Prediction of survival rate for breast cancer patients

Yixin Zheng, Shangzi Gao, Khue Nguyen

Abstract

Breast cancer is a leading cause of cancer-related mortality among women worldwide. This study develops a predictive model for survival outcomes in breast cancer patients using logistic regression, leveraging demographic, clinical, and pathological factors from a prospective cohort dataset. The analysis focuses on identifying key predictors of mortality, evaluating model performance across racial groups, and addressing fairness in prediction accuracy. Tumor stage, grade, and hormone receptor status emerged as significant predictors. The initial logistic regression model achieved a moderate performance, with an area under the receiver operating characteristic curve (ROC-AUC) of 0.74. However, disparities in model performance were observed between racial groups, prompting the implementation of reweighting strategies to enhance fairness. These findings highlight the importance of equitable modeling approaches to improve prognostic accuracy and clinical outcomes in breast cancer care.

Introduction

Breast cancer is highly prevalence with 2.3 million new cases worldwide reported in 2022¹. Approximately 13.1 % of female are diagnosed with breast cancer at some point in their lifetime. This type of cancer accounts for 23% of total cancer cases and 14% of cancer deaths². Advances diagnosis and tailored treatments have reduced mortality rates3. Survival of breast cancer depends on various factors including the tumor size, the grade of tumor⁴, cancer stage, lymph node stages, and socioeconomic and race⁵. Younger patients often have better outcomes⁶ while disparities exist for lower-income and African American populations. This report examines data from a prospective cohort of breast cancer patients, which includes variables including demographic, clinical, and pathological factors, aiming to improve outcomes and address disparities.

Methods

Data Description The dataset includes 10 categorical variables: race (Black, White, Other), marital status (Divorced, Married, Separated, Single, Widowed), tumor stage (T1, T2, T3, T4), lymph node stage (N1, N2, N3), adjusted AJCC 6th stage (IIA, IIB, IIIA, IIIB, IIIC), tumor differentiation (Well, Moderately, Poorly, Undifferentiated), grade (1–4), tumor spread stage (Regional, Distant), estrogen receptor status (Positive, Negative), and progesterone receptor status (Positive, Negative). Additionally, there are 4 continuous variables: age, tumor size, regional nodes examined, and regional nodes positive.

Data Cleaning Column headers were renamed, categorical variables were converted to factors. tumor grade levels were recoded to ensure interpretability. Missing values were assessed, and log transformations were applied to highly skewed continuous variables (tumor size, regional nodes examined, and regional nodes positive) to normalize their distributions.

EDA Methods skim() function is applied to the dataset (model_data) to compute detailed summary statistics for all variables. (table.1 & 2), Group-wise key statistics are calculated based on survival status(table.3) Cramér's V was used to quantify the strength of the association between categorical variables and the binary outcome (Alive/Dead), with values ranging from 0 (no association) to 1 (perfect association). Distributional plots, including proportional bar plots for race groups and histograms for continuous variables, were created to visualize the data. Boxplots stratified by survival status were used to explore relationships between continuous variables and the binary outcome. A correlation matrix for continuous variables was generated to assess pairwise relationship between variables.

Modeling Assumptions and Transformations Logistic regression was chosen as the primary method due to the binary nature of the outcome. Assumptions checked are: 1. The response variable (status) was confirmed to be binary by code.

2. The alias() function identified collinearity between models: grade2, grade3, and grade4 with other predictors, x6th_stageIIIC with n_stage, and differentiate with grade. For simplification, some variables were removed: x6th_stage captures n_stage's information, n_stage was dropped. t_stage (linked to tumor size), differentiate (overlapping with grade), and regional_node_positive (redundant with tumor size and regional_node_examined) were removed.

VIF were calculated, to ensure that all values were below 5 (table.4), indicating no multicollinearity. (dataset updated with dropped variable)

- 3. Continuous predictors were log-transformed, and their relationships with the log odds were examined (fig.12). This confirmed linearity. (dataset updated with transformed variable)
- 4. Independence of Errors: Since there were no group-level structures, the independence assumption was satisfied.
- 5. Outliers: Cook's Distance identified potential outliers exceeding 4/n, which flagged numerous points as influential, likely reflecting population variability rather than errors (fig.13). Models with and without these points were compared. Removing these points destabilized coefficients like grade, making them unreliable. Robust logistic regression (model_robust) mitigated outliers impact, providing stable estimates for predictors (table.5). About 12% of data had reduced influence, while most observations are unaffected. Despite its advantages, we chose the original logistic regression for simplicity and familiarity.

Model Construction and Selection Models were constructed using predictors identified during EDA and assumption checks. Forward, backward, and stepwise selection were applied based on the AIC to select the final model. Interaction effects were tested by examining pairwise interactions between predictors.

Model Validation and Fairness The model was validated using 10-fold cross-validation, evaluating ROC-AUC, sensitivity, and specificity. Fairness was assessed by evaluating model performance across racial subgroups (White, Black, Other) based on subgroup-specific ROC-AUC values. To address disparities, inverse probability weighting (IPW) was applied: the proportions were computed as the size of each group divided by the total sample size (White: 3413, Other: 320, Black: 291) (table.1). The weights W were then derived as follows: $W = \frac{1}{\text{Proportion}}$. The calculated weights were 1.18 for White, 13.89 for Black, and 12.50 for Other. To keep weights manageable, we normalized them such that the maximum weight was scaled to 2, resulting in final weights of 0.17 for White, 2.00 for Black, and 1.80 for Other. The reweighted model's performance was then compared to the original, using subgroup-specific and overall AUC values to assess predictive fairness. Predictor coefficients were interpreted as odds ratios to quantify their impact on survival outcomes.

Results

EDA Cramér's V analysis identified x6th_stage, n_stage, and hormone receptor statuses as the strongest predictors of survival, while marital status, race, and a_stage showed weaker associations (fig.1). Bar plots highlight racial disparities, with higher mortality among Non-White, particularly

Black patients, and combining groups simplifies comparisons (figs.2–3). Histograms revealed significant right skewness for most continuous variables (except age), improved by log transformations for the other continuous vars (figs.4–5). Boxplots confirm these patterns (figs.6–10). A correlation matrix showed weak overall relationships and no multicollinearity, except regional_node_positive is moderately associated with both tumor size (0.24) and regional nodes examined (0.41), guiding modeling choices (fig.11).

Model Selection and Interretation

AIC values differed slightly (Full: 3039.8; Forward: 3039.8; Backward: 3037.5; Stepwise: 3037.5), but all methods yielded the same model. The stepwise model was chosen for its lower AIC (full results attached):

$$Logit(P) = \beta_0 + \beta_1 \times raceBlack + \beta_2 \times raceOther + \ldots + \beta_i \times X_i$$

Where β_i values are taken from Table 10, X_i represents other significant predictors, and $Logit(P) = log(\frac{P}{1-P})$, P is the probability of the status being 1.

