

Title: Integrated spatial model estimates the fish distribution using environmental DNA and catch data

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Abstract

Keywords

1 Introduction

Understanding of spatial distribution of species and underlying its mechanism is an essential issue in ecology. Field surveys using environmental DNA (eDNA) are widely used for detecting invasive or rare species and hotspot of biodiversity (面倒なのでレビュー論文を引用) because the surveys of eDNA are easy to detect presence/absence of target species, non-invasiveness, and high cost effectiveness rather than previous direct sampling method (Rees et al. 2014; Thomsen & Willerslev 2015). However, the presence/absence of eDNA includes many types of uncertainties due to relating to environmental factors such as temperature and advection (). For example, in aquatic habitats, it is not sure whether target species are in a location or not when eDNA of target species is detected because eDNA are transported passively. Therefore, the consideration to the influence of environmental factors on eDNA is necessary for estimation of species distribution when we use eDNA methods.

One step towards overcoming these uncertainties is a understanding of the "ecology of eDNA": (Barnes & Turner 2016). Previous studies

Integrated species distribution models (IDMs) are now common spatial model to predict spatial pattern of species (Issac et al. 2020). The model use the different type of data with strengths and weaknesses, such as scientific survey data which is restricted spatially and quantitatively and opportunistic citizen data which is widely collected and abundant, and combine in a single model (Isaac et al. 2020; Miller et al. 2019).

The models combine the different type of data with strengths and weaknesses in a single model (). For example, scientific survey data are high quality but less abundant due to restriction of spatially costly while opportunistic data such as citizen data are widely

23 collected and abundant but may be low quality due to not using consistent field methods.
24 Combining both types of data can capitalize on the strengths of each data and perform better
25 prediction than models when we use single data (Pacifi et al. 2017; Miller et al. 2019).

26 Tokyo Bay is a large enclosed coastal sea in Japan. In Tokyo Bay, there are many
27 commercially important species for fisheries that are called "Edomae" because these species
28 have been used for Sushi since Edo Era (about 400 years ago). Catch of some Edomae have
29 been decreased because of habitat modification due to urbanization (e.g., landfill of tidal
30 flats and water pollution). Catch statistics (total catch in each species, efforts, and
31 geographic location of fishing) have been collected for stock assessment since 1990 by
32 prefectures around Tokyo Bay. The strengths of this data are the direct evidence that a focal
33 species occupies a location of fishing and abundant because of widely collected in Tokyo
34 Bay. On the other hand, weakness of this data is like a opportunistic data because the data is
35 likely to be biased towards areas to high density of focal species due to commercially fishes,
36 consequently less zero data. In addition to this catch statistics, scientific survey of eDNA
37 has been conducted monthly since 2018 for biodiversity monitoring because biodiversity
38 also may decreased due to human-induced environmental changes in Tokyo Bay (Hongo et
39 al., submitted). The strengths are that the data is systematically collected by scientific survey
40 data and includes zero data, while the weaknesses are that the data is less abundant due to
41 spatial restriction of the survey and includes uncertainties in presence/absence as description
42 in above.

43 In this paper, to predict spatial distribution of species from eDNA, we first make a
44 model which considers uncertainties of eDNA caused by environmental factors without
45 additional laboratory experiments and numerical hydrodynamic models, by using an

integrated spatial distribution model (eDNA-IDM). We then apply the model to both eDNA data and catch statistics for four Edomae fish in Tokyo Bay, Japan. The predicted spatial distribution of four fish from our model reduced

2 Materials and Methods

2.1 A general model to estimate species distribution from eDNA

2.2 An application to a eDNA and catch data in Tokyo Bay

2.2.1 eDNA data

2.2.1.1 Field survey

Field surveys were conducted at 14 sites in Tokyo Bay from April to December in 2018 using R/V Fusanami or R/V Fusami-maru of Chiba Prefecture and R/V Enoshima-maru of Kanagawa Prefecture (Fig. 1). In each station, seawater for eDNA analysis and environmental data were simultaneously collected. For eDNA analysis, two liter of bottom seawater was collected using a Niskin water sampler and were separated for two 1L samples. Each samples filtered glass fiber membrane GF/F (0.7 μm pore size; Cytiva, Sheffield, UK) onboard and then the filters were frozen on a block of dry ice. These frozen filters were transported and stored at -30° in the laboratory until eDNA extraction. To lower the levels of cross-contamination, equipments for eDNA sampling were changed new one or washed in each site. During sampling the bottom seawater, water temperature, salinity, pH, and dissolved oxygen (DO) were measured by CTD (メーカー).

66 2.2.1.1 Laboratory experiments

67 In laboratory, eDNA extraction, eDNA amplification, and eDNA sequence were conducted.
68 Total eDNA was extracted from the frozen filters using a DNeasy Blood and Tissue Kit
69 (Qiagen, Hilden, Germany) following Yamamoto et al. 2019. Mitochondrial 12S rRNA
70 gene was amplified using MiFish universal primers referring to Miya et al. 2015 with slight
71 modification. The details was shown in Hongo et al. (受理されてないようだったら書くし
72 かない). eDNA sequence were

73 2.2.2 Catch statistics

74 A part of catch statistics of small-scale bottom trawl fisheries recorded by several
75 representative boats of Chiba Prefecture were provided by Chiba Prefecture. This data
76 included date, geographic location, efforts (number of tows), gear, and catch weight (kg) in
77 each fish. Almost of all gear was beam trawl although dredge net also used. The species
78 which also detected by eDNA was *Conger myriaster* (マアナゴ), *Kareius bicoloratus* (イシ
79 ガレイ), *Lateolabrax japonicus* (スズキ), and *Konosirus punctatus* (コノシロ). Thus, we
80 estimated the spatial distribution of these four species using the eDNA-IDM. マコガレイ,
81 カマス類, クロダイ, イシモチ類も解析できる??

82 2.2.3 Estimation of spatial distribution

83 equation examples

$$\begin{aligned}\text{logit } p_i &= \beta_p(t_i) + \omega_p(s_i) + \varepsilon_p(s_i, t_i) + \eta_p(v_i) + \lambda_p Q(i) \\ \log d_i &= \beta_d(t_i) + \omega_d(s_i) + \varepsilon_d(s_i, t_i) + \eta_d(v_i) + \lambda_d Q(i)\end{aligned}\tag{1}$$

84 where $\beta(t_i)$ is the intercept for year t , and $\omega(s_i)$ and $\varepsilon(s_i, t_i)$ are the spatial and

85 spatio–temporal random effects for year t and location s , respectively. $\eta(v_i)$ is the
86 overdispersion random effect of factor v_i , which is the interaction of year and month. λ is
87 the effect of the hatchability covariate $Q(i)$:

$$Q(i) = \log(d_{chub}(s_i) + 0.1).$$

88 That is, this term considers the effect of species misidentification between chub mackerel
89 and spotted mackerel; as mentioned earlier, we suspected overestimation of egg density of
90 spotted mackerel because the difference in egg diameter has become ambiguous according
91 to increase in egg density of chub mackerel and the distributions of egg diameters between
92 species have overlapped (Yukami et al., 2019). The constant 0.1 was added because $\log 0$
93 (i.e., no chub mackerel eggs) is undefined, and the same result was obtained when using 1 in
94 place of 0.1.

95

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98 **Authorship**

99 YK conceived of the research idea. YH, YU, HM, MI, KA, and AK conducted field
100 sampling. YH performed the laboratory experiments. YK, HO, and SN designed statistical
101 analyses. YK wrote programs and performed the analyses. YK wrote the manuscript with
102 input from all co-authors' comments.