Chapter 8
Meta-Analysis of Multiple GWAS Data

Genome-wide association scans allow millions of single-nucleotide polymorphisms (SNPs) to be genotyped or imputed and studied simultaneously using hypothesis-free agnostic approaches. Although single genome-wide association studies (GWASs) have identified many variants associated with complex diseases, these variants currently explain little of the risk variability for most diseases. Genetic effects due to common alleles are small, and detection of signals requires large sample sizes. As single GWASs are underpowered, meta-analysis — the statistical synthesis of information from multiple independent studies — increases power and reduces false-positive findings. Also, the techniques developed for meta-analysis can use summary data, diminishing the limitations that are imposed by restrictions on sharing individual-level data. Therefore, meta-analysis has become a popular approach for the discovery of new genetic loci for common diseases and phenotypes. Most genetic risk variants discovered in the past few years have come from large-scale meta-analyses of GWASs7. Several hundred GWAS meta-analyses have already been published. In the period between the beginning of 2009 and 15th June 2012, 139 of these meta-analyses had sample sizes in the discovery phase exceeding 10,000 participants. These efforts have dramatically increased the yield of discovered and validated genetic risk loci, thus making a case that the manpower and time required for such a design are justified. Large meta-analyses may continue to increase the yield of loci in proportion to total sample size.

As GWAS meta-analysis has become so popular and as new waves of even more extensive data are accumulating from sequencing efforts, it is timely to review the meta-analysis methods that have successfully been applied so far and also to highlight novel approaches that may be useful for the discovery of new variants. We cover the main models along with their extensions and the inferential tools that are being used, providing guidance for the most appropriate methods in particular situations. Moreover, we address issues of heterogeneity and describe methodological approaches that take into account heterogeneity introduced by various sources, such as phenotype definition, different ancestral groups, different genotyping platforms or imputation software. Given that research is increasingly shifting towards low-frequency and rare variants, we also describe methods for the meta-analysis of these variants.

This introduction was adopted from Evangelou and Ioannidis (2013, Nature Review Genetics)

Meta-analysis requires results of GWAS from separate environments (environment specific analyses) or independent studies. For each locus, we only need the p-value or estimated effect and the estimation error of the effect. Any method of GWAS can be used in metag-analysis. However, we used the most advanced method of GWAS by controlling the polygenic background noise (XU 2013). When the polygenic background only contains the additive effects, the method is equivalent to the genome-wide efficient

47   mixed model analysis for association studies (GEMMA) (ZHOU AND STEPHENS 2012).
48   Methods of estimating marker effects for the independent studies are not the main
49   concerns of this study, but for the paper to be self-contained, we briefly introduce the
50   model here. Let y be an $n \times 1$ vector for the phenotypic values of a quantitative trait
51   from one environment. The linear mixed model is

52
$$y = X\beta + Z_k\gamma_k + \xi + \varepsilon \tag{1}$$

53   The model effects are defined as follows: X and β are the design matrix and effects
54   for some systematic errors (treated as fixed effects); Z_k is a genotype indicator variable
55   for marker k and γ_k is the effect for this marker (treated as fixed effect), where
56     $k = 1,...,m$ and m is the number of markers; ξ is a polygenic effect following a $N(0, K\sigma_\xi^2)$
57   distribution, where K is an $n \times n$ kinship matrix calculated from genome-wide marker
58   information and σ_ξ^2 is a polygenic variance; ε is a vector of residual errors following a
59     $N(0, I\sigma^2)$ distribution, where σ^2 is the residual error variance. Let us denote the
60   estimated effect and its estimation error variance for marker k by $\tilde{\gamma}_k$ and $\mathrm{var}(\tilde{\gamma}_k) = \tilde{v}_k$,
61   where $k = 1,..,m$ for all markers in the genome. For T separate studies, we will have T
62   such estimates and error variances for each marker. We now focus our analysis on one
63   marker since the genome-wide marker analysis is simply calling a loop of the single
64   marker analysis over all markers. We now drop subscript k and replace it by t for the
65   same marker but over $t = 1,...,T$ environments. In other words, $\tilde{\gamma}_t$ and $\tilde{v}_t$ are the
66   estimated effect and its error variance for the current marker of interest from the tth
67   environment.
68
69   We will learn several methods of meta-analysis in this chapter. One method requires the
70   p-values of all markers from all independent studies (Fisher's method) and the
71   remaining methods require the estimated effects and the estimation errors of the
72   effects.
73
74   **1. Fisher's method**
75
76   This method is very simple, but is still very effective. We now consider only one locus
77   and introduce the method for this locus. The entire genome scan requires looping over
78   all loci. Let p_t be the p-value from the t-th study for $t = 1,...,T$ where T is the number of
79   independent studies. Fisher (19xx) proposed a combined test statistic

80
$$X^2 = -2\sum_{t=1}^{T}\ln(p_t)$$

81   where the "ln" represents the natural log. He showed that under the null model (the p's
82   are independent uniform distribution between 0 and 1), this combined test statistic
83   follows a chi-square distribution with $df = 2T$ degrees of freedom. Therefore, the new p-
84   value from the meta-analysis is calculated using

85
$$p_{\mathrm{Fisher}} = 1 - \mathrm{Pr}(\chi_{2T}^2 \leq X^2)$$

86   where χ^2_{2T} is a chi-square variable with $2T$ degrees of freedom. This method is
87   extremely simple and only requires the p-values. Therefore, it has been widely adopted
88   in meta-analysis.
89
90   An alternative method similar to the Fisher's method is to use probit transformation of
91   the p-values. Let $z_t = \Phi^{-1}(1-0.5p_t)$ be the probit (quantile) of $q_t = 1-0.5p_t$. The 0.5 in
92   front of p_t is due to the two-tailed nature of the normal test. If p_t is uniformly distributed
93   between 0 and 1, z_t should be the positive half of the standardized normal variable
94   (truncated standardized normal distribution). As a result,

$$X^2 = \sum_{t=1}^{T} z_t^2 = \sum_{t=1}^{T} \left[\Phi^{-1}(1-0.5p_t) \right]^2$$

96   should follow a chi-square distribution with T degrees of freedom. The new p-value
97   should be calculated using

$$p_{\text{Probit}} = 1 - \Pr(\chi^2_T \le X^2)$$

99   where χ^2_T is a chi-square variable with T degrees of freedom. This method is more
100 intuitive than the Fisher's method because there is no need to prove it. However, the
101 calculation is complicated because you need the probit function, which is not available
102 in many computing platforms. The combined p-value calculated this way is different
103 from that of the Fisher's method. For example, if the four p-values from four separate
104 GWAS are

$$p = \begin{bmatrix} p_1 & p_2 & p_3 & p_4 \end{bmatrix} = \begin{bmatrix} 0.7570 & 0.2907 & 0.12511 & 0.5745 \end{bmatrix}$$

106 then the combined test statistics from the Fisher's method and the Probit method are

$$X^2_{\text{Fisher}} = 8.292636 \text{ and } X^2_{\text{Probit}} = 3.879056$$

108 respectively. The corresponding combined p-values are

$$p_{\text{Fisher}} = 0.4054192 \text{ and } p_{\text{Probit}} = 0.4226212$$

110 The probit method has not been tested. Therefore, we do not have an answer whether
111 this new test is better or worse than the Fisher's test.
112
113 **2. Fixed model methodology**
114
115 This method is called the "main" effect test, assuming that all effects have the same
116 direction. We are actually testing the mean effect from all independent studies. This
117 method requires the estimated marker effect and its estimation error (variance) for each
118 study, denoted by $\tilde{\gamma}_t$ and $\tilde{v}_t = \text{var}(\tilde{\gamma}_t)$, respectively, for the tth study. The data are
119 represented by $\{\tilde{\gamma}_t, \tilde{v}_t\}$ for $t = 1,...,T$. Let $\tilde{\gamma} = \{\tilde{\gamma}_1,...,\tilde{\gamma}_T\}$ be a vector of the data points and
120 $\tilde{v} = \text{diag}\{\tilde{v}_1,...,\tilde{v}_T\}$ be a matrix of the error variances. We now write a linear model for $\tilde{\gamma}$

$$\tilde{\gamma} = \mu_\gamma + e_\gamma \tag{2}$$

122 where μ_γ is a mean value representing the main effect for the marker of interest and e_γ
123 is an $T \times 1$ vector of errors with a known $N(0,\hat{v})$ distribution. The parameter is μ_γ and
124 the data are $\tilde{\gamma}$ and $\tilde{v}$. The log likelihood function is

3

125
$$L(\mu_\gamma) = -\frac{1}{2}\sum_{t=1}^{T}\ln(\tilde{v}_t) - \frac{1}{2}\sum_{t=1}^{T}\frac{(\tilde{\gamma}_t - \mu_\gamma)^2}{\tilde{v}_t}$$
(3)

126 whose MLE is denoted by $\hat{\mu}_\gamma$, which happens to have an explicit solution,

127
$$\hat{\mu}_\gamma = \left(\sum_{t=1}^{T}\frac{1}{\tilde{v}_t}\right)^{-1}\left(\sum_{t=1}^{T}\frac{\tilde{\gamma}_t}{\tilde{v}_t}\right)$$
(4)

128 You can try it by taking the derivative of $L(\mu_\gamma)$ with respect to μ_γ, letting the derivative
129 be zero and solving for the parameter. The variance of the estimate happens to be

130
$$\text{var}(\hat{\mu}_\gamma) = \left(\sum_{t=1}^{T}\frac{1}{\tilde{v}_t}\right)^{-1}$$
(5)

