

Fecal Microbiome Bioinformatics Analysis GEN711 Final Project

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Background: Original Data

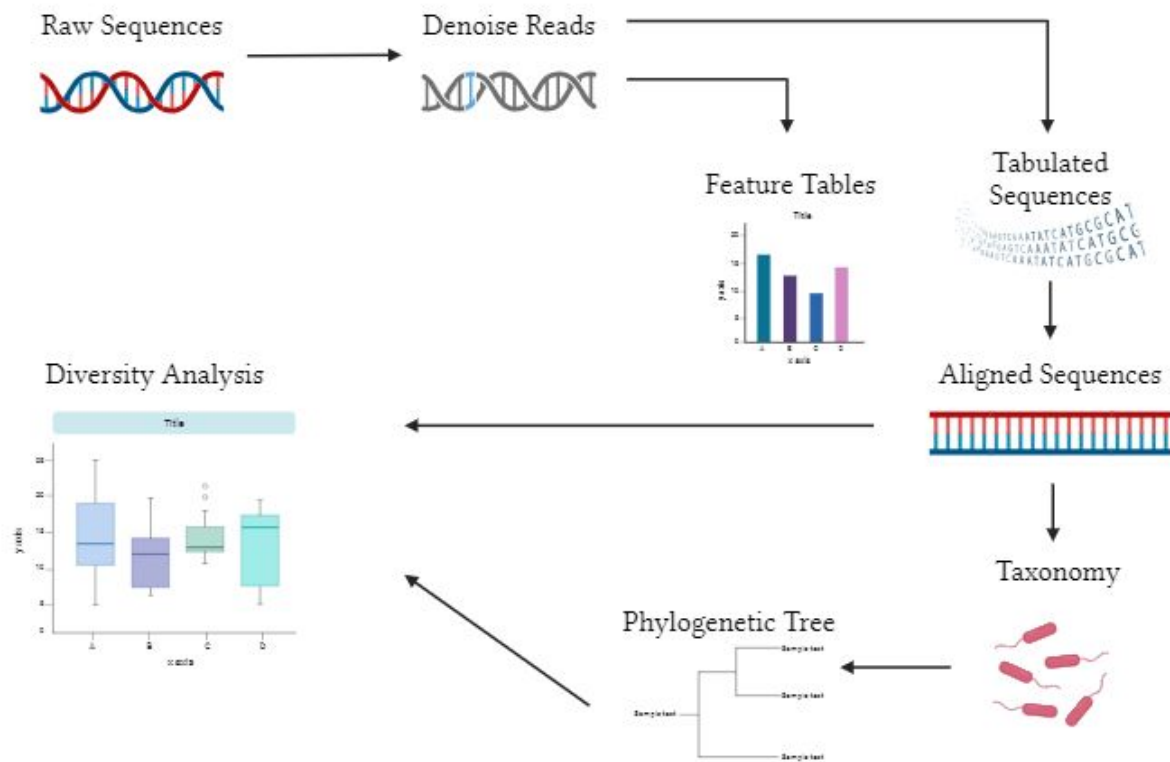
- This data was taken from a study on the correlation between the human gut microbiome and autism spectrum disorder (ASD) gastrointestinal issues
- The main goal of this study was to evaluate the effect of Microbiota Transfer Therapy (MTT) on the GI of ASD-diagnosed children and overall how that impacted ASD symptoms

Kang, DW., Adams, J.B., Gregory, A.C. et al. Microbiota Transfer Therapy alters gut ecosystem and improves gastrointestinal and autism symptoms: an open-label study. *Microbiome* 5, 10 (2017). <https://doi.org/10.1186/s40168-016-0225-7>

Background: Data Collection

- One group received the MTT fecal transplant (treatment group) and one group did not receive the transplant (control group)
- Stool collections and fecal swab were collected bi-weekly for 12 weeks
- DNA was isolated from both sample collection avenues with PowerSoil® DNA Isolation Kit
- A 16S rRNA library prep from Illumina MiSeq was performed to amplify specifically bacterial and archaeal DNA
 - Primer tag: 515f-806r
- 10% of total data was analyzed

