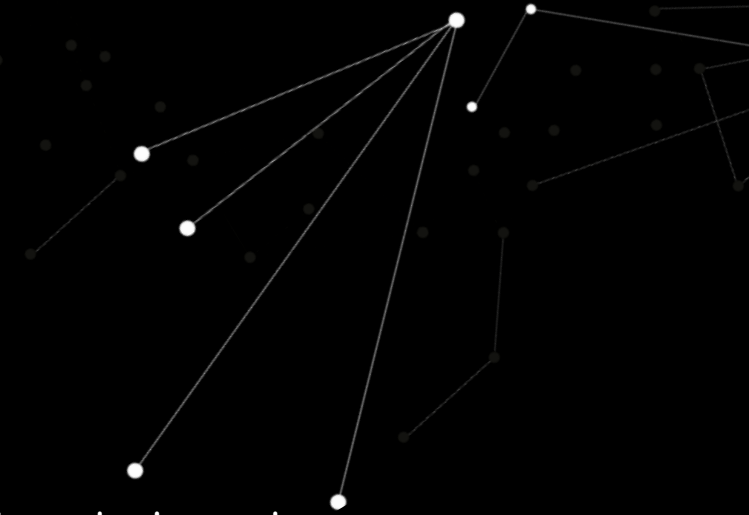


2020 T2 COMP9517

Computer Vision



Project Goal

Develop an automated cell tracking framework that can consistently and accurately:

- **Detect Cells**: Unet for Segmentation.
- **Track Cells**: Euclidean distance of centroids and 3D trajectory.
- **Detect Cell Division**: Changing colors of the cell's bounding box.
- **Analyze Cell Motion**: Euclidean, Total and Net distance travelled.

Motivation

- Demonstrate the importance of computer vision methods in tracking biological cells in time-lapsed microscopy images.
- Help to reduce human errors, effort and expertise required in manual annotation process.
- Assist biologists or medical researchers in their area of research.

Challenges

- Low contrast between cells and image background.
- Large image sequences with dense cell population.
- Irregular and deformable cell shapes, overlapping cells.

