

# Assignment 3: Data Exploration

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## OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

## Directions

1. Rename this file `<FirstLast>_A03_DataExploration.Rmd` (replacing `<FirstLast>` with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Assign a useful **name to each code chunk** and include ample **comments** with your code.
5. Be sure to **answer the questions** in this assignment document.
6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Canvas.

**TIP:** If your code extends past the page when knit, tidy your code by manually inserting line breaks.

**TIP:** If your code fails to knit, check that no `install.packages()` or `View()` commands exist in your code.

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## Set up your R session

1. Load necessary packages (tidyverse, lubridate, here), check your current working directory and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX\_Neonicotinoids\_Insects\_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON\_NIWO\_Litter\_massdata\_2018-08\_raw.csv). Name these datasets “Neonics” and “Litter”, respectively. Be sure to include the sub-command to read strings in as factors.

```
#Import packages
library(tidyverse); library(lubridate); library(here); library(ggplot2)

#Check work directory
here()
```

```
## [1] "/home/guest/EDE_Fall2024"
```

```

setwd(here())
#Upload the two datasets
Neonics <- read.csv(
  file = here('Data', 'Raw', 'ECOTOX_Neonicotinoids_Insects_raw.csv'),
  stringsAsFactors = T
)

Litter <- read.csv(
  file = here('Data', 'Raw', 'NEON_NIWO_Litter_massdata_2018-08_raw.csv'),
  stringsAsFactors = T
)

```

## Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: Because neonicotinoids can be risky. They can be not only toxic to insects but also human beings and food. Since neonicotinoids are widely used in agriculture, it is important to know if these neonicotinoids are effective. According to the report from NRDC: <https://www.nrdc.org/stories/neonicotinoids-101-effects-humans-and-bees>, neonicotinoids can hurt the ecosystem, so it is important to study on it.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: According to the report from USDA: [https://www.srs.fs.usda.gov/pubs/gtr/gtr\\_srs038/gtr\\_srs038-scheungrab001.pdf](https://www.srs.fs.usda.gov/pubs/gtr/gtr_srs038/gtr_srs038-scheungrab001.pdf) litter and woody debris is important because it can store carbon, recycles nutrients and plays an essential role for aquatic ecosystems. Litter and woody debris is biomass, which can be a type of renewable energy.

4. How is litter and woody debris sampled as part of the NEON network? Read the NEON\_Litterfall\_UserGuide.pdf document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. Randomly selected locations of tower plots 2. The percentage of vegetation covered is also important because it determines whether the plots should be targeted or randomized 3. Frequency for elevated traps is salient for Temporal Sampling Design.

## Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
dim(Neonics)
```

```
## [1] 4623 30
```

```
#There are 4623 rows and 30 columns
```

6. Using the `summary` function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest? [Tip: The `sort()` command is useful for listing the values in order of magnitude...]

```
sort(summary(Neonics$Effect))
```

```
##      Hormone(s)      Histology      Physiology      Cell(s)
##           1           5           7           9
##      Biochemistry      Accumulation      Intoxication      Immunological
##           11          12          12          16
##      Morphology      Growth      Enzyme(s)      Genetics
##           22          38          62          82
##      Avoidance      Development      Reproduction      Feeding behavior
##          102         136          197          255
##      Behavior      Mortality      Population
##          360        1493          1803
```

Answer: The most common effects that are studied are “Population” and “Mortality”, which are bigger than 1,000. These two effects show that researchers are more interested in how the neonicotinoids affect insects’ population and mortality. They are interested in that because these two can directly show if the neonicotinoids decrease insects’ population and increase their mortality. If neonicotinoids do, it means that they impact the growth of insects.

7. Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed. [TIP: Explore the help on the `summary()` function, in particular the `maxsum` argument...]