Methods: Flowchart

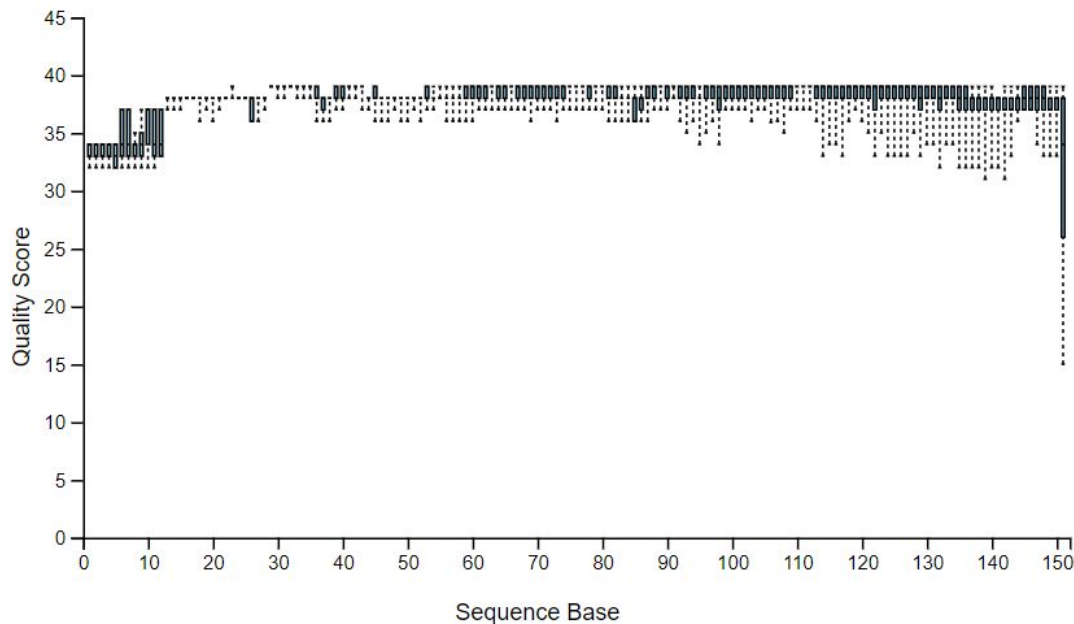


Methods: Data Import and Qiime Installation

- Performed “wget” command to download metadata and raw reads
 - Downloaded forward and reverse reads
- Installed “qiime2-2022.2” onto local device using “wget” and “conda env create”
 - Enabled all “qiime2” analysis to be done in terminal

Methods: Denoising

- “qiime demux summarize”
 - first 13 bases will be trimmed
 - 150 bases total length



