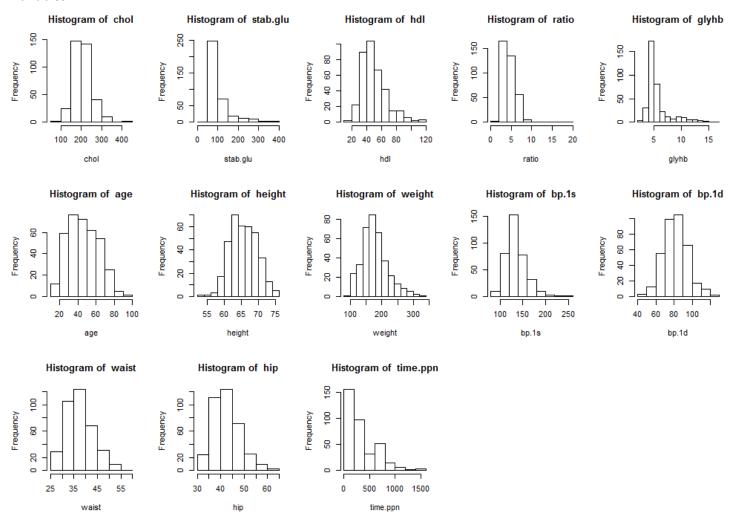
Statistics 108, Project 1

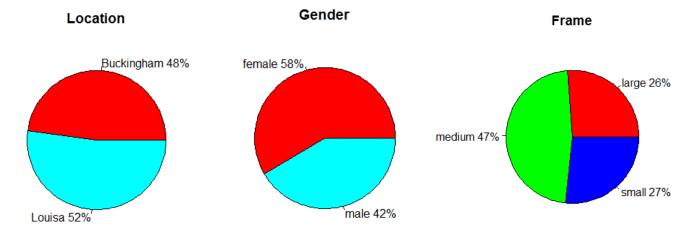
Data exploration and split data for validation later on:

1.

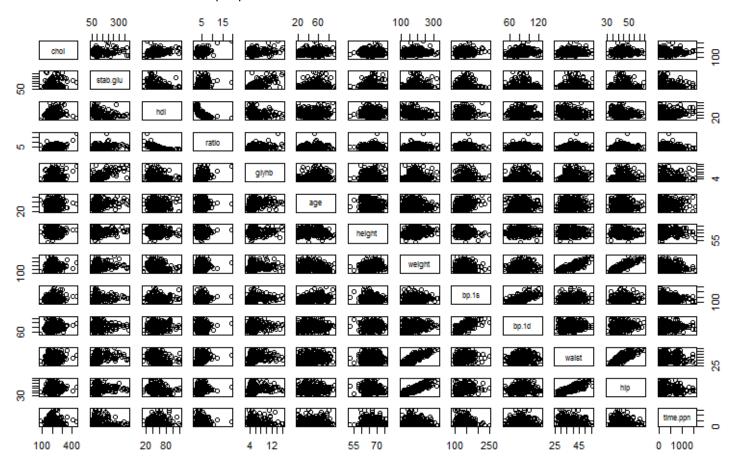
Among all variables, location, gender and frame are qualitative variables. The rest of the variables are quantitative variables.



The distribution of the histogram of chol has heavy tails. The distribution of the histogram of stab.glu, hdl, ratio, glyhd, weight, bp.1s, and time.ppn are right-skewed. The distribuion of the histogram of age, height, bp.1d, waist and hip are approximately normal.

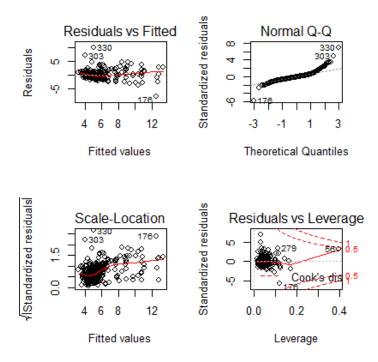


The pie chart of the location shows that the distribution of Buckingham and Louisa are about the same. The pie chart of the gender shows that the there are more females than male in the dataset. The pie chart of the frame shows that about half of the data are collected from people have a medium frame.



