

CYP1A2 DDI Qualification

Version	2.0-OSP12.1
Qualification Plan Release	https://github.com/Open-Systems-Pharmacology/Qualification-DDI-CYP1A2/releases/tag/v2.0
OSP Version	12.1
Qualification Framework Version	3.4

This qualification report is filed at:

<https://github.com/Open-Systems-Pharmacology/OSP-Qualification-Reports>

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1 Introduction

1.1 Objective

This qualification report evaluates the developed PBPK drug-drug interactions (DDI) models network for the ability to perform simulations with the intended purpose to predict cytochrome P450 1A2 (**CYPA12**)-mediated DDI.

To demonstrate the level of confidence, the predictive performance of the platform for this intended purpose is assessed via a network of PBPK models of selected index CYP1A2 DDI perpetrators, and respective sensitive CYP1A2 victim drugs and a comprehensive dataset from published clinical DDI studies. All PBPK models represent whole-body PBPK models, which allow dynamic DDI simulations in organs expressing CYP1A2.

The respective *qualification plan* to produce this *qualification report* is transparently documented and provided open-source (<https://github.com/Open-Systems-Pharmacology/OSP-Qualification-Reports>). The same applies for all presented PBPK models including *evaluation reports* on model building and evaluation of each model (<https://github.com/Open-Systems-Pharmacology/OSP-PBPK-Model-Library>).

Evaluation reports including descriptions on model building and detailed evaluations of the included models are documented separately (see [Section 1.2](#)).

Please refer to the [Appendix](#) to learn more details:

- An overview over the Open Systems Pharmacology Suite is given in chapter [Section 6.1](#)
- [Section 6.2](#) shows the implementation of the underlying mathematical equations for drug-drug interactions in the OSP suite.
- A detailed general description of the performed qualification workflow (*qualification plan*, *qualification report*, etc.) can be found in chapter [Section 6.3](#).

1.2 CYP1A2 DDI Network

CYP1A2 is involved in the elimination of about 15% of all therapeutic drugs (e.g., clozapine, tacrine, tizanidine, and theophylline), a number of procarcinogens (e.g. benzo[a]pyrene and aflatoxin B1), and several important endogenous compounds (e.g. steroids and arachidonic acids) ([Zhou 2009](#), [Goldstein 2001](#)). This enzyme is exclusively expressed in the liver and can be markedly induced by smoking. Well-known substrates of CYP1A2 include caffeine and tizanidine.

Like other CYPs, CYP1A2 is subject to induction and/or inhibition by a number of compounds, which can result in significant drug interactions in clinical practice.

The U.S. Food and Drug Administration (FDA) lists several perpetrator and victim drugs of interactions in the CYP1A2 network ([FDA](#)). For instance, caffeine and tizanidine are classified as sensitive index substrates for CYP1A2, and fluvoxamine is listed as a strong clinical index inhibitor of CYP1A2.

To qualify the developed models for the prediction of the CYP1A2 DDI potential of new drugs, a set of verified PBPK models of index perpetrators and respective CYP1A2 DDI victim drugs is specified to set up a CYP1A2-mediated DDI modeling network.

The following perpetrator compounds were selected:

- **Fluvoxamine** (strong CYP1A2 inhibitor) Model snapshot and evaluation plan (*release alt_v1.0*):
https://github.com/Open-Systems-Pharmacology/Fluvoxamine-Model/releases/tag/alt_v1.0
- **Ethinylestradiol** (moderate CYP1A2 inhibitor) Model snapshot and evaluation plan (*release v2.0*):
<https://github.com/Open-Systems-Pharmacology/Ethinylestradiol-Model/releases/tag/v2.0>

- **Mexiletine** (moderate CYP1A2 inhibitor) Model snapshot and evaluation plan (*release v2.0*):
<https://github.com/Open-Systems-Pharmacology/Mexiletine-Model/releases/tag/v2.0>

The following sensitive CYP1A2 substrates as victim drugs were selected:

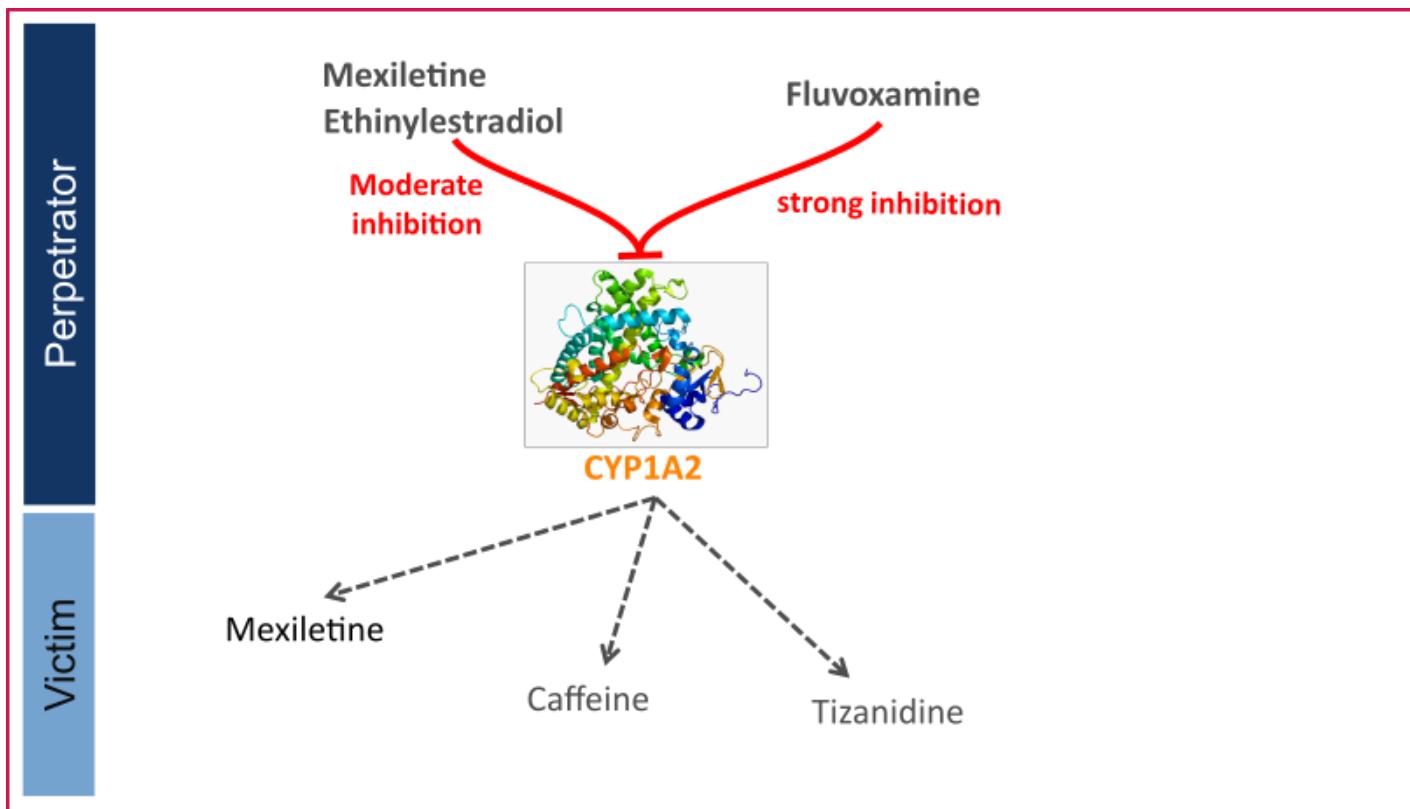
- **Caffeine** PK-Sim compound template
- **Tizanidine** Model snapshot and evaluation plan (*release v2.0*): <https://github.com/Open-Systems-Pharmacology/Tizanidine-Model/releases/tag/v2.0>
- **Mexiletine** Model snapshot and evaluation plan (*release v2.0*): <https://github.com/Open-Systems-Pharmacology/Mexiletine-Model/releases/tag/v2.0>

The following interaction studies were predicted and used to qualify/optimize the final network:

- Strong CYP1A2 inhibition
 - Fluvoxamine - caffeine
 - Fluvoxamine - tizanidine
 - Fluvoxamine - mexiletine
- Moderate CYP1A2 inhibition
 - Mexiletine - caffeine
 - Mexiletine - tizanidine
 - Ethinylestradiol - caffeine
 - Ethinylestradiol - tizanidine

Figure 1 shows the specified and developed DDI modeling network of interacting perpetrator and victim drugs.

Figure 1: CYP1A2 DDI modeling network



The Ki values used to predict the interactions are listed in [Table 1](#).

Inhibitor category	Inhibitor	Substrate	Ki	Reference
Strong CYP1A2	Fluvoxamine	caffeine	2.97 nM	Iga 2016
		tizanidine	0.8697 nM	Fit ¹
		mexiletine	2.97 nM	Iga 2016
Moderate CYP1A2	Mexiletine	caffeine	0.28 µM	Wei 1999
		tizanidine	0.28 µM	Wei 1999
		Ethinylestradiol (EE)	0.48 µM	Fit ²
		tizanidine	0.48 µM	Fit ²

Table 1: Ki values used in CYP1A2 DDI network. ¹Lowest literature value = 0.9 nM; ²Literature value = 10.6 µM

The published DDI studies between the respective perpetrators and victim drugs were simulated and compared to observed data. The following sections give an overview of the clinical studies being part of this qualification report.

1.2.1 Caffeine - Ethinylestradiol DDI

A dynamical DDI simulation with ethinylestradiol as CYP1A2 inhibitor and caffeine as victim was conducted and compared to literature data. No profiles of caffeine under this interaction were available, but [Balogh 1995](#) reported a 55% decrease in caffeine CL due to ethinylestradiol co-administration and report AUCR and CmaxR.

1.2.2 Tizanidine - Ethinylestradiol DDI

The tizanidine-ethinylestradiol interaction was evaluated using clinical DDI studies listed in [Table 2](#).

Source	Route	Dose [mg] / Schedule *	Pop.	Sex	N	Form.
Granfors 2005	p.o.	EE: 0.02 Tizanidine: 4	HV	f	15	tablet

Table 2: Literature sources of clinical concentration data of tizanidine used for DDI prediction qualification with ethinylestradiol. *EE*: *ethinylestradiol*

A dynamical DDI simulation with ethinylestradiol as moderate CYP1A2 inhibitor and tizanidine as victim drug was conducted and compared to literature data. Clinical observations were derived from [Granfors 2005](#) in which 15 healthy women using Oral contraceptives (OCs) (ethinylestradiol + gestodene) and 15 healthy women without OCs (control subjects) ingested a single dose of 4 mg tizanidine.

1.2.3 Caffeine - Fluvoxamine DDI

The caffeine-fluvoxamine interaction was evaluated using clinical DDI studies listed in [Table 3](#).

Source	Route	Dose [mg] / Schedule *	Pop.	Age [yrs] (mean)	Weight [kg] (mean)	Sex	N	Form.
Jeppesen 1996	p.o.	200 caffeine, 50 mg 4 days, 100 mg 8 days fluvoxamine	HV	27	-	-	8	Tablet
Culm-Merdek 2005	p.o.	250 caffeine, 100 b.i.d. fluvoxamine	HV	50	82	m/f	7	Capsule

Table 3: Literature sources of clinical concentration data of caffeine used for DDI prediction qualification with fluvoxamine.

A dynamical DDI simulation with fluvoxamine as CYP1A2 inhibitor and caffeine as victim was conducted and compared to literature data. For caffeine the template model included in PK-Sim was used. Two published clinical studies were found where fluvoxamine and caffeine were given together. Fluvoxamine is a strong inhibitor of CYP1A2 and increases the AUC of caffeine by 7 to 14-fold.

In [Jeppesen 1996](#), subjects in one group received 50 mg fluvoxamine-maleate on the first 4 days followed by 100 mg q.d. for 8 days. On day 8 they received a single dose of 200 mg caffeine. The other group received caffeine without fluvoxamine co-treatment.

In [Culm-Merdek 2005](#), seven healthy subjects received single 250 mg dose of caffeine (or matching placebo) together with fluvoxamine (four doses of 100 mg over 2 days) or with matching placebo in a cross-over fashion.

The simulated caffeine levels with co-administered fluvoxamine were underpredicted. However, the pre-dose concentrations of caffeine were not 0 in the test group. It may be speculated that subjects may not have refrained completely from caffeine containing beverages before the test period. To investigate this hypothesis, a simulation was conducted where a dose of 100 mg caffeine (corresponding approximately to the caffeine content of one cup of coffee ([Caffeine quantities, FDA](#))) was given 24 hours before the administration of caffeine-tablets as per the study protocol. The resulting simulation results support this hypothesis. Hence it was not deemed necessary to adjust the underlying caffeine or fluvoxamine models but rather conclude that the clinical study potentially was facing issues with subjects not compliant with the protocol rules and drank coffee the morning before the study day. The final DDI simulations were therefore conducted with administration of 100 mg caffeine 24 hours prior study protocol.

1.2.4 Tizanidine - Fluvoxamine DDI

The tizanidine-fluvoxamine interaction was evaluated using clinical DDI studies listed in [Table 4](#).

Source	Route	Dose [mg] / Schedule *	Pop.	Age [yrs]	Weight [kg]	Sex	N	Form.
Granfors 2004	p.o.	4 mg tizanidine, 100 mg fluvoxamine	HV	21-31	65-83	m	10	Tablet

Table 4: Literature sources of clinical concentration data of tizanidine used for DDI prediction qualification with fluvoxamine. *:*single dose*

In the clinical study reported by [Granfors 2004](#), a single oral dose of 4 mg tizanidine was given after treatment with fluvoxamine (100mg fluvoxamine-maleate ~73.3 mg free base, q.d. for 4 days).

Initially, the Ki value for the inhibition of CYP1A2 by fluvoxamine reported by [Iga 2016](#) was used. While the predicted Cmax ratio matched closely the observed value, the AUC ratio was underpredicted by a factor of 2.1. Since the predictions of the tizanidine profiles without co-administration of fluvoxamine matched the observations very well, it was concluded that the most plausible reason for the under prediction was the value of Ki, as the fraction metabolized via CYP1A2 was

already 99%. Hence Ki was optimized using the data from [Granfors 2004](#). A Ki value of 0.8697 +/- 0.1935 nmol/L was estimated, which is still in line with the Ki values derived for other CYP1A2 substrates.

1.2.5 Tizanidine - Mexiletine DDI

The tizanidine-mexiletine interaction was evaluated using clinical DDI studies listed in [Table 5](#).

Source	Route	Dose [mg] / Schedule *	Pop.	Sex	N	Form.
Momo 2010	p.o.	2 mg tizanidine, 50 mg b.i.d mexiletine	HV	m	12	Tablet

Table 5: Literature sources of clinical concentration data of tizanidine used for DDI prediction qualification with mexiletine.

A dynamical DDI simulation with mexiletine as CYP1A2 inhibitor and tizanidine as victim was conducted and compared to literature data from [Momo 2010](#). The same Ki as for the caffeine interaction simulation was used (0.28 μ M). The predefined "Standard European Male for DDI" individual (age = 30 y, weight = 73 kg, height = 176 cm, BMI = 23.57 kg/m²) was used.

The pharmacokinetics of tizanidine was examined in an open-label study in 12 healthy participants after a single dose of tizanidine (2 mg) with and without mexiletine co-administration (50 mg, 3 times as a pretreatment for a day and 2 times on the study day).

1.2.6 Caffeine - Mexiletine DDI

The caffeine-mexiletine interaction was evaluated using clinical DDI studies listed in [Table 6](#).

Source	Route	Dose [mg] / Schedule *	Pop.	N
Joeres 1987	p.o.	366 caffeine, 200 mexiletine	HV	6

Table 6: Literature sources of clinical concentration data of caffeine used for DDI prediction qualification with mexiletine.

A dynamical DDI simulation with mexiletine as CYP1A2 inhibitor and caffeine as victim was conducted and compared to literature data.

Clinical observations of caffeine-mexiletine interaction were derived from [Joeres 1987](#) where 6 healthy volunteers received 366 mg caffeine (400 mg caffeine monohydrate) after an overnight fast, together with 200 mg mexiletine orally. One week later caffeine was administered alone.

The in-built caffeine template model included in PK-Sim was used for caffeine predictions.

Competitive inhibition was assumed on CYP1A2 enzyme between caffeine (substrate) and mexiletine (inhibitor) with an inhibitory constant of 0.28 μ M. No reported values were found for mexiletine Ki on CYP1A2 and caffeine as substrate, so the inhibitory constant for methoxyresofurin ([Ko 1997](#)) was used [calculated *in vivo*, unbound Ki for methoxyresofurin].

1.2.7 Mexiletine - Fluvoxamine DDI

The mexiletine-fluvoxamine interaction was evaluated using clinical DDI studies listed in [Table 7](#).

Source	Route	Dose [mg] / Schedule *	Pop.	Sex	N	Form.
Kusumoto 2001	p.o.	166.62 mexiletine 50 b.i.d. fluvoxamine	HV japanese	m	6	-

Table 7: Literature sources of clinical concentration data of mexiletine used for DDI prediction qualification with fluvoxamine. *:*single dose*

A dynamical DDI simulation with fluvoxamine as CYP1A2 inhibitor and mexiletine as victim was conducted and compared to literature data ([Kusumoto 2001](#)). A typical Japanese subject (age = 30 y, weight = 61.87 kg, height = 168.99 cm, BMI = 21.67 kg/m²) was created in PK-Sim from predefined database "Japanese (2015)" by adding CYP2D6 and CYP1A2 expressions from PK-Sim RT PCR database.

A randomized crossover design with two phases was used. Each subject received an oral dose of mexiletine (200 mg) or fluvoxamine (50 mg twice a day) for 7 days, and on the eighth day they received mexiletine and fluvoxamine concomitantly.

2 Qualification of Use Case CYP1A2-mediated DDI

The following section shows the correlations between observed and model-predicted AUC and C_{max} ratios, respectively.

Specifically, the PBPK model performance for the PK parameters **AUC ratio (AUCR)** and **C_{max} ratio (CMAXR)** is assessed via:

- predicted (*Pred*) vs. observed (*Obs*) plots
- *Pred/Obs* vs. *Obs* plots
- geometric mean fold error (GMFE):

$$10^{\frac{\sum |\log(\frac{Pred}{Obs})|}{n}}$$

- number of AUCR and CMAXR falling within 2-fold error range and within the limits as suggested by [Guest et al. 2011](#)
- detailed table of results for each study

In the plots,

- the dotted lines denote 0.50–2.00 (2-fold) criterion,
- the solid lines denote the limits as suggested by [Guest et al. 2011](#),
- the bold solid line denotes the unity line,
- each color represents one combination of drugs

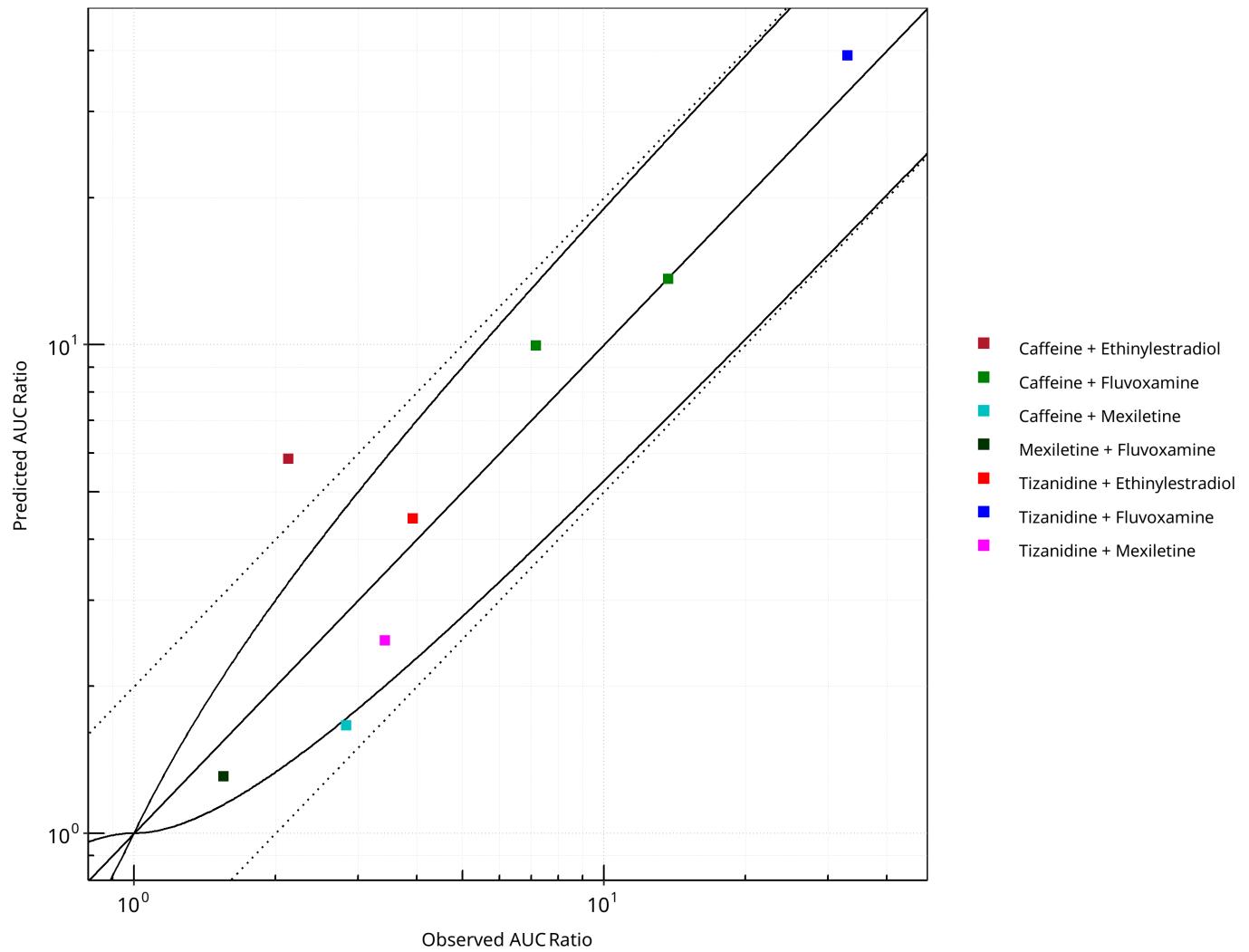


Figure 2-1: CYP1A2 DDI. Predicted vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