Interaction Effects Pairwise interactions between predictors were tested, but none of the interaction terms had p-values below 0.05, consistent with earlier analyses showing weak or absent interaction effects (results attached)

Key Findings on Coefficients of Final Model-Original Table 6 highlights the coefficients for key predictors. Race was significant, with raceBlack (0.47961) associated with higher odds of death and raceOther (-0.42884) linked to lower odds compared to White patients. x6th_stage was the strongest predictor, with higher stages (e.g., IIIC) significantly increasing mortality risk. Poorly differentiated tumors (Grades 3: 0.91053, Grade 4: 1.87333) also raised mortality odds, while positive hormone receptor status reduced the likelihood of death. Log-transformed predictors showed rn_examined_log (-0.29822) improved survival odds, while age_log (1.09581) indicated higher mortality risk for older patients. Marital status had no strong associations.

Model Performance Cross-validation (10-fold) revealed an overall ROC-AUC of 0.7400, indicating moderately good performance with an acceptable ability to distinguish between "Alive" and "Dead" outcomes. The model demonstrated high sensitivity (0.985), meaning it correctly identifies most of the "Dead" cases, but low specificity (0.122), indicating difficulty in correctly identifying "Alive" cases (full results attached).

The model performance varies across different racial groups. It demonstrates high ROC-AUC for the White group (0.7504), while the lower scores were evaluated for the Black group (0.7021) and Other

group (0.6584). When Black and Other groups are combined as a single minority group, the model ROC-AUC improves to 0.7313, though it remains lower than the majority White group. Reweighting narrowed this gap, increasing the model's ROC-AUC for the minority group to 0.7358, closer to the White group (0.7486), improving fairness while maintaining accuracy (table.8). The model equation must be adjusted accordingly if the reweighted model is chosen:

$$Logit(P) = \beta_0 + \beta_1 \times raceBlack + \ldots + \beta_i \times X_i$$

where β_i values are from Table 7, and X_i represents the same significant predictors as in the original stepwise model.

Conclusion and Limitation

This study developed a logistic regression model to predict breast cancer survival, highlighting key predictors and addressing racial disparities. Tumor stage and grade were the strongest predictors, with higher stages (e.g., IIIA, IIIB, IIIC) and poorly differentiated tumors (Grades 3 and 4) significantly increasing mortality risk. Positive hormone receptor status improved survival odds, while older age and inadequate lymph node evaluation increased mortality risk. Racial disparities were evident: Black patients faced higher odds of death (OR = 1.5) compared to White patients, while the "Other" race group had lower odds (OR = 0.6).

Reweighting improved fairness for underrepresented groups, increasing the minority group's ROC-AUC from 0.7313 to 0.7358 and narrowing the gap with the White group (0.7486). However, it may reduce generalizability to majority-dominant populations and alter predictor importance, requiring careful interpretation. Stepwise selection treated factor levels as independent variables (e.g., x6th_stageIIIA as a dummy), potentially excluding some levels and raising interpretability concerns. Marital status was retained in the final model despite weak significance due to its potential theoretical relevance, confounding effects, or fairness considerations. All three aspects—reweighting, factor level treatment, and marital status inclusion—require further research to ensure robustness and interpretability.

Contribution

All members participated in group discussions, and editing reports. Ada Guo drafted the abstract, introduction, and results-eda. Khue Nguyen analyzed and draft results, conclusion and limitations,

refined results structure, add references. Yixin Zheng wrote the methods, revise the results, conclusion, limitation, and constructed the Rmd file(cleaning, correlation matrix, Cramér's V, assumption checks, feature selection, model, cross-validation, fairness reweight, appendix).

Reference

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Table, Plots, and Code Results

Tables

Table 1: Skim Summary for Categorical Variables

skim_variable	n_missingcom	plete_ra	ttactor.orde	eredfactor.n_u	niq	ukactor.top_counts
race	0	1	FALSE		3	Whi: 3413, Oth: 320, Bla: 291
marital_status	0	1	FALSE		5	Mar: 2643, Sin: 615, Div: 486,
						Wid: 235
t_stage	0	1	FALSE		4	T2: 1786, T1: 1603, T3: 533,
						T4: 102
n_stage	0	1	FALSE		3	N1: 2732, N2: 820, N3: 472
x6th_stage	0	1	FALSE		5	IIA: 1305, IIB: 1130, III: 1050,
						III: 472
differentiate	0	1	FALSE		4	Mod: 2351, Poo: 1111, Wel:
						543, Und: 19
grade	0	1	FALSE		4	2: 2351, 3: 1111, 1: 543, 4: 19
a_stage	0	1	FALSE		2	Reg: 3932, Dis: 92
estrogen_status	0	1	FALSE		2	Pos: 3755, Neg: 269
progesterone_stat	tus 0	1	FALSE		2	Pos: 3326, Neg: 698
status	0	1	FALSE		2	Ali: 3408, Dea: 616

Table 2: Skim Summary for Numeric Variables

skim_variable n	_missing complet	e_rate	mean	sd	p0	p25	p50	p75	p100	hist
age	0	1 5	53.972167	8.963134	30	47	54	61	69	_
tumor_size	0	1 3	30.473658	21.119696	1	16	25	38	140	-
regional_node_examir	ned 0	1 1	14.357107	8.099675	1	9	14	19	61	
reginol_node_positive	0	1 4	1.158052	5.109331	1	1	2	5	46	

Table 3: Summary Statistics Grouped by Survival Status

status	mean_age sd_a	ge mean_tumor_siz s o	d_tumor_size p	prop_white prop_	_black_other	n_obs
Alive	53.75910 8.8084	29.26878	20.30317	0.8518192	0.1481808	3408
Dead	55.15097 9.6982	91 37.13961	24.11611	0.8279221	0.1720779	616

Table 4: Variance Inflation Factors for Predictors

Variable	GVIF	Df	GVIF_Ratio
age	1.106908	1	1.052097
race	1.058083	2	1.014215
marital_status	1.127489	4	1.015112
x6th_stage	1.967732	4	1.088293
grade	1.118473	3	1.018836
a_stage	1.210950	1	1.100432
tumor_size	1.365304	1	1.168462
estrogen_status	1.484914	1	1.218570
progesterone_status	1.434692	1	1.197786
regional_node_examined	1.225729	1	1.107127

Table 5: Unstable Coefficients

Variable	Full_Coef	Full_SE	No_Outliers_Coef	No_Outliers_SE	Robust_Coef	Robust_SE
(Intercept)	-5.8924387	1.3069930	-30.590617	438.7266805	-5.0273109	1.3799307
raceOther	-0.4272352	0.2010365	-2.877142	0.7253748	-0.5745467	0.2281736
grade2	0.5309961	0.1831397	16.658587	438.7223333	0.4079614	0.1935342
grade3	0.9060030	0.1917058	17.208447	438.7223392	0.7801358	0.2013946
grade4	1.8504135	0.5421565	17.639998	438.7259010	1.6966710	0.5472605