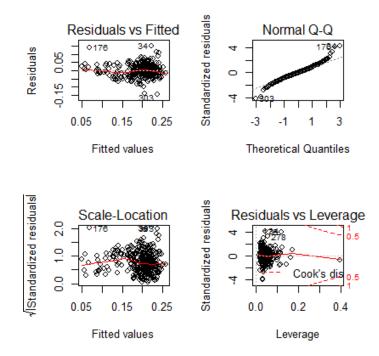
From the pairwise correlation matrix for all quantitative variables, we can see that some of the variables are correlated. Because some of the correlation plots show linear patterns or particular patterns.

2.



In Residuals vs Fitted plot, the reference line is approximately horizontal at x=0. Thus, the linearity assumption is hold. In the Normal Q-Q plot, we can see that points are spread below the refence line the left end and above the reference line at the right end, so the residuals are not normally distributed, and it has heavy tails. In the Scale-Location plot, we can see that the reference line is not horizonal, so the equal variance assumption doesn't hold. The Residuals vs Leverage plot shows no outlier.

3.



First applied boxcox function to the model1, the boxcox plot indicates that $(glyhd)^{-1}$ is a necessary transformation on glyhd. After applied to transformation, new diagnostic plots were plotted. We can tell that the equal variance assumption approximately holds on the new model; however, the normality assumption is still not hold. By applied the boxcox function to model2 again, the boxcox plot suggest that no further transformation on glyhd is needed.

4.

```
#problem 4#
set.seed(10)
N=nrow(data)
index=sample(1:N, size=N/2, replace=FALSE)
data.t=data[index,]
data.v=data[-index,]
```

Selection of first-order effects

5.

```
#problem5#
model3=lm(glyhb~.,data.t)
summary(model3)
length(model3$coefficients)
MSE=sum((data.t$glyhb-model3$fitted.values)^2)/166
```

There are 17 regression coefficients in model 3, and the MSE ≈ 0.00138

6.

Return of top 1 best subset of each subset size :

Subset sizes	Best subsets
null	null
1	Stab.glu
2	Stab.glu, age
3	Stab.glu, age, waist,
4	Stab.glu, age, waist, ratio
5	Stab.glu, age, waist, ratio, frame\$small
6	Stab.glu, age, waist, ratio, frame\$small, time.ppn
7	Stab.glu, age, waist, ratio, frame\$small, time.ppn, bp.1s
8	Stab.glu, age, waist, ratio, frame\$small, time.ppn, bp.1s,
	height
9	Stab.glu, age, waist, ratio, frame\$small, time.ppn, heap,
	height, weight,
10	Stab.glu, age, waist, ratio, frame\$small, time.ppn, bp.1s,
	height, weight, heap
11	Stab.glu, age, waist, ratio, frame\$small, time.ppn, bp.1s,
	height, weight, heap, chol
12	Stab.glu, age, waist, ratio, frame\$small, time.ppn, bp.1s,
	height, weight, heap, chol, location\$louisa,
13	Stab.glu, age, waist, ratio, frame\$small, time.ppn, bp.1s,
	height, weight, heap, chol, location\$louisa, hdl
14	Stab.glu, age, waist, ratio, frame\$small, time.ppn, bp.1s,
	height, weight, heap, chol, location\$louisa, hdl,
	frame\$medium

Zuck Liu	313020133	11/30/17
15	Stab.glu, age, waist, ratio, frame\$small,	time.ppn, bp.1s,
	height, weight, heap, chol, location\$lou	isa, hdl,
	frame\$medium, bp.1d	
16	Stab.glu, age, waist, ratio, frame\$small,	time.ppn, bp.1s,
	height, weight, heap, chol, location\$lou	isa, hdl,
	frame\$medium, bp.1d, gender\$male	