Methods: Denoising continued

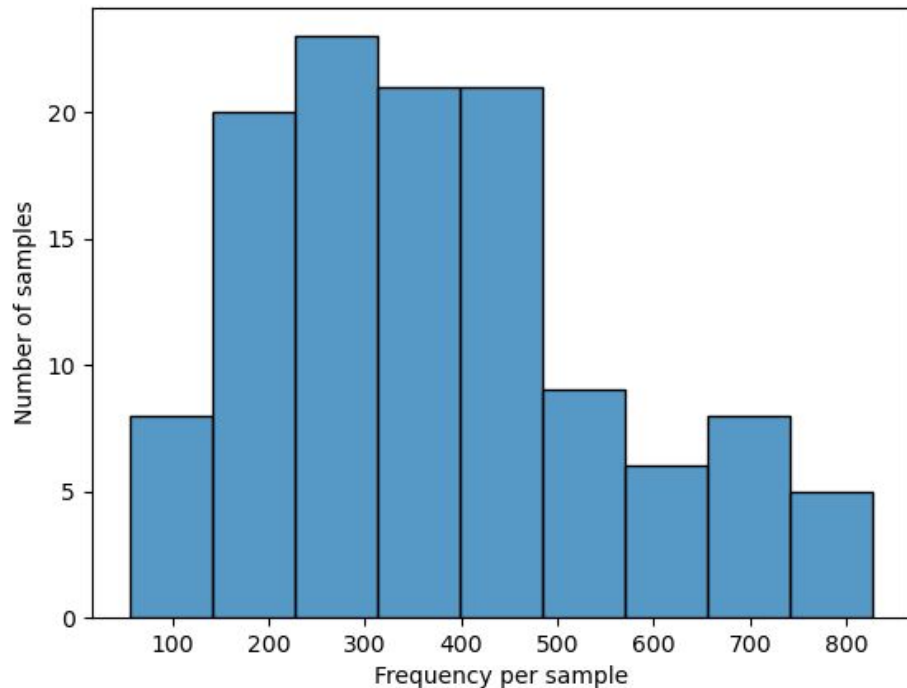
- “qiime dada2 denoise-single”
 - trimmed and truncated both raw reads
- “qiime metadata tabulate”
 - output table containing all read information (feature ID, filtering statistics, how much of the data was denoised/passed filter)

sample-id #q2:types	input numeric	filtered numeric	percentage of input passed filter numeric	denoised numeric	non-chimeric numeric	percentage of input non-chimeric numeric
0bb5e24a.aa34.48e7.b1f9.e761ac2dc6b4	6621	6488	97.99	6394	6141	92.75
101c02ce.4f8d.4394.83a3.0c406831d934	6634	6545	98.66	6473	6473	97.57
104e5902.1c3e.417f.bb22.124bfca61a61	7952	7795	98.03	7663	7040	88.53

Methods: Merging Reads and Alignment

- “qiime feature-table merge”
 - merge denoised sequences together and create new aligned clean DNA
- “qiime feature-table summarize”
 - create frequency table from merged sequences
- “qiime feature-table tabulate-seqs”
 - show table of merged DNA

Methods: Merging Reads and Alignment continued



Feature ID	Sequence Length	Sequence
002e78333d6cf2b11aa7a5ba03dd2c68	137	AGCGTTATCCGGATTTATTGGGTTTAAAGGGCGCGTAGGCCGGGAGC
0381153be965e62e229b3a5abb22386a	137	AGCGTTGTCCGGAATTATTGGGCGTAAAGCGCGCAGGCCGCTTT
039054e9b9aeede42b5d6dcd141dad55	137	GGCGTTATCCGGATTTATTGGGTTTAAAGGGAGCGTAGGCCGTGAG
0410886f68af125d1ae58b5ab40ae05f	137	AGCGTTATCCGGATTTACTGGGTGTAAGGGAGCGTAGACGGTTAA
04aa965c153b62d0f35b450bb1472f9f	137	AGCGTTATCCGGATTTATTGGGTTTAAAGGGAGCGTAGGTGGATTG
050d9d1c576251d7653dd965fd603f58	137	AGCGTTGTCCGGATTTACTGGGTGTAAGGGCGGTAGCCGGGAGG
059bed64e31fbc55d7c1f19e9837bf5f	137	AGCGTTGTCCGGATTTACTGGGTGTAAGGGCGGTAGCCGGGAAG
05ddf0ef80a162cf33bffa987d2490e	137	AGCGTTATCCGGATTTACTGGGTGTAAGGGAGGTAGGTGGTATC
0677a32afd11e65443b18c0fc4192708	137	AGCGTTGTCCGGAATGATTGGGCGTAAAGGGCGCGTAGGCCGCTG
073945437c2d32d92ca4f8a9ca6b1103	137	AGCGTTATCCGGAATGACTGGGCGTAAAGGGTGCGTAGGTGGCCTA
07ca0db2bf8582a081f72bd1d6cb1eed	137	AGCGTTATCCGGATTTACTGGGTGTAAGGGAGCGTAGACGCCGCA
0817bbd38bcdcc2c5e83b94e69301ab8	137	AGCGTTGTCCGGAATTACTGGGCGTAAAGGGTGCGTAGCGGTTAT