```
summary(Neonics$Species.Common.Name) #Summarize the data
```

```
##      Honey Bee      Parasitic Wasp
##           667          285
##      Buff Tailed Bumblebee      Carniolan Honey Bee
##           183          152
##      Bumble Bee      Italian Honeybee
##           140          113
##      Japanese Beetle      Asian Lady Beetle
##           94           76
##      Euonymus Scale      Wireworm
##           75           69
##      European Dark Bee      Minute Pirate Bug
##           66           62
##      Asian Citrus Psyllid      Parastic Wasp
##           60           58
```

##	Colorado Potato Beetle	Parasitoid Wasp
##	57	51
##	Erythrina Gall Wasp	Beetle Order
##	49	47
##	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
##	47	46
##	True Bug Order	Buff-tailed Bumblebee
##	45	39
##	Aphid Family	Cabbage Looper
##	38	38
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ground Beetle Family
##	29	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23
##	Ladybird Beetle	Mason Bee
##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order
##	17	17
##	Egg Parasitoid	Insect Class
##	17	17
##	Moth And Butterfly Order	Oystershell Scale Parasitoid
##	17	17

## Hemlock Woolly Adelgid Lady Beetle	Hemlock Woolly Adelgid
## 16	16
## Mite	Onion Thrip
## 16	16
## Western Flower Thrips	Corn Earworm
## 15	14
## Green Peach Aphid	House Fly
## 14	14
## Ox Beetle	Red Scale Parasite
## 14	14
## Spined Soldier Bug	Armoured Scale Family
## 14	13
## Diamondback Moth	Eulophid Wasp
## 13	13
## Monarch Butterfly	Predatory Bug
## 13	13
## Yellow Fever Mosquito	Braconid Parasitoid
## 13	12
## Common Thrip	Eastern Subterranean Termite
## 12	12
## Jassid	Mite Order
## 12	12
## Pea Aphid	Pond Wolf Spider
## 12	12
## Spotless Ladybird Beetle	Glasshouse Potato Wasp
## 11	10
## Lacewing	Southern House Mosquito
## 10	10
## Two Spotted Lady Beetle	Ant Family
## 10	9
## Apple Maggot	(Other)
## 9	670

```
help(summary) #Check what maxsum means
summary(Neonics$Species.Common.Name, maxsum = 7 )
```

## Honey Bee	Parasitic Wasp	Buff Tailed Bumblebee
## 667	285	183
## Carniolan Honey Bee	Bumble Bee	Italian Honeybee
## 152	140	113
## (Other)		
## 3083		

*#The question is asking for the six most commonly studied species*  
*#We make the levels of 7*  
*#because there would be one level shows "Other" besides the six most commonly studied species.*

Answer: All of the six most commonly studied species are belong to the Apocrita suborder (wasps and bees). Studies show that neonicotinoids not only kill pests but also many other kinds of insects, especially bees. Therefore, researchers might want to know how the neonicotinoids can hurt bees and wasps, which are not targeted pest but are important to the ecosystem and food chain.

8. Concentrations are always a numeric value. What is the class of `Conc.1..Author.` column in the dataset, and why is it not numeric? [Tip: Viewing the dataframe may be helpful...]

