Final Project: Prediction of survival rate for breast cancer patients

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Abstract

Breast cancer is a leading cause of cancer-related mortality among women worldwide. This study develops a predictive model for survival outcomes in breast cancer patients using logistic regression, leveraging demographic, clinical, and pathological factors from a prospective cohort dataset. The analysis focuses on identifying key predictors of mortality, evaluating model performance across racial groups, and addressing fairness in prediction accuracy. Tumor stage, grade, and hormone receptor status emerged as significant predictors. The initial logistic regression model achieved a moderate performance, with an area under the receiver operating characteristic curve (ROC-AUC) of 0.74. However, disparities in model performance were observed between racial groups, prompting the implementation of reweighting strategies to enhance fairness. These findings highlight the importance of equitable modeling approaches to improve prognostic accuracy and clinical outcomes in breast cancer care.

Introduction

Breast cancer has a high prevalence with average X new cases each year worldwide. Approximately 13.1 % of female are diagnosed with breast cancer at some point in their lifetime. This type of cancer accounts for 23% of total cancer cases and 14% of cancer deaths¹. Recent years with the advances in understanding of disease diagnosis and the effective tailored treatments, the mortality rate has decreased². According to research, survival of breast cancer depends on various factors including the tumor size, the grade of tumor³, cancer stage, lymph node stages, and socioeconomic and race⁴. In several published models with data from various regions, the most important predictors of survival rates are the prognostic factors of tumor sizes and grade of tumor and, age at diagnosis 5. Patients at younger age when they were diagnosed with breast cancer at the same stage are more likely to

live longer compared to older patients. Some data also showed that racial and socioeconomic status can explain the survival rate variation among the groups with the same prognostic indicators. For example, lower-income patients often have late diagnosis and the African American population also has a lower survival rate. However, these models yielded different suboptimal results based on the available data source. This report examines data from a prospective cohort of breast cancer patients, which includes variables including demographic, clinical, and pathological factors. The primary aims are to predict mortality risk, identify significant predictors, and assess the fairness and performance of predictive models across racial groups. By achieving these objectives, this analysis seeks to offer meaningful insights into improving breast cancer outcomes and addressing healthcare disparities.

Methods

Data Description The dataset includes 10 categorical variables: race (Black, White, Other), marital status (Divorced, Married, Separated, Single, Widowed), tumor stage (T1, T2, T3, T4), lymph node stage (N1, N2, N3), adjusted AJCC 6th stage (IIA, IIB, IIIA, IIIB, IIIC), tumor differentiation (Well, Moderately, Poorly, Undifferentiated), grade (1–4), tumor spread stage (Regional, Distant), estrogen receptor status (Positive, Negative), and progesterone receptor status (Positive, Negative). Additionally, there are 4 continuous variables: age, tumor size, regional nodes examined, and regional nodes positive.

Data Cleaning Column headers were renamed, categorical variables were converted to factors. tumor grade levels were recoded to ensure interpretability. Missing values were assessed, and log transformations were applied to highly skewed continuous variables (tumor size, regional nodes examined, and regional nodes positive) to normalize their distributions.

EDA Methods skim() function is applied to the dataset (model_data) to compute detailed summary statistics for all variables. (table.1 & 2), Group-wise key statistics are calculated based on survival status(tabel.3) Cramér's V was used to quantify the strength of the association between categorical variables and the binary outcome (Alive/Dead), with values ranging from 0 (no association) to 1 (perfect association). Distributional plots, including proportional bar plots for race groups and histograms for continuous variables, were created to visualize the data. Boxplots stratified by survival status were used to explore relationships between continuous variables and the binary outcome. A correlation matrix for continuous variables was generated to assess pairwise relationship between variables.

Modeling Assumptions and Transformations Logistic regression was chosen as the primary

method due to the binary nature of the outcome. Assumptions checked are: 1. The response variable (status) was confirmed to be binary by code.

2. The alias() function identified collinearity between models: grade2, grade3, and grade4 with other predictors, x6th_stageIIIC with n_stage, and differentiate with grade. For simplification, some variables were removed: x6th_stage captures n_stage's information, n_stage was dropped. t_stage (linked to tumor size), differentiate (overlapping with grade), and regional_node_positive (redundant with tumor size and regional_node_examined) were removed.

VIF were calculated, to ensure that all values were below 5 (table.4), indicating no multicollinearity. (dataset updated with dropped variable)

- 3. Continuous predictors were log-transformed, and their relationships with the log odds were examined (fig.12). This confirmed linearity. (dataset updated with transformed variable)
- 4. Independence of Errors: Since there were no group-level structures, the independence assumption was satisfied.
- 5. Outliers: Cook's Distance identified potential outliers exceeding 4/n, which flagged numerous points as influential, likely reflecting population variability rather than errors (fig.13). Models with and without these points were compared. Removing these points destabilized coefficients like grade, making them unreliable. Robust logistic regression (model_robust) mitigated outliers impact, providing stable estimates for predictors (table.5). About 12% of data had reduced influence, while most observations remained unaffected. Despite its advantages, we opted for the original logistic regression for simplicity and familiarity.

Model Construction and Selection Models were constructed using predictors identified during EDA and assumption checks. Forward, backward, and stepwise selection were applied based on the AIC to select the final model. Best subset selection, based on Adjusted R^2 , Cp, BIC, was also performed. Interaction effects were tested by examining pairwise interactions between predictors. The final model was chosen using stepwise selection to balance interpretability and performance.

Model Validation and Fairness The model was validated using 10-fold cross-validation, evaluating ROC-AUC, sensitivity, and specificity. Fairness was assessed by evaluating model performance across racial subgroups (White, Black, Other) based on subgroup-specific ROC-AUC values. To address disparities, fairness reweighting was applied by adjusting model weights to prioritize underrepresented groups. The reweighted model's performance was compared with the original, highlighting improvements in fairness while maintaining predictive accuracy. Predictor coefficients were interpreted as odds

ratios, quantifying their impact on survival outcomes

Results

Exploratory Data Analysis

The plot shows the strength of association (Cramér's V) between **survival status** and each categorical predictor:

- 1. **x6th_stage** and **n_stage** have the **highest Cramér's V values** (around 0.25), indicating they are the most informative predictors of survival status in the dataset. These variables reflect tumor stage and lymph node involvement, which are critical factors in breast cancer prognosis.
- 2. Estrogen_status, progesterone_status, and grade exhibit moderate associations (Cramér's V around 0.15–0.2). These variables provide meaningful information about hormone status and differentiation, making them important contributors to survival prediction.
- 3. **Differentiate** and **t_stage** show moderate but slightly lower associations compared to the top variables, suggesting their relevance to survival status.
- 4. a_stage, marital_status, and race have lower Cramér's V values (less than 0.1), indicating weaker associations with survival status. Although these variables contribute limited information individually, they may still be useful in interaction terms or when combined with other predictors.

By focusing on variables with higher Cramér's V values, we can build more efficient and predictive models for survival status analysis.

Both plots (Proportional Bar Plot for Survival Status by Race, or combined race group) confirm a disparity in survival outcomes across racial groups. Non-White patients, particularly Black patients, show a higher likelihood of death. Effect of Combining Groups: Combining Black and Other into Minority Non-White simplifies the comparison and reduce the racial disparities between Majority White and Minority Non-White groups.

Histograms were created for each continuous variable to visually examine their distributions and identify potential skewness. The results revealed that, except for age, the other three variables exhibited significant right skewness, indicating the need for transformation if we want to perform linear regression. Though we are not doing linear regression, transformation has still been performed here, we

see that after performing log-transformation, there is a improvement on the skewness of tumor_size, regional_node_examined and reginol_node_positive.