“qiime feature-table summarize” (left) and “qiime feature-table tabulate-seqs” (right)

Methods: Taxonomy Assignment

- Downloaded a reference classifier for human stool from SILVA rRNA database using “wget”
- “qiime feature-classifier classify-sklearn” and “qiime metadata tabulate”
 - uses reference classifier to compare sequences to known bacteria in human stool and create a list of taxa found

Feature ID #q2:types	Taxon categorical	Confidence categorical
002e78333d6cf2b11aa7a5ba03dd2c68	d__Bacteria; p__Bacteroidota; c__Bacteroidia; o__Bacteroidales; f__Marinifilaceae; g__Butyricimonas	0.9997788497367014
0046913ae6f9e12dbd889671ed26c09d	d__Bacteria; p__Firmicutes; c__Negativicutes; o__Veillonellales-Selenomonadales; f__Veillonellaceae; g__Megasphaera; s__uncultured_bacterium	0.9685055485925435

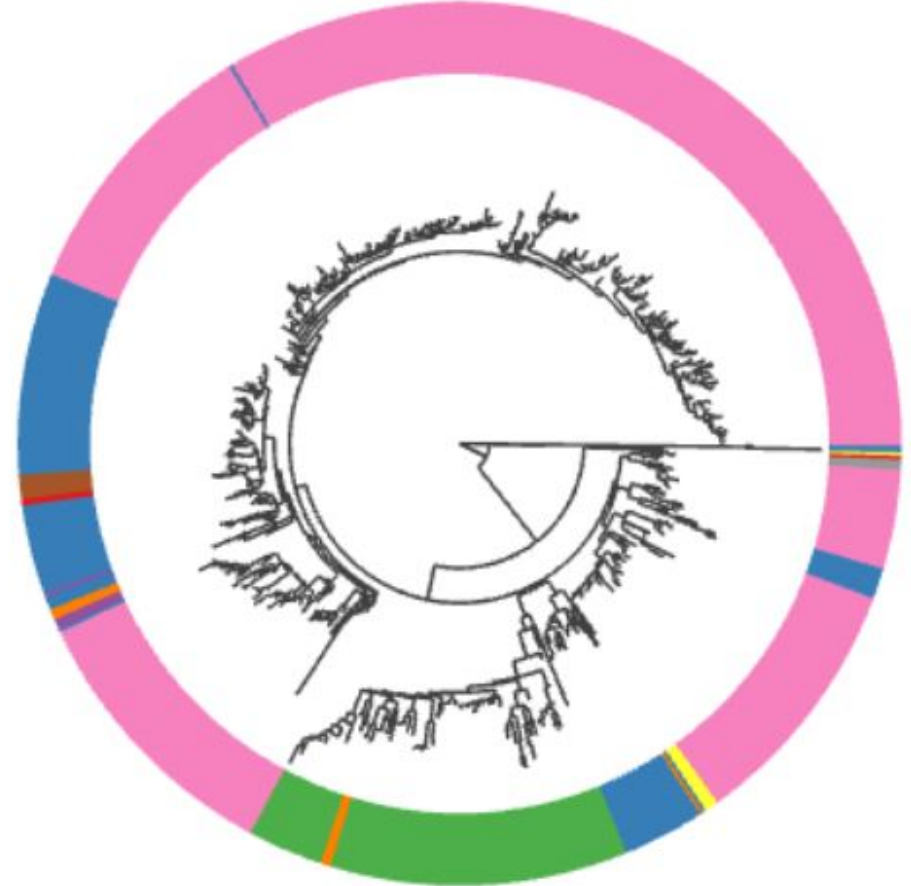
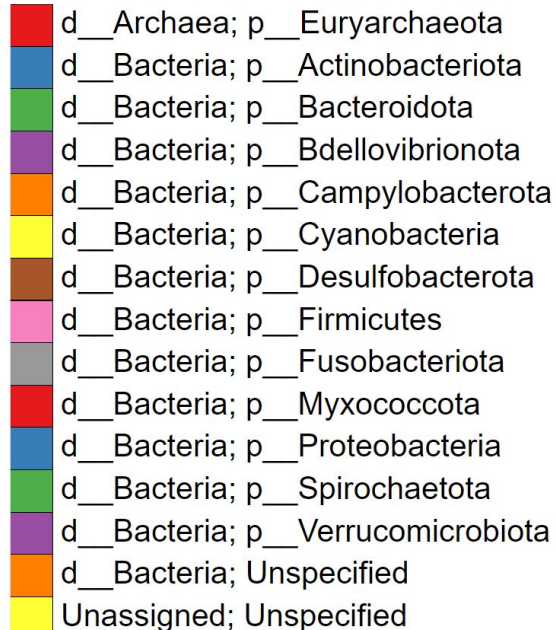
Methods: Phylogenetic Tree Assembly

