

scVMAP: a comprehensive platform for integrating single-cell chromatin accessibility regions with causal variants

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Abstract

Integrating causal variant effects with single-cell assay for transposase-accessible chromatin with high-throughput sequencing (scATAC-seq) enables a more effective elucidation of the roles and impacts of genetic variations at the single-cell level. With the accumulation of genome-wide association studies (GWAS) and single-cell genomic data, there is an urgent need for comprehensive analysis and efficient exploration of these data to uncover the underlying biological processes. To address this, we developed scVMAP (<https://bio.liclab.net/scvmap/>), a user-friendly database, which aims to provide trait-relevant cell populations at single-cell resolution. The current version of scVMAP integrated ~180 scATAC-seq datasets and ~15,800 fine-mapping results, generating 15.95 billion trait-cell pairs, offering valuable resources for exploring the functional localization of single-cell variations. To enhance the understanding of how phenotypic associations are mapped to single-cell data, scVMAP provides a wealth of detailed information, including trait-relevance scores (TRSs) for each cell, cell-type-specific differential gene and transcription factor (TF) activities, trait-relevant gene and TF interactions, and regulatory networks linking traits to cell types. Based on these comprehensive analysis results, scVMAP offers users convenient interfaces to search, browse, analyse, and visualize relationships between traits and cell populations at single-cell resolution.

Introduction

Genome-wide association studies (GWAS) provide a broad foundation for the study of genetic variant mechanisms underlying human complex phenotypes or diseases, such as the manifestation of genetic factors in various phenotypes (1-6), and risk prediction for various complex diseases (3,7-15). Given that the functional effects of genetic variants vary across tissues and cell types, identifying associations between genetic variants and cell types is of great significance for in-depth understanding of disease mechanisms and the development of targeted therapeutic strategies (4,16-20). Single-cell sequencing technologies offer unprecedented opportunities to identify cellular heterogeneity within complex diseases. To date, several databases integrating single-cell data with GWAS summary statistics have emerged, such as sc2GWAS and SC2disease (21,22). However, existing resources and methods primarily focus on single-cell RNA sequencing (scRNA-seq) data. Notably, it has been confirmed that most of the identified disease-relevant genetic variants are located outside the exome (1,23). Approximately 80-90% of the relevant variant sites are found in non-coding regions (24-26). Therefore, in relevant traits or diseases, the activity of regulatory elements often overlaps with causal genetic variant sites to identify the relevant cell types, i.e., enrichment in functional cell types (24,27,28). In practice, a major limitation is the computational burden imposed by a large number of significant variants due to the linkage disequilibrium relationship between effect variants (1,6,27,29). Therefore, the method that assumes the presence of a causal variant effect among multiple linked variant sites is most commonly used, which facilitates large-scale data analysis (5,27,29). In the context of the above observations, genetic fine-mapping can effectively address the problem and help to elucidate the role and impact of genetic variation at the cellular level of association (25,30).

Currently, the advancement of single-cell genomics has given rise to scATAC-seq technology (31), facilitating the exploration of the functional roles of genetic variants in non-coding regions and at the single-cell level (32). The integration of causal variant effects with scATAC-seq enables the precise identification of genetic variants that play pivotal roles in specific cell types (25). However, compared to scRNA-seq, scATAC-seq data commonly exhibits high sparsity and technical noise (33), significantly increasing the difficulty of performing cell-type-specific genetic inference based on such data. To address this challenge, recently developed innovative methods, g-chromVAR (27) and SCAVENGE (25), effectively integrate causal variant effects with scATAC-seq maps using co-localization techniques, successfully identifying cell types significantly implicated in specific traits. The SCAVENGE method, building upon g-chromVAR, incorporates network propagation to mitigate the sparsity of single-cell data, thereby enhancing analytical performance. These pioneering methods systematically infer the specific cellular contexts in which phenotype-relevant variants exert their effects, laying a solid foundation for elucidating the causal mechanisms of phenotypic variation. However, challenges such as lengthy computation times and dependence on high-performance computing in large-scale acquisition and exploration of genetic variation at the single-cell level have hindered convenient utilization for researchers. Simultaneously, the rapid accumulation of large-scale scATAC-seq

data and causal variant data has created an urgent need for centralized integration, allowing researchers to effectively utilize these resources to explore the functional roles and impacts of genetic association loci at single-cell resolution.