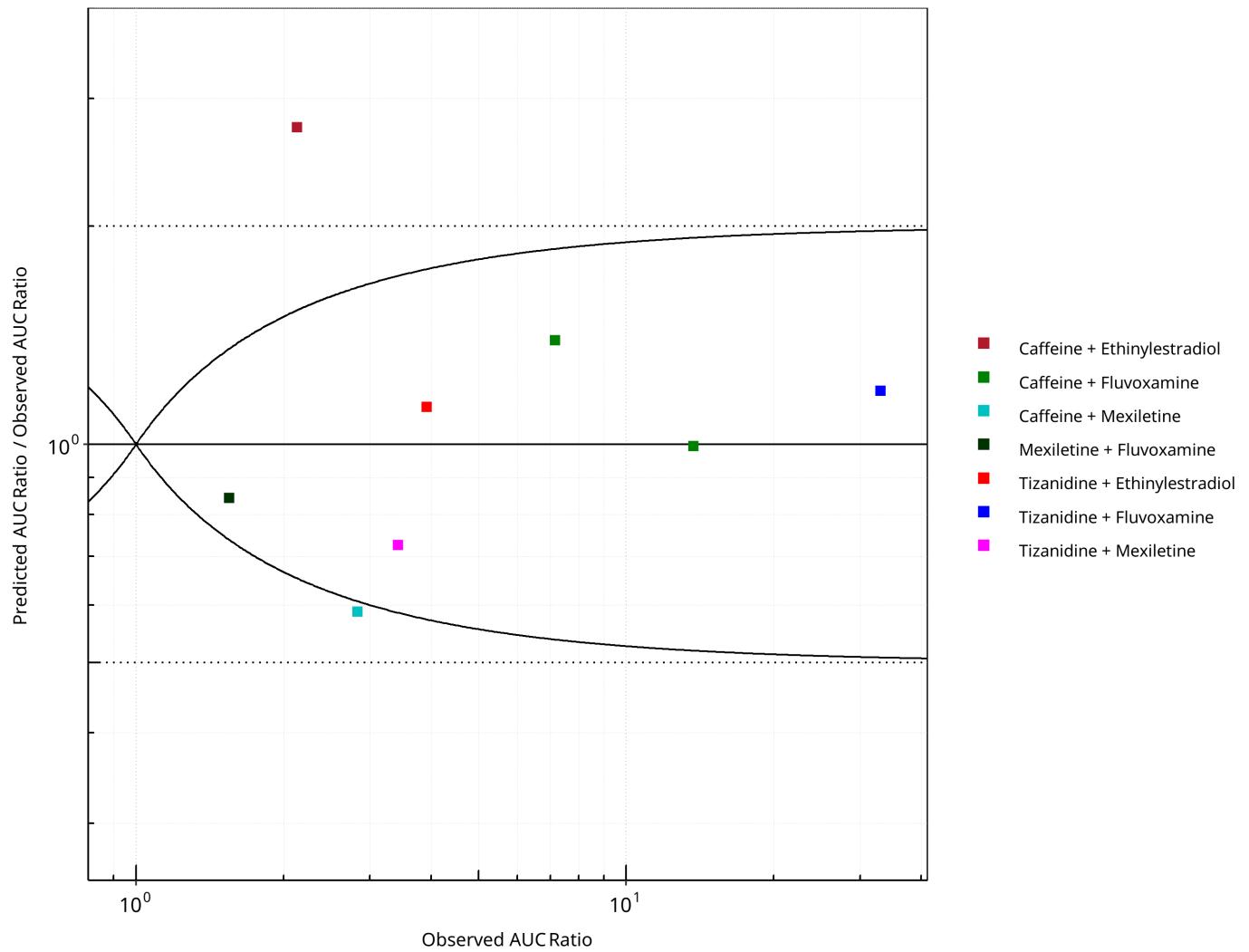


Figure 2-2: CYP1A2 DDI. Predicted/Observed vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

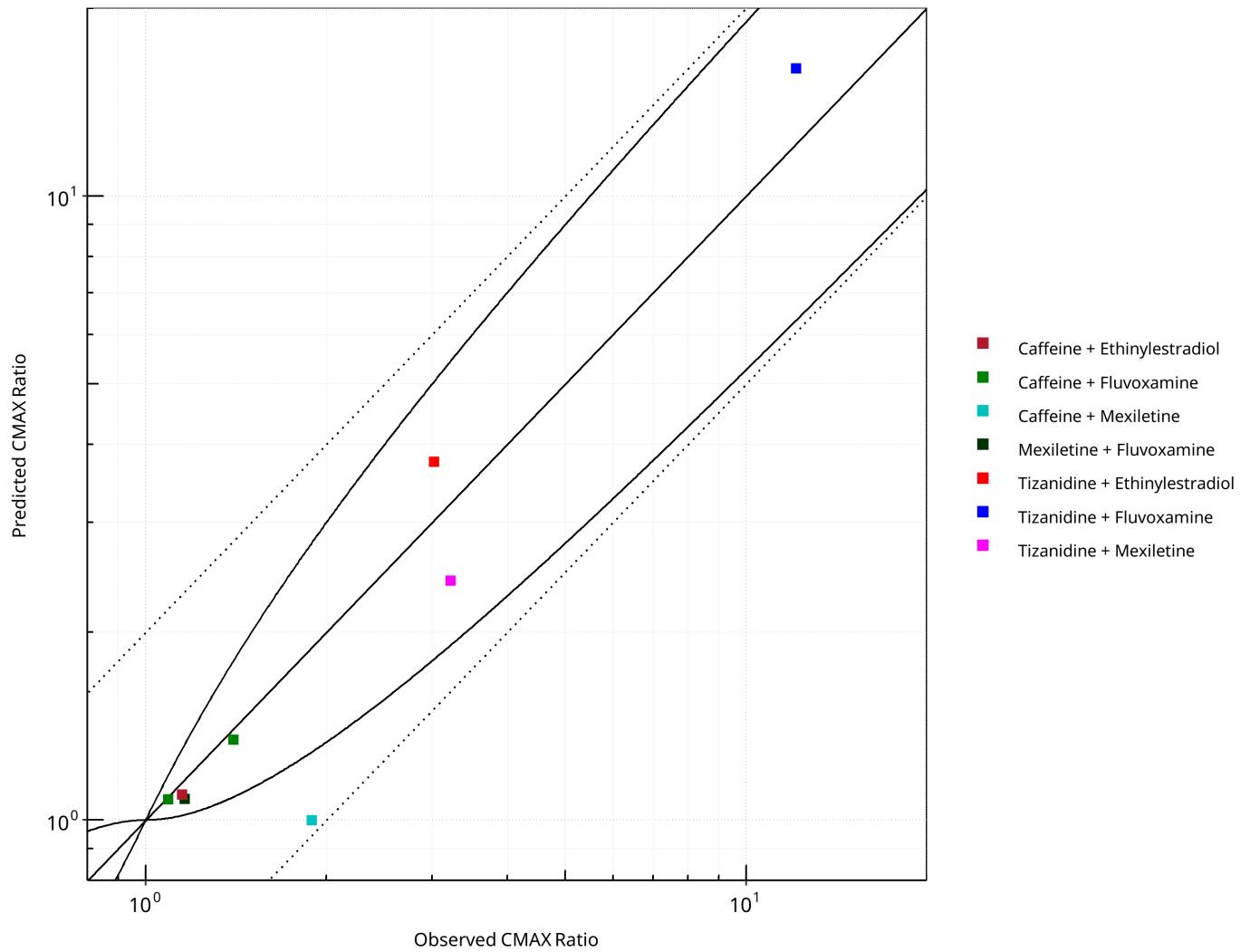


Figure 2-3: CYP1A2 DDI. Predicted vs. Observed CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

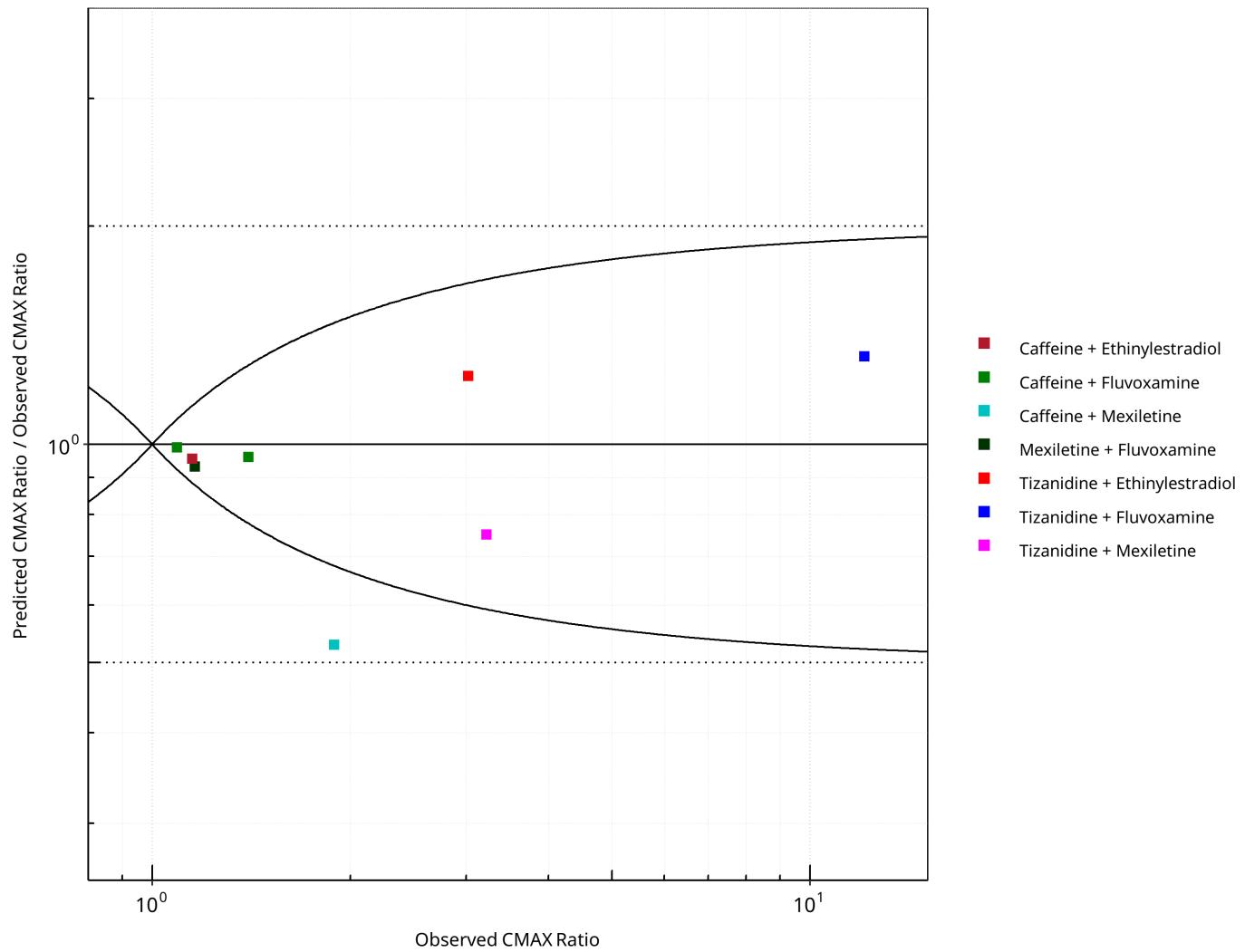


Figure 2-4: CYP1A2 DDI. Predicted/Observed vs. Observed CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

Table 2-1: GMFE for CYP1A2 DDI Ratio

PK parameter	GMFE
AUC	1.39
CMAX	1.22

Table 2-2: Summary table for CYP1A2 DDI - AUC Ratio. ($\delta = 1$ in Guest *et al.* formula)

AUC	Number	Ratio [%]
Points total	8	-
Points within Guest <i>et al.</i>	6	75.00
Points within 2 fold	7	87.50

Table 2-3: Summary table for CYP1A2 DDI - CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

CMAX	Number	Ratio [%]
Points total	8	-
Points within Guest <i>et al.</i>	7	87.50
Points within 2 fold	8	100.00

Table 2-4: Summary table for CYP1A2 DDI

DataID	Perpetrator	Victim	Predicted AUC Ratio	Observed AUC Ratio	Pred/Obs AUC Ratio	Predicted CMAX Ratio	Observed CMAX Ratio	Pred/Obs CMAX Ratio	Reference
6110	Ethinylestradiol, 0.03 mg, PO,	Caffeine, PO	5.84	2.13	2.74	1.10	1.15	0.96	Balogh 1995
13017	Fluvoxamine, 100 mg, PO,	Tizanidine, PO	39.12	33.00	1.19	16.00	12.10	1.32	Granfors 2004
13018	Mexiletine, 50 mg, PO,	Tizanidine, PO	2.48	3.42	0.73	2.42	3.22	0.75	Momo 2010
14002	Ethinylestradiol, 20 mg, PO,	Tizanidine, PO	4.41	3.92	1.13	3.75	3.02	1.24	Granfors 2005
14004	Fluvoxamine, 100 mg, PO,	Caffeine, PO	9.97	7.16	1.39	1.08	1.09	0.99	Jeppesen 1996
14006	Fluvoxamine, 100 mg, PO,	Caffeine, PO	13.64	13.71	0.99	1.34	1.40	0.96	Culm-Merdek 2005
14008	Mexiletine, 200 mg, PO,	Caffeine, PO	1.66	2.83	0.59	1.00	1.89	0.53	Joeres 1987
14010	Fluvoxamine, 50 mg, PO,	Mexiletine, PO	1.31	1.55	0.84	1.08	1.16	0.93	Kusumoto 2001

2.1 Perpetrator

2.1.1 Ethinylestradiol

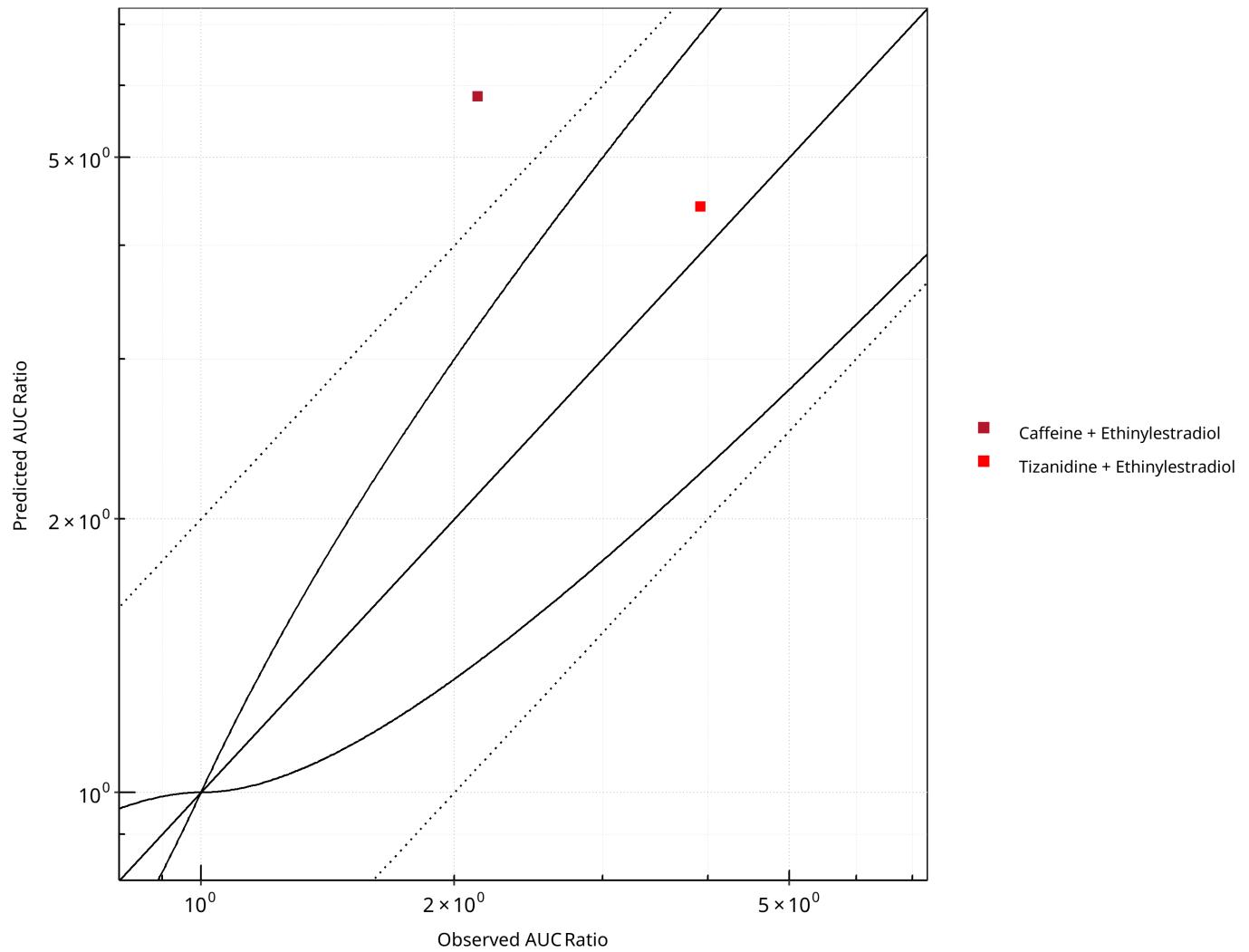


Figure 2-5: CYP1A2 DDI. Perpetrator: Ethinylestradiol. Predicted vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

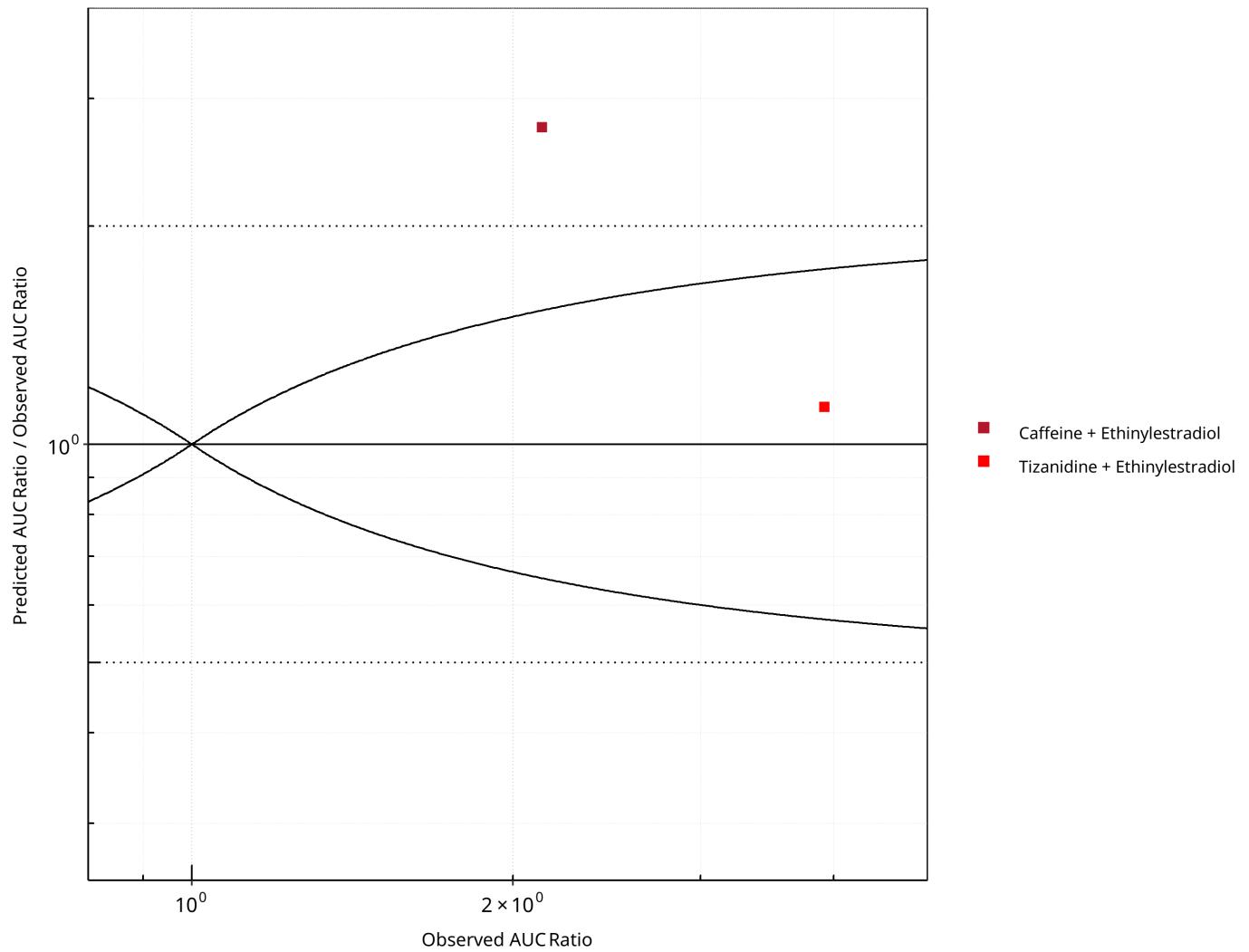


Figure 2-6: CYP1A2 DDI. Perpetrator: Ethinylestradiol. Predicted/Observed vs. Observed AUC Ratio. ($\delta = 1$ in Guest *et al.* formula)