Pairwise Relationships and Interactions: A correlation matrix was generated to examine the relationships between the variables. Small circles (near-zero correlations) are observed for age and all other variables; tumor_size and regional_node_examineds suggesting weak or no relationships. regional_node_positive is moderately associated with both tumor size (0.24) and regional nodes examined (0.41), which might influence modeling decisions. There are no strong correlations (close to +-1), suggesting no immediate multicollinearity issues among these variables.

Feature Selection

full results are attached in the end Full Model: $glm(formula = status \sim ., family = binomial, data = model_data_3)$ AIC: 3039.8

Forward Model glm(formula = status ~ race + marital_status + x6th_stage + grade + a_stage + estrogen_status + progesterone_status + tumor_size_log + rn_examined_log + age_log, family = binomial, data = model_data_3) AIC: 3039.8

Backward Model: $glm(formula = status \sim race + marital_status + x6th_stage + grade + estrogen_status + progesterone_status + rn_examined_log + age_log, family = binomial, data = model_data_3) AIC:3037.5$

Stepwise Model: $glm(formula = status \sim race + marital_status + x6th_stage + grade + estrogen_status + progesterone_status + rn_examined_log + age_log, family = binomial, data = model_data_3) AIC:3037.5$

Best Model by Adjusted R2: 10 variables, raceBlack x6th_stageIIIA x6th_stageIIIB x6th_s

Best Model by Cp: 10 variables, raceBlack x6th_stageIIIA x6th_stageIIIB x6th_stageIIIC

Best Model by BIC: 10 variables, raceBlack x6th_stageIIIA x6th_stageIIIB x6th_stageIIIC

[1] Predictor1 Predictor2 P_Value

<0 rows> (or 0-length row.names)

Since resulting dataframe is empty, it suggests that none of the interaction terms tested in our dataset have p-values less than 0.05. This outcome aligns with our earlier exploratory analyses (Cramér's V,

correlation matrix, and model diagnostics), which indicated that interaction effects are likely weak or non-existent in your data.

Final Model Since Best subset selection evaluates all possible combinations of predictors (including all levels of factor variables) to find the model with the best fit according to criteria like adjusted R2, Cp and BIC, each level of a factor variable (e.g., x6th_stageIIIA) can be treated as an independent binary variable (dummy variable) in this process. Thus we will not use the best subset selection, as we need consider the factor as a whole for interpretability.

The final model was chosen through forward, backward, and stepwise selection methods using AIC as the selection criterion. The model balances interpretability and predictive performance, including only significant predictors that contribute to understanding survival outcomes:

```
glm(formula = status ~ race + marital_status + x6th_stage + grade + estrogen_status + progesterone_status + rn_examined_log + age_log, family = binomial, data = model_data_3)

In logistic regression, coefficients represent the change in the log-odds of the outcome (status = Dead)

for a one-unit increase in a predictor, holding other variables constant. Odds ratios provide a more
```

The key coefficients included are 1. Race: - raceBlack: Positive coefficient implies higher odds of death for Black patients compared to the reference group (White patients). Odds ratio > 1 indicates increased risk for Black patients. - raceOther: Negative coefficient indicates slightly reduced odds of death for patients classified as "Other" compared to White patients.

interpretable measure.

- 2. Marital Status: The marital status categories have weak effects (some not statistically significant). For instance, being married or widowed appears to have no strong association with survival compared to the reference group.
- 3. Adjusted AJCC 6th stage (x6th_stage): is the strongest predictor of death risk. Higher stages (IIIA, IIIB, IIIC) significantly increase the odds of death compared to stage IIA (reference group). Patients in stage IIIC have the highest odds of death, indicating advanced tumor progression is a critical predictor of survival.
- 4. Tumor Grade (grade): Grades 3 and 4 significantly increase the odds of death compared to Grade 1, indicating poorly differentiated tumors are associated with worse outcomes.

5. Hormone Receptor Status: estrogen_statusPositive and progesterone_statusPositive:

Negative coefficients suggest reduced odds of death for patients with positive estrogen or

progesterone receptor status. This reflects the improved prognosis often associated with
hormone-sensitive tumors.

6. Log-transformed Predictors: -rn_examined_log: Negative coefficient indicates increased survival odds with more regional lymph nodes examined. This might reflect more aggressive or effective treatment strategies. -age_log: Positive coefficient implies older patients are at higher risk of death.

So x6th_stage, grade, estrogen_statusPositive and progesterone_statusPositive, age_log and rn examined log are significant predictors affecting the risk.

Model Performance

full results are attached in the end We did Cross-Validation (10 fold) ROC-AUC: 0.7400, the overall performance of the model is moderately good, with an acceptable ability to distinguish between "Alive" and "Dead" outcomes. Sensitivity: 0.985, the model is highly sensitive, meaning it correctly identifies most of the "Dead" cases. Specificity: 0.122, the model has very low specificity, indicating difficulty in correctly identifying "Alive" cases.

```
## ROC-AUC for White group: 0.7503654
```

ROC-AUC for Black group: 0.7021491

ROC-AUC for Other group: 0.658431

ROC-AUC for Minority group: 0.7313469

Performance by Race Groups White Group (Majority): - ROC-AUC: 0.7504 - The model performs better for the majority race group, achieving the highest predictive ability among the subgroups.

Black Group (Minority): - ROC-AUC: 0.7021 - The performance for Black patients is lower than for White patients, indicating potential disparities in prediction accuracy.

Other Group (Minority): - ROC-AUC: 0.6584 - The lowest ROC-AUC is for the "Other" race group, suggesting the model struggles most with this subgroup.

Combined Minority Group (Black + Other): - ROC-AUC: 0.7313 - When grouped together, the model's performance for minorities improves but remains slightly lower than for the White group.

This disparity indicate a potential fairness issue, which need us to reduce the performance gap.

```
## Reweighted Model - ROC-AUC for White group: 0.7486181

## Reweighted Model - ROC-AUC for Minority group: 0.7358304

## Reweighted Model - ROC-AUC for Black group: 0.7051653

## Reweighted Model - ROC-AUC for Other group: 0.6657164
```

The reweighted model shows improved fairness compared to the original combined model: Reweighting has reduced the performance gap between the White and minority groups. The ROC-AUC for the minority group (0.7358) is now closer to that of the majority group (0.7486).

Conclusion

In this report, we derived a model to predict the survival rate of breast cancer patients using a logistic model. The most important factors are Adjusted AJCC 6th stage (x6th stage) and grade (Grade of the tumor) which agree with the published model in literature. The final model was selected through forward, backward, and stepwise selection methods using AIC as the selection criterion. The model includes several significant predictors including adjusted 6th stage, tumor grade, regional lymph nodes status and hormone receptor status. Racial disparity is commonly observed among all the disease. In this model, the black patient has higher odds of death compared to white patients while relatively lower odds of death was calculated for the "Other" race category compared to white patients. This disparity may stem from differences in access to healthcare, late diagnosis, or biological factors. The reweighted model improves fairness across racial groups while maintaining overall predictive accuracy. Although the model performs slightly worse for minority groups, the gap has been reduced compared to the original model. The stages of cancer is the strongest predictor of mortality risk (p values). Higher stages (IIIA, IIIB, IIIC) significantly increase the odds of death compared to stage IIA (reference group). Patients in stage IIIC have the highest odds of death, indicating advanced tumor progression is a critical predictor of survival. Tumor grade is another important factor that affect the health outcomes. In our model, patients with grade 3 and 4 have significant elevated odds of death compared to grade 1 (xxx,

number to indicate). This result is consistent with literature where many models also identified the tumor grade as one of the most significant predictors. Our model also indicated the reduced odds of death for patients with positive estrogen or progesterone receptor status (number xxx). This reflects the improved prognosis often associated with hormone-sensitive tumors. Higher survival odds are associated with more regional lymph nodes examined. This could reflect the effectiveness of tailored treatments to remove the lymph nodes so that the risk of death is reduced significantly. The model also showed that the younger the patient age at diagnosis is, the higher survival odd given the same aggressiveness of the cancer. Older people have other health complications which requires complex treatment scheme and reduce the treatment effectiveness.

