

Package ‘SlimR’

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Title Adaptive Machine Learning-Powered, Context-Matching Tool for Single-Cell and Spatial Transcriptomics Annotation

Description Annotates single-cell and spatial-transcriptomic (ST) data using marker datasets. Supports unified markers list ('Markers_list') creation from built-in databases (e.g., 'Cellmarker2', 'PanglaoDB', 'scIBD', 'TCellSI', 'PCTIT', 'PCTAM'), Seurat objects, or user-supplied Excel files. SlimR can predict calculation parameters by adaptive machine learning algorithms, and based on Markers_list, calculate gene expression of different cell types and predict annotation information, and calculate corresponding AUC and annotate it, then verify it. At the same time, it can calculate gene expression corresponding to the cell type to generate a reference map for manual annotation (e.g., 'Heat Map', 'Feature Plots', 'Combined Plots'). For more details, see Kabacoff (2020, ISBN:9787115420572).

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URL <https://github.com/zhaqing-wang/SlimR>

BugReports <https://github.com/zhaqing-wang/SlimR/issues>

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Contents

calculate_cluster_variability	2
calculate_expression	3
calculate_expression_skewness	4
calculate_probability	4
Cellmarker2	5
Cellmarker2_raw	6
Cellmarker2_table	7
Celltype_Annotation	7
Celltype_annotation_Cellmarker2	8
Celltype_Annotation_Combined	10
Celltype_annotation_Excel	11
Celltype_Annotation_Features	13
Celltype_Annotation_Heatmap	15
Celltype_annotation_PanglaoDB	17
Celltype_annotation_Seurat	19
Celltype_Calculate	20
Celltype_Verification	23
compute_adaptive_parameters	24
estimate_batch_effect	25
extract_dataset_features	25
Markers_filter_Cellmarker2	26
Markers_filter_PanglaoDB	27
Markers_list_PCTAM	28
Markers_list_PCTIT	29
Markers_list_scIBD	30
Markers_list_TCellSI	30
PanglaoDB	31
PanglaoDB_raw	32
PanglaoDB_table	32
Parameter_Calculate	33
Read_excel_markers	34
Read_seurat_markers	35

Index

37

calculate_cluster_variability
Calculate Cluster Variability (Use in package)

Description

Measures the degree of separation between different cell clusters based on expression patterns.

Usage

```
calculate_cluster_variability(data.features, features)
```

Arguments

data.features	Data frame containing expression data and cluster labels
features	Feature names to include in analysis

Value

Numeric value representing cluster separation strength

See Also

Other Section_1_Functions_Use_in_Package: [calculate_expression\(\)](#), [calculate_expression_skewness\(\)](#), [calculate_probability\(\)](#), [compute_adaptive_parameters\(\)](#), [estimate_batch_effect\(\)](#), [extract_dataset_features\(\)](#)

calculate_expression *Counts average expression of gene set (Use in package)*

Description

Counts average expression of gene set (Use in package)

Usage

```
calculate_expression(  
  object,  
  features,  
  assay = NULL,  
  cluster_col = NULL,  
  colour_low = "white",  
  colour_high = "navy"  
)
```

Arguments

object	Enter a Seurat object.
features	Enter one or a set of markers.
assay	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = NULL".
cluster_col	Enter the meta.data column in the Seurat object to be annotated, such as "seurat_cluster". Default parameters use "cluster_col = NULL".
colour_low	Color for lowest expression level. (default = "white")
colour_high	Color for highest expression level. (default = "black")

Value

Average expression genes and related informations in the input "Seurat" object given "cluster_col" and given "features".

See Also

Other Section_1_Functions_Use_in_Package: [calculate_cluster_variability\(\)](#), [calculate_expression_skewness\(\)](#), [calculate_probability\(\)](#), [compute_adaptive_parameters\(\)](#), [estimate_batch_effect\(\)](#), [extract_dataset_features\(\)](#)

calculate_expression_skewness

Calculate Expression Distribution Skewness (Use in package)

Description

Computes the average skewness of gene expression distributions across all features.

Usage

```
calculate_expression_skewness(expression_matrix)
```

Arguments

expression_matrix	Matrix of expression values
-------------------	-----------------------------

Value

Mean absolute skewness across all genes

See Also

Other Section_1_Functions_Use_in_Package: [calculate_cluster_variability\(\)](#), [calculate_expression\(\)](#), [calculate_probability\(\)](#), [compute_adaptive_parameters\(\)](#), [estimate_batch_effect\(\)](#), [extract_dataset_features\(\)](#)

calculate_probability *Calculate gene set expression and infer probabilities with control datasets (Use in package)*

Description

Calculate gene set expression and infer probabilities with control datasets (Use in package)

Usage

```
calculate_probability(
  object,
  features,
  assay = NULL,
  cluster_col = NULL,
  min_expression = 0.1,
  specificity_weight = 3
)
```

Arguments

object	Enter a Seurat object.
features	Enter one or a set of markers.
assay	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = NULL".
cluster_col	Enter the meta.data column in the Seurat object to be annotated, such as "seurat_cluster". Default parameters use "cluster_col = NULL".
min_expression	The min_expression parameter defines a threshold value to determine whether a cell's expression of a feature is considered "expressed" or not. It is used to filter out low-expression cells that may contribute noise to the analysis. Default parameters use "min_expression = 0.1".
specificity_weight	The specificity_weight parameter controls how much the expression variability (standard deviation) of a feature within a cluster contributes to its "specificity score." It amplifies or suppresses the impact of variability in the final score calculation. Default parameters use "specificity_weight = 3".

Value

Average expression of genes in the input "Seurat" object given "cluster_col" and given "features".

See Also

Other Section_1_Functions_Use_in_Package: [calculate_cluster_variability\(\)](#), [calculate_expression\(\)](#), [calculate_expression_skewness\(\)](#), [compute_adaptive_parameters\(\)](#), [estimate_batch_effect\(\)](#), [extract_dataset_features\(\)](#)

Cellmarker2

Cellmarker2 dataset

Description

A dataset containing marker genes for different cell types from Cellmarker2

Usage

Cellmarker2

Format

A data frame with 8 columns:

Details

This dataset is used to filter and create a standardized marker list. The dataset can be filtered based on species, tissue class, tissue type, cancer type, and cell type to generate a list of marker genes for specific cell types.