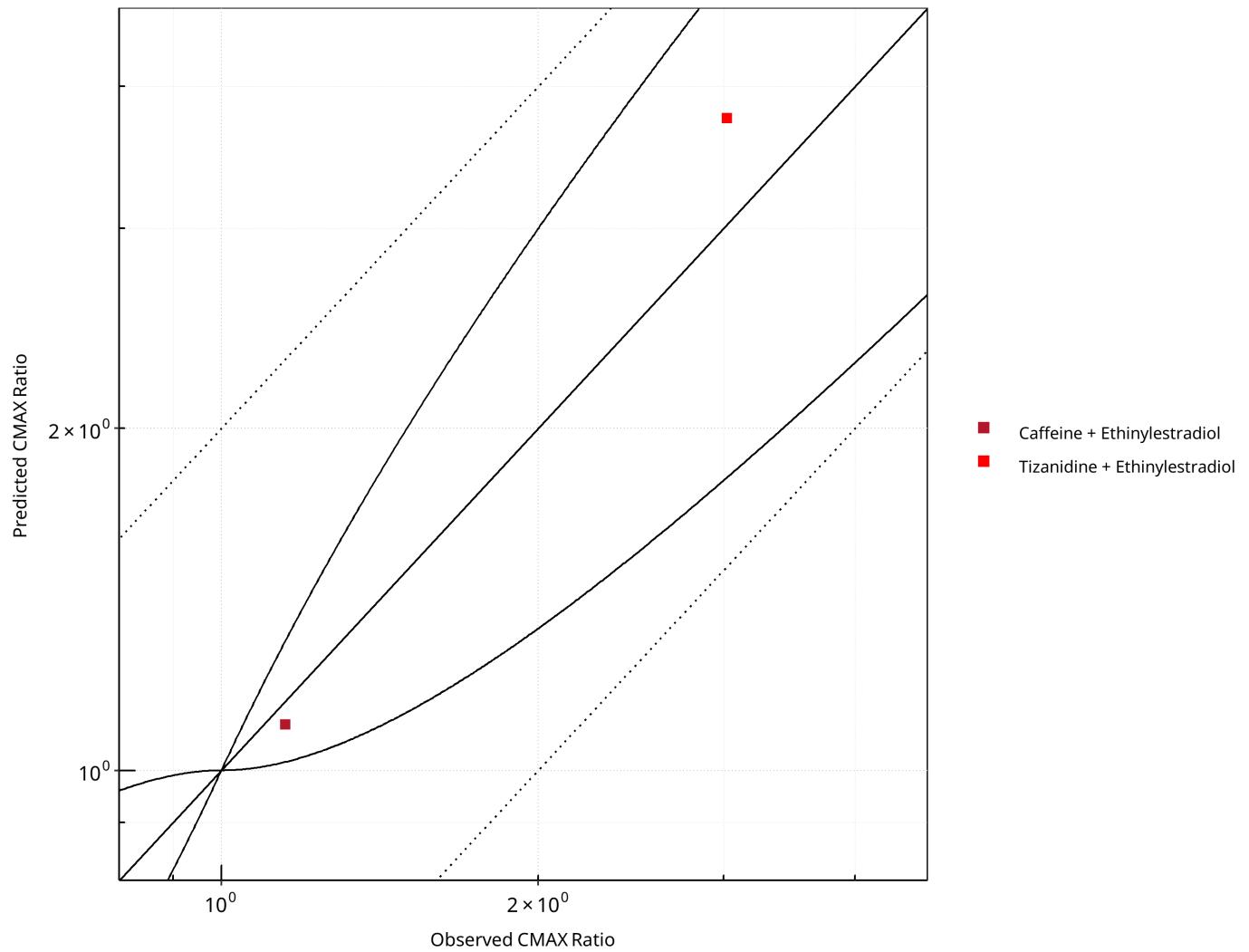


Figure 2-7: CYP1A2 DDI. Perpetrator: Ethinylestradiol. Predicted vs. Observed CMAX Ratio. ($\delta = 1$ in Guest et al. formula)

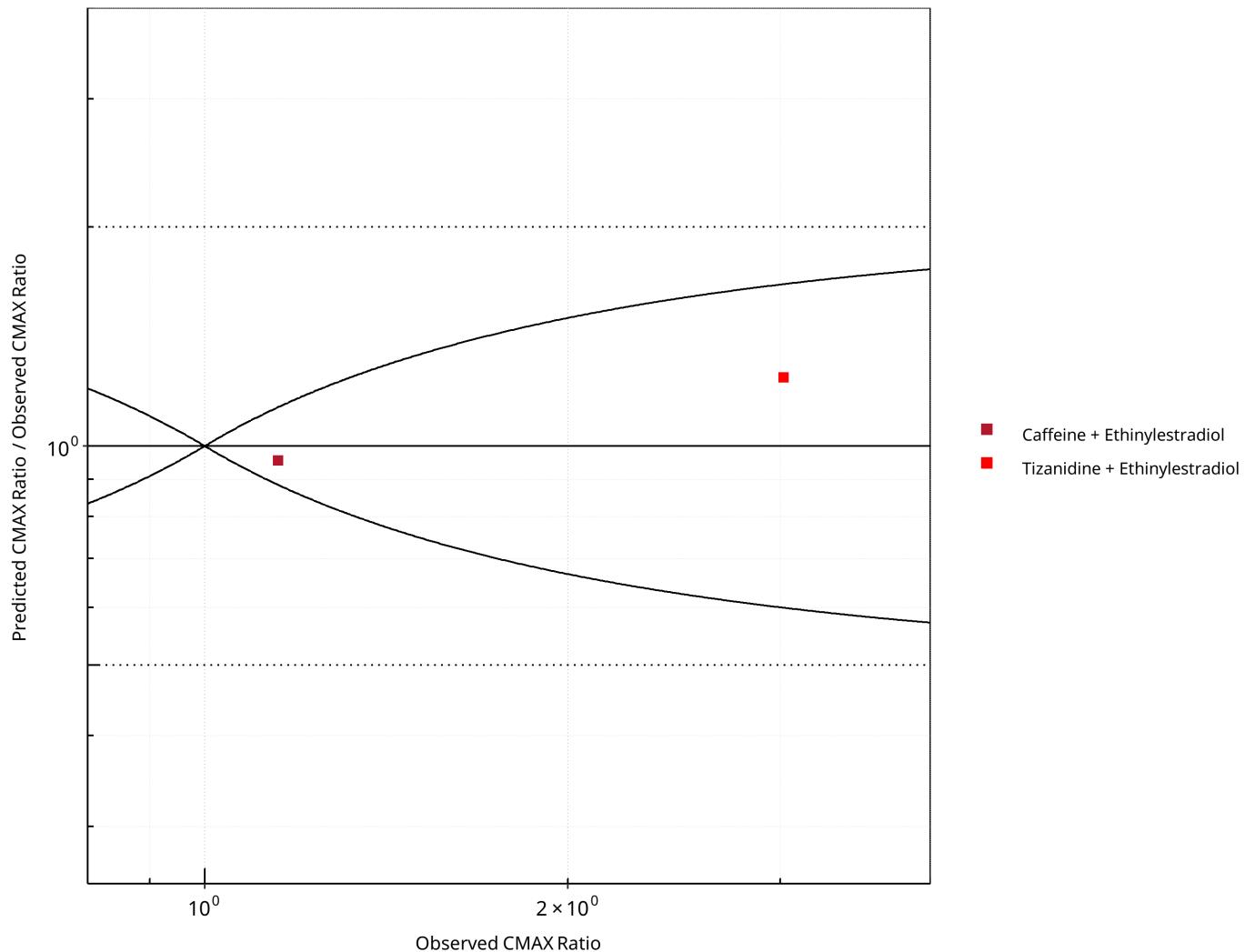


Figure 2-8: CYP1A2 DDI. Perpetrator: Ethinylestradiol. Predicted/Observed vs. Observed CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

Table 2-5: GMFE for CYP1A2 DDI Ratio

PK parameter	GMFE
AUC	1.76
CMAX	1.14

Table 2-6: Summary table for CYP1A2 DDI - AUC Ratio. ($\delta = 1$ in Guest *et al.* formula)

AUC	Number	Ratio [%]
Points total	2	-
Points within Guest <i>et al.</i>	1	50
Points within 2 fold	1	50

Table 2-7: Summary table for CYP1A2 DDI - CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

CMAX	Number	Ratio [%]
Points total	2	-
Points within Guest <i>et al.</i>	2	100
Points within 2 fold	2	100

2.1.2 Fluvoxamine

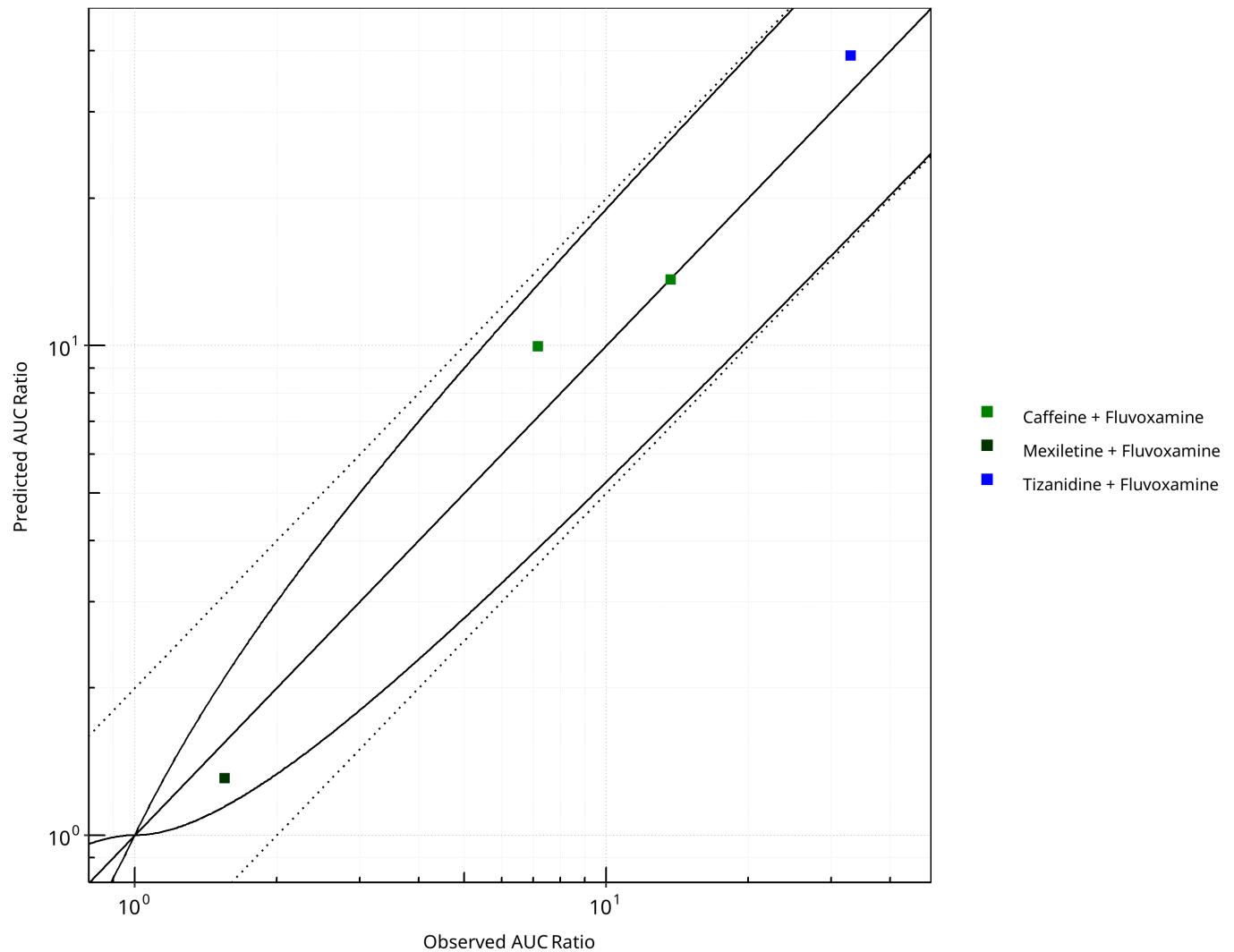


Figure 2-9: CYP1A2 DDI. Perpetrator: Fluvoxamine. Predicted vs. Observed AUC Ratio. ($\delta = 1$ in Guest *et al.* formula)

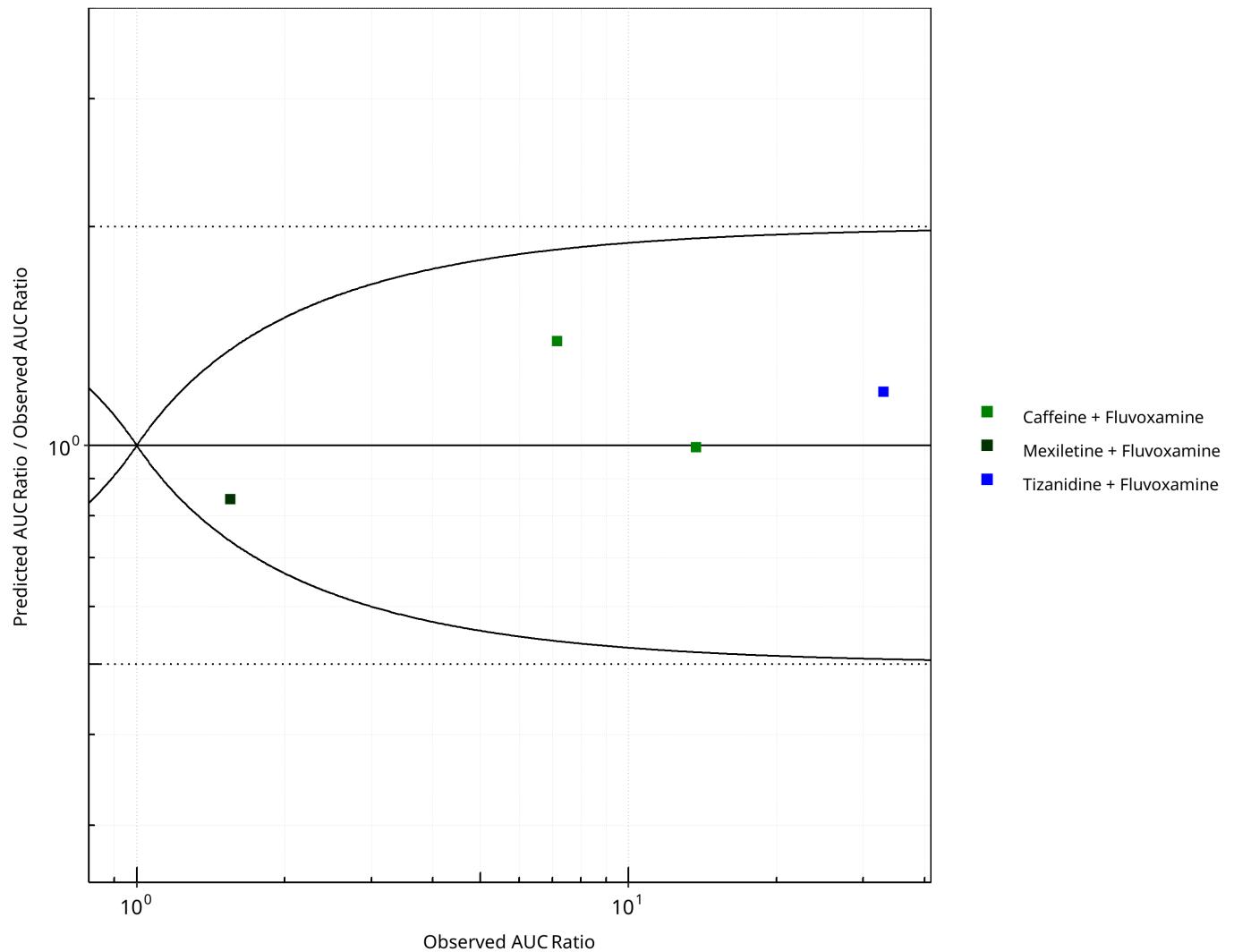


Figure 2-10: CYP1A2 DDI. Perpetrator: Fluvoxamine. Predicted/Observed vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

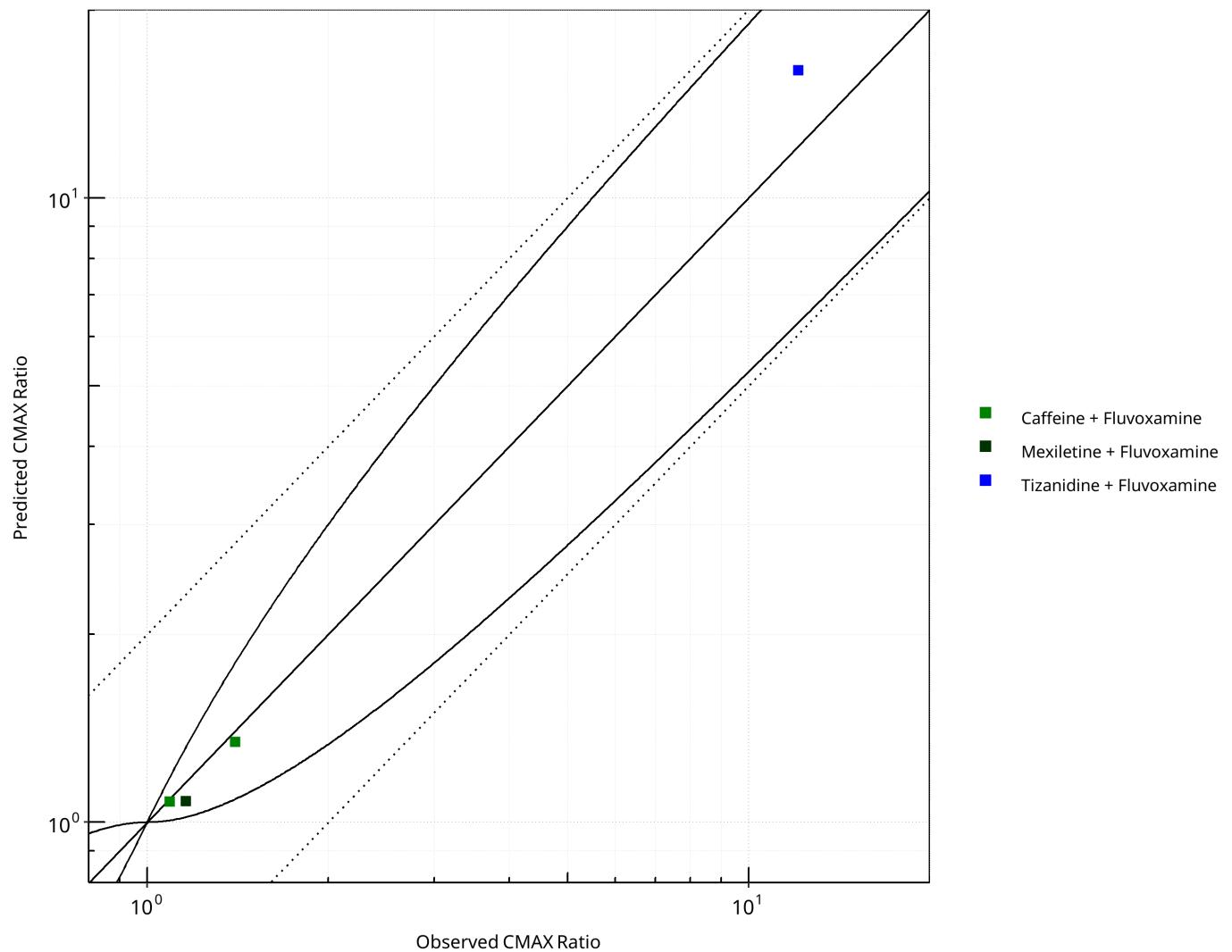


Figure 2-11: CYP1A2 DDI. Perpetrator: Fluvoxamine. Predicted vs. Observed CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

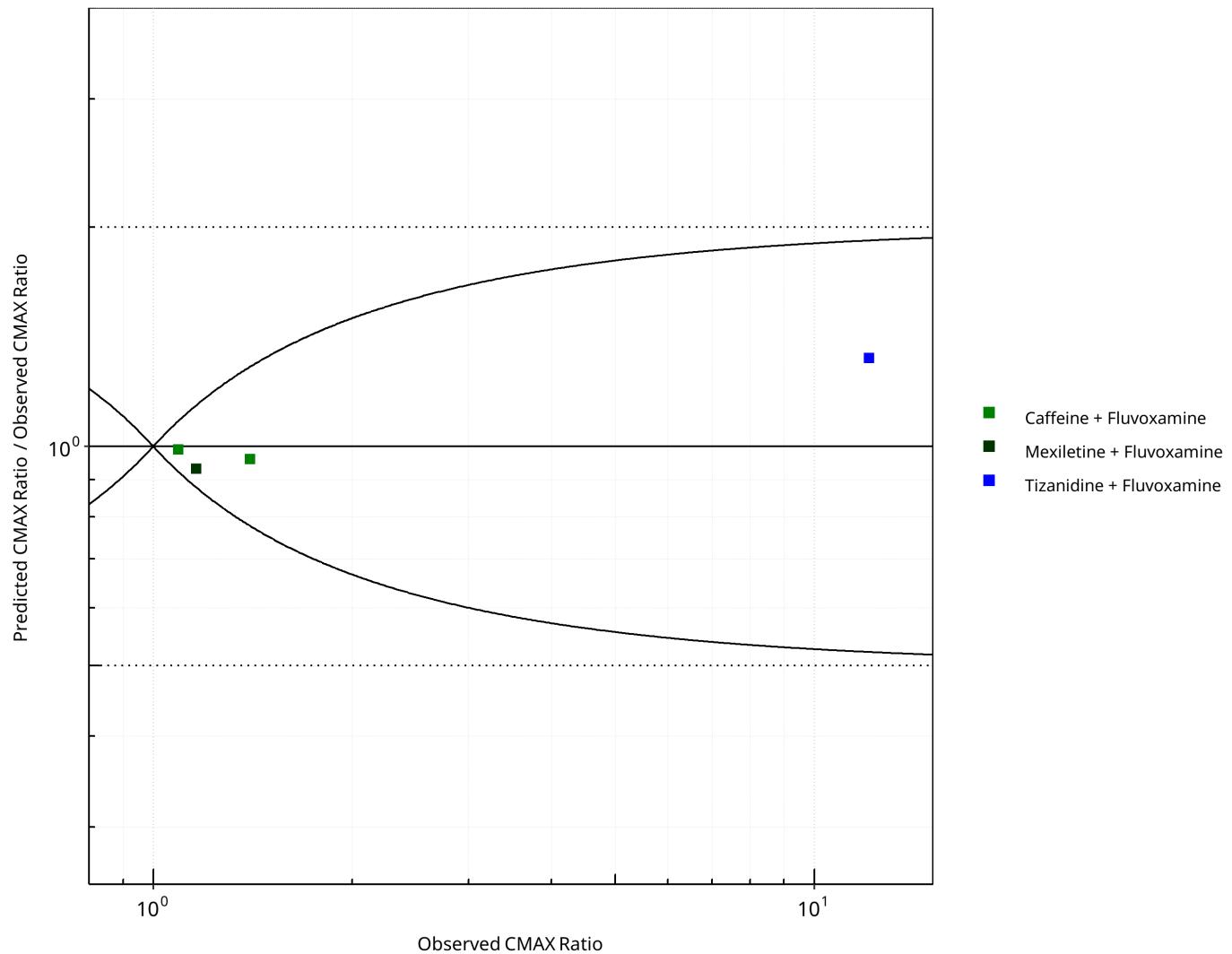


Figure 2-12: CYP1A2 DDI. Perpetrator: Fluvoxamine. Predicted/Observed vs. Observed CMAX Ratio. ($\delta = 1$ in Guest et al. formula)