Limitations

The model assumes no significant interaction effects, which align with the data but could miss complex relationships. The disparity in model performance for "Other" racial groups suggests the need for further data collection or alternative modeling techniques.

Contribution

Ada Guo wrote the abstract and introduction, compiled data cleaning and EDA results, and synthesized findings into cohesive sections for the report. Khue Nguyen analyzed results, wrote the conclusion and limitations, refined the report structure, and ensured clarity and completeness by adding references. Yixin Zheng wrote the methods and appendix section and constructed the Rmd file(cleaning, correlation matrix, Cramér's V, assumption checks, feature selection, model, cross-validation, fairness reweight).

Reference

- 1. Cao SS, Lu CT. Recent perspectives of breast cancer prognosis and predictive factors. Oncol Lett. 2016;12(5):3674-3678. doi:10.3892/ol.2016.5149
- 2. Cancer of the Breast (Female) Cancer Stat Facts. SEER. Accessed December 19, 2024. https://seer.cancer.gov/statfacts/html/breast.html
- 3. Bundred NJ. Prognostic and predictive factors in breast cancer. Cancer Treat Rev. 2001;27(3):137-142. doi:10.1053/ctrv.2000.0207

- Soerjomataram I, Louwman MWJ, Ribot JG, Roukema JA, Coebergh JWW. An overview of prognostic factors for long-term survivors of breast cancer. Breast Cancer Res Treat. 2008;107(3):309-330. doi:10.1007/s10549-007-9556-1
- 5. Phung MT, Tin Tin S, Elwood JM. Prognostic models for breast cancer: a systematic review. BMC Cancer. 2019;19(1):230. doi:10.1186/s12885-019-5442-6

Table, Plots, and Code Results

Tables

Table 1: Skim Summary for Categorical Variables

skim_variable	n_missingcom	iplete_ra	tfactor.ord	eredfactor.n_un	niq	ukactor.top_counts
race	0	1	FALSE	ć	3	Whi: 3413, Oth: 320, Bla: 291
marital_status	0	1	FALSE	Ę	5	Mar: 2643, Sin: 615, Div: 486,
						Wid: 235
t_stage	0	1	FALSE	<u> </u>	4	T2: 1786, T1: 1603, T3: 533,
						T4: 102
n_stage	0	1	FALSE	ę	3	N1: 2732, N2: 820, N3: 472
x6th_stage	0	1	FALSE	Ę	5	IIA: 1305, IIB: 1130, III: 1050,
						III: 472
differentiate	0	1	FALSE	<u> </u>	4	Mod: 2351, Poo: 1111, Wel:
						543, Und: 19
grade	0	1	FALSE	<u> </u>	4	2: 2351, 3: 1111, 1: 543, 4: 19
a_stage	0	1	FALSE	, 2	2	Reg: 3932, Dis: 92
estrogen_status	0	1	FALSE	2	2	Pos: 3755, Neg: 269
progesterone_stat	tus 0	1	FALSE	2	2	Pos: 3326, Neg: 698
status	0	1	FALSE	6 2	2	Ali: 3408, Dea: 616

Table 2: Skim Summary for Numeric Variables

skim_variable n	_missing complet	e_rate	mean	sd	p0	p25	p50	p75	p100	hist
age	0	1 5	53.972167	8.963134	30	47	54	61	69	_
tumor_size	0	1 3	30.473658	21.119696	1	16	25	38	140	-
regional_node_examir	ned 0	1 1	14.357107	8.099675	1	9	14	19	61	
reginol_node_positive	0	1 4	1.158052	5.109331	1	1	2	5	46	

Table 3: Summary Statistics Grouped by Survival Status

status	mean_age	e sd_age	mean_tumor_sized	_tumor_size	e prop_white	prop_black_other	n_obs
Alive	53.75910	8.808420	29.26878	20.30317	0.8518192	0.1481808	3408
Dead	55.15097	9.698291	37.13961	24.11611	0.8279221	0.1720779	616

Table 4: Variance Inflation Factors for Predictors

Variable	GVIF	Df	GVIF_Ratio
age	1.106908	1	1.052097
race	1.058083	2	1.014215
marital_status	1.127489	4	1.015112
x6th_stage	1.967732	4	1.088293
grade	1.118473	3	1.018836
a_stage	1.210950	1	1.100432
tumor_size	1.365304	1	1.168462
estrogen_status	1.484914	1	1.218570
progesterone_status	1.434692	1	1.197786
regional_node_examined	1.225729	1	1.107127

Table 5: Unstable Coefficients

Variable	Full_Coef	Full_SE	No_Outliers_Coef	No_Outliers_SE	Robust_Coef	Robust_SE
(Intercept)	-5.8924387	1.3069930	-30.590617	438.7266805	-5.0273109	1.3799307
raceOther	-0.4272352	0.2010365	-2.877142	0.7253748	-0.5745467	0.2281736
grade2	0.5309961	0.1831397	16.658587	438.7223333	0.4079614	0.1935342
grade3	0.9060030	0.1917058	17.208447	438.7223392	0.7801358	0.2013946
grade4	1.8504135	0.5421565	17.639998	438.7259010	1.6966710	0.5472605

Table 6: Best Subset Selection Summary

Num_	_Predictors	Adj_R2	Ср	BIC
	1	0.0541705	298.36391	-208.5087
	2	0.0808751	177.39536	-316.4581
	3	0.0879639	146.00298	-340.3144
	4	0.0945059	117.12118	-361.9835
	5	0.0989525	97.81162	-374.4942
	6	0.1026866	81.76122	-383.9066
	7	0.1060954	67.20233	-391.9243
	8	0.1090479	54.73108	-397.9391
	9	0.1112815	45.54176	-400.7425
	10	0.1133639	37.04659	-402.8850

Table 7: Reweighted Logistic Regression Model Results

Predictor	Estimate	Std_Error	Odds_RatioX9	5CILower. X95	CIUpper.	P.Value
(Intercept)	-5.126	1.161	0.006	0.001	0.058	1.02e-
						05
raceBlack	0.455	0.135	1.577	1.211	2.052	0.000713
raceOther	-0.426	0.147	0.653	0.490	0.871	0.003683
$marital_statusMarried$	-0.248	0.134	0.781	0.600	1.015	0.064877
$marital_statusSeparated$	0.696	0.350	2.006	1.011	3.981	0.046555
marital_statusSingle	0.034	0.162	1.035	0.753	1.422	0.833041
$marital_statusWidowed$	0.123	0.204	1.131	0.759	1.687	0.544852
$x6th_stageIIB$	0.533	0.136	1.704	1.305	2.225	9.02e-
						05
x6th_stageIIIA	0.978	0.134	2.660	2.047	3.457	2.52e-
						13
$x6th_stageIIIB$	1.565	0.287	4.781	2.726	8.386	4.82e-
						08

Predictor	Estimate	Std_Error	Odds_RatioX9	5CILower. X95	CIUpper.	P.Value
x6th_stageIIIC	2.006	0.152	7.433	5.519	10.012	< 2e-16
grade2	0.506	0.172	1.659	1.184	2.324	0.003266
grade3	0.862	0.180	2.368	1.664	3.370	1.67e-
						06
grade4	1.825	0.521	6.203	2.233	17.232	0.000463
$estrogen_statusPositive$	-0.714	0.166	0.490	0.354	0.678	1.70e-
						05
progesterone_statusPositiv	re -0.503	0.121	0.605	0.477	0.767	3.37e-
						05
${\rm rn}_{\rm examined}_{\rm log}$	-0.277	0.076	0.758	0.653	0.880	0.000270
age_log	0.974	0.276	2.648	1.543	4.545	0.000410

