

# Assignment 3: Data Exploration

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## OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

## Directions

1. Rename this file `<FirstLast>_A03_DataExploration.Rmd` (replacing `<FirstLast>` with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Assign a useful **name to each code chunk** and include ample **comments** with your code.
5. Be sure to **answer the questions** in this assignment document.
6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai.

**TIP:** If your code extends past the page when knit, tidy your code by manually inserting line breaks.

**TIP:** If your code fails to knit, check that no `install.packages()` or `View()` commands exist in your code.

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## Set up your R session

1. Check your working directory, load necessary packages (tidyverse, lubridate), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX\_Neonicotinoids\_Insects\_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON\_NIWO\_Litter\_massdata\_2018-08\_raw.csv). Name these datasets “Neonics” and “Litter”, respectively. Be sure to include the subcommand to read strings in as factors.

```
library(tidyverse)
```

```
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr      1.1.3      v readr      2.1.4
## v forcats    1.0.0      v stringr   1.5.0
## v ggplot2    3.4.3      v tibble    3.2.1
## v lubridate  1.9.2      v tidyr     1.3.0
## v purrr      1.0.2
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()     masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors
```

```
library(lubridate)
```

```
Neonics <- read.csv('D:/ENV872_DataExploration/ENV872_DataExploration_Fall2023/Data/Raw/ECOTOX_Neonicot.  
Litter <- read.csv('D:/ENV872_DataExploration/ENV872_DataExploration_Fall2023/Data/Raw/NEON_NIWO_Litter
```

## Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: the specificity of these neonicotinoids is very important. If these neonicotinoids have a high specificity, other insects (that have important ecological impacts) can still be alive. Human and other mammals will also be safe. The knowing the ecotoxicology helps us to understand the specificity and protect our food and water resources for the current and future generations.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: Woody debris has important roles in carbon recycle and can provide habitat to terrestrial and aquatic creatures. Litter debris, however, can be a source of plastic pollution. These litter debris can affect soil quality, negatively influence human and other animals' health. Therefore, studying litter and woody debris is important to forest ecology and health of different creatures.

4. How is litter and woody debris sampled as part of the NEON network? Read the [NEON\\_Litterfall\\_UserGuide.pdf](#) document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. Litter and fine woody debris sampling is executed at terrestrial NEON sites that contain woody vegetation >2m tall. 2. Ground traps are sampled once per year. Target sampling frequency for elevated traps varies by vegetation present at the site, with frequent sampling (1x every 2 weeks) in deciduous forest sites during senescence, and infrequent year-round sampling (1x every 1-2 months) at evergreen sites. 3. In sites with forested tower airsheds, the litter sampling is targeted to take place in 20 40m x 40m plots.

## Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
colnames(Neonics)
```

```
## [1] "CAS.Number" "Chemical.Name"  
## [3] "Chemical.Grade" "Chemical.Analysis.Method"  
## [5] "Chemical.Purity" "Species.Scientific.Name"  
## [7] "Species.Common.Name" "Species.Group"
```

```
## [9] "Organism.Lifestage"      "Organism.Age"
## [11] "Organism.Age.Units"      "Exposure.Type"
## [13] "Media.Type"              "Test.Location"
## [15] "Number.of.Doses"         "Conc.1.Type..Author."
## [17] "Conc.1..Author."         "Conc.1.Units..Author."
## [19] "Effect"                  "Effect.Measurement"
## [21] "Endpoint"                "Response.Site"
## [23] "Observed.Duration..Days." "Observed.Duration.Units..Days."
## [25] "Author"                  "Reference.Number"
## [27] "Title"                   "Source"
## [29] "Publication.Year"        "Summary.of.Additional.Parameters"
```

*# column names refer to the dimensions of dataset*

6. Using the `summary` function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest?

```
summary(Neonics$Effect)
```

```
##      Accumulation      Avoidance      Behavior      Biochemistry
##           12           102           360           11
##      Cell(s)      Development      Enzyme(s) Feeding behavior
##           9           136           62           255
##      Genetics      Growth      Histology      Hormone(s)
##          82           38           5           1
##      Immunological      Intoxication      Morphology      Mortality
##          16           12           22           1493
##      Physiology      Population      Reproduction
##           7           1803           197
```

Answer: Population seems to be of interest as 1803 studies are population studies and mortality is the second (1493). The reason why population study is the most popular category may be that population study is the basic study for ecotoxicity. Most of the studies need to be done in population level to confirm the toxicity level before further research.

7. Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed. [TIP: The `sort()` command can sort the output of the summary command...]