Table 2-8: GMFE for CYP1A2 DDI Ratio

PK parameter	GMFE
AUC	1.18
CMAX	1.11

Table 2-9: Summary table for CYP1A2 DDI - AUC Ratio. ($\delta = 1$ in Guest et al. formula)

AUC	Number	Ratio [%]
Points total	4	-
Points within Guest <i>et al.</i>	4	100
Points within 2 fold	4	100

Table 2-10: Summary table for CYP1A2 DDI - CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

CMAX	Number	Ratio [%]
Points total	4	-
Points within Guest <i>et al.</i>	4	100
Points within 2 fold	4	100

2.1.3 Mexiletine

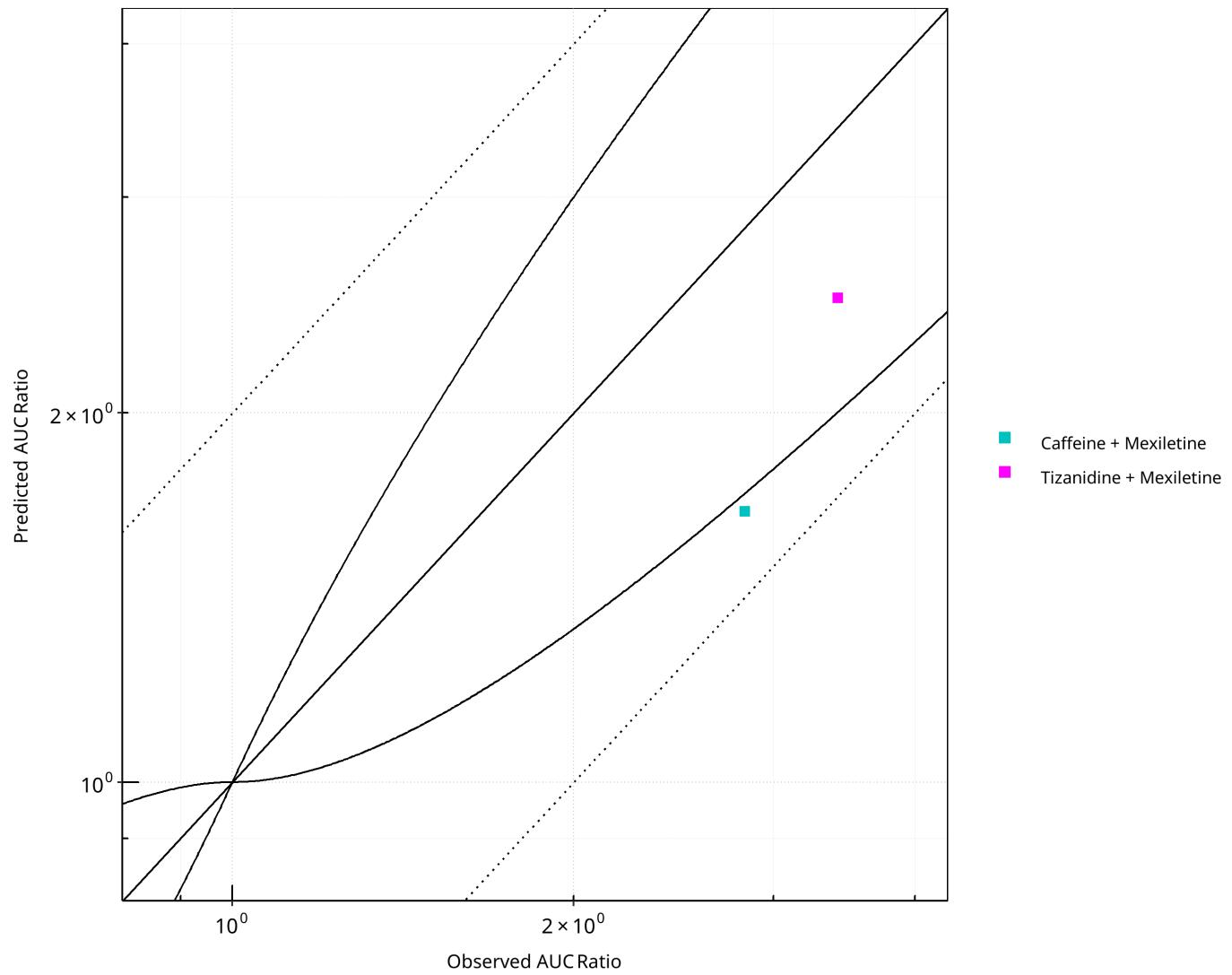


Figure 2-13: CYP1A2 DDI. Perpetrator: Mexiletine. Predicted vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

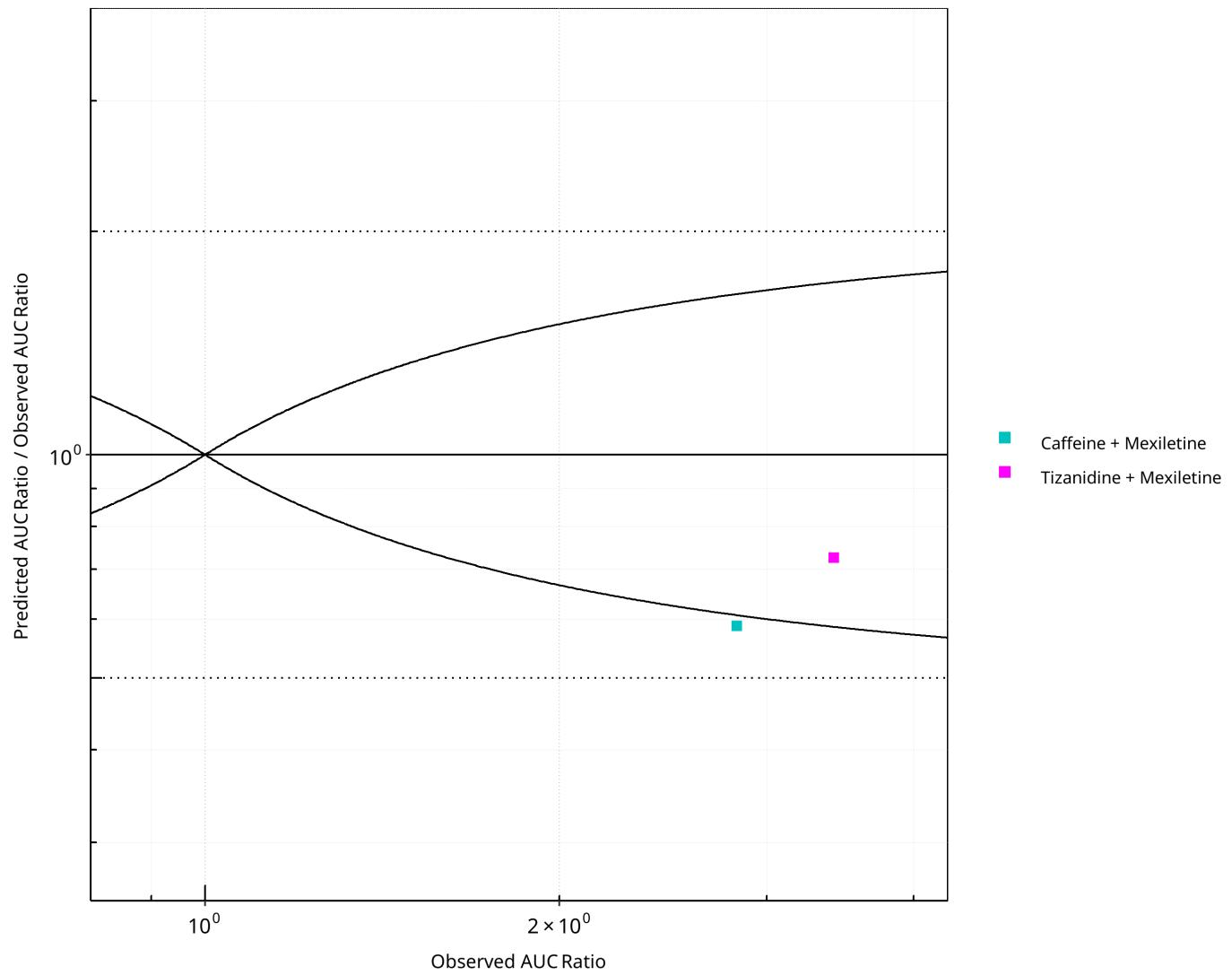


Figure 2-14: CYP1A2 DDI. Perpetrator: Mexiletine. Predicted/Observed vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

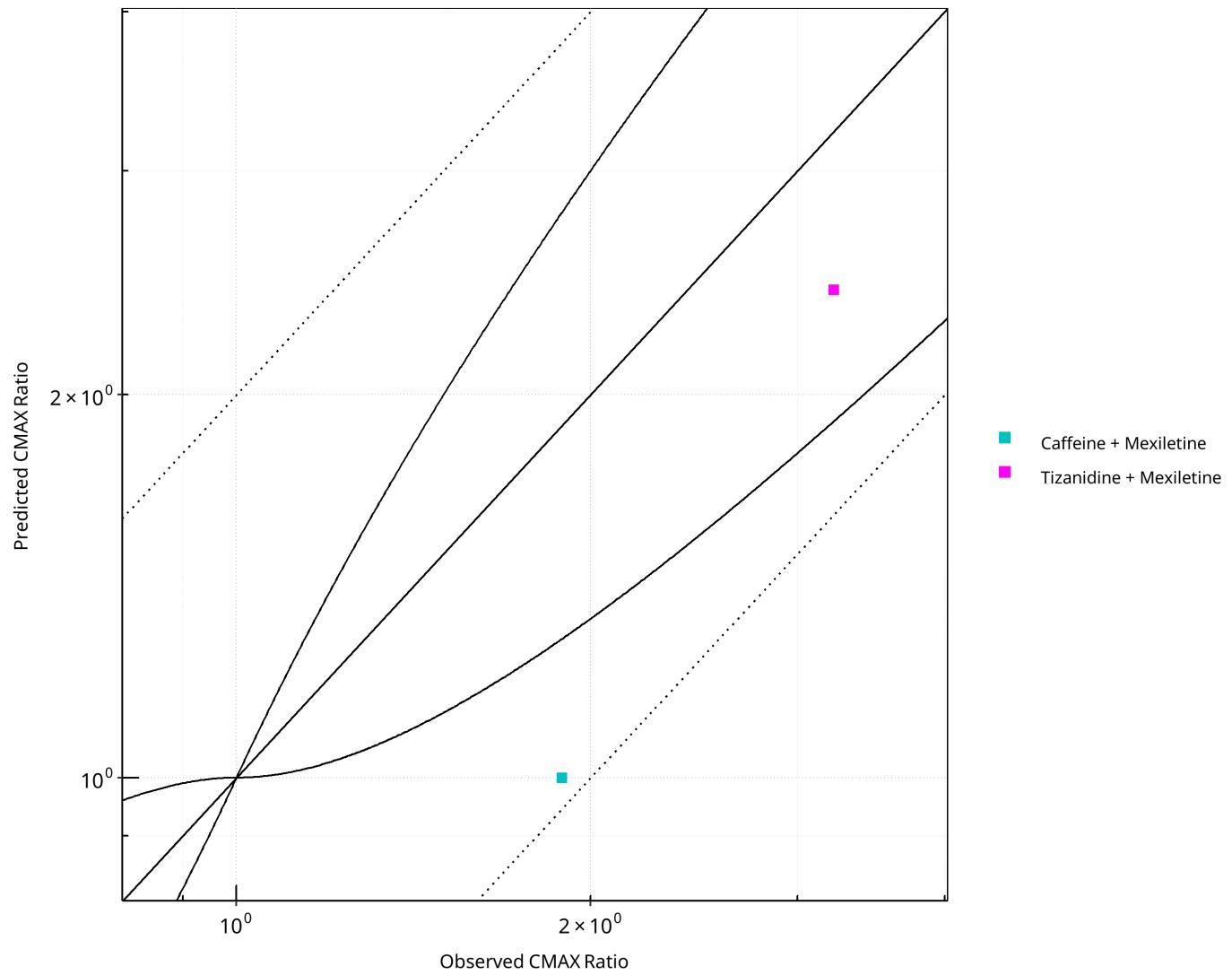


Figure 2-15: CYP1A2 DDI. Perpetrator: Mexiletine. Predicted vs. Observed CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

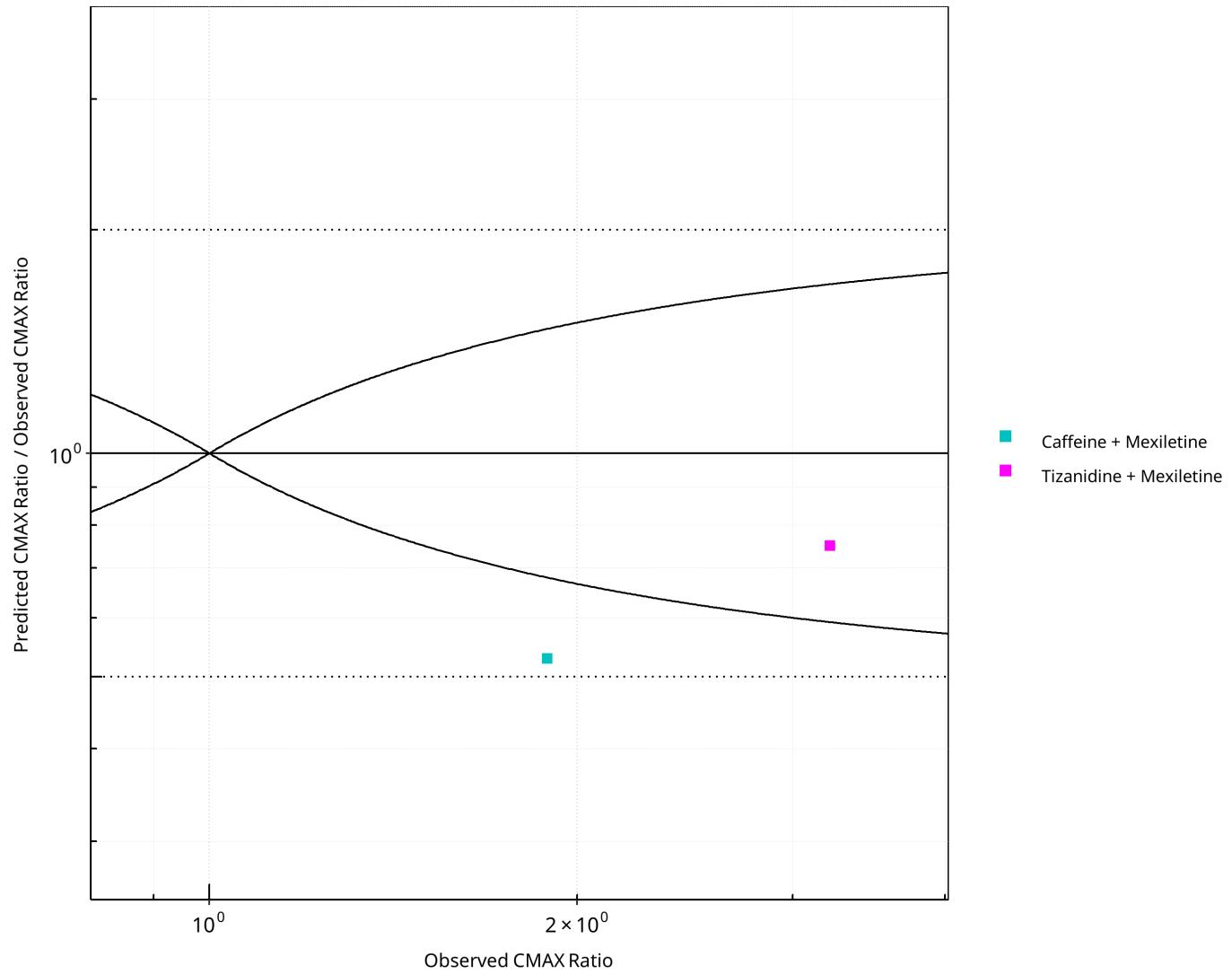


Figure 2-16: CYP1A2 DDI. Perpetrator: Mexiletine. Predicted/Observed vs. Observed CMAX Ratio. ($\delta = 1$ in Guest et al. formula)

Table 2-11: GMFE for CYP1A2 DDI Ratio

PK parameter	GMFE
AUC	1.53
CMAX	1.59

Table 2-12: Summary table for CYP1A2 DDI - AUC Ratio. ($\delta = 1$ in Guest et al. formula)

AUC	Number	Ratio [%]
Points total	2	-
Points within Guest <i>et al.</i>	1	50
Points within 2 fold	2	100

Table 2-13: Summary table for CYP1A2 DDI - CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

CMAX	Number	Ratio [%]
Points total	2	-
Points within Guest <i>et al.</i>	1	50
Points within 2 fold	2	100

2.2 Victim

2.2.1 Caffeine

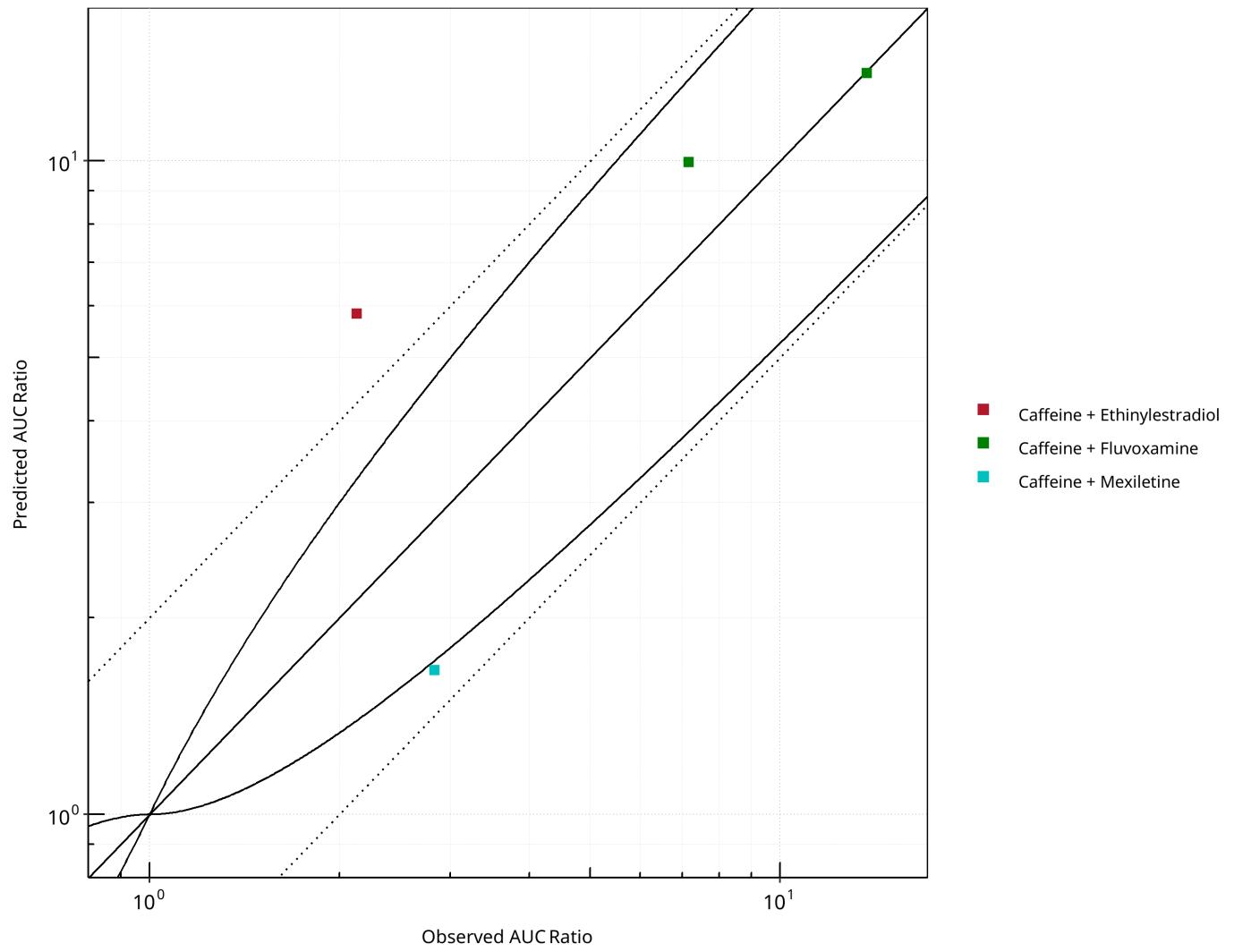


Figure 2-17: CYP1A2 DDI. Victim: Caffeine. Predicted vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