Plots

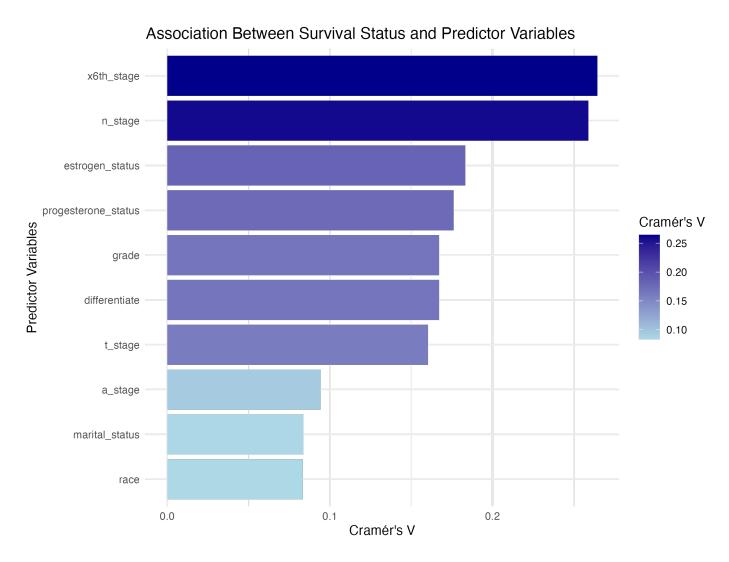


Figure 1: Cramér's V Associations

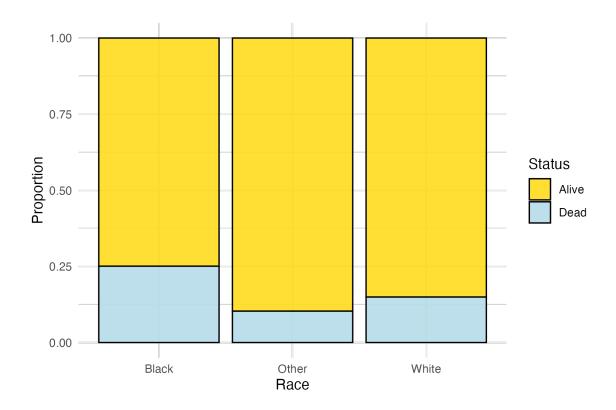


Figure 2: Survival Status by Race

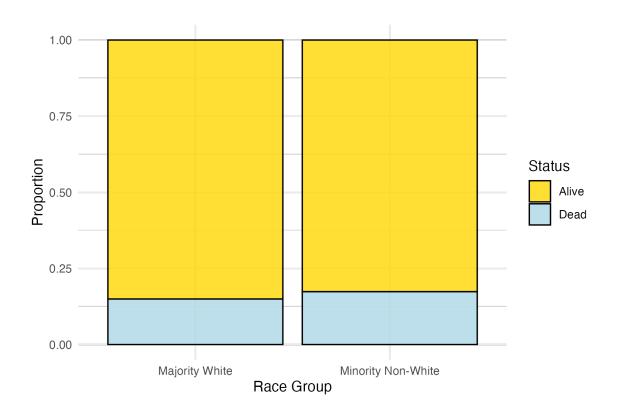


Figure 3: Survival Status by Combined Race Groups

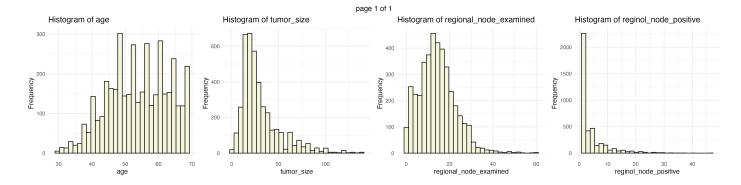


Figure 4: Histograms for Original Continuous Variables

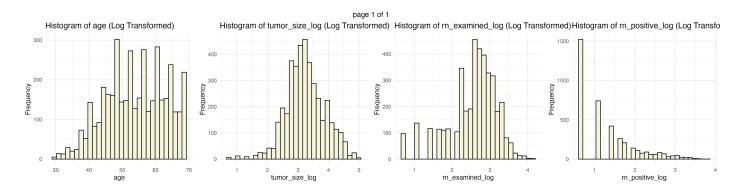


Figure 5: Histograms for Log-Transformed Continuous Variables

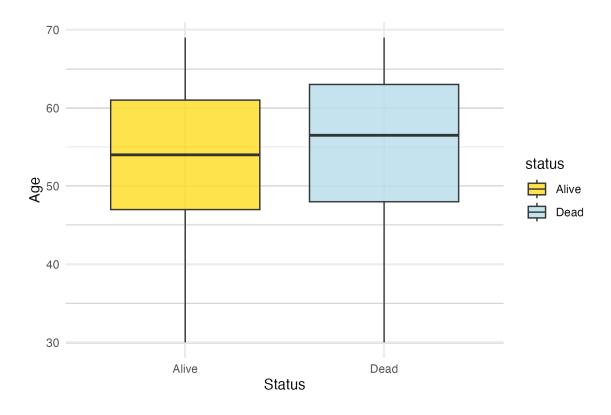


Figure 6: Age by Survival Status

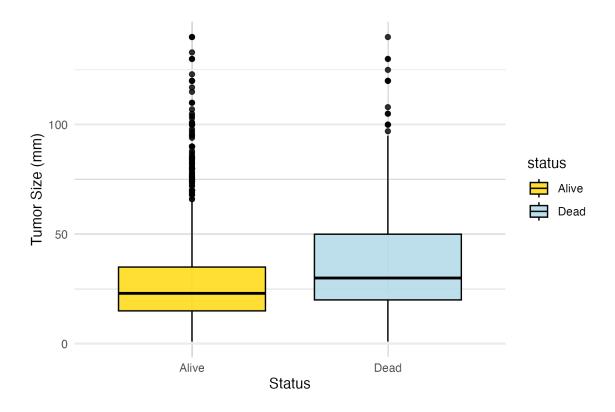


Figure 7: Tumor Size by Survival Status

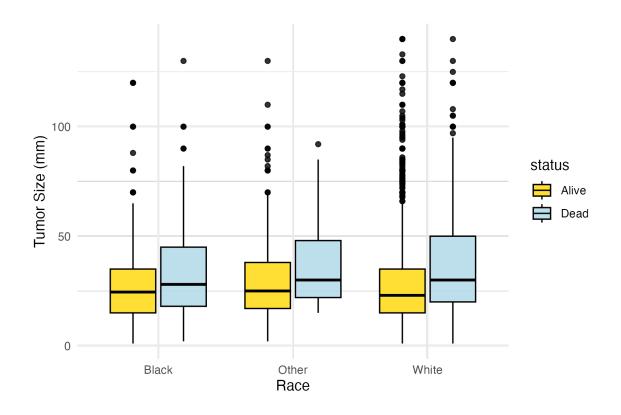


Figure 8: Tumor Size by Race and Survival Status