```
summary(Neonics$Species.Common.Name)
```

```
##      Honey Bee      Parasitic Wasp
##           667           285
##      Buff Tailed Bumblebee      Carniolan Honey Bee
##           183           152
##      Bumble Bee      Italian Honeybee
##          140           113
##      Japanese Beetle      Asian Lady Beetle
##           94           76
##      Euonymus Scale      Wireworm
```

##	75	69
##	European Dark Bee	Minute Pirate Bug
##	66	62
##	Asian Citrus Psyllid	Parastic Wasp
##	60	58
##	Colorado Potato Beetle	Parasitoid Wasp
##	57	51
##	Erythrina Gall Wasp	Beetle Order
##	49	47
##	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
##	47	46
##	True Bug Order	Buff-tailed Bumblebee
##	45	39
##	Aphid Family	Cabbage Looper
##	38	38
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ground Beetle Family
##	29	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23
##	Ladybird Beetle	Mason Bee
##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order

##		17		17
##		Egg Parasitoid		Insect Class
##		17		17
##	Moth And Butterfly Order		Oystershell Scale Parasitoid	
##		17		17
##	Hemlock Woolly Adelgid Lady Beetle		Hemlock Woolly Adelgid	
##		16		16
##		Mite		Onion Thrip
##		16		16
##	Western Flower Thrips		Corn Earworm	
##		15		14
##	Green Peach Aphid		House Fly	
##		14		14
##	Ox Beetle		Red Scale Parasite	
##		14		14
##	Spined Soldier Bug		Armoured Scale Family	
##		14		13
##	Diamondback Moth		Eulophid Wasp	
##		13		13
##	Monarch Butterfly		Predatory Bug	
##		13		13
##	Yellow Fever Mosquito		Braconid Parasitoid	
##		13		12
##	Common Thrip		Eastern Subterranean Termite	
##		12		12
##	Jassid		Mite Order	
##		12		12
##	Pea Aphid		Pond Wolf Spider	
##		12		12
##	Spotless Ladybird Beetle		Glasshouse Potato Wasp	
##		11		10
##	Lacewing		Southern House Mosquito	
##		10		10
##	Two Spotted Lady Beetle		Ant Family	
##		10		9
##	Apple Maggot		(Other)	
##		9		670

```
sort(summary(Neonics$Species.Common.Name), decreasing = TRUE) # sort the output of species common names
```

##	(Other)		Honey Bee	
##		670		667
##	Parasitic Wasp		Buff Tailed Bumblebee	
##		285		183
##	Carniolan Honey Bee		Bumble Bee	
##		152		140
##	Italian Honeybee		Japanese Beetle	
##		113		94
##	Asian Lady Beetle		Euonymus Scale	
##		76		75
##	Wireworm		European Dark Bee	
##		69		66
##	Minute Pirate Bug		Asian Citrus Psyllid	
##		62		60

##	Parastic Wasp	Colorado Potato Beetle
##	58	57
##	Parasitoid Wasp	Erythrina Gall Wasp
##	51	49
##	Beetle Order	Snout Beetle Family, Weevil
##	47	47
##	Sevenspotted Lady Beetle	True Bug Order
##	46	45
##	Buff-tailed Bumblebee	Aphid Family
##	39	38
##	Cabbage Looper	Sweetpotato Whitefly
##	38	37
##	Braconid Wasp	Cotton Aphid
##	33	33
##	Predatory Mite	Ladybird Beetle Family
##	33	30
##	Parasitoid	Scarab Beetle
##	30	29
##	Spring Tiphia	Thrip Order
##	29	29
##	Ground Beetle Family	Rove Beetle Family
##	27	27
##	Tobacco Aphid	Chalcid Wasp
##	27	25
##	Convergent Lady Beetle	Stingless Bee
##	25	25
##	Spider/Mite Class	Tobacco Flea Beetle
##	24	24
##	Citrus Leafminer	Ladybird Beetle
##	23	23
##	Mason Bee	Mosquito
##	22	22
##	Argentine Ant	Beetle
##	21	21
##	Flatheaded Appletree Borer	Horned Oak Gall Wasp
##	20	20
##	Leaf Beetle Family	Potato Leafhopper
##	20	20
##	Tooth-necked Fungus Beetle	Codling Moth
##	20	19
##	Black-spotted Lady Beetle	Calico Scale
##	18	18
##	Fairyfly Parasitoid	Lady Beetle
##	18	18
##	Minute Parasitic Wasps	Mirid Bug
##	18	18
##	Mulberry Pyralid	Silkworm
##	18	18
##	Vedalia Beetle	Araneoid Spider Order
##	18	17
##	Bee Order	Egg Parasitoid
##	17	17
##	Insect Class	Moth And Butterfly Order
##	17	17