Here, we developed scVMAP, a platform for documenting variant-to-function mapping relationships at single-cell resolution (Figure 1). It aims to accommodate an extremely vast number of trait-relevant cell pairs at the single-cell level by integrating large-scale scATAC-seq data with complex traits. scVMAP provides comprehensive annotations, gene pathway enrichment analysis, and context-specific regulatory networks for these associated pairs. The current version of scVMAP accommodates over 1,342,000 single cells (derived from ~180 manually collected scATAC-seq samples) and over 15,800 traits (filtered based on fine-mapping results). By applying g-chromVAR and SCAVENGE, the database has established over 15.9 billion trait-cell relationships. The fine-mapping results encompass a comprehensive disease spectrum, including all 22 major categories and over 250 subcategories of the International Classification of Diseases (ICD-10) (34). Moreover, the dataset spans comprehensive phenotypic information, covering indicators, drugs and chemical compounds, etc. To ensure the reliability of results, we processed scATAC-seq profiles and genetic fine-mapping results independently via standardized pipelines, using consistent parameters tailored to each data type. To empower researchers to access the landscape underlying potential enrichment associations between phenotypes or diseases and cell types, we provide detailed and abundant regulation information, including TRSs for cells, cell-type-specific differential gene and TF activities, trait-relevant genes and TFs, and systematically constructed trait-cell type association networks. In conclusion, scVMAP is a user-centric database that facilitates the exploration and interpretation of single-cell genetic data through structured displays and dynamic visualizations. It offers intuitive workflows, customizable parameters, and comprehensive data accessibility.

Material and methods

Collection and processing of fine-mapping data

We collected fine-mapping results of 16,445 traits from four major resources: CAUSALdb2 (35), FinnGen (36,37), UKBB (37,38) and BBJ (37,39) (Figure 1). In order to ensure the reliability of the fine-mapping data in subsequent integrated analyses, we performed uniform preprocessing of the data from different cohorts. For each trait, we retained the variants with a causal variant probability value (PP) > 0.001 calculated by FINEMAP (25,40). As a result, we retained 15,805 traits from the initial 16,445 traits and used them for subsequent analysis. To harmonize genomic coordinates between variants and scATAC-seq data, we performed LiftOver (41) to convert variant positions to match the reference genome version used in single-cell analysis. Next, we manually categorized them into a broad array of classifications, including diseases, indicators, drugs, chemical compounds, health care, treatments, therapies and symptoms. For the disease category, we annotated diseases according to ICD-10, encompassing all 22 major

disease categories, and further subclassifying them into more than 250 specific subcategories to provide an intuitive and convenient reference.

Collection and processing of single-cell chromatin accessibility datasets

We obtained ~180 scATAC-seq datasets with cell type labels from multiple sources: scATAC-Ref (42), scBlood (43), GreenleafLab laboratory (<https://github.com/GreenleafLab/MPAL-Single-Cell-2019>) (25) and the PlaqView single-cell data portal (<https://www.plaqview.com/>) (44) (Figure 1). These data involve approximately 20 tissue types including blood, brain, skin, heart, kidney, and liver, encompassing around 200 cell types such as lymphocytes, cardiomyocytes, fibroblasts, and tumor cells, and covering more than 1,342,000 cells in total. To ensure data standardization, we used the SnapATAC2 (45) software for unified preprocessing and quality control, including feature selection and doublets identification. Next, we used SnapATAC2 to derive cellular gene activity profiles and identify differential TFs across cell types. To identify differentially active genes among cell types, we employed SCANPY (46), performing the differential activity analysis using the Wilcoxon test. For each cell type in each sample, we conducted gene set enrichment analysis using GSEAPy (47) on the top 500 genes ranked by differential gene scores.