```
class(Neonics$Conc.1..Author.)
```

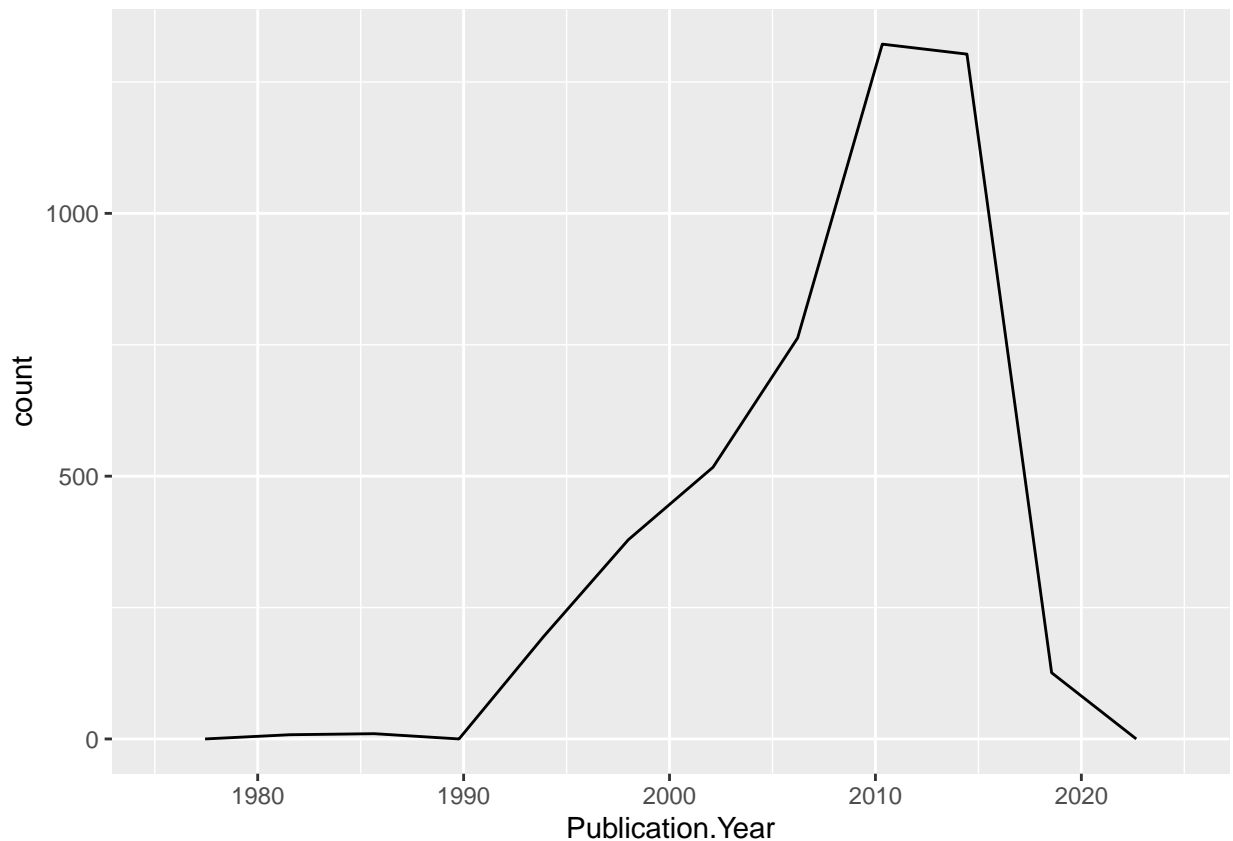
```
## [1] "factor"
```

Answer: The “Conc.1..Author.” is classified as factor. It is not numeric because it includes ranges such as smaller/bigger than specific numbers instead of just numbers, and it includes some special figures such as “<” “>” “/” which are not numeric.

## Explore your data graphically (Neonics)

9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

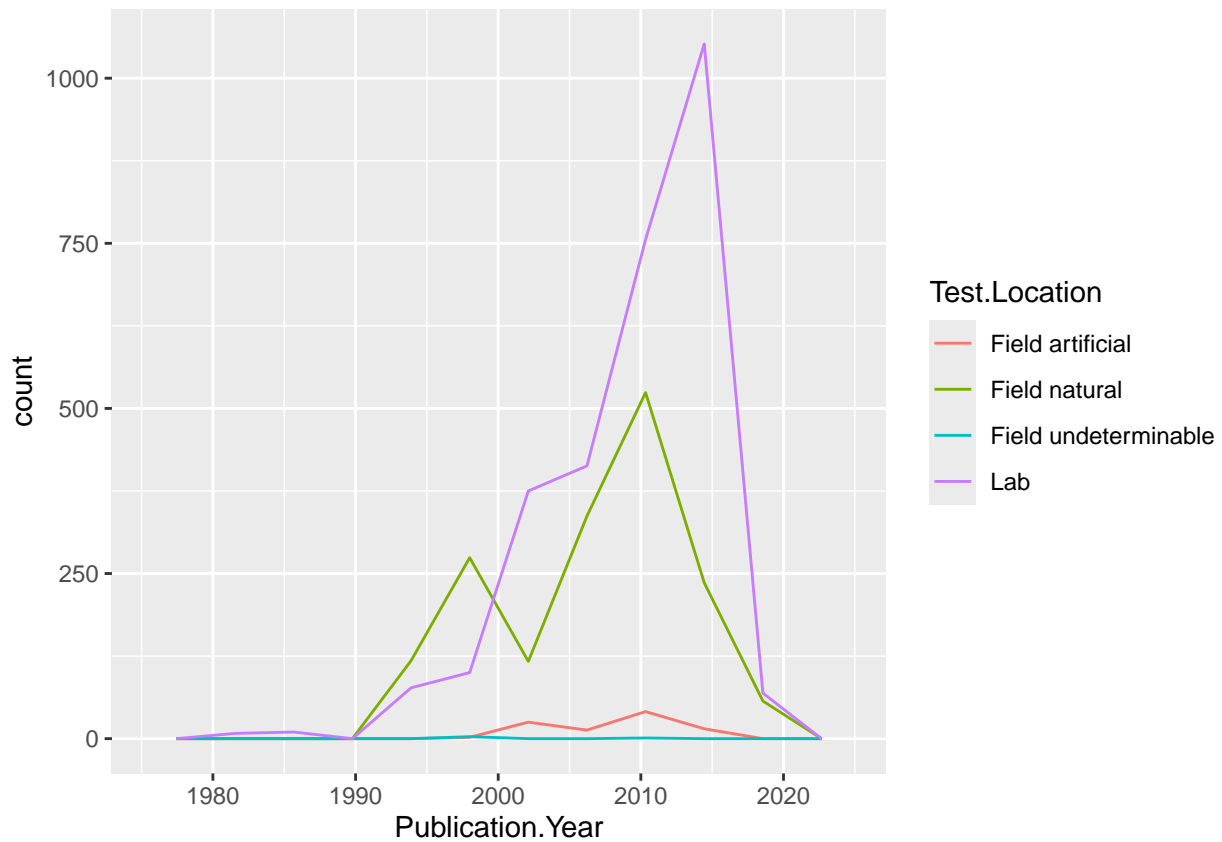
```
ggplot(Neonics)+  
  geom_freqpoly(aes(x=Publication.Year), bins = 10)
```



*#The bin is 10, showing the number of publications in around each three to five years.*

10. Reproduce the same graph but now add a color aesthetic so that different `Test.Location` are displayed as different colors.