```
##      Oystershell Scale Parasitoid Hemlock Woolly Adelgid Lady Beetle
##                                17                                16
##      Hemlock Woolly Adelgid                                Mite
##                                16                                16
##                                Onion Thrip      Western Flower Thrips
##                                16                                15
##                                Corn Earworm      Green Peach Aphid
##                                14                                14
##                                House Fly          Ox Beetle
##                                14                                14
##                                Red Scale Parasite  Spined Soldier Bug
##                                14                                14
##                                Armoured Scale Family Diamondback Moth
##                                13                                13
##                                Eulophid Wasp      Monarch Butterfly
##                                13                                13
##                                Predatory Bug      Yellow Fever Mosquito
##                                13                                13
##                                Braconid Parasitoid Common Thrip
##                                12                                12
##      Eastern Subterranean Termite                                Jassid
##                                12                                12
##                                Mite Order          Pea Aphid
##                                12                                12
##                                Pond Wolf Spider    Spotless Ladybird Beetle
##                                12                                11
##                                Glasshouse Potato Wasp Lacewing
##                                10                                10
##                                Southern House Mosquito Two Spotted Lady Beetle
##                                10                                10
##                                Ant Family          Apple Maggot
##                                9                                9
```

Answer: Honey bees are the most popular research subject because honey bees are ecologically important making them easy to transport pollutants. They are resilient to environmental stress. References: Cunningham MM, Tran L, McKee CG, et al. Honey bees as biomonitors of environmental contaminants, pathogens, and climate change. Ecol Ind. 2022;134:108457. <https://www.sciencedirect.com/science/article/pii/S1470160X21011225>. doi: 10.1016/j.ecolind.2021.108457.

8. Concentrations are always a numeric value. What is the class of `Conc.1..Author.` column in the dataset, and why is it not numeric?