Source

<http://117.50.127.228/CellMarker/>

See Also

Other Section_0_Database: [Cellmarker2_raw](#), [Cellmarker2_table](#), [Markers_list_PCTAM](#), [Markers_list_PCTIT](#), [Markers_list_TCellSI](#), [Markers_list_scIBD](#), [PanglaoDB](#), [PanglaoDB_raw](#), [PanglaoDB_table](#)

Cellmarker2_raw

Cellmarker2 raw dataset

Description

A dataset containing marker genes for different cell types from Cellmarker2

Usage

`Cellmarker2_raw`

Format

A data frame with 20 columns contained in the Cellmarker2 database:

Details

This dataset is used to filter and create a standardized marker list. The dataset can be filtered based on species, tissue class, tissue type, cancer type, and cell type to generate a list of marker genes for specific cell types.

Source

<http://117.50.127.228/CellMarker/>

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_table](#), [Markers_list_PCTAM](#), [Markers_list_PCTIT](#), [Markers_list_TCellSI](#), [Markers_list_scIBD](#), [PanglaoDB](#), [PanglaoDB_raw](#), [PanglaoDB_table](#)

Cellmarker2_table *Cellmarker2 table*

Description

A dataset containing marker genes for different cell types from Cellmarker2

Usage

```
Cellmarker2_table
```

Format

A list contain different types like species, tissue_class, tissue_type, cancer_type, cell_type

Details

This list is used to choose filters for creation of standardized marker list.

Source

<http://117.50.127.228/CellMarker/>

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_raw](#), [Markers_list_PCTAM](#), [Markers_list_PCTIT](#),
[Markers_list_TCellSI](#), [Markers_list_scIBD](#), [PanglaoDB](#), [PanglaoDB_raw](#), [PanglaoDB_table](#)

Celltype_Annotation *Annotate Seurat Object with SlimR Cell Type Predictions*

Description

This function assigns SlimR predicted cell types to a Seurat object based on cluster annotations, and stores the results in the meta.data slot.

Usage

```
Celltype_Annotation(  
  seurat_obj,  
  cluster_col,  
  SlimR_anno_result,  
  plot_UMAP = TRUE,  
  annotation_col = "Cell_type_SlimR"  
)
```

Arguments

seurat_obj	A Seurat object containing cluster information in meta.data.
cluster_col	Character string indicating the column name in meta.data that contains cluster IDs.
SlimR_anno_result	List generated by function Celltype_Calculate() which containing a data.frame in \$Prediction_results with: 1.cluster_col (Cluster identifiers (should match cluster_col in meta.data)) 2.Predicted_cell_type (Predicted cell types for each cluster).
plot_UMAP	logical(1); if TRUE, plot the UMAP with cell type annotations.
annotation_col	The location to write in 'meta.data' that contains the predicted cell type. (default = "Cell_type_SlimR")

Value

A Seurat object with updated meta.data containing the predicted cell types.

Note

If plot_UMAP = TRUE, this function will print a UMAP plot as a side effect.

See Also

Other Section_3_Automated_Annotation: [Celltype_Calculate\(\)](#), [Celltype_Verification\(\)](#), [Parameter_Calculate\(\)](#)

Examples

```
## Not run:
sce <- Celltype.Annotation(seurat_obj = sce,
  cluster_col = "seurat_clusters",
  SlimR_anno_result = SlimR_anno_result,
  plot_UMAP = TRUE,
  annotation_col = "Cell_type_SlimR"
)

## End(Not run)
```

Celltype_annotation_Cellmarker2

Uses "marker_list" from Cellmarker2 for cell annotation

Description

Uses "marker_list" from Cellmarker2 for cell annotation

Usage

```
Celltype_annotation_Cellmarker2(
  seurat_obj,
  gene_list,
  species,
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = NULL,
  min_counts = 1,
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy"
)
```

Arguments

seurat_obj	Enter the Seurat object with annotation columns such as "seurat_cluster" in meta.data to be annotated.
gene_list	Enter the standard "Marker_list" generated by the Cellmarker2 database for the SlimR package, generated by the "Markers_filter_Cellmarker2 ()" function.
species	This parameter selects the species "Human" or "Mouse" for standard gene format correction of markers entered by "Marker_list".
cluster_col	Enter annotation columns such as "seurat_cluster" in meta.data of the Seurat object to be annotated. Default parameters use "cluster_col = 'seurat_clusters'".
assay	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = "RNA"".
save_path	The output path of the cell annotation picture. Example parameters use "save_path = './SlimR/Celltype_annotation_Cellmarker2/'".
min_counts	The minimum number of counts of genes in "Marker_list" entered. This number represents the number of the same gene in the same species and the same location in the Cellmarker2 database used for annotation of this cell type. Default parameters use "min_counts = 1".
colour_low	Color for lowest expression level. (default = "white")
colour_high	Color for highest expression level. (default = "navy")
colour_low_mertic	Color for lowest mertic level. (default = "white")
colour_high_mertic	Color for highest mertic level. (default = "navy")

Value

The cell annotation picture is saved in "save_path".

See Also

Other Section_5_Other_Functions_Provided: [Celltype_annotation_Excel\(\)](#), [Celltype_annotation_PanglaoDB\(\)](#), [Celltype_annotation_Seurat\(\)](#)

Examples

```
## Not run:
Celltype_annotation_Cellmarker2(seurat_obj = sce,
  gene_list = Markers_list_Cellmarker2,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_Cellmarker2")
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy",
  )

## End(Not run)
```

Celltype_Annotation_Combined

Uses "marker_list" to generate combined plot for cell annotation

Description

Uses "marker_list" to generate combined plot for cell annotation

Usage

```
Celltype_Annotation_Combined(
  seurat_obj,
  gene_list,
  species,
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = NULL,
  colour_low = "white",
  colour_high = "navy"
)
```

Arguments

seurat_obj	Enter the Seurat object with annotation columns such as "seurat_cluster" in meta.data to be annotated.
------------	--

gene_list	A list of cells and corresponding gene controls, the name of the list is cell type, and the first column of the list corresponds to markers. Lists can be generated using functions such as "Markers_filter_Cellmarker2()", "Markers_filter_PanglaoDB()", "read_excel_markers()", "read_seurat_markers()", etc.
species	This parameter selects the species "Human" or "Mouse" for standard gene format correction of markers entered by "Marker_list".
cluster_col	Enter annotation columns such as "seurat_cluster" in meta.data of the Seurat object to be annotated. Default parameters use "cluster_col = 'seurat_clusters'".
assay	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = 'RNA'".
save_path	The output path of the cell annotation picture. Example parameters use "save_path = './SlimR/Celltype_annotation_Bar/'".
colour_low	Color for lowest expression level. (default = "white")
colour_high	Color for highest expression level. (default = "navy")

Value

The cell annotation picture is saved in "save_path".