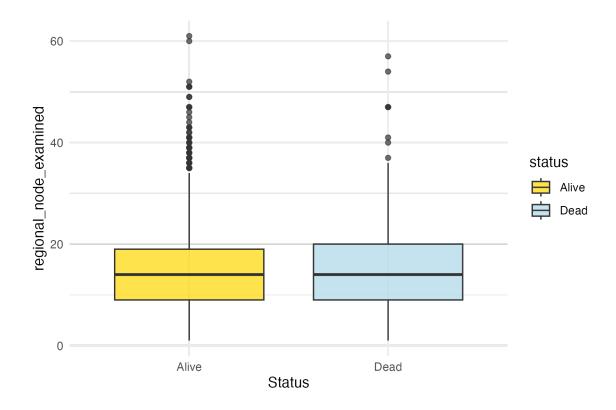


Figure 9: Regional Node Examined by Survival Status

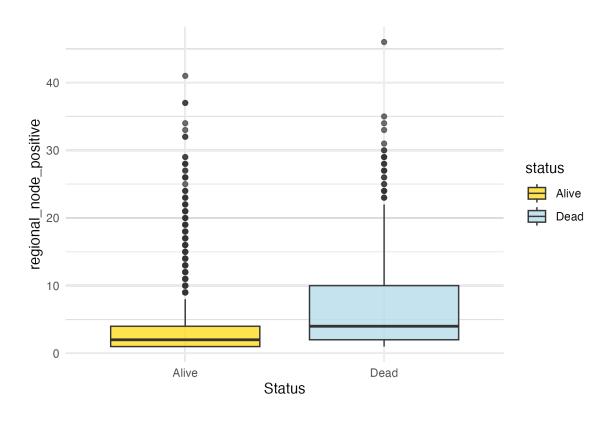


Figure 10: Regional Node Positive by Survival Status

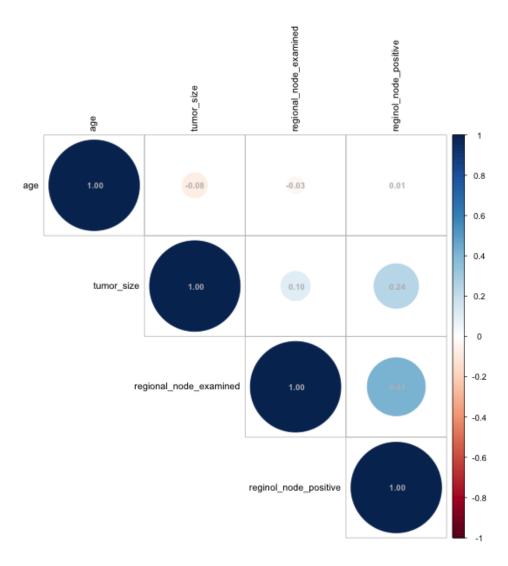


Figure 11: Correlation Matrix for Continuous Variables

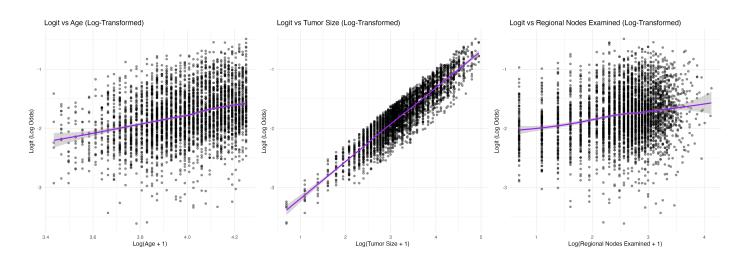


Figure 12: Log Odds Relationship with Predictors

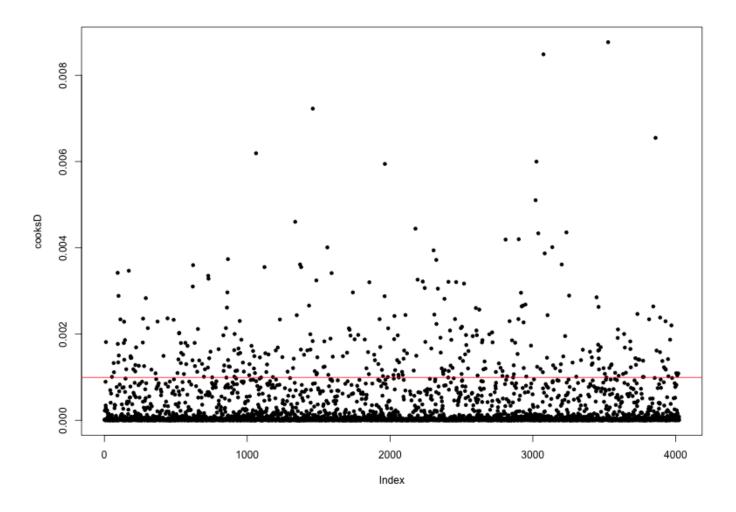


Figure 13: Cook's Distance for Outlier Detection

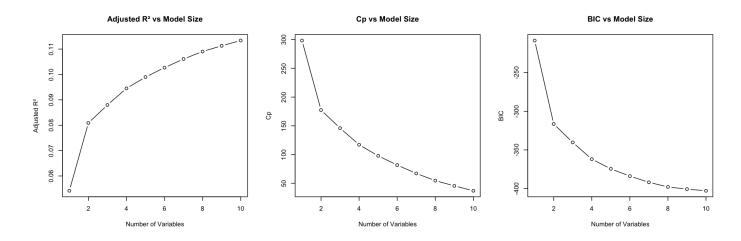


Figure 14: Best Subset Selection Performance Metrics

Code Results

For full code results, please refer to the .txt files available in the results folder of the GitHub repository: Yixin-Zheng/p8130_finalproject.

