

ClusterProfiler-LLM: NSCLC Case Study

Automated Interpretation of Nature Genetics (2025) Data

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1 Introduction

This document demonstrates the capability of `clusterProfiler-LLM` to reproduce and interpret key findings from a high-complexity single-cell study. We analyze the dataset from *Nature Genetics* (2025): “Multi-omic profiling highlights factors associated with resistance to immuno-chemotherapy in non-small-cell lung cancer”.

We compare the automated output of our multi-agent framework against the manual conclusions drawn by the original authors, specifically focusing on:

- 1. **Mechanism Interpretation:** Tumor cell states (e.g., NRF2-mediated stress response).
- 2. **Cell Type Annotation:** Identification of specific subsets (e.g., SPP1+ Macrophages).
- 3. **Phenotypic Characterization:** Immune states (e.g., TIGIT/CTLA4-driven exhaustion).

Note

Online HTML version: https://yulab-smu.top/clusterProfiler-LLM-Supplementary/NSCLC_Case_Study.html

2 Materials and Methods

2.1 Data Source

- **Study:** Yan et al., *Nature Genetics*, 2025.
- **Context:** NSCLC treated with neoadjuvant ICB + chemotherapy.
- **Key Original Findings:**
 - **SPP1+ Macrophages:** Interact with COL11A1+ CAFs to form a physical barrier.
 - **Tumor Cells:** “Basal/Stress” state characterized by NRF2 pathway activation.
 - **T Cells:** Distinct exhaustion trajectories vs. activation.

2.2 Setup

```
library(clusterProfiler)
library(dplyr)
```

This Quarto document renders from pre-computed objects (RDS) to avoid re-running LLM calls during build. To reproduce the results end-to-end, you will need the development version of `clusterProfiler` that provides `interpret()`, `interpret_agent()`, and `interpret_hierarchical()`, and an LLM backend configured via `fanyi`.

```
library(fanyi)

api_key <- Sys.getenv("DEEPSEEK_API_KEY")
set_translate_option(source = "dsk", key = api_key)
```

3 Analysis Workflow

3.1 1. Data Preparation

We load the pre-processed marker genes and metadata.

The case study is distributed with the following inputs in the same directory as this QMD:

File	What it contains	Used for
<code>scobj.markers.rds</code>	FindAllMarkers output (per-cluster marker genes)	Selecting top markers for enrichment
<code>md.rds</code>	Cell-level metadata from the Seurat object	Building context and priors
<code>x.rds</code>	Cell-marker enrichment (<code>compareCluster(..., fun=enricher)</code>)	LLM interpretation input
<code>anno.rds</code> , <code>pheno.rds</code> , <code>deep.rds</code> , <code>hier.rds</code>	Pre-computed LLM outputs	Rendering results without live API calls

The `md.rds` file is a cell-level metadata table (i.e., `scobj@meta.data`) saved separately so this case study does not need to bundle the full Seurat object. It is expected to contain, at minimum:

- Study/clinical fields used to summarize dataset context: `PathType`, `Timepoint`, `PathRes`, `Drug`, `Group`
- Cluster ID used to aggregate priors: `seurat_clusters`
- A coarse lineage/cell class label used as prior knowledge: `all_cluster_annotation`

```
scobj.markers <- readRDS("./scobj.markers.rds")

# Filter for top 10 markers per cluster
scobj.markers %>%
  group_by(cluster) %>%
  dplyr::filter(avg_log2FC > 1) %>%
  slice_head(n = 10) %>%
  ungroup() -> top10

md <- readRDS("./md.rds")
```

3.2 2. Context Construction

We construct a prompt context based on the study design.

```
majority <- function(x) names(sort(table(x), decreasing = TRUE))[1]

ctx <- paste0(
  "Study setting: NSCLC single-cell RNA-seq in the context of ",
  "neoadjuvant ICB + chemotherapy. ",
  "This dataset includes paired/longitudinal sampling ",
  "(pre- vs post-treatment) and response stratification by pathology. ",
  "Current object summary: ",
  "PathType=", majority(md$PathType), "; ",
  "Timepoint=", majority(md$Timepoint), "; ",
  "PathologicResponse=", majority(md$PathRes), "; ",
  "Response labels include NMPR/MPR/pCR. ",
```

```

"Drug=", majority(md$Drug), "; ",
"Group=", majority(md$Group), ". ",
"Goal: annotate clusters/cell states and interpret ",
"marker-enrichment results in this context."
)

```

We also construct two optional inputs used by the interpretation functions:

1. A prior vector (`prior_vec`) derived from the dataset's coarse lineage annotation (`all_cluster_annotation`).
2. A per-gene fold-change lookup (`gene_fold_change`) derived from marker statistics.

```

prior_tbl <- md %>%
  mutate(cluster = .data[["seurat_clusters"]]) %>%
  group_by(cluster) %>%
  summarise(prior = majority(all_cluster_annotation), .groups = "drop")
prior_vec <- setNames(prior_tbl$prior, as.character(prior_tbl$cluster))

fc_col <- if ("avg_log2FC" %in% names(top10)) "avg_log2FC" else "avg_logFC"
gene_fc <- top10 %>%
  group_by(gene) %>%
  summarise(fc = max(.data[[fc_col]], na.rm = TRUE), .groups = "drop")
gene_fold_change <- setNames(gene_fc$fc, gene_fc$gene)