```
class(Neonics$Conc.1..Author.)
```

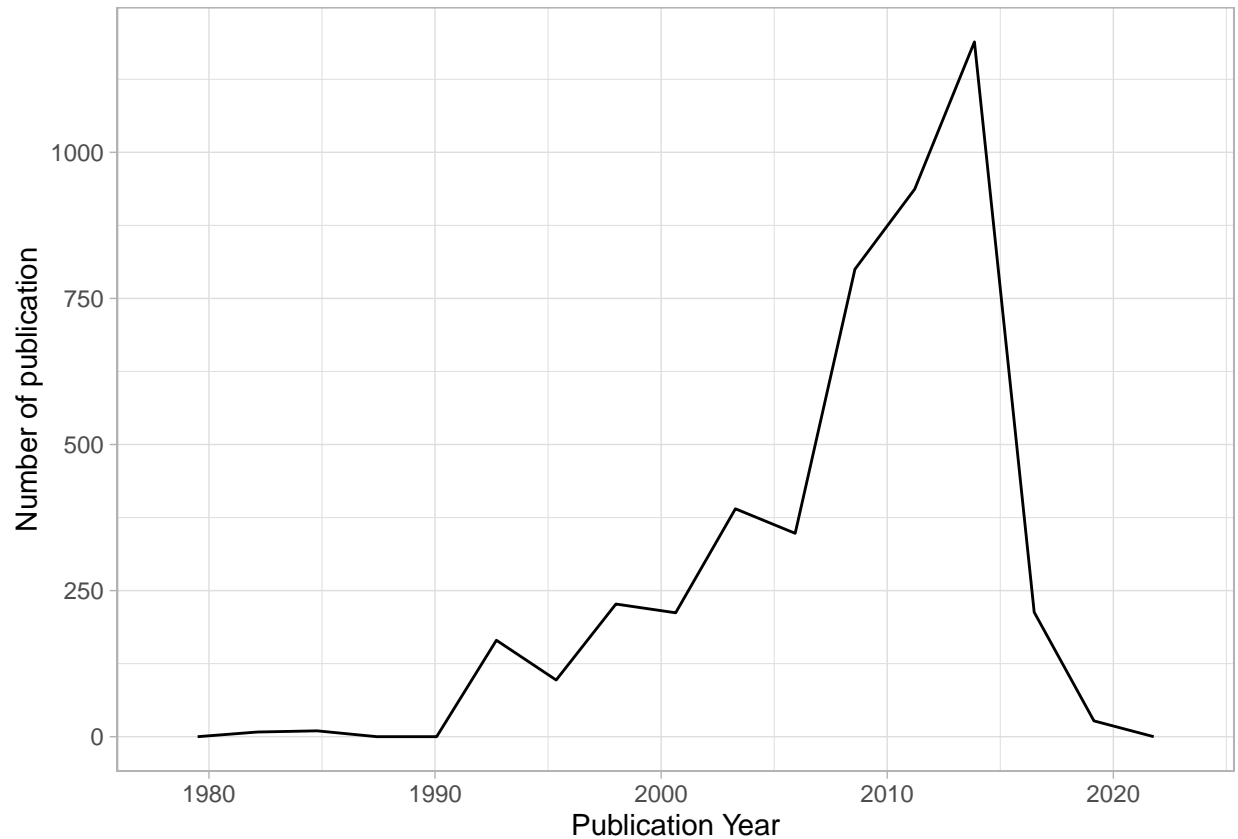
```
## [1] "factor"
```

Answer: The class of `Conc.1..Author.` column is factor because some of the values are 'NR' and some values contains '/'.

## Explore your data graphically (Neonics)

9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

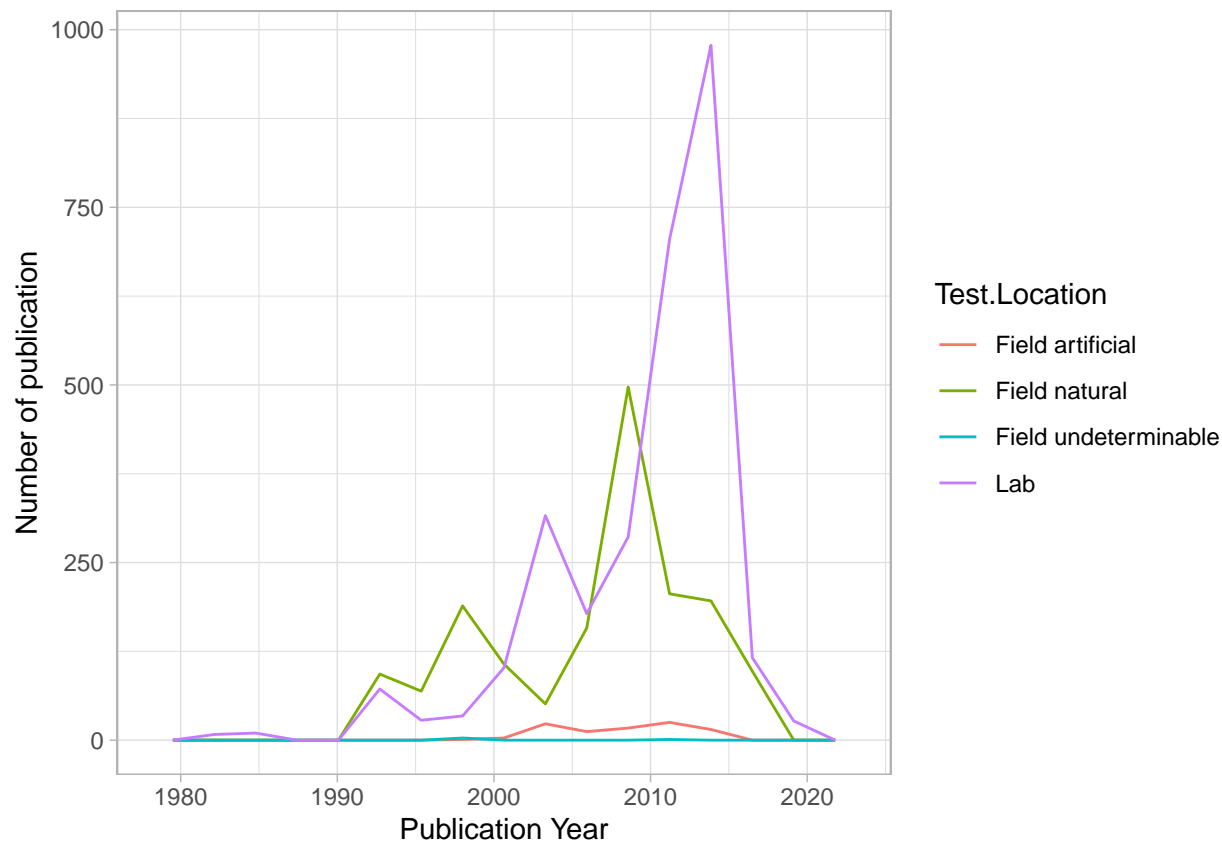
```
plot1_NeoFreq <- ggplot(Neonics) +
  geom_freqpoly(aes(x = Publication.Year, bins = 15) + theme_light() + labs(x = 'Publication Year', y =
plot1_NeoFreq
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
plot2_NeoFreqColor <- ggplot(Neonics) +
  geom_freqpoly(aes(x = Publication.Year, color = Test.Location), bins = 15) + theme_light() + labs(x =