Estimation of trait relevance score for each single cell

Identifying causal variants relevant cell types underlying the observed associations will provide important reference resources for understanding the complex phenotypic manifestations and disease pathogenesis. Therefore, building upon the collection and preprocess of fine-mapping data across 15,805 traits and scATAC-seq datasets from ~180 samples, we next linked key genetic variants to cellular functions, which will help to discover the cell-specific mechanisms behind traits. Specifically, we systematically link genetic variants to their functional cellular contexts at single-cell resolution using g-chromVAR and SCAVENGE methods (25,27), with peak-by-cell matrix generated from scATAC-seq data and fine-mapping results of traits as inputs. For each trait, we compute bias-corrected Z-scores of per cell using g-chromVAR by integrating variant posterior probabilities and chromatin accessibility at linked peaks (27). SCAVENGE was applied with default parameters to calculate TRSs for each cell. Briefly, cells ranked high based on g-chromVAR Z-scores were selected as seed cells. A mutual k-nearest neighbor (M-kNN) graph was constructed based on latent semantic indexing (LSI) features of the scATAC-seq data, and network propagation was performed to propagate seed cell information, generating TRSs that reflect each cell's relevance to the trait (25). As a result, with ~180 scATAC-seq samples and 15,805 traits, a total of ~2,844,900 trait-sample associations were generated, where each trait is linked to every cell within each sample. Each association is annotated with both g-chromVAR Z-scores and SCAVENGE TRSs to quantify trait relevance at single-cell resolution. A small fraction of these results shows entirely zero TRS values across all cells in specific samples for certain traits, indicating no detectable enrichment of the corresponding trait in those samples. In contrast, a total of over 1,923,500 trait-sample

associations exhibited non-zero TRS values in at least one cell per sample, demonstrating detectable trait enrichment. Among these, there are over 13.06 million associations linking traits to distinct cell types within specific samples and more than 15.95 billion trait/disease-cell associations at the single-cell level.

Trait-relevant genes and TFs

To systematically study causal variant effects, we provide comprehensive trait-related analysis resources: (1) Gene-based analysis: We performed gene-based association analysis using MAGMA (48) to help investigate the potential effects of causal variants on associated genes. In brief, we first use Entrez eUtils to match variant sites lacking rsID annotations with dbSNP to ensure the integrity of MAGMA enrichment (49). Then, variant loci were mapped to genes using MAGMA's annotation function based on GENCODE v41 (50), ensuring consistency with the single-cell data processing pipeline. Gene-level association analysis was performed using MAGMA, incorporating LD structure from the 1000 Genomes Project to account for linkage disequilibrium among variants. (2) Functional enrichment analysis: To characterize trait-associated functions, we performed gene set enrichment analysis using GSEApy (47) across Gene Ontology (BP/CC/MF) (51), KEGG pathways (52), and GWAS Catalog (53) annotations. (3) Motif analysis: We performed motif enrichment analysis using HOMER (54) with the parameter *-size* 1000 to assess variant impact on TF binding affinity.

Construction of the trait-cell type association network

To help understand how traits affect biological functions and disease susceptibility through specific cell types, we constructed trait-cell type association network linking by gene and TF, referred to as "Gene hub network" and "TF hub network" respectively. The both networks integrate data from two dimensions genetic variation and cell type specificity. (1) Gene hub network: The genetic variation dimension refers to the variant-relevant genes obtained through MAGMA analysis (48), which represent the variant-gene associations network. Since a trait or disease have recorded numerous susceptibility causal variant loci, this constitutes the trait-variant association network. The cell type specificity dimension is derived from cell type-specific differentially active genes in single-cell samples, forming the gene-cell type association network. These two dimensions of data are used to construct a global regulatory landscape of the trait-variant-gene-cell type-single cell sample network. The key genes, obtained by overlapping the genes contained in two dimensions, represent candidate marker genes that play enriched functional roles and effects in specific cell types for a trait or disease. (2) TF hub network: The genetic variation dimension refers to the trait- or disease-relevant TFs obtained through HOMER analysis (54), forming the trait-TF association network. From the cell type-specific dimension, we obtained the TF-cell type association network. From these two-dimensional data, we construct a trait-TF-cell type-single cell sample network regulatory landscape. The key TFs were obtained by overlapping the TFs included in the two dimensions.