131 The null hypothesis is $H_0 : \mu_\gamma = 0$, which is tested via the following Wald test,

132
$$W = \frac{\hat{\mu}_\gamma^2}{\text{var}(\hat{\mu}_\gamma)}$$

133 Under the null hypothesis, the Wald test statistic follows a chi-square distribution with
134 one degree of freedom.
135
136 The fixed model method only tests the so called "main" effect because it assumes that
137 the effects of all independent studies have the same direction. If this assumption is
138 violated, positive and negative effects may be cancelled out each other, leading to a low
139 power of the meta-analysis. Therefore, this is not the optimal method for some data.
140
141 **3. Random model methodology**
142
143 Let $\gamma = \begin{bmatrix} \gamma_1 & \gamma_2 & \cdots & \gamma_T \end{bmatrix}^T$ be the true effects from T separate analyses. We now assume
144 $\gamma \sim N(0, I\sigma_\gamma^2)$, i.e., the true effects are independent normal variables with mean zero and
145 a common variance. We can write the following random model
146
$$\tilde{\gamma} = \gamma + e_\gamma$$
(6)
147 where the residual error variances are known, $\text{var}(e_\gamma) = \text{diag}(\tilde{v})$. The likelihood function
148 is

149
$$L(\sigma_\gamma^2) = -\frac{1}{2}\sum_{t=1}^{T}\ln(\sigma_\gamma^2 + \tilde{v}_t) - \frac{1}{2}\sum_{t=1}^{T}\frac{\tilde{\gamma}_t^2}{\sigma_\gamma^2 + \tilde{v}_t}$$
(7)

150 The maximum likelihood estimates (MLE) of σ_γ^2 can be obtained by numerically solving

151
$$\frac{\partial}{\partial\theta}L(\sigma_\gamma^2) = 0$$
(8)

152 for the solution of σ_γ^2, denoted by $\hat{\sigma}_\gamma^2$. The null hypothesis is $H_0 : \sigma_\gamma^2 = 0$, which is tested
153 via the following likelihood ratio test,
154
$$\lambda = -2\left[L(\sigma_\gamma^2 = 0) - L(\hat{\sigma}_\gamma^2)\right]$$
(9)
155 where

$$156 \qquad L(\sigma_\gamma^2 = 0) = -\frac{1}{2}\sum_{t=1}^{T}\ln(\tilde{v}_t) - \frac{1}{2}\sum_{t=1}^{T}\frac{\tilde{\gamma}_t^2}{\tilde{v}_t} \qquad (10)$$

157   is the likelihood value evaluated at $\sigma_\gamma^2 = 0$. Under the null model, λ follows

158   approximately a mixture of two chi-square distribution,

$$159 \qquad \lambda \sim 0.5\chi_0^2 + 0.5\chi_1^2$$

160   Therefore, the p-value is calculated using

$$161 \qquad p = \begin{cases} 1 & \text{if} \quad \lambda = 0 \\ \frac{1}{2}\left[1 - \Pr(\chi_1^2 < \lambda)\right] & \text{otherwise } \lambda > 0 \end{cases} \qquad (11)$$

162   Interpretation of the random model approach may not be intuitive to many people. The

163   null hypothesis $\sigma_\gamma^2 = 0$ means that effects from all independent studies are the same

164   and, in addition, they are all equal to zero. One non-zero effect will cause $\sigma_\gamma^2 > 0$. If the

165   effects have different directions, they will not cancel out each other. Therefore, the

166   random model approach is more robust than the fixed model approach.

167

168   **4. Mixed model methodology**

169

170   We now use a linear mixed model to describe $\tilde{\gamma}$

$$171 \qquad \tilde{\gamma} = \mu_\gamma + \gamma + e_\gamma \qquad (12)$$

172   where μ_γ is a mean value representing the main effect for the marker of interest, γ is a

173   vector of true effects following an assumed $N(0, I\sigma_\gamma^2)$ distribution, and e_γ is an $T \times 1$

174   vector of errors with a known $N(0, \tilde{v})$ distribution, where $\tilde{v} = \mathrm{diag}(\tilde{v}_1, \cdots \tilde{v}_T)$. The

175   parameters are μ_γ and σ_γ^2. The log likelihood function is

$$176 \qquad L(\mu_\gamma, \sigma_\gamma^2) = -\frac{1}{2}\sum_{t=1}^{T}\ln(\sigma_\gamma^2 + \tilde{v}_t) - \frac{1}{2}\sum_{t=1}^{T}\frac{(\tilde{\gamma}_t - \mu_\gamma)^2}{\sigma_\gamma^2 + \tilde{v}_t} \qquad (13)$$

177   Numerical solutions can be obtained via the Newton-Raphson method. Let $\hat{\theta} = \{\hat{\mu}_\gamma, \hat{\sigma}_\gamma^2\}$

178   be the MLE of the parameters. We now have two hypotheses to test, one for the main

179   effect $H_0 : \mu_\gamma = 0$ and one for the marker-by-population interaction $H_0 : \sigma_\gamma^2 = 0$. Rejection

180   of either hypothesis means a significant association. If we are not interested in

181   differentiation of whether the association is due to the main effect or interaction, we can

182   combine the two tests together to develop a composite test $H_0 : \mu_\gamma = \sigma_\gamma^2 = 0$. Let

$$183 \qquad L(0,0) = -\frac{1}{2}\sum_{t=1}^{T}\ln(\tilde{v}_t) - \frac{1}{2}\sum_{t=1}^{T}\frac{\tilde{\gamma}_t^2}{\tilde{v}_t} \qquad (14)$$

184   be the likelihood value under the null model, the likelihood ratio test statistic is

$$185 \qquad \lambda_{\mu,\sigma^2} = -2\left[L(0,0) - L(\hat{\mu}_\gamma, \hat{\sigma}_\gamma^2)\right] \qquad (15)$$

186   Under the null model, this test statistic follows approximately a mixture of chi-square

187   one and chi-square two distributions with an equal weight. Therefore, the p-value is

188   calculated using

189
$$p_{\mu,\sigma^2} = 0.5 \Pr(\chi_1^2 \geq \lambda_{\mu,\sigma^2}) + 0.5 \Pr(\chi_2^2 \geq \lambda_{\mu,\sigma^2}) \qquad (16)$$

190 where χ_1^2 and χ_2^2 are variables of chi-square one and chi-square two, respectively. The
191 approximation can be very good if T is relatively large (citation).
192
193 We now introduce separate tests for the main effect (mean) and interaction effect
194 (variance). For the main effect hypothesis, $H_0 : \mu_\gamma = 0$, the Wald test is

195
$$\text{Wald} = \frac{\hat{\mu}_\gamma^2}{\text{var}(\hat{\mu}_\gamma)} = \hat{\mu}_\gamma^2 \sum_{t=1}^{T} (\hat{\sigma}_\gamma^2 + \tilde{v}_t)^{-1} \qquad (17)$$

196 Under the null hypothesis, the Wald test follows approximately a chi-square distribution
197 with one degree of freedom. For the GxE interaction hypothesis, $H_0 : \sigma_\gamma^2 = 0$, a likelihood
198 ratio test is appropriate,

199
$$\lambda_{\sigma^2} = -2 \left[L(\hat{\mu}_\gamma, 0) - L(\hat{\mu}_\gamma, \hat{\sigma}_\gamma^2) \right] \qquad (18)$$

200 where $L(\hat{\mu}_\gamma, 0)$ is the likelihood value evaluated at $\sigma_\gamma^2 = 0$. This test statistic follows

201 approximately a mixture distribution of $\chi_{(0)}^2$ and $\chi_{(1)}^2$ with an equal weight. Therefore, the

202 p-value is calculated using

203
$$p_{\sigma^2} = 0.5 \Pr(\chi_0^2 \geq \lambda_{\sigma^2}) + 0.5 \Pr(\chi_1^2 \geq \lambda_{\sigma^2}) \qquad (19)$$

204
205 The mixed model approach seems to be redundant compared with the random model
206 approach. However, the test results can be different. Simulation studies are required to
207 compare the performance of the two models.
208
209 **5. Application of meta-analysis to GxE interaction**
210
211 Kang et al. (2014) demonstrated that methods of meta-analysis can be applied to
212 analyze GxE interaction. Here, we used an RIL population of rice evaluated in four
213 environments as an example to demonstrate such an application. The rice population
214 consists of 210 RILs derived from a hybrid rice, Shanyou63, which was the cross of
215 Zhenshan97 and Minghui63. The experiment was replicated in two years in two
216 locations (four environments). The trait of interest is yield per plant. The markers
217 (270,000) were converted into 1619 bins that are evenly distributed across all 12
218 chromosomes.
219

Table 1. Estimated mean and variance of rice in four environments

Environment	Mean	Residual variance	Polygenic variance	Heritability
1	19.5412	8.5988	4.1860	0.3274
2	20.8201	9.1668	20.3685	0.6896
3	29.9017	35.0289	31.3583	0.4723
4	23.5324	42.4509	39.2117	0.4801
Average	23.4489	23.8114	23.7811	0.4923

The estimated bin effects of yield across the entire genome are depicted in Figure 1 for all four environments. The second half of chromosome 1 and the middle of chromosome 8 appear to have some evidence of QTL. Figure 2 shows the test statistic, $-\log_{10}(p)$, plotted against genome location for each of the four environments. Only one region in chromosome 8 from environment 4 has reached the Bonferroni corrected significance level.