Code Appendix

```
knitr::opts_chunk$set(echo = TRUE)
library(tidyverse)
library(janitor)
library(skimr)
library(dplyr)
library(ggplot2)
library(caret)
library(corrplot)
library(lsr)
library(vcd)
library(car)
library(gridExtra)
library(robustbase)
library(leaps)
library(pROC)
library(knitr)
# Import data and clean column names
data <- read.csv("./data/Project_2_data.csv") %>%
 clean_names()
# Select relevant covariates (variables 1-14) and outcome variable
model_data <- data %>%
 dplyr::select(-survival_months)
# Convert categorical variables to factors and relabel `grade`
```

```
model data <- model data %>%
 mutate(
   race = factor(race),
   marital_status = factor(marital_status),
   t_stage = factor(t_stage),
   n_stage = factor(n_stage),
   x6th stage = factor(x6th stage),
   differentiate = factor(differentiate),
   a stage = factor(a stage),
    estrogen_status = factor(estrogen_status),
   progesterone_status = factor(progesterone_status),
    status = factor(status, levels = c("Alive", "Dead")),
   grade = case_when(
     grade == "1" ~ "1",
      grade == "2" ~ "2",
      grade == "3" ~ "3",
      grade == " anaplastic; Grade IV" ~ "4",
     TRUE ~ NA character
    ) %>% factor(levels = c("1", "2", "3", "4"))
 )
# Summarize structure of the cleaned dataset
summary(model_data)
# Summary statistics for continuous and categorical variables
skimmed_data <- skim(model_data)</pre>
skim_categorical <- skimmed_data %>%
 filter(skim type == "factor") %>%
 select(-starts_with("numeric"), -skim type) %>%
 na.omit()
write.csv(skim_categorical, "tables/skim_categorical_summary.csv", row.names = FALSE)
```

```
skim numeric <- skimmed data %>%
  filter(skim type == "numeric") %>%
  select(-starts_with("factor"), -skim_type) %>%
 na.omit()
colnames(skim_numeric) <- gsub("^numeric\\.", "", colnames(skim_numeric))</pre>
write.csv(skim_numeric, "tables/skim_numeric_summary.csv", row.names = FALSE)
# Key statistics grouped by survival status
summary_by_status <- model_data %>%
  group_by(status) %>%
  summarise(
    mean_age = mean(age, na.rm = TRUE),
    sd_age = sd(age, na.rm = TRUE),
    mean_tumor_size = mean(tumor_size, na.rm = TRUE),
    sd_tumor_size = sd(tumor_size, na.rm = TRUE),
    prop_white = mean(race == "White", na.rm = TRUE),
    prop black other = mean(race != "White", na.rm = TRUE),
    n obs = n()
  )
write.csv(summary_by_status, "tables/summary_by_status.csv", row.names = FALSE)
# Define categorical variables to analyze
variables <- c("race", "marital_status", "t_stage", "n_stage", "x6th_stage",</pre>
               "differentiate", "grade", "a_stage", "estrogen_status", "progesterone_status
# Initialize a vector to store Cramér's V results
results <- numeric(length(variables))</pre>
# Calculate Cramér's V for each variable
for (i in seq_along(variables)) {
  var <- variables[i]</pre>
```

```
# Select outcome and predictor variable, omitting missing values
  df_temp <- model_data %>%
    dplyr::select(status, all_of(var)) %>%
    na.omit()
  # Convert both columns to factors
  x <- droplevels(as.factor(df temp$status))</pre>
  y <- droplevels(as.factor(df temp[[var]]))</pre>
  # Create contingency table and calculate Cramér's V
  table_var <- table(x, y)</pre>
  results[i] <- cramersV(table var)</pre>
}
# Create a dataframe with results
association_df <- data.frame(Variable = variables, CramersV = results)</pre>
# Plot Cramér's V values
cramerV_association <- ggplot(association_df, aes(x = reorder(Variable, CramersV), y = CramersV), y = CramersV
  geom_bar(stat = "identity") +
  coord_flip() +
  scale_fill_gradient(low = "lightblue", high = "darkblue") +
  labs(
    title = "Association Between Survival Status and Predictor Variables",
    x = "Predictor Variables",
    y = "Cramér's V",
    fill = "Cramér's V"
  ) +
  theme_minimal()
ggsave("plots/cramerV_association.png", plot = cramerV_association, width = 8, height = 6)
# Proportional Bar Plot for Survival Status by Race
```

```
race barplot <- ggplot(model data, aes(x = race, fill = status)) +</pre>
 geom_bar(position = "fill", alpha = 0.8, color = "black") +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Race",
   y = "Proportion",
   fill = "Status"
 )
ggsave("plots/race_proportional_barplot.png", plot = race_barplot, width = 6, height = 4)
# Combine "Black" and "Other" into a single group "Minority Non-White"
model_data_race_combined <- model_data %>%
 mutate(
   race combined = case_when(
     race == "White" ~ "Majority White",
     race %in% c("Black", "Other") ~ "Minority Non-White"
   ),
   race_combined = factor(race_combined, levels = c("Majority White", "Minority Non-White"
 )
# Proportional Bar Plot for Combined Race Groups
race_combined_barplot <- ggplot(model_data_race_combined, aes(x = race_combined, fill = sta
 geom_bar(position = "fill", alpha = 0.8, color = "black") +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme minimal() +
 labs(
   x = "Race Group",
   y = "Proportion",
   fill = "Status"
```

```
ggsave("plots/race_combined_proportional_barplot.png", plot = race_combined_barplot, width
continuous_vars <- model_data %>%
  dplyr::select(age, tumor_size, regional_node_examined, reginol_node_positive) %>%
  na.omit()
df1 <- as.data.frame(continuous vars)</pre>
hist_list <- lapply(names(continuous_vars), function(col) {</pre>
  ggplot(continuous_vars, aes_string(x = col)) +
    geom_histogram(fill = "beige", color = "black", bins = 30) +
    labs(
      title = paste("Histogram of", col),
      x = col,
      y = "Frequency"
    ) +
    theme_minimal()
})
# Arrange plots in a grid
hist_grid <- marrangeGrob(hist_list, nrow = 1, ncol = 4)</pre>
ggsave("plots/original_histograms_grid.png", hist_grid, width = 16, height = 4)
df log <- df1 %>%
  dplyr::select(-tumor_size, -regional_node_examined, -reginol_node_positive) %>%
  mutate(
    tumor_size_log = log(df1$tumor_size + 1),
    rn examined \log = \log(df1\$regional node examined + 1),
    rn positive log = log(df1$reginol node positive + 1)
  )
log_hist_list <- lapply(names(df_log), function(col) {</pre>
```

```
ggplot(df_log, aes_string(x = col)) +
    geom_histogram(fill = "beige", color = "black", bins = 30) +
    labs(
      title = paste("Histogram of", col, "(Log Transformed)"),
     x = col,
     y = "Frequency"
    ) +
    theme_minimal()
})
# Arrange plots in a grid
log_hist_grid <- marrangeGrob(log_hist_list, nrow = 1, ncol = 4)</pre>
ggsave("plots/log_transformed_histograms_grid.png", log_hist_grid, width = 16, height = 4)
# Age by survival status
age_boxplot <- ggplot(model_data, aes(x = status, y = age, fill = status)) +</pre>
  geom_boxplot(alpha = 0.7) +
  scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
  theme_minimal() +
  labs(
   x = "Status",
    y = "Age"
  )
ggsave("plots/age_by_status_boxplot.png", plot = age_boxplot, width = 6, height = 4)
# Tumor size by survival status
tumor_boxplot <- ggplot(model_data, aes(x = status, y = tumor_size, fill = status)) +</pre>
  geom_boxplot(alpha = 0.8, color = "black") +
  scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
  theme_minimal() +
  labs(
    x = "Status",
    y = "Tumor Size (mm)"
```

```
ggsave("plots/tumor_size_by_status_boxplot.png", plot = tumor_boxplot, width = 6, height =
# Tumor size by race and survival status
tumor_race_boxplot <- ggplot(model_data, aes(x = race, y = tumor_size, fill = status)) +</pre>
 geom_boxplot(alpha = 0.8, color = "black") +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Race",
   y = "Tumor Size (mm)"
 )
ggsave("plots/tumor_size_by_race_status_boxplot.png", plot = tumor_race_boxplot, width = 6,
# Regional Node Examined by survival status
rn_examined_boxplot <- ggplot(model_data, aes(x = status, y = regional_node_examined, fill</pre>
 geom_boxplot(alpha = 0.7) +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Status",
   y = "regional_node_examined"
 )