SSEp, $R_{\rm p}^2$, $R_{\rm a,p}^2$, Cp, AICp, BICp for the best model of each subset:

```
R^2_a
none 0.5158646 0.0000000 0.0000000 191.73453170 -1072.466 -1069.256
     0.2864076 0.4448009 0.4417335 27.96351331 -1178.148 -1171.729
2
     0.2574112 0.5010102 0.4954659
                                     9.01014928 -1195.682 -1186.053
3
     0.2428890 0.5291612 0.5212701
                                     0.51619889 -1204.309 -1191.471
     0.2401432 0.5344840 0.5240230
                                     0.53201659 -1204.389 -1188.342
     0.2367131 0.5411332 0.5281708
                                     0.05337754 -1205.022 -1185.765
     0.2343460 0.5457220 0.5302352
                                     0.34280455 -1204.861 -1182.395
7
     0.2331725 0.5479966 0.5299165
                                     1.49487219 -1203.780 -1178.104
8
     0.2326634 0.5489836 0.5282473
                                     3.12693590 -1202.180 -1173.294
9
     0.2314193 0.5513952 0.5280574
                                     4.22797088 -1201.161 -1169.066
     0.2303187 0.5535287 0.5275711
10
                                     5.43265348 -1200.033 -1164.729
     0.2300477 0.5540541 0.5253676
                                     7.23678869 -1198.249 -1159.735
11
12
     0.2299216 0.5542986 0.5228374
                                     9.14564365 -1196.349 -1154.626
13
     0.2298166 0.5545020 0.5202329 11.06983181 -1194.433 -1149.500
14
     0.2297510 0.5546292 0.5175150 13.02241521 -1192.485 -1144.343
15
     0.2297274 0.5546751 0.5146758 15.00531267 -1190.504 -1139.152
     0.2297200 0.5546893 0.5117678 17.00000000 -1188.510 -1133.948
16
```

Best models According to different criterions:

Criterions	Best model
SSEp	Full model
R_p^2	Full model
$R_{a,p}^2$	Best Model of subset size 6
C_p	Best model of subset size 5
AIC_p	Best model of subset size 5
BIC_p	Best model of subset size 3

The C_p value of the best model according C_p criterion is 0.05337754, and the p is 6. The C_p value is quite smaller than the p. One possible explanation is that we use MSE of full model as an unbiased estimator of sigma in measuring C_p , and the MSE is overestimated.

```
\label{local-control} $$ model3.1=lm(glyhb\sim stab.glu+age+waist+ratio+framesmall,data.t) $$ model3.2=lm(glyhb\sim stab.glu+age+waist,data.t) $$ model3.3=lm(glyhb\sim stab.glu+age+waist+ratio+framesmall+time.ppn,data.t) $$
```

Selection of first- and second- order effects

7.

There are 136 regression coefficients in this model. The $MSE \approx 0.001036$. Model 4 has too many regression coefficients and we would lose lots of degree of freedom by fitting the data into this model.

```
#problem 7#
model4=lm(glyhb~.^2,data.t)
summary(model4)
length(model4$coefficients)
MSE=sum((data.t$glyhb-model4$fitted.values)^2)/47
```

8.

The model being selected:

```
glyhb ~ stab.glu + age + waist + ratio + stab.glu:ratio + age:ratio
```

The AIC of model.fs1 is -1205.14, it is slightly smaller than the AIC of model3.1.

```
\label{localization} $$  \mbox{model.fs1=stepAIC(fit0, scope=list(upper=lm(glyhb\sim. \land 2, data=data.t), lower=$\sim$1), direction="both", k=2) $$  \mbox{lower} = \mbox{lower}
```

9.