```
ggplot(Neonics)+
  geom_freqpoly(aes(x=Publication.Year, color=Test.Location), bins = 10)
```



Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are lab and field natural. The number of tests at field natural increased a lot between 2000s to 2010s, the number of tests at lab increased dramatically from 2005s to 2015s. From around 2010, the tests done at field natural decreased alot, and from around 2015, the tests done at lab dramatically decreased.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX\_CodeAppendix for more information.

[**TIP:** Add `theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))` to the end of your plot command to rotate and align the X-axis labels...]

```
ggplot(Neonics, aes(x=Endpoint)) + geom_bar()+
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))
```



Answer: The two most common end points are LOEL and NOEL. The LOEL represents the Lowest-observable-effect-level, which is the lowest dose (concentration) producing effects that were significantly different from responses of controls. The NOEL represents No-observable-effect-level, which is highest dose (concentration) producing effects not significantly different from responses of controls.

## Explore your data (Litter)

- Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate)
```

```
## [1] "factor"
```

```
#It is a factor, not a data
```

```
Litter$collectDate <- as.Date(Litter$collectDate, format='%Y-%m-%d')
class(Litter$collectDate)
```

```
## [1] "Date"
```

```
#Now it is a date
```

```
unique(Litter$collectDate)
```



```
## [1] "2018-08-02" "2018-08-30"
```

