

Title: Integrated spatial model estimates the fish distribution using environmental DNA and catch data

Yuki Kanamori^{1*}, Hiroshi Okamura², Shota Nishijima², Yuki Hongo², Yasuyuki Uto³, Hisatoku Mita⁴, Mitsuhiyo Ishii⁴, Kiyoharu Akimoto⁵, and Akane Kusano⁶

¹ Fisheries Resources Institute, Japan Fisheries Research and Education Agency, 25-259 Shimomekurakubo, Samemachi, Hachinohe, Aomori 031-0841, Japan

² Fisheries Resources Institute, Japan Fisheries Research and Education Agency, 2-12-4 Fukuura, Kanazawa, Yokohama, Kanagawa 236-8648, Japan

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* Corresponding author

Email: kana.yuki@fra.affrc.go.jp

Abstract

Keywords

1 Introduction

Understanding of spatial distribution of species and underlying its mechanism is an essential issue in ecology. Field surveys using environmental DNA (eDNA) are widely used for detecting invasive or rare species and hotspot of biodiversity (面倒なのでレビュー論文を引用) because the surveys of eDNA are easy to detect presence/absence of target species, non-invasiveness, and high cost effectiveness rather than previous direct sampling method (Rees et al. 2014; Thomsen & Willerslev 2015). However, the presence/absence of eDNA includes many types of uncertainties due to relating to environmental factors such as temperature and advection (). For example, in aquatic habitats, it is not sure whether target species are in a location or not when eDNA of target species is detected because eDNA are transported passively. Therefore, the consideration to the influence of environmental factors on eDNA is necessary for estimation of species distribution when we use eDNA methods.

One step towards overcoming these uncertainties is a understanding of the "ecology of eDNA": (Barnes & Turner 2016). Previous studies

Integrated species distribution models are now common spatial model to predict spatial pattern of species (Issac et al. 2020). The model use the different type of data with strengths and weaknesses, such as scientific survey data which is restricted spatially and quantitatively and opportunistic citizen data which is widely collected and abundant, and combine in a single model (Isaac et al. 2020; Miller et al. 2019).

The models combine the different type of data with strengths and weaknesses in a single model (). For example, scientific survey data are high quality but less abundant due to restriction of spatially costly while opportunistic data such as citizen data are widely

23 collected and abundant but may be low quality due to not using consistent field methods.
24 Combining both types of data can capitalize on the strengths of each data and perform better
25 prediction than models when we use single data (Pacifi et al. 2017; Miller et al. 2019).

26 Tokyo Bay is a large enclosed coastal sea in Japan. In Tokyo Bay, there are many
27 commercially important species for fisheries that are called "Edomae" because these species
28 have been used for Sushi since Edo Era (about 400 years ago). Catch of some Edomae have
29 been decreased because of habitat modification due to urbanization (e.g., landfill of tidal
30 flats and water pollution). Catch statistics (total catch in each species, efforts, and
31 geographic location of fishing) have been collected for stock assessment since 1990 by
32 prefectures around Tokyo Bay. The strengths of this data are the direct evidence that a focal
33 species occupies a location of fishing and abundant because of widely collected in Tokyo
34 Bay. On the other hand, weakness of this data is like a opportunistic data because the data is
35 likely to be biased towards areas to high density of focal species due to commercially fishes,
36 consequently less zero data. In addition to this catch statistics, scientific survey of eDNA
37 has been conducted monthly since 2018 for biodiversity monitoring because biodiversity
38 also may decreased due to human-induced environmental changes in Tokyo Bay (Hongo et
39 al., submitted). The strengths are that the data is systematically collected by scientific survey
40 data and includes zero data, while the weaknesses are that the data is less abundant due to
41 spatial restriction of the survey and includes uncertainties in presence/absence as description
42 in above.

