable_coeffs, "Std. Error"],
  No_Outliers_Coef = no_outliers_coefficients[unstable_coeffs, "Estimate"],
  No_Outliers_SE = no_outliers_coefficients[unstable_coeffs, "Std. Error"],
  Robust_Coef = robust_coefficients[rownames(full_coefficients)[unstable_coeffs], "Estimate
  Robust_SE = robust_coefficients[rownames(full_coefficients)[unstable_coeffs], "Std. Error
write.csv(unstable_coeffs_df, "tables/unstable_coefficients.csv", row.names = FALSE)
# Extract robustness weights
robust_summary <- capture.output(summary(model_robust))</pre>
start_line <- grep("Robustness weights w.r \\* w.x:", robust_summary)
end_line <- start_line + 5</pre>
desired section <- robust summary[start line:end line]</pre>
cat(paste(desired section, collapse = "\n"))
# Forward selection
forward_model <- step(glm(status ~ ., data = model_data_3, family = binomial), scope = list
```

```
# Backward elimination
backward_model <- step(glm(status ~ ., data = model_data_3, family = binomial), direction =
# Stepwise selection
stepwise_model <- step(glm(status ~ ., data = model_data_3, family = binomial), scope = lis</pre>
writeLines(capture.output(summary(full model)), "results/full model summary.txt")
writeLines(capture.output(summary(forward_model)), "results/forward_model_summary.txt")
writeLines(capture.output(summary(backward_model)), "results/backward_model_summary.txt")
writeLines(capture.output(summary(stepwise_model)), "results/stepwise_model_summary.txt")
# Define predictors
predictors <- c("race", "marital status", "x6th stage", "grade",</pre>
                "estrogen status", "progesterone status", "rn examined log", "age log")
# Create a dataframe to store results
interaction results <- data.frame(</pre>
  Predictor1 = character(),
  Predictor2 = character(),
  P Value = numeric(),
  stringsAsFactors = FALSE
)
# Loop through each pair of predictors
for (i in seq_along(predictors)) {
  for (j in seq_along(predictors)) {
    if (i < j) {
      predictor1 <- predictors[i]</pre>
      predictor2 <- predictors[j]</pre>
      # Fit interaction model
      formula <- as.formula(paste("status ~", paste(predictors, collapse = " + "),</pre>
                                   "+", predictor1, "*", predictor2))
```

```
interaction model <- glm(formula, family = binomial, data = model data 3)
      # Extract interaction term
      interaction_term <- paste(predictor1, predictor2, sep = ":")</pre>
      coef_names <- names(coef(interaction_model))</pre>
      if (interaction_term %in% coef_names) {
        # Extract p-value for the interaction term
        p value <- coef(summary(interaction model))[interaction term, "Pr(>|z|)"]
        # Append to results if p-value < 0.05
        if (p_value < 0.05) {</pre>
          interaction_results <- rbind(interaction_results,</pre>
                                         data.frame(Predictor1 = predictor1,
                                                     Predictor2 = predictor2,
                                                     P_Value = p_value,
                                                     stringsAsFactors = FALSE))
        }
      }
    }
  }
}
# Sort results by p-value
interaction_results <- interaction_results[order(interaction_results$P_Value), ]</pre>
print(interaction_results)
stepwise_summary <- summary(stepwise_model)</pre>
stepwise coefficients <- as.data.frame(stepwise summary$coefficients)</pre>
colnames(stepwise coefficients) <- c("Estimate", "Std Error", "Z Value", "P Value")</pre>
stepwise coefficients <- tibble::rownames_to_column(stepwise coefficients, "Predictor")</pre>
write.csv(stepwise_coefficients, "tables/stepwise_coefficients.csv", row.names = FALSE)
```

```
set.seed(123)
control <- trainControl(method = "cv", number = 10, classProbs = TRUE, summaryFunction = tw
cv_model <- train(</pre>
  status ~ race + marital_status + x6th_stage + grade + estrogen_status +
    progesterone_status + rn_examined_log + age_log,
  data = model data 3,
  method = "glm",
  family = "binomial",
  trControl = control,
 metric = "ROC"
)
performance_summary <- capture.output(cv_model)</pre>
writeLines(performance_summary, "results/cv_model_performance.txt")
# Split data by race group
white data <- model data 3 %>% filter(race == "White")
minority data <- model data 3 %>% filter(race != "White")
black data <- model data 3 %>% filter(race == "Black")
other data <- model data 3 %>% filter(race == "Other")
# Predict probabilities for White group
pred_white <- predict(cv_model, newdata = white_data, type = "prob")[, "Dead"]</pre>
roc_white <- roc(white_data$status, pred_white)</pre>
auc_white <- auc(roc_white)</pre>
# Predict probabilities for Minority group
pred_minority <- predict(cv_model, newdata = minority_data, type = "prob")[, "Dead"]</pre>
roc_minority <- roc(minority_data$status, pred_minority)</pre>
auc minority <- auc(roc minority)</pre>
# Predict probabilities for Black group
```