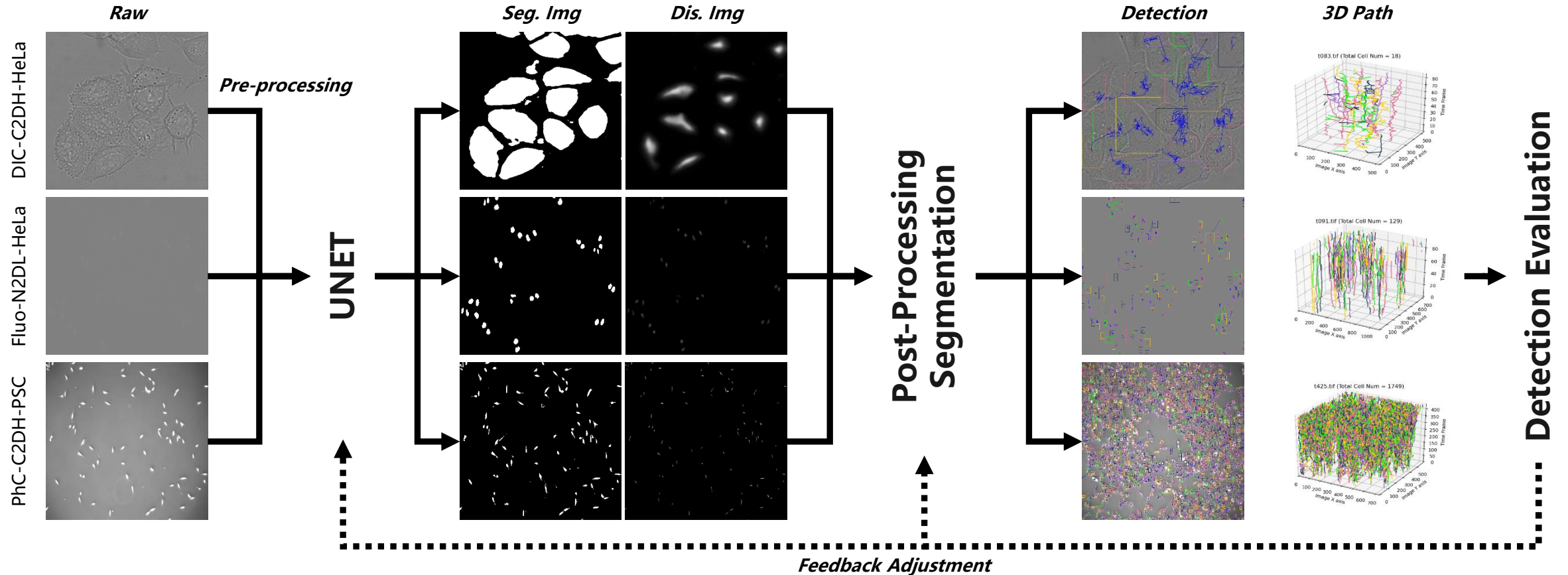
Team Name: Group 1

- Rafael Formoso
- Sebastian Castillo Castro
- Xinli Wang (Kevin Wang)
- Yuzhong Duan (Jack Duan)
- Kan-Lin Lu (Gary Lu)

Detection methodology

Pre-processing, UNET, Post-Processing

Detection Methodology Overview



UNET Set up:

Pre-processing	➡	Dataset specific mode + normalization
Data augmentation	➡	Random flip, rotation, skew, crop
Targets	➡	Semantic segmentation + Distance est.

Post-Processing Methods:

DIC-C2DH-HeLa and Fluo-N2DL-HeLa	➡	Watershed + Morphological Operation
PhC-C2DH-PSC	➡	Morphological Operation

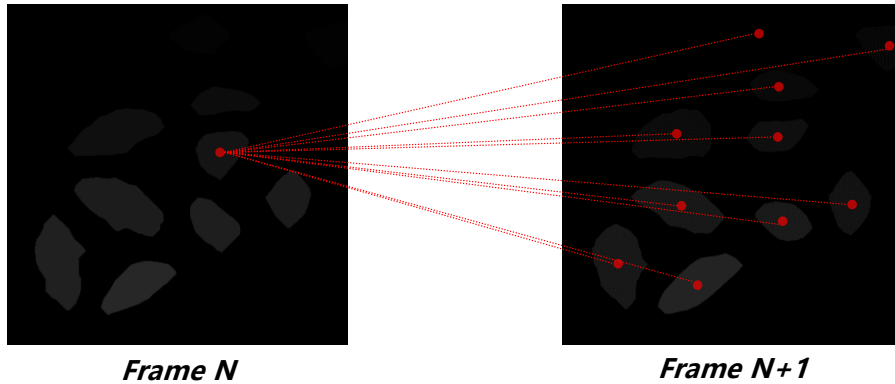
Tracking and Division methodology

Distance Matrix, Division distance definition, Backward patching.