13. Using the `unique` function, determine how many different plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

```
unique(Litter$plotID)
```

```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

```
summary(Litter$plotID)
```

```
## NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 NIWO_058 NIWO_061
##      20      19      18      15      14      8      16      17
## NIWO_062 NIWO_063 NIWO_064 NIWO_067
##      14      14      16      17
```

```
length(unique(Litter$plotID))
```

```
## [1] 12
```

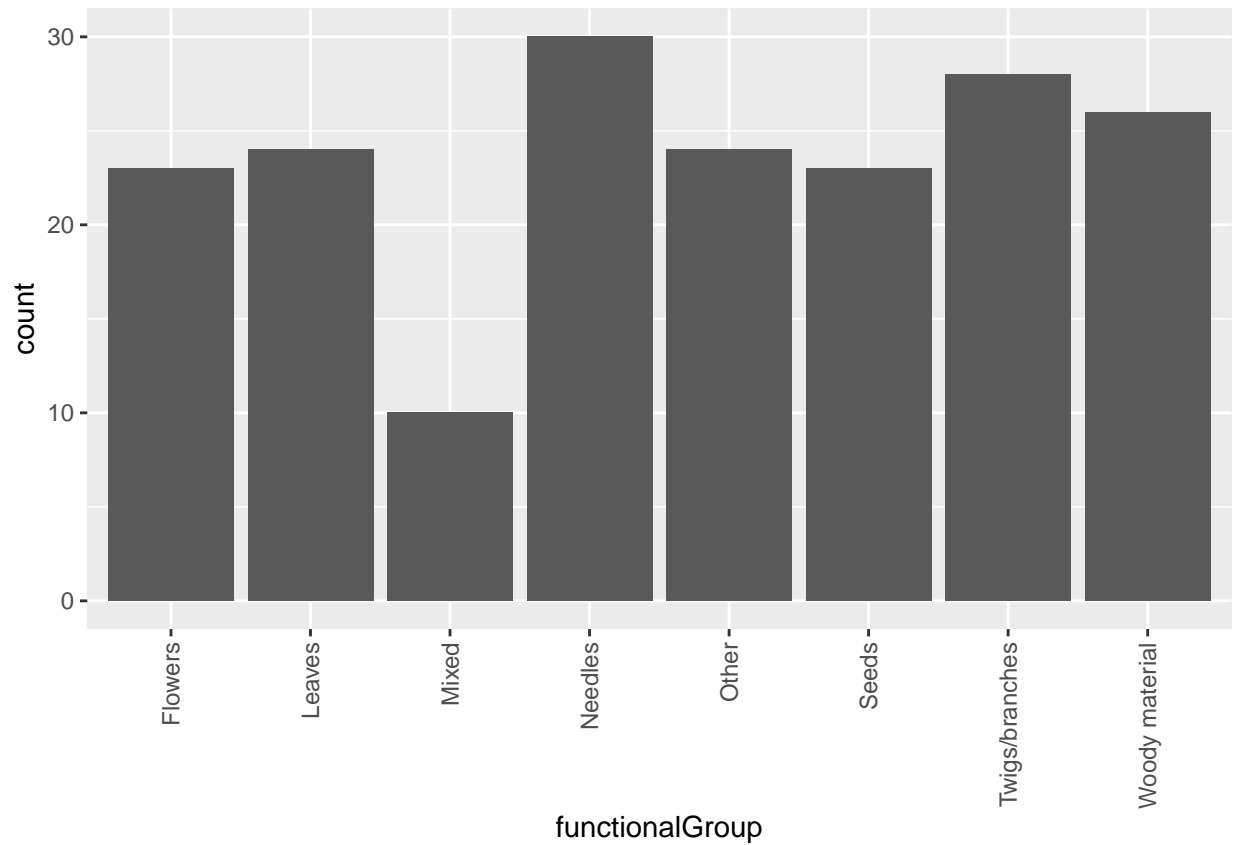
```
length(summary(Litter$plotID))
```