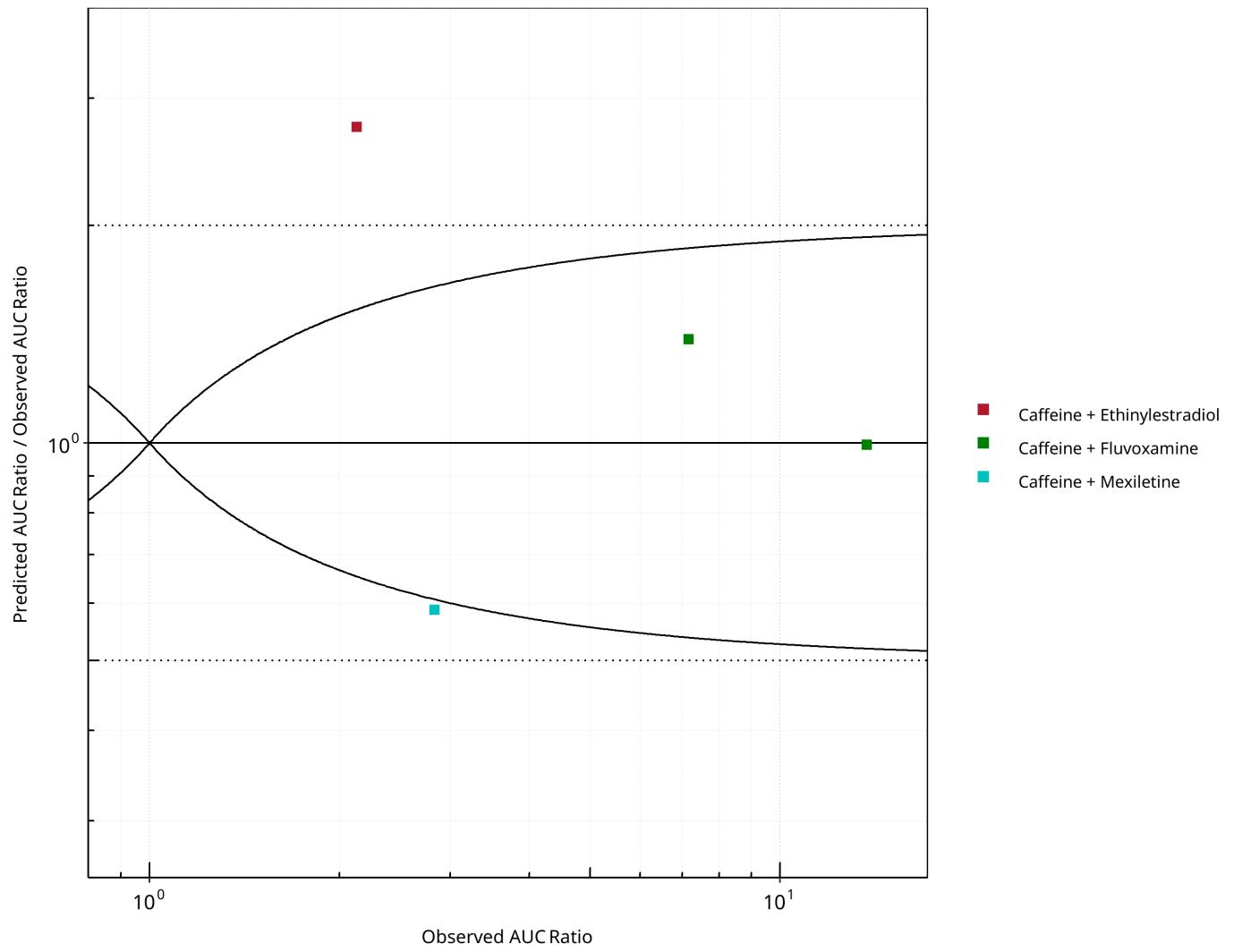


Figure 2-18: CYP1A2 DDI. Victim: Caffeine. Predicted/Observed vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

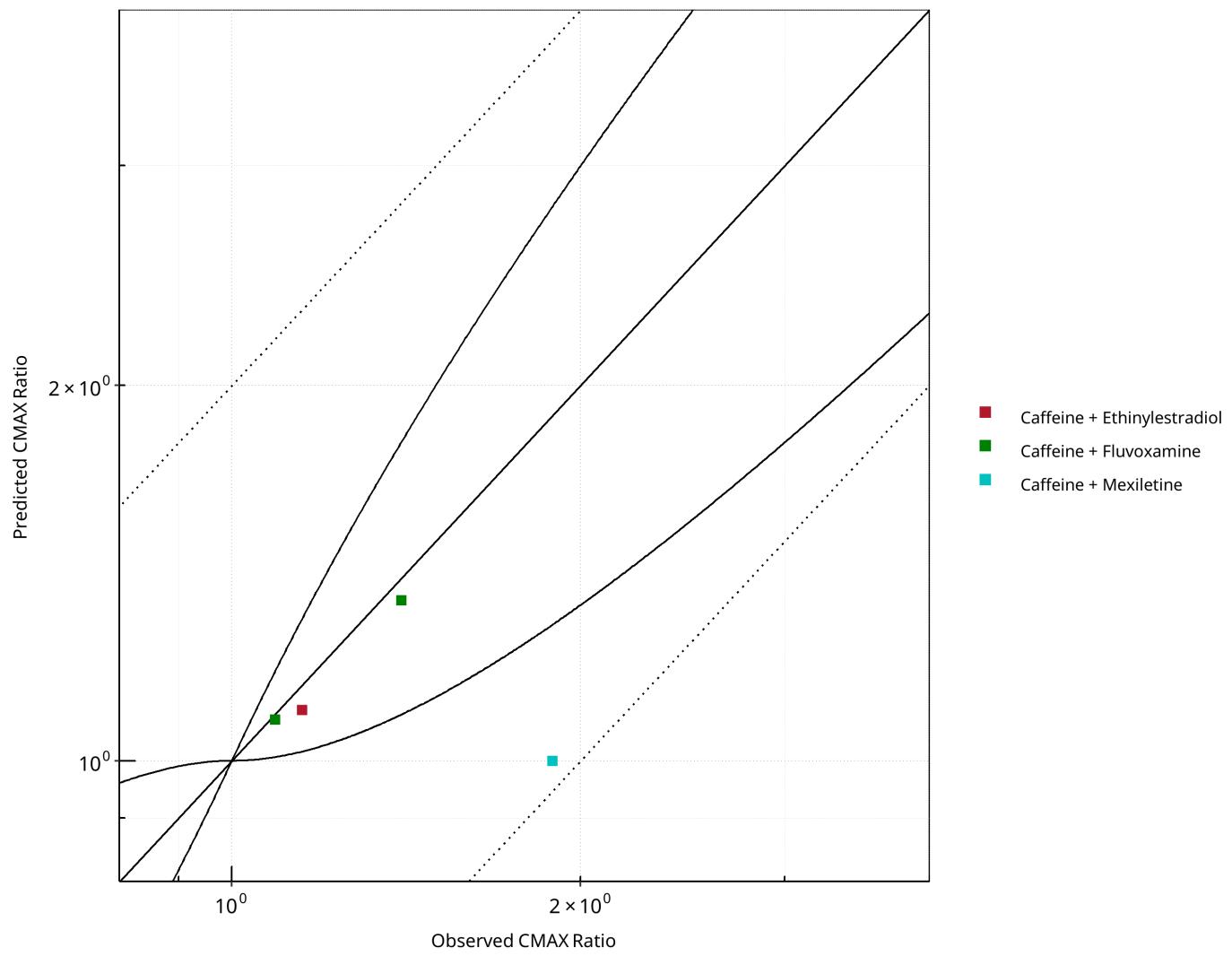


Figure 2-19: CYP1A2 DDI. Victim: Caffeine. Predicted vs. Observed CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

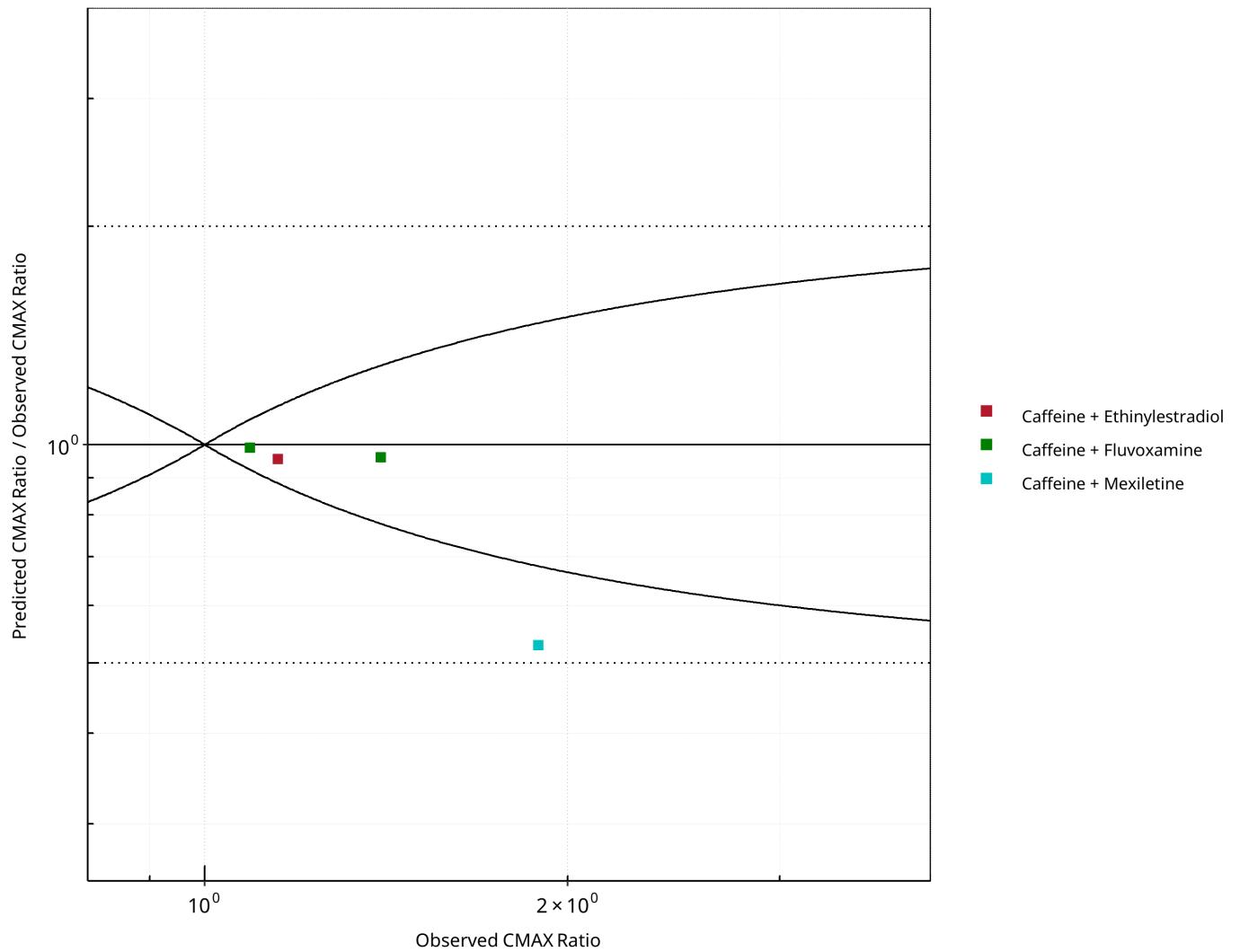


Figure 2-20: CYP1A2 DDI. Victim: Caffeine. Predicted/Observed vs. Observed CMAX Ratio. ($\delta = 1$ in Guest et al. formula)

Table 2-14: GMFE for CYP1A2 DDI Ratio

PK parameter	GMFE
AUC	1.60
CMAX	1.20

Table 2-15: Summary table for CYP1A2 DDI - AUC Ratio. ($\delta = 1$ in Guest et al. formula)

AUC	Number	Ratio [%]
Points total	4	-
Points within Guest <i>et al.</i>	2	50
Points within 2 fold	3	75

Table 2-16: Summary table for CYP1A2 DDI - CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

CMAX	Number	Ratio [%]
Points total	4	-
Points within Guest <i>et al.</i>	3	75
Points within 2 fold	4	100

2.2.2 Mexiletine

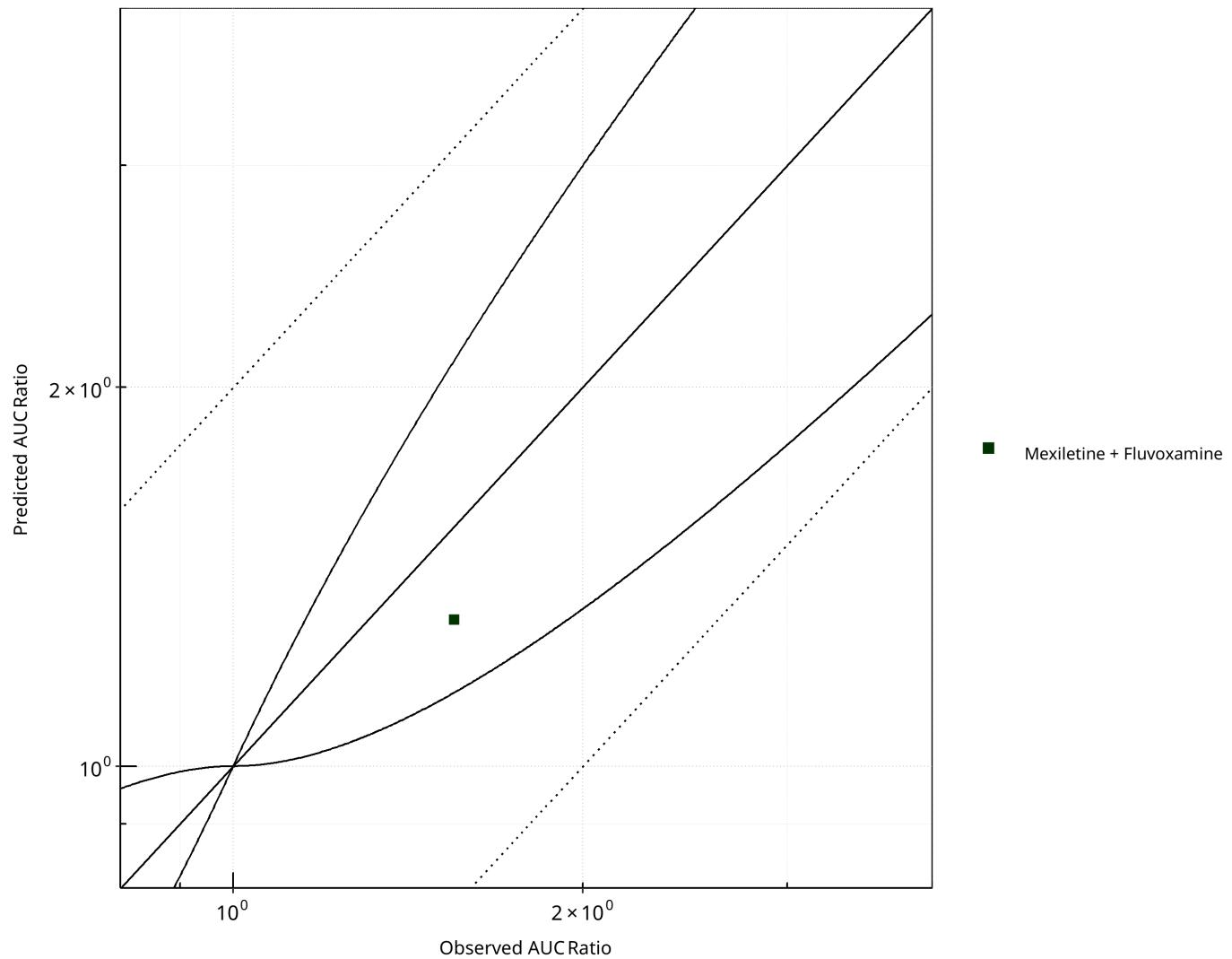


Figure 2-21: CYP1A2 DDI. Victim: Mexiletine. Predicted vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

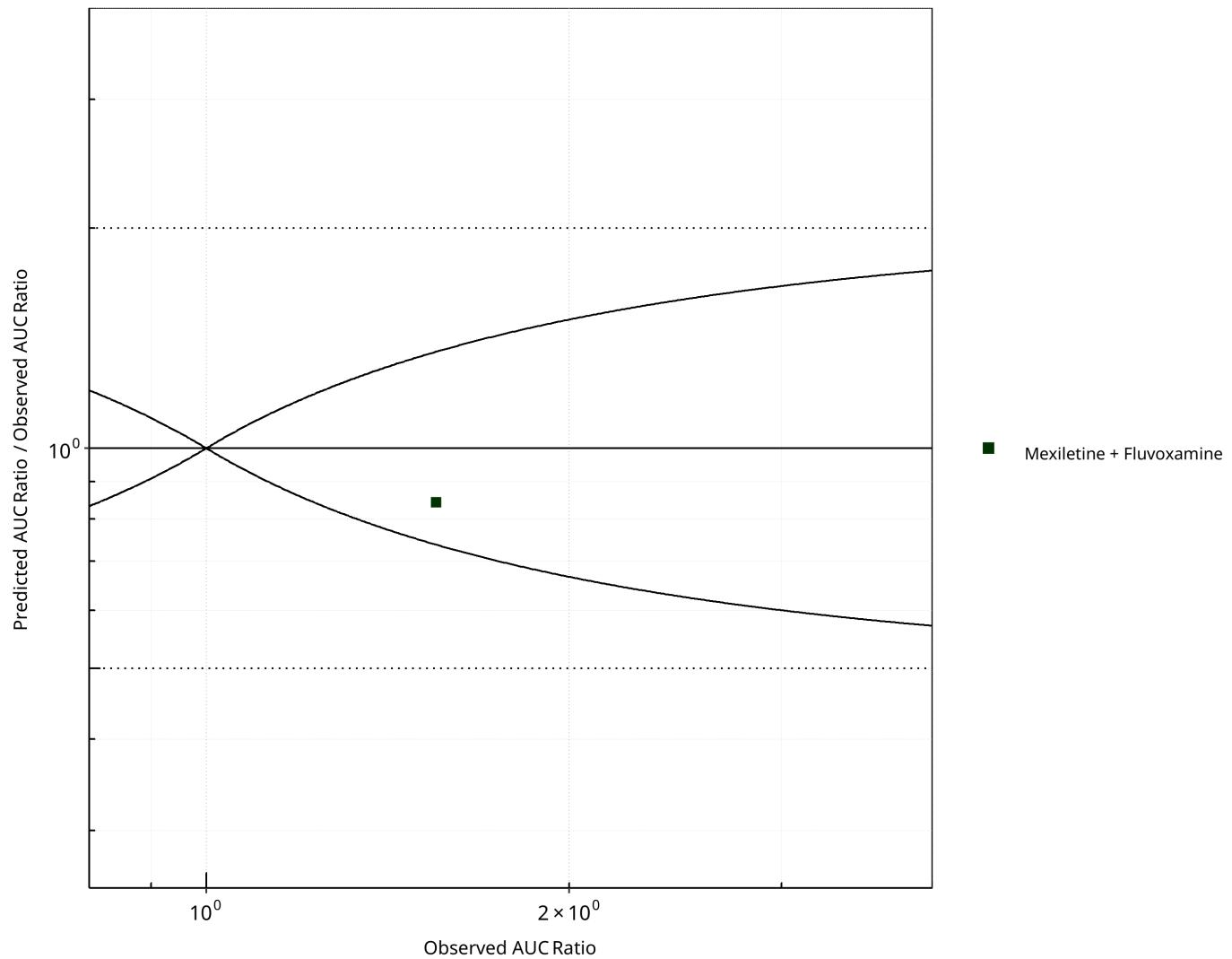


Figure 2-22: CYP1A2 DDI. Victim: Mexiletine. Predicted/Observed vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

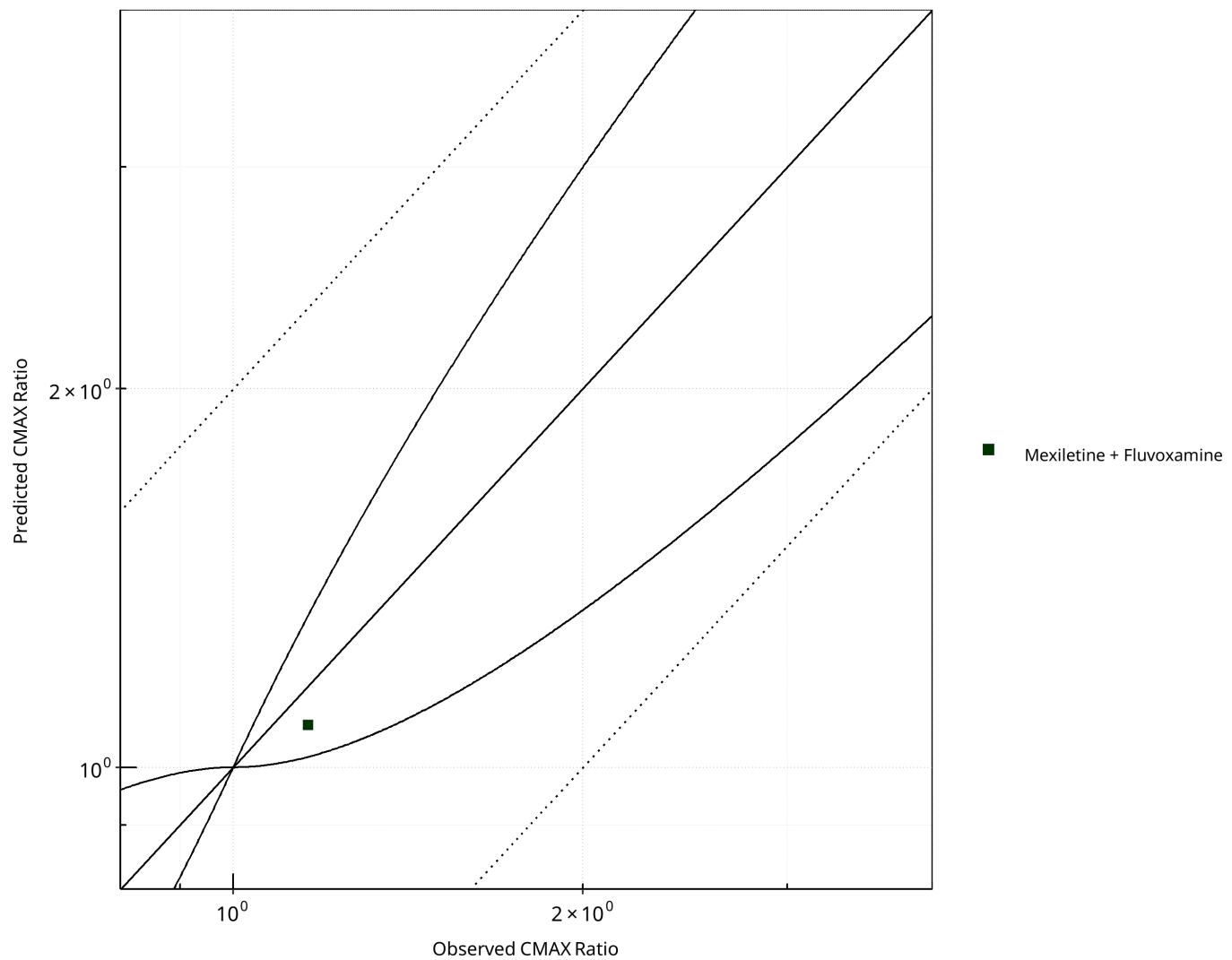


Figure 2-23: CYP1A2 DDI. Victim: Mexiletine. Predicted vs. Observed CMAX Ratio. ($\delta = 1$ in Guest et al. formula)