Tracking Methodology Overview

Euclidean Distance: $d = \sqrt{(x_1 - x_2)^2 + (y_1 - y_2)^2}$

illustration

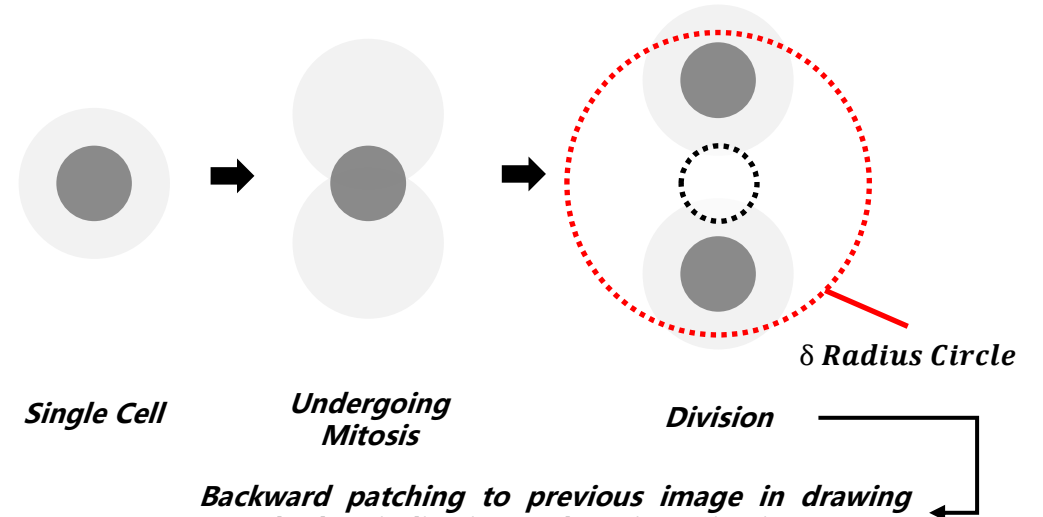


Distance Matrix Example

		Frame N+1 Cell IDs			
		1	2	3	4
Frame N Cell IDs	1	32	12	2	15
	2	2	17	3	41
	3	35	4	1	11
	4	4	12	84	1
	5	21	34	9	5
	6	31	1	12	3

Division Methodology Overview

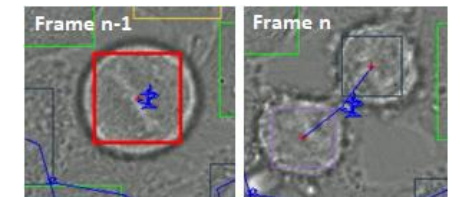
δ division distance



Backward patching to previous image in drawing
"Red" box indicating undergoing mitosis

*Selected Distance:
(Trial and Error)*

	δ
DIC-C2DH-HeLa	80
Fluo-N2DL-HeLa	10
PhC-C2DH-PSC	3



Motion Analysis

Centroids per frame
stored in dataframe

Per Frame Motion
Calculation

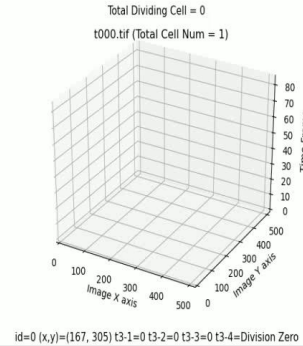
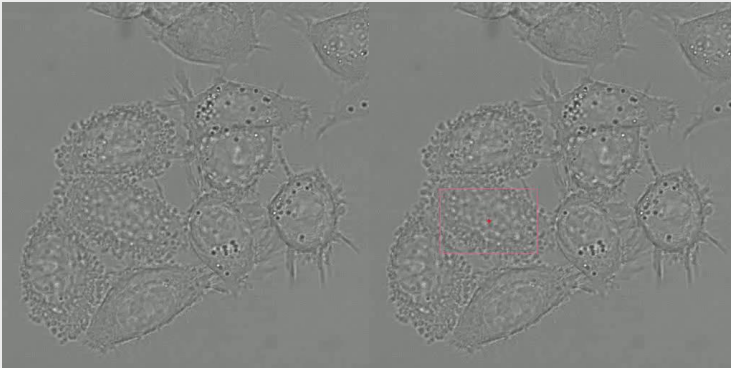
Per Frame
Annotation Html

Any single cell can be selected by user prompt in plotting 3D pathology graph,
along with motion analysis printed in output image.

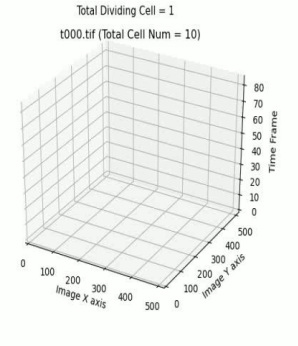
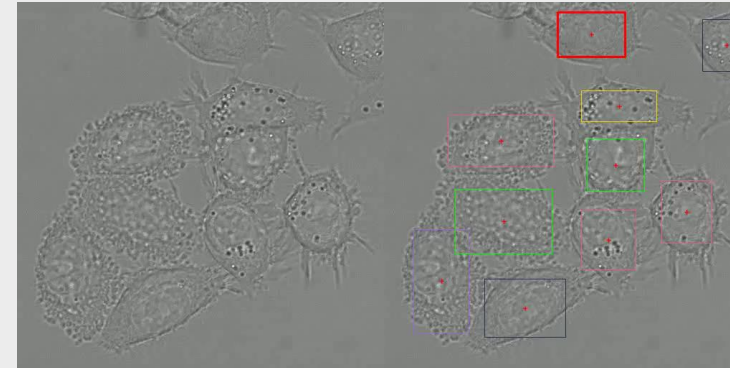
Demonstration and Limitations

Cell Tracking, Mitosis Tracking.