43 In this paper, to predict spatial distribution of species from eDNA, we first make a
44 model which considers uncertainties of eDNA caused by environmental factors without
45 additional laboratory experiments and numerical hydrodynamic models, by using an

integrated spatial distribution model. We then apply the model to both eDNA data and catch statistics for four Edomae fish in Tokyo Bay, Japan. The predicted spatial distribution of four fish from our model reduced

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2 Materials and Methods

2.1 A general model to estimate species distribution from eDNA

2.2 An application to a eDNA and catch data in Tokyo Bay

2.2.1 eDNA data

2.2.1.1 Field survey

Field surveys were conducted at 14 sites in Tokyo Bay from April to December in 2018 using R/V Fusanami or R/V Fusami-maru of Chiba Prefecture and R/V Enoshima-maru of Kanagawa Prefecture (Fig. 1). In each station, seawater for eDNA analysis and environmental data were simultaneously collected. For eDNA analysis, two liter of bottom seawater was collected using a Niskin water sampler and were separated for two 1L samples. Each samples filtered glass fiber membrane GF/F (0.7 μm pore size; Cytiva, Sheffield, UK) onboard and then the filters were frozen on a block of dry ice. These frozen filters were transported and stored at -30° in the laboratory until eDNA extraction. To lower the levels of cross-contamination, equipments for eDNA sampling were changed new one or washed in each site. During sampling the bottom seawater, water temperature, salinity, pH, and dissolved oxygen (DO) were measured by CTD (メーカー?).

66 **2.2.1.1 Laboratory experiments**

67 In laboratory, eDNA extraction, eDNA amplification, and eDNA sequence were conducted.
68 Total eDNA was extracted from the frozen filters using a DNeasy Blood and Tissue Kit
69 (Qiagen, Hilden, Germany) following Yamamoto et al. 2019. Mitochondrial 12S rRNA
70 gene was amplified using MiFish universal primers referring to Miya et al. 20

71 **2.2.2 Catch statistics**

72 To estimate the chub– and the chub+ indices of egg density by considering sampling effects
73 (i.e., spatio–temporal changes in survey design) as well as the effect of the egg density of
74 chub mackerel on the catchability of egg density of spotted mackerel, we used the
75 multivariate vector autoregressive spatio-temporal (VAST) model (Thorson and Barnett,
76 2017), which accounts for spatio-temporal changes in survey design, survey effort, and
77 observation rates and can accurately estimate relative local densities at high resolution by
78 standardizing sampling designs (Thorson and Barnett, 2017; Thorson, 2019). The model
79 includes two potential components because it is designed to support delta-models: (i) the
80 encounter probability p_i for each sample i and (ii) the expected egg density d_i for each
81 sample i when spawning occurs (i.e., egg density is not zero). The encounter probability p_i
82 and the expected egg density d_i are, respectively, approximated using a logit-linked linear
83 predictor and a log-linked linear predictor as follows (Thorson and Barnett, 2017):

$$\begin{aligned}\text{logit } p_i &= \beta_p(t_i) + \omega_p(s_i) + \varepsilon_p(s_i, t_i) + \eta_p(v_i) + \lambda_p Q(i) \\ \log d_i &= \beta_d(t_i) + \omega_d(s_i) + \varepsilon_d(s_i, t_i) + \eta_d(v_i) + \lambda_d Q(i)\end{aligned}\tag{1}$$

84 where $\beta(t_i)$ is the intercept for year t , and $\omega(s_i)$ and $\varepsilon(s_i, t_i)$ are the spatial and

85 spatio-temporal random effects for year t and location s , respectively. $\eta(v_i)$ is the
86 overdispersion random effect of factor v_i , which is the interaction of year and month. λ is
87 the effect of the catchability covariate $Q(i)$:

$$Q(i) = \log(d_{chub}(s_i) + 0.1).$$

88 That is, this term considers the effect of species misidentification between chub mackerel
89 and spotted mackerel; as mentioned earlier, we suspected overestimation of egg density of
90 spotted mackerel because the difference in egg diameter has become ambiguous according
91 to increase in egg density of chub mackerel and the distributions of egg diameters between
92 species have overlapped (Yukami et al., 2019). The constant 0.1 was added because $\log 0$
93 (i.e., no chub mackerel eggs) is undefined, and the same result was obtained when using 1 in
94 place of 0.1.

95 **2.2.3 Estimation of spatial distribution**

96

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99 **Authorship**

100 YK conceived of the research idea. YH, YU, HM, MI, KA, and AK conducted field
101 sampling. YH performed the laboratory experiments. YK, HO, and SN designed statistical
102 analyses. YK wrote programs and performed the analyses. YK wrote the manuscript with
103 input from all co-authors' comments.