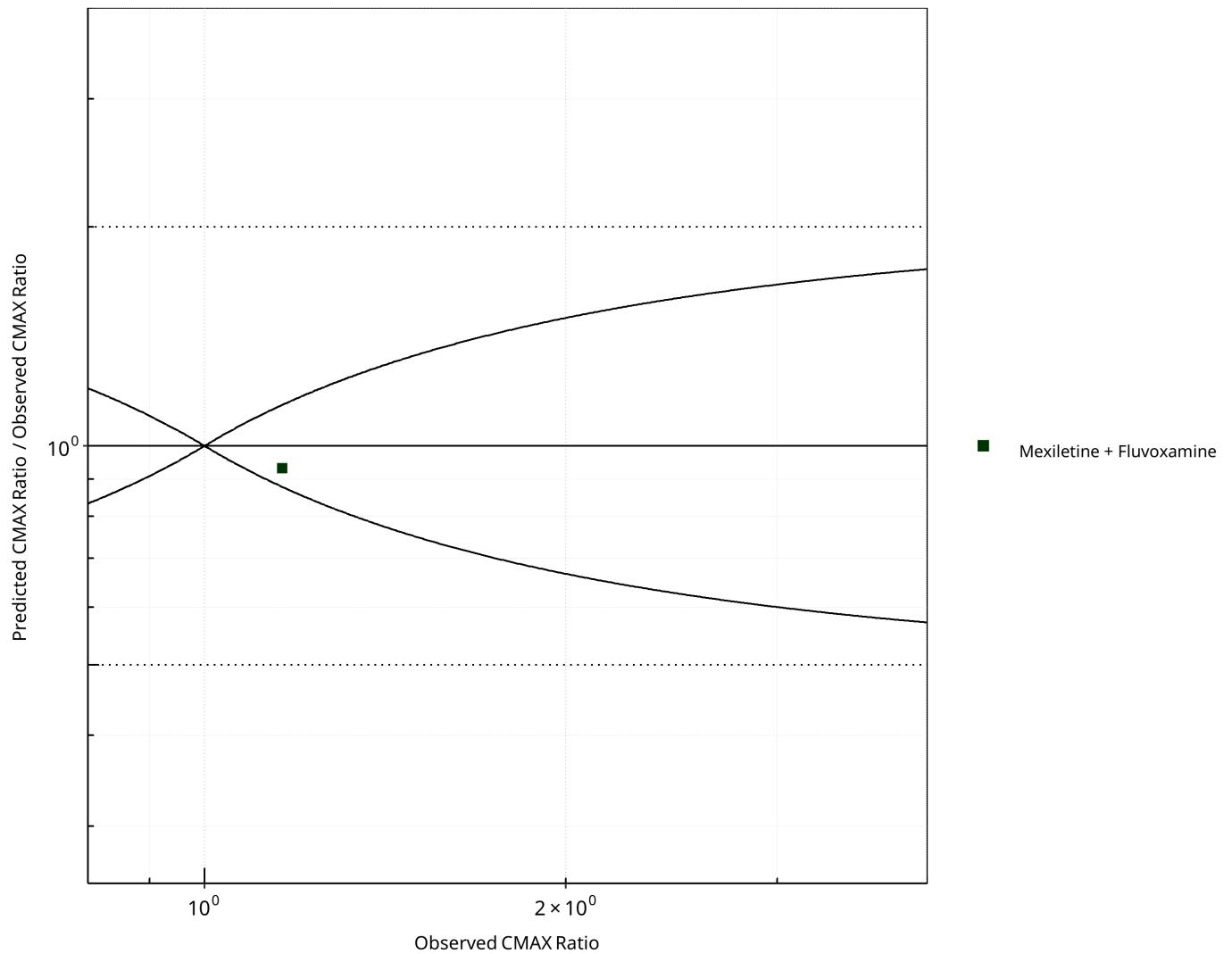


Figure 2-24: CYP1A2 DDI. Victim: Mexiletine. Predicted/Observed vs. Observed CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

Table 2-17: GMFE for CYP1A2 DDI Ratio

PK parameter	GMFE
AUC	1.19
CMAX	1.07

Table 2-18: Summary table for CYP1A2 DDI - AUC Ratio. ($\delta = 1$ in Guest *et al.* formula)

AUC	Number	Ratio [%]
Points total	1	-
Points within Guest <i>et al.</i>	1	100
Points within 2 fold	1	100

Table 2-19: Summary table for CYP1A2 DDI - CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

CMAX	Number	Ratio [%]
Points total	1	-
Points within Guest <i>et al.</i>	1	100
Points within 2 fold	1	100

2.2.3 Tizanidine

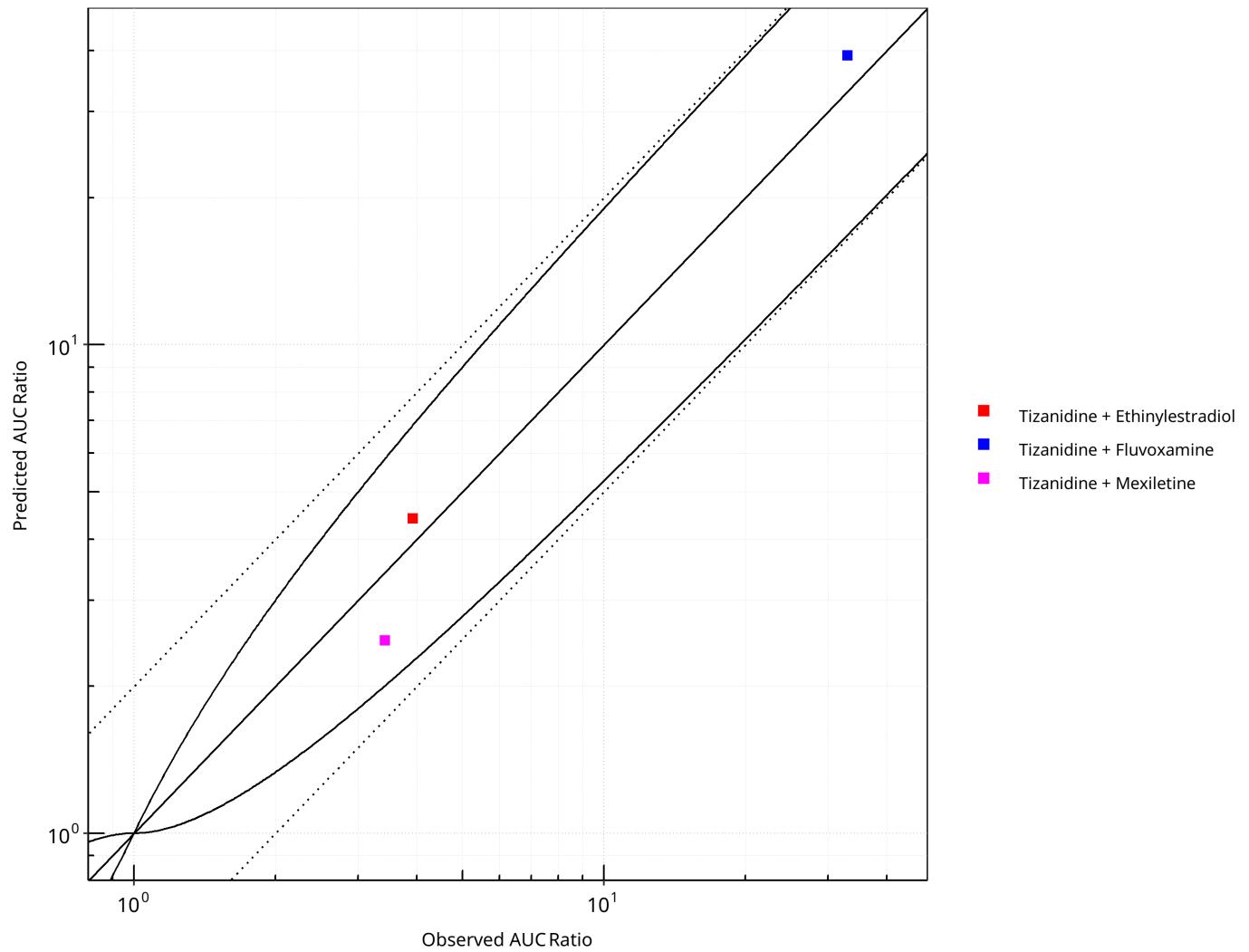


Figure 2-25: CYP1A2 DDI. Victim: Tizanidine. Predicted vs. Observed AUC Ratio. ($\delta = 1$ in Guest *et al.* formula)

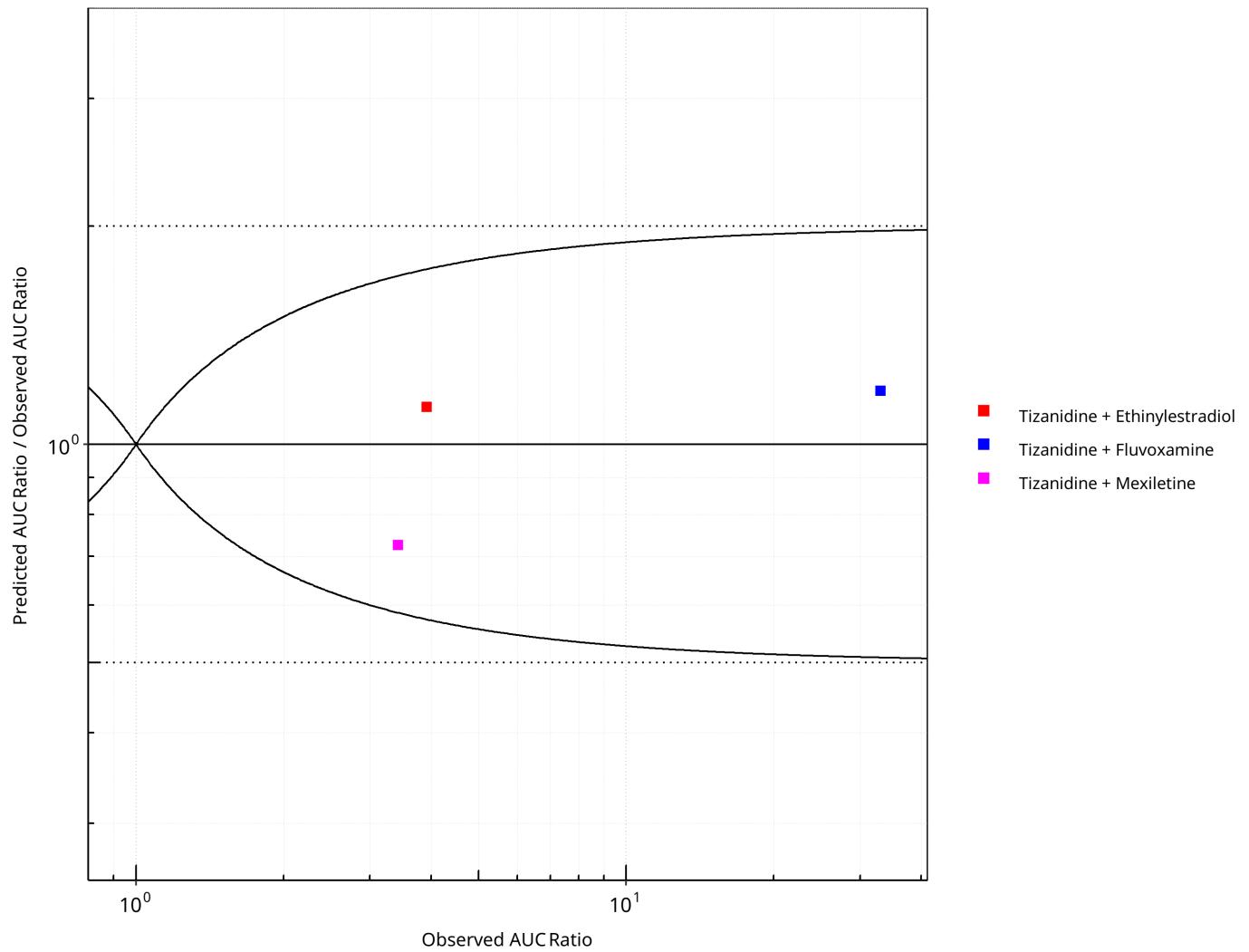


Figure 2-26: CYP1A2 DDI. Victim: Tizanidine. Predicted/Observed vs. Observed AUC Ratio. ($\delta = 1$ in Guest et al. formula)

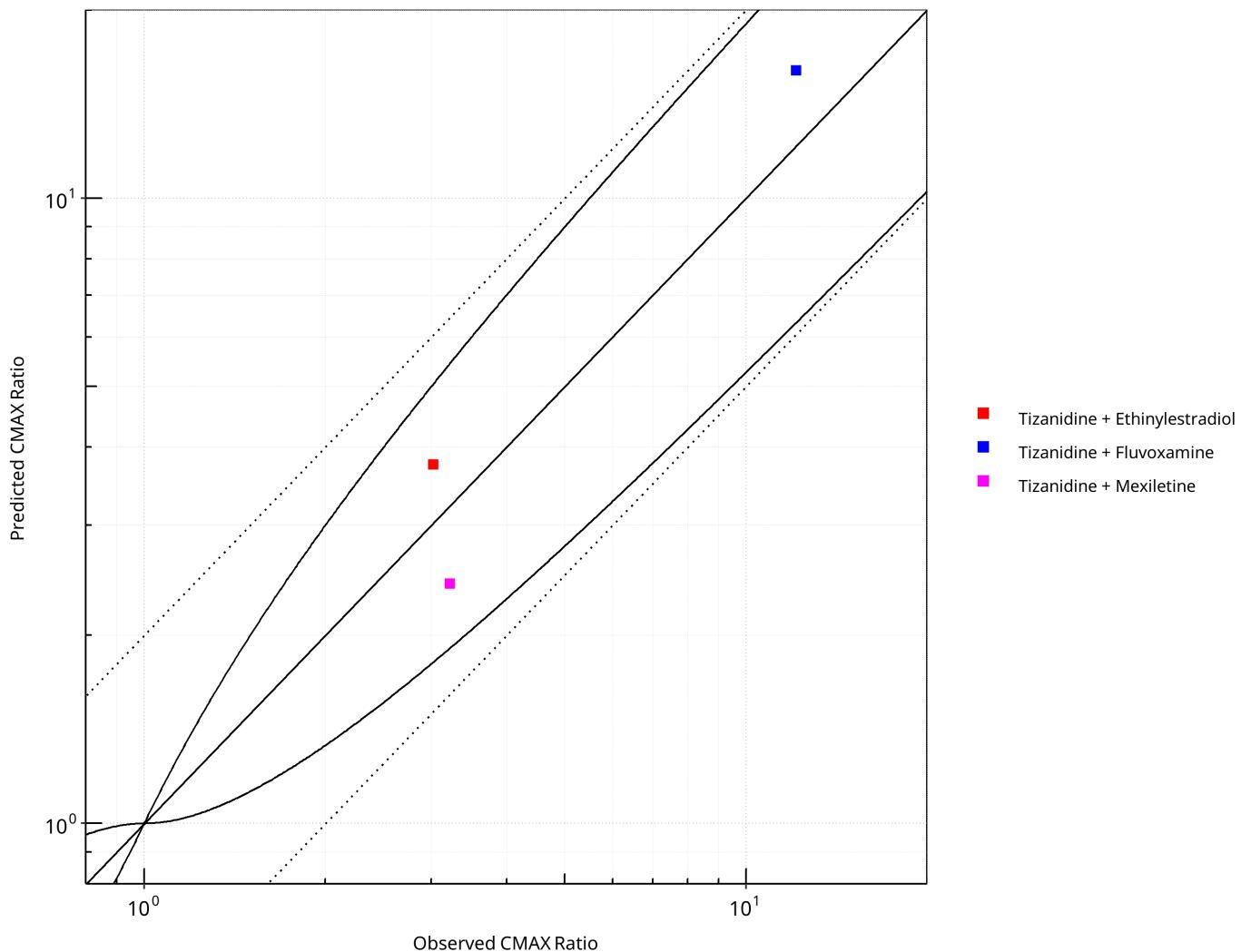


Figure 2-27: CYP1A2 DDI. Victim: Tizanidine. Predicted vs. Observed CMAX Ratio. ($\delta = 1$ in Guest et al. formula)

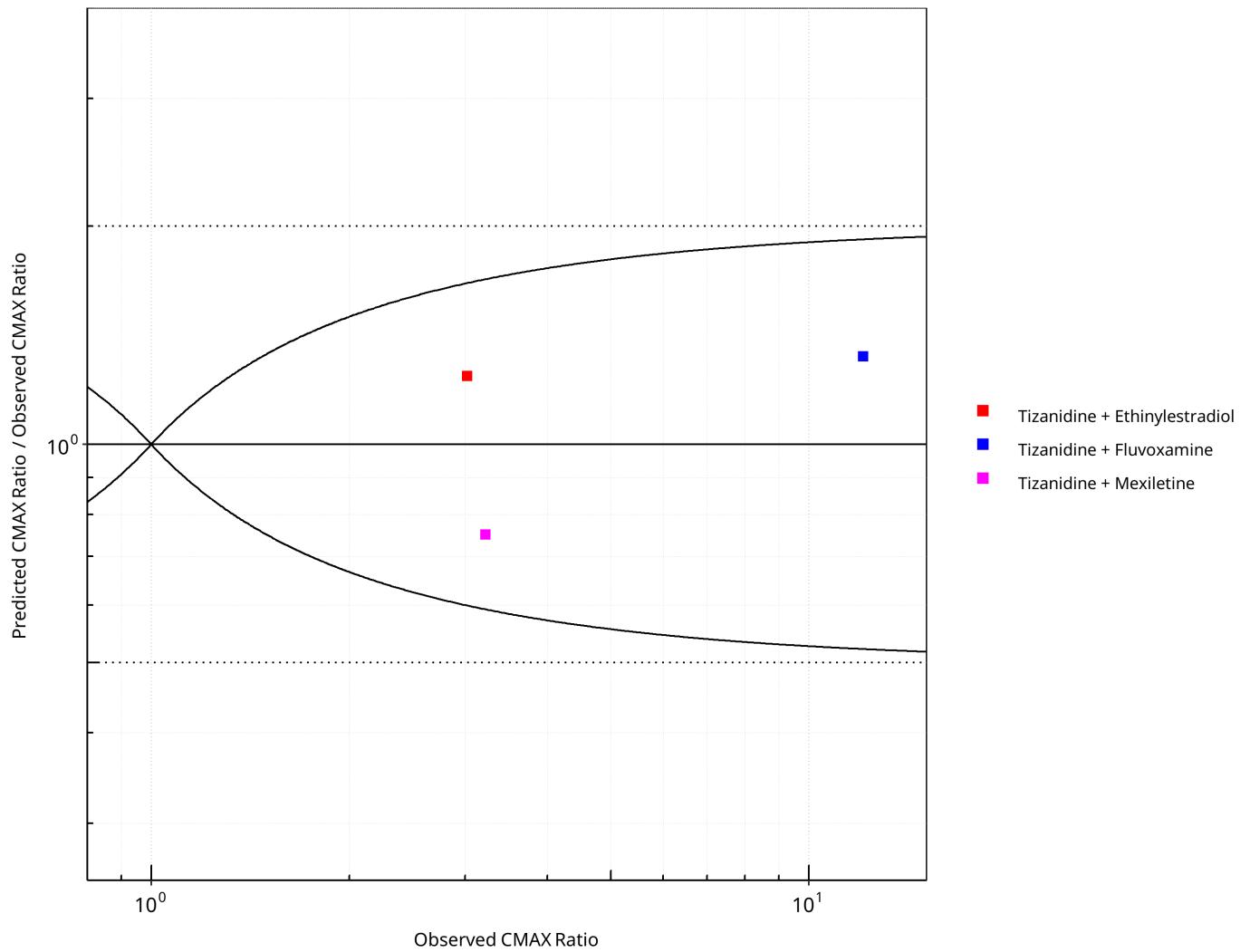


Figure 2-28: CYP1A2 DDI. Victim: Tizanidine. Predicted/Observed vs. Observed CMAX Ratio. ($\delta = 1$ in Guest et al. formula)

Table 2-20: GMFE for CYP1A2 DDI Ratio

PK parameter	GMFE
AUC	1.23
CMAX	1.30

Table 2-21: Summary table for CYP1A2 DDI - AUC Ratio. ($\delta = 1$ in Guest et al. formula)

AUC	Number	Ratio [%]
Points total	3	-
Points within Guest <i>et al.</i>	3	100
Points within 2 fold	3	100

Table 2-22: Summary table for CYP1A2 DDI - CMAX Ratio. ($\delta = 1$ in Guest *et al.* formula)

CMAX	Number	Ratio [%]
Points total	3	-
Points within Guest <i>et al.</i>	3	100
Points within 2 fold	3	100

3 Concentration-Time Profiles

The following section shows concentration time profiles of the victim drugs of the simulated DDI studies in comparison to observed data.