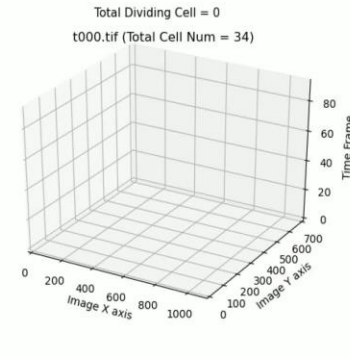
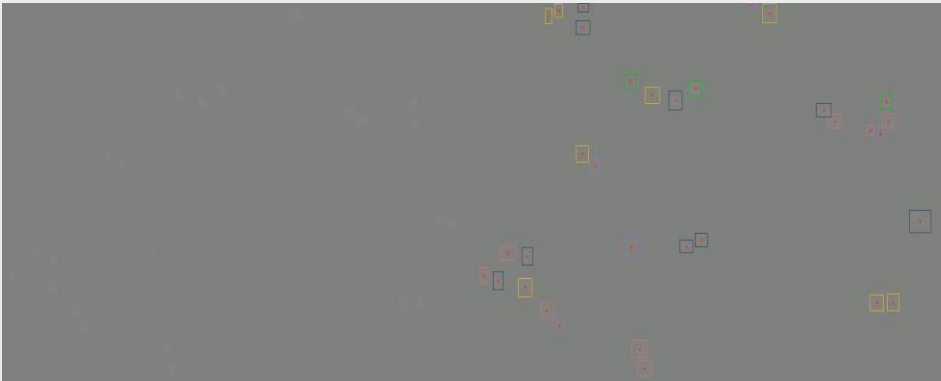
DIC-C2DH-HeLa
(Selected Cell)



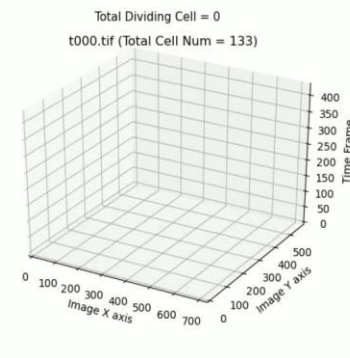
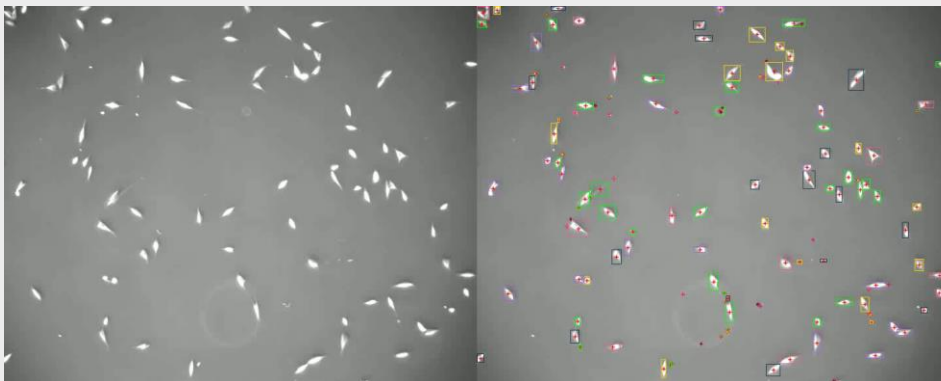
DIC-C2DH-HeLa



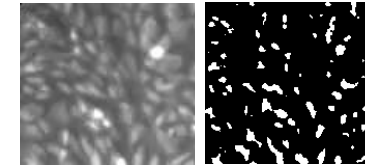
Fluo-N2DL-HeLa



PhC-C2DH-PSC



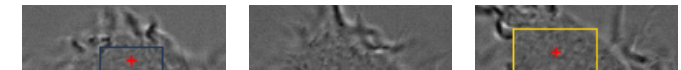
Limitations observed:



Over-Segmentation



Overlapped Cells results in mislabelling of mitosis event



Appeared then reappeared cells caused by over-segmentation results in extra cell ID

Mismatching caused by cell travelling over another cell across frames

Dataset 1, extreme large cell segmented to 2 pieces due to irregular shape

Result, Discussion and Conclusion

Discuss outcome, present conclusion and possible future work.

