Reconstruction

2023-03-10

Create a Seurat object for the reconstructed data

```
library(Seurat)
## Attaching SeuratObject
## Attaching sp
library(SeuratObject)
library(SeuratData)
## -- Installed datasets ------ SeuratData v0.2.2 --
## v bmcite
                 0.3.0
                                      v panc8
                                                   3.0.2
                                      v pancreasref 1.0.0
                 3.1.4
## v cbmc
                                      v pbmcref 1.0.0
v pbmcsca 3.0.0
               3.0.0
## v hcabm40k
## v ifnb
                 3.1.0
             2.0.0
## v lungref
                                      v thp1.eccite 3.1.5
## v mousecortexref 1.0.0
## ------ Key ------ Key ------
## v Dataset loaded successfully
## > Dataset built with a newer version of Seurat than installed
## (?) Unknown version of Seurat installed
library(dplyr)
##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
      filter, lag
##
## The following objects are masked from 'package:base':
##
##
      intersect, setdiff, setequal, union
```

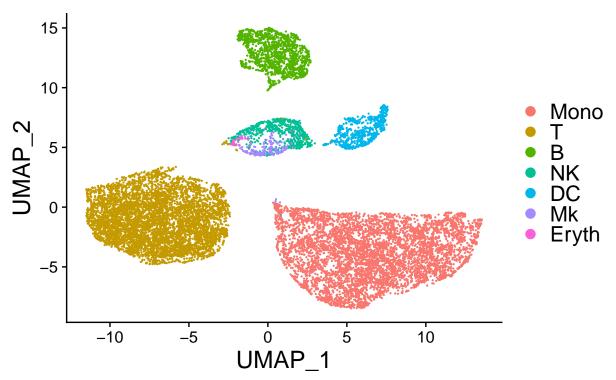
```
library(useful)
## Loading required package: ggplot2
library(data.table)
## Attaching package: 'data.table'
## The following objects are masked from 'package:dplyr':
##
       between, first, last
LoadData('ifnb')
## An object of class Seurat
## 14053 features across 13999 samples within 1 assay
## Active assay: RNA (14053 features, 0 variable features)
ifnb$cell_type <- plyr::mapvalues(ifnb$seurat_annotations,</pre>
                                    from = c('CD8 T', 'CD4 Memory T', 'T activated', 'CD4 Naive T', 'B',
                                    to = c('T', 'T', 'T', 'B', 'B', 'Mono', 'Mono', 'DC'))
ifnb$cell_subtype <- ifnb$seurat_annotations</pre>
data <- fread('ifnb_celltype_denoised_expression.txt',data.table = FALSE)</pre>
rownames(data) <- data[,1]</pre>
data <- data[,-1]</pre>
data <- t(data)</pre>
counts <- expm1(data)</pre>
recon <- CreateSeuratObject(counts, meta.data = ifnb@meta.data[colnames(data), ])</pre>
recon <- SetAssayData(recon, slot = 'data', data)</pre>
recon <- FindVariableFeatures(recon, nfeatures = 5000)</pre>
recon <- ScaleData(recon)</pre>
## Centering and scaling data matrix
recon <- RunPCA(recon, verbose=FALSE)</pre>
recon <- RunUMAP(recon, dims=1:30)</pre>
## Warning: The default method for RunUMAP has changed from calling Python UMAP via reticulate to the R
## To use Python UMAP via reticulate, set umap.method to 'umap-learn' and metric to 'correlation'
## This message will be shown once per session
## 21:29:05 UMAP embedding parameters a = 0.9922 b = 1.112
```

21:29:05 Read 13999 rows and found 30 numeric columns

```
## 21:29:05 Using Annoy for neighbor search, n_neighbors = 30
## 21:29:05 Building Annoy index with metric = cosine, n_trees = 50
## 0%
       10
           20
                         50
                                  70
                                           90
                                                100%
## [----|----|----|
## **************
## 21:29:06 Writing NN index file to temp file /tmp/RtmpVSnhj6/file255e7334adc6b
## 21:29:06 Searching Annoy index using 1 thread, search_k = 3000
## 21:29:09 Annoy recall = 100%
## 21:29:10 Commencing smooth kNN distance calibration using 1 thread with target n_neighbors = 30
## 21:29:10 Initializing from normalized Laplacian + noise (using irlba)
## 21:29:11 Commencing optimization for 200 epochs, with 495230 positive edges
## 21:29:15 Optimization finished
```

Plot cell type

Factor combination 1



Plot cell subtype

```
DimPlot(recon, reduction = 'umap', group.by = 'cell_subtype') +
    ggtitle('Factor combination 1') +
    theme(plot.title = element_text(size=24),
        axis.title = element_text(size=18),
        legend.text = element_text(size=16))
```

Factor combination 1