The model being selected:

```
glyhb ~ chol + stab.glu + hdl + ratio + age + gender + height +
   weight + bp.1s + bp.1d + waist + hip + time.ppn + stab.glu:gender +
   hdl:ratio + age:bp.1d + weight:bp.1s + age:hip + hip:time.ppn +
   gender:height + stab.glu:bp.1s + stab.glu:time.ppn + stab.glu:waist +
   age:waist + chol:time.ppn + hdl:weight + bp.1d:waist + weight:hip
```

The AIC of model.fs2 is -1230.61 and it is significantly smaller than the AIC of model.fs1.

```
#problem9#
model.fs2=s|tepAIC(model3, scope=list(upper=lm(glyhb~.^2,data=data.t), lower=~1), direction="both", k=2)
```

10.

```
BIC of model.fs1= -1182.677
BIC of model.fs2= -1137.536
```

The BIC of model.fs1 has a smaller value than the BIC of model.fs2. BIC and AIC choose different models.

```
\label{eq:bic=n*log} bic=n*log(sse.fs1)+log(n)*length(model.fs1$coefficients)-n*log(n)\\ bic=n*log(sse.fs2)+log(n)*length(model.fs2$coefficients)-n*log(n)\\ model4.1=model.fs2\\ model4.2=model.fs1
```

Model validation

11.

PRESS of model3.1=0.25278

PRESS of model3.2=0.25398

PRESS of model3.3=0.25257

PRESS of model4.1=0.21719

PRESS of model4.2=0.25348

From the result of PRESS, we can see that the value of PRESS is close to the value of SSE respectively.

By comparing the PRESS of each model, the model4.1 has a smallest PRESS values.

```
press.3.1=sum((model3.1$residuals/(1-lm.influence(model3.1)$hat))^2)
press.3.2=sum((model3.2$residuals/(1-lm.influence(model3.2)$hat))^2)
press.3.3=sum((model3.3$residuals/(1-lm.influence(model3.3)$hat))^2)
press.4.1=sum((model4.1$residuals/(1-lm.influence(model4.1)$hat))^2)
press.4.2=sum((model4.2$residuals/(1-lm.influence(model4.2)$hat))^2)
```

12.

MSPR of model3.1=0.001368	PRESS/n= 0.001381295
MSPR of model3.2=0.001377	PRESS/n= 0.001387887
MSPR of model3.3=0.001341	PRESS/n= 0.001380191
MSPR of model4.1=0.001798	PRESS/n= 0.001186856
MSPR of model4.2=0.001526	PRESS/n= 0.001385155

By comparing the MSPR of the models to the respective PRESS/n, the MSPR of model3.1, model3.2, and model3.3 are slightly smaller than their respective PRESS/n, and the MSPR of model4.1 and model4.2 are respectively much larger than their respective PRESS/n.

Model3.1 has the smallest MSPR.

```
MSPR3.1=sum((data.v$glyhb-predict(model3.1,data.v))^2)/n
MSPR3.2=sum((data.v$glyhb-predict(model3.2,data.v))^2)/n
MSPR3.3=sum((data.v$glyhb-predict(model3.3,data.v))^2)/n
MSPR4.1=sum((data.v$glyhb-predict(model4.1,data.v))^2)/n
MSPR4.2=sum((data.v$glyhb-predict(model4.2,data.v))^2)/n
```

13.

The final model I would choose is model3.3, because it has a smallest MSPR. It indicates that model3.3 has a better predictive capacity. Also, model3.3 has significantly less predictor variables than model 4.1, which has the smallest PRESS.

Fitted Regression function:

```
lm(formula = glyhb ~ stab.glu + age + waist + ratio + framesmall +
    time.ppn, data = data)
```

```
> summary(finalmodel)
call:
lm(formula = glyhb ~ stab.glu + age + waist + ratio + framesmall +
    time.ppn, data = data)
Residuals:
      Min
                 10
                       Median
                                      30
                                               Max
-0.154503 -0.020705 -0.001382 0.019680 0.150207
Coefficients:
              Estimate Std. Error t value Pr(>|t|)
(Intercept) 3.414e-01 1.536e-02 22.221 < 2e-16 ***
           -4.947e-04 3.824e-05 -12.937 < 2e-16 ***
            -6.525e-04 1.230e-04 -5.306 1.97e-07 ***
-1.061e-03 3.737e-04 -2.839 0.00479 **
-3.665e-03 1.187e-03 -3.088 0.00217 **
age
waist
ratio
framesmall 2.008e-03 4.774e-03 0.421 0.67422
time.ppn
            -1.328e-05 6.176e-06 -2.150 0.03223 *
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 0.03628 on 359 degrees of freedom
Multiple R-squared: 0.5075, Adjusted R-squared: 0.4993
F-statistic: 61.66 on 6 and 359 DF, p-value: < 2.2e-16
> anova(finalmodel)
Analysis of Variance Table
Response: glyhb
               Df Sum Sq Mean Sq F value
                                                    Pr(>F)
```