233
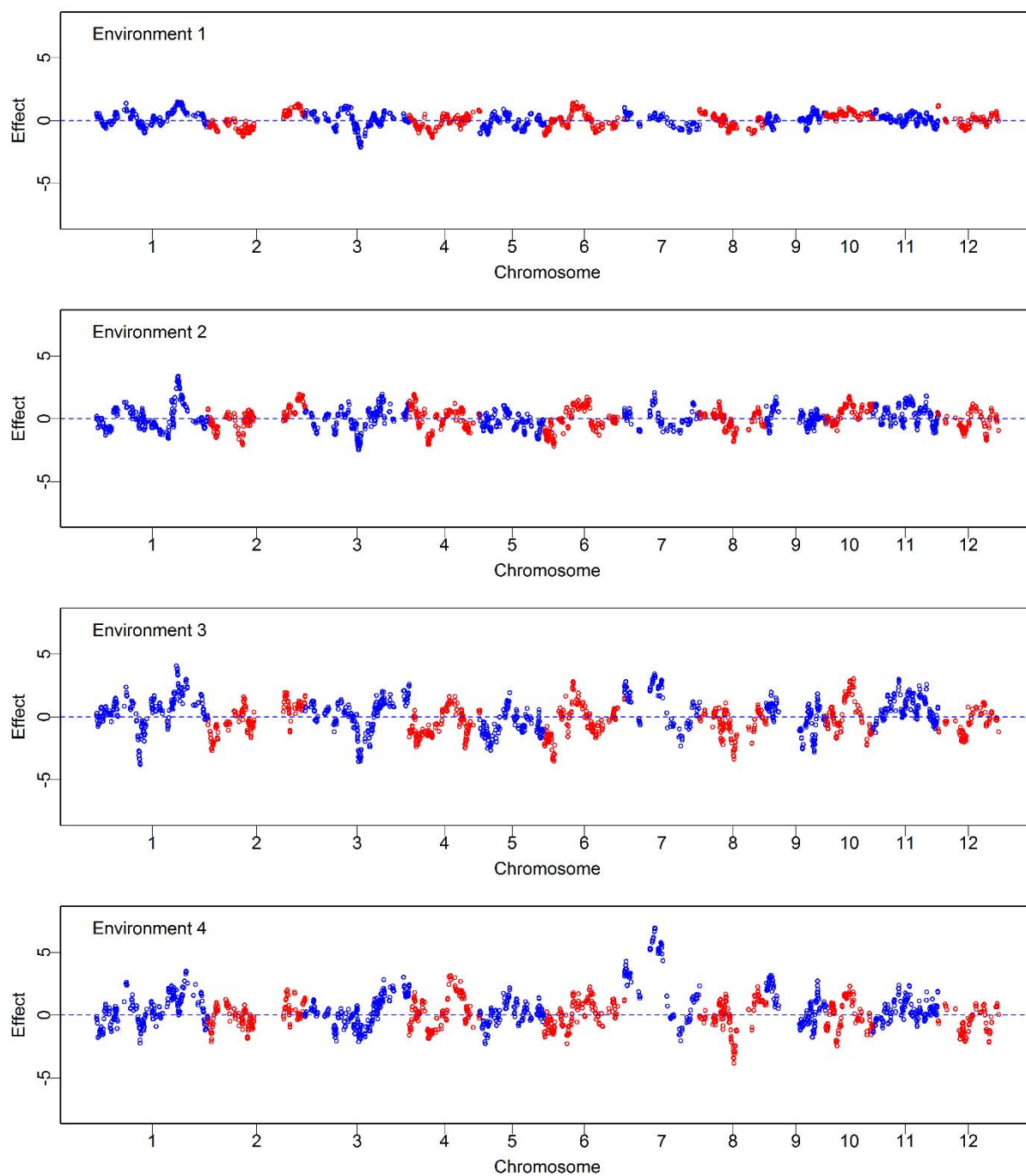
234
235 **Figure 1.** Estimated effects of bins for yield of rice across the entire genome for all four
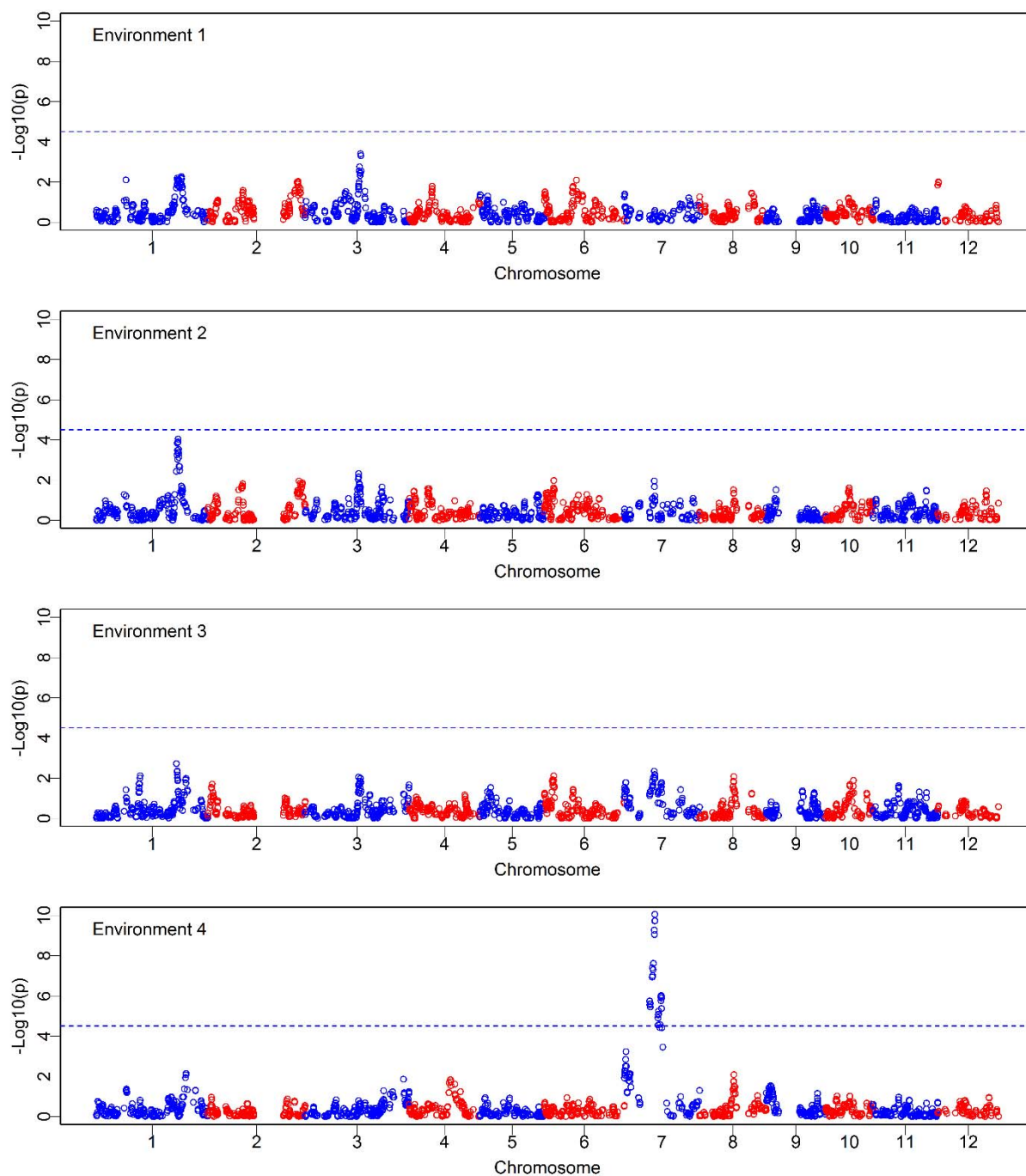236 environments.
237

238

239
240 **Figure 2.** Test statistics of bins for yield of rice across the entire genome for all four
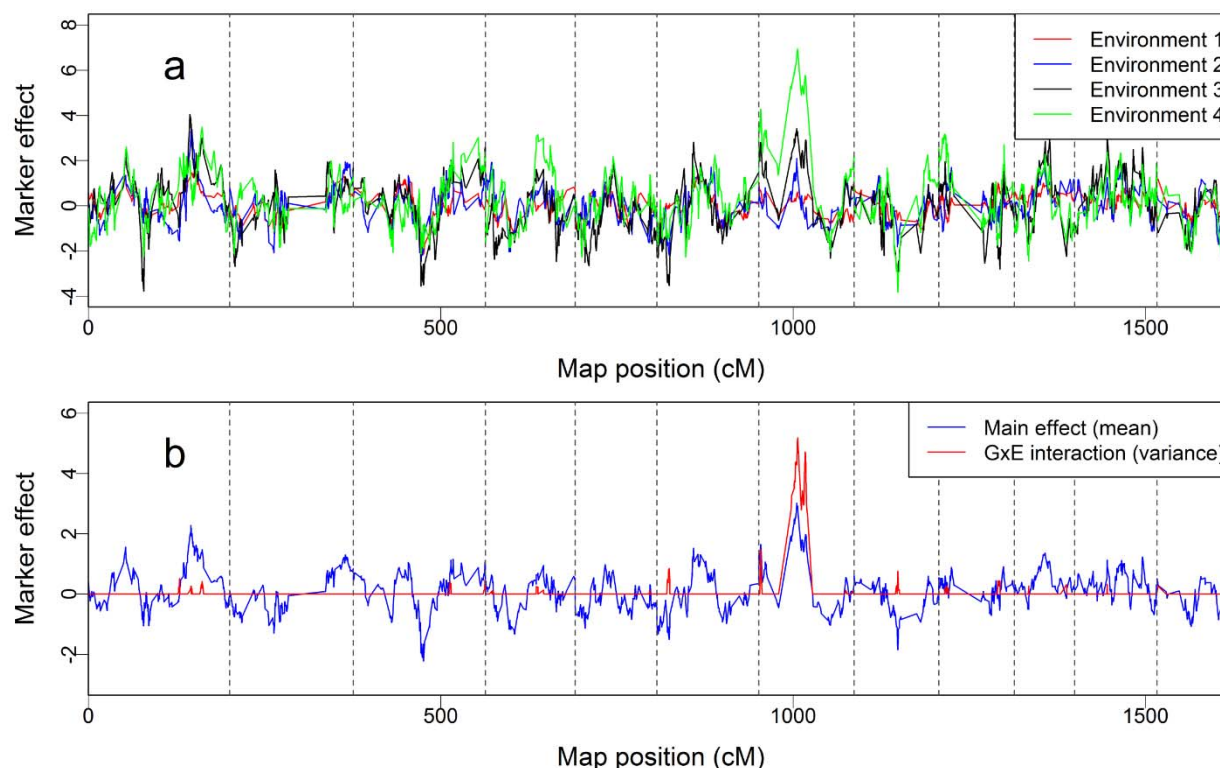241 environments.
242

243

244
245
Figure 3. Additional plots of GxE interaction of rice yield from the RIL population
evaluated in four environments. Panel a: estimated marker effects of four environments
plotted together; Panel b: estimated main effects (mean) and interaction (variance)
plotted against genome location.