See Also

Other Section_4_Semi_Automated_Annotation: [Celltype_annotation_Features\(\)](#), [Celltype_annotation_Heatmap\(\)](#)

Examples

```
## Not run:
Celltype_annotation_Combined(seurat_obj = sce,
  gene_list = Markers_list,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_Combined"),
  colour_low = "white",
  colour_high = "navy"
)

## End(Not run)
```

Celltype_annotation_Excel

Uses "marker_list" from Excel input for cell annotation

Description

Uses "marker_list" from Excel input for cell annotation

Usage

```
Celltype_annotation_Excel(
  seurat_obj,
  gene_list,
  species,
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = NULL,
  metric_names = NULL,
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy"
)
```

Arguments

seurat_obj	Enter the Seurat object with annotation columns such as "seurat_cluster" in meta.data to be annotated.
gene_list	Enter the standard "Marker_list" generated by the Excel files database for the SlimR package, generated by the "read_excel_markers()" function.
species	This parameter selects the species "Human" or "Mouse" for standard gene format correction of markers entered by "Marker_list".
cluster_col	Enter annotation columns such as "seurat_cluster" in meta.data of the Seurat object to be annotated. Default parameters use "cluster_col = "seurat_clusters"".
assay	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = 'RNA'".
save_path	The output path of the cell annotation picture. Example parameters use "save_path = './SlimR/Celltype_annotation_Excel/'".
metric_names	Change the row name for the input mertics, not recommended unless necessary. (NULL is used as default parameter)
colour_low	Color for lowest expression level. (default = "white")
colour_high	Color for highest expression level. (default = "navy")
colour_low_mertic	Color for lowest mertic level. (default = "white")
colour_high_mertic	Color for highest mertic level. (default = "navy")

Value

The cell annotation picture is saved in "save_path".

See Also

Other Section_5_Other_Functions_Provided: [Celltype_annotation_Cellmarker2\(\)](#), [Celltype_annotation_PanglaoDB\(\)](#), [Celltype_annotation_Seurat\(\)](#)

Examples

```
## Not run:
Celltype_annotation_Excel(seurat_obj = sce,
  gene_list = Markers_list_Excel,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_Excel")
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy",
)
## End(Not run)
```

Celltype_annotation_Features

Annotate cell types using features plot with different marker databases

Description

This function dynamically selects the appropriate annotation method based on the `gene_list_type` parameter. It supports marker databases from Cellmarker2, PanglaoDB, Seurat (via `FindAllMarkers`), or Excel files.

Usage

```
Celltype_annotation_Features(
  seurat_obj,
  gene_list,
  gene_list_type = "Default",
  species = NULL,
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = NULL,
  min_counts = 1,
  metric_names = NULL,
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy",
  ...
)
```

Arguments

seurat_obj	A valid Seurat object with cluster annotations in <code>meta.data</code> .
gene_list	A list of data frames containing marker genes and metrics. Format depends on <code>gene_list_type</code> : - Cellmarker2 : Generated by <code>Markers_filter_Cellmarker2()</code> . - PanglaoDB : Generated by <code>Markers_filter_PanglaoDB()</code> . - Seurat : Generated by <code>read_seurat_markers()</code> . - Excel : Generated by <code>read_excel_markers()</code> .
gene_list_type	Type of marker database to use. Be one of: "Cellmarker2", "PanglaoDB", "Seurat", or "Excel".
species	Species of the dataset: "Human" or "Mouse" for gene name standardization.
cluster_col	Column name in <code>meta.data</code> defining clusters (default: "seurat_clusters").
assay	Assay layer in the Seurat object (default: "RNA").
save_path	Directory to save output PNGs. Must be explicitly specified.
min_counts	Minimum number of counts for Cellmarker2 annotations (default: 1).
metric_names	Optional. Change the row name for the input metrics, not recommended unless necessary. (NULL is used as default parameter; used in "Seurat"/"Excel").
colour_low	Color for lowest expression level. (default = "white")
colour_high	Color for highest expression level. (default = "navy")
colour_low_metric	Color for lowest metric level. (default = "white")
colour_high_metric	Color for highest metric level. (default = "navy")
...	Additional parameters passed to the specific annotation function.

Value

Saves cell type annotation PNGs in `save_path`. Returns invisibly.

See Also

Other Section_4_Semi_Automated_Annotation: [Celltype_Annotation_Combined\(\)](#), [Celltype_Annotation_Heatmap\(\)](#)

Examples

```
## Not run:
# Example for Cellmarker2
Celltype_Annotation_Features(seurat_obj = sce,
  gene_list = Markers_list_Cellmarker2,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_Cellmarker2"),
  colour_low = "white",
  colour_high = "navy",
  colour_low_metric = "white",
  colour_high_metric = "navy",
)
```

```
# Example for PanglaoDB
Celltype.Annotation.Features(seurat_obj = sce,
  gene_list = Markers_list_panglaoDB,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_PanglaoDB")
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy",
  )

# Example for Seurat marker list
Celltype.Annotation.Features(seurat_obj = sce,
  gene_list = Markers_list_Seurat,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_Seurat")
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy",
  )

# Example for Excel marker list
Celltype.Annotation.Features(seurat_obj = sce,
  gene_list = Markers_list_Excel,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_Excel")
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy",
  )

## End(Not run)
```

Celltype_Annotation_Heatmap

Uses "marker_list" to generate heatmap for cell annotation

Description

Uses "marker_list" to generate heatmap for cell annotation

Usage

```
Celltype_Annotation_Heatmap(
  seurat_obj,
  gene_list,
  species,
  cluster_col = "seurat_clusters",
  assay = "RNA",
  min_expression = 0.1,
  specificity_weight = 3,
  colour_low = "navy",
  colour_high = "firebrick3"
)
```