Table 6: Stepwise Model Results

	Estimate	Std. Error	z value	$\Pr(> z)$
(Intercept)	-5.5402471	1.2360055	-4.4823805	0.0000074
raceBlack	0.4796129	0.1604322	2.9895047	0.0027943
raceOther	-0.4288428	0.2008699	-2.1349280	0.0327669
$marital_statusMarried$	-0.2257394	0.1398532	-1.6141167	0.1065021
$marital_statusSeparated$	0.6844028	0.3807017	1.7977402	0.0722182
$marital_statusSingle$	-0.0355339	0.1726281	-0.2058410	0.8369152
$marital_statusWidowed$	0.0321840	0.2181767	0.1475133	0.8827269
x6th_stageIIB	0.5232540	0.1439353	3.6353418	0.0002776
x6th_stageIIIA	0.9870602	0.1411669	6.9921488	0.0000000
x6th_stageIIIB	1.5728667	0.3020784	5.2068165	0.0000002
$x6th_stageIIIC$	2.0388975	0.1606928	12.6881737	0.0000000
grade2	0.5325726	0.1829927	2.9103483	0.0036103
grade3	0.9105293	0.1915207	4.7542082	0.0000020
grade4	1.8733288	0.5418545	3.4572545	0.0005457
estrogen_statusPositive	-0.7232933	0.1754783	-4.1218401	0.0000376
progesterone_statusPositive	-0.5680958	0.1266501	-4.4855520	0.0000073
rn_examined_log	-0.2982186	0.0800080	-3.7273614	0.0001935
age_log	1.0958051	0.2938018	3.7297427	0.0001917

Table 7: Reweighted Logistic Regression Model Results

Predictor	Estimate	Std_Error	· Odds_RatioX	95CILower. X95	CIUpper	. P.Value
(Intercept)	-4.065	1.775	0.017	0.001	0.556	0.0220000
raceBlack	0.353	0.166	1.423	1.027	1.971	0.0337700
raceOther	-0.405	0.189	0.667	0.460	0.967	0.0324200
$marital_statusMarried$	-0.328	0.222	0.720	0.466	1.113	0.1397100
$marital_statusSeparated$	0.820	0.474	2.270	0.896	5.748	0.0838600
marital_statusSingle	0.418	0.243	1.519	0.944	2.443	0.0850800

Predictor	Estimate	Std_Error	Odds_RatioX9	5CILower. X9	5CIUpper.	P.Value
marital_statusWidowed	0.427	0.301	1.533	0.850	2.766	0.1560400
x6th_stageIIB	0.516	0.209	1.675	1.113	2.522	0.0133900
$x6th_stageIIIA$	0.918	0.206	2.505	1.672	3.754	0.0000086
$x6th_stageIIIB$	1.426	0.449	4.163	1.727	10.032	0.0014800
$x6th_stageIIIC$	1.833	0.234	6.255	3.956	9.889	0.0000000
grade2	0.389	0.263	1.476	0.881	2.472	0.1395800
grade3	0.656	0.274	1.926	1.126	3.295	0.0167000
grade4	1.626	0.826	5.081	1.007	25.647	0.0490700
estrogen_statusPositive	-0.559	0.258	0.572	0.345	0.947	0.0300200
progesterone_statusPositi	ve -0.288	0.197	0.750	0.509	1.104	0.1449600
rn_examined_log	-0.172	0.121	0.842	0.665	1.067	0.1551800
age_log	0.607	0.418	1.835	0.808	4.164	0.1467700

 $\label{thm:comparison} \mbox{Table 8: Comparison of ROC-AUC Values Across Models} \\ \mbox{and Racial Groups}$

Model Type	White	Black	Other	Minority
Original	0.7504	0.7021	0.6584	0.7313
Reweighted	0.7302	0.7189	0.6962	0.7526