Gene annotation

To further explore the manifestations of complex phenotypes and the underlying mechanisms of diseases, scVMAP provides more (epi) genetic annotation data for variant-relevant genes. We curated genomic regulatory elements including 26.75 million typical enhancers (TEs) and 1.33 million super-enhancers (SEs). Specifically, TEs were compiled from SEA v3 (55) and SEdb v2 (56), while SEs were aggregated from SEA v3, dbSUPER (57), and SEdb v2. We also incorporated over 37.3 million common SNPs from dbSNP b151 (49) and over 96,900 risk SNPs from GWAS ATLAS (6). These annotations were mapped to variant-relevant genes using BEDTools (58) software. Additionally, we further annotated genes with 101.5 million expression quantitative trait loci (eQTLs) sourced from GTEx v10 (59), directly linking genetic variants to genes.

Database implementation

The latest version of the scVMAP database operates on CentOS 7.7.1908. We deploy and release the front-end and back-end projects to the remote server separately using Dockerfile in the integrated development environments WebStorm 2025.1.1.1 and IntelliJ IDEA 2025.1.1.1. The remote server has pre-installed [Docker](#) 19.03.5, and the back-end API along with the front-end pages are reverse-proxied through [Nginx](#) 1.22.0. The project employs a technical architecture that separates the front-end from the back-end. On the back-end side, business logic processing is built upon the [Spring Boot](#) 3.0.5 framework, which is based on [Java](#) 17.0.1. For database management, [MyBatis-plus](#) 3.5.7 serves as the ORM framework, connecting to a [MySQL](#) structured database set up through the Docker container version mysql:8.0.32. To enhance system performance, we have introduced the [Redis](#) 6.2.11-alpine Docker container version as a caching mechanism. The front-end is developed using the [Vue](#) 3.2.4 framework within a [Node.js](#) v16.13.0 environment. For front-end page development, [Axios](#) 0.21.4 is used for data interaction with the back-end API, while [Bootstrap](#) v5.1.3 and [Element-UI](#) (element-plus 2.2.0) provide page layout and style design tools. [Font Awesome](#) 6.1.1 provides icon style support. [Echarts](#) 5.3.1, [Plotly](#) 2.23.0 and [CanvasXpress](#) 38.4.1 are used for graph visualization. To ensure the smoothest browsing experience, it is recommended that users access the website with modern web browsers that support HTML5 standard, such as Firefox, Google Chrome and Edge.

Database content and usage

A user-friendly interface for browsing datasets

The "Data Browser" page of scVMAP offers users two intuitive ways to explore detailed data: browsing by scATAC-seq datasets and by traits (Figure 2A). The scATAC-seq dataset-based browsing is further divided into sample-based and cell type-based browsing modes. Users can filter samples by "Tissue type" and "Health type", while cell type-based browsing can filter by "Tissue type" and "Cell type". Trait-based browsing supports filtering by "Type", "ICD-10 category", "ICD-10 subcategory" and "Source cohort". Upon clicking on any interesting "Sample

ID" or "Trait ID", detailed information is presented in a new webpage featuring the following eight panels: (1) Overview module: This module provides comprehensive descriptions and visualizations for selected entries. For single-cell datasets, it details key information such as the tissue origin of samples, the number of cell types, cell count, etc., and visually presents the proportional distribution of cell types via pie charts (Figure 2B). For trait data, it offers information such as trait type, ICD-10 disease categories, the number of associated genetic variant sites, etc., along with pie charts showing the chromatin distribution of variant sites (Figure 2C). (2) Trait relevance score module: This module provides information and visualizations on the associations between target traits and single-cell data calculated by g-chromVAR and SCAVENGE. It includes pie charts of enrichment proportions for single-cell samples or traits (Figure 2D), UMAP plots annotated by cell type and TRSs, and box plots showing the distribution of TRSs across different cell types (Figure 2E). By presenting data from multiple perspectives, this module serves as an important basis for identifying key regulatory cell types associated with genetic traits. (3) Cell type-specific differential genes module: This module provides a heatmap of cell type-specific differential genes and a bubble plot of function enrichment analysis results, offering biological functional insights to support the exploration of trait-relevant cell types (Figure 2F). (4) Trait-relevant genes module: For selected traits, this module presents MAGMA gene-level analysis results and performs function enrichment analysis on related genes. Clicking on a gene provides access to detailed annotation information, including enhancers, super-enhancers, risk SNPs, among other annotations. (5) Gene hub network module: Trait- or disease-cell type association networks primarily document and reveal key genes underlying phenotypic manifestations or pathogenic mechanisms, as well as the potential regulatory relationships of key genes (Figure 2G). (6) Cell type-specific differential TFs module: This module provides tables and heatmaps of differential TFs across different cell types. (7) Trait-relevant TFs module: This module provides a list of TFs enriched in trait-relevant variants predicted by HOMER. (8) TF hub network module: Trait- or disease-cell type association networks primarily document and reveal key TFs underlying phenotypic manifestations or pathogenic mechanisms (Figure 2H).