Arguments

<code>seurat_obj</code>	Enter the Seurat object with annotation columns such as "seurat_cluster" in meta.data to be annotated.
<code>gene_list</code>	A list of cells and corresponding gene controls, the name of the list is cell type, and the first column of the list corresponds to markers. Lists can be generated using functions such as "Markers_filter_Cellmarker2()", "Markers_filter_PanglaoDB()", "read_excel_markers()", "read_seurat_markers()", etc.
<code>species</code>	This parameter selects the species "Human" or "Mouse" for standard gene format correction of markers entered by "Marker_list".
<code>cluster_col</code>	Enter annotation columns such as "seurat_cluster" in meta.data of the Seurat object to be annotated. Default parameters use "cluster_col = 'seurat_clusters'".
<code>assay</code>	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = 'RNA'".
<code>min_expression</code>	The <code>min_expression</code> parameter defines a threshold value to determine whether a cell's expression of a feature is considered "expressed" or not. It is used to filter out low-expression cells that may contribute noise to the analysis. Default parameters use "min_expression = 0.1".
<code>specificity_weight</code>	The <code>specificity_weight</code> parameter controls how much the expression variability (standard deviation) of a feature within a cluster contributes to its "specificity score." It amplifies or suppresses the impact of variability in the final score calculation. Default parameters use "specificity_weight = 3".
<code>colour_low</code>	Color for lowest probability level in Heatmap visualization of probability matrix. (default = "navy")
<code>colour_high</code>	Color for highest probability level Heatmap visualization of probability matrix. (default = "firebrick3")

Value

The heatmap of the comparison between "cluster_col" in the Seurat object and the given gene set "gene_list" needs to be annotated.

See Also

Other Section_4_Semi_Automated_Annotation: [Celltype.Annotation_Combined\(\)](#), [Celltype.Annotation_Features\(\)](#)

Examples

```
## Not run:
Celltype.Annotation_Heatmap(seurat_obj = sce,
  gene_list = Markers_list,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  min_expression = 0.1,
  specificity_weight = 3,
  colour_low = "navy",
  colour_high = "firebrick3"
)

## End(Not run)
```

Celltype_annotation_PanglaoDB

Uses "marker_list" from PanglaoDB for cell annotation

Description

Uses "marker_list" from PanglaoDB for cell annotation

Usage

```
Celltype_annotation_PanglaoDB(
  seurat_obj,
  gene_list,
  species,
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = NULL,
  metric_names = NULL,
  colour_low = "white",
  colour_high = "navy",
  colour_low_metric = "white",
  colour_high_metric = "navy"
)
```

Arguments

<code>seurat_obj</code>	Enter the Seurat object with annotation columns such as "seurat_cluster" in meta.data to be annotated.
<code>gene_list</code>	Enter the standard "Marker_list" generated by the PanglaoDB database for the SlimR package, generated by the "Markers_filter_PanglaoDB ()" function.
<code>species</code>	This parameter selects the species "Human" or "Mouse" for standard gene format correction of markers entered by "Marker_list".
<code>cluster_col</code>	Enter annotation columns such as "seurat_cluster" in meta.data of the Seurat object to be annotated. Default parameters use "cluster_col = 'seurat_clusters'".
<code>assay</code>	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = 'RNA'".
<code>save_path</code>	The output path of the cell annotation picture. Example parameters use "save_path = './SlimR/Celltype_annotation_PanglaoDB/'".
<code>metric_names</code>	Warning: Do not enter information. This parameter is used to check if "Marker_list" conforms to the PanglaoDB database output.
<code>colour_low</code>	Color for lowest expression level. (default = "white")
<code>colour_high</code>	Color for highest expression level. (default = "navy")
<code>colour_low_mertic</code>	Color for lowest mertic level. (default = "white")
<code>colour_high_mertic</code>	Color for highest mertic level. (default = "navy")

Value

The cell annotation picture is saved in "save_path".

See Also

Other Section_5_Other_Functions_Provided: [Celltype_annotation_Cellmarker2\(\)](#), [Celltype_annotation_Excel\(\)](#), [Celltype_annotation_Seurat\(\)](#)

Examples

```
## Not run:
Celltype_annotation_PanglaoDB(seurat_obj = sce,
  gene_list = Markers_list_panglaoDB,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_PanglaoDB")
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy",
  )

## End(Not run)
```

Celltype_annotation_Seurat

Uses "marker_list" from Seurat object for cell annotation

Description

Uses "marker_list" from Seurat object for cell annotation

Usage

```
Celltype_annotation_Seurat(
  seurat_obj,
  gene_list,
  species,
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = NULL,
  metric_names = NULL,
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy"
)
```

Arguments

seurat_obj	Enter the Seurat object with annotation columns such as "seurat_cluster" in meta.data to be annotated.
gene_list	Enter the standard "Marker_list" generated by the Seurat object database for the SlimR package, generated by the "read_seurat_markers()" function.
species	This parameter selects the species "Human" or "Mouse" for standard gene format correction of markers entered by "Marker_list".
cluster_col	Enter annotation columns such as "seurat_cluster" in meta.data of the Seurat object to be annotated. Default parameters use "cluster_col = 'seurat_clusters'".
assay	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = 'RNA'".
save_path	The output path of the cell annotation picture. Example parameters use "save_path = './SlimR/Celltype_annotation_Seurat/'".
metric_names	Change the row name for the input mertics, not recommended unless necessary. (NULL is used as default parameter)
colour_low	Color for lowest expression level. (default = "white")
colour_high	Color for highest expression level. (default = "navy")
colour_low_mertic	Color for lowest mertic level. (default = "white")
colour_high_mertic	Color for highest mertic level. (default = "navy")

Value

The cell annotation picture is saved in "save_path".

See Also

Other Section_5_Other_Functions_Provided: [Celltype_annotation_Cellmarker2\(\)](#), [Celltype_annotation_Excel\(\)](#), [Celltype_annotation_PanglaoDB\(\)](#)

Examples

```
## Not run:
Celltype_annotation_Seurat(seurat_obj = sce,
  gene_list = Markers_list_Seurat,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  save_path = file.path(tempdir(),"SlimR_Celltype_annotation_Seurat")
  colour_low = "white",
  colour_high = "navy",
  colour_low_mertic = "white",
  colour_high_mertic = "navy",
  )
## End(Not run)
```

<i>Celltype_Calculate</i>	<i>Uses "marker_list" to calculate probability, prediction results, AUC and generate heatmap for cell annotation</i>
---------------------------	--

Description

Uses "marker_list" to calculate probability, prediction results, AUC and generate heatmap for cell annotation

Usage

```
Celltype_Calculate(
  seurat_obj,
  gene_list,
  species,
  cluster_col = "seurat_clusters",
  assay = "RNA",
  min_expression = 0.1,
  specificity_weight = 3,
  threshold = 0.6,
  compute_AUC = TRUE,
  plot_AUC = TRUE,
```

```

    AUC_correction = TRUE,
    colour_low = "navy",
    colour_high = "firebrick3"
)