Plots

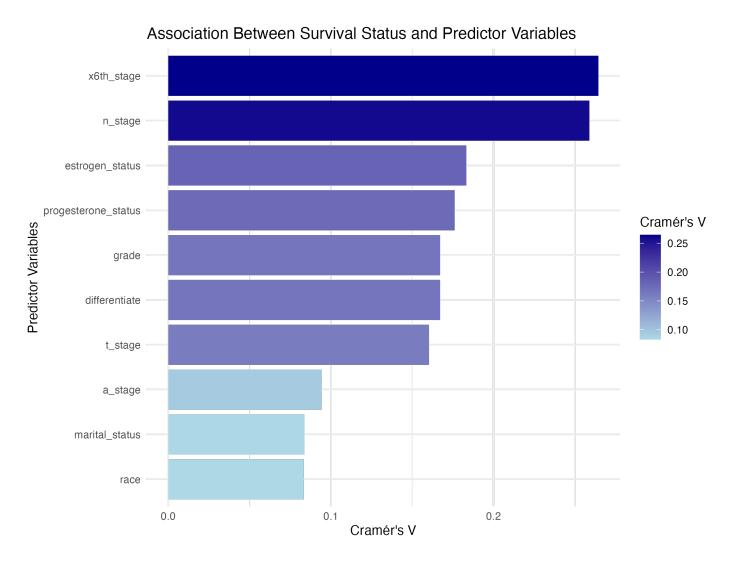


Figure 1: Cramér's V Associations

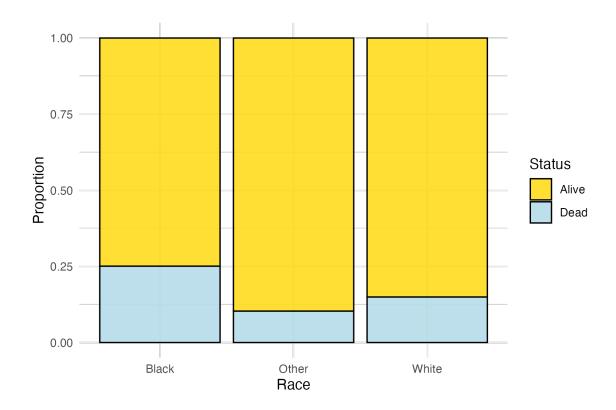


Figure 2: Survival Status by Race

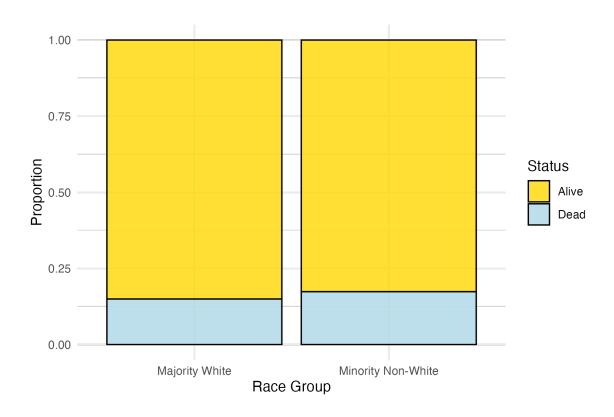


Figure 3: Survival Status by Combined Race Groups

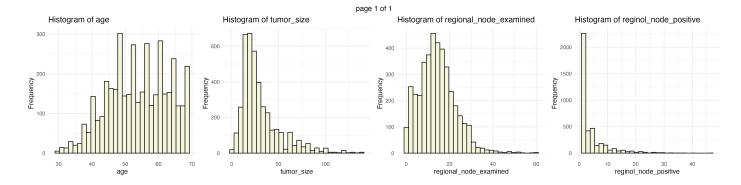


Figure 4: Histograms for Original Continuous Variables

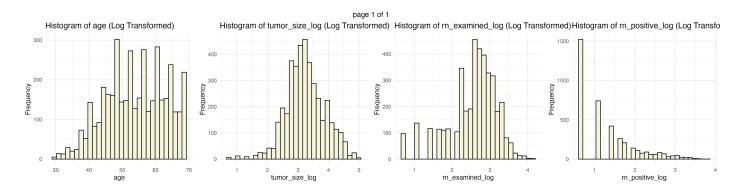


Figure 5: Histograms for Log-Transformed Continuous Variables

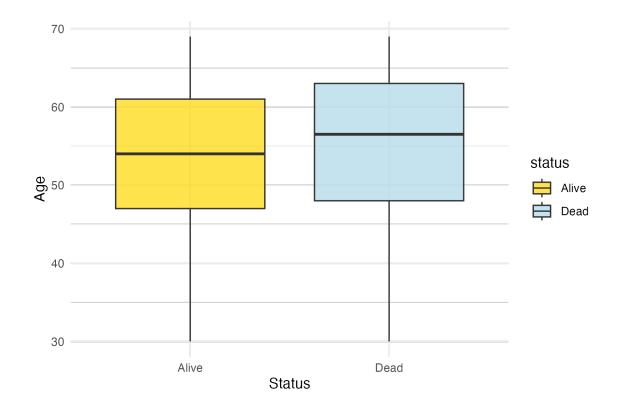


Figure 6: Age by Survival Status

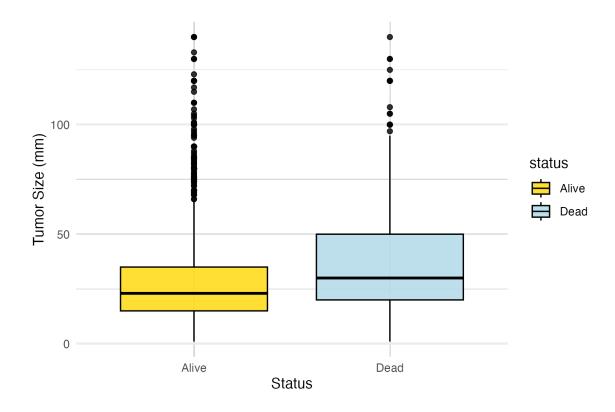


Figure 7: Tumor Size by Survival Status

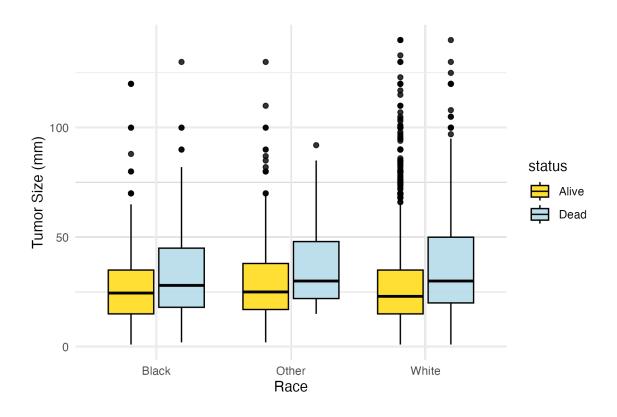