An interface for convenient retrieving in the "Search" page

The scVMAP search interface provides five query modes for exploring trait-cell associates, including "Search by category or source cohort", "Search by SNP", "Search by tissue or cell type", "Search by gene", and "Search by TF" (Figure 2I). With the "Search by category or source cohort" query, users can flexibly select search strategies by switching between the "Source cohort" and "Category" tabs. After entering a specific "Source cohort" or "Category" entry, the system will return the corresponding list of traits. By clicking a "Trait ID", users can view detailed information about cells associated with that trait. The "Search by SNP" mode allows users to enter an rsID number to retrieve trait information linked to that particular genetic variant and the SNP-relevant regulatory network. The "Search by tissue or cell type" mode enables rapid retrieval of relevant scATAC-seq samples. Clicking on the "Sample ID" in the search results

allows users to enter detailed sample information. In the "Search by gene" mode, users can query a target gene to obtain its basic information, associated traits and scATAC-seq samples, and regulatory annotations. Similarly, in the "Search by TF" mode, users can retrieve the basic information of a specific TF, related traits and scATAC-seq samples.

Online analysis tools

scVMAP provides three analysis tools with multi-level genetic regulation research, including "Variant-to-function mapping", "Variant-to-function mapping with genes", and "Variant-to-function mapping with TFs" (Figure 2J). In "Variant-to-function mapping", users can input multiple traits and select a specific sample to obtain regulatory potential scores for these traits across various cell types. The "Variant-to-function mapping with genes" and "Variant-to-function mapping with TFs" enable users to input custom genes or TFs, and flexibly set differential analysis parameters (Log2(Fold change) and p-value) and trait associated parameters (SNP number and p-value). With a single-click analysis, the tool provides scATAC-seq samples and traits related to the target genes or TFs, facilitating the interpretation of genetic regulatory mechanisms of genes or TFs in specific cell types.

Data download

The "Download" page provides a centralized access portal for all downloadable data from scVMAP, designed to help users conveniently obtain the research resources (Figure 2K). Users can quickly locate target datasets through categorized navigation, including: (i) TRS for each single cell in each sample generated through g-chromVAR and SCAVENGE methods; (ii) gene and TF associated analysis results; (iii) gene regulation annotation results, and (iv) scATAC-seq associated analysis results. Additionally, we also provide downloads of all table data and all visualization graphics in detail page.

Case study

Basal cell carcinoma (BCC) is the most common cancer in humans worldwide, and its incidence is on the rise (60-63). Previous fine-mapping studies have identified several genetic variants with potential causal effects within susceptibility regions for BCC. To further observe the potential pathogenic mechanisms of causal variant effects in the single-cell context of BCC, we utilized the "Search information by tissue type or cell type" function of scVMAP to analyse and identify disease-relevant cell populations. We entered "Skin" into the search box and clicked "Search", retrieving the scATAC-seq data table. We selected the single-cell sample "sample_id_1", derived from BCC (Figure 3A). Then, in the "Overview" module, detailed information for "sample_id_1" is displayed, which contains 20 cell types and over 3,600 cells (Figure 3B). The "Trait-relevant cell score" module provides a trait table related to this BCC sample, as well as UMAP plots and boxplots showing the TRSs for each specific trait across various cells and cell types. Next, we queried the trait table for "BCC" and selected the relevant

BCC disease "trait_id_894" to further investigate the cell populations associated with it (Figure 3C).