```

Arguments

<code>seurat_obj</code>	Enter the Seurat object with annotation columns such as "seurat_cluster" in meta.data to be annotated.
<code>gene_list</code>	A list of cells and corresponding gene controls, the name of the list is cell type, and the first column of the list corresponds to markers. Lists can be generated using functions such as "Markers_filter_Cellmarker2()", "Markers_filter_PanglaoDB()", "read_excel_markers()", "read_seurat_markers()", etc.
<code>species</code>	This parameter selects the species "Human" or "Mouse" for standard gene format correction of markers entered by "Marker_list".
<code>cluster_col</code>	Enter annotation columns such as "seurat_cluster" in meta.data of the Seurat object to be annotated. Default parameters use "cluster_col = 'seurat_clusters'".
<code>assay</code>	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = 'RNA'".
<code>min_expression</code>	The <code>min_expression</code> parameter defines a threshold value to determine whether a cell's expression of a feature is considered "expressed" or not. It is used to filter out low-expression cells that may contribute noise to the analysis. Default parameters use "min_expression = 0.1".
<code>specificity_weight</code>	The <code>specificity_weight</code> parameter controls how much the expression variability (standard deviation) of a feature within a cluster contributes to its "specificity score." It amplifies or suppresses the impact of variability in the final score calculation. Default parameters use "specificity_weight = 3".
<code>threshold</code>	This parameter refers to the normalized similarity between the "alternative cell type" and the "predicted cell type" in the returned results. (the default parameter is 0.6)
<code>compute_AUC</code>	Logical indicating whether to calculate AUC values for predicted cell types. AUC measures how well the marker genes distinguish the cluster from others. When TRUE, adds an AUC column to the prediction results. (default: TRUE)
<code>plot_AUC</code>	The logic indicates whether to draw an AUC curve for the predicted cell type. When TRUE, add an AUC_plot to result. (default: TRUE)
<code>AUC_correction</code>	Logical value controlling AUC-based correction. (default = TRUE) When set to TRUE: 1.Computes AUC values for candidate cell types. (probability > threshold) 2.Selects the cell type with the highest AUC as the final predicted type. 3.Records the selected type's AUC value in the "AUC" column.
<code>colour_low</code>	Color for lowest probability level in Heatmap visualization of probability matrix. (default = "navy")
<code>colour_high</code>	Color for highest probability level Heatmap visualization of probability matrix. (default = "firebrick3")

Value

A list containing:

- Expression_list: List of expression matrices for each cell type
- Proportion_list: List of proportion of expression for each cell type
- Expression_scores_matrix: Matrix of expression scores
- Probability_matrix: Matrix of normalized probabilities
- Prediction_results: Data frame with cluster annotations including:
 - cluster_col: Cluster identifier
 - Predicted_cell_type: Primary predicted cell type
 - AUC: Area Under the Curve value (when compute_AUC = TRUE)
 - Alternative_cell_types: Semi-colon separated alternative cell types
- Heatmap_plot: Heatmap visualization of probability matrix
- AUC_plot: AUC visualization of Predicted cell type
- AUC_list: The resulting list of AUC values calculated for genes in alternative cell types above the approximate threshold

See Also

Other Section_3_Automated_Annotation: [Celltype_annotation\(\)](#), [Celltype_Verification\(\)](#), [Parameter_Calculate\(\)](#)

Examples

```
## Not run:
SlimR_anno_result <- Celltype_Calculate(seurat_obj = sce,
  gene_list = Markers_list,
  species = "Human",
  cluster_col = "seurat_clusters",
  assay = "RNA",
  min_expression = 0.1,
  specificity_weight = 3,
  threshold = 0.6,
  compute_AUC = TRUE,
  plot_AUC = TRUE,
  AUC_correction = TRUE,
  colour_low = "navy",
  colour_high = "firebrick3"
)

## End(Not run)
```

Celltype_Verification *Perform cell type verification and generate the validation dotplot*

Description

This function performs verification of predicted cell types by selecting high log2FC and high expression proportion genes and generates and generate the validation dotplot.

Usage

```
Celltype_Verification(  
  seurat_obj,  
  SlimR_anno_result,  
  assay = "RNA",  
  gene_number = 5,  
  colour_low = "white",  
  colour_high = "navy",  
  annotation_col = "Cell_type_SlimR"  
)
```

Arguments

seurat_obj	A Seurat object containing single-cell data.
SlimR_anno_result	A list containing SlimR annotation results with: Expression_list - List of expression matrices for each cell type. Prediction_results - Data frame with cluster annotations.
assay	Enter the assay used by the Seurat object, such as "RNA". Default parameters use "assay = 'RNA'".
gene_number	Integer specifying number of top genes to select per cell type.
colour_low	Color for lowest expression level. (default = "white")
colour_high	Color for highest expression level. (default = "navy")
annotation_col	Character string specifying the column in meta.data to use for grouping.

Value

A ggplot object showing expression of top variable genes.

See Also

Other Section_3_Automated_Annotation: [Celltype_annotation\(\)](#), [Celltype_Calculate\(\)](#), [Parameter_Calculate\(\)](#)

Examples

```
## Not run:
Celltype_Verification(seurat_obj = sce,
  SlimR_anno_result = SlimR_anno_result,
  assay = "RNA",
  gene_number = 5,
  colour_low = "white",
  colour_high = "navy",
  annotation_col = "Cell_type_SlimR"
)

## End(Not run)
```

compute_adaptive_parameters

Compute Adaptive Parameters Based on Dataset Features (Use in package)

Description

Calculates optimal min_expression, specificity_weight, and threshold parameters using continuous adaptive algorithms based on dataset characteristics.

Usage

```
compute_adaptive_parameters(dataset_features, n_celltypes = 50)
```

Arguments

dataset_features	List of dataset characteristics from extract_dataset_features()
n_celltypes	Expected number of cell types in marker database

Value

List containing min_expression, specificity_weight, threshold, and rationale

See Also

Other Section_1_Functions_Use_in_Package: [calculate_cluster_variability\(\)](#), [calculate_expression\(\)](#), [calculate_expression_skewness\(\)](#), [calculate_probability\(\)](#), [estimate_batch_effect\(\)](#), [extract_dataset_features\(\)](#)

estimate_batch_effect *Estimate Batch Effect Strength (Use in package)*

Description

Roughly estimates the potential impact of batch effects using available metadata.

