DAMPSA: Domain-aware Multiple Protein Sequence Aligner

Roll number: 820

DAMPSA is a multiple sequence analysis (MSA) tool that utilises protein domain annotations as biological constraints.

DAMPSA generates MSA outputs where protein domains are anchored. This means:

- 1. Domain and linker segments of the protein will not be mixed by random similarity.
- 2. Potentially, distant sequences with similar structures can be aligned.

Installation

```
conda create -n dampsa python=3.9 conda activate dampsa
```

In project folder (cd DAMPSA),

1. Install python libraries through pip.

```
pip install -r py_requirements.txt
```

2. Install command line programs through bioconda.

```
conda install --file=cmd_requirements.txt -c bioconda
```

- 3. Prepare the **local Pfam database** for domain annotation.
 - The following code download Pfam database files and then use hmmpress to index them for efficient processing.
 - Note: The ready database will takes ~3.4 GB space.
 - Note: The links are updated on 30/05/2022. Check Pfam if any link fails.

```
mkdir data/Pfam_scan_db
cd data/Pfam_scan_db
wget http://ftp.ebi.ac.uk/pub/databases/Pfam/current_release/Pfam-A.hn
wget http://ftp.ebi.ac.uk/pub/databases/Pfam/current_release/Pfam-A.hn
wget http://ftp.ebi.ac.uk/pub/databases/Pfam/current_release/active_si
gunzip *.gz
hmmpress Pfam-A.hmm
```

:) You are now ready to run DAMPSA main pipeline.

Further notes:

- To run visualisation scripts in R (not the main pipeline), you need the following packages.
 - ▼ click here

```
tidyverse
msa
ggmsa
RColorBrewer
Biostrings
stringr
getopt
```

• DAMPSA is developed and tested on MacOS 12.1, Python 3.9.12, and R 4.1.2.

Getting started (or see a walkthrough tutorial here)

```
In project folder (cd DAMPSA),
  python bin/main.py -h
```

▼ parameter descriptions

--log

```
[--n-thread N_THREAD]
DAMPSA input arguments.
optional arguments:
-h, --help
                      show this help message and exit
--input INPUT
                      Path to the input .fasta file.
--output OUTPUT
                      Path to the alignment .fasta file output.
--domain-out DOMAIN_OUT
Path to output domain annotation results.
--refine-edge
                      Refine alignments at the edge between domain and
                      Not to check if the linker is too long - likely
--no-check-linker
--focus-clan FOCUS_CLAN
Only consider the specified Clan IDs (domain superfamily) - comma sepo
--cache-dom CACHE_DOM
Skip hmmscan, use supplied filepath to cached domain table (TSV-like).
--domain-app DOMAIN_APP
Aligner for domain segments (clustalo or mafft), default clustalo.
--linker-app LINKER_APP
Aligner for linker segments (clustalo or mafft), default clustalo.
                      Store log file in the same folder as the alignme
```

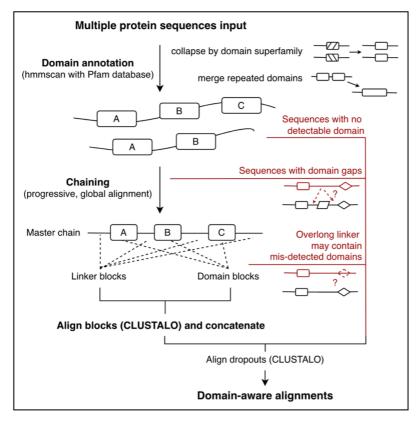
usage: main.py [-h] [--input INPUT] [--output OUTPUT] [--domain-out [

[--focus-clan FOCUS_CLAN] [--cache-dom CACHE_DOM] [--domain-apr

Run an example command (align the RASSF family)

```
python bin/main.py --input tutorial/RASSF/raw.fasta \
   --output tutorial/RASSF/aligned.fasta \
   --domain-out tutorial/RASSF/domain.txt \
   --no-check-linker --log
```

Architecture



The DAMPSA pipeline involves three stages:

- 1. Annotating domains with hmmscan and Pfam database.
- 2. Chaining domain sequence using progressive global alignment implemented here.
- 3. Align blocks defined by the chain, using Clustal-Omega (CLUSTALO). These blocks are concantenated to generate full alignment.

In three cases, sequences cannot be considered by DAMPSA. They are dropped out and aligned finally using *sequence-to-profile* method in CLUSTALO.

- Sequences with no domain detected.
- Domain sequences that are gapped (e.g. -A-C- vs. -A-B-C-, the first sequence will be dropped out)
- Sequences with overlong linker which may indicate domain misdetection.

All dropout cases are logged. Please check log.txt in DAMPSA output.

API documentation

See here.