UMAP and box plot visualizations show that for "trait_id_894", the TRSs in tumor cells (including the four subtypes Tumor 1-4) are significantly higher than those of other cell types (Figure 3D). These findings indicate the identification of tumor cells closely relevant to BCC within the tumor microenvironment (60). This demonstrated that scVMAP has the capability to accurately map traits to biologically relevant cell types. Notably, when selecting other BCC diseases (e.g., trait_id_895), these analysis results also show good reproducibility (Supplementary data). To explore the functional impact of this trait on tumor cells, we identified genes significantly associated with "trait_id_894" based on MAGMA gene-level analysis, including *RCC2*, *CASC15*, *OCA2*, *IRF4*, and *TP53* (62,64,65). GWAS (Catalog) term enrichment analysis results of "trait_id_894"-relevant genes demonstrated that "trait_id_894" was significantly enriched for multiple skin tumor-relevant functional terms, including "Basal Cell Carcinoma", "Cutaneous Melanoma" and "Cutaneous Squamous Cell Carcinoma" (Figure 3F). Importantly, the "Cell type-specific differential genes module" further identified *RCC2*, *CASC15*, *OCA2*, and *IRF4* as significantly differentially active genes in tumor cells (Figure 3E). These genes are considered candidate marker genes potentially involved in the pathogenesis of BCC. To obtain these candidate marker genes, scVMAP utilized the "Gene hub network" module to construct a gene hub regulatory network from "trait_id_894" to a tumor cell subtype with high TRSs (Figure 3G). The key genes in this network are the candidate marker genes, among which we found multiple reported BCC susceptibility genes, such as *RCC2*, *PADI6*, *CASC15*, *RGS22*, *LINC00111*, and *ZBTB10* (61,62,64,66). Additionally, it also includes genes reported to influence pigmentation in Europeans, such as *ASIP*, *SLC45A2*, and *OCA2* (61,67-69).

Next, we used the identified candidate marker gene *RCC2* as an anchor to elucidate its potential regulatory mechanism in BCC (64). In the "Trait-relevant genes" module, click on the *RCC2* gene to enter the gene details page. On this page, the epigenomic annotation of the *RCC2* gene in the "Regulation regions" panel, including risk SNPs, eQTLs, TEs, and SEs, provides potential evidence linking *RCC2* to skin-relevant diseases (Figure 3I). Meanwhile, in the "Gene-relevant traits or diseases" panel, by clicking the corresponding "View" button, we accessed the detailed enrichment information of "trait_id_894"-relevant *RCC2*. In the "*RCC2*-relevant SNP regulatory network" section, we identified a causal variant *rs57142672* (Figure 3J), which is located within 500kb of the *RCC2* transcription start site (TSS). In the identification of risk loci for BCC, the regulatory relationship involving *RCC2*-*rs57142672* has been reported in the literature (64). scVMAP allows for additional assessment of causal variants. After clicking on *rs57142672* to access the SNP details page, we found that *rs57142672* primarily exhibits causal effects in BCC, malignant skin tumors, and cancer (Figure 3K). These findings fully demonstrate that scVMAP exhibits high reliability in the identification of cell types associated with traits or diseases, providing researchers with a trustworthy data resource.

Furthermore, within the TF-hub association network from "trait_id_894" to tumor cell subtype with high TRSs, we observed a significant correlation between Hypoxia-Inducible Factor 1 Alpha (*HIF1A*) and tumor cell subpopulations (Figure 3H). It is noteworthy that during cancer development, HIFs are described as oncogenes or tumor suppressors under different environmental conditions (70). Previous articles have reported that the *HIF1A* gene plays a potential downregulating role in BCC (71). This result provides guidance for TFs that perform potential regulatory functions in cell types with significant enrichment of traits or diseases.