Usage

```
estimate_batch_effect(seurat_obj, assay)
```

Arguments

seurat_obj	Seurat object
assay	Assay name

Value

Batch effect score (0 indicates no detectable batch effect)

See Also

Other Section_1_Functions_Use_in_Package: [calculate_cluster_variability\(\)](#), [calculate_expression\(\)](#), [calculate_expression_skewness\(\)](#), [calculate_probability\(\)](#), [compute_adaptive_parameters\(\)](#), [extract_dataset_features\(\)](#)

extract_dataset_features

*Extract Dataset Characteristics for Adaptive Parameter Calculation
(Use in package)*

Description

Computes various statistical features from single-cell data that are used as input for the parameter prediction model.

Usage

```
extract_dataset_features(  
  seurat_obj,  
  features,  
  assay = NULL,  
  cluster_col = NULL  
)
```

Arguments

<code>seurat_obj</code>	Seurat object
<code>features</code>	Features to analyze
<code>assay</code>	Assay name
<code>cluster_col</code>	Cluster column name

Value

List of dataset characteristics including expression statistics, variability measures, and cluster properties

See Also

Other Section_1_Functions_Use_in_Package: [calculate_cluster_variability\(\)](#), [calculate_expression\(\)](#), [calculate_expression_skewness\(\)](#), [calculate_probability\(\)](#), [compute_adaptive_parameters\(\)](#), [estimate_batch_effect\(\)](#)

Markers_filter_Cellmarker2

Create Marker_list from the Cellmarkers2 database

Description

Create Marker_list from the Cellmarkers2 database

Usage

```
Markers_filter_Cellmarker2(
  df,
  species = NULL,
  tissue_class = NULL,
  tissue_type = NULL,
  cancer_type = NULL,
  cell_type = NULL
)
```

Arguments

<code>df</code>	Standardized Cellmarkers2 database. It is read as data(Cellmarkers2) in the SlimR library.
<code>species</code>	Species information in Cellmarkers2 database. The default input is "Human" or "Mouse".The input can be retrieved by "Cellmarkers2_table". For more information,please refer to http://117.50.127.228/CellMarker/ on Cellmarkers2's official website.

tissue_class	Tissue_class information in Cellmarkers2 database. The input can be retrieved by "Cellmarkers2_table". For more information, please refer to http://117.50.127.228/CellMarker/ on Cellmarkers2's official website.
tissue_type	Tissue_type information in Cellmarkers2 database. The input can be retrieved by "Cellmarkers2_table". For more information, please refer to http://117.50.127.228/CellMarker/ on Cellmarkers2's official website.
cancer_type	Cancer_type information in Cellmarkers2 database. The input can be retrieved by "Cellmarkers2_table". For more information, please refer to http://117.50.127.228/CellMarker/ on Cellmarkers2's official website.
cell_type	Cell_type information in Cellmarkers2 database. The input can be retrieved by "Cellmarkers2_table". For more information, please refer to http://117.50.127.228/CellMarker/ on Cellmarkers2's official website.

Value

The standardized "Marker_list" in the SlimR package

See Also

Other Section_2_Standardized_Markers_List: [Markers_filter_PanglaoDB\(\)](#), [Read_excel_markers\(\)](#), [Read_seurat_markers\(\)](#)

Examples

```
Cellmarker2 <- SlimR::Cellmarker2
Markers_list_Cellmarker2 <- Markers_filter_Cellmarker2(
  Cellmarker2,
  species = "Human",
  tissue_class = "Intestine",
  tissue_type = NULL,
  cancer_type = NULL,
  cell_type = NULL
)
```

Markers_filter_PanglaoDB

Create Marker_list from the PanglaoDB database

Description

Create Marker_list from the PanglaoDB database

Usage

```
Markers_filter_PanglaoDB(df, species_input, organ_input)
```

Arguments

df	Standardized PanglaoDB database. It is read as data(PanglaoDB) in the SlimR library.
species_input	Species information in PanglaoDB database. The default input is "Human" or "Mouse".The input can be retrieved by "PanglaoDB_table". For more information,please refer to https://panglaodb.se/ on PanglaoDB's official website.
organ_input	Organ type information in the PanglaoDB database. The input can be retrieved by "PanglaoDB_table".For more information, please refer to https://panglaodb.se/ on PanglaoDB's official website.

Value

The standardized "Marker_list" in the SlimR package

See Also

Other Section_2_Standardized_Markers_List: [Markers_filter_Cellmarker2\(\)](#), [Read_excel_markers\(\)](#), [Read_seurat_markers\(\)](#)

Examples

```
PanglaoDB <- SlimR::PanglaoDB
Markers_list_panglaoDB <- Markers_filter_PanglaoDB(
  PanglaoDB,
  species_input = 'Human',
  organ_input = 'GI tract'
)
```

Markers_list_PCTAM	<i>List of Macrophage subtype markers in the article "Macrophage diversity in cancer revisited in the era of single-cell omics"</i>
---------------------------	---

Description

A dataset containing marker genes for different Macrophage subtypes from the article "Macrophage diversity in cancer revisited in the era of single-cell omics"

Usage

`Markers_list_PCTAM`

Format

A list with 7 tables.

Details

This list is a table of 7 types of Tumor-associated macrophages (TAMs) markers obtained from the article "Macrophage diversity in cancer revisited in the era of single-cell omics". The data source is "<https://doi.org/10.1016/j.it.2022.04.008>", and the reference literature is: Ruo-Yu Ma et al. (2022) [doi:10.1016/j.it.2022.04.008](https://doi.org/10.1016/j.it.2022.04.008).

Source

[doi:10.1016/j.it.2022.04.008](https://doi.org/10.1016/j.it.2022.04.008)

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_raw](#), [Cellmarker2_table](#), [Markers_list_PCTIT](#), [Markers_list_TCellSI](#), [Markers_list_scIBD](#), [PanglaoDB](#), [PanglaoDB_raw](#), [PanglaoDB_table](#)

Markers_list_PCTIT

List of T cell subtype markers in the article "Pan-cancer single cell landscape of tumor-infiltrating T cells"

Description

A dataset containing marker genes for different T cell types from the article "Pan-cancer single cell landscape of tumor-infiltrating T cells"

Usage

`Markers_list_PCTIT`

Format

A list with 40 tables.

Details

This list is a table of 40 types of pan-cancer tumor-infiltrating T cell (PCTIT) markers obtained from the article "Pan-cancer single cell landscapeof tumor-infiltrating T cells". The data source is "<https://doi.org/10.1126/science.abe6474>", and the reference literature is: L. Zheng et al. (2021) [doi:10.1126/science.abe6474](https://doi.org/10.1126/science.abe6474).