Figure 4 shows the plots of test statistics against genome location for the four meta-
analysis methods. It appears that the mixed model approach detected four significant
regions. The fixed model also detected the same four regions, but the peaks of the test
statistic are lower than the mixed model analysis. The Fisher's method and the random
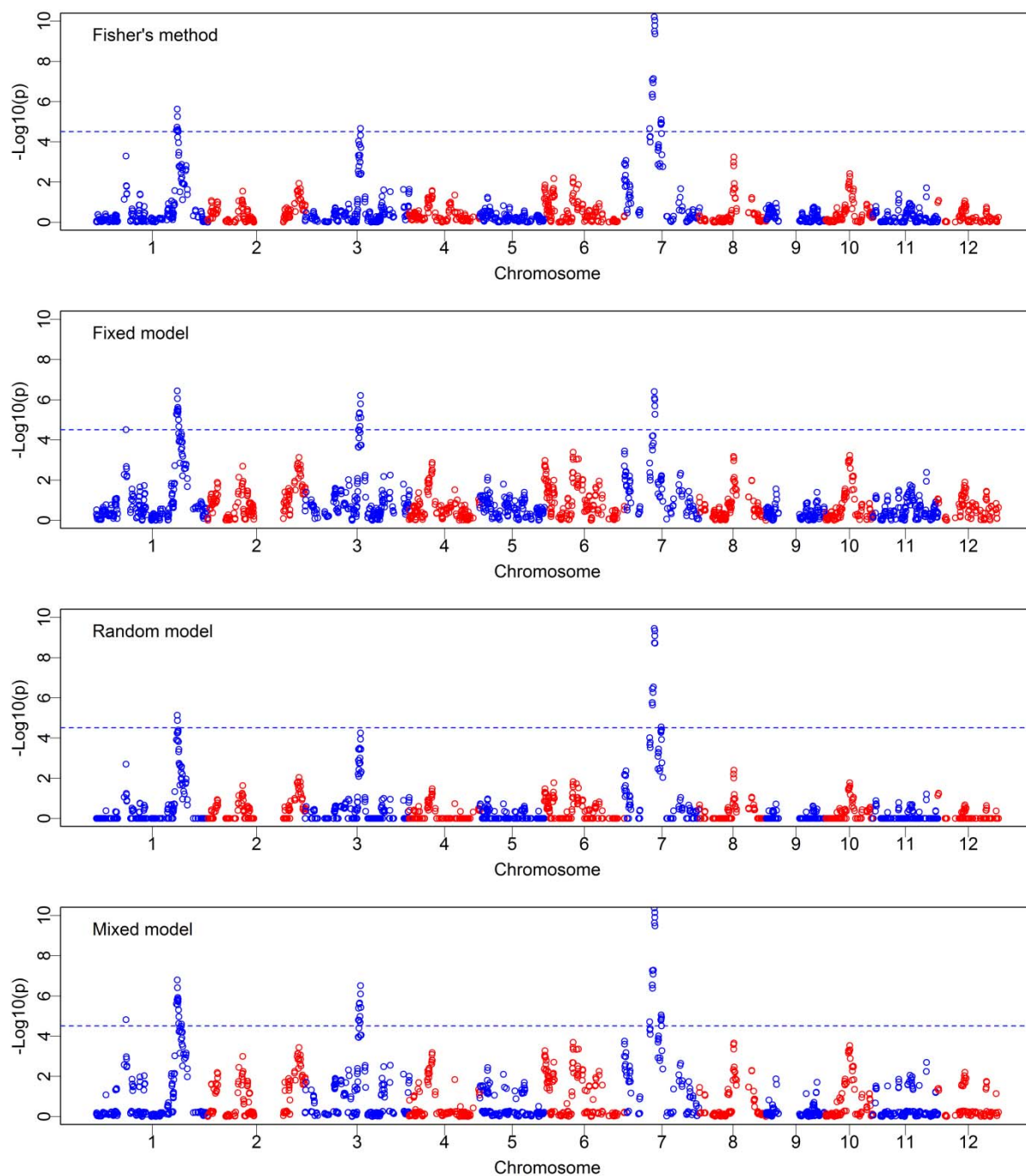model only detected three significant regions.

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258
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Figure 4. Test statistics of four methods for GxE studies of yield from the RIL population of rice.

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264   LITERATURE CITED
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281

GENOME-WIDE ASSOCIATION STUDIES

Meta-analysis methods for genome-wide association studies and beyond

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Abstract | Meta-analysis of genome-wide association studies (GWASs) has become a popular method for discovering genetic risk variants. Here, we overview both widely applied and newer statistical methods for GWAS meta-analysis, including issues of interpretation and assessment of sources of heterogeneity. We also discuss extensions of these meta-analysis methods to complex data. Where possible, we provide guidelines for researchers who are planning to use these methods. Furthermore, we address special issues that may arise for meta-analysis of sequencing data and rare variants. Finally, we discuss challenges and solutions surrounding the goals of making meta-analysis data publicly available and building powerful consortia.

Meta-analysis
A statistical method for the combination of different studies to provide a summary result.

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Genome-wide association scans allow millions of single-nucleotide polymorphisms (SNPs) to be genotyped or imputed and studied simultaneously using hypothesis-free agnostic approaches[1,2]. Although single genome-wide association studies (GWASs) have identified many variants associated with complex diseases, these variants currently explain little of the risk variability for most diseases. Genetic effects due to common alleles are small, and detection of signals requires large sample sizes[3–6]. As single GWASs are underpowered, meta-analysis — the statistical synthesis of information from multiple independent studies — increases power and reduces false-positive findings. Also, the techniques developed for meta-analysis can use summary data, diminishing the limitations that are imposed by restrictions on sharing individual-level data. Therefore meta-analysis has become a popular approach for the discovery of new genetic loci for common diseases and phenotypes. Most genetic risk variants discovered in the past few years have come from large-scale meta-analyses of GWASs[7]. Several hundred GWAS meta-analyses have already been published. In the period between the beginning of 2009 and 15th June 2012, 139 of these meta-analyses had sample sizes in the discovery phase exceeding 10,000 participants[8]. These efforts have dramatically increased the yield of discovered and validated genetic risk loci, thus making a case that the manpower and time required for such a design are justified. Large meta-analyses may continue to increase the yield of loci in proportion to total sample size[9,10].

As GWAS meta-analysis has become so popular and as new waves of even more extensive data are accumulating from sequencing efforts, it is timely to review the meta-analysis methods that have successfully been applied so far and also to highlight novel approaches that may be useful for the discovery of new variants. We cover the main models along with their extensions and the inferential tools that are being used, providing guidance for the most appropriate methods in particular situations. Moreover, we address issues of heterogeneity and describe methodological approaches that take into account heterogeneity introduced by various sources, such as phenotype definition, different ancestral groups, different genotyping platforms or imputation software. Given that research is increasingly shifting towards low-frequency and rare variants, we also describe methods for the meta-analysis of these variants.

Meta-analysis stages
Previous reviews have described in detail the organizational stages of a GWAS meta-analysis[7,11,12], and only a brief overview is provided here (FIG. 1). In BOX 1, we provide a more detailed description of different steps of meta-analysis, including setting up an analysis plan, dealing with heterogeneity, data storage, prioritization of variants for follow up and other secondary analysis[13–19]. Some successful high-profile consortia for which workflows can be followed and for which useful material describing their operations can be downloaded are summarized in TABLE 1.
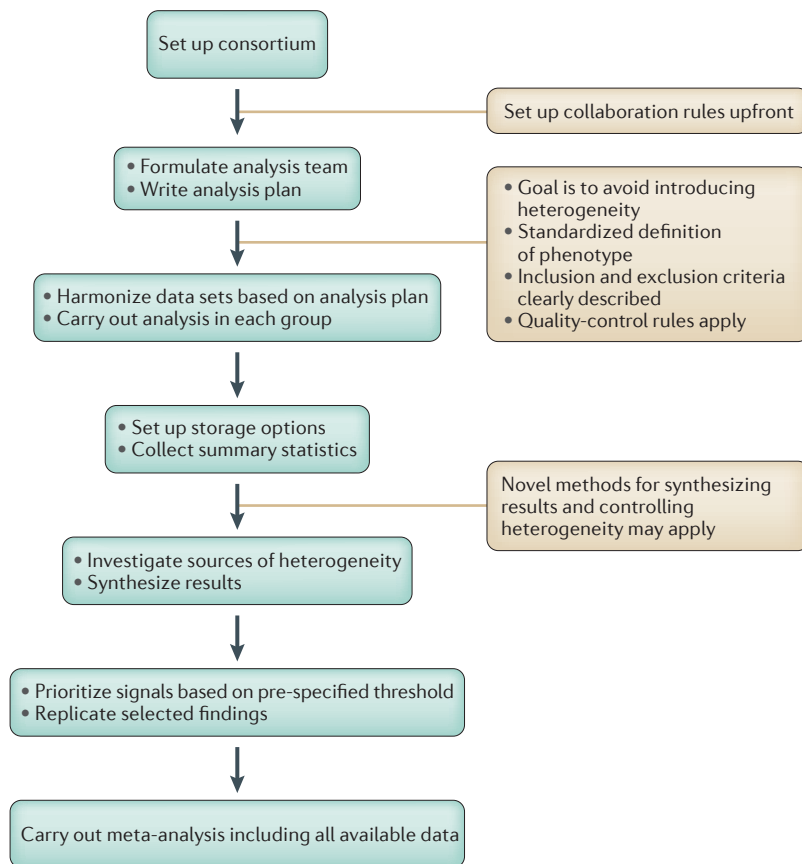
Figure 1 | **Stages in a meta-analysis.** A typical plan for a meta-analysis of genome-wide and next-generation sequence data.

size. Moreover, between-data-set heterogeneity cannot be assessed, and there is disagreement on the optimal weighting of studies. Furthermore, combining *P* values may be spurious when the direction of effects in the combined studies is not consistent.