```
## [1] 12
```

Answer: The ‘unique’ only lists each plot’s ID and how many different plots, without showing other information, while the ‘summary’ shows not only each plot’s ID but also the number of each plot. However, the summary does not directly give the levels of the plot’s ID, we need to count it one by one. If we use ‘length’, we can directly see how many different plots were sampled.

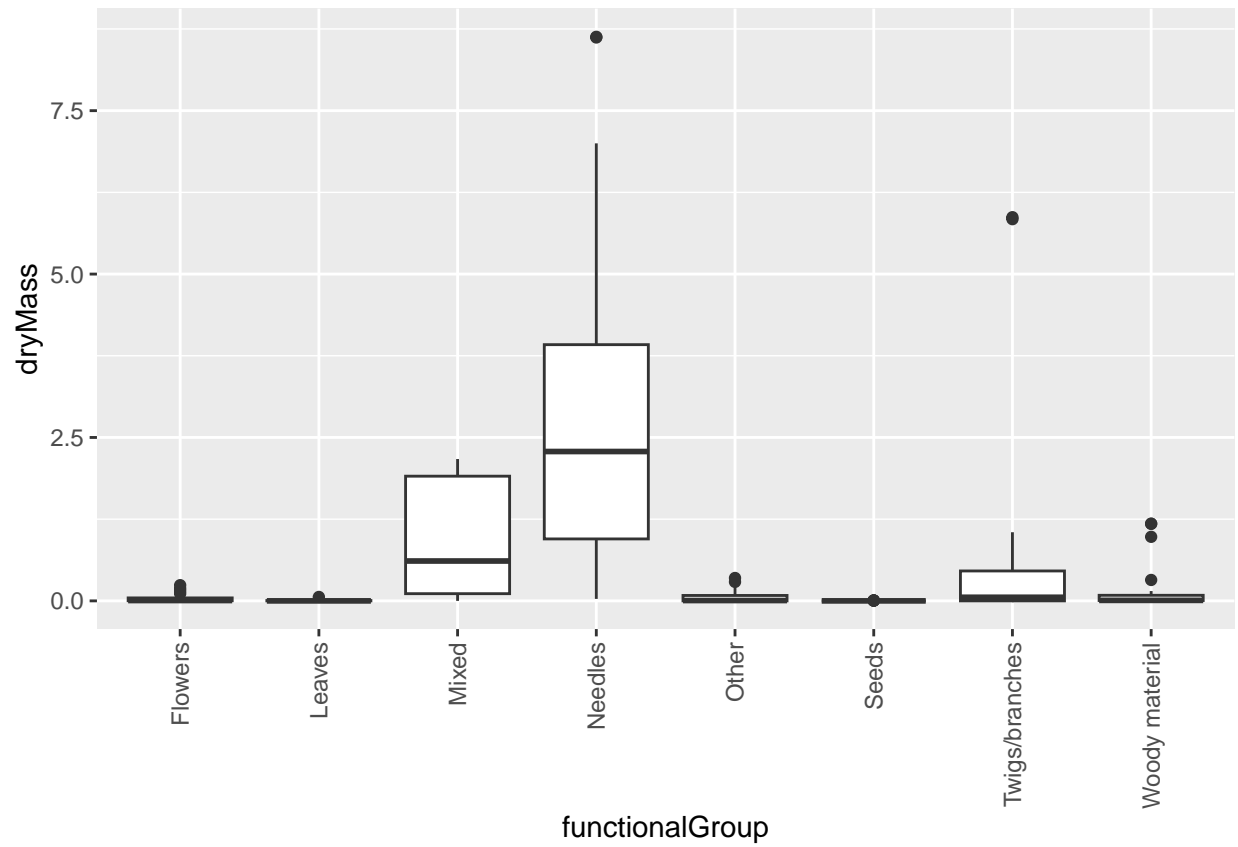
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter, aes(x=functionalGroup)) + geom_bar() +
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))
```