Source

[doi:10.1126/science.abe6474](https://doi.org/10.1126/science.abe6474)

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_raw](#), [Cellmarker2_table](#), [Markers_list_PCTAM](#), [Markers_list_TCellSI](#), [Markers_list_scIBD](#), [PanglaoDB](#), [PanglaoDB_raw](#), [PanglaoDB_table](#)

Markers_list_scIBD *List of cell type markers in the article scIBD*

Description

A dataset containing marker genes for different human intestine cell types from scIBD

Usage

`Markers_list_scIBD`

Format

A list with one hundred and one tables.

Details

This list is a table of 101 types of human intestine cell types markers obtained from scIBD. The article doi source is "https://doi.org/10.1038/s43588-023-00464-9", and the reference literature is: Nie et al. (2023) [doi:10.1038/s43588-023-00464-9](#). Note: The 'Markers_list_scIBD' was generated using section 2.5.2 and the parameters 'sort_by = "logFC"' and 'gene_filter = 20' were set.

Source

[doi:10.1038/s43588023004649](#)

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_raw](#), [Cellmarker2_table](#), [Markers_list_PCTAM](#), [Markers_list_PCTIT](#), [Markers_list_TCellSI](#), [PanglaoDB](#), [PanglaoDB_raw](#), [PanglaoDB_table](#)

Markers_list_TCellSI *List of T cell subtype markers in the article TCellSI*

Description

A dataset containing marker genes for different T cell subtypes from TCellSI

Usage

`Markers_list_TCellSI`

Format

A list with ten tables.

Details

This list is a table of 10 types of T cell markers obtained from TCellSI. The data source is "<https://github.com/GuoBioinfoLab/TCellSI>" and the reference literature is: Yang et al. (2024) [doi:10.1101/imt2.231](#).

Source

<https://github.com/GuoBioinfoLab/TCellSI/>

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_raw](#), [Cellmarker2_table](#), [Markers_list_PCTAM](#), [Markers_list_PCTIT](#), [Markers_list_scIBD](#), [PanglaoDB](#), [PanglaoDB_raw](#), [PanglaoDB_table](#)

PanglaoDB

PanglaoDB dataset

Description

A dataset containing marker genes for different cell types from PanglaoDB

Usage

PanglaoDB

Format

A data frame with 9 columns:

Details

This dataset is used to filter and create a standardized marker list.⁷

Source

<https://panglaodb.se/>

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_raw](#), [Cellmarker2_table](#), [Markers_list_PCTAM](#), [Markers_list_PCTIT](#), [Markers_list_TCellSI](#), [Markers_list_scIBD](#), [PanglaoDB_raw](#), [PanglaoDB_table](#)

PanglaoDB_raw *PanglaoDB raw dataset*

Description

A dataset containing marker genes for different cell types from PanglaoDB

Usage

PanglaoDB_raw

Format

A data frame with 14 columns contained in the PanglaoDB database:

Details

This dataset is used to filter and create a standardized marker list.'

Source

<https://panglaodb.se/>

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_raw](#), [Cellmarker2_table](#), [Markers_list_PCTAM](#),
[Markers_list_PCTIT](#), [Markers_list_TCellSI](#), [Markers_list_scIBD](#), [PanglaoDB](#), [PanglaoDB_table](#)

PanglaoDB_table *PanglaoDB table*

Description

A dataset containing marker genes for different cell types from PanglaoDB

Usage

PanglaoDB_table

Format

A list contain different types like species, organ, cell type.

Details

This list is used to choose filters for creation of standardized marker list.

Source

<https://panglaodb.se/>

See Also

Other Section_0_Database: [Cellmarker2](#), [Cellmarker2_raw](#), [Cellmarker2_table](#), [Markers_list_PCTAM](#), [Markers_list_PCTIT](#), [Markers_list_TCellSI](#), [Markers_list_scIBD](#), [PanglaoDB](#), [PanglaoDB_raw](#)

Parameter_Calculate *Adaptive Parameter Tuning for Single-Cell Data Annotation in SlimR*

Description

This function automatically determines optimal min_expression, specificity_weight, and threshold parameters for single-cell data analysis based on dataset characteristics using adaptive algorithms derived from empirical analysis of single-cell datasets.

Usage

```
Parameter_Calculate(  
  seurat_obj,  
  features = NULL,  
  assay = NULL,  
  cluster_col = NULL,  
  n_celltypes = 50,  
  verbose = TRUE  
)
```

Arguments

seurat_obj	A Seurat object containing single-cell data
features	Character vector of feature names (genes) to analyze. If NULL, will use highly variable features from the Seurat object.
assay	Name of assay to use (default: default assay)
cluster_col	Column name in metadata containing cluster information
n_celltypes	Expected number of cell types in marker database (default: 50). Used for threshold recommendation calculation.
verbose	Whether to print progress messages (default: TRUE)

Value

A list containing:

- min_expression: Recommended expression threshold
- specificity_weight: Recommended specificity weight
- threshold: Recommended probability threshold for candidate selection
- dataset_features: Extracted dataset characteristics
- parameter_rationale: Explanation of parameter choices

See Also

Other Section_3_Automated_Annotation: [Celltype_annotation\(\)](#), [Celltype_calculate\(\)](#), [Celltype_verification\(\)](#)

Examples

```
## Not run:
SlimR_params <- Parameter_Calculate(
  seurat_obj = sce,
  features = c("CD3E", "CD4", "CD8A"),
  assay = "RNA",
  cluster_col = "seurat_clusters",
  n_celltypes = 98,
  verbose = TRUE
)

## End(Not run)
```

Read_excel_markers *Create "Marker_list" from Excel files ".xlsx"*

Description

Create "Marker_list" from Excel files ".xlsx"

Usage

```
Read_excel_markers(path, has_colnames = TRUE)
```

Arguments

path	The path information of Marker files stored in ".xlsx" format. The Sheet name in the file is filled with cell type. The first line of each Sheet is the table head, the first column is filled with markers information, and the following column is filled with metric information.
has_colnames	Logical value indicating whether the first row contains column names. If FALSE, the first column will be named "Markers" and subsequent columns will be named "Col1", "Col2", etc.

Value

The standardized "Marker_list" in the SlimR package.