A closely related approach to Fisher's method is based on *Z* scores[24]. As Fisher's method is based on the average of $-\log(P_i)$ across all studies (where P_i is the *P* value of the ith study) and as the *Z* scores method is based on the average of the Z_i values, these two approaches are highly correlated. One advantage of the *Z* score approach is that it takes into account the direction of the effect, and it is rather straightforward to introduce weights. A review has shown that meta-analysis of *Z* scores and *P* values (obtained using Fisher's method) has been applied in 14% and 3% of the published GWAS meta-analyses, respectively[25]. Both *P* values and *Z* scores meta-analysis might be useful tools for the synthesis of low-frequency variants (that is, those with a minor allele frequency (MAF) of 1–5%) or rare variants (that is, those with an MAF of <1%) when the association is based on tests that provide *P* values or *Z* test statistics rather than effect estimates[19,26] (see later for a discussion of methods for low-frequency and rare variants).

Fixed effects. Fixed effects meta-analysis is the most popular approach for synthesizing GWAS data and the most powerful approach for prioritizing and discovering phenotype-associated SNPs[27]. Fixed effects meta-analysis assumes that the true effect of each risk allele is the same in each data set. This is a rather tenuous assumption, but fixed effects models have the major advantage of maximizing discovery power compared to random effects models (see below)[28]. There are different models for fixed effects meta-analysis, but the inverse variance weighting[29], in which each study is weighted according to the inverse of its squared standard error, is predominantly used[7]. Another common method used in genetics is the Cochran–Mantel–Haenszel approach, which provides almost identical results to the inverse variance weighting method[24]. All bioinformatics tools that have been developed for GWAS meta-analysis carry out inverse variance meta-analysis[20–23].

Random effects. By far the most popular estimator of the between-study variance for the random effects approach is the DerSimonian and Laird estimator[30], but many others approaches also exist (for example, Sidik–Jonkman, Hedges–Vevea, Hunter–Schmidt and Schuster methods). The DerSimonian and Laird approach may give less robust results with small numbers (for example, rare variants)[24,31,32]. Random effects models are not used in discovery efforts owing to far more limited power than fixed effects models; however, random effects models are more appropriate than fixed effects models when the aim is to consider the generalizability of the observed association and estimate the average effect size of the associated variant and its uncertainty across different populations: for example, for predictive purposes[28,33].

Models for data synthesis

There are several approaches for GWAS meta-analysis. Here, we discuss methods that have been widely applied so far and alternative or new methods that have been proposed. For each model, issues such as weighing, power and ability to detect heterogeneity are discussed. TABLE 2 presents commonly used bioinformatics tools and software[20–23]. TABLE 3 presents the statistical properties of the most widely used methods.

P *values and Z scores.* Meta-analysis methods based on *P* values were widely used in different scientific fields until the 1980s but then became unpopular and were almost abandoned in biomedical sciences. This was because they suffered from several limitations, such as an inability to provide a summary effect and difficulties in addressing heterogeneity issues. The best known method of this type is Fisher's approach, but over a dozen other methods exist for combining *P* values in the traditional meta-analysis literature[24], although the large majority of these has never been applied with *P* values in the genome-wide significance range. In these approaches, the null hypothesis is that the true effect is null in each of the combined data sets (the alternative hypothesis is that there is a non-null association in at least one data set). The major disadvantage of *P* values meta-analysis is that it cannot provide an overall estimate of effect

Imputation
In genetics, the inference of genotypes of markers that have not been directly genotyped by making use of information from haplotype reference panels such as the HapMap or the 1000 Genomes panels.

Genome-wide significance
The significance threshold for rejecting the null hypothesis in genome-wide association studies.

Minor allele frequency
(MAF). The frequency of the less common allele of a polymorphic locus. It has a value that lies between 0 and 0.5 and can vary between populations.

Box 1 | Stages in a GWAS meta-analysis

Setting up an analysis plan

Each genome-wide association study (GWAS) meta-analysis initiative should be based on strong collaborative agreements and should be carefully designed and organized. An analysis team should design and draft a detailed plan that explicitly describes all of the steps of the anticipated analysis. Independent performance of some core statistical analyses by at least two analysts and using different software is not uncommon and may also allow for cross-verification and quality checks of processes and results. The analysis plan should be adopted by all teams, which should try to avoid deviations that introduce unnecessary between-study heterogeneity.

Dealing with heterogeneity

Despite careful planning to avoid heterogeneity, sometimes differences are inevitable, even in prospective designs: for example, when some samples have a family structure or when designs include extreme values[13]. Also, differences in phenotype definition may affect the estimated magnitude of the genetic effects, a factor that needs to be considered in terms of optimizing power for discovering new associations[6]. Ideally, phenotype definitions should be standardized according to stringent definitions applied in all data sets; if perfect standardization is impossible, participating teams should decide what kind of harmonization of definitions is desirable and feasible[14]. Inclusion and exclusion criteria of subjects and variants should be described in detail. Popular exclusion thresholds are >5% missing data, $P < 10^{-5}$ for Hardy–Weinberg equilibrium and quality index <0.3 for imputation metrics (BOX 2 summarizes the challenges of the imputation efforts using 1000 Genome Project panels[2,15]). Also, strand issues should be considered during quality control for the proper alignment of the alleles. Most GWAS meta-analyses to date focus on common variants and exclude variants with minor allele frequency <1%. However, this is likely to change as low-frequency and rare variants become the focus of interest. Statistical methods that account for between-study heterogeneity introduced by various sources are described in the main text.

Data storage

Data storage is an important aspect of meta-analysis as the individual-level data collected by each partner and also single participants' genotypes should be kept secured and unidentifiable. Most collaborative meta-analyses use online storage options to deposit summary data, giving access to members of the analysis team. This enables the partners to retain control of individual-level primary data. In most settings, summary data are statistically as efficient for meta-analysis as individual-level data[16]. The major drawback of working with summary data comes when more detailed investigations are required, such as conditional analyses, gene–gene interactions or adjusted analyses.

Prioritization of variants for follow-up and other secondary analyses

A GWAS meta-analysis may suggest variants to test in additional follow-up efforts. Prioritization of variants should follow rules that are pre-specified in the analysis plan. These typically include thresholds of statistical significance, but additional information may also be incorporated (for example, biological plausibility, evidence from other GWASs and other meta-analyses in the same field, text-mining-based methods or information on pathways[17]). The analysis plan should also specify how to choose among potentially multiple correlated variants based on linkage disequilibrium considerations.

When a variant passes the agreed level of genome-wide significance (typically set at $P < 5 \times 10^{-8}$), other secondary analyses might follow to assess the importance and the mode of action of the variant. For example, conditional analysis may be carried out for: other variants in the vicinity; adjustments for other traditional risk factors; gene–gene, gene–environment or protein–protein interactions; and diverse functional tests that may be carried out *in silico*, *in vitro* or *in vivo*[19].

Hardy–Weinberg equilibrium
A principle stating that the genetic variation in a population will remain constant from one generation to the next in the absence of disturbing factors.

Bayesian approaches
Fully probabilistic methods for describing models, parameters and data. They are so called because extensive use is made of Bayes' theorem to compute the probability distribution of model parameters given the experimental data.

Some new random effects methods have been proposed for improving discovery power in the presence of between-study heterogeneity in effect sizes. Han and Eskin[34] assume that there is no heterogeneity under the null hypothesis, in contrast to the traditional random effects models. Their statistic can be decomposed into two parts, one that is equal to the fixed effects statistic and another that is equal to the test statistic for heterogeneity. Further empirical validation of loci discovered by this method would be useful.

Optimal weights. Intuitively, it makes sense that studies with more data should count more (that is, have a larger weight) in the meta-analysis calculations, but the definition of weights depends on the statistical model used. The optimal weight for meta-analysis is the inverse variance weighting[35]. The favourable properties of such weighting in GWAS meta-analysis are well-known for initial screenings and discovery[28]. For imputed SNPs,

simulations show that the optimal weight is proportional to the inverse variance and the expected value of the effect estimate[36], but the inverse variance modelling scheme approximates the optimal weights well. Other weights derived from score statistics can have practical advantages when only *P* values and direction of effects have been provided, and they might be easier to interpret when there are differences with respect to covariates or definitions of the phenotypes that are modelled. For small effect sizes, the optimal weights from score statistics have essentially the same power as inverse variance weighting[37].