1 0.39753 0.39753 302.0648 < 2.2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

36.9817 3.053e-09 ***

16.1450 7.151e-05 ***

9.6925 0.001999 **

4.6223 0.032227 *

0.4640 0.496221

1 0.04867 0.04867

1 0.02125 0.02125

1 0.01276 0.01276

1 0.00608 0.00608

framesmall 1 0.00061 0.00061

Residuals 359 0.47245 0.00132

stab.glu

time.ppn

age

waist ratio R code:

```
data=read.table("diabetes.txt",header=T)
    attach(data)
 3
 4
   #problem 1#
 5 str(data)
 6 name=names(data)
 7
   #histogram#
 8 par(mfrow=c(3,5))
 9 par(mfrow=c(1,1))
10 - for (i in seq_along(data)){
11 · if (i==6||i==8||i==11){
12
        next
13
      variable=as.name(name[i])
14
15
      hist(eval(variable),xlab=name[i],main=paste("Histogram of ", name[i], sep = ""))
16
17
   #pie chart#
18 table(location)
19 count=c(175,191)
20 pct <- round(count/sum(count)*100)</pre>
21 lbls=c("Buckingham","Louisa")
22 lbls=paste(lbls,pct,"%",sep="")
23 pie(count, labels=lbls, main="Location", col=rainbow(length(lbls)))
24 table(gender)
25 count=c(214,152)
26 pct <- round(count/sum(count)*100)
   lbls=c("female ","male ")
27
28 lbls=paste(lbls,pct,"%",sep="")
   pie(count, labels=lbls, main="Gender", col=rainbow(length(lbls)))
29
30 table(frame)
31 count=c(96,172,98)
32 pct <- round(count/sum(count)*100)</pre>
33
    pct
   lbls=c("large ","medium ","small ")
35 lbls=paste(lbls,pct,"%",sep="")
    pie(count, labels=lbls, main="Frame", col=rainbow(length(lbls)))
36
37
    #scatterplot matrix and pairwise correlation matrix#
38
    pairs(data[-c(6,8,11)])
39
   #problem 2#
40
41 model1=lm(glyhb~.,data)
42 summary(model1)
43 par(mfrow=c(2,2))
44 plot(model1)
45
```

```
46 #problem 3#
47 boxcox(model1)
48 data$glyhb=glyhb^-1
49 model2=lm(glyhb~.,data)
50 summary(mode12)
51 plot(model2)
52 boxcox(model2)
53
54 #problem 4#
55 set.seed(10)
56 N=nrow(data)
57 index=sample(1:N, size=N/2, replace=FALSE)
58 data.t=data[index,]
59 data.v=data[-index,]
60
61 #problem5#
62 model3=lm(glyhb~.,data.t)
63 summary(model3)
64 length(model3$coefficients)
65 MSE=sum((data.t$glyhb-model3$fitted.values)^2)/166
66 MSE
67 #problem 6#
68 library(leaps)
69 library(MASS)
70 best=regsubsets(glyhb~., data=data.t, nbest=1, nvmax=16)
71 sum_sub=summary(best)
72 sum_sub$which
73 n=nrow(data.t)
74 p.m=2:17
75 sse=sum_sub$rss
76 sse
77 aic=n*log(sse)+2*p.m-n*log(n)
78 bic=n*log(sse)+log(n)*p.m-n*log(n)
79 fit0=lm(glyhb~1,data=data.t)
80 sse1=sum(fit0$residuals^2)
81 p=1
82 c1=sse1/0.001384-(n-2*p)
83 aic1=n*log(sse1)+2*p-n*log(n)
84 bic1=n*log(sse1)+log(n)*p-n*log(n)