```

3.3 3. Standard Enrichment Analysis

```

x <- readRDS("./x.rds")

```

The enrichment objects were generated from `top10` using `compareCluster`. Cell-marker enrichment used an external TERM2GENE table (`Cell_marker_Human.xlsx`), which is not bundled in this directory.

```

cm <- readxl::read_excel("./Cell_marker_Human.xlsx")

x <- compareCluster(
  gene ~ cluster,
  data = top10,
  fun = enricher,
  TERM2GENE = cm[, c("cell_name", "marker")]
)
saveRDS(x, "./x.rds")

```

4 Results: LLM-Driven Interpretation

4.1 Task 1: Cell Type Annotation (Hypothesis Verification)

We use `interpret(task = "celltype")` with prior knowledge injection.

```
library(fanyi)

api_key <- Sys.getenv("DEEPSEEK_API_KEY")
set_translate_option(source = "dsk", key = api_key)

anno <- interpret(
  x = x,
  context = ctx,
  task = "celltype",
  prior = prior_vec,
  n_pathways = 20,
  add_ppi = FALSE,
  gene_fold_change = gene_fold_change,
  model = "deepseek-chat",
  api_key = api_key
)
saveRDS(anno, "./anno.rds")
```

Result:

Full results (online): https://yulab-smu.top/clusterProfiler-LLM-Supplementary/Task1_Cell-Type_Annotation.md

Comparison with *Nature Genetics* (2025):

Biological Feature	clusterProfiler-LLM Interpretation	Nature Genetics (Original Findings)	Key Match?	Significance
Macrophage Identity	Identified SPI1+ M2-like Macrophages with immunosuppressive signatures (<i>C1QA/B/C</i> , <i>CD163</i>).	Described SPP1+ TAMs as a major immunosuppressive population interacting with CAFs.	Yes (M2/TAM overlap)	Major Resistance Mechanism

Biological Feature	clusterProfiler-LLM Interpretation	Nature Genetics (Original Findings)	Key Match?	Significance
Tumor Cell State	Detected Basal-like Progenitor state with <i>GPX2</i> and <i>AKR1C1</i> (Oxidative Stress).	Defined “Basal/Stress” tumor cell state characterized by NRF2 activation.	Direct (Stress/Basal)	Poor Prognosis Driver
Stromal Niche	Annotated Activated Myofibroblasts (<i>ACTA2</i> , <i>COL1A2</i>) and Vascular Progenitors.	Detailed COL11A1+ CAFs forming a barrier at tumor boundaries.	Yes (CAF Activation)	Immune Exclusion
T Cell States	Found IL7R+ Memory T cells and Exhausted subsets.	Highlighted dysfunctional CD8+ T cells and potential for revival.	Yes	Therapy Response Determinant

4.2 Task 2: Phenotypic Characterization

We use `interpret(task = "phenotype")` to define the functional state of each cluster.

```
library(fanyi)

api_key <- Sys.getenv("DEEPSEEK_API_KEY")
set_translate_option(source = "dsk", key = api_key)

pheno <- interpret(
  x = x,
  context = ctx,
  task = "phenotype",
  n_pathways = 30,
  add_ppi = FALSE,
  gene_fold_change = gene_fold_change,
  model = "deepseek-chat",
  api_key = api_key
)
saveRDS(pheno, "./pheno.rds")
```

Result:

Full results (online): https://yulab-smu.top/clusterProfiler-LLM-Supplementary/Task2_Phenotypic_Characterization.md

Comparison with *Nature Genetics* (2025):

Biological Feature	clusterProfiler-LLM Interpretation	Nature Genetics (Original Findings)	Key Match?	Significance
Tumor Cell Plasticity	Identified Basal-like Progenitor state expressing <i>AKR1C1</i> , <i>GPX2</i> (Detoxification/Stress).	Defined “Basal/Stress” state driven by NRF2 (<i>NFE2L2</i>) and oxidative stress response.	Direct (AKR1C1/Stress)	Chemoresistance Mechanism
Fibroblast Activation	Characterized Activated Myofibroblasts with high <i>COL1A1</i> , <i>ACTA2</i> , <i>TGFB1</i> .	Described COL11A1+ CAFs forming immune-excluded niches.	Yes (TGF-()) (CAFs)	Immune Exclusion Barrier
Myeloid Polarization	Detected Immunosuppressive Macrophages (<i>C1QA</i> , <i>APOE</i>) with M2-like features.	Highlighted SPP1+ TAMs as the dominant immunosuppressive myeloid population.	Yes (M2/TAMs)	T-cell Suppression
B Cell Function	Noted Germinal Center B cells (<i>BCL6</i> , <i>AICDA</i>) indicating humoral immunity.	Discussed TLS (Tertiary Lymphoid Structure) presence and B cell maturity.	Consistent	Anti-tumor Immunity Potential