Bayesian meta-analysis. Some consortia have applied Bayesian approaches for GWAS meta-analyses. Specifically, the Wellcome Trust Case Control Consortium has used the Bayes factor[38], whereas the Coronary Artery Disease Consortium has presented results as posterior probabilities that a variant is null[39].

Table 1 | **Examples of high-profile consortia for various disease phenotypes**

Consortium (acronym)	Phenotype (or phenotypes)	Publicly available genome-wide data?	Website
AMD	Age-related macular degeneration	Yes, accessible through the website	http://www.sph.umich.edu/csg/abecasis/public/amdgene2012
BCAC	Breast cancer	No	http://ccge.medschl.cam.ac.uk/consortia/bcac
CHARGE	Heart disease and ageing	No	http://web.chargeconsortium.com
GEFOS	Osteoporosis	Yes, accessible through the website	http://www.gefos.org
GIANT	Anthropometric traits	Yes, accessible through the website	http://www.broadinstitute.org/collaboration/giant/index.php/GIANT_consortium
GLGC	TC, HDL-C, LDL-C, triglycerides	Yes, accessible through the website	http://www.sph.umich.edu/csg/abecasis/public/lipids2010
IIBDGC	Inflammatory bowel disease	Yes, accessible through the website	http://www.ibdgenetics.org
IMSGC	Multiple sclerosis	Yes, accessible through the website	https://www.imsgenetics.org/
ISC	Schizophrenia	No	http://pngu.mgh.harvard.edu/isc
MAGIC	Glycaemic traits	Yes, accessible through the website	http://www.magicinvestigators.org
NARAC-III	Rheumatoid arthritis	No	http://www.naracstudy.org/NaracStudy/narac.aspx
TREATOA	Osteoarthritis	Yes, accessible through the website	http://treatoa.eu
WTCCC	Various phenotypes	Yes, accessible through the website	http://www.wtccc.org.uk

HDL-C: high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; TC, total cholesterol.

Bayesian methods have also been developed to characterize the best fitting inheritance model for variants emerging from GWAS meta-analyses[40], to characterize the polygenic architecture of complex traits for which there are thousands of top SNPs[41] and for multivariate methods (see relevant section below). Bayesian models are straightforward and intuitive, and the models used (for example, fixed, random effects) can be similar to the models used in the frequentist approaches described above. However, Bayesian models may depend on assumptions made about the prior distributions of parameters of interest, and their genome-wide implementation can become computationally intensive.

Inferential tools and their interpretation

P *values and genome-wide significance thresholds.* The most common measure used for flagging a SNP as 'noteworthy' is the *P* value[42]. The selection of the most suitable genome-wide significance threshold should take into account the multiplicity of comparisons carried out. Some early GWASs used a threshold of $P \leq 10^{-7}$, but the current practice, in which more SNPs are actually typed and the imputation quality is higher, accepts a threshold of $P \leq 5 \times 10^{-8}$ as reaching genome-wide significance. This roughly corresponds to a simple Bonferroni correction to maintain a 5% genome-wide type I error rate that is based on an estimated burden of 1 million independent comparisons for common sequence variation[18,43]. However, a study has shown that associations with borderline genome-wide significance (that is, $P \geq 5 \times 10^{-8}$ and $P \leq 10^{-7}$) are successfully replicated 73% of the time when additional data are accumulated[44]. The most suitable genome-wide significance threshold may vary for different populations, SNPs, MAFs, linkage disequilibrium (LD) patterns, arrays

or different types of genetic data, but the differences are not major. For example, for populations with lower LD, such as Africans, stricter thresholds would be most appropriate[45].

Bayes factors. The Bayes factor[46] represents the ratio of the probability of the data under the null hypothesis, H_0, to the probability of data under the alternative hypothesis. Following the Bayes theorem, the posterior odds of $H_0 = $ Bayes factor × prior odds of H_0. Bayes factor and *P* values will often provide similar rankings for MAFs in the range 0.1–0.5 (REF. 47). However, differences can be observed for SNPs with low MAFs, which will be more often assessed as next-generation sequencing data become more commonly used in GWASs[47].

Q values (false discovery rate). *Q* values and false discovery rate (FDR)[48] are very popular in 'omics' fields such as gene expression profiling, in which multiplicity issues are prominent, but surprisingly they have rarely been used in GWASs and meta-analyses thereof. The FDR estimates the proportion of associations that are seemingly discovered but are nevertheless expected to be false positives. The *Q* value is the minimum FDR that can be attained when declaring an association. *Q* values can be calculated for a list of *P* values using publicly available software called QVALUE[48]. An efficient process for selecting top SNPs using false-discovery rates following by further screening with least absolute shrinkage and selection operator (LASSO) regression has been proposed[49].

Reliability of signals from imputed genotypes. Imputation can allow data to be combined from studies using different genotyping platforms. Imputation is

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Bonferroni correction
A method to counteract the problem of multiple comparisons. It is the simplest and most conservative approach to control for type I error.

Type I error
The probability of rejecting the null hypothesis when it is true. For genetic association studies, type I errors reflect false-positive findings of associations between allele or genotype and disease.

Linkage disequilibrium
The nonrandom association of alleles of different linked polymorphisms in a population.

Table 2 | **Comparison of meta-analysis software packages**

	METAL	GWAMA	MetABEL	PLINK	R packages
Ability to process files from GWAS analysis tools; software used	No	Yes; SNPTEST, PLINK	Yes; ABEL	Yes; PLINK	No
Fixed effects implemented?	Yes	Yes	Yes	Yes	Yes
Random effects implemented?	No	Yes	No	No	Yes
Heterogeneity metrics generated	Q, I^2	Q, I^2	Q, I^2	Q, I^2	Q, I^2
Graphical illustration of meta-analysis results	No	Manhattan and QQ plots	Forest plots	No	Yes

GWAS, genome-wide association study.

problematic for polymorphisms with low MAFs and those without at least modestly linked markers. Often, imputed variants with MAFs <5% (and certainly those with MAFs <1%) are disregarded and excluded from the analysis[11]. Imputation quality is usually measured by imputation accuracy. For most SNPs, imputation algorithms implemented in IMPUTE, MACH, MINIMAC or BEAGLE have very high imputation accuracy provided that the study population is similar to the reference population in terms of LD patterns and MAFs[50–52]. A metric to estimate the imputation quality is r^2, which is the ratio of the empirical variance of the imputed genotypes to the expected variance given the imputation estimate of the MAF. However, with low-frequency SNPs, the use of r^2 to evaluate the quality of the imputation can be misleading. Reliability of imputation in these cases can be assessed with imputation quality score (IQS)[53]. IQS takes chance agreement into account and thus controls for allele frequencies. Meta-analytical methods that take into account the errors in imputation by weighing not only in the inverse variance of each study but also in the accuracy of the imputed genotypes have been developed in each study; in some simulation studies, they may slightly outperform standard meta-analytical approaches[54], but this is not seen by all authors[37]. Challenges for imputations using panels from the 1000 Genomes Project are presented in BOX 2.

Dealing with heterogeneity

Phenotype-based heterogeneity. GWASs have been less successful for diseases in which phenotypes have been more difficult to define and to standardize, such as cognitive traits and mental-health-related diseases[55], behavioural traits[56] and osteoarthritis[57,58]. Evidence from other fields, such as obesity, also suggests that the establishment of associations may be dependent on phenotype definition[59] and that variability in definitions may cause heterogeneity in effect size or even spurious associations[60]. In some cases, harmonization of different phenotype definitions[61,62] is possible, whereas in other situations this may not be feasible: for example, if phenotypes have already been collected and it is not possible to go back and remeasure them. The process must balance the need to augment the sample size (to increase power for gene discovery) with the likelihood of increased heterogeneity, which dilutes the average genetic effect and thus leads to loss of

power[6]. Methods have been proposed to improve power in the presence of heterogeneous traits[63,64] but have not been extensively used.

Ancestry-based heterogeneity. Synthesizing data from populations of different ancestry may increase the observed heterogeneity. The agnostic GWAS approach usually captures common markers that are likely to be in linkage only with the functional or causative culprits; the most strongly associated SNP may not be the functional or the causative variant. An assessment of GWAS-discovered variants shows modest correlation in MAF of the variants between ancestries and different genetic effects in different ancestries[65]. However, consistency across different ancestries may be higher for some common diseases[66]. A proposed transethnic meta-analysis approach takes into account the similarity in allelic effects between the most closely related populations while allowing for heterogeneity between more diverse ethnic groups[67]; this approach may occasionally improve power to detect a novel association and localize causal variants[67].

Other sources of heterogeneity. There are several other sources that can introduce heterogeneity in meta-analysis of genetic data. Population stratification may exist even in populations that are assumed to be fairly homogeneous: for example, Europeans[68–70]. One study showed different stratification patterns in the UK population for rare and low-frequency variants[71]. Adjusting for principal components can handle heterogeneity for common variants, but additional methods are required to address stratification issues for rare variants[71]. Heterogeneity can also be introduced by using different genotyping platforms, imputation software or quality-control criteria across studies[33]. Gene–gene interactions and gene–environment interactions with differential non-genetic environmental exposures across different populations may also introduce heterogeneity — see below for a discussion of meta-analysis of joint estimation of interaction effects.

Finally, sex differences may induce heterogeneity, and some studies suggest differential genetic effects with respect to sex for many common variants[72], although this has not been borne out in other large-scale empirical studies[73]. Methods that optimize power for meta-analysis of sex-specific GWASs have been proposed[74].

Population stratification
The presence of several population subgroups that show limited interbreeding. When such subgroups differ both in allele frequency and in disease prevalence, this can lead to erroneous results in association studies.