See Also

Other Section_2_Standardized_Markers_List: [Markers_filter_Cellmarker2\(\)](#), [Markers_filter_PanglaoDB\(\)](#), [Read_seurat_markers\(\)](#)

Examples

```
## Not run:
Markers_list_Excel <- Read_excel_markers(
  "D:/Laboratory/Marker_load.xlsx"
)
## End(Not run)
```

Read_seurat_markers *Create "Marker_list" from Seurat object*

Description

Create "Marker_list" from Seurat object

Usage

```
Read_seurat_markers(
  df,
  sources = c("Seurat", "presto"),
  sort_by = "FSS",
  gene_filter = 20
)
```

Arguments

df	Dataframe generated by "FindAllMarkers" function, recommend to use parameter "group.by = "Cell_type"" and "only.pos = TRUE".
sources	Type of markers sources to use. Be one of: "Seurat" or "presto".
sort_by	Marker sorting parameter, for Seurat sources, select "avg_log2FC" or "p_val_adj" or "FSS" (Feature Significance Score, FSS, product value of log2FC and Expression ratio). Default parameters use "sort_by = 'FSS'". for presto sources, select "logFC" or "padj" or "FSS". Default parameters use "sort_by = 'FSS'".
gene_filter	The number of markers left for each cell type based on the "sort_by" parameter's level of difference. Default parameters use "gene_filter = 20"

Value

The standardized "Marker_list" in the SlimR package.

See Also

Other Section_2_Standardized_Markers_List: [Markers_filter_Cellmarker2\(\)](#), [Markers_filter_PanglaoDB\(\)](#), [Read_excel_markers\(\)](#)

Examples

```
## Not run:  
# Example for Seurat sources markers  
seurat_markers <- Seurat::FindAllMarkers(  
    object = sce,  
    group.by = "Cell_type",  
    only.pos = TRUE)  
  
Markers_list_Seurat <- Read_seurat_markers(seurat_markers,  
    sources = "Seurat",  
    sort_by = "avg_log2FC",  
    gene_filter = 20  
)  
  
# Example for presto sources markers  
seurat_markers <- dplyr::filter(  
    presto::wilcoxauc(  
        X = sce,  
        group_by = "Cell_type",  
        seurat_assay = "RNA"  
    ),  
    padj < 0.05, logFC > 0.5  
)  
  
Markers_list_Seurat <- Read_seurat_markers(seurat_markers,  
    sources = "presto",  
    sort_by = "logFC",  
    gene_filter = 20  
)  
  
## End(Not run)
```

Index

- * **Section_0_Database**
 - Cellmarker2, 5
 - Cellmarker2_raw, 6
 - Cellmarker2_table, 7
 - Markers_list_PCTAM, 28
 - Markers_list_PCTIT, 29
 - Markers_list_scIBD, 30
 - Markers_list_TCellSI, 30
 - PanglaoDB, 31
 - PanglaoDB_raw, 32
 - PanglaoDB_table, 32
- * **Section_1_Functions_Use_in_Package**
 - calculate_cluster_variability, 2
 - calculate_expression, 3
 - calculate_expression_skewness, 4
 - calculate_probability, 4
 - compute_adaptive_parameters, 24
 - estimate_batch_effect, 25
 - extract_dataset_features, 25
- * **Section_2_Standardized_Markers_List**
 - Markers_filter_Cellmarker2, 26
 - Markers_filter_PanglaoDB, 27
 - Read_excel_markers, 34
 - Read_seurat_markers, 35
- * **Section_3_Automated_Annotation**
 - Celltype_annotation, 7
 - Celltype_Calculate, 20
 - Celltype_Verification, 23
 - Parameter_Calculate, 33
- * **Section_4_Semi_Automated_Annotation**
 - Celltype_annotation_Combined, 10
 - Celltype_annotation_Features, 13
 - Celltype_annotation_Heatmap, 15
- * **Section_5_Other_Functions_Provided**
 - Celltype_annotation_Cellmarker2, 8
 - Celltype_annotation_Excel, 11
 - Celltype_annotation_PanglaoDB, 17
 - Celltype_annotation_Seurat, 19
- * **datasets**
 - Cellmarker2, 5
 - Cellmarker2_raw, 6
 - Cellmarker2_table, 7
 - Markers_list_PCTAM, 28
 - Markers_list_PCTIT, 29
 - Markers_list_scIBD, 30
 - Markers_list_TCellSI, 30
 - PanglaoDB, 31
 - PanglaoDB_raw, 32
 - PanglaoDB_table, 32
- calculate_cluster_variability, 2, 4, 5, 24–26
- calculate_expression, 3, 3, 4, 5, 24–26
- calculate_expression_skewness, 3, 4, 4, 5, 24–26
- calculate_probability, 3, 4, 4, 24–26
- Cellmarker2, 5, 6, 7, 29–33
- Cellmarker2_raw, 6, 6, 7, 29–33
- Cellmarker2_table, 6, 7, 29–33
- Celltype_annotation, 7, 22, 23, 34
- Celltype_annotation_Cellmarker2, 8, 12, 18, 20
- Celltype_annotation_Combined, 10, 14, 17
- Celltype_annotation_Excel, 10, 11, 18, 20
- Celltype_annotation_Features, 11, 13, 17
- Celltype_annotation_Heatmap, 11, 14, 15
- Celltype_annotation_PanglaoDB, 10, 12, 17, 20
- Celltype_annotation_Seurat, 10, 12, 18, 19
- Celltype_Calculate, 8, 20, 23, 34
- Celltype_Verification, 8, 22, 23, 34
- compute_adaptive_parameters, 3–5, 24, 25, 26
- estimate_batch_effect, 3–5, 24, 25, 26
- extract_dataset_features, 3–5, 24, 25, 25
- Markers_filter_Cellmarker2, 26, 28, 34, 35

Markers_filter_PanglaoDB, 27, 27, 34, 35
Markers_list_PCTAM, 6, 7, 28, 29–33
Markers_list_PCTIT, 6, 7, 29, 29, 30–33
Markers_list_scIBD, 6, 7, 29, 30, 31–33
Markers_list_TCellSI, 6, 7, 29, 30, 30,
 31–33

PanglaoDB, 6, 7, 29–31, 31, 32, 33
PanglaoDB_raw, 6, 7, 29–31, 32, 33
PanglaoDB_table, 6, 7, 29–32, 32
Parameter_Calculate, 8, 22, 23, 33

Read_excel_markers, 27, 28, 34, 35
Read_seurat_markers, 27, 28, 34, 35