4.3 Task 3: Mechanism Interpretation (Multi-Agent Deep Mode)

We use `interpret_agent()` to reconstruct causal networks, integrating PPI and LogFC data.

```
library(fanyi)

api_key <- Sys.getenv("DEEPSEEK_API_KEY")
set_translate_option(source = "dsk", key = api_key)

deep <- interpret_agent(
  x = x,
  context = ctx,
```

```

n_pathways = 50,
add_ppi = TRUE,
gene_fold_change = gene_fold_change,
model = "deepseek-chat",
api_key = api_key
)
saveRDS(deep, "./deep.rds")

```

Result:

Full results (online): https://yulab-smu.top/clusterProfiler-LLM-Supplementary/Task3_Mechanism_Interpretation.md

Comparison with *Nature Genetics* (2025):

Biological Feature	clusterProfiler-LLM Interpretation	Nature Genetics (Original Findings)	Key Match?	Significance
T Cell Exhaustion	TOX identified as a key regulator of Exhausted CD8+ T cells (<i>PDCD1</i> , <i>HAVCR2</i>).	Dysfunctional CD8+ T cells are a major feature of non-responders; exhaustion is a key barrier.	Direct (TOX/Exhaustion)	Checkpoint Blockade Target
Myeloid Regulation	SPI1 (PU.1) inferred as the master regulator for M2-like TAMs (<i>C1QA</i> , <i>MRC1</i>).	SPP1+ TAMs recruit regulatory T cells and suppress adaptive immunity.	Yes (Myeloid Driver)	Immunosuppressive Hub
B Cell Identity	PAX5 and BCL6 regulatory network defines Follicular/GC B cells .	Presence of mature B cells in TLS correlates with better prognosis.	Consistent	Prognostic Marker
Tumor Proliferation	E2F targets and MYC signaling active in Cycling Tumor Cells .	High proliferation rates in specific tumor subclones drive progression.	Yes	Tumor Aggressiveness

4.4 Task 4: Hierarchical Interpretation

Refining annotations from Major lineage to Minor states using `interpret_hierarchical()`.


```

library(fanyi)

api_key <- Sys.getenv("DEEPSEEK_API_KEY")
set_translate_option(source = "dsk", key = api_key)

map_tbl <- md %>%
  mutate(minor = as.character(minor), major = as.character(major)) %>%
  count(minor, major, name = "n") %>%
  group_by(minor) %>%
  slice_max(n, n = 1, with_ties = FALSE) %>%
  ungroup()
mapping <- setNames(map_tbl$major, map_tbl$minor)

top10_minor <- readRDS("./top10_minor.rds")
top10_major <- readRDS("./top10_major.rds")

cm <- readxl::read_xlsx("./Cell_marker_Human.xlsx")

x_minor <- compareCluster(
  gene ~ cluster,
  data = top10_minor,
  fun = enricher,
  TERM2GENE = cm[, c("cell_name", "marker")]
)
saveRDS(x_minor, "./x_minor.rds")

x_major <- compareCluster(
  gene ~ cluster,
  data = top10_major,
  fun = enricher,
  TERM2GENE = cm[, c("cell_name", "marker")]
)
saveRDS(x_major, "./x_major.rds")

hier <- interpret_hierarchical(
  x_minor = x_minor,
  x_major = x_major,
  mapping = mapping,
  model = "deepseek-chat",
  api_key = api_key,
  task = "cell_type"
)
saveRDS(hier, "./hier.rds")

```

Result:

Full results (online): https://yulab-smu.top/clusterProfiler-LLM-Supplementary/Task4_Hierarchical_Interpretation.md

Comparison with *Nature Genetics* (2025):

Biological Feature	clusterProfiler-LLM Interpretation	Nature Genetics (Original Findings)	Key Match?	Significance
CD8+ T Cell Subsets	Distinctly separated Naïve , Memory (<i>IL7R</i>), and Exhausted (<i>TOX</i> , <i>LAG3</i>) T cells.	Emphasized the spectrum from Pre-dysfunctional to Dysfunctional CD8+ T cells.	Precise	Therapy Response Continuum
Macrophage Heterogeneity	Hierarchically resolved Alveolar Macrophages vs. Tumor-Associated Macrophages (<i>C1QA</i>).	Distinguished resident macrophages from tumor-infiltrating SPP1+ TAMs .	Yes	Origin Matters (Tissue vs. Tumor)
Tumor Heterogeneity	Sub-classified tumor cells into Cycling (<i>MKI67</i>) and Stress/Basal (<i>AKR1C1</i>) states.	Mapped tumor macro-clusters to distinct Cellular States (Cycling, Stress, Interferon-high).	Direct	Intratumoral Heterogeneity
Endothelial States	Identified Tip cells and Stalk cells indicative of angiogenesis.	Noted PLVAP+ Endothelial cells associated with tumor vascularization.	Consistent	Angiogenesis Targets

5 Conclusion

The clusterProfiler-LLM framework successfully reproduced the key biological insights of the 2025 *Nature Genetics* study without manual curation. By automating the interpretation of mechanism, cell identity, and phenotype, it accelerates the transition from data to knowledge.