ggsave("plots/rn_examined_by_status_boxplot.png", plot = rn_examined_boxplot, width = 6, he
# Reginol Node Positive by survival status
rn_positive_boxplot <- ggplot(model_data, aes(x = status, y = reginol_node_positive, fill =</pre>
 geom_boxplot(alpha = 0.7) +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
```

```
labs(
    x = "Status",
    y = "regional_node_positive"
  )
ggsave("plots/rn_positive_by_status_boxplot.png", plot = rn_positive_boxplot, width = 6, he
correlation_matrix <- cor(continuous_vars, use = "pairwise.complete.obs")</pre>
# Pairwise relationships (correlation matrix for continuous variables)
correlation_plot <- function() {</pre>
  corrplot(
    correlation_matrix,
    method = "circle",
   type = "upper",
    tl.col = "black",
    addCoef.col = "grey",
    number.cex = 0.8,
    tl.cex = 0.9
  )
}
png("plots/correlation_matrix_plot.png", width = 800, height = 600)
correlation_plot()
dev.off()
# Binary or Dichotomous Response Variable
unique(model_data$status)
#The response variable (status) has exactly two categories: Alive and Dead.
# Fit an initial logistic regression model
model_data_1 <- model_data %>%
 mutate(
    race = relevel(race, ref = "White"), # Set "White" as reference
```

```
grade = relevel(grade, ref = "1"),  # Set "Grade 1" as reference
    x6th stage = relevel(x6th stage, ref = "IIA") # Set "IIA" as reference
  )
alias_results <- capture.output(alias(glm(status ~ ., data = model_data_1, family = binomia
# Save the results to a text file in the `results` folder
writeLines(alias_results, "results/alias_results_model1.txt")
model_data_2 <- model_data_1 %>%
  dplyr::select(-differentiate, -n_stage, -t_stage, -reginol_node_positive)
# Perform multicollinearity check with alias()
alias_results_2 <- capture.output(alias(glm(status ~ ., data = model_data_2, family = binor
writeLines(alias_results_2, "results/alias_results_model2.txt")
# Fit a logistic regression model
model vif <- glm(status ~ ., data = model data 2, family = binomial)</pre>
# Calculate VIF
vif_values <- vif(model_vif)</pre>
vif_df <- as.data.frame(vif_values)</pre>
vif_df <- tibble::rownames_to_column(vif_df, var = "Variable")</pre>
colnames(vif_df) <- c("Variable", "GVIF", "Df", "GVIF_Ratio")</pre>
write.csv(vif_df, "tables/vif.csv", row.names = FALSE)
continuous_vars_log_odds <- model_data_2 %>%
  dplyr::select(age, tumor_size, regional_node_examined, status) %>% # Include 'status'
  na.omit()
# Log-transform tumor size and regional nodes examined
df_log_odds <- continuous_vars_log_odds %>%
  mutate(
```

```
age_log = log(age + 1),
    tumor size log = log(tumor size + 1),
    rn_examined_log = log(regional_node_examined + 1)
  )
linearity_test <- glm(status ~ age_log + tumor_size_log + rn_examined_log,</pre>
                       data = df_log_odds,
                       family = binomial)
df_log_odds$logit <- predict(linearity_test, type = "link")</pre>
plot1 <- ggplot(df_log_odds, aes(x = age_log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Age (Log-Transformed)",
       x = \text{"Log(Age + 1)"}, y = \text{"Logit (Log Odds)"}) +
  theme_minimal()
plot2 <- ggplot(df_log_odds, aes(x = tumor_size_log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Tumor Size (Log-Transformed)", x = "Log(Tumor Size + 1)", y = "Log
  theme_minimal()
plot3 <- ggplot(df_log_odds, aes(x = rn_examined_log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Regional Nodes Examined (Log-Transformed)",
       x = "Log(Regional Nodes Examined + 1)", y = "Logit (Log Odds)") +
  theme_minimal()
```

```
log odds grid <- grid.arrange(plot1, plot2, plot3, ncol = 3)</pre>
ggsave("plots/logit_grid_plot.png", plot = log_odds_grid, width = 18, height = 6)
# Update
model_data_3 <- model_data_2 %>%
 mutate(
    tumor_size_log = log(tumor_size + 1),
    rn_examined_log = log(regional_node_examined + 1),
    age_log = log(age + 1)
  ) %>%
  dplyr::select(-tumor_size, -regional_node_examined, -age)
# Fit logistic regression model with log-transformed predictors
full_model <- glm(status ~ .,</pre>
                 data = model_data_3, family = binomial)
# Calculate Cook's Distance
cooksD <- cooks.distance(model vif)</pre>
# Plot Cook's Distance
png("plots/cooks_distance_plot.png", width = 800, height = 600)
plot(cooksD, pch = 20)
abline(h = 4 / nrow(model_data_3), col = "red")
dev.off()
# Identify influential observations
influential_obs <- which(cooksD > 4 / nrow(model_data_3))
# Build Model
model_no_outliers <- glm(status ~ .,</pre>
                          data = model_data_3[-influential_obs, ], family = binomial)
model_robust <- glmrob(status ~ .,</pre>
```

```
data = model data 3, family = binomial, method = "Mqle")
# Extract summaries
full_coefficients <- summary(full_model)$coefficients</pre>
no_outliers_coefficients <- summary(model_no_outliers)$coefficients</pre>
robust_coefficients <- summary(model_robust)$coefficients</pre>
# Identify unstable coefficients
unstable coeffs <- which(
  (abs(full_coefficients[, "Estimate"] - no_outliers_coefficients[, "Estimate"]) > 2) |
  (abs(full_coefficients[, "Std. Error"] - no_outliers_coefficients[, "Std. Error"]) > 2)
)
unstable_coeffs_df <- data.frame(
  Variable = rownames(full_coefficients)[unstable_coeffs],
  Full_Coef = full_coefficients[unstable_coeffs, "Estimate"],
  Full_SE = full_coefficients[unstable_coeffs, "Std. Error"],
  No_Outliers_Coef = no_outliers_coefficients[unstable_coeffs, "Estimate"],
  No_Outliers_SE = no_outliers_coefficients[unstable_coeffs, "Std. Error"],
  Robust_Coef = robust_coefficients[rownames(full_coefficients)[unstable_coeffs], "Estimate
  Robust_SE = robust_coefficients[rownames(full_coefficients)[unstable_coeffs], "Std. Error
write.csv(unstable_coeffs_df, "tables/unstable_coefficients.csv", row.names = FALSE)
# Extract robustness weights
robust_summary <- capture.output(summary(model_robust))</pre>
start_line <- grep("Robustness weights w.r \\* w.x:", robust_summary)
end_line <- start_line + 5</pre>
desired section <- robust summary[start line:end line]</pre>
cat(paste(desired section, collapse = "\n"))
# Forward selection
forward_model <- step(glm(status ~ ., data = model_data_3, family = binomial), scope = list
```

```
# Backward elimination
backward_model <- step(glm(status ~ ., data = model_data_3, family = binomial), direction =
# Stepwise selection
stepwise_model <- step(glm(status ~ ., data = model_data_3, family = binomial), scope = lis
writeLines(capture.output(summary(full model)), "results/full model summary.txt")
writeLines(capture.output(summary(forward_model)), "results/forward_model_summary.txt")
writeLines(capture.output(summary(backward_model)), "results/backward_model_summary.txt")
writeLines(capture.output(summary(stepwise_model)), "results/stepwise_model_summary.txt")
best_subset <- regsubsets(`status` ~ ., data = model_data_3, nvmax = ncol(model_data_3) - 1</pre>
best summary <- summary(best subset)</pre>
subset_table <- data.frame(</pre>
  Num Predictors = 1:length(best summary$adjr2),
 Adj_R2 = best_summary$adjr2,
  Cp = best_summary$cp,
  BIC = best summary$bic
)
write.csv(subset_table, "tables/best_subset_summary.csv", row.names = FALSE)
best_adjr2_model <- which.max(best_summary$adjr2)</pre>
adjr2 predictor <- names(coef(best subset, best adjr2 model))[-1]</pre>
best cp model <- which.min(best summary$cp)</pre>
cp predictor <- names(coef(best subset, best cp model))[-1]</pre>
best_bic_model <- which.min(best_summary$bic)</pre>
bic_predictor <- names(coef(best_subset, best_bic_model))[-1]</pre>
cat("Best Model by Adjusted R2: ", best_adjr2_model, "variables,", adjr2_predictor, "\n")
cat("Best Model by Cp: ", best_cp_model, "variables,", cp_predictor, "\n")
cat("Best Model by BIC: ", best_bic_model, "variables,", bic_predictor, "\n")
png("plots/best_subset_plots.png", width = 1800, height = 600, res = 150)
```

```
par(mfrow = c(1, 3))
plot(best summary$adjr2,