85 none=c(1,rep(0,16),sse1,0,0,c1,aic1,bic1)
86 res_sub=cbind(sum_sub$which,sse,sum_sub$rsq,sum_sub$adjr2,sum_sub$cp,aic, bic)
87 res_sub=rbind(none,res_sub)
88 colnames(res_sub)=c(colnames(sum_sub$which), "sse", "R^2", "R^2_a", "Cp", "aic", "bic")
89 res_sub
90 frametype=model.matrix(~data.t$frame-1)
```

```
frametype
 91
     framesmall=frametype[,3]
 92
 93 framesmall
     model3.1=lm(glyhb~stab.glu+age+waist+ratio+framesmall,data.t)
 95 model3.2=lm(glyhb~stab.glu+age+waist,data.t)
 96 model3.3=lm(glyhb~stab.glu+age+waist+ratio+framesmall+time.ppn,data.t)
 97
 98 #problem 7#
 99 model4=lm(glyhb~.^2,data.t)
100 summary(model4)
101 length(model4$coefficients)
102
     MSE=sum((data.t$glyhb-model4$fitted.values)^2)/47
103
     #problem8#
104
     library(leaps)
     library(MASS)
105
106 model.fs1=stepAIC(fit0, scope=list(upper=lm(glyhb~.^2,data=data.t), lower=~1), direction="both", k=2)
107
     #problem9#
model.fs2=stepAIC(model3, scope=list(upper=lm(glyhb\sim.^2,data=data.t), lower=\sim1), direction="both", k=2)
109 #problem10#
110 sse.fs1=sum(model.fs1$residuals^2)
111 sse.fs2=sum(model.fs2$residuals^2)
112 bic=n*log(sse.fs1)+log(n)*length(model.fs1$coefficients)-n*log(n)
     bic=n*log(sse.fs2)+log(n)*length(model.fs2$coefficients)-n*log(n)
113
114 bic
115
     model4.1=model.fs2
116
     model4.2=model.fs1
117 #problem11#
118 press.3.1=sum((model3.1$residuals/(1-lm.influence(model3.1)$hat))^2)
119 press.3.2=sum((model3.2$residuals/(1-lm.influence(model3.2)$hat))^2)
120 press.3.3=sum((model3.3$residuals/(1-lm.influence(model3.3)$hat))^2)
press.4.1=sum((model4.1$residuals/(1-lm.influence(model4.1)$hat))^2)
press.4.2=sum((model4.2$residuals/(1-lm.influence(model4.2)$hat))^2)
sum(model3.1$residuals^2)
124 sum(model3.2$residuals^2)
125 sum(model3.3$residuals^2)
126 sum(model4.1$residuals^2)
127 sum(model4.2$residuals^2)
128 #problem12#
129 MSPR3.1=sum((data.v$glyhb-predict(model3.1,data.v))^2)/n
     MSPR3.2=sum((data.v$glyhb-predict(model3.2,data.v))^2)/n
130
131 MSPR3.3=sum((data.v$glyhb-predict(model3.3,data.v))^2)/n
132 MSPR4.1=sum((data.v$g]yhb-predict(model4.1,data.v))^2)/n
133 MSPR4.2=sum((data.v$g]yhb-predict(model4.2,data.v))^2)/n
134 press.3.1/n
135 press.3.2/n
 136 press. 3.3/n
 137
        press.4.1/n
 138
        press.4.2/n
 139
        #problem13#
 140
        frametype=model.matrix(~data$frame-1)
 141
        framesmall=frametype[,3]
        finalmodel=lm(glyhb~stab.glu+age+waist+ratio+framesmall+time.ppn,data)
 142
 143
        summary(finalmodel)
 144
        anova(finalmodel)
 145
```