- “qiime phylogeny align-to-tree-mafft-fasttree”
 - utilizes all of the aligned sequences and taxonomic analysis to construct data for a phylogenetic tree
- “qiime empress tree-plot”
 - had to install the “empress” extension for qiime
 - creates a physical phylogenetic tree for the data

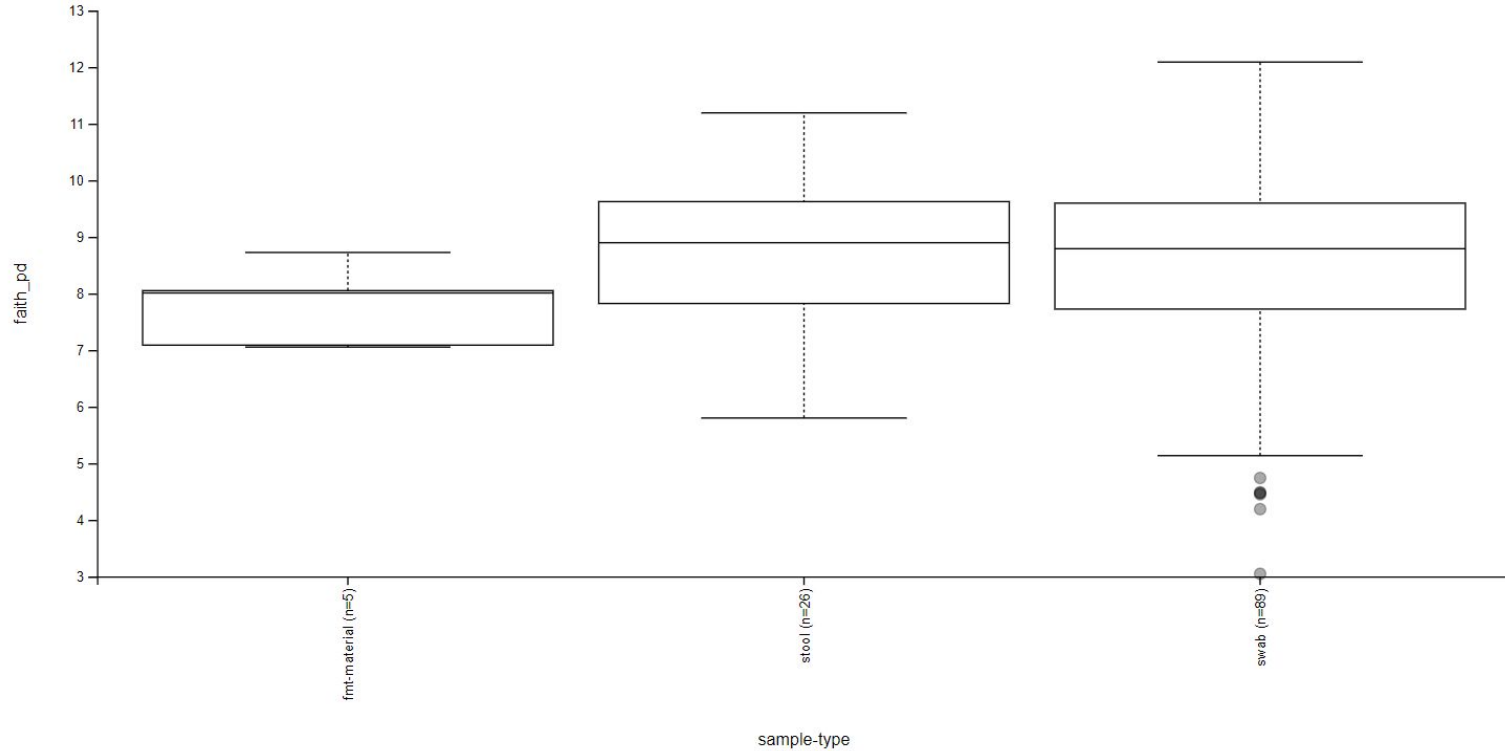
Methods: Alpha and Beta Diversity Analysis

