Random forest

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Replicate original random forest analysis in the paper

The followings are codes modified from scripts_from_original_paper/7_Random_forest/random_forest_NEW_3.r.

Load CSV data and exclude redundant variables

```
data <- read_csv("../data/AppendixS1.csv") %>%
# Exclude low-level taxonomic variables
 select(-Species, -Genus, -Family, -Order) %>%
# Exclude redundant statistical variables
 select(-pmtr, -dgr, -dgp, -der, -dep, -ger, -gep) %>%
# Exclude variables of genetic sequence
 select(-bp, -pi) %>%
# Exclude variables generated by downstream analysis
 select(-r, -c, -t, -p,
                         -f, -fp, -cgeo, -tgeo, -pgeo, -cenv, -tenv, -penv) %>%
# Exclude rows with NAs
 na.omit()
## Rows: 19197 Columns: 67
## -- Column specification -------
## Delimiter: ","
## chr (10): Species, Gene, Genus, Family, Order, Class, Phylum, Kingdom, Metab...
## dbl (57): n, pmtr, pmtp, dgr, dgp, der, dep, ger, gep, bp, pi, Postfloodingo...
##
## i Use 'spec()' to retrieve the full column specification for this data.
## i Specify the column types or set 'show_col_types = FALSE' to quiet this message.
dim(data)
```

[1] 12252

In total there are 41 predictors, 1 response variable and 12252 data points.

Convert p-value to labels

pmtp is generated by mantel.partial(mydata.dnadist, mydata.geodist, mydata.envdist, method = "pearson", permutations = 999) from scripts_from_original_paper/8_IBE/IBE.r.

```
data_labeled <- data %>%
# Convert p-value to reject/accept null hypothesis by a threshold of 0.05
  mutate(pmtp = ifelse(pmtp <= 0.05, "yes", "no") %>% as.factor())
```

The data set is highly unbalanced because the positive cases are only 14.9118511%.

```
# Calculate percentage of yes/no
data_labeled %>%
  group_by(pmtp) %>%
  summarize(n = n(), p = n/12252)
## # A tibble: 2 x 3
    pmtp
             n
     <fct> <int> <dbl>
## 1 no
           10425 0.851
## 2 yes
           1827 0.149
```

Run random forest and imporance analysis

```
rf_unfiltered_unbalanced <- randomForest(pmtp ~ .,</pre>
                    data=data_labeled,
                    ntree=1000, importance=TRUE, nPerm=100)
rf_unfiltered_unbalanced
\#saveRDS(rf\_unfiltered\_unbalanced, "rf\_unfiltered\_unbalanced.RDS")
##
  randomForest(formula = pmtp ~ ., data = data_labeled, ntree = 1000,
                                                                                 importance = TRUE, nPerm =
                   Type of random forest: classification
##
                         Number of trees: 1000
##
## No. of variables tried at each split: 6
##
##
           OOB estimate of error rate: 14.5%
## Confusion matrix:
          no yes class.error
## no 10177 248 0.02378897
## yes 1528 299 0.83634373
If no filtering and balancing is applied, the overall accuracy is relatively high (0.1467067) but highly biased
```

(false negative rate = 0.8363437). See important variables:

```
importance(rf_unfiltered_unbalanced) %>%
 as_tibble(rownames = "var") %>%
# Sort by MDA descendantly
  arrange(desc(MeanDecreaseAccuracy)) %>%
  select(var, MeanDecreaseAccuracy)
```

```
## # A tibble: 41 x 2
##
                 MeanDecreaseAccuracy
     var
```

```
##
      <chr>
                                   <dbl>
##
    1 n
                                   100.
##
    2 abs mid lat
                                    51.9
                                    50.2
##
    3 area
##
    4 abs_min_lat
                                    48.3
    5 max lon
                                    47.0
##
##
    6 length
                                    46.7
##
    7 min_lat
                                    45.9
##
    8 min lon
                                    45.8
##
    9 abs_max_lat
                                    45.2
## 10 max_lat
                                    43.3
## # ... with 31 more rows
```

Re-run random forest using balanced data and set $n \ge 20$

Apparently n is an important predictor, but it is an artifect that has nothing to do with the scientific question I'm addressing. n is the number of individuals whose genetic sequences were used to infer genetic structure. n too low means the inference of genetic structure is not reliable. In the paper, they claimed that n>=20 is a good threshold.

```
data_labeled_20 <- data_labeled %>%
  filter(n >= 20)
dim(data_labeled_20)
```

```
## [1] 2363 42
```