Figure 8: Tumor Size by Race and Survival Status

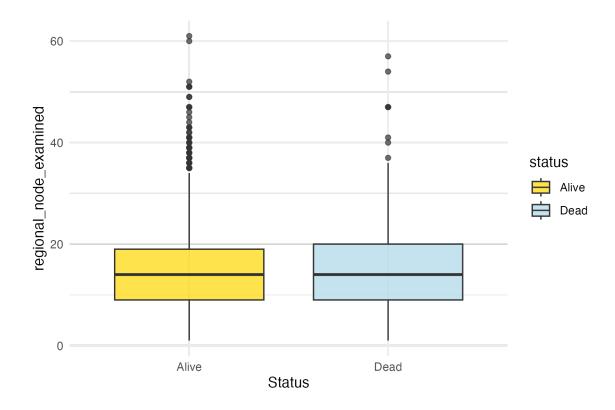


Figure 9: Regional Node Examined by Survival Status

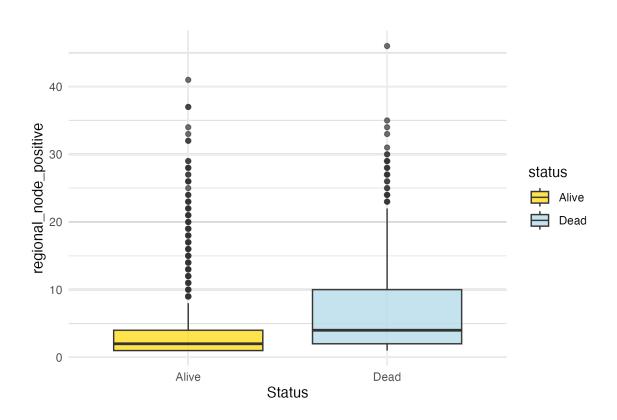


Figure 10: Regional Node Positive by Survival Status

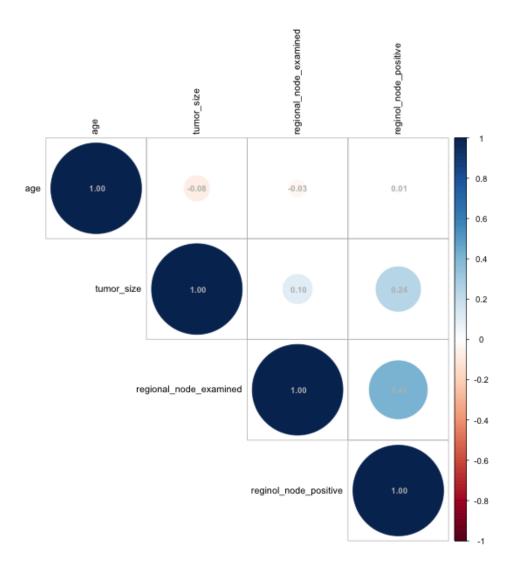


Figure 11: Correlation Matrix for Continuous Variables

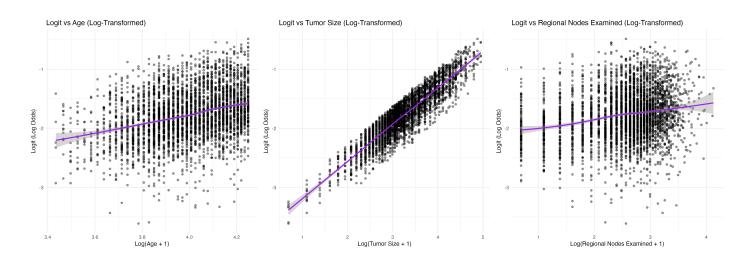


Figure 12: Log Odds Relationship with Predictors

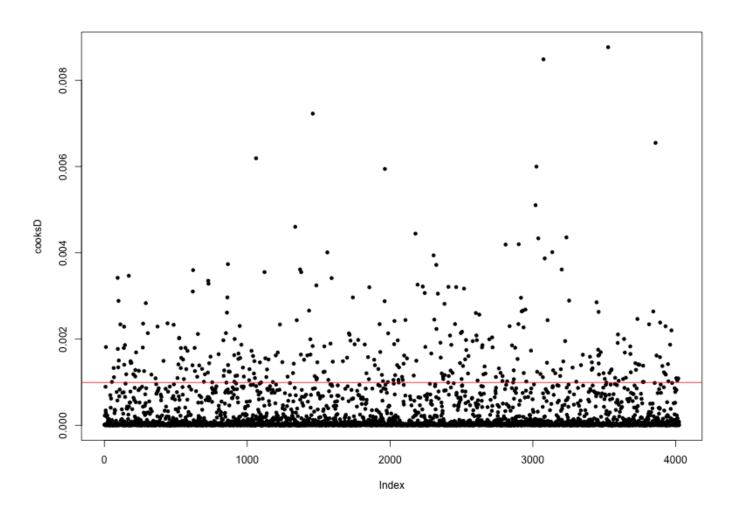


Figure 13: Cook's Distance for Outlier Detection

Code Results

For full code results, please refer to the .txt files available in the results folder of the GitHub repository: https://github.com/Yixin-Zheng/p8130_finalproject.