Principal components
A composite variable that summarizes the variation across a larger number of variables, each represented by a column of a matrix.

Table 3 | Summary of methods for meta-analysis of genome-wide data

Method	Description	Advantages	Disadvantages	Main software used
P value meta-analysis	Simplest meta-analytical approach	Allows meta-analysis when effects are not available	Direction of effect is not always available; inability to provide effect sizes; difficulties in interpretation	METAL, GWAMA, R packages
Fixed effects	Synthesis of effect sizes. Between-study variance is assumed to be zero	Effects readily available through specialized software	Results may be biased if a large amount of heterogeneity exists	METAL, GWAMA, R packages
Random effects	Synthesis of effect sizes. Assumes that the individual studies estimate different effects	Generalizability of results	Power deserts in discovery efforts; may yield spuriously large summary effect estimates when there are selection biases	GWAMA, R packages
Bayesian approach	Incorporates prior assessment of the genetic effects	Most direct method for interpretation of results as posterior probabilities given the observed data	Methodologically challenging; GWAS-tailored routine software not available; subjective prior information used	R packages
Multivariate approaches	Incorporates the possible correlation between outcomes or genetic variants	Increased power can identify variants that conventional meta-analysis do not reveal using the same data sets	Computationally intensive; software not available for all analyses; some may require individual-level data	GCTA for multi-locus approaches
Other extensions	A set of different approaches that allows for the identification of multiple variants across different diseases	Summary results of previous meta-analyses can be used	May need additional exploratory analyses for the identification of variants; prone to systematic biases	Software developed by the authors of the proposed methodologies

GCTA, genome-wide complex trait analysis; GWAS, genome-wide association study.

Statistical metrics of heterogeneity. Typical heterogeneity metrics that are extensively used in GWAS meta-analyses are Cochran's Q statistic and I^2. Cochran's Q statistic follows a χ^2 distribution with $k–1$ degrees of freedom, where k is the number of studies and is typically considered to be significant at $\alpha = 0.10$ (where α is the type I error)[35]. However, the test is often underpowered when there are few studies (for example, <20), which is a common scenario in GWAS meta-analysis. The power of the Q statistic can be estimated[75], and it can be used in the estimation of the posterior odds of heterogeneity in a Bayesian framework[76]. Q is also used in the estimation of the between-study variance; the ratio of the effect size over τ (which is defined in BOX 3) offers an estimate of how big the effect is against the typical variability seen across studies and populations. Finally, I^2 quantifies the heterogeneity by measuring the amount of heterogeneity that is not due to chance[77]. It ranges from 0–100% and is considered low, moderate, large and very large for values 0–25%, 25–50%, 50–75% and >75%, respectively[78]. Statistical properties of heterogeneity metrics are presented in BOX 3.

Other extensions

Cross-phenotype meta-analysis. Cross-phenotype meta-analysis (CPMA) aims to detect multiple associations at a single marker across different diseases that may share a common genetic background or that involve a common biological process, such as autoimmunity[79]. CPMA assumes that the P values used for the individual traits come from different non-overlapping cohorts; as such, it cannot be applied in the case of large consortia that investigate many phenotypes but usually share the same control samples[79]. Modest overlap of the control samples

(<50%) is tolerable, but large overlaps erode the power of this statistic. CPMA has been applied in the study of immune-mediated and autoimmune diseases; of 107 previously identified variants across seven diseases, almost half were shared across diseases[79]. The CPMA statistic is agnostic to the direction of effect in each disease and can detect markers for at least some, but not necessarily all, phenotypes. It has one degree of freedom as it measures a deviation in P value behaviour instead of testing all possible combinations of diseases for association to each SNP, and therefore provides high power to reject the null hypothesis. However, power comes at the price of not knowing to which phenotypes the marker is associated, thus requiring further evidence.

Exploratory cross-phenotype checks. A common practice in many GWAS meta-analyses is to carry out cross-phenotype checks. Here, SNPs that have been identified as being significantly associated with one phenotype are also tested for association with other phenotypes in the same or other consortia. Typically, related phenotypes are examined: for example, SNPs that have been found to be associated with coronary artery disease may be specifically tested for association with phenotypes that represent diverse components of the metabolic syndrome[80]. Such checks could offer interesting insights about the pathophysiological paths implicated. The risk is that these analyses are exploratory, and thus their reporting may be less than comprehensive, potentially leading to selective reporting bias.

Joint meta-analysis for main and interaction effects. Main effects analysis may fail to identify additional susceptibility loci that interact with environmental factors

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Box 2 | Challenges for imputation using data from the 1000 Genomes Project

Genotype imputation has widely been used in meta-analysis of genome-wide association studies (GWASs). In the first generation of GWASs, most imputation analysis used panels from Phase 2 of the International HapMap project, which contained a total of 210 unrelated individuals with ancestry from West Africa, East Asia and Europe. The 1000 Genomes Project[15] has now extended this resource by applying whole-genome shotgun sequencing to an even larger number of individuals (2,500) sampled from around 25 different locations. Direct sequencing can identify variants that are not represented on the genotyping arrays that were used in the HapMap Project and therefore can include more reference panels with most of the new variants occurring at low population frequencies. However, this might create enormous databases that require more exhaustive computer power to be analysed.

For original single-nucleotide polymorphism (SNP) sets from the 1000 Genomes Project, the estimated imputation quality criteria should be treated with caution and should be more conservative, given that early haplotype sets were of lower quality. However, in mid-2012, the 1000 Genomes Project released a powerful reference panel known as 'Phase I v3'. Different approaches for imputing the panels are proposed from publicly available imputation software such as IMPUTE2, MACH and BEAGLE.

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if the genetic effects for distinct levels of an environmental exposure are not investigated. A method of joint analysis of SNP and SNP–environment interaction coefficients has been proposed using a multivariate approach that provides confidence intervals for the two estimates and a test of homogeneity across samples. This method performs better when both main and interaction effects are present[81], when joint meta-analysis may have a potential power gain of over 50% compared with classical meta-analysis[82]. Application of joint meta-analysis is hampered by a lack of sufficient environmental exposure data in most genetic data sets.

Meta-analysis of multiple correlated phenotypes. Consideration of several correlated phenotypes together in the same studies can boost power[83]. Different methods for meta-analyses of correlated phenotypes have been proposed[84,85]. The principle is that of bivariate meta-analysis, which is adapted for handling the correlation between outcomes[84]. The method can be applied both to binary and to continuous outcomes. One of the drawbacks is that individual-level data might be needed; however, methods for calculation of the covariance between two correlated effect sizes from summary data have also been proposed[85]. These approaches have not been applied yet in GWAS data sets, but they may offer the advantage of increased rigor to exploratory cross-phenotype checks within the same consortium.

Meta-analysis of multiple correlated variants. This multivariate approach is also known as multi-point, multi-locus, multi-marker or joint multiple-SNP analysis. The causal variant may not necessarily emerge as the top SNP in a GWAS meta-analysis, and there may be multiple causal variants at the same locus, each independently contributing to genetic association with the phenotype[86,87]. Conditional analysis has been used as a tool to detect secondary independent signals at a locus, but individual-level data are required, which are usually unavailable for large-scale meta-analysis of GWASs.

An approximate conditional approach using summary data from a meta-analysis and LD correlations between SNPs estimated from a reference sample (here, a subset of the meta-analysis sample) was successfully applied in a meta-analysis for height and body mass index[88]. This method identified 36 loci with multiple associated variants for height adding 49 additional SNPs on top of the already known variants. In this approach, a genome-wide stepwise selection procedure selects SNPs on the basis of conditional *P* values and estimates the joint effects of all selected SNPs after the model has been optimized. The method assumes that the reference sample is from the same population as the samples from which the genotype–phenotype associations are estimated (that is, that linkage correlation estimates in the reference sample are unbiased). Simulations show that *P* values from this approximate approach are consistent with those from conditional analysis using individual-level data[88]. A reference sample of at least 2,000 is required to estimate linkage correlations with little error.

Mendelian randomization in GWAS meta-analysis. Mendelian randomization entails the use of genetic variants as proxies ('instrumental variables') for modifiable or environmental exposures under investigation[89]. Meta-analysis methods have been proposed for Mendelian randomization studies[90]. Meta-analysis can help to bypass the problem that most genetic variants used as instrumental variables have small effects with large uncertainty in small studies, and thus they also leave large uncertainty about the presence or absence of causal relationships. For example, a large Mendelian randomization study based on a GWAS meta-analysis managed to demonstrate conclusively that high-density lipoprotein (HDL) is not causally related to coronary artery disease, as variants that affect specifically HDL levels do not have an impact on coronary artery disease risk[91].

Sequence data and low-frequency variants

Low event rates and zero counts. As discussed above, it has been common practice in GWAS meta-analyses to exclude low-frequency and rare variants. The main reason for these exclusions was related to imputation and genotyping quality, but this practice also worked as a safety net for avoiding false-positive or other spurious findings that would mostly be statistical artefacts. This situation is now changing as projects such as the 1000 Genomes Project[15] and the UK10K, along with the advent of next-generation sequencing technologies[92],

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Main effects
The effects of a variable assuming no dependency or conditionality of other variables.

Bivariate meta-analysis
Joint synthesis of two phenotypes by using their correlation.

Box 3 | Statistical properties of common GWAS meta-analysis approaches

The simplest genome-wide association study (GWAS) meta-analysis approach is to combine P values using Fisher's method. The formula for the statistic is

$$X^2 = -2\sum_{i=1}^{k}\log(P_i)$$

where P_i is the P value for the ith study, and k is the number of studies in the meta-analysis. Under the null hypothesis, X^2 follows a χ^2 distribution with $2k$ degrees of freedom. The Z scores meta-analysis can be implemented using the equation

$$Z = \frac{\sum_i Z_i w_i}{\sqrt{\sum_i w_i^2}}$$

where w_i is the square root of sample size of the ith study and

$$Z_i = \Phi^{-1}\left(1 - \frac{P_i}{2}\right)^*(\text{effect direction for study } i)$$

where Φ is the standard normal cumulative distribution function. For fixed effects models, inverse variance weighting is widely used. The weighted average of the effect sizes can be calculated as

$$\hat{\theta}_F = \frac{\sum_i w_i \hat{\theta}_i}{\sum_i w_i}$$

and the variance is

$$\text{var}(\hat{\theta}_F) = \frac{1}{\sum_i w_i}$$

where $\hat{\theta}_i$ is the ith study normalized effect (for example, logarithm of odds ratio or β-coefficient for a logistic regression for a binary phenotype or mean difference or standardized mean difference for a continuous phenotype), and w_i is the reciprocal of the estimated variance of the effect study. The random effects model incorporates the between-study variance of heterogeneity, and therefore the weight for the random effects model is calculated as

$$w_i^R = \frac{1}{\left(\frac{1}{w_i} + \hat{\tau}^2\right)}$$

where

$$\tau^2 = \frac{(Q - (k-1))}{\left(\sum_i w_i - \left(\frac{\sum_i w_i^2}{\sum_i w_i}\right)\right)}$$

and Q is Cochran's Q statistic, which is given by

$$Q = \sum_i w_i (\hat{\theta}_i - \hat{\theta}_F).$$

Another popular heterogeneity metric, I^2, is given by

$$I^2 = \frac{100^*(Q - (k-1))}{Q}.$$

The multivariate meta-analysis approaches are based on the calculation of a variance–covariance matrix for the correlated phenotypes or the single-nucleotide polymorphisms in linkage disequilibrium that will allow the calculation of the marginal effects. In cross-phenotype meta-analysis, the developed statistic measures the likelihood of the null hypothesis, given the data. The test is asymptotically distributed as

$$\chi^2_{df=1}.$$

will facilitate the study of low-frequency and rare variants. This translates, however, to larger sample size requirements to robustly identify associations involving variants in these classes that have modest effects. The problem is magnified for uncommon phenotypes. Asymptotic assumptions may not hold for rare variants or even low-frequency ones. Zero counts in 2×2 tables may become a common scenario. Typically used solutions use continuity corrections: for example, adding 0.5 to all four cells of the 2×2 table. However, most meta-analysis methods, including inverse variance fixed effects, DerSimonian and Laird random effects, and Mantel–Haenszel, perform poorly with such corrections to zero counts. The Peto method seems the least biased[93], but this method is also problematic when the two groups that are compared are unequal in terms of observed events and sample sizes. Other approaches besides continuity corrections may be considered (for example, arcsine transformation[94]), but there is no empirical evidence yet that relates to their application in genetics. Mega-analysis (which is the pooling of all individual-level data from all studies into a common single data set; see below) may also be useful, but in this case, analyses need to account carefully for ancestry and other sources of potential heterogeneity (as discussed above).

Merging rare variation. Single-point analysis of low-frequency and rare variants requires hundreds of thousands of individuals in order to detect weak effect sizes. Several approaches for the aggregate analysis of multiple low-frequency or rare variants across a locus that may exhibit heterogeneity have been proposed[95–102]. Locus-based methods that incorporate variant quality scores available from next-generation sequencing data using a regression-based collapsing approach and an allele-matching method have been also proposed[103]. These quality-incorporating approaches are more powerful than their unweighted counterparts, when the causal variants are of high quality[103]. A sequencing kernel association test (SKAT) seems to outperform previously proposed collapsing methods[104] when a large fraction of the variants in a region are non-causal or the effects of causal variants are in different directions. Collapsing methods seem to be more powerful when most variants in a region are causal, and the effects are in the same direction. Of course, further studies are needed to address power issues, given that few true-positive associations with rare variants have been identified so far. SKAT-O is another interesting unified approach with application in small case–control sequencing studies; it maximizes power by adaptively using the data to combine optimally the burden test and non-burden sequence kernel tests[105]. Besides the known approaches for single-variant tests, such as the typical logistic Wald test, other exact approaches could be applied, such as the Firth-bias-corrected likelihood ratio test[106].

Most of the methods described above provide a P value or a test statistic as a result for a locus-based association, and these can easily be combined in a meta-analysis when more than one data set is available, as described above. Software has been developed for

Glossary

Asymptotic assumptions
When the sample size in a data set grows indefinitely, then the distribution of the estimators becomes approximately normal.

2 × 2 tables
A 2 × 2 table that describes the cross-classification of data that are divided into two groups with two categories in each.

Collapsing approach
Statistical methods for association analysis in which multiple low-frequency or rare variants are collapsing into a single locus.

Lambda inflation factor
A metric used in genetic association studies to correct for spurious associations (which may arise owing to population stratification) by estimating the extent of inflation in the statistical evidence and appropriately down-weighting this inflation.

the meta-analysis of rare variants (such as MetaSKAT, SKAT-O and Rare-METAL). However, it has been shown that meta-analyses based on *P* values or test statistics for rare variants lose important information. To retain full efficiency, the test can be reformulated using summary statistics from each cohort on both the per-variant score test and the genotype covariance matrix[107]. The individual-variant test statistics are routinely shared for meta-analysis in GWASs anyhow, and the genotype covariance matrix contains no information about genotype–phenotype associations. Thus, data sharing for these quantities should be no more difficult to arrange than in GWASs.

Another important issue to consider is that for rare variants, the identity of the carriers and their privacy may be jeopardized. A method has been developed (MetaSeq) for conducting the meta-analysis in which privacy is maintained through cryptographic means[108].

Genome-wide significance thresholds for sequencing. With the inclusion of low-frequency and rare variants from the 1000 Genomes Project imputations and sequencing efforts, in which up to ~50 million markers may be typed, the number of the independent loci under study will be significantly larger than the 1 million markers estimated for GWASs. Therefore, it may be argued that an even more stringent threshold, compared to the typical 5×10^{-8}, may be required to secure robust findings that will be considered replicated. Additional studies are needed to indicate the optimal threshold. This threshold may depend on sample size, asymptotical properties of the tests, number of low-frequency variants known and other factors.

Issues related to data sources

Publicly available data. The benefits of data sharing are obvious. The National Center for Biotechnology Information has created a public repository (called dbGaP) for individual-level phenotype, exposure, genotype and sequence data and the associations between them[109]. Moreover, several funders in Europe and the United States promote sharing of summary data from already published GWASs or meta-analyses of GWASs. This facilitates a wide range of secondary research, both methodological and applied, by investigators who can access these data. There is evidence that participants are in favour of providing a re-consent for submission of their data to dbGaP[110].

Meta-analyses of meta-analyses. For some phenotypes of interest, several independent or partially overlapping consortia may exist, such as those that are funded by different mechanisms and that are assembled by different initiatives. The successful cooperation between different consortia will allow for mega-analyses, which can include both previously published data and data from meta-analyses that are conducted prospectively. This will significantly increase the power to detect weak signals, especially for low-frequency and rare variants. Consortia studying anthropometric traits such as body mass index have now accumulated almost a quarter of a million participants[111], and this number is expected to increase markedly in the near future as it has been shown that meta-analysis sample size requirements increase steeply when small genetic effects are considered[4]. Big data in terms of sample sizes may be merged from existing and emerging research teams, consortia, biobanks and other population-level efforts.

The impressive increase in the numbers of included participants may result in an artificial increase of the lambda inflation factor, in many cases with values >2, thus reducing the novel variants assessed in the discovery effort if a genomic control for the observed value is taken in account. Approaches that correct for this inflated lambda and that are based on mixed models are being developed, such as EMMAX[112]. Furthermore, in such efforts, the researchers should always be cautious when they combine data with different sequencing depths and from different platforms. Given that expectations for finding many low-frequency and rare variants with large effects have not been validated in sequencing studies to date, meta-analytical methods will probably continue to be the key approach taken towards exploring further a genetic architecture that is characterized mostly by small effects.

Conclusions and future directions

Multiple methods have been developed for the meta-analysis of available genome-wide data improving our understanding of complex diseases and explaining part of the missing heritability that we were not able to identify using simple association metrics. Merging of existing research teams and consortia may further allow for the identification of novel variants and represent a significant advance in our understanding of genetic susceptibility. iGOGS, a recent example of such a collaborative effort including five consortia studying hormone-mediated cancers, revealed 74 new susceptibility loci for these cancers[113–117]. Such multi-consortium efforts may become a necessity in the future to identify more low-frequency and rare variants with small or even moderate effects. More empirical and simulation studies are needed to assess the advantages and disadvantages of some of the proposed techniques that were presented here and to delineate their optimal application and interpretation of the results that they produce.

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FURTHER INFORMATION
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dbGaP: http://www.ncbi.nlm.nih.gov/gap
GWAMA: http://www.well.ox.ac.uk/gwama
IMPUTE2: http://mathgen.stats.ox.ac.uk/impute/impute_v2.html
metABEL: http://www.genabel.org/packages/MetABEL
METAL: http://www.sph.umich.edu/csg/abecasis/metal
metaSKAT/SKAT-O: http://people.hsph.harvard.edu/~sglee/MetaSKAT
PLINK: http://pngu.mgh.harvard.edu/~purcell/plink
QVALUE: http://genomine.org/qvalue
Rare-METAL: http://genome.sph.umich.edu/wiki/Rare-Metal
SKAT: http://www.bios.unc.edu/~mcwu/software/index.html#SKAT
UK10K: http://www.uk10k.org
Wellcome Trust Case Control Consortium: http://www.wtccc.org.uk

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