1463 173

yes 507 220

0.1057457

0.6973865

After applying $n \ge 20$, there are only 2363 (0.1928665%) data points left. See how filtering by $n \ge 20$ changes accuracy:

```
##
## Call:
##
    randomForest(formula = pmtp ~ ., data = data_labeled_20, ntree = 1000,
                                                                                  importance = TRUE, nPer
                  Type of random forest: classification
##
##
                        Number of trees: 1000
## No. of variables tried at each split: 6
##
           OOB estimate of error rate: 28.78%
##
  Confusion matrix:
##
         no yes class.error
```

As shown above, the model is still predicting badly on positive cases. What's more, it's predicting worse on true negative cases and the overall error rate is even higher. It could be because of the low number of positive cases in the training data set and applying filter makes both positive cases and negative cases rarer, so the model does not have much data to train. We can (partially) solve this problem by resampling from the original data set and sample equal numbers of positive and negative cases.

```
# Sample from original data set to get balanced data set.
# By default the resampled data set will be 2 times the size of the original one.
# Return the resampled dataframe
generate_balanced_data <- function(data, N = 2*nrow(data)){
    data_pos <- data %>%
        filter(pmtp == "yes")
    data_neg <- data %>%
        filter(pmtp == "no")
    data_pos_sample <- data_pos %>%
        slice_sample(n = round(N/2), replace = TRUE)
    data_neg_sample <- data_neg %>%
        slice_sample(n = N - round(N/2), replace = TRUE)
    data_pos_sample %>%
        bind_rows(data_neg_sample)
}
```

See how balanced training data set can affect accuracy:

```
##
## Call:
   randomForest(formula = pmtp ~ ., data = generate_balanced_data(data_labeled),
                                                                                        ntree = 1000, im
##
                  Type of random forest: classification
##
                        Number of trees: 1000
## No. of variables tried at each split: 6
##
##
           OOB estimate of error rate: 1.46%
## Confusion matrix:
##
          no
               yes class.error
## no 11918
               334 0.027260855
          24 12228 0.001958864
## yes
```

As shown above, the accuracy is elevated so much and the bias is also eliminated!

Putting it together, I made a pipeline function that does balancing sampling, random forest building and importance analysis:

```
select(var, MeanDecreaseAccuracy) %>%
mutate(index = i) %>%
select(index, type = var, value = MeanDecreaseAccuracy)
error_rate %>%
bind_rows(imp)
}
```

Here I did resampling 100 times, pooled the results and only showed the mean of all metrics.

Summarizing the error rates from 100 resamplings

```
imp_filtered_balanced <- random_forests_tbl %>%
  filter(type == "yes" | type == "no") %>%
  group_by(type) %>%
  summarize(mean_error = mean(value))
imp_filtered_balanced
```

Summarizing the mean decrease accuracy of predictors from 100 resamplings

```
MDA <- random_forests_tbl %>%
  filter(type != "yes" & type != "no") %>%
  group_by(type) %>%
  summarize(meanDecreaseAccuracy = mean(value)) %>%
  arrange(desc(meanDecreaseAccuracy))
saveRDS(MDA, "MDA.RDS")
```

Comparing accuracies based on four data set:

```
as_tibble_row() %>%
              gather(key = true, value = filtered_unbalanced)
            ) %>%
  left_join(imp_filtered_balanced %>%
              select(true = type, filtered_balanced = mean_error)
## Joining, by = "true"
## Joining, by = "true"
## Joining, by = "true"
saveRDS(summary_accuracy, "summary_accuracy_randomForest.RDS")
summary_accuracy
## # A tibble: 2 x 5
##
   true unfiltered_unbalanced unfiltered_balanced filtered_unbalanced filtered~1
                           <dbl>
                                               <dbl>
                                                                   <dbl>
## 1 no
                          0.0238
                                             0.0273
                                                                   0.106
                                                                             0.0563
## 2 yes
                          0.836
                                             0.00196
                                                                   0.697
                                                                             0.0261
## # ... with abbreviated variable name 1: filtered_balanced
```

Plotting the error rate with combination of filtered and balanced