Code Appendix

```
knitr::opts_chunk$set(echo = TRUE)
library(tidyverse)
library(janitor)
library(skimr)
library(dplyr)
library(ggplot2)
library(caret)
library(corrplot)
library(lsr)
library(vcd)
library(car)
library(gridExtra)
library(robustbase)
library(leaps)
library(pROC)
library(knitr)
# Import data and clean column names
data <- read.csv("./data/Project_2_data.csv") %>%
 clean_names()
# Select relevant covariates (variables 1-14) and outcome variable
model_data <- data %>%
 dplyr::select(-survival_months)
# Convert categorical variables to factors and relabel `grade`
```

```
model data <- model data %>%
 mutate(
   race = factor(race),
   marital_status = factor(marital_status),
   t_stage = factor(t_stage),
   n_stage = factor(n_stage),
   x6th stage = factor(x6th stage),
   differentiate = factor(differentiate),
   a_stage = factor(a_stage),
    estrogen_status = factor(estrogen_status),
   progesterone_status = factor(progesterone_status),
    status = factor(status, levels = c("Alive", "Dead")),
   grade = case_when(
     grade == "1" ~ "1",
      grade == "2" ~ "2",
      grade == "3" ~ "3",
      grade == " anaplastic; Grade IV" ~ "4",
     TRUE ~ NA character
    ) %>% factor(levels = c("1", "2", "3", "4"))
 )
# Summarize structure of the cleaned dataset
summary(model_data)
# Summary statistics for continuous and categorical variables
skimmed_data <- skim(model_data)</pre>
skim_categorical <- skimmed_data %>%
 filter(skim type == "factor") %>%
 select(-starts_with("numeric"), -skim type) %>%
 na.omit()
write.csv(skim_categorical, "tables/skim_categorical_summary.csv", row.names = FALSE)
```

```
skim numeric <- skimmed data %>%
  filter(skim type == "numeric") %>%
  select(-starts_with("factor"), -skim_type) %>%
 na.omit()
colnames(skim_numeric) <- gsub("^numeric\\.", "", colnames(skim_numeric))</pre>
write.csv(skim_numeric, "tables/skim_numeric_summary.csv", row.names = FALSE)
# Key statistics grouped by survival status
summary_by_status <- model_data %>%
  group_by(status) %>%
  summarise(
    mean_age = mean(age, na.rm = TRUE),
    sd_age = sd(age, na.rm = TRUE),
    mean_tumor_size = mean(tumor_size, na.rm = TRUE),
    sd_tumor_size = sd(tumor_size, na.rm = TRUE),
    prop_white = mean(race == "White", na.rm = TRUE),
    prop black other = mean(race != "White", na.rm = TRUE),
    n obs = n()
  )
write.csv(summary_by_status, "tables/summary_by_status.csv", row.names = FALSE)
variables <- c("race", "marital_status", "t_stage", "n_stage", "x6th_stage",</pre>
               "differentiate", "grade", "a_stage", "estrogen_status", "progesterone_status
# Initialize a vector
results <- numeric(length(variables))</pre>
for (i in seq_along(variables)) {
  var <- variables[i]</pre>
  # Select outcome and predictor variable, omit N/A
  df_temp <- model_data %>%
```

```
dplyr::select(status, all_of(var)) %>%
    na.omit()
  x <- droplevels(as.factor(df_temp$status))</pre>
  y <- droplevels(as.factor(df_temp[[var]]))</pre>
  # calculate Cramér's V
  table var <- table(x, y)
  results[i] <- cramersV(table var)</pre>
}
association_df <- data.frame(Variable = variables, CramersV = results)</pre>
# Plot Cramér's V values
cramerV_association <- ggplot(association_df, aes(x = reorder(Variable, CramersV), y = CramersV)
  geom_bar(stat = "identity") +
  coord_flip() +
  scale_fill_gradient(low = "lightblue", high = "darkblue") +
  labs(
    title = "Association Between Survival Status and Predictor Variables",
    x = "Predictor Variables",
    y = "Cramér's V",
   fill = "Cramér's V"
  ) +
  theme_minimal()
ggsave("plots/cramerV_association.png", plot = cramerV_association, width = 8, height = 6)
# Proportional Bar Plot for Survival Status by Race
race barplot <- ggplot(model data, aes(x = race, fill = status)) +</pre>
  geom_bar(position = "fill", alpha = 0.8, color = "black") +
  scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
  theme_minimal() +
```

```
labs(
   x = "Race",
   y = "Proportion",
   fill = "Status"
 )
ggsave("plots/race_proportional_barplot.png", plot = race_barplot, width = 6, height = 4)
# Combine "Black" and "Other" into a single group "Minority Non-White"
model_data_race_combined <- model_data %>%
 mutate(
   race_combined = case_when(
     race == "White" ~ "Majority White",
     race %in% c("Black", "Other") ~ "Minority Non-White"
   ),
   race combined = factor(race combined, levels = c("Majority White", "Minority Non-White"
 )
# Proportional Bar Plot for Combined Race Groups
race_combined_barplot <- ggplot(model_data_race_combined, aes(x = race_combined, fill = sta
 geom_bar(position = "fill", alpha = 0.8, color = "black") +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Race Group",
   y = "Proportion",
   fill = "Status"
 )
ggsave("plots/race_combined_proportional_barplot.png", plot = race_combined_barplot, width
continuous_vars <- model_data %>%
```

```
dplyr::select(age, tumor size, regional node examined, reginol node positive) %>%
  na.omit()
df1 <- as.data.frame(continuous_vars)</pre>
hist_list <- lapply(names(continuous_vars), function(col) {</pre>
  ggplot(continuous_vars, aes_string(x = col)) +
    geom_histogram(fill = "beige", color = "black", bins = 30) +
    labs(
      title = paste("Histogram of", col),
      x = col,
      y = "Frequency"
    theme_minimal()
})
# Arrange plots in a grid
hist grid <- marrangeGrob(hist list, nrow = 1, ncol = 4)
ggsave("plots/original_histograms_grid.png", hist_grid, width = 16, height = 4)
df log <- df1 %>%
  dplyr::select(-tumor_size, -regional_node_examined, -reginol_node_positive) %>%
  mutate(
    tumor_size_log = log(df1$tumor_size + 1),
    rn_examined_log = log(df1$regional_node_examined + 1),
    rn_positive_log = log(df1$reginol_node_positive + 1)
  )
log hist list <- lapply(names(df log), function(col) {</pre>
  ggplot(df log, aes_string(x = col)) +
    geom_histogram(fill = "beige", color = "black", bins = 30) +
    labs(
      title = paste("Histogram of", col, "(Log Transformed)"),
```

```
x = col,
      y = "Frequency"
    ) +
    theme_minimal()
})
# Arrange plots in a grid
log_hist_grid <- marrangeGrob(log_hist_list, nrow = 1, ncol = 4)</pre>
ggsave("plots/log_transformed_histograms_grid.png", log_hist_grid, width = 16, height = 4)
# Age by survival status
age_boxplot <- ggplot(model_data, aes(x = status, y = age, fill = status)) +</pre>
  geom_boxplot(alpha = 0.7) +
  scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
  theme_minimal() +
  labs(
   x = "Status",
    y = "Age"
  )
ggsave("plots/age_by_status_boxplot.png", plot = age_boxplot, width = 6, height = 4)
# Tumor size by survival status
tumor_boxplot <- ggplot(model_data, aes(x = status, y = tumor_size, fill = status)) +</pre>
  geom_boxplot(alpha = 0.8, color = "black") +
  scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
  theme_minimal() +
  labs(
   x = "Status",
    y = "Tumor Size (mm)"
  )
ggsave("plots/tumor_size_by_status_boxplot.png", plot = tumor_boxplot, width = 6, height =
```

```
# Tumor size by race and survival status
tumor_race_boxplot <- ggplot(model_data, aes(x = race, y = tumor_size, fill = status)) +</pre>
 geom_boxplot(alpha = 0.8, color = "black") +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Race",
   y = "Tumor Size (mm)"
 )
ggsave("plots/tumor_size_by_race_status_boxplot.png", plot = tumor_race_boxplot, width = 6,
# Regional Node Examined by survival status
rn examined boxplot <- ggplot(model data, aes(x = status, y = regional node examined, fill
 geom_boxplot(alpha = 0.7) +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Status",