- “qiime diversity core-metrics-phylogenetic”
 - set the initial parameters for alpha and beta analysis
- “qiime diversity alpha-group-significance”
 - produced graphs regarding the data richness and evenness
- “qiime diversity beta-group-significance”
 - produced several graphs to compare how the treatment groups and sample types are associated

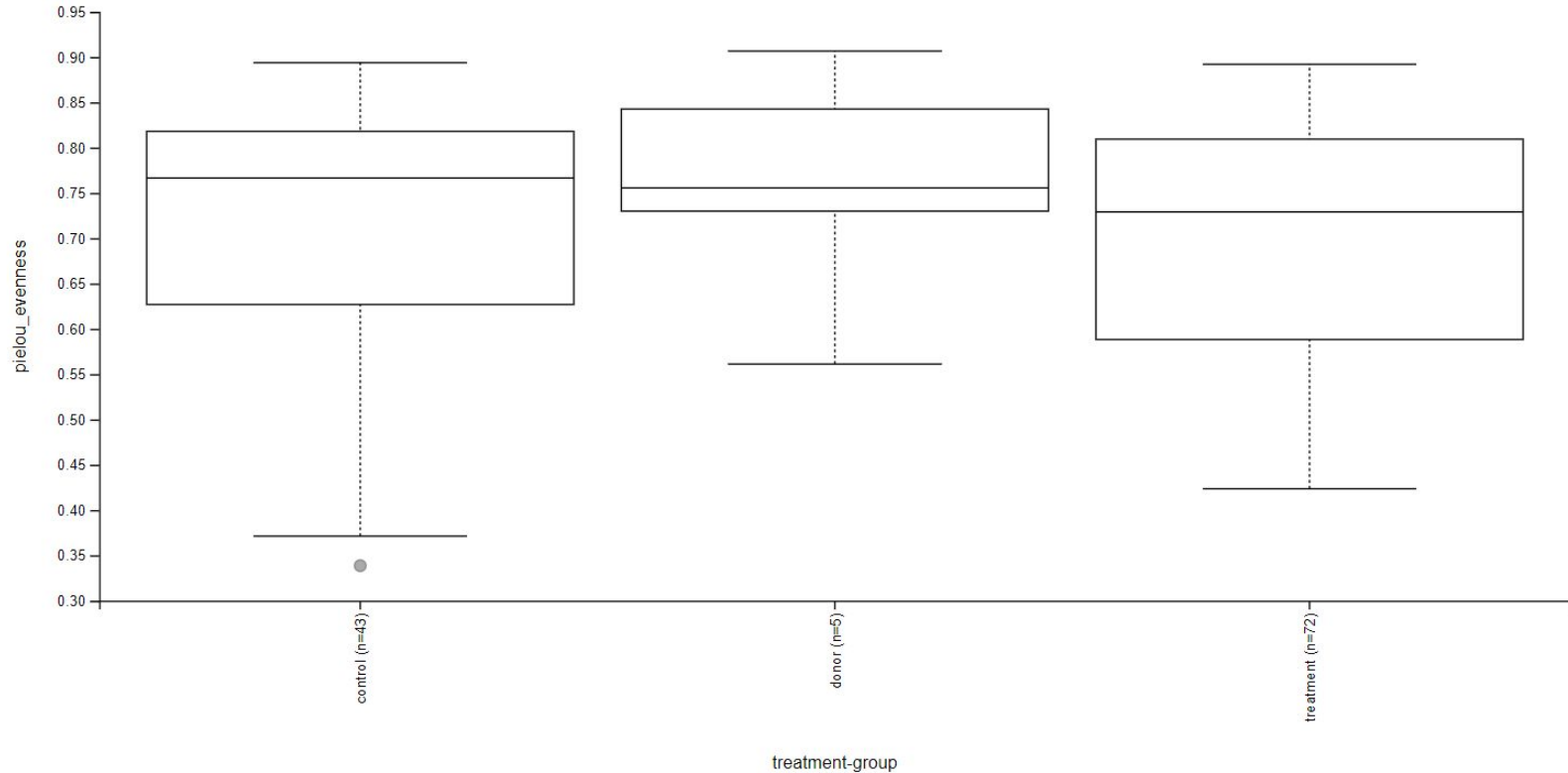
Results: Phylogenetic Tree



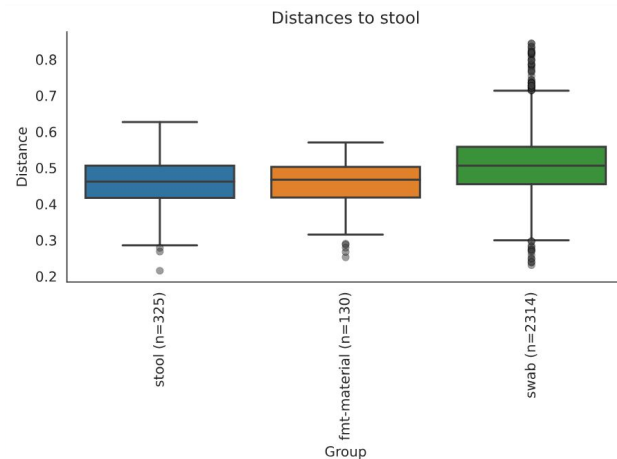
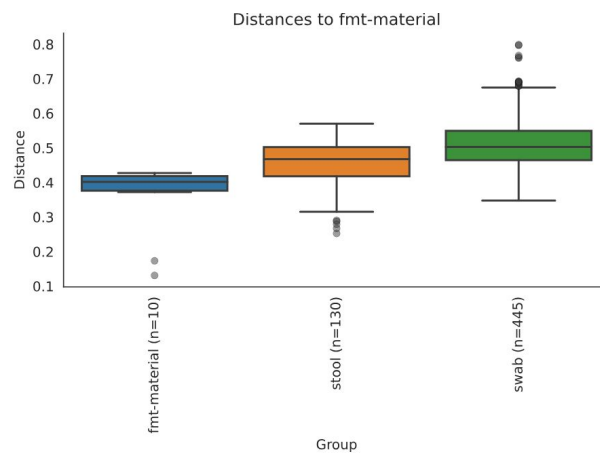
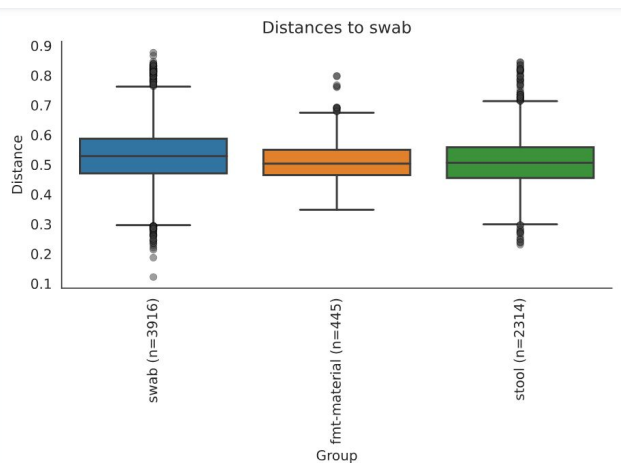
Results: Alpha Diversity (Richness)