3.1 Tizanidine - Ethinylestradiol DDI

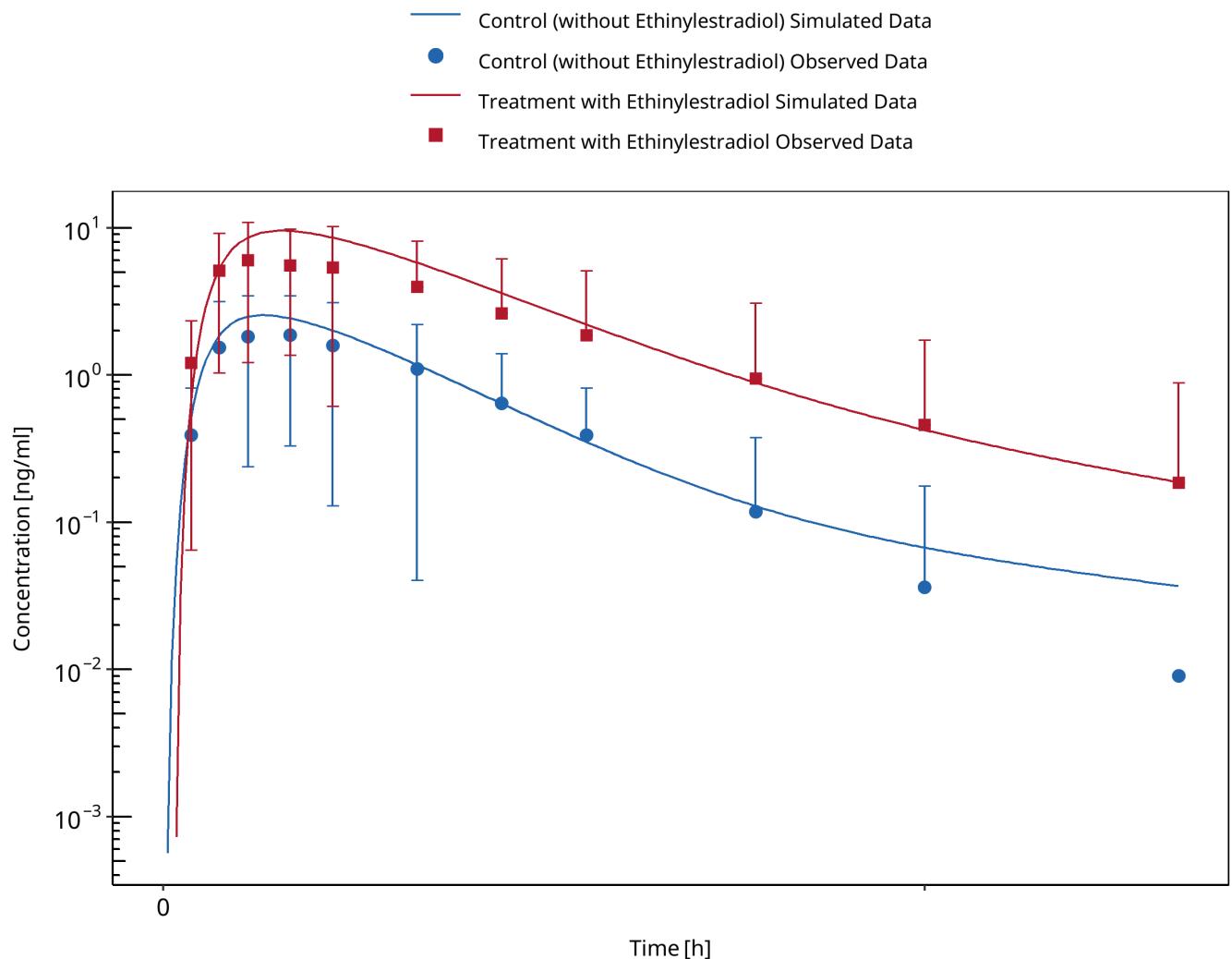


Figure 3-1: Granfors 2005 (Tizanidine 4 mg po)

3.2 Caffeine - Fluvoxamine DDI

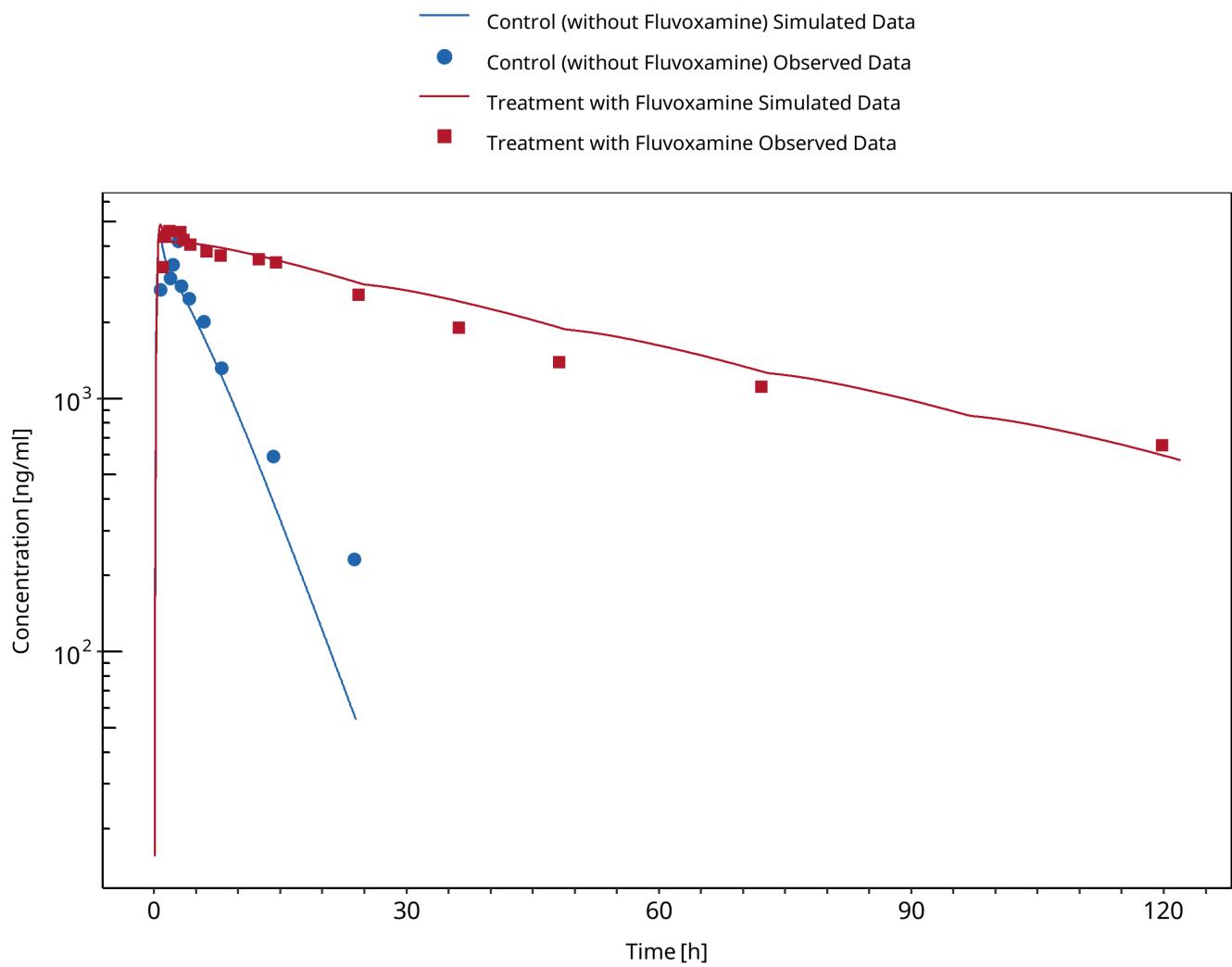


Figure 3-2: Jeppesen 1996 (Caffeine 200 mg po)

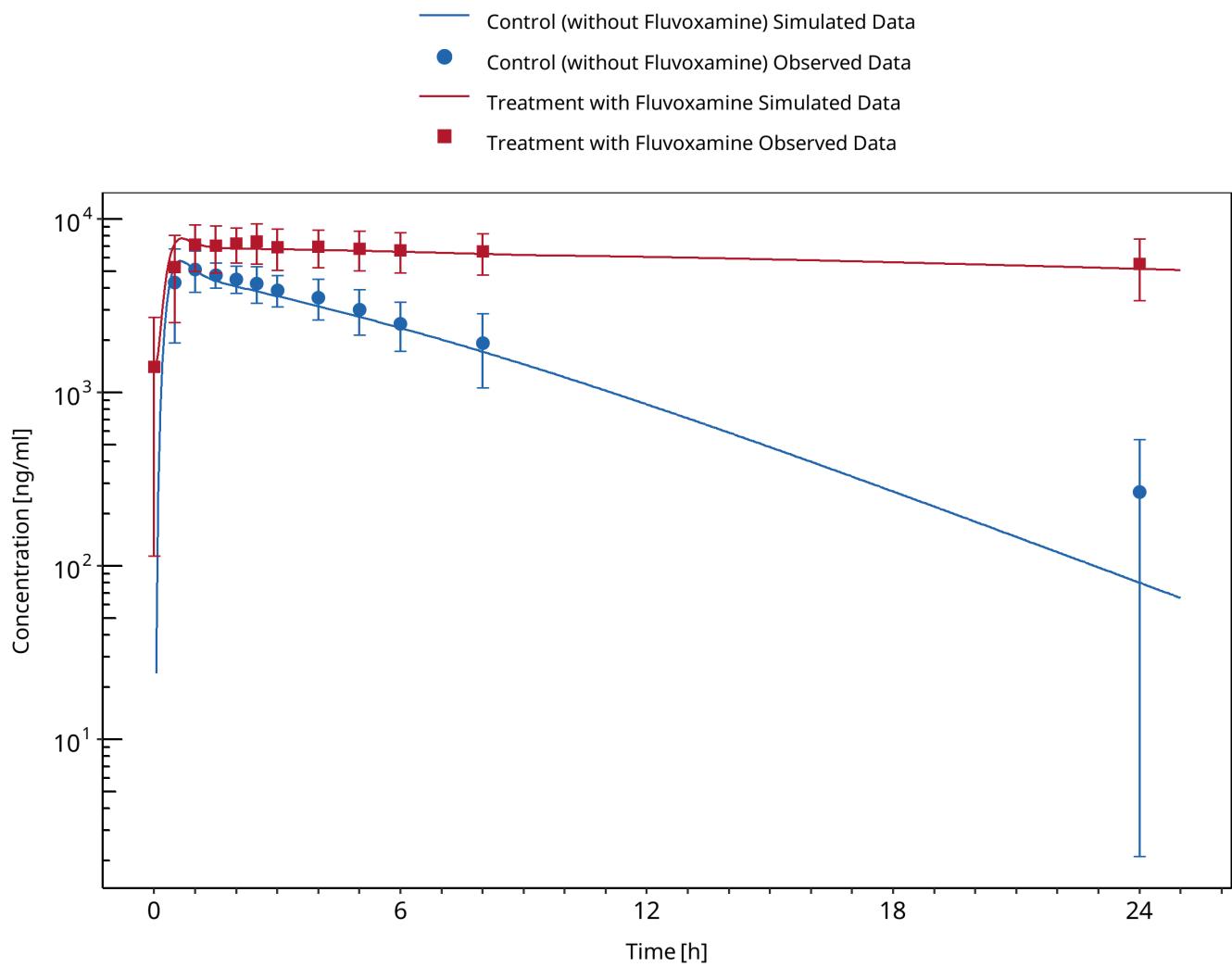


Figure 3-3: Culm-Merdeck 2005 (Caffeine 250 mg po)

3.3 Tizanidine - Fluvoxamine DDI

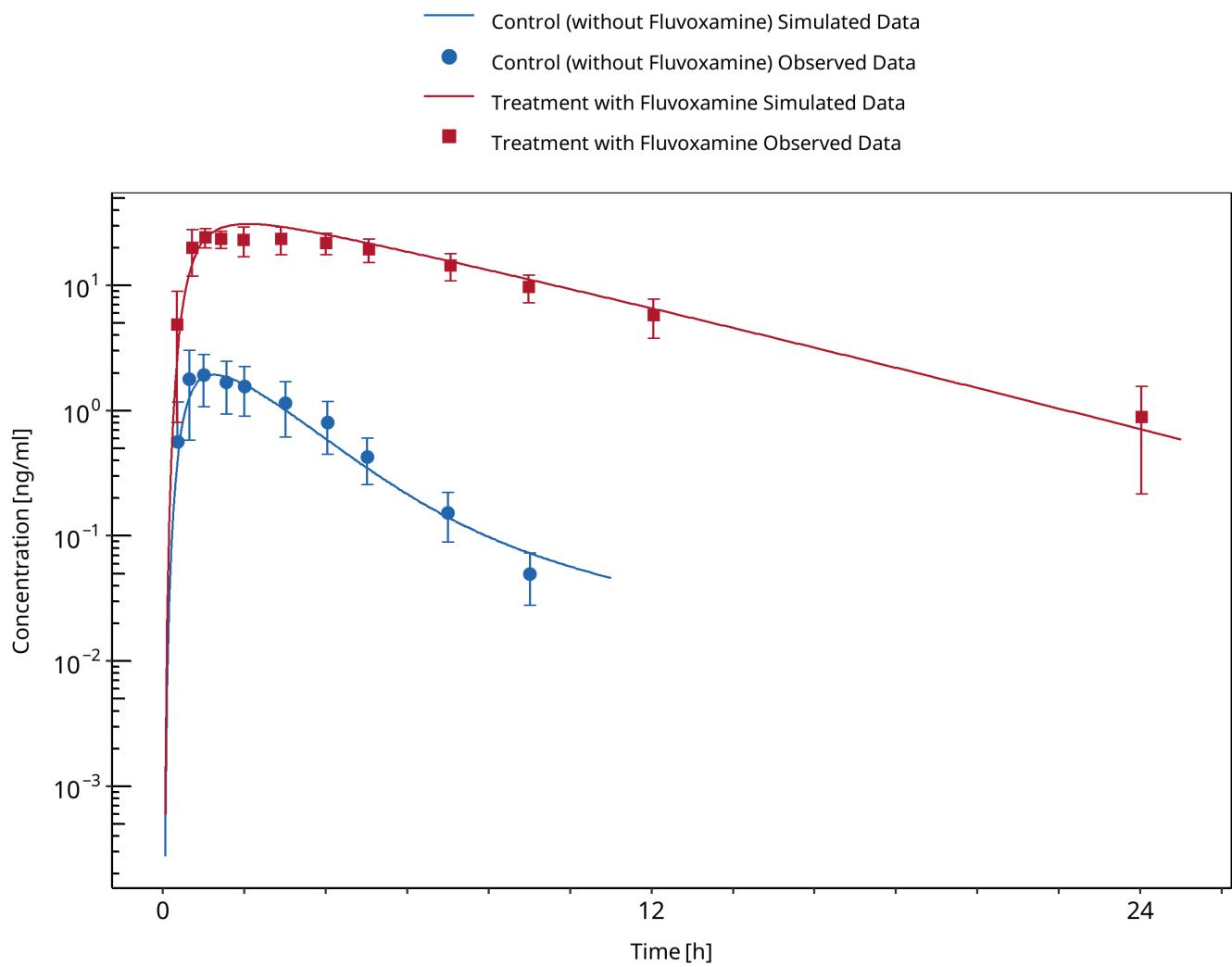


Figure 3-4: Granfors 2004 (Tizanidine 4 mg po)

3.4 Tizanidine - Mexiletine DDI

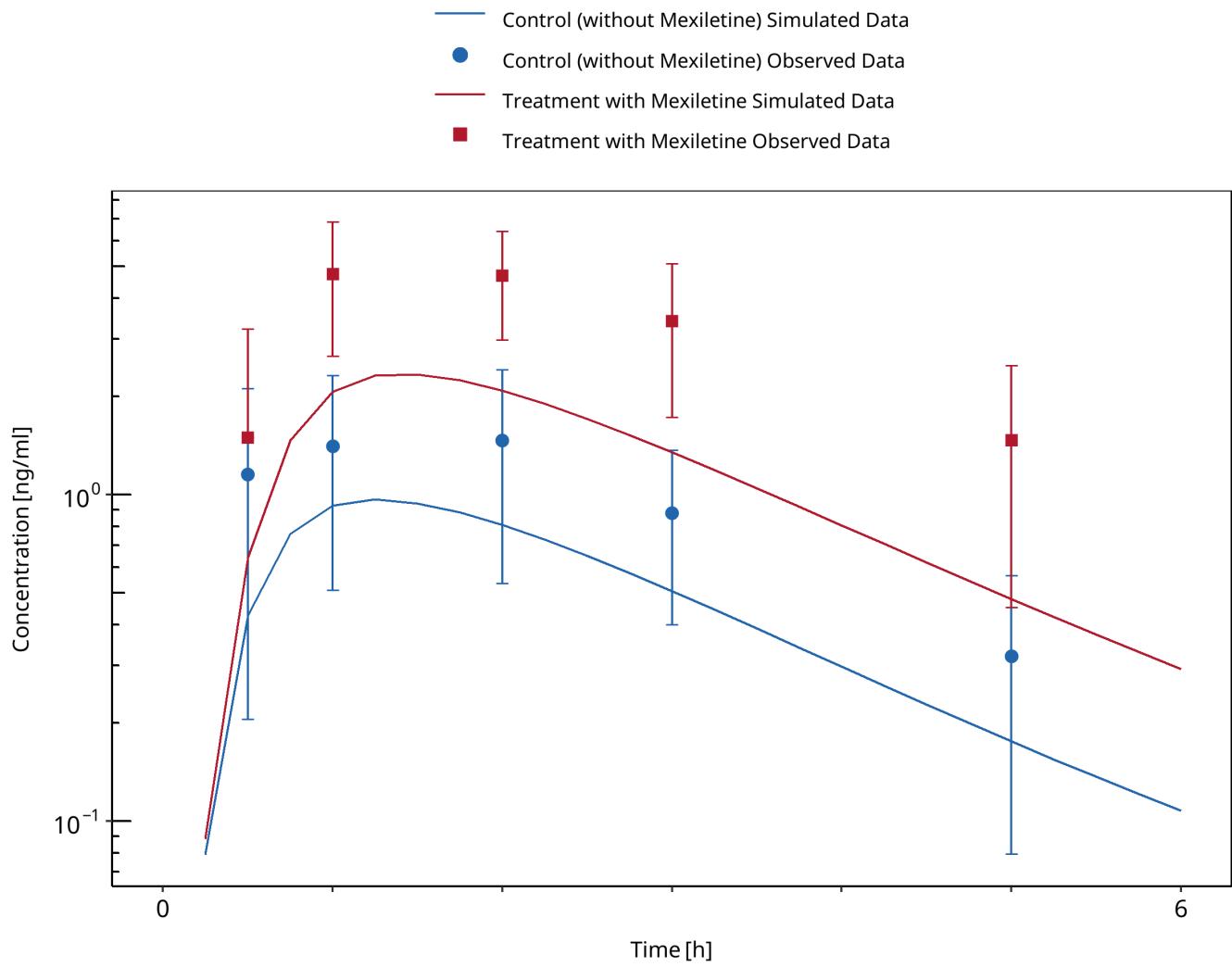


Figure 3-5: Momo 2010 (Tizanidine 2 mg po)

3.5 Caffeine - Mexiletine DDI

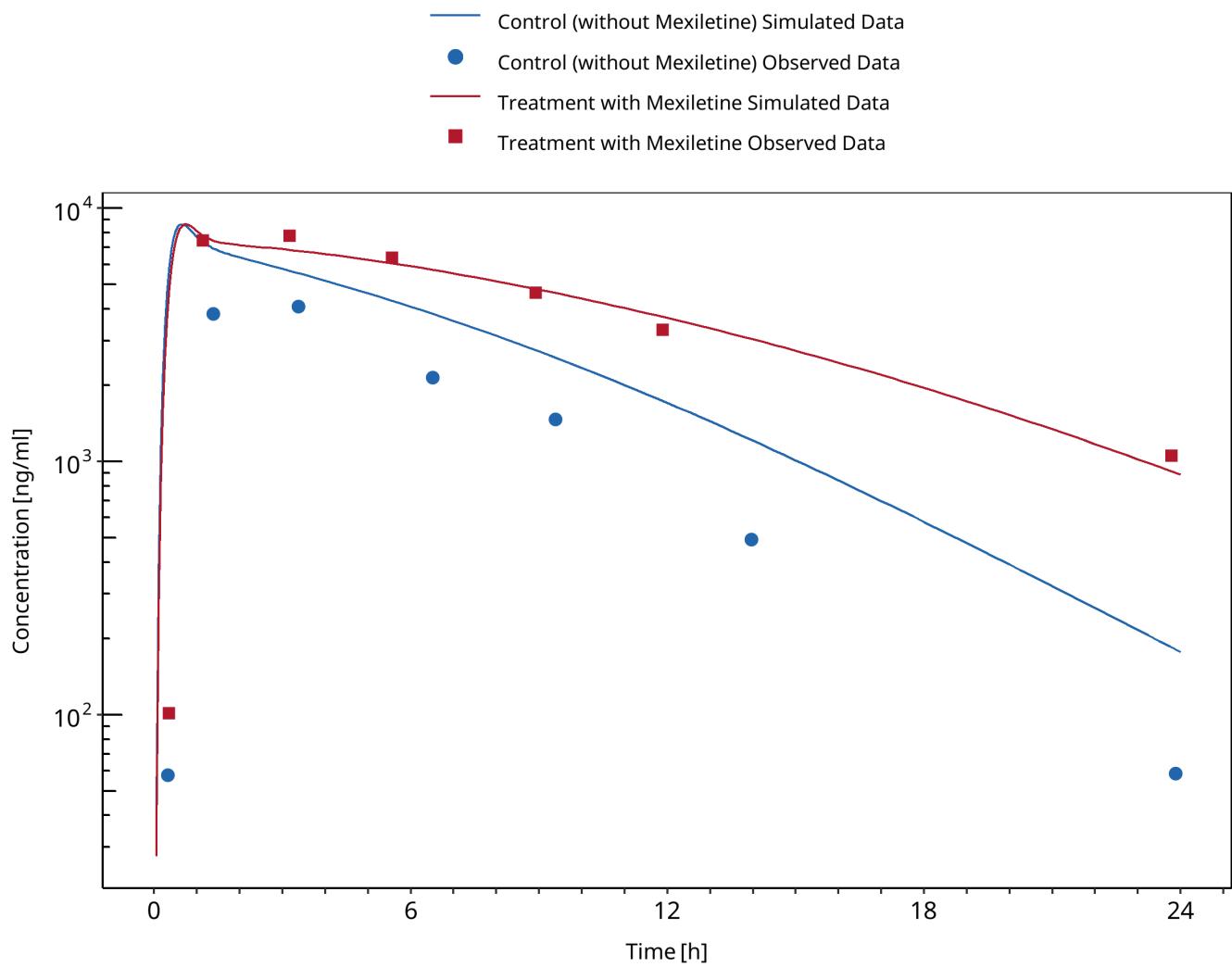


Figure 3-6: Joeres 1987 (Caffeine 336 mg po)