Results

Per Frame Outcome Analysis:

DIC-C2DH-HeLa / Sequence 1 (Frame 0 - 14)					
Cell Detection Analysis					
TP	157	FP	4	FN	1
Precision (%)	97.52 %		Recall (%)	97 %	
Cell Division Analysis					
TP	1	FP	3	FN	0
Precision (%)	25.00 %		Recall (%)	80%	
DIC-C2DH-HeLa / Sequence 3 (Frame 0 - 14)					
Cell Detection Analysis					
TP	179	FP	8	FN	0
Precision (%)	95.72 %		Recall (%)	100%	
Cell Division Analysis					
TP	2	FP	5	FN	0
Precision (%)	28.57 %		Recall (%)	100 %	

DIC-C2DH-HeLa: Division took place 48% average of total Cell Number

Fluo-N2DL-HeLa: Division took place 3.1% average of total Cell Number

PhC-C2DH-PSC: Division took place 2.6 % average of total Cell Number

Per Sequence Outcome Behaviour Analysis:

	DIC-C2DH-HeLa				Fluo-N2DL-HeLa				PhC-C2DL-PSC			
Sequence Number	1	2	3	4	1	2	3	4	1	2	3	4
Total Cell Number in Entire sequence	71	96	165	64	770	2768	1020	1207	51452	43864	27369	26923
Total Cell Number in Last Frame	18	17	9	12	129	334	213	229	1749	1423	865	795
Total Division Occurrence	38	55	67	27	16	112	21	51	1740	1306	519	604
Average Cell Speed (Pixel/Frame)	12.37	14.07	12.96	9.21	2.4	2.25	2.22	1.9	0.705	0.68	0.66	0.65
Speed 90% CI	(11.77,12.97)	(13.33,14.82)	(12.25,13.67)	(8.72,9.71)	(2.36,2.44)	(2.24,2.28)	(2.19,2.24)	(1.88,1.92)	(0.7,0.71)	(0.68,0.69)	(0.66,0.66)	(0.65,0.66)
Average Cell Cumulative Distance (Pixel)	213.97	154.23	120.01	190.97	23.07	19.43	24.92	24.22	3.73	3.44	2.57	2.41
Cumulative Distance 90% CI	(151.15,276.8)	(111.46,197.0)	(90.1,149.92)	(129.8,252.1)	(21.04,25.1)	(18.44,20.43)	(22.96,26.88)	(22.49,25.96)	(3.67,3.78)	(3.38,3.5)	(2.51,2.63)	(2.35,2.47)
Average Cell Net Distance (Pixel):	49.67	46.1	38.09	45.84	6.99	6.56	7.34	6.88	1.44	1.4	1.11	1.02
Net Distance 90% CI	(42.34,56.99)	(40.54,51.66)	(33.04,43.14)	(36.91,54.78)	(6.46,7.52)	(6.26,6.87)	(6.86,7.82)	(6.47,7.3)	(1.42,1.46)	(1.38,1.42)	(1.09,1.14)	(1.01,1.05)
Average Confinement Ratio	5.15	3.46	5.06	5.61	3.33	3.36	3.47	3.85	2.53	2.39	2.24	2.28
Confinement Ratio 90% CI	(3.5,6.79)	(2.62,4.3)	(3.49,6.62)	(3.32,7.89)	(3.11,3.55)	(3.18,3.54)	(3.21,3.74)	(3.54,4.15)	(2.51,2.57)	(2.36,2.43)	(2.2,2.38)	(2.23,2.33)

Discussion

- High computational time for PhC-C2DH-PSC, due to matching process
- Decrease in reliability when cells are clustered with extreme high population
- Motion analysis highly depends on outcome of segmentation and tracking

Conclusion and Future Work

- An insight to cell detection and a reasonable outcome is achieved
- Fine tuning UNET and Post-Process method in tackling stated limitation
- Include extra features in matching across frames
- Include extra features in division matching across frames