   y = "regional node examined"
 )
ggsave("plots/rn_examined_by_status_boxplot.png", plot = rn_examined_boxplot, width = 6, he
# Reginol Node Positive by survival status
rn_positive_boxplot <- ggplot(model_data, aes(x = status, y = reginol_node_positive, fill =</pre>
 geom_boxplot(alpha = 0.7) +
 scale_fill_manual(values = c("Alive" = "gold", "Dead" = "lightblue")) +
 theme_minimal() +
 labs(
   x = "Status",
   y = "regional_node_positive"
 )
```

```
ggsave("plots/rn_positive_by_status_boxplot.png", plot = rn_positive_boxplot, width = 6, he
correlation matrix <- cor(continuous vars, use = "pairwise.complete.obs")</pre>
# Pairwise relationships (correlation matrix for continuous variables)
correlation_plot <- function() {</pre>
 corrplot(
   correlation_matrix,
   method = "circle",
   type = "upper",
   tl.col = "black",
   addCoef.col = "grey",
   number.cex = 0.8,
   tl.cex = 0.9
 )
}
png("plots/correlation matrix plot.png", width = 800, height = 600)
correlation_plot()
dev.off()
# Binary or Dichotomous Response Variable
unique(model_data$status)
#The response variable (status) has exactly two categories: Alive and Dead.
# Fit an initial logistic regression model
model_data_1 <- model_data %>%
 mutate(
   race = relevel(race, ref = "White"), # Set "White" as reference
   grade = relevel(grade, ref = "1"),  # Set "Grade 1" as reference
   x6th_stage = relevel(x6th_stage, ref = "IIA") # Set "IIA" as reference
 )
```

```
alias results <- capture.output(alias(glm(status ~ ., data = model data 1, family = binomia
writeLines(alias_results, "results/alias_results_model1.txt")
model_data_2 <- model_data_1 %>%
  dplyr::select(-differentiate, -n_stage, -t_stage, -reginol_node_positive)
# Multicollinearity check with alias()
alias_results_2 <- capture.output(alias(glm(status ~ ., data = model_data_2, family = binor
writeLines(alias_results_2, "results/alias_results_model2.txt")
# Fit a logistic regression model
model_vif <- glm(status ~ ., data = model_data_2, family = binomial)</pre>
# Calculate VIF
vif_values <- vif(model_vif)</pre>
vif_df <- as.data.frame(vif_values)</pre>
vif df <- tibble::rownames_to_column(vif df, var = "Variable")</pre>
colnames(vif df) <- c("Variable", "GVIF", "Df", "GVIF Ratio")</pre>
write.csv(vif df, "tables/vif.csv", row.names = FALSE)
continuous_vars_log_odds <- model_data_2 %>%
  dplyr::select(age, tumor_size, regional_node_examined, status) %>% # Include 'status'
 na.omit()
# Log-transform tumor size and regional nodes examined
df_log_odds <- continuous_vars_log_odds %>%
  mutate(
    age_log = log(age + 1),
    tumor size log = log(tumor size + 1),
   rn examined log = log(regional node examined + 1)
  )
linearity_test <- glm(status ~ age_log + tumor_size_log + rn_examined_log,</pre>
```

```
data = df log odds,
                       family = binomial)
df_log_odds$logit <- predict(linearity_test, type = "link")</pre>
plot1 <- ggplot(df log odds, aes(x = age log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Age (Log-Transformed)",
       x = \text{"Log(Age + 1)"}, y = \text{"Logit (Log Odds)"}) +
  theme_minimal()
plot2 <- ggplot(df_log_odds, aes(x = tumor_size_log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Tumor Size (Log-Transformed)", x = "Log(Tumor Size + 1)", y = "Log
  theme_minimal()
plot3 <- ggplot(df_log_odds, aes(x = rn_examined_log, y = logit)) +</pre>
  geom_point(alpha = 0.4) +
  geom_smooth(method = "loess", color = "purple") +
  labs(title = "Logit vs Regional Nodes Examined (Log-Transformed)",
       x = "Log(Regional Nodes Examined + 1)", y = "Logit (Log Odds)") +
  theme_minimal()
log_odds_grid <- grid.arrange(plot1, plot2, plot3, ncol = 3)</pre>
ggsave("plots/logit grid plot.png", plot = log odds grid, width = 18, height = 6)
# Update
model_data_3 <- model_data_2 %>%
  mutate(
```

```
tumor size log = log(tumor size + 1),
    rn_examined_log = log(regional_node_examined + 1),
    age_log = log(age + 1)
  ) %>%
  dplyr::select(-tumor_size, -regional_node_examined, -age)
# Fit logistic regression model with log-transformed predictors
full_model <- glm(status ~ .,</pre>
                 data = model data 3, family = binomial)
# Cook's Distance
cooksD <- cooks.distance(model vif)</pre>
# Plot
png("plots/cooks_distance_plot.png", width = 800, height = 600)
plot(cooksD, pch = 20)
abline(h = 4 / nrow(model_data_3), col = "red")
dev.off()
# Identify
influential_obs <- which(cooksD > 4 / nrow(model_data_3))
# Build Model
model_no_outliers <- glm(status ~ .,</pre>
                          data = model_data_3[-influential_obs, ], family = binomial)
model_robust <- glmrob(status ~ .,</pre>
                        data = model_data_3, family = binomial, method = "Mqle")
# Extract summaries
full_coefficients <- summary(full_model)$coefficients</pre>
no_outliers_coefficients <- summary(model_no_outliers)$coefficients</pre>
robust_coefficients <- summary(model_robust)$coefficients</pre>
```

```
# Identify unstable coefficients
unstable_coeffs <- which(
  (abs(full_coefficients[, "Estimate"] - no_outliers_coefficients[, "Estimate"]) > 2) |
  (abs(full_coefficients[, "Std. Error"] - no_outliers_coefficients[, "Std. Error"]) > 2)
)
unstable_coeffs_df <- data.frame(
 Variable = rownames(full_coefficients)[unstable_coeffs],
 Full_Coef = full_coefficients[unstable_coeffs, "Estimate"],
 Full_SE = full_coefficients[unstable_coeffs, "Std. Error"],
 No_Outliers_Coef = no_outliers_coefficients[unstable_coeffs, "Estimate"],
 No_Outliers_SE = no_outliers_coefficients[unstable_coeffs, "Std. Error"],
 Robust_Coef = robust_coefficients[rownames(full_coefficients)[unstable_coeffs], "Estimate
 Robust_SE = robust_coefficients[rownames(full_coefficients)[unstable_coeffs], "Std. Error
)
write.csv(unstable_coeffs_df, "tables/unstable_coefficients.csv", row.names = FALSE)
# Extract robustness weights
robust_summary <- capture.output(summary(model_robust))</pre>
start_line <- grep("Robustness weights w.r \\* w.x:", robust_summary)
end line <- start line + 5
desired_section <- robust_summary[start_line:end_line]</pre>
cat(paste(desired_section, collapse = "\n"))
# Forward selection
forward_model <- step(glm(status ~ ., data = model_data_3, family = binomial), scope = list
# Backward elimination
backward_model <- step(glm(status ~ ., data = model_data_3, family = binomial), direction =
# Stepwise selection
stepwise_model <- step(glm(status ~ ., data = model_data_3, family = binomial), scope = lis
```

```
writeLines(capture.output(summary(full_model)), "results/full_model_summary.txt")
writeLines(capture.output(summary(forward model)), "results/forward model summary.txt")
writeLines(capture.output(summary(backward_model)), "results/backward_model_summary.txt")
writeLines(capture.output(summary(stepwise_model)), "results/stepwise_model_summary.txt")
# Define predictors
predictors <- c("race", "marital_status", "x6th_stage", "grade",</pre>
                 "estrogen status", "progesterone status", "rn examined log", "age log")
interaction results <- data.frame(</pre>
  Predictor1 = character(),
  Predictor2 = character(),
  P Value = numeric(),
  stringsAsFactors = FALSE
)
# Loop through pair of predictors
for (i in seq_along(predictors)) {
  for (j in seq_along(predictors)) {
    if (i < j) {
      predictor1 <- predictors[i]</pre>
      predictor2 <- predictors[j]</pre>
      # Fit interaction model
      formula <- as.formula(paste("status ~", paste(predictors, collapse = " + "),</pre>
                                   "+", predictor1, "*", predictor2))
      interaction_model <- glm(formula, family = binomial, data = model_data_3)</pre>