3.6 Mexiletine - Fluvoxamine DDI

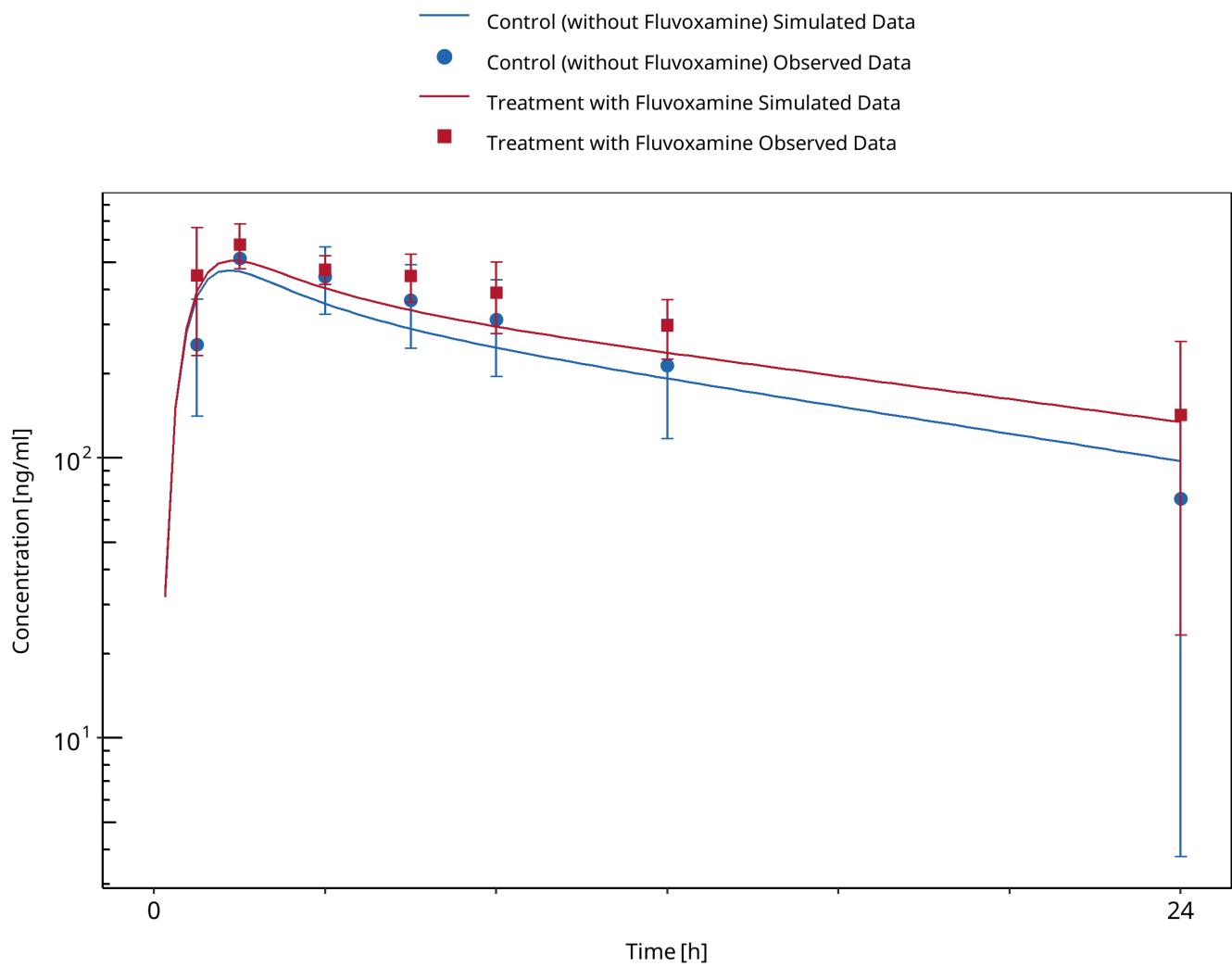


Figure 3-7: Kusumoto 2001 (Mexiletine 166.62 mg po)

4 Conclusion

The predicted perpetrator/victim drug concentration-time profiles, DDI AUC and Cmax ratios confirmed that the developed PBPK models are well suited to characterize the CYP1A2 DDI network over the full range of reported DDI studies. The same Ki values could be used by the moderate CYP1A2 inhibitors EE (0.48 μ M) and mexiletine (0.28 μ M) with regards to all tested substrates (caffeine and tizanidine). For the strong inhibition of CYP1A2 by fluvoxamine, the same Ki value (2.97 nM) could be used for both caffeine and mexiletine as substrate. In contrast, for tizanidine the Ki (0.9 nM) needed to be refitted to capture the data.

Fluvoxamine

- CYP1A2 inhibition:
 - DDI simulations with caffeine, tizanidine, and mexiletine demonstrate an excellent prediction (ratio pred/obs = around 1 and within 2-fold) of the inhibitory potential of fluvoxamine on CYP1A2.

Tizanidine

- Substrate:
 - DDI simulations with fluvoxamine as inhibitor of tizanidine demonstrated a good prediction of tizanidine levels (pred/obs AUCR and pred/obs CmaxR within 2-fold) when using an optimized value of fluvoxamine Ki.
 - DDI simulations with mexiletine as inhibitor of tizanidine demonstrated an underprediction of tizanidine levels but were within 2-fold of observed ratios. Tizanidine is underpredicted both with and without mexiletine, probably due to an un-modelled food effect
 - DDI predictions with ethinylestradiol (EE) using a TDI mechanism was necessary to describe the observed interaction with tizanidine. DDI simulations with ethinylestradiol demonstrated a good prediction of tizanidine levels when using TDI.

Mexiletine

- Perpetrator:
 - DDI simulations with mexiletine as inhibitor of caffeine demonstrated an underprediction of caffeine levels but were within 2-fold of observed ratios.
 - DDI simulations with mexiletine as inhibitor of tizanidine demonstrated an underprediction of tizanidine levels but were within 2-fold of observed ratios.
 - Overall, DDI predictions with mexiletine as an inhibitor tend to lead to underprediction of both AUCR and CmaxR but were within 2-fold of observed ratios.
- Substrate:
 - DDI simulations with fluvoxamine as inhibitor of mexiletine demonstrated an excellent prediction of mexiletine levels.

Ethinylestradiol

- A TDI mechanism on CYP1A2 was introduced to describe tizanidine DDI study data.
- Perpetrator:
 - DDI simulations with EE as inhibitor of caffeine demonstrated a good prediction of caffeine levels (pred/obs AUCR and pred/obs CmaxR within 2-fold).
 - DDI simulations with EE as inhibitor of tizanidine demonstrated a good prediction of tizanidine levels (pred/obs AUCR and pred/obs CmaxR within 2-fold).

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6 Appendix

6.1 Open Systems Pharmacology Suite (OSPS) Introduction

Open Systems Pharmacology Suite (OSP suite) is a tool for PBPK modeling and simulation of drugs in laboratory animals and humans. PK-Sim® and MoBi® are part of the OSP suite [1]. PK-Sim® is based on a generic PBPK-model with 18 organs and tissues. One of the main assumptions is that all compartments are well-stirred. Represented organs/tissues include arterial and venous blood, adipose tissue (separable adipose, excluding yellow marrow), brain, lung, bone (including yellow marrow), gonads, heart, kidneys, large intestine, liver, muscle, portal vein, pancreas, skin, small intestine, spleen and stomach, as shown in [Figure Appendix-1](#).

Each organ consists of four sub-compartments namely the plasma, blood cells (which together build the vascular space), interstitial space, and cellular space. Distribution between the plasma and blood cells as well as between the interstitial and cellular compartments can be permeability-limited. In the brain, the permeation barrier is located between the vascular and the interstitial space. PK-Sim® estimates model parameters (intestinal permeability [2] organ partition coefficients (tissue-to-plasma partition coefficients) [3,4], and permeabilities) from physico-chemical properties of compounds (molecular weight, pKa, acid/base properties) and the composition of each tissue compartment (lipids, water and proteins). Partition coefficients can be calculated using a variety of methods available in PK-Sim®, for example the internal PK-Sim® method [3,4] or that of Rodgers and Rowland [5-7].

Physiological databases included in the software incorporate the dependencies of organ composition, organ weights, organ blood flows and gastrointestinal parameters (gastrointestinal length, radius of each section, intestinal surface area, gastrointestinal transit times, and pH in different intestinal segments [2]), with the user-defined body weight and height and ethnicity of the individual [8]. Thereby, PK Sim® allows generating realistic virtual populations. For a detailed description of the PBPK model structure implemented in PK Sim®, see Willmann et al. [2,4,8,9] or the OSP Suite homepage (<https://docs.open-systems-pharmacology.org/mechanistic-modeling-of-pharmacokinetics-and-dynamics/modeling-concepts>).

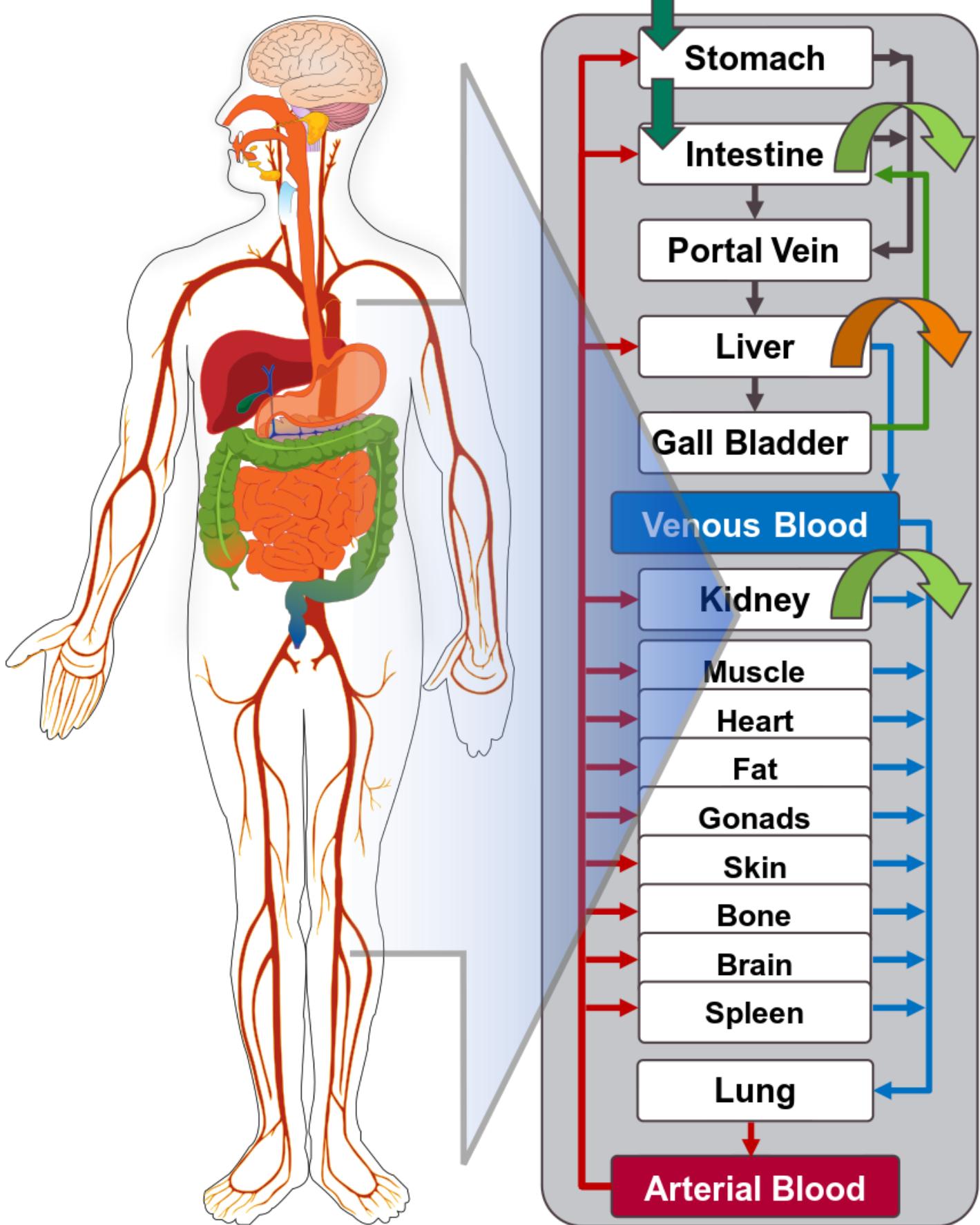


Figure Appendix-1: Structure of the Whole Body PBPK Model integrated in PK-Sim®

References for OSPS introduction

- [1] www.open-systems-pharmacology.org
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6.2 Mathematical Implementation of Drug-Drug Interactions

DDI modeling: Competitive inhibition

A detailed representation of the mathematical implementation of competitive enzyme inhibition can be found in the OSP manual (<https://docs.open-systems-pharmacology.org/working-with-pk-sim/pk-sim-documentation/pk-sim-compounds-defining-inhibition-induction-processes#competitive-inhibition-simple-setting-with-one-inhibitor>).

DDI modeling: Mechanism-based inhibition

A detailed representation of the mathematical implementation of mechanism-based enzyme inhibition can be found in the OSP manual (<https://docs.open-systems-pharmacology.org/working-with-pk-sim/pk-sim-documentation/pk-sim-compounds-defining-inhibition-induction-processes#irreversible-inhibition>).

DDI modeling: Induction

A detailed representation of the mathematical implementation of enzyme induction can be found in the OSP manual (<https://docs.open-systems-pharmacology.org/working-with-pk-sim/pk-sim-documentation/pk-sim-compounds-defining-inhibition-induction-processes#enzyme-induction>).

6.3 Automatic (re)-qualification workflow

Open Systems Pharmacology ([https://www.open-systems-pharmacology.org/](http://www.open-systems-pharmacology.org/)) provides a dynamic landscape of model repositories and a database of observed clinical data. Additionally, a technical framework to assess confidence of a specific intended use has been developed (qualification runner and reporting engine). This framework allows for an automatic (re)-qualification workflow of the OSP suite, comprising the following steps [Figure Appendix-2](#):

- PBPK model development and verification with observed data,

- Qualification plan generation,
- Qualification plan execution,
- Qualification report generation.

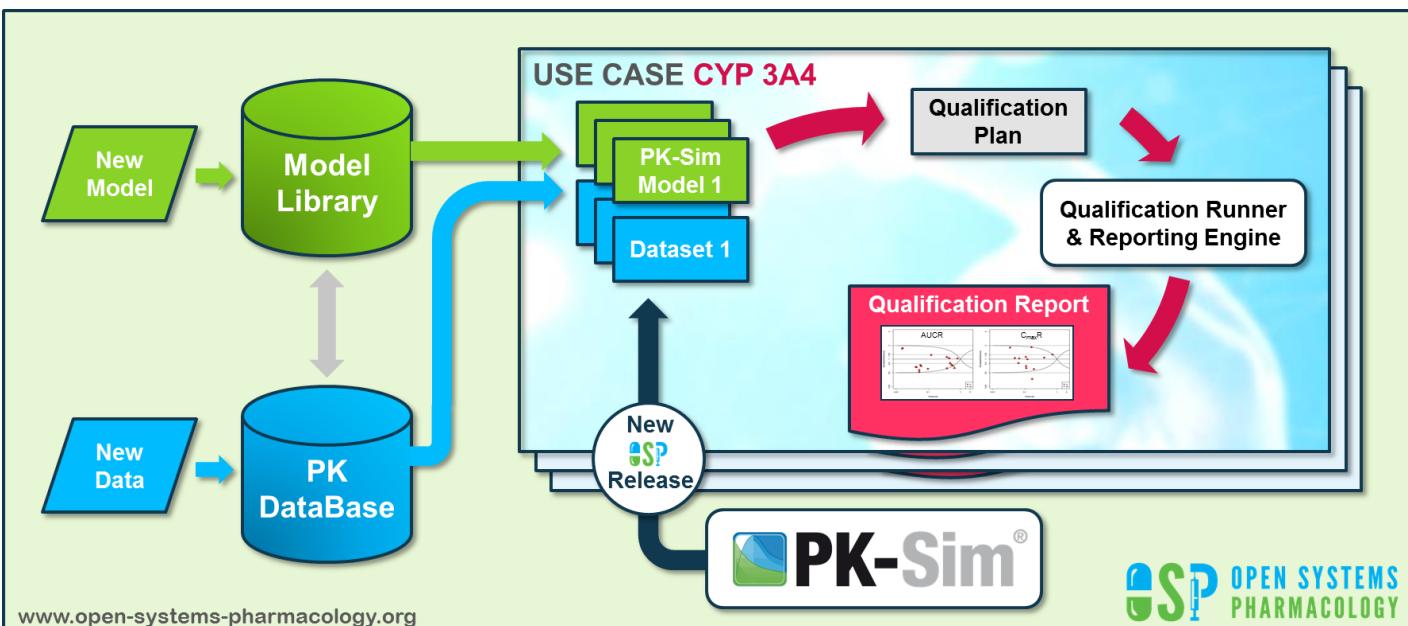
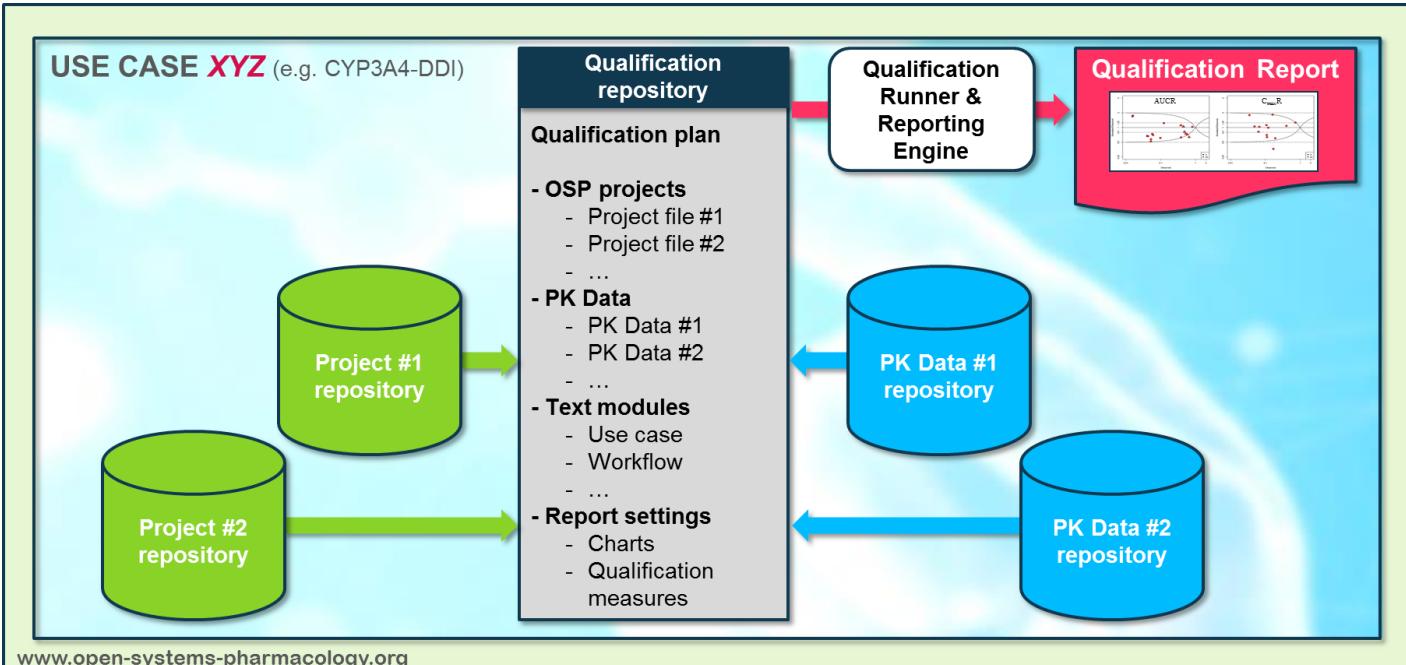


Figure Appendix-2: OSP suite automatic (re)-qualification workflow

In a first step, the respective qualification scenario is saved in a special qualification repository on OSP GitHub (<https://github.com/Open-Systems-Pharmacology/>). This qualification scenario repository contains a detailed qualification plan that links and combines respective models and data to address the use case that shall be qualified. Therefore, the qualification plan consists of:

- PK-Sim project files,
- Additional model building steps (if applicable),
- Description of potential cross-dependencies between PK-Sim project files (if applicable),
- Observed data (needed for model development and verification),
- Qualification scenario description text modules
- Detailed report settings to describe the generation of charts and qualification measures.

PK-Sim projects, observed data sets, and qualification scenario text modules are deposited in distinct repositories and are referenced by the qualification plan ([Figure Appendix-3](#)).



www.open-systems-pharmacology.org

Figure Appendix-3: Qualification scenario repository landscape on GitHub

In a second step the qualification runner (<https://github.com/Open-Systems-Pharmacology/QualificationRunner>) processes the qualification plan, i.e. all project parts are exported and prepared for the reporting engine (<https://github.com/Open-Systems-Pharmacology/Reporting-Engine>). The reporting engine provides a validated environment (implemented in R) for model execution and finally generates the qualification report. This report contains the evaluation of the individual PBPK models with observed data (i.e. standard goodness of fit plots, visual predictive checks) and a comprehensive qualification of the specific use case assessing the predictive performance of the OSP suite by means of a predefined set of qualification measures and charts.

The automated execution of the described workflow can be triggered to assess re-qualification in case new data, changes in model structure or parameterization, or new OSP suite releases arise.

7 Glossary

ADME		Absorption, Distribution, Metabolism, Excretion
AUC		Area under the plasma concentration versus time curve
AUCinf		AUC until infinity
AUClast		AUC until last measurable sample
AUCR		Area under the plasma concentration versus time curve Ratio
b.i.d.		Twice daily (bis in diem)
CL		Clearance
Clint		Intrinsic liver clearance
Cmax		Maximum concentration
CmaxR		Maximum concentration Ratio
CYP		Cytochrome P450 oxidase
CYP1A2		Cytochrome P450 1A2 oxidase
CYP2C19		Cytochrome P450 2C19 oxidase
CYP3A4		Cytochrome P450 3A4 oxidase
DDI		Drug-drug interaction
e.c.		Enteric coated
EE		Ethinylestradiol
EM		Extensive metabolizers
fm		Fraction metabolized
FMO		Flavin-containing monooxygenase
fu		Fraction unbound
FDA		Food and Drug administration
GFR		Glomerular filtration rate
HLM		Human liver microsomes
hm		homozygous
ht		heterozygous
IM		Intermediate metabolizers

ADME	Absorption, Distribution, Metabolism, Excretion
i.v.	Intravenous
IVIVE	In Vitro to In Vivo Extrapolation
Ka	Absorption rate constant
kcat	Catalyst rate constant
Ki	Inhibitor constant
Kinact	Rate of enzyme inactivation
Km	Michaelis Menten constant
m.d.	Multiple dose
OSP	Open Systems Pharmacology
PBPK	Physiologically-based pharmacokinetics
PK	Pharmacokinetics
PI	Parameter identification
PM	Poor metabolizers
RT-PCR	Reverse transcription polymerase chain reaction
p.o.	Per os
q.d.	Once daily (queaque diem)
SD	Single Dose
SE	Standard error
s.d.SPC	Single dose Summary of Product Characteristics
SD	Standard deviation
TDI	Time dependent inhibition
t.i.d	Three times a day (ter in die)
UGT	Uridine 5'-diphospho-glucuronosyltransferase
UM	Ultra-rapid metabolizers