```
pred black <- predict(cv model, newdata = black data, type = "prob")[, "Dead"]</pre>
roc black <- roc(black data$status, pred black)</pre>
auc_black <- auc(roc_black)</pre>
# Predict probabilities for Other group
pred_other <- predict(cv_model, newdata = other_data, type = "prob")[, "Dead"]</pre>
roc_other <- roc(other_data$status, pred_other)</pre>
auc_other <- auc(roc_other)</pre>
model_data_4 <- model_data_3 %>%
  mutate(weight = case_when(
   race == "White" ~ 1,
    race == "Black" ~ 1.5,
    race == "Other" ~ 2
  ))
minority_data <- model_data_4 %>% filter(race != "White")
# Refit the model with weights
reweighted model <- train(
  status ~ race + marital_status + x6th_stage + grade + estrogen_status +
    progesterone_status + rn_examined_log + age_log,
  data = model_data_4,
  method = "glm",
  family = "binomial",
  trControl = control,
  weights = weight
)
# Evaluate performance on subgroups
pred_white_weighted <- predict(reweighted_model, newdata = white_data, type = "prob")[, "De</pre>
roc_white_weighted <- roc(white_data$status, pred_white_weighted)</pre>
auc_white_weighted <- auc(roc_white_weighted)</pre>
```

```
pred_minority_weighted <- predict(reweighted_model, newdata = minority_data, type = "prob")</pre>
roc minority weighted <- roc(minority data$status, pred minority weighted)
auc_minority_weighted <- auc(roc_minority_weighted)</pre>
pred_black_weighted <- predict(reweighted_model, newdata = black_data, type = "prob")[, "De</pre>
roc_black_weighted <- roc(black_data$status, pred_black_weighted)</pre>
auc_black_weighted <- auc(roc_black_weighted)</pre>
pred_other_weighted <- predict(reweighted_model, newdata = other_data, type = "prob")[, "De</pre>
roc other weighted <- roc(other data$status, pred other weighted)
auc other weighted <- auc(roc other weighted)</pre>
auc_comparison_summary <- data.frame(</pre>
  Model = factor(c("Original", "Reweighted"), levels = c("Original", "Reweighted")),
  White = c(auc_white, auc_white_weighted),
  Black = c(auc_black, auc_black_weighted),
  Other = c(auc other, auc other weighted),
  Minority = c(auc_minority, auc_minority_weighted)
)
auc comparison summary[, 2:5] <- round(auc comparison summary[, 2:5], 4)
write.csv(auc_comparison_summary, "tables/auc_comparison_summary.csv", row.names = FALSE)
# Extract coefficients and confidence intervals from the reweighted model
summary_reweighted <- summary(reweighted_model$finalModel)</pre>
coefficients <- coef(summary reweighted)</pre>
# Calculate Odds Ratios and 95% Confidence Intervals
odds ratios <- exp(coefficients[, "Estimate"])</pre>
lower_ci <- exp(coefficients[, "Estimate"] - 1.96 * coefficients[, "Std. Error"])</pre>
upper ci <- exp(coefficients[, "Estimate"] + 1.96 * coefficients[, "Std. Error"])
```

```
# Combine into a table
results table <- data.frame(
  Predictor = rownames(coefficients),
  Estimate = round(coefficients[, "Estimate"], 3),
  Std_Error = round(coefficients[, "Std. Error"], 3),
  Odds_Ratio = round(odds_ratios, 3),
  `95% CI (Lower)` = round(lower_ci, 3),
  `95% CI (Upper)` = round(upper_ci, 3),
  `P-Value` = format.pval(coefficients[, "Pr(>|z|)"], digits = 3)
)
# Clean up row names
rownames(results_table) <- NULL</pre>
write.csv(results_table, "tables/reweighted_model_results.csv", row.names = FALSE)
skim categorical <- read.csv("tables/skim categorical summary.csv")</pre>
knitr::kable(skim_categorical, caption = "Skim Summary for Categorical Variables")
skim_numeric <- read.csv("tables/skim_numeric_summary.csv")</pre>
knitr::kable(skim numeric, caption = "Skim Summary for Numeric Variables")
summary_by_status <- read.csv("tables/summary_by_status.csv")</pre>
knitr::kable(summary_by_status, caption = "Summary Statistics Grouped by Survival Status")
vif_table <- read.csv("tables/vif.csv")</pre>
knitr::kable(vif_table, caption = "Variance Inflation Factors for Predictors")
unstable table <- read.csv("tables/unstable coefficients.csv")</pre>
knitr::kable(unstable_table, caption = "Unstable Coefficients")
stepwise_coefficients <- read.csv("tables/stepwise_coefficients.csv")</pre>
```

```
knitr::kable(stepwise_coefficients, caption = "Stepwise Model Coefficients", col.names = c(
reweighted_model_results <- read.csv("tables/reweighted_model_results.csv")
knitr::kable(reweighted_model_results, caption = "Reweighted Logistic Regression Model Results)
auc_table <- read.csv("tables/auc_comparison_summary.csv")
knitr::kable(auc_table, caption = "Comparison of ROC-AUC Values Across Models and Racial Gr</pre>
```