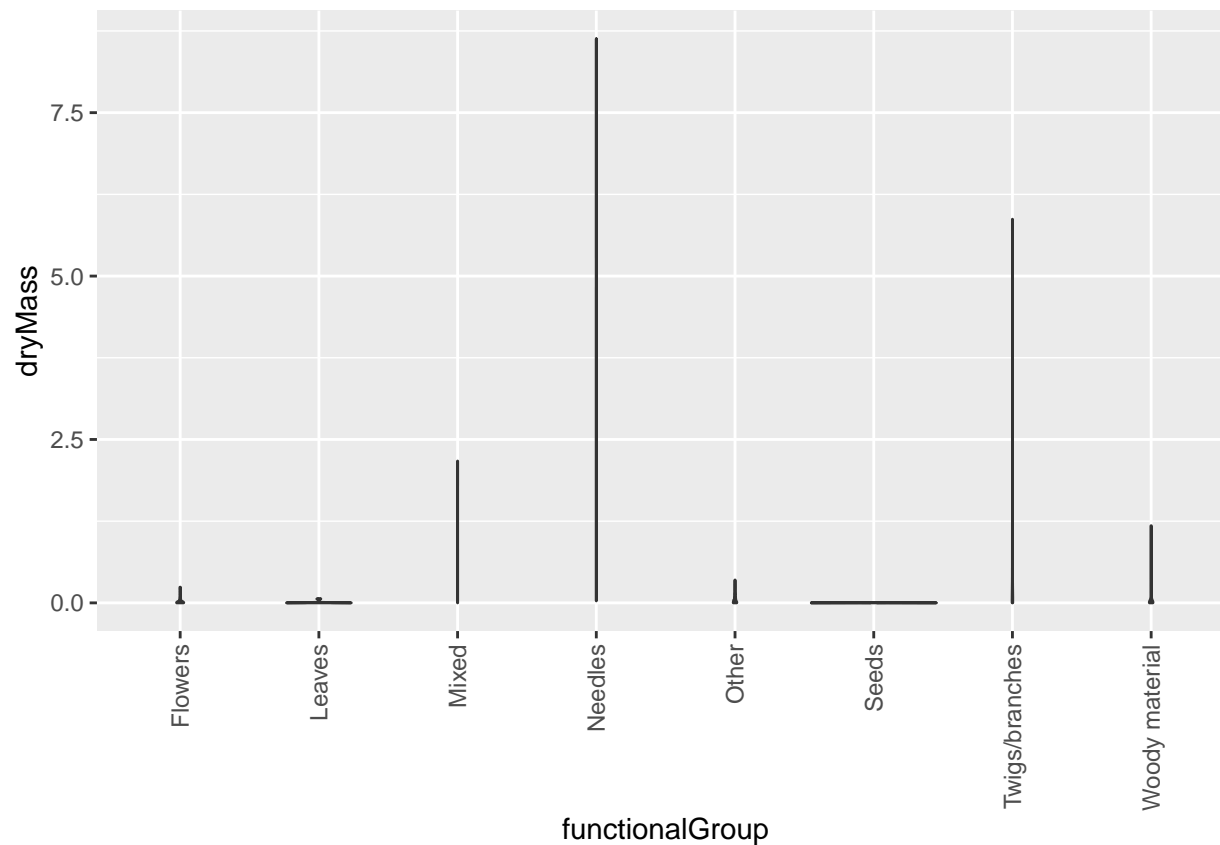


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by `functionalGroup`.

```
ggplot(Litter, aes(y=dryMass, x=functionalGroup)) +geom_boxplot()+  
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))
```



```
ggplot(Litter, aes(y=dryMass, x=functionalGroup)) +geom_violin()+  
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: Because most of the functional groups' dry mass is smaller than 1, while some of the groups have dry mass larger than 5, the medians of functional groups are all smaller than 2.5. A boxplot can clearly show the median, range, and outliers. While the violin plot cannot show the median and only shows the range.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles tend to have the highest biomass at these sites.