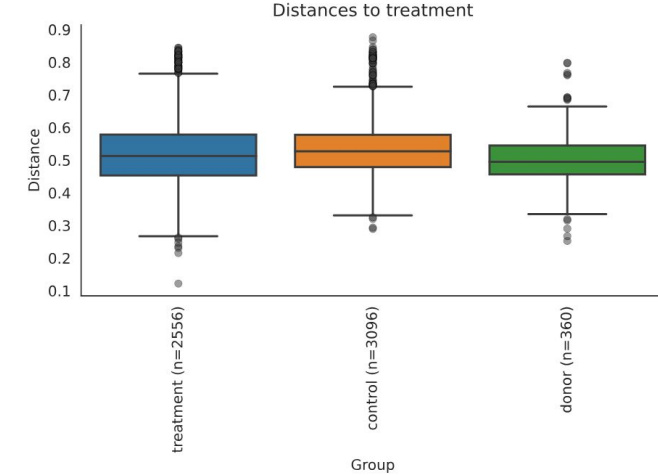
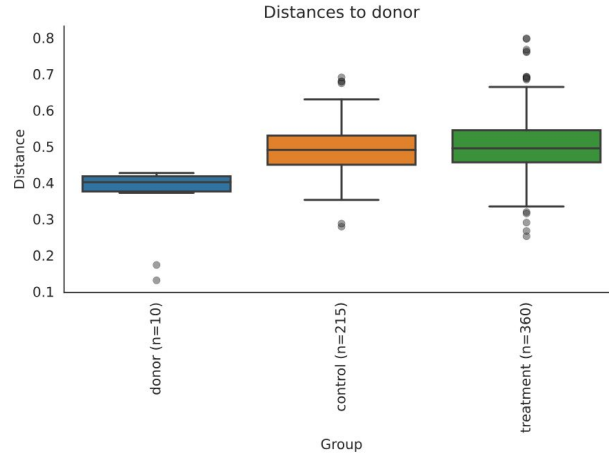
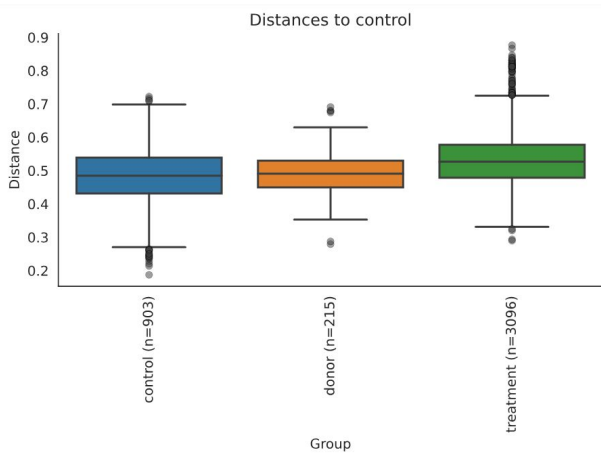
Results: Alpha Diversity (Evenness)



Results: Beta Diversity (Sample Group)



Results: Beta Diversity (Treatment Group)



Major Takeaways:

- qiime2 is an incredibly useful tool to analyze raw reads and transform them into visual statistics
- The most difficult part of this project was trying to download all files onto a local device to be viewed
- Best analysis and result: taxonomic assignment and the phylogenetic tree
 - Best visualization of bacteria present

References

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