plot2_NeoFreqColor
```





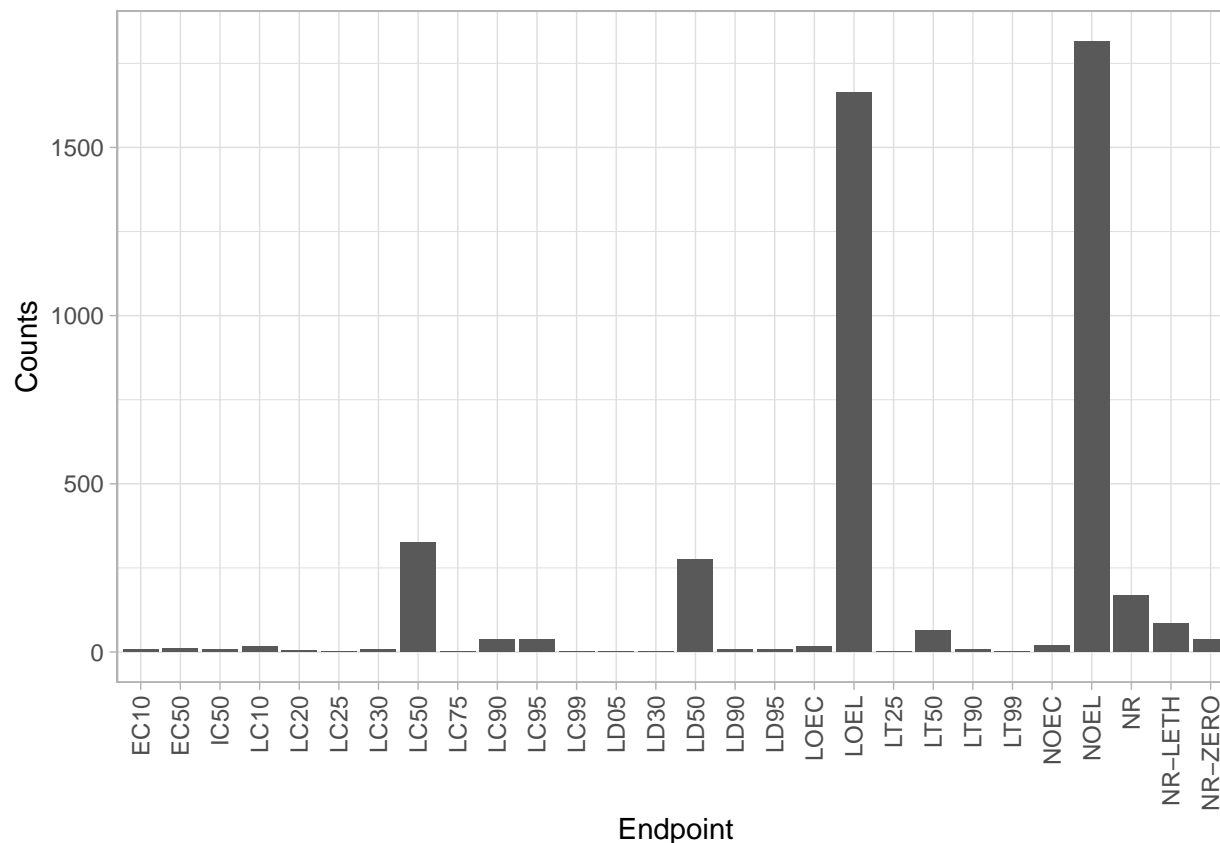
Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are lab and field natural. Lab experiments have a trend of increasing from 1980 to around 2014 and decreasing afterwards. Similarly, the number of natural field experiments increases and decreases. The peak of the publications of natural field experiment is around 2009.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX\_CodeAppendix for more information.

[**TIP:** Add `theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))` to the end of your plot command to rotate and align the X-axis labels...]