In summary, scVMAP encompasses multi-level, multi-faceted regulatory annotations for variant-to-function mapping relationships, primarily through cellular trait or disease enrichment analysis, differential gene and differential TF analysis across cell types, MAGMA gene-level analysis, HOMER motif analysis, gene pathway enrichment analysis, gene and TF hub regulatory networks and SNP-relevant regulatory networks. This assists researchers in exploring and revealing the phenotypic manifestations and pathogenic mechanisms of causal variant effects within the cellular environment.

Discussion

Deciphering cell-specific regulatory mechanisms of causal variants on traits or diseases, and exploring the regulatory links between these traits or diseases and fate-relevant cell types, is crucial for genetic medicine progress. To meet this challenge, we developed the scVMAP database platform, leveraging g-chromVAR and SCAVENGE algorithms. scVMAP effectively integrates scATAC-seq and fine-mapping data, providing causal variant to function mapping at single-cell resolution, which elucidates the role and mechanisms of genetic variation at the cellular level. scVMAP not only offers comprehensive trait- or disease-cell and trait- or disease-cell type association relationships but also identifies potential key regulatory genes and TFs involved in these associations. Additionally, it provides epigenetic annotations and gene functional enrichment analyses. These features deliver detailed insights into the functional exploration of complex phenotypes and diseases within cellular contexts. In summary, scVMAP is a user-friendly database that facilitates the searching, browsing, analysis and visualizing of information related to single-cell genetics. As a comprehensive platform that integrates human single-cell chromatin accessibility data with causal variant effects, scVMAP provides researchers with access to a wealth of scATAC-seq datasets, fine-mapping results and TRS data, along with a variety of analytical tools. scVMAP is expected to significantly advance research in single-cell genetics and clinical medicine.

To further enhance the value of the scVMAP database, we are currently constrained by the limited availability of scATAC-seq data with clear cell-type labels, as well as by the accuracy of methods like SCAVENGE. Moving forward, we will continue to incorporate emerging scATAC-seq data while refining our algorithmic pipeline to improve accuracy. We are confident that through continuous improvements, scVMAP will better serve the scientific community by providing researchers with richer and more valuable information resources.

Data availability

The research community can access information freely in the scVMAP without registration or logging in. The URL for scVMAP is <https://bio.liclab.net/scvmap/>.

Supplementary data

Supplementary Data are available at NAR Online.

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Conflict of interest statement

None declared.

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Figure legends

Figure 1. | The architecture of scVMAP. The scVMAP platform has amassed a comprehensive collection of scATAC-seq data annotated with cell type labels, as well as extensive fine-mapping results, all preprocessed through a unified pipeline. scVMAP supports multiple functions, including browsing, searching, analysis, statistics, and downloading.

Figure 2. | Main functions and usage of scVMAP. (A) An interface for efficient browsing of both scATAC-seq data and fine-mapping results. (B) Detailed information for each scATAC-seq sample. (C) Detailed information for each trait or disease. (D) Enriching trait or disease information for each single-cell scATAC-seq sample. (E) Visualization of trait or disease-enriched cells. (F) Heatmap and table of differentially active genes across cell types for each scATAC-seq sample, along with functional enrichment bubble plots for highly scored genes. (G) Gene hub regulatory network for each trait/disease-single cell sample pair. (H) TF hub regulatory network for each trait/disease-single cell sample pair. (I) Five search modes are provided. (J) Three online analysis tools are provided. (K) scVMAP provides a comprehensive collection of downloadable data, including scATAC-seq profiles, trait/disease data, TRS result data, MAGMA gene-level association data, HOMER motif analysis data, differential gene activity and TF activity data, genome annotation data and additional resources.

Figure 3. | Case study of basal cell carcinoma (BCC) sample. (A) Search for BCC using "Search information by tissue type or cell type". (B) Detailed information for BCC sample. (C) Enriching trait or disease information for BCC sample. (D) Visualization of trait or disease-enriched cells. (E) Differential gene heatmap of BCC sample. (F) Bubble plot of MAGMA gene-level enrichment analysis for "trait_id_894" disease. (G) Gene hub regulatory pathway network from "trait_id_894" to "Tumor 3". (H) TF hub regulatory network from "trait_id_894" to "Tumor 3". (I) Gene-relevant regulatory annotations. (J) Gene-relevant SNP network. (K) SNP-relevant regulatory network.