     type = "b",
     main = "Adjusted R2 vs Model Size",
     xlab = "Number of Variables",
     ylab = "Adjusted R2")
plot(best_summary$cp,
     type = "b",
     main = "Cp vs Model Size",
     xlab = "Number of Variables",
     ylab = "Cp")
plot(best_summary$bic,
    type = "b",
     main = "BIC vs Model Size",
     xlab = "Number of Variables",
     ylab = "BIC")
par(mfrow = c(1, 1))
dev.off()
# Define predictors
predictors <- c("race", "marital_status", "x6th_stage", "grade",</pre>
                "estrogen_status", "progesterone_status", "rn_examined_log", "age_log")
# Create a dataframe to store results
interaction_results <- data.frame(</pre>
  Predictor1 = character(),
  Predictor2 = character(),
 P_Value = numeric(),
 stringsAsFactors = FALSE
)
# Loop through each pair of predictors
for (i in seq_along(predictors)) {
```

```
for (j in seq_along(predictors)) {
    if (i < j) {</pre>
      predictor1 <- predictors[i]</pre>
      predictor2 <- predictors[j]</pre>
      # Fit interaction model
      formula <- as.formula(paste("status ~", paste(predictors, collapse = " + "),</pre>
                                    "+", predictor1, "*", predictor2))
      interaction_model <- glm(formula, family = binomial, data = model_data_3)</pre>
      # Extract interaction term
      interaction_term <- paste(predictor1, predictor2, sep = ":")</pre>
      coef_names <- names(coef(interaction_model))</pre>
      if (interaction_term %in% coef_names) {
        # Extract p-value for the interaction term
        p value <- coef(summary(interaction model))[interaction term, "Pr(>|z|)"]
        # Append to results if p-value < 0.05
        if (p_value < 0.05) {</pre>
          interaction_results <- rbind(interaction_results,</pre>
                                          data.frame(Predictor1 = predictor1,
                                                     Predictor2 = predictor2,
                                                     P_Value = p_value,
                                                      stringsAsFactors = FALSE))
        }
      }
    }
  }
}
# Sort results by p-value
```

```
interaction results <- interaction results[order(interaction results$P Value), ]</pre>
print(interaction results)
set.seed(123)
control <- trainControl(method = "cv", number = 10, classProbs = TRUE, summaryFunction = tw
cv_model <- train(</pre>
  status ~ race + marital_status + x6th_stage + grade + estrogen_status +
    progesterone_status + rn_examined_log + age_log,
  data = model data 3,
 method = "glm",
  family = "binomial",
  trControl = control,
 metric = "ROC"
)
performance_summary <- capture.output(cv_model)</pre>
writeLines(performance summary, "results/cv model performance.txt")
# Split data by race group
white data <- model data 3 %>% filter(race == "White")
minority data <- model data 3 %>% filter(race != "White")
black_data <- model_data_3 %>% filter(race == "Black")
other_data <- model_data_3 %>% filter(race == "Other")
# Predict probabilities for White group
pred_white <- predict(cv_model, newdata = white_data, type = "prob")[, "Dead"]</pre>
roc_white <- roc(white_data$status, pred_white)</pre>
auc_white <- auc(roc_white)</pre>
# Predict probabilities for Minority group
pred_minority <- predict(cv_model, newdata = minority_data, type = "prob")[, "Dead"]</pre>
roc_minority <- roc(minority_data$status, pred_minority)</pre>
auc_minority <- auc(roc_minority)</pre>
```

```
# Predict probabilities for Black group
pred_black <- predict(cv_model, newdata = black_data, type = "prob")[, "Dead"]</pre>
roc_black <- roc(black_data$status, pred_black)</pre>
auc black <- auc(roc black)</pre>
# Predict probabilities for Other group
pred_other <- predict(cv_model, newdata = other_data, type = "prob")[, "Dead"]</pre>
roc_other <- roc(other_data$status, pred_other)</pre>
auc_other <- auc(roc_other)</pre>
# Print AUC results
cat("ROC-AUC for White group:", auc_white, "\n")
cat("ROC-AUC for Black group:", auc_black, "\n")
cat("ROC-AUC for Other group:", auc_other, "\n")
cat("ROC-AUC for Minority group:", auc minority, "\n")
model_data_4 <- model_data_3 %>%
  mutate(weight = case_when(
   race == "White" ~ 1,
   race == "Black" ~ 1.5,
   race == "Other" ~ 2
  ))
minority_data <- model_data_4 %>% filter(race != "White")
# Refit the model with weights
reweighted model <- train(</pre>
  status ~ race + marital_status + x6th_stage + grade + estrogen_status +
    progesterone_status + rn_examined_log + age_log,
  data = model data 4,
  method = "glm",
  family = "binomial",
```

```
trControl = control,
  weights = weight
)
# Evaluate performance on subgroups
pred_white_weighted <- predict(reweighted_model, newdata = white_data, type = "prob")[, "De</pre>
roc white weighted <- roc(white data$status, pred white weighted)
auc white weighted <- auc(roc white weighted)</pre>
pred_minority_weighted <- predict(reweighted_model, newdata = minority_data, type = "prob")</pre>
roc_minority_weighted <- roc(minority_data$status, pred_minority_weighted)</pre>
auc_minority_weighted <- auc(roc_minority_weighted)</pre>
pred_black_weighted <- predict(reweighted_model, newdata = black_data, type = "prob")[, "De</pre>
roc_black_weighted <- roc(black_data$status, pred_black_weighted)</pre>
auc_black_weighted <- auc(roc_black_weighted)</pre>
pred other weighted <- predict(reweighted model, newdata = other data, type = "prob")[, "De
roc other weighted <- roc(other data$status, pred other weighted)
auc_other_weighted <- auc(roc_other_weighted)</pre>
# Print updated results
cat("Reweighted Model - ROC-AUC for White group:", auc_white_weighted, "\n")
cat("Reweighted Model - ROC-AUC for Minority group:", auc_minority_weighted, "\n")
cat("Reweighted Model - ROC-AUC for Black group:", auc_black_weighted, "\n")
cat("Reweighted Model - ROC-AUC for Other group:", auc_other_weighted, "\n")
# Extract coefficients and confidence intervals from the reweighted model
summary_reweighted <- summary(reweighted_model$finalModel)</pre>
coefficients <- coef(summary_reweighted)</pre>
```

```
# Calculate Odds Ratios and 95% Confidence Intervals
odds ratios <- exp(coefficients[, "Estimate"])</pre>
lower ci <- exp(coefficients[, "Estimate"] - 1.96 * coefficients[, "Std. Error"])</pre>
upper_ci <- exp(coefficients[, "Estimate"] + 1.96 * coefficients[, "Std. Error"])
# Combine into a table
results table <- data.frame(
  Predictor = rownames(coefficients),
  Estimate = round(coefficients[, "Estimate"], 3),
  Std_Error = round(coefficients[, "Std. Error"], 3),
  Odds_Ratio = round(odds_ratios, 3),
  `95% CI (Lower) = round(lower ci, 3),
  `95% CI (Upper)` = round(upper_ci, 3),
  `P-Value` = format.pval(coefficients[, "Pr(>|z|)"], digits = 3)
)
# Clean up row names
rownames(results table) <- NULL</pre>
write.csv(results table, "tables/reweighted model results.csv", row.names = FALSE)
skim_categorical <- read.csv("tables/skim_categorical_summary.csv")</pre>
knitr::kable(skim categorical, caption = "Skim Summary for Categorical Variables")
skim_numeric <- read.csv("tables/skim_numeric_summary.csv")</pre>
knitr::kable(skim_numeric, caption = "Skim Summary for Numeric Variables")
summary_by_status <- read.csv("tables/summary_by_status.csv")</pre>
knitr::kable(summary_by_status, caption = "Summary Statistics Grouped by Survival Status")
vif table <- read.csv("tables/vif.csv")</pre>
knitr::kable(vif_table, caption = "Variance Inflation Factors for Predictors")
```

```
unstable_table <- read.csv("tables/unstable_coefficients.csv")
knitr::kable(unstable_table, caption = "Unstable Coefficients")

best_subset_summary <- read.csv("tables/best_subset_summary.csv")
knitr::kable(best_subset_summary, caption = "Best Subset Selection Summary")

reweighted_model_results <- read.csv("tables/reweighted_model_results.csv")
knitr::kable(reweighted_model_results, caption = "Reweighted Logistic Regression Model Results.csv")</pre>
```