```
plot3_NeoBar <- ggplot(Neonics) +
  geom_bar(aes(x = Endpoint)) + theme_light() + theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))
plot3_NeoBar
```



Answer: NOEL and LOEL are the two most common end points. NOEL (No-observable-effect-level) and LOEL (Lowest-observable-effect-level) are common Endpoints for terrestrial database.

## Explore your data (Litter)

12. Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate) # the class of collectDate is factor
```

```
## [1] "factor"
```

```
Litter$collectDate <- as.Date(Litter$collectDate, Format = "%Y-%m-%d") # change factor into date
class(Litter$collectDate) # check the class again
```

```
## [1] "Date"
```

```
unique(Litter$collectDate) # to see which dates litter was sampled
```

```
## [1] "2018-08-02" "2018-08-30"
```

13. Using the `unique` function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

```
unique(Litter$fieldSampleID) # to see how many unique values are in the column fieldSampleID
```

```
## [1] NEON.LTR.NIW0061169.20180802 NEON.LTR.NIW0064103.20180802
## [3] NEON.LTR.NIW0067017.20180802 NEON.LTR.NIW0040205.20180802
## [5] NEON.LTR.NIW0041059.20180802 NEON.LTR.NIW0063062.20180802
## [7] NEON.LTR.NIW0047197.20180802 NEON.LTR.NIW0051045.20180802
## [9] NEON.LTR.NIW0058101.20180802 NEON.LTR.NIW0046155.20180802
## [11] NEON.LTR.NIW0062050.20180802 NEON.LTR.NIW0040205.20180830
## [13] NEON.LTR.NIW0041059.20180830 NEON.LTR.NIW0047197.20180830
## [15] NEON.LTR.NIW0051045.20180830 NEON.LTR.NIW0058101.20180830
## [17] NEON.LTR.NIW0063062.20180830 NEON.LTR.NIW0046155.20180830
## [19] NEON.LTR.NIW0062050.20180830 NEON.LTR.NIW0061169.20180830
## [21] NEON.LTR.NIW0064103.20180830 NEON.LTR.NIW0057081.20180830
## [23] NEON.LTR.NIW0067017.20180830
## 23 Levels: NEON.LTR.NIW0040205.20180802 ... NEON.LTR.NIW0067017.20180830
```

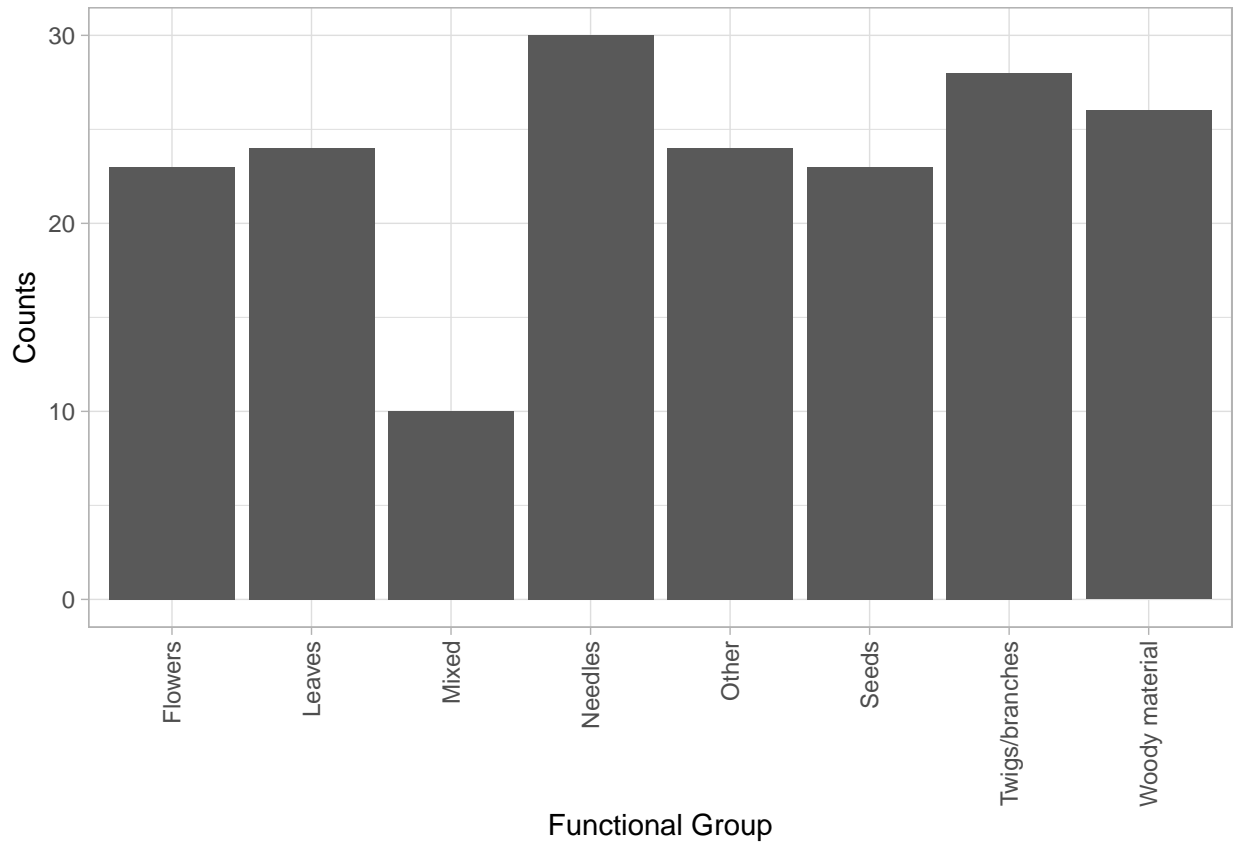
```
summary(Litter$fieldSampleID)
```

```
## NEON.LTR.NIW0040205.20180802 NEON.LTR.NIW0040205.20180830
##                               10                               10
## NEON.LTR.NIW0041059.20180802 NEON.LTR.NIW0041059.20180830
##                               8                               11
## NEON.LTR.NIW0046155.20180802 NEON.LTR.NIW0046155.20180830
##                               10                               8
## NEON.LTR.NIW0047197.20180802 NEON.LTR.NIW0047197.20180830
##                               8                               7
## NEON.LTR.NIW0051045.20180802 NEON.LTR.NIW0051045.20180830
##                               7                               7
## NEON.LTR.NIW0057081.20180830 NEON.LTR.NIW0058101.20180802
##                               8                               9
## NEON.LTR.NIW0058101.20180830 NEON.LTR.NIW0061169.20180802
##                               7                               9
## NEON.LTR.NIW0061169.20180830 NEON.LTR.NIW0062050.20180802
##                               8                               7
## NEON.LTR.NIW0062050.20180830 NEON.LTR.NIW0063062.20180802
##                               7                               7
## NEON.LTR.NIW0063062.20180830 NEON.LTR.NIW0064103.20180802
##                               7                               8
## NEON.LTR.NIW0064103.20180830 NEON.LTR.NIW0067017.20180802
##                               8                               8
## NEON.LTR.NIW0067017.20180830
##                               9
```