      # Extract interaction term
      interaction term <- paste(predictor1, predictor2, sep = ":")</pre>
      coef names <- names(coef(interaction model))</pre>
      if (interaction_term %in% coef_names) {
```

```
# Extract p-value for the interaction term
        p_value <- coef(summary(interaction_model))[interaction_term, "Pr(>|z|)"]
        # if p-value < 0.05
        if (p_value < 0.05) {</pre>
          interaction_results <- rbind(interaction_results,</pre>
                                         data.frame(Predictor1 = predictor1,
                                                    Predictor2 = predictor2,
                                                    P_Value = p_value,
                                                    stringsAsFactors = FALSE))
        }
      }
    }
  }
}
# Sort by p-value
interaction_results <- interaction_results[order(interaction_results$P_Value), ]</pre>
print(interaction_results)
stepwise_summary <- summary(stepwise_model)</pre>
stepwise_coefficients <- as.data.frame(stepwise_summary$coefficients)</pre>
# Calculate Odds Ratios and Confidence Intervals
odds ratios <- exp(stepwise coefficients[, "Estimate"])</pre>
lower_ci <- exp(stepwise_coefficients[, "Estimate"] - 1.96 * stepwise_coefficients[, "Std.</pre>
upper_ci <- exp(stepwise_coefficients[, "Estimate"] + 1.96 * stepwise_coefficients[, "Std.
stepwise_result_table <- data.frame(</pre>
  Predictor = rownames(stepwise_coefficients),
  Estimate = round(stepwise_coefficients[, "Estimate"], 3),
  Std_Error = round(stepwise_coefficients[, "Std. Error"], 3),
  Odds_Ratio = round(odds_ratios, 3),
```

```
`95% CI (Lower)` = round(lower ci, 3),
  `95% CI (Upper)` = round(upper ci, 3),
  `P-Value` = format.pval(stepwise_coefficients[, "Pr(>|z|)"], digits = 3)
)
rownames(stepwise_result_table) <- NULL</pre>
write.csv(stepwise_result_table, "tables/stepwise_model_results.csv", row.names = FALSE)
set.seed(123)
control <- trainControl(method = "cv", number = 10, classProbs = TRUE, summaryFunction = two</pre>
cv_model <- train(</pre>
  status ~ race + marital_status + x6th_stage + grade + estrogen_status +
    progesterone_status + rn_examined_log + age_log,
  data = model_data_3,
  method = "glm",
 family = "binomial",
  trControl = control,
 metric = "ROC"
)
performance_summary <- capture.output(cv_model)</pre>
writeLines(performance_summary, "results/cv_model_performance.txt")
# Split by race group
white_data <- model_data_3 %>% filter(race == "White")
minority_data <- model_data_3 %>% filter(race != "White")
black_data <- model_data_3 %>% filter(race == "Black")
other_data <- model_data_3 %>% filter(race == "Other")
# Predict for White group
pred_white <- predict(cv_model, newdata = white_data, type = "prob")[, "Dead"]</pre>
roc_white <- roc(white_data$status, pred_white)</pre>
auc_white <- auc(roc_white)</pre>
```

```
# Predict for Minority group
pred_minority <- predict(cv_model, newdata = minority_data, type = "prob")[, "Dead"]</pre>
roc_minority <- roc(minority_data$status, pred_minority)</pre>
auc minority <- auc(roc minority)</pre>
# Predict for Black group
pred_black <- predict(cv_model, newdata = black_data, type = "prob")[, "Dead"]</pre>
roc_black <- roc(black_data$status, pred_black)</pre>
auc_black <- auc(roc_black)</pre>
# Predict for Other group
pred_other <- predict(cv_model, newdata = other_data, type = "prob")[, "Dead"]</pre>
roc_other <- roc(other_data$status, pred_other)</pre>
auc_other <- auc(roc_other)</pre>
# Assign weights based on normalized inverse probabilities
model_data_4 <- model_data_3 %>%
  mutate(weight = case_when(
    race == "White" ~ 0.17,
    race == "Black" ~ 2.00,
    race == "Other" ~ 1.80
  ))
minority_data <- model_data_4 %>% filter(race != "White")
# Refit the model with weights
reweighted model <- train(</pre>
  status ~ race + marital_status + x6th_stage + grade + estrogen_status +
    progesterone_status + rn_examined_log + age_log,
  data = model data 4,
  method = "glm",
  family = "binomial",
```

```
trControl = control,
  weights = weight
)
# Evaluate performance on subgroups
pred_white_weighted <- predict(reweighted_model, newdata = white_data, type = "prob")[, "De</pre>
roc white weighted <- roc(white data$status, pred white weighted)
auc white weighted <- auc(roc white weighted)</pre>
pred_minority_weighted <- predict(reweighted_model, newdata = minority_data, type = "prob")</pre>
roc_minority_weighted <- roc(minority_data$status, pred_minority_weighted)
auc_minority_weighted <- auc(roc_minority_weighted)</pre>
pred_black_weighted <- predict(reweighted_model, newdata = black_data, type = "prob")[, "De</pre>
roc_black_weighted <- roc(black_data$status, pred_black_weighted)</pre>
auc_black_weighted <- auc(roc_black_weighted)</pre>
pred other weighted <- predict(reweighted model, newdata = other data, type = "prob")[, "De
roc other weighted <- roc(other data$status, pred other weighted)
auc_other_weighted <- auc(roc_other_weighted)</pre>
auc_comparison_summary <- data.frame(</pre>
  Model = factor(c("Original", "Reweighted"), levels = c("Original", "Reweighted")),
  White = c(auc_white, auc_white_weighted),
  Black = c(auc_black, auc_black_weighted),
  Other = c(auc_other, auc_other_weighted),
  Minority = c(auc_minority, auc_minority_weighted)
)
auc_comparison_summary[, 2:5] <- round(auc_comparison_summary[, 2:5], 4)</pre>
write.csv(auc_comparison_summary, "tables/auc_comparison_summary.csv", row.names = FALSE)
# Extract coefficients and CI from the reweighted model
```

```
summary reweighted <- summary(reweighted model$finalModel)</pre>
coefficients <- coef(summary reweighted)</pre>
# Calculate Odds Ratios and 95% CI
odds_ratios <- exp(coefficients[, "Estimate"])</pre>
lower_ci <- exp(coefficients[, "Estimate"] - 1.96 * coefficients[, "Std. Error"])</pre>
upper_ci <- exp(coefficients[, "Estimate"] + 1.96 * coefficients[, "Std. Error"])
# Combine
results_table <- data.frame(</pre>
  Predictor = rownames(coefficients),
  Estimate = round(coefficients[, "Estimate"], 3),
  Std_Error = round(coefficients[, "Std. Error"], 3),
  Odds_Ratio = round(odds_ratios, 3),
  `95% CI (Lower)` = round(lower_ci, 3),
  `95% CI (Upper)` = round(upper_ci, 3),
  `P-Value` = format.pval(coefficients[, "Pr(>|z|)"], digits = 3)
)
rownames(results table) <- NULL</pre>
write.csv(results_table, "tables/reweighted_model_results.csv", row.names = FALSE)
skim_categorical <- read.csv("tables/skim_categorical_summary.csv")</pre>
knitr::kable(skim_categorical, caption = "Skim Summary for Categorical Variables")
skim_numeric <- read.csv("tables/skim_numeric_summary.csv")</pre>
knitr::kable(skim_numeric, caption = "Skim Summary for Numeric Variables")
summary by status <- read.csv("tables/summary by status.csv")</pre>
knitr::kable(summary by status, caption = "Summary Statistics Grouped by Survival Status")
vif_table <- read.csv("tables/vif.csv")</pre>
knitr::kable(vif_table, caption = "Variance Inflation Factors for Predictors")
```

```
unstable_table <- read.csv("tables/unstable_coefficients.csv")
knitr::kable(unstable_table, caption = "Unstable Coefficients")

stepwise_result_table <- read.csv("tables/stepwise_model_results.csv")
knitr::kable(stepwise_coefficients, caption = "Stepwise Model Results")

reweighted_model_results <- read.csv("tables/reweighted_model_results.csv")
knitr::kable(reweighted_model_results, caption = "Reweighted Logistic Regression Model Results")

auc_table <- read.csv("tables/auc_comparison_summary.csv")
knitr::kable(auc_table, caption = "Comparison of ROC-AUC Values Across Models and Racial Gr</pre>
```