From aCIBER de Enfermedades Respiratorias, Instituto de Salud Carlos III, Madrid, Spain; bthe Research Unit, Hospital Universitario N.S. de Candelaria, Tenerife, Spain; ^cthe Multidisciplinary Organ Dysfunction Evaluation Research Network (MOD-ERN), Research Unit, and ^fthe Allergy Unit, Hospital Universitario Dr. Negrin, Gran Canaria, Spain; ^dthe Department of Medicine, University of California, San Francisco, Calif: ethe Section of Pulmonary and Critical Care Medicine, University of Chicago, Chicago, Ill; gthe Allergy Unit, Hospital Universitario N.S. de Candelaria, Tenerife, Spain; hU.G.C. Allergy, Regional University Hospital of Málaga-IBIMA, Málaga, Spain; ⁱGrupo de Medicina Xenómica, CEGEN-ISCIII-Universidade de Santiago de Compostela, Santiago de Compostela, Spain; ^jGrupo de Medicina Xenómica, CIBERER-Universidade de Santiago de Compostela-Fundación Galega de Medicina Xenómica (SERGAS), Santiago de Compostela, Spain; kthe Arizona Health Sciences Center, University of Arizona, Tucson, Ariz; ¹the Department of Bioengineering and Therapeutic Sciences, University of California, San Francisco, Calif; and mthe Applied Genomics Group (G2A), Genetics Laboratory, Instituto Universitario de Enfermedades Tropicales y Salud Pública de Canarias, Universidad de La Laguna, Tenerife, Spain. E-mail: cflores@ull.edu.es.

*These authors contributed equally to this work.

Supported by Instituto de Salud Carlos III (FIS PI11/00623) and cofinanced by the European Regional Development Funds, "A way of making Europe" from the European Union; by the University of Chicago Core Subsidy Mini Award (ITM/CTSA UL1 RR024999); and by a grant from the 7th Framework Programme (FP7-REGPOT-2012-CT2012-31637-IMBRAIN). This work was also supported by grants from the National Institutes of Health (to E.G.B.): the National Heart, Lung, and Blood Institute (RC2 HL101651, HL088133, HL078885, HL004464, HL104608, and HL117004); the National Institute of Environmental Health Sciences (ES015794); the National Institute on Minority Health and Health Disparities (MD006902); and the National Institute of General Medical Sciences (GM007546). E.G.B. was also funded by the American Asthma Foundation, the RWJF Amos Medical Faculty Development Award, the Sandler Foundation, and the Flight Attendant Medical Research Institute. M.P.-Y. was supported by a postdoctoral fellowship from Fundación Ramón Areces (http://www.fundacionareces.es). M.A.-H. and A.B.-L. were supported by fellowships from the Instituto de Salud Carlos III (FI11/00074 and FI12/00493, respectively).

Disclosure of potential conflict of interest: M. Pino-Yanes has received payment for lectures from Affymetrix. J. Villar has received research support from MAQUET. A. Carracedo is employed by the University of Santiago and has received research support from the Ministry of Justice, the Ministry of Health, and the European Union. E. G. Burchard has received research support from the National Institutes of Health (the National Heart, Lung, and Blood Institute; National Institute of Environmental Health Sciences; and National Institute on Minority Health and Health Disparities), the American Asthma Foundation, the Sandler Foundation, and the Flight Attendant Medical Research Institute. C. Flores has received research support from Instituto de Salud Carlos III (Grant FIS PI11/00623). The rest of the authors declare that they have no relevant conflicts of interest.

REFERENCES

- Benayoun L, Druilhe A, Dombret MC, Aubier M, Pretolani M. Airway structural alterations selectively associated with severe asthma. Am J Respir Crit Care Med 2003;167:1360-8.
- Flores C, Ma SF, Maresso K, Ober C, Garcia JG. A variant of the myosin light chain kinase gene is associated with severe asthma in African Americans. Genet Epidemiol 2007;31:296-305.
- Gao L, Grant AV, Rafaels N, Stockton-Porter M, Watkins T, Gao P, et al. Polymorphisms in the myosin light chain kinase gene that confer risk of severe sepsis are associated with a lower risk of asthma. J Allergy Clin Immunol 2007; 119:1111-8.
- Galanter JM, Gignoux CR, Torgerson DG, Roth LA, Eng C, Oh SS, et al. Genome-wide association study and admixture mapping identify different asthma-associated loci in Latinos: the Genes-environments & Admixture in Latino Americans study. J Allergy Clin Immunol 2014;134:295-305.
- Torgerson DG, Gignoux CR, Galanter JM, Drake KA, Roth LA, Eng C, et al. Case-control admixture mapping in Latino populations enriches for known asthma-associated genes. J Allergy Clin Immunol 2012;130:76-82.e12.
- Pino-Yanes M, Corrales A, Acosta-Herrera M, Perez-Rodriguez E, Cumplido J, Campo P, et al. HLA-DRB1*15:01 allele protects from asthma susceptibility. J Allergy Clin Immunol 2014;134:1201-3.
- Garcia JG, Lazar V, Gilbert-McClain LI, Gallagher PJ, Verin AD. Myosin light chain kinase in endothelium: molecular cloning and regulation. Am J Respir Cell Mol Biol 1997;16:489-94.
- Moffatt MF, Gut IG, Demenais F, Strachan DP, Bouzigon E, Heath S, et al. A large-scale, consortium-based genomewide association study of asthma. N Engl J Med 2010;363:1211-21.

- Torgerson DG, Ampleford EJ, Chiu GY, Gauderman WJ, Gignoux CR, Graves PE, et al. Meta-analysis of genome-wide association studies of asthma in ethnically diverse North American populations. Nat Genet 2011;43:887-92.
- Bonnelykke K, Sleiman P, Nielsen K, Kreiner-Moller E, Mercader JM, Belgrave D, et al. A genome-wide association study identifies CDHR3 as a susceptibility locus for early childhood asthma with severe exacerbations. Nat Genet 2014;46:51-5.
- Botigue LR, Henn BM, Gravel S, Maples BK, Gignoux CR, Corona E, et al. Gene flow from North Africa contributes to differential human genetic diversity in southern Europe. Proc Natl Acad Sci U S A 2013;110:11791-6.

Available online May 27, 2015. http://dx.doi.org/10.1016/j.jaci.2015.04.025

Do human rhinovirus infections and food allergy modify grass pollen-induced asthma hospital admissions in children?

To the Editor:

Asthma prevalence in children has remained relatively constant in many Western countries, but hospital admissions for younger age groups have increased over time. Although the role of outdoor aeroallergens as triggers for asthma exacerbations requiring hospitalization in children and adolescents is complex, there is evidence that increasing concentrations of grass pollen are associated with an increased risk of asthma exacerbations in children. Human rhinovirus (HRV) infections are implicated in most of the serious asthma exacerbations in school-age children. In previous research, HRV infections and aeroallergen exposure have usually been studied independently. To our knowledge, only 1 study has examined interactions between these 2 factors, but lack of power prevented any meaningful interpretation.

Furthermore, although sensitization to aeroallergens is an important risk factor for the development of asthma in children,⁵ little is known about the role of allergic sensitization to food and the risk of asthma admissions.⁶ The aims of this study were to assess the effect of outdoor grass pollen levels on the incidence of asthma admissions in children and to assess whether the presence of HRV and allergic sensitization modified this effect.

Data from the Melbourne Air Pollen Children and Adolescent Health study (see this article's Methods section in the Online Repository at www.jacionline.org^{E1}) were analyzed. A total of 644 children and adolescents aged between 2 and 17 years with a principal diagnosis of asthma were enrolled between September

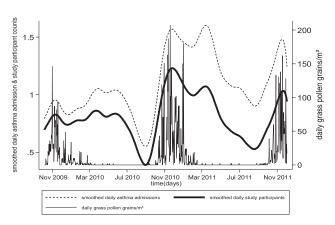


FIG 1. The total numbers of asthma admissions to the Royal Children's Hospital and participants enrolled in the Melbourne Air Pollen Children and Adolescent Health study (smoothed) and the seasonal distribution (in days) of ambient grass pollen during the study period.

TABLE I. Adjusted ORs for a per 50 grains/m³ increase in grass pollen, stratified by HRV and sensitization to pollen and food

	AII (N = 644)	Boys (N = 407; 63%)	Girls (N = 237; 37%)
Grass pollen	1.11 (1.00-1.22)*	1.22 (1.05-1.35)*	1.00 (0.84-1.21)
HRV infection status			
No HRV infection	1.04 (0.61-1.28)	1.11 (0.61-1.42)	0.89 (0.60-1.34)
HRV infection	1.22 (1.02-1.57)*	1.42 (1.11-1.64)*	0.58 (0.26-1.09)
Pollen sensitization			
Not sensitized to pollen	1.11 (0.61-1.28)	1.16 (0.61-1.42)	1.02 (0.77-1.36)
Sensitized to grass pollen	1.16 (1.02-1.28)*	1.22 (1.11-1.42)*	1.01 (0.77-1.32)
Food sensitization			
Not sensitized to food	1.11 (0.61-1.22)	1.22 (1.11-1.42)*	0.87 (0.71-1.05)
Sensitized to food	1.07 (0.83-1.39)	0.94 (0.68-1.30)	1.52 (1.03-2.24)*

Adjusted for age, HRV infection status (when not stratified by), pollutants, relative humidity, rainfall, and temperature. Adjusted models included only those variables that were significant at the 5% level. Values represent OR (95% CI).

2009 and December 2011. The design was case-crossover. The case day was defined as the day of admission. The control period was the same day of the week as the case day in all weeks within the same month of the same year in which the case day occurred. At admission, respiratory viral infections and skin prick test response to common allergens were tested. Daily ambient concentrations of grass pollen and air pollutants and weather data were available during the study period.

Population-averaged conditional regression models with robust standard errors were used to investigate the association between grass pollen and asthma admissions. E2 The primary exposure variable was daily concentrations of grass pollen fitted as a continuous variable. HRV infection status at admission, sex, and positive skin prick test result for allergens were considered as effect modifiers, and stratified analysis has been presented in tables. All models of the association between grass pollen and hospital admissions were adjusted for age, air pollutants, and weather variables. Potential confounders were retained if they changed the estimated associations between pollen exposure and the outcome by 10% or more, or were significant at the 5% level in adjusted models. Results are presented as odds ratios (ORs) with 95% CIs; these can be interpreted as an increment of 50 grains/m³ in airborne grass pollen (defined as high pollen days). Analyses were performed using Stata, release 10.1 (StataCorp, College Station, Tex).

Of 644 participants in the Melbourne Air Pollen Children and Adolescent Health study, 407 (63%) were male and the median (range) age was 5.2 years (2-17 years). There were 249 patients (39%) admitted during the peak grass pollen season, which occurred between October and December in Melbourne, Australia (Fig 1). Of this group, 164 were male, similar in age to girls, and showed comparable infection rates with HRV and other viruses. Although the rate of sensitization to food was similar in boys and girls, the rate of sensitization to grass pollen differed, with 47% in boys and 37% in girls (see Table E1 in this article's Online Repository at www.jacionline.org).

Presence of grass pollen in the atmosphere was significantly associated with an increased risk of admission in an adjusted model (Table I; OR = 1.11; 95% CI, 1.003-1.22). High levels of airborne grass pollen were associated with increased admission in boys (OR = 1.22; 95% CI, 1.05-1.35) but not in girls (OR = 1.00; 95% CI, 0.84-1.21). When stratified by HRV infection status, daily exposure to 50 grains/m³ of grass pollen or more increased admission only in boys with HRV infection (OR = 1.42; 95% CI, 1.11-1.64). Grass pollen exposure was associated with increased

admission among girls who were sensitized only to food (OR = 1.52; 95% CI, 1.03-2.24).

It is known that boys have higher risks of asthma exacerbations requiring emergency treatment or admission, particularly before puberty. Sex differences in lung physiology may partly contribute to early life increase in asthma admission for boys. Hitherto, the risk of asthma exacerbations requiring hospitalization associated with pollen exposure and cosensitization to food and aeroallergens was largely unknown.

Our results indicate that the combination of grass pollen exposure and HRV infection increases asthma exacerbations in boys. The reasons need further exploration because these factors are presumably acting through different biological pathways. Murray et al⁴ previously reported an increased risk for admission following a combination of allergen exposure and rhinovirus infection, although they did not separately report results for grass pollen exposure and rhinovirus. The combination of these 2 triggers particularly in boys needs further exploration. The "two-hit hypothesis" is one such possible mechanism where viral infections combined with atopy in early life may increase asthma exacerbations in children.

Our findings further indicate that food sensitization presents an increased risk of admission related to pollen exposure in girls, and highlights the value of identifying concomitant food sensitization/ allergy in children with asthma. Significant effects of grass pollen exposure in girls sensitized to food could suggest a more severe atopic phenotype, rather than cross-reactivity to allergens as was previously thought. ^{10,11} Additional studies are required to confirm our findings and investigate mechanisms.

A major strength of our study is the case-crossover design, which was chosen to address the hypothesis of pollen exposure, cosensitization, and the presence of respiratory viral infection. One potential limitation should be considered when interpreting the results: pollen measurements were performed at only 1 outdoor site, and it is possible that the counts did not reflect airborne pollen levels across the city. However, in previous studies, the exposures to grass pollen were similar for residents across Melbourne.²

In summary, boys with HRV infection at admission on days with high concentrations of outdoor grass pollen are at risk of asthma exacerbations requiring hospitalization. In contrast, girls sensitized to food were at an increased risk of admission on high pollen days. With climatic conditions continuing to have an impact on the duration and intensity of the pollen season coupled with likely increases in allergen exposures, ¹² it is imperative to

1120 LETTERS TO THE EDITOR

J ALLERGY CLIN IMMUNOL

OCTOBER 2015

better understand the relationship between pollen exposure and risk for adverse respiratory health outcomes. Interventions to reduce pollen exposure to better manage asthma and allergies may reduce the potential for these factors to interact, and can help to prevent serious asthma exacerbations in children and adolescents.

HRV identification was performed by the Victorian Infectious Diseases Reference Laboratory, Melbourne, Australia.

Bircan Erbas, PhD^a
Shyamali C. Dharmage, MD, PhD^b
Mimi L. K. Tang, PhD^{c,d,e}
Muhammad Akram, PhD^f
Katrina J. Allen, PhD^g
Don Vicendese, BSc(Hons)^a
Janet M. Davies, PhD^{h,i}
Rob J. Hyndman, PhD^f
Ed J. Newbigin, PhD^k
Philip E. Taylor, PhD^f
Philip G. Bardin, MD^m
Michael J. Abramson, PhD^f

From athe School of Public Health and Human Biosciences, La Trobe University, bthe Centre for Molecular, Environmental, Genetic and Analytic Epidemiology, School of Population Health, the University of Melbourne, the Department of Allergy & Immunology, Royal Children's Hospital, the Department of Allergy and Immune Disorders, Murdoch Children's Research Institute, the Department of Paediatrics, University of Melbourne, the Department of Epidemiology and Preventive Medicine, School of Public Health and Preventive Medicine, Monash University, and the Department of Gastro and Food Allergy, Murdoch Children's Research Institute, The Royal Children's Hospital, Melbourne, Australia; the School of Medicine and the Translational Research Institute, the University of Queensland, Brisbane, Australia; and the Department of Econometrics and Business Statistics, Monash University, the School of Botany, the University of Melbourne, the School of Life and Environmental Sciences, Deakin University, and the Department of Respiratory and Sleep Medicine, Monash Medical Centre, Melbourne, Australia. E-mail: berbas@latrobe.edu.au.

The Melbourne Air Pollen Children and Adolescent Health study and S.C.D. were funded by the National Health and Medical Research Council.

Disclosure of potential conflict of interest: B. Erbas, D. Vicendese, and E. J. Newbigin have received research support from the National Health and Medical Research Council (NHMRC) (project grant ID 541934). M. L. K. Tang has received research support from the NHMRC (541934), has received payment for lectures from Nestle Nutrition Institute and Nutricia; and has received travel support from the World Allergy Organization. K. J. Allen has received consulting fees from Pfizer, Abbott, Nutricia, Aspencare, Alphafarm, Wyeth, Danone, and Nestle and is a board member for Ilhan Food Allergy Foundation. J. M. Davies has received research support from the University of Queensland Collaborative Industry Engagement Fund, the NHMRC (1043311 and 1017441), the Asthma Foundation of Queensland, and the Australian Society for Clinical Immunology and Allergy; has consultant arrangements with Stallergenes Australia; is employed by the University of Queensland; has received payment for lectures from Stallergenes Pty Ltd and GlaxoSmithKline; has received travel support from Thermofisher; is a named inventor on a patent granted in Australia (2008316301) and applied for in the United States (12/738618) but no funds have been received from this: "Novel immunogenic molecules and uses thereof: Immunogenic protein Pas n 1 from Bahia grass pollen"; receives in kind support for the development of her research from Thermofisher (Uppsala, Sweden) by way of provision of materials; and receives in kind support for the development of her research from Sullivan NIcolaides Pathology (Taringa, Australia) by way of provision of pathology services. M. J. Abramson has received research support from the NHMRC (project grant ID 541934), Pfizer, and Boehringer Ingelheim; has consultant arrangements with AstraZeneca; has received payment for lectures from Novartis; and has received travel support from Boehringer Ingelheim and Sanofi. The rest of the authors declare that they have no relevant conflicts of interest.

REFERENCES

- Vicendese D, Olenko A, Dharmage SC, Tang MLK, Abramson MJ, Erbas B. Modelling and predicting low count child asthma hospital readmissions using General Additive Models. Open J Epidemiol 2013;3:125-34.
- Erbas B, Akram M, Dharmage SC, Tham R, Dennekamp M, Newbigin E, et al. The role of seasonal grass pollen on childhood asthma emergency department presentations. Clin Exp Allergy 2012;42:799-805.

- Rawlinson WD, Waliuzzaman Z, Carter IW, Belessis YC, Gilbert KM, Morton JR, et al. Asthma exacerbations in children associated with rhinovirus but not human metapneumovirus infection. J Infect Dis 2003;187:1314-8.
- Murray CS, Poletti G, Kebadze T, Morris J, Woodcock A, Johnston SL, et al. Study
 of modifiable risk factors for asthma exacerbations: virus infection and allergen
 exposure increase the risk of asthma hospital admissions in children. Thorax
 2006;61:376-82.
- Guilbert TW, Morgan WJ, Zeiger RS, Bacharier LB, Boehmer SJ, Krawiec M, et al. Atopic characteristics of children with recurrent wheezing at high risk for the development of childhood asthma. J Allergy Clin Immunol 2004;114:1282-7.
- Liu AH, Jaramillo R, Sicherer SH, Wood RA, Bock SA, Burks AW, et al. National prevalence and risk factors for food allergy and relationship to asthma: results from the National Health and Nutrition Examination Survey 2005-2006. J Allergy Clin Immunol 2010;126:798-806.e13.
- Sears MR. Epidemiology of asthma exacerbations. J Allergy Clin Immunol 2008; 122:662-8.
- Turner DJ, Stick SM, Lesouëf KL, Sly PD, Lesouëf PN. A new technique to generate and assess forced expiration from raised lung volume in infants. Am J Respir Crit Care Med 1995:151:1441-50.
- Sly PD, Boner AL, Björksten B, Bush A, Custovic A, Eigenmann PA, et al. Early identification of atopy in the prediction of persistent asthma in children. Lancet 2008;372:1100-6.
- de Jong AB, Dikkeschei LD, Brand PL. Sensitization patterns to food and inhalant allergens in childhood: a comparison of nonsensitized, monosensitized, and polysensitized children. Pediatr Allergy Immunol 2011;22:166-71.
- Ghunaim N, Wickman M, Almqvist C, Söderström L, Ahlstedt S, van Hage M. Sensitization to different pollens and allergic disease in 4-year-old Swedish children. Clin Exp Allergy 2006;36:722-7.
- Albertine JM, Manning WJ, DaCosta M, Stinson KA, Muilenberg ML, Rogers CA. Projected carbon dioxide to increase grass pollen and allergen exposure despite higher ozone levels. PLoS One 2014;9:e111712.

Available online June 3, 2015. http://dx.doi.org/10.1016/j.jaci.2015.04.030

Alternate methods of nasal epithelial cell sampling for airway genomic studies

To the Editor:

Recent translational studies of airway inflammation have shown that nasal epithelial cells are a good surrogate for bronchial epithelial cells^{1,2} in asthmatic patients.^{3,4} However, the standard method of nasal sampling requires use of a nasal speculum and specialized training. In pediatric studies requiring longitudinal specimen collection, sampling by this method can be limited by subject refusal and technical challenges.

Alternate methods of nasal sampling have been proposed. Different instruments for collection have been used, ranging from polyester-tipped swabs to plastic curettes to cytology brushes. Different sampling locations have been proposed, such as beneath the inferior turbinate or the anterior nares, where respiratory epithelial cells are also located. Obtaining nasal epithelial cells beneath the inferior turbinate with a cytology brush has been the most commonly used method. These cells have been validated as a surrogate for bronchial epithelial cells and have been shown to be clinically important in translational asthma studies. This method has also been shown in preliminary studies to be more difficult to tolerate. Whether a more comfortable method of sampling exists and whether this method can provide equivalent cytologic, gene expression, and epigenetic results is undetermined.

Here we compared nasal epithelial cells sampled from the anterior nares with either a polyester swab or a cytology brush with the standard collection method of cytology brush sampling from beneath the inferior turbinate. The benefit of the former method is that it does not require the use of a speculum to visualize nasal anatomy, is technically easy to perform, and, with

REFERENCES

- E1. Erbas B, Dharmage SC, O'Sullivan M, Akram M, Newbigin E, Taylor P, et al. A case-crossover design to examine the role of aeroallergens and respiratory viruses on childhood asthma exacerbations requiring hospitalization: the MAPCAH study. J Biomet Biostat 2012;S7-018.
- E2. Navidi W, Weinhandl E. Risk set sampling for case-crossover designs. Epidemiology 2002;13:100-5.

TABLE E1. Selected characteristics of Melbourne Air Pollen Children and Adolescent Health study participants

Variable	N = 644 (%)	Boys, n (%)	Girls, n (%)	P value
Sex		407 (63)	237 (37)	<.00005
Age (y), mean ± SD	5.2 ± 3.3	5.0 ± 3.2	5.5 ± 3.5	.11
Age (y), categories				
2-5	416 (65)	273 (66)	143 (34)	<.00005
6-12	201 (31)	119 (59)	82 (41)	.01
13+	27 (4)	15 (56)	12 (44)	.56
Presence of respiratory	N = 642			
virus at admission				
HRV	447 (70)	292 (72)	155 (66)	.12
Other viruses	16 (2)	10(2)	6 (3)	.94
Allergen sensitization (wheal size ≥3 mm)	N = 643	N = 406		
Grass pollen	269 (42)	181 (45)	88 (37)	.07
Any pollen	311 (48)	206 (51)	105 (44)	.12
7 1	N = 640	N = 405	` ′	
Food	145 (23)	87 (21)	58 (25)	.35
	N = 642	N = 406		
Cat	179 (28)	118 (29)	57 (24)	.18