Answer: Total of 23 plots were sampled at Niwot Ridge. Summary() returns the sample sites but also how many results are in the same site while unique() only returns the name of each unique sample site.

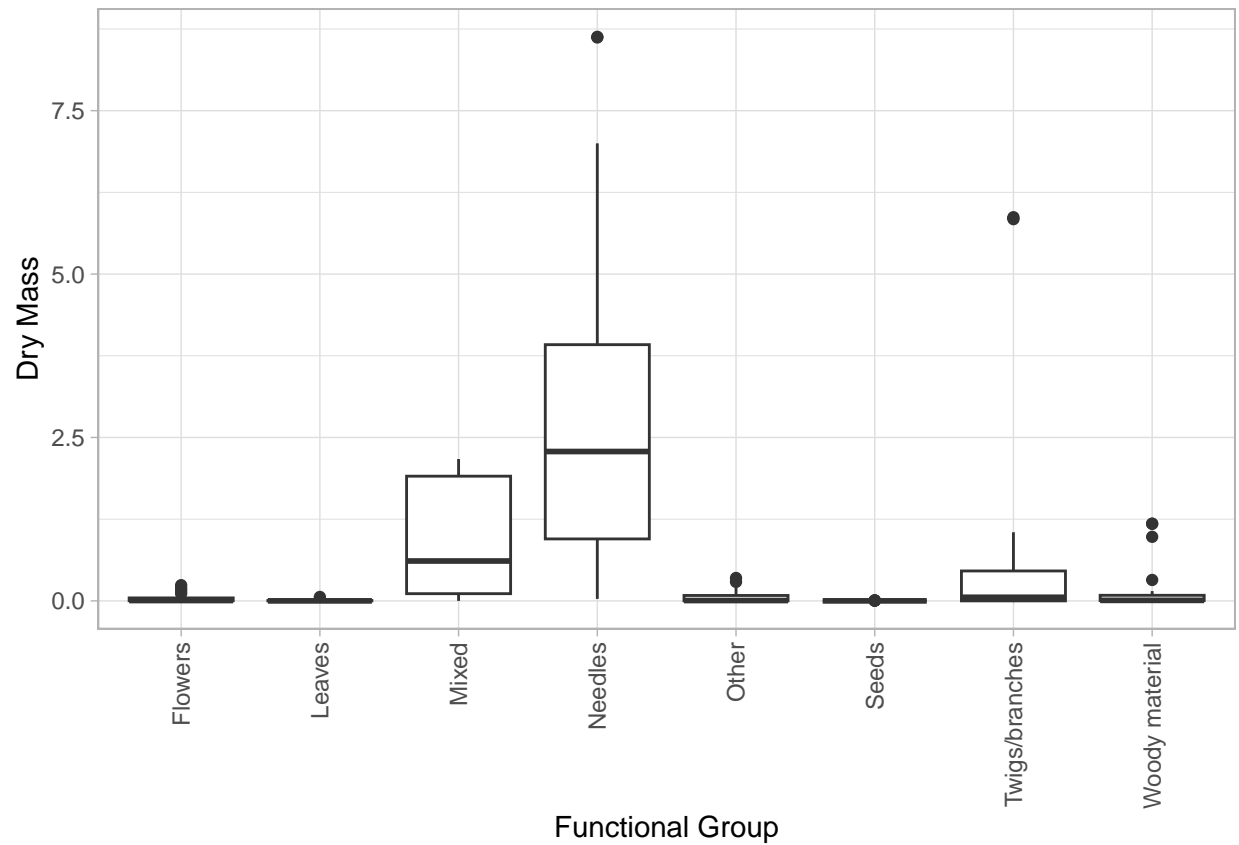
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
plot4_litterBar <- ggplot(Litter) +
  geom_bar(aes(x = functionalGroup)) + theme_light() + theme(axis.text.x = element_text(angle = 90, vjust = 0.5))
plot4_litterBar
```

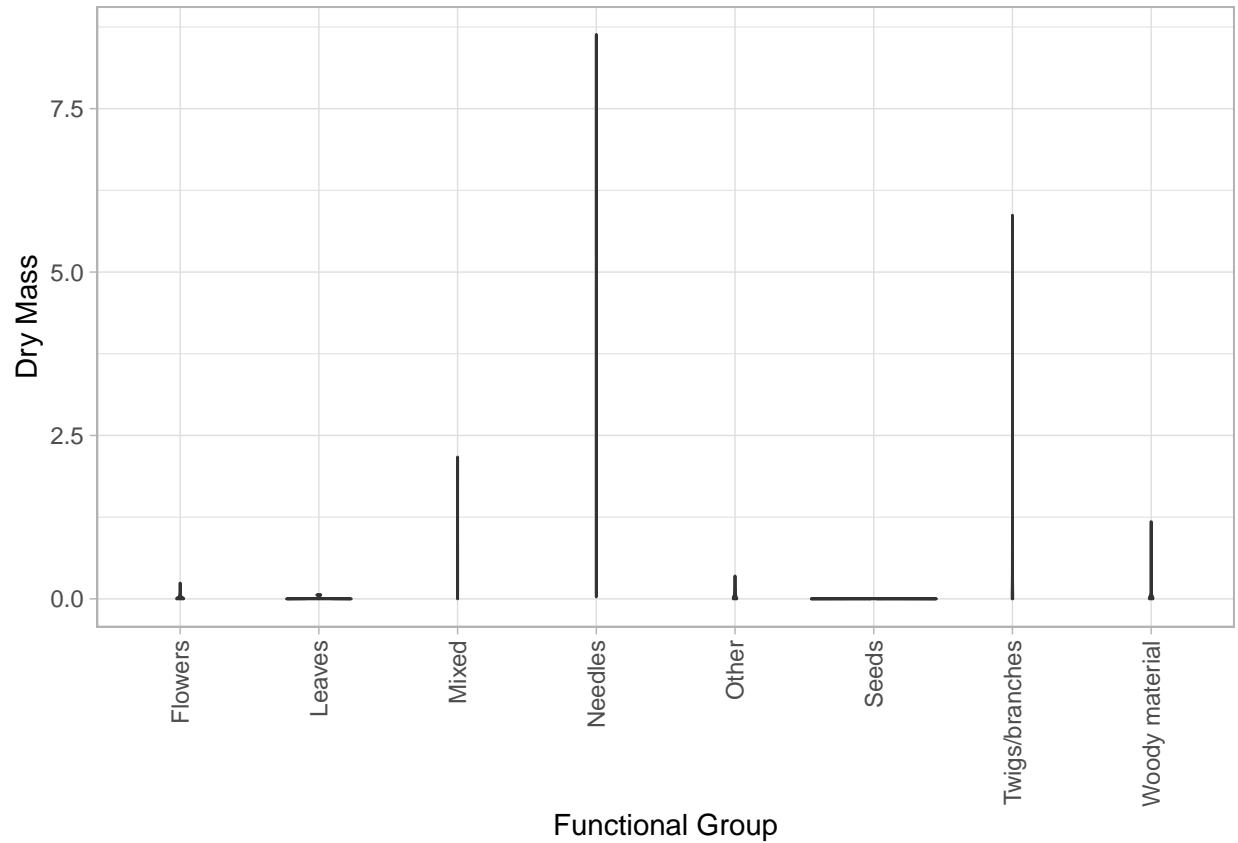


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by functional-Group.

```
# box plot
plot5_litterBox <- ggplot(Litter) +
  geom_boxplot(aes(x = functionalGroup, y = dryMass)) + theme_light() + theme(axis.text.x = element_text(angle = 90, vjust = 0.5))
plot5_litterBox
```



```
# violin plot
plot6_litterVio <- ggplot(Litter) +
  geom_violin(aes(x = functionalGroup, y = dryMass)) + theme_light() + theme(axis.text.x = element_text(
plot6_litterVio
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: Violin plots may not work well when the sample size is small or there are multiple peaks rather than having a unimodal distribution. Therefore, in our case, the boxplot can be a more effective option which can also clearly show outliers.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles