

# Personalized Biopsy Schedules for Men With Low-Risk Prostate Cancer Under Active Surveillance\*

Anirudh Tomer, MSc<sup>a,\*</sup>, Daan Nieboer, MSc<sup>b,c</sup>, Monique J. Roobol, PhD<sup>c</sup>, Anders Bjartell, MD, PhD<sup>d</sup>, Ewout W. Steyerberg, PhD<sup>b,e</sup>, Dimitris Rizopoulos, PhD<sup>a</sup>, Movember Foundation's Global Action Plan Prostate Cancer Active Surveillance (GAP3) consortium<sup>f</sup>

<sup>a</sup>*Department of Biostatistics, Erasmus University Medical Center, Rotterdam, the Netherlands*

<sup>b</sup>*Department of Public Health, Erasmus University Medical Center, Rotterdam, the Netherlands*

<sup>c</sup>*Department of Urology, Erasmus University Medical Center, Rotterdam, the Netherlands*

<sup>d</sup>*Department of Urology, Skåne University Hospital, Malmö, Sweden*

<sup>e</sup>*Department of Biomedical Data Sciences, Leiden University Medical Center, Leiden, the Netherlands*

<sup>f</sup>*The Movember Foundation's Global Action Plan Prostate Cancer Active Surveillance (GAP3) consortium members presented in Appendix A*

---

## Abstract

**Background:** Prostate cancer active surveillance (AS) patients undergo repeat biopsies. Active treatment is advised when biopsy Gleason grade group  $\geq 2$  (*upgrading*). Many patients never experience upgrading, yet undergo biopsies frequently. Personalized biopsy decisions based on upgrading-risk may reduce patient burden.

---

\*Word count: abstract (headings excluded) 300; Text 2488; Total 2788

\*Corresponding author (Anirudh Tomer): Erasmus MC, kamer flex Na-2823, PO Box 2040, 3000 CA Rotterdam, the Netherlands. Tel: +31 10 70 43393

*Email addresses:* [a.tomer@erasmusmc.nl](mailto:a.tomer@erasmusmc.nl) (Anirudh Tomer, MSc), [d.nieboer@erasmusmc.nl](mailto:d.nieboer@erasmusmc.nl) (Daan Nieboer, MSc), [m.roobol@erasmusmc.nl](mailto:m.roobol@erasmusmc.nl) (Monique J. Roobol, PhD), [anders.bjartell@med.lu.se](mailto:anders.bjartell@med.lu.se) (Anders Bjartell, MD, PhD), [e.w.steyerberg@lumc.nl](mailto:e.w.steyerberg@lumc.nl) (Ewout W. Steyerberg, PhD), [d.rizopoulos@erasmusmc.nl](mailto:d.rizopoulos@erasmusmc.nl) (Dimitris Rizopoulos, PhD)

**Objective:** Develop a risk prediction model and web-application to assist patients/doctors in personalized biopsy decisions.

**Design, Setting, and Participants:** Model development: world's largest AS study PRIAS, 7813 patients, 1134 experienced upgrading; External validation: largest six cohorts of Movember Foundation's GAP3 database ( $> 20,000$  patients, 27 centers worldwide); Data: repeat prostate-specific antigen (PSA) and biopsy Gleason grade group.

**Outcome Measurements, and Statistical Analysis:** A Bayesian joint model fitted to PRIAS dataset. This model was validated in GAP3 cohorts using risk prediction error, calibration, area under ROC (AUC). Model and personalized biopsy schedules based on predicted risks were implemented in a web-application.

**Results and Limitations:** Cause-specific cumulative upgrading-risk at year five of follow-up: 35% in PRIAS, at most 50% in GAP3 cohorts. PRIAS based model: PSA velocity was stronger predictor of upgrading (Hazard Ratio: 2.47, 95%CI: 1.93–2.99) than PSA value (Hazard Ratio: 0.99, 95%CI: 0.89–1.11). Validation: Moderate AUC (0.55–0.75) in PRIAS and GAP3 cohorts. Moderate prediction error (0.1–0.3) in GAP3 cohorts where impact of PSA value and velocity on upgrading-risk was similar to PRIAS, but large (0.3–0.45) otherwise. Recalibration of baseline risk required for external cohorts.

**Conclusions:** We successfully developed and validated a model for predicting upgrading-risk, and providing risk-based personalized biopsy decisions, in prostate cancer AS. The model made available via a web-application enables shared decision making of biopsy schedules by comparing fixed and personalized schedules on total biopsies and expected time delay in detecting upgrading.

**Patient Summary:** Personalized prostate biopsies are a novel alternative to fixed one-size-fits-all schedules. The underlying statistical models are made available through a user-friendly web-application and may help to reduce unnecessary prostate biopsies while maintaining cancer control.

**Keywords:** Active Surveillance, Biopsies, Personalized Medicine, Prostate Cancer, Shared Decision Making

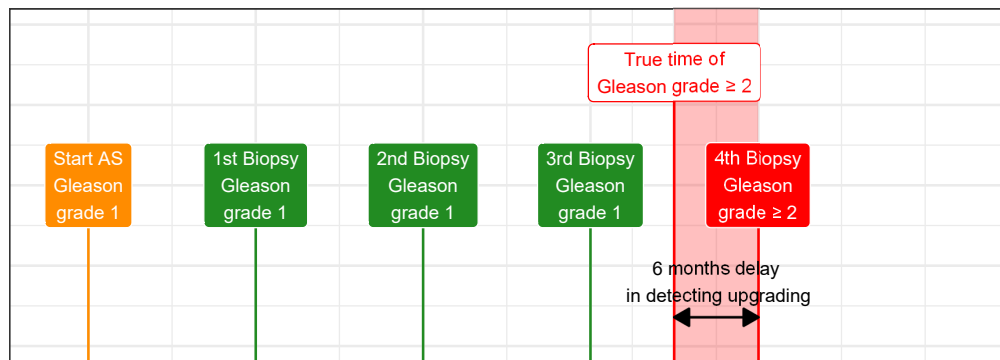
---

## 1. Introduction

Patients with low- and very low-risk screening-detected localized prostate cancer are usually recommended active surveillance (AS) instead of immediate radical treatment [1]. In AS, cancer progression is routinely monitored via prostate-specific antigen (PSA), digital rectal examination, and repeat biopsies. Among these, the strongest indicator of cancer-related outcomes is the biopsy Gleason grade group [2]. When it increases from group 1 (Gleason 3+3) to 2 (Gleason 3+4) or higher, called *upgrading* [3], patients are commonly advised curative treatment [4].

Usually, AS protocols schedule biopsies periodically. Consequently, upgrading is always detected with a time delay (Figure 1). For detecting

### A Biopsy every year



### B Biopsy every 2 years

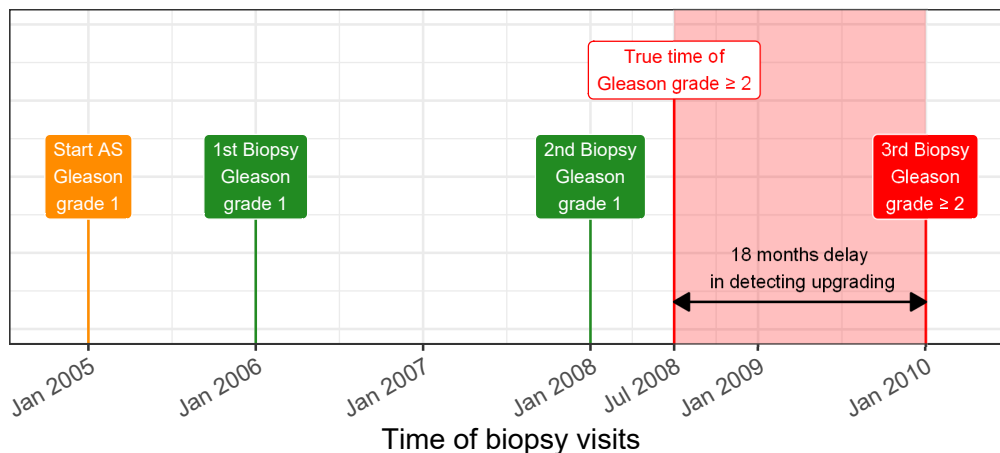


Figure 1: **Trade-off between the number of biopsies and time delay in detecting upgrading (Increase in Gleason grade group from 1 to 2 or higher):** The true time of upgrading for the patient in this figure is July 2008. When biopsies are scheduled annually (**Panel A**), upgrading is detected in January 2009 with a time delay of six months, and a total of four biopsies are scheduled. When biopsies are scheduled biennially (**Panel B**), upgrading is detected in January 2010 with a time delay of 18 months, and a total of three biopsies are scheduled. Since biopsies are conducted periodically, the time of upgrading is observed as an interval. For example, between Jan 2008–Jan 2009 in **Panel A** and between Jan 2008–Jan 2010 in **Panel B**. The phrase ‘Gleason grade group’ is shortened to ‘Gleason grade’ for brevity.

12 upgrading timely, many AS programs schedule fixed and frequent biopsies  
 13 (e.g., annually) for all patients [5, 6]. However, this leads to unnecessary biop-  
 14 sies in slow/non-progressing patients. Biopsies are invasive, may be painful,  
 15 and are prone to medical complications such as bleeding and septicemia[7].  
 16 Biopsy burden and patient non-compliance to frequent biopsies [8] have raised  
 17 concerns regarding the optimal biopsy schedule [9, 10]. In this regard, in  
 18 some cohorts, magnetic resonance imaging (MRI) is employed for targeted  
 19 biopsies and to study its value for tumor monitoring. Although, due to cur-  
 20 rently limited AS data, MRI’s value is not clear. Others have proposed the  
 21 option of scheduling biopsies infrequently (e.g., biennially) [9, 11]. However,  
 22 due to differences in baseline upgrading-risk across cohorts [9], fixed biopsy  
 23 schemes can still lead to many unnecessary biopsies. A promising alternative  
 24 to fixed schedules are personalized biopsy schedules based on the patient-  
 25 specific upgrading-risk (Figure 2).

26 The first challenge in creating personalized biopsy schedules is developing  
 27 a statistical model to consolidate accumulated patient data (e.g., PSA, pre-  
 28 vious biopsy results) into predictions for upgrading-risk. Existing upgrading-  
 29 risk [12, 13] calculators use only the latest PSA measurement of a patient.  
 30 Comparatively, more information is captured by considering all repeatedly  
 31 measured PSA, previous biopsy results, and baseline characteristics of a pa-  
 32 tient. To this end, a suitable model is the joint model for time-to-event and  
 33 longitudinal data [14, 15, 16]. A joint model predicts upgrading-risk in a  
 34 personalized manner. However, a subsequent challenge is translating pre-  
 35 dicted risks into clinical decisions. For example, a 10% upgrading-risk can  
 36 be perceived high/low depending upon the patient’s age. Patients may also

**A** Should a biopsy be conducted at current visit?

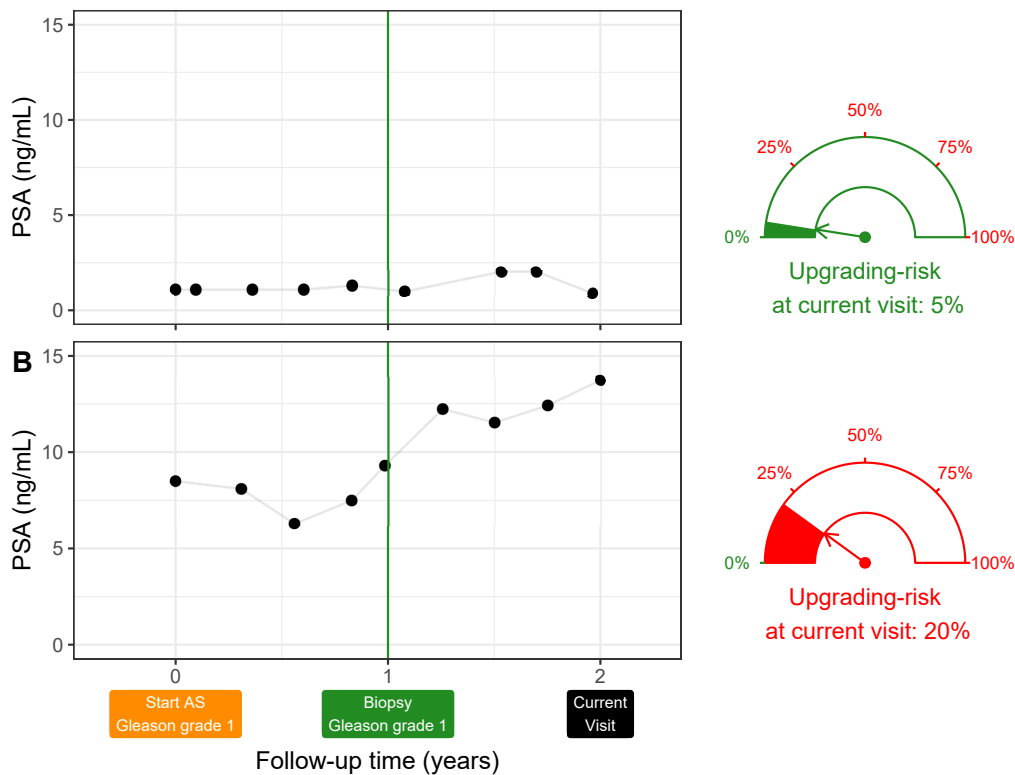


Figure 2: **Motivation for personalized upgrading-risk based decisions of biopsy:** Patient A (**Panel A**) and B (**Panel B**) had their latest biopsy at year one of follow-up (green vertical line). Patient A's prostate-specific antigen (PSA) profile remained stable until his current visit at year two, whereas patient B's profile has shown a rise. Consequently, patient B's upgrading-risk at the current visit (year two) is higher than that of patient A. This makes patient B a more suitable candidate for biopsy than Patient A. Risk estimates in this figure are only illustrative.

weigh risks of upgrading with the potential *consequences* of another biopsy. Two such relevant consequences (Figure 1) are the timing and the total number of planned biopsies (burden), and the time delay in detecting upgrading (smaller is beneficial). The relative importance of these consequences can vary between the patients, and also within a patient over the follow-up period.

The goal of this work is two-fold. First, to develop a robust, generalizable model that gives reliable estimates for individual upgrading-risk in AS. Second, to utilize the predicted upgrading-risks to create personalized biopsy schedules. To facilitate shared decision making of biopsy schedules, we also intend to provide quantitative estimates of the aforementioned *consequences* of opting for a personalized versus the standard fixed schedule. For developing our model, we will use the world’s largest AS dataset PRIAS. Subsequently, we want to externally validate our model in the largest six AS cohorts from the Movember Foundation’s GAP3 database [17]. Last, we intend to implement our model and methodology in a web-application.

## 2. Patients and Methods

### 2.1. Study Cohort

For developing a statistical model to predict upgrading-risk, we used the world’s largest AS dataset, Prostate Cancer International Active Surveillance or PRIAS [4], dated April 2019 (Table 1). In PRIAS, PSA was measured quarterly for the first two years of follow-up and semiannually thereafter. Biopsies were scheduled at year one, four, seven, and ten of follow-up. Additional yearly biopsies were scheduled when PSA doubling time was between

61 zero and ten years.

62 We selected all 7813 patients who had Gleason grade group 1 at the time  
 63 of inclusion in PRIAS. Our primary event of interest is an increase in this  
 64 Gleason grade group observed upon repeat biopsy, called *upgrading* (1134 pa-  
 65 tients). Upgrading is a trigger for treatment advice in PRIAS. Also, 2250 pa-  
 66 tients were provided treatment based on their PSA, number of biopsy cores  
 67 with cancer, or anxiety/other reasons. Our reasons for focusing solely on  
 68 upgrading are, namely, upgrading is strongly associated with cancer-related  
 69 outcomes, and other treatment triggers vary between cohorts [5].

70 For model validation, we selected the following largest (by number of  
 71 repeated measurements) six cohorts from Movember Foundation’s GAP3  
 72 database version 3.1 [17]: University of California San Francisco AS (UCSF,  
 73 version 3.2), University of Toronto AS (Toronto), Johns Hopkins AS (Hop-  
 74 kins), Memorial Sloan Kettering Cancer Center AS (MSKCC), King’s College  
 75 London AS (KCL), and Michigan Urological Surgery Improvement Collabo-  
 76 rative AS (MUSIC). Only patients with a Gleason grade group 1 at the time  
 77 of inclusion in these cohorts were selected. Summary statistics are presented  
 78 in Supplementary A.2.

## 79 2.2. Statistical Model

80 For developing an upgrading-risk prediction model, the data we utilized  
 81 from the PRIAS cohort was patient age at inclusion in AS, longitudinally  
 82 measured PSA, timing of repeat biopsies and Gleason grades, and observed  
 83 time of upgrading. Analysis of this data required modeling the within-patient  
 84 correlation for PSA, the association between the Gleason grades and PSA  
 85 profiles of a patient, and handling missing PSA measurements after a patient



Table 1: **Summary of the PRIAS dataset as of April 2019.** The primary event of interest is upgrading, that is, increase in Gleason grade group from group 1 [2] to 2 or higher. IQR: interquartile range, PSA: prostate-specific antigen. Study protocol URL: <https://www.prias-project.org>

Characteristic	Value
Total centers	> 100
Total patients	7813
Upgrading (primary event)	1134
Treatment	2250
Watchful waiting	334
Loss to follow-up	249
Death (unrelated to prostate cancer)	95
Death (related to prostate cancer)	2
Median age at diagnosis (years)	66 (IQR: 61–71)
Median maximum follow-up per patient (years)	1.8 (IQR: 0.9–4.0)
Total PSA measurements	67578
Median number of PSA measurements per patient	6 (IQR: 4–12)
Median PSA value (ng/mL)	5.7 (IQR: 4.1–7.7)
Total biopsies	15686
Median number of biopsies per patient	2 (IQR: 1–2)

86 experienced upgrading. In such situations, a commonly used model is the  
 87 joint model for time-to-event and longitudinal data [14, 15, 16].

88 Our joint model consisted of two sub-models. First, a linear mixed sub-  
 89 model [18] for longitudinally measured PSA (log-transformed). Second, a  
 90 relative-risk sub-model (similar to the Cox model) for obtaining the cause-  
 91 specific upgrading-risk. We included patient age in both sub-models. In the  
 92 PSA sub-model, we fitted a unique curve to the PSA measurements of each  
 93 patient (Panel A, Figure 3). Subsequently, we calculated the mathematical  
 94 derivative of the patient’s fitted PSA profile (Equation 2, Supplementary A),  
 95 to obtain his follow-up time specific instantaneous PSA velocity (Panel B,  
 96 Figure 3). This instantaneous velocity is a stronger predictor of upgrading  
 97 than the widely used average PSA velocity [19]. We modeled the impact  
 98 of PSA on upgrading-risk by employing fitted PSA value and instantaneous  
 99 velocity as predictors in the risk sub-model. Also, we included the time of  
 100 the latest negative biopsy in the risk sub-model (Panel C, Figure 3). The  
 101 parameters of the two sub-models were estimated jointly (Supplementary A)  
 102 using the R package **JMbayes** [20].

### 103 *2.3. Risk Prediction and Model Validation*

104 Our model provides predictions for upgrading-risk over the entire fu-  
 105 ture follow-up period of a patient. Predictions also automatically update  
 106 over follow-up as more patient data becomes available (Figure 5, Supple-  
 107 mentary B). We validated our PRIAS based model internally in the PRIAS  
 108 cohort, and externally in the largest six GAP3 database cohorts. We em-  
 109 ployed calibration plots [21, 22] and follow-up *time-dependent* mean absolute  
 110 risk prediction error or MAPE [23] to graphically and quantitatively evaluate

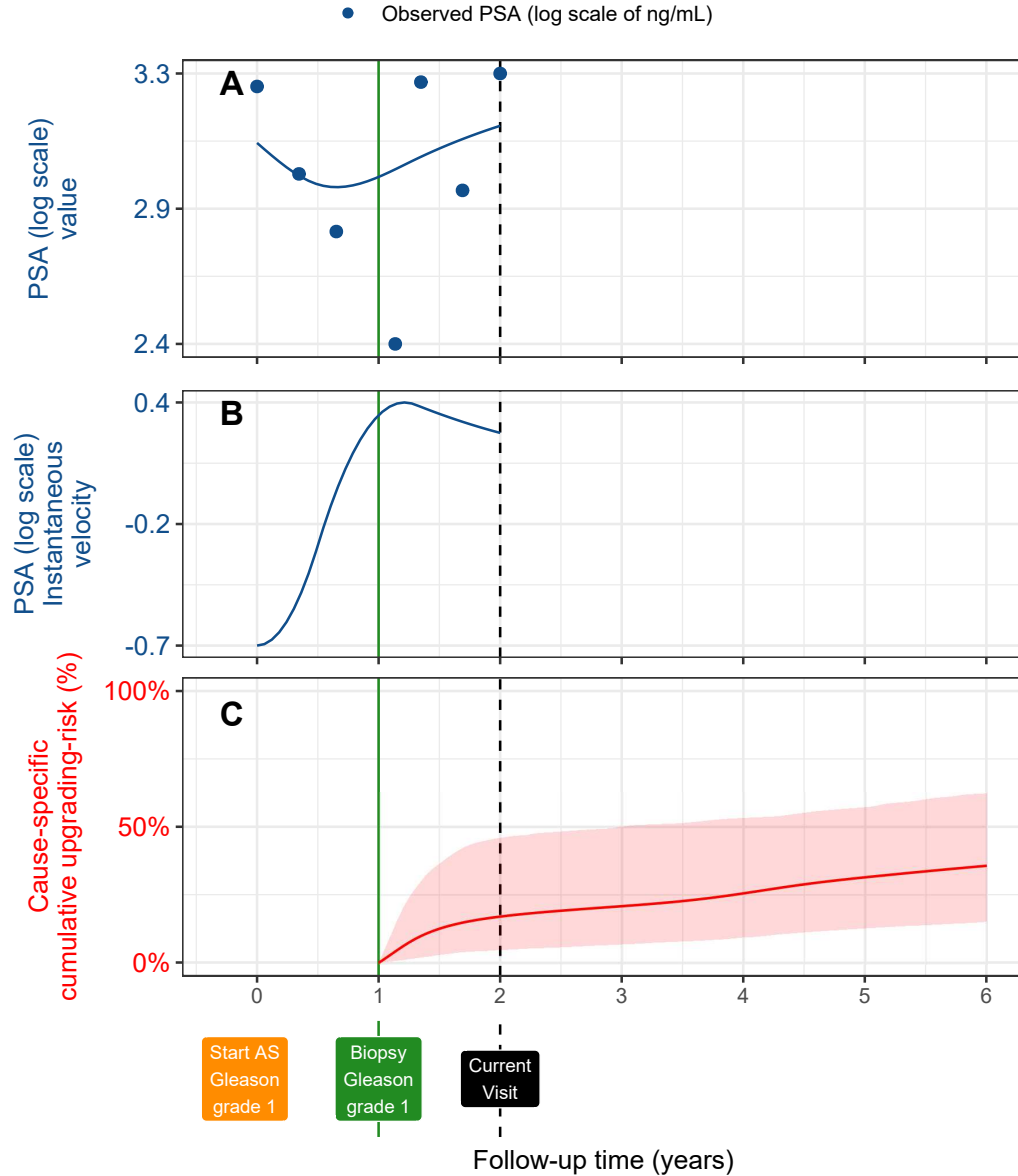


Figure 3: **Illustration of the joint model on a real PRIAS patient.** **Panel A:** Observed PSA (blue dots) and fitted PSA (solid blue line), log-transformed from ng/mL. **Panel B:** Estimated instantaneous velocity of PSA (log-transformed). **Panel C:** Predicted cause-specific cumulative upgrading-risk (95% credible interval shaded). Upgrading is defined as an increase in the Gleason grade group from group 1 [2] to 2 or higher. This upgrading-risk is calculated starting from the time of the latest negative biopsy (vertical green line at year one of follow-up). The joint model estimated it by combining the fitted PSA (log scale) value and instantaneous velocity, and time of the latest negative biopsy. Black dashed line at year two denotes the time of current visit.

our model’s risk prediction accuracy, respectively. We assessed our model’s ability to discriminate between patients who experience/do not experience upgrading via the time-dependent area under the receiver operating characteristic curve or AUC [23].

The aforementioned *time-dependent* AUC and MAPE [23] are temporal extensions of their standard versions [22] in a longitudinal setting. Specifically, at every six months of follow-up, we calculated a unique AUC and MAPE for predicting upgrading-risk in the subsequent one year (Supplementary B.1). For emulating a realistic situation, we calculated the AUC and MAPE at each follow-up using only the validation data available until that follow-up. Last, to resolve any potential model miscalibration in validation cohorts, we aimed to recalibrate our model’s baseline hazard of upgrading (Supplementary B.1), individually for each cohort.

### 3. Results

The cause-specific cumulative upgrading-risk at year five of follow-up was 35% in PRIAS and at most 50% in validation cohorts (Panel B, Figure 4). Hence, many patients do not require all biopsies planned in the first five years of AS. In the fitted PRIAS model, the adjusted hazard ratio (aHR) of upgrading for an increase in patient age from 61 to 71 years (25-th to 75-th percentile) was 1.45 (95%CI: 1.30–1.63). For an increase in fitted PSA value from 2.36 to 3.07 (25-th to 75-th percentile, log scale), the aHR was 0.99 (95%CI: 0.89–1.11). The strongest predictor of upgrading-risk was instantaneous PSA velocity, with an increase from -0.09 to 0.31 (25-th to 75-th percentile), giving an aHR of 2.47 (95%CI: 1.93–2.99). The aHR for PSA

135 value and velocity varied between GAP3 cohorts (Supplementary Table 8).

136 The time-dependent AUC, calibration plot, and time-dependent MAPE  
 137 of our model are shown in Figure 4, and Supplementary Figure 8. In all  
 138 cohorts, time-dependent AUC was moderate (0.55 to 0.75) over the whole  
 139 follow-up period. Time-dependent MAPE was large (0.3 to 0.45) in those co-  
 140 horts where the impact of PSA on upgrading-risk was different from PRIAS  
 141 (e.g., MUSIC cohort, Supplementary Table 8), and moderate (0.1 to 0.3) oth-  
 142 erwise. In all cohorts, the MAPE decreased rapidly after year one of follow-  
 143 up. Our model was miscalibrated for validation cohorts (Panel B, Figure 4).  
 144 Recalibrating the baseline hazard of upgrading in validation cohorts resolved  
 145 this issue (Supplementary Figure 6). We compared risk predictions from the  
 146 recalibrated models, with predictions from separately fitted cohort-specific  
 147 joint models (Supplementary Figure 7). The difference in predictions was  
 148 lowest in Johns Hopkins cohort (impact of PSA on upgrading-risk similar to  
 149 PRIAS). Comprehensive results are in Supplementary A.2 and B.

### 150 3.1. *Personalized Biopsy Schedules*

151 We employed the PRIAS based fitted model to create personalized biopsy  
 152 schedules for real PRIAS patients. Specifically, first using the model and pa-  
 153 tient’s observed data, we predicted his cumulative upgrading-risk (Figure 5)  
 154 on all of his future follow-up visits (biannually in PRIAS). Subsequently,  
 155 we planned biopsies on those future visits where his conditional cumulative  
 156 upgrading-risk was more than a certain threshold (Supplementary Figure 9).  
 157 Example personalized schedules based on 5% and 10% risk thresholds are  
 158 shown in Figure 5, and in Supplementary Figure 10–12. For both personal-  
 159 ized and fixed schedules, we estimated the expected time delay in detecting

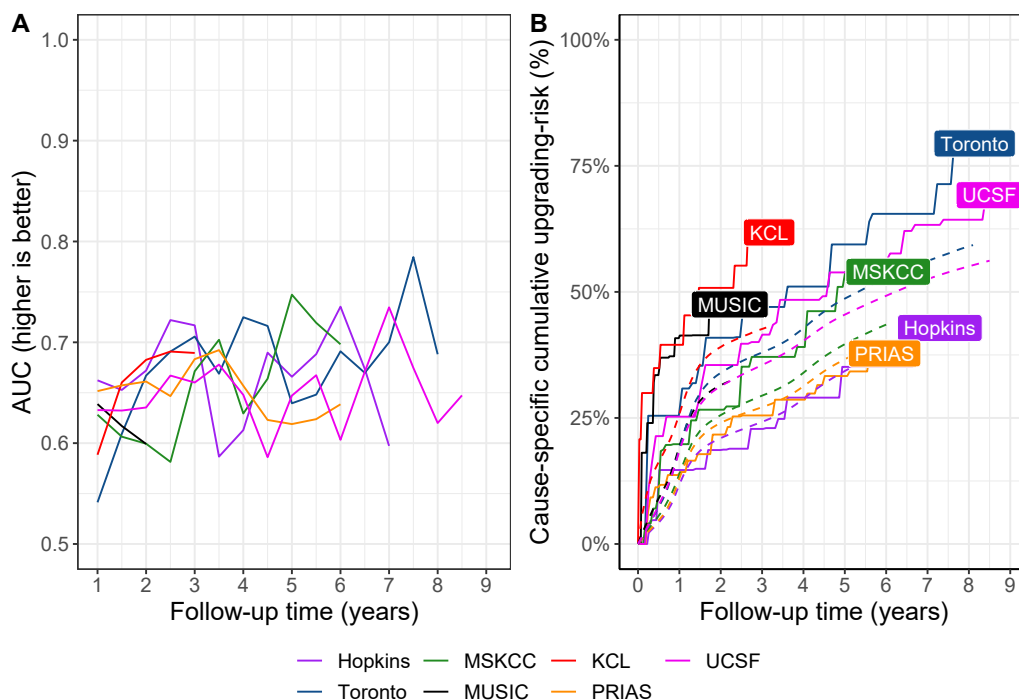


Figure 4: **Model Validation Results.** **Panel A:** time-dependent area under the receiver operating characteristic curve or AUC (measure of discrimination). **Panel B:** calibration-at-large indicates model miscalibration. This is because solid lines depicting the non-parametric estimate of the cause-specific cumulative upgrading-risk [24], and dashed lines showing the average cause-specific cumulative upgrading-risk obtained using the joint model fitted to the PRIAS dataset, are not overlapping. Recalibrating the baseline hazard of upgrading resolved this issue (Supplementary Figure 6). Full names of Cohorts are *PRIAS*: Prostate Cancer International Active Surveillance, *Toronto*: University of Toronto Active Surveillance, *Hopkins*: Johns Hopkins Active Surveillance, *MSKCC*: Memorial Sloan Kettering Cancer Center Active Surveillance, *KCL*: King’s College London Active Surveillance, *MUSIC*: Michigan Urological Surgery Improvement Collaborative Active Surveillance, *UCSF*: University of California San Francisco AS.

160 upgrading if the patient progresses before the time of the last planned biopsy  
 161 (Panel C, Figure 5). This delay is also personalized (Supplementary C.1).  
 162 That is, even if two different patients are prescribed the same biopsy schedule,  
 163 their expected delays will depend on their individual upgrading-risk profiles.  
 164 Patients/doctors can utilize the expected delay and schedule of biopsies as  
 165 criteria to compare fixed, and different risk-based personalized schedules.

### 166 3.2. *Web-Application*

167 We implemented our model and personalized schedules in a user-friendly  
 168 web-application [https://emcbiostatistics.shinyapps.io/prias\\_biopsy\\_](https://emcbiostatistics.shinyapps.io/prias_biopsy_recommender/)  
 169 **recommender/**. Currently, the web-application supports PRIAS and the six  
 170 validation cohorts. Patient data can be entered manually and in Microsoft  
 171 Excel format. Predictions for upgrading-risk are available for a currently  
 172 limited, cohort-specific, follow-up period (Supplementary Table 9). The web-  
 173 application visualizes the timing of biopsies, and expected time delay in de-  
 174 tecting upgrading, for personalized schedules based on 5%, 10%, and 15%  
 175 risk threshold; annual biopsies; biennial biopsies; and PRIAS schedule.

## 176 4. Discussion

177 We successfully developed and externally validated a model for predicting  
 178 upgrading-risk [3] in prostate cancer AS, and providing risk-based personal-  
 179 ized biopsy decisions. Our work has four novel features over earlier risk  
 180 calculators [15, 25]. First, our model was fitted to the world’s largest AS  
 181 dataset PRIAS and externally validated in the largest six cohorts of the  
 182 Movember Foundation’s GAP3 database [17]. Second, the model predicts a  
 183 patient’s current and future upgrading-risk in a personalized manner. Third,

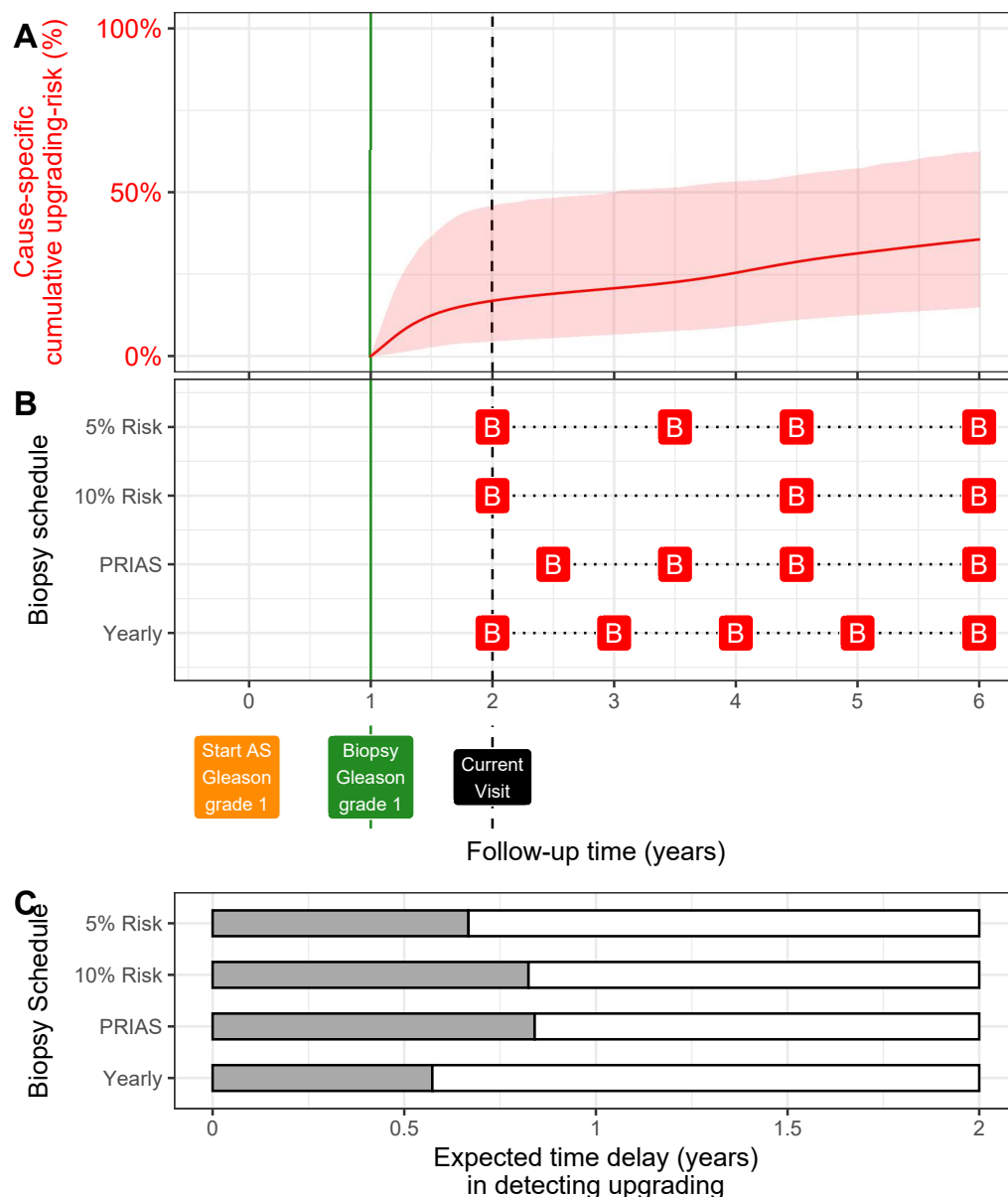


Figure 5: **Illustration of personalized and fixed schedules of biopsies for patient from Figure 3.** **Panel A:** Predicted cumulative upgrading-risk (95% credible interval shaded). **Panel B:** Different biopsy schedules with a red 'B' indicating a future biopsy. Risk: 5% and Risk: 10% are personalized schedules in which a biopsy is planned whenever the conditional cause-specific cumulative upgrading-risk is above 5% or 10% risk, respectively. Green vertical line at year one is the time of the latest negative biopsy. Black dashed line at year two denotes the time of the current visit. **Panel C:** Expected time delay in detecting upgrading (years) if patient progresses before year six. A compulsory biopsy was scheduled at year six (maximum biopsy scheduling time in PRIAS, Supplementary C) in all schedules for a meaningful comparison between them.



184 using the predicted risks, we created personalized biopsy schedules and also  
 185 calculated the expected time delay in detecting upgrading (less is beneficial)  
 186 if that schedule was followed. Thus, patients/doctors can compare sched-  
 187 ules before making a choice. Fourth, we implemented our methodology in a  
 188 user-friendly web-application ([https://emcbiostatistics.shinyapps.io/  
 189 prias\\_biopsy\\_recommender/](https://emcbiostatistics.shinyapps.io/prias_biopsy_recommender/)) for both PRIAS and validated cohorts.

190 Our model is useful for numerous patients from PRIAS and validated  
 191 cohorts. The discrimination ability of our model, exhibited by the *time-*  
 192 *dependent* AUC, was moderate (0.55–0.75). This is possible because, unlike  
 193 the standard AUC [22], the time-dependent AUC utilizes only the validation  
 194 data available until the time at which it is calculated. The same holds for the  
 195 time-dependent MAPE (mean absolute prediction error). Although, MAPE  
 196 varied much more between cohorts than AUC. In cohorts where the effect  
 197 size for the impact of PSA value and velocity on upgrading-risk was similar  
 198 to that for PRIAS (e.g., Hopkins cohort), MAPE was moderate. Otherwise,  
 199 MAPE was large (e.g., KCL and MUSIC cohorts). In all cohorts, MAPE  
 200 decreased rapidly after year one of follow-up. A plausible reason is that at  
 201 year one, the validation data also contains those patients who may have been  
 202 misclassified as Gleason grade group 1 at the time of inclusion in AS. This  
 203 issue can be obviated by scheduling a compulsory biopsy at year one for all  
 204 patients (current PRIAS recommendation). Last, we required recalibration  
 205 of our model’s baseline hazard of upgrading for all validation cohorts.

206 The clinical implications of our work are as follows. First, the cause-  
 207 specific cumulative upgrading-risk at year five of follow-up was at most 50%  
 208 in all cohorts (Panel B, Figure 4). That is, many patients may not require

all biopsies planned in the first five years of AS. Given the non-compliance and burden of frequent biopsies [8], the availability of our methodology as a web-application may encourage patients/doctors to consider upgrading-risk based personalized schedules instead. An additional advantage of personalized schedules is that they update as more patient data becomes available over follow-up. We have shown via a simulation study [26] that personalized schedules may reduce, on average, six biopsies compared to annual schedule and two biopsies compared to PRIAS schedule in slow/non-progressing AS patients, while maintaining almost the same time delay in detecting upgrading as PRIAS schedule. Personalized schedules with different risk thresholds indeed have different performance. In this regard, to assist patients/doctors in choosing between fixed schedules and personalized schedules based on different risk thresholds, the web-application provides a patient-specific estimate of the expected time delay in detecting upgrading, for both personalized and fixed schedules. We hope that this will objectively address patient apprehensions regarding adverse outcomes in AS.

This work has certain limitations. Predictions for upgrading-risk, and personalized schedules are available only for a currently limited, cohort-specific, follow-up period (Supplementary Table 9). This problem can be mitigated by refitting the model with new follow-up data in the future. Recently, some cohorts started utilizing MRI to explore the possibility of targeting visible lesions by biopsy. Presently, the GAP3 database has limited MRI follow-up data available. As more such data becomes available, the current model can be extended to include MRI based predictors. We scheduled biopsies using cause-specific cumulative upgrading-risk, which ignores com-

234 peting events such as treatment based on the number of positive biopsy cores.  
 235 Employing a competing-risk model may lead to improved personalized sched-  
 236 ules. Upgrading is susceptible to inter-observer variation too. Models which  
 237 account for this variation [15, 27] will be interesting to investigate further.  
 238 However, even with an enhanced risk prediction model, the methodology for  
 239 personalized scheduling and calculation of expected time delay (Supplemen-  
 240 tary C) need not change.

## 241 5. Conclusions

242 We successfully developed and externally validated a model for predict-  
 243 ing upgrading-risk, and providing risk-based personalized biopsy decisions,  
 244 in prostate cancer AS. The model made available via a user-friendly web-  
 245 application ([https://emcbiostatistics.shinyapps.io/prias\\_biopsy\\_recommender/](https://emcbiostatistics.shinyapps.io/prias_biopsy_recommender/))  
 246 enables shared decision making of biopsy schedules by comparing fixed and  
 247 personalized schedules on total biopsies and expected time delay in detecting  
 248 upgrading. Novel biomarkers and MRI data can be added as predictors in  
 249 the model to improve predictions in the future. Recalibration of baseline  
 250 upgrading-risk is advised for external cohorts.

## 251 Author Contributions

252 Anirudh Tomer had full access to all the data in the study and takes  
 253 responsibility for the integrity of the data and the accuracy of the data anal-  
 254 ysis.

255 *Study concept and design:* Tomer, Nieboer, Roobol, Bjartell, and Rizopoulos

256 *Acquisition of data:* Tomer, Nieboer, and Roobol

257 *Analysis and interpretation of data:* Tomer, Nieboer, and Rizopoulos  
 258 *Drafting of the manuscript:* Tomer, and Rizopoulos  
 259 *Critical revision of the manuscript for important intellectual content:* Tomer,  
 260 Nieboer, Roobol, Bjartell, Steyerberg, and Rizopoulos  
 261 *Statistical analyses:* Tomer, Nieboer, Steyerberg, and Rizopoulos  
 262 *Obtaining funding:* Roobol, Steyerberg, and Rizopoulos  
 263 *Administrative, technical or material support:* Nieboer  
 264 *Supervision:* Roobol, and Rizopoulos  
 265 *Other:* none

## 266 **Acknowledgments**

267 We thank Jozien Helleman from the Department of Urology, Erasmus  
 268 University Medical Center, for coordinating the project. The first and last  
 269 authors would like to acknowledge support by Nederlandse Organisatie voor  
 270 Wetenschappelijk Onderzoek (the national research council of the Nether-  
 271 lands) VIDI grant nr. 016.146.301, and Erasmus University Medical Cen-  
 272 ter funding. Part of this work was carried out on the Dutch national e-  
 273 infrastructure with the support of SURF Cooperative. The authors also  
 274 thank the Erasmus University Medical Center's Cancer Computational Biol-  
 275 ogy Center for giving access to their IT-infrastructure and software that was  
 276 used for the computations and data analysis in this study.

277 The PRIAS website is funded by the Prostate Cancer Research Foun-  
 278 dation, Rotterdam (SWOP). This work was supported by the Movember  
 279 Foundation. The funder did not play any role in the study design, collection,  
 280 analysis or interpretation of data, or in the drafting of this paper.

281 **Appendix A. Members of The Movember Foundation’s Global Ac-**  
 282 **tion Plan Prostate Cancer Active Surveillance (GAP3) consortium**

283 *Principle Investigators:* Bruce Trock (Johns Hopkins University, The  
 284 James Buchanan Brady Urological Institute, Baltimore, USA), Behfar Ehdaie  
 285 (Memorial Sloan Kettering Cancer Center, New York, USA), Peter Car-  
 286 roll (University of California San Francisco, San Francisco, USA), Christo-  
 287 pher Filson (Emory University School of Medicine, Winship Cancer Insti-  
 288 tute, Atlanta, USA), Jeri Kim / Christopher Logothetis (MD Anderson  
 289 Cancer Centre, Houston, USA), Todd Morgan (University of Michigan and  
 290 Michigan Urological Surgery Improvement Collaborative (MUSIC), Michi-  
 291 gan, USA), Laurence Klotz (University of Toronto, Sunnybrook Health Sci-  
 292 ences Centre, Toronto, Ontario, Canada), Tom Pickles (University of British  
 293 Columbia, BC Cancer Agency, Vancouver, Canada), Eric Hyndman (Uni-  
 294 versity of Calgary, Southern Alberta Institute of Urology, Calgary, Canada),  
 295 Caroline Moore (University College London & University College London  
 296 Hospital Trust, London, UK), Vincent Gnanapragasam (University of Cam-  
 297 bridge & Cambridge University Hospitals NHS Foundation Trust, Cam-  
 298 bridge, UK), Mieke Van Hemelrijck (King’s College London, London, UK  
 299 & Guy’s and St Thomas’ NHS Foundation Trust, London, UK), Prokar Das-  
 300 gupta (Guy’s and St Thomas’ NHS Foundation Trust, London, UK), Chris  
 301 Bangma (Erasmus Medical Center, Rotterdam, The Netherlands/ represen-  
 302 tative of Prostate cancer Research International Active Surveillance (PRIAS)  
 303 consortium), Monique Roobol (Erasmus Medical Center, Rotterdam, The  
 304 Netherlands/ representative of Prostate cancer Research International Active  
 305 Surveillance (PRIAS) consortium), Arnauld Villers (Lille University Hospi-

tal Center, Lille, France), Antti Rannikko (Helsinki University and Helsinki  
 University Hospital, Helsinki, Finland), Riccardo Valdagni (Department of  
 Oncology and Hemato-oncology, Università degli Studi di Milano, Radia-  
 tion Oncology 1 and Prostate Cancer Program, Fondazione IRCCS Istituto  
 Nazionale dei Tumori, Milan, Italy), Antoinette Perry (University College  
 Dublin, Dublin, Ireland), Jonas Hugosson (Sahlgrenska University Hospital,  
 Göteborg, Sweden), Jose Rubio-Briones (Instituto Valenciano de Oncología,  
 Valencia, Spain), Anders Bjartell (Skåne University Hospital, Malmö, Swe-  
 den), Lukas Hefermehl (Kantonsspital Baden, Baden, Switzerland), Lee Lui  
 Shiong (Singapore General Hospital, Singapore, Singapore), Mark Fryden-  
 berg (Monash Health; Monash University, Melbourne, Australia), Yoshiyuki  
 Kakehi / Mikio Sugimoto (Kagawa University Faculty of Medicine, Kagawa,  
 Japan), Byung Ha Chung (Gangnam Severance Hospital, Yonsei University  
 Health System, Seoul, Republic of Korea)

*Pathologist:* Theo van der Kwast (Princess Margaret Cancer Centre,  
 Toronto, Canada). *Technology Research Partners:* Henk Obbink (Royal  
 Philips, Eindhoven, the Netherlands), Wim van der Linden (Royal Philips,  
 Eindhoven, the Netherlands), Tim Hulsén (Royal Philips, Eindhoven, the  
 Netherlands), Cees de Jonge (Royal Philips, Eindhoven, the Netherlands).

*Advisory Regional statisticians:* Mike Kattan (Cleveland Clinic, Cleve-  
 land, Ohio, USA), Ji Xinge (Cleveland Clinic, Cleveland, Ohio, USA), Ken-  
 neth Muir (University of Manchester, Manchester, UK), Artitaya Lophatananon  
 (University of Manchester, Manchester, UK), Michael Fahey (Epworth Health-  
 Care, Melbourne, Australia), Ewout Steyerberg (Erasmus Medical Center,  
 Rotterdam, The Netherlands), Daan Nieboer (Erasmus Medical Center, Rot-

terdam, The Netherlands); Liying Zhang (University of Toronto, Sunnybrook  
Health Sciences Centre, Toronto, Ontario, Canada)

*Executive Regional statisticians:* Ewout Steyerberg (Erasmus Medical  
Center, Rotterdam, The Netherlands), Daan Nieboer (Erasmus Medical Cen-  
ter, Rotterdam, The Netherlands); Kerri Beckmann (King's College London,  
London, UK & Guy's and St Thomas' NHS Foundation Trust, London, UK),  
Brian Denton (University of Michigan, Michigan, USA), Andrew Hayen (Uni-  
versity of Technology Sydney, Australia), Paul Boutros (Ontario Institute of  
Cancer Research, Toronto, Ontario, Canada).

*Clinical Research Partners' IT Experts:* Wei Guo (Johns Hopkins Uni-  
versity, The James Buchanan Brady Urological Institute, Baltimore, USA),  
Nicole Benfante (Memorial Sloan Kettering Cancer Center, New York, USA),  
Janet Cowan (University of California San Francisco, San Francisco, USA),  
Dattatraya Patil (Emory University School of Medicine, Winship Cancer In-  
stitute, Atlanta, USA), Emily Tolosa (MD Anderson Cancer Centre, Hous-  
ton, Texas, USA), Tae-Kyung Kim (University of Michigan and Michigan  
Urological Surgery Improvement Collaborative, Ann Arbor, Michigan, USA),  
Alexandre Mamedov (University of Toronto, Sunnybrook Health Sciences  
Centre, Toronto, Ontario, Canada), Vincent LaPointe (University of British  
Columbia, BC Cancer Agency, Vancouver, Canada), Trafford Crump (Uni-  
versity of Calgary, Southern Alberta Institute of Urology, Calgary, Canada),  
Vasilis Stavriniades (University College London & University College Lon-  
don Hospital Trust, London, UK), Jenna Kimberly-Duffell (University of  
Cambridge & Cambridge University Hospitals NHS Foundation Trust, Cam-  
bridge, UK), Aida Santaolalla (King's College London, London, UK & Guy's

and St Thomas' NHS Foundation Trust, London, UK), Daan Nieboer (Erasmus Medical Center, Rotterdam, The Netherlands), Jonathan Olivier (Lille University Hospital Center, Lille, France), Tiziana Rancati (Fondazione IRCCS Istituto Nazionale dei Tumori di Milano, Milan, Italy), Helén Ahlgren (Sahlgrenska University Hospital, Göteborg, Sweden), Juanma Mascarós (Instituto Valenciano de Oncología, Valencia, Spain), Annica Löfgren (Skåne University Hospital, Malmö, Sweden), Kurt Lehmann (Kantonsspital Baden, Baden, Switzerland), Catherine Han Lin (Monash University and Epworth HealthCare, Melbourne, Australia), Hiromi Hiram (Kagawa University, Kagawa, Japan), Kwang Suk Lee (Yonsei University College of Medicine, Gangnam Severance Hospital, Seoul, Korea).

*Research Advisory Committee:* Guido Jenster (Erasmus MC, Rotterdam, the Netherlands), Anssi Auvinen (University of Tampere, Tampere, Finland), Anders Bjartell (Skåne University Hospital, Malmö, Sweden), Masoom Haider (University of Toronto, Toronto, Canada), Kees van Bochove (The Hyve B.V. Utrecht, Utrecht, the Netherlands), Ballentine Carter (Johns Hopkins University, Baltimore, USA – until 2018).

*Management team:* Sam Gledhill (Movember Foundation, Melbourne, Australia), Mark Buzza / Michelle Koussou (Movember Foundation, Melbourne, Australia), Chris Bangma (Erasmus Medical Center, Rotterdam, The Netherlands), Monique Roobol (Erasmus Medical Center, Rotterdam, The Netherlands), Sophie Bruinsma / Joziën Helleman (Erasmus Medical Center, Rotterdam, The Netherlands).



## 379 References

- 380 [1] Briganti A, Fossati N, Catto JW, Cornford P, Montorsi F, Mottet N,  
 381 et al. Active surveillance for low-risk prostate cancer: the European As-  
 382 sociation of Urology position in 2018. *European urology* 2018;74(3):357–  
 383 68.
- 384 [2] Epstein JI, Egevad L, Amin MB, Delahunt B, Srigley JR, Humphrey PA.  
 385 The 2014 international society of urological pathology (isup) consensus  
 386 conference on gleason grading of prostatic carcinoma. *The American*  
 387 *journal of surgical pathology* 2016;40(2):244–52.
- 388 [3] Bruinsma SM, Roobol MJ, Carroll PR, Klotz L, Pickles T, Moore CM,  
 389 et al. Expert consensus document: semantics in active surveillance for  
 390 men with localized prostate cancer—results of a modified delphi consen-  
 391 sus procedure. *Nature reviews urology* 2017;14(5):312.
- 392 [4] Bul M, Zhu X, Valdagni R, Pickles T, Kakehi Y, Rannikko A, et al.  
 393 Active surveillance for low-risk prostate cancer worldwide: the prias  
 394 study. *European urology* 2013;63(4):597–603.
- 395 [5] Nieboer D, Tomer A, Rizopoulos D, Roobol MJ, Steyerberg EW. Active  
 396 surveillance: a review of risk-based, dynamic monitoring. *Translational*  
 397 *andrology and urology* 2018;7(1):106–15.
- 398 [6] Loeb S, Carter HB, Schwartz M, Fagerlin A, Braithwaite RS, Lepor H.  
 399 Heterogeneity in active surveillance protocols worldwide. *Reviews in*  
 400 *urology* 2014;16(4):202–3.

- 401 [7] Loeb S, Vellekoop A, Ahmed HU, Catto J, Emberton M, Nam R, et al.  
402 Systematic review of complications of prostate biopsy. *European urology*  
403 2013;64(6):876–92.
- 404 [8] Bokhorst LP, Alberts AR, Rannikko A, Valdagni R, Pickles T, Kakehi  
405 Y, et al. Compliance rates with the Prostate Cancer Research Interna-  
406 tional Active Surveillance (PRIAS) protocol and disease reclassification  
407 in noncompliers. *European Urology* 2015;68(5):814–21.
- 408 [9] Inoue LY, Lin DW, Newcomb LF, Leonardson AS, Ankerst D, Gulati R,  
409 et al. Comparative analysis of biopsy upgrading in four prostate cancer  
410 active surveillance cohorts. *Annals of internal medicine* 2018;168(1):1–9.
- 411 [10] Bratt O, Carlsson S, Holmberg E, Holmberg L, Johansson E, Josefsson  
412 A, et al. The study of active monitoring in sweden (sams): a ran-  
413 domized study comparing two different follow-up schedules for active  
414 surveillance of low-risk prostate cancer. *Scandinavian journal of urology*  
415 2013;47(5):347–55.
- 416 [11] de Carvalho TM, Heijnsdijk EA, de Koning HJ. Estimating the risks  
417 and benefits of active surveillance protocols for prostate cancer: a mi-  
418 crosimulation study. *BJU international* 2017;119(4):560–6.
- 419 [12] Partin AW, Yoo J, Carter HB, Pearson JD, Chan DW, Epstein JI, et al.  
420 The use of prostate specific antigen, clinical stage and gleason score to  
421 predict pathological stage in men with localized prostate cancer. *The*  
422 *Journal of urology* 1993;150(1):110–4.

- 423 [13] Makarov DV, Trock BJ, Humphreys EB, Mangold LA, Walsh PC, Ep-  
424 stein JI, et al. Updated nomogram to predict pathologic stage of prostate  
425 cancer given prostate-specific antigen level, clinical stage, and biopsy  
426 gleason score (partin tables) based on cases from 2000 to 2005. *Urology*  
427 2007;69(6):1095–101.
- 428 [14] Tomer A, Nieboer D, Roobol MJ, Steyerberg EW, Rizopoulos D. Per-  
429 sonalized schedules for surveillance of low-risk prostate cancer patients.  
430 *Biometrics* 2019;75(1):153–62.
- 431 [15] Coley RY, Zeger SL, Mamawala M, Pienta KJ, Carter HB. Prediction  
432 of the pathologic gleason score to inform a personalized management  
433 program for prostate cancer. *European urology* 2017;72(1):135–41.
- 434 [16] Rizopoulos D. *Joint Models for Longitudinal and Time-to-Event Data:*  
435 *With Applications in R.* CRC Press; 2012. ISBN 9781439872864.
- 436 [17] Bruinsma SM, Zhang L, Roobol MJ, Bangma CH, Steyerberg EW,  
437 Nieboer D, et al. The movember foundation’s gap3 cohort: a profile of  
438 the largest global prostate cancer active surveillance database to date.  
439 *BJU international* 2018;121(5):737–44.
- 440 [18] Laird NM, Ware JH, et al. Random-effects models for longitudinal data.  
441 *Biometrics* 1982;38(4):963–74.
- 442 [19] Cooperberg MR, Brooks JD, Faino AV, Newcomb LF, Kearns JT, Car-  
443 roll PR, et al. Refined analysis of prostate-specific antigen kinetics to  
444 predict prostate cancer active surveillance outcomes. *European urology*  
445 2018;74(2):211–7.

- 446 [20] Rizopoulos D. The R package JMBayes for fitting joint models for lon-  
447 gitudinal and time-to-event data using MCMC. *Journal of Statistical*  
448 *Software* 2016;72(7):1–46.
- 449 [21] Royston P, Altman DG. External validation of a cox prognostic  
450 model: principles and methods. *BMC medical research methodology*  
451 2013;13(1):33.
- 452 [22] Steyerberg EW, Vickers AJ, Cook NR, Gerds T, Gonen M, Obuchowski  
453 N, et al. Assessing the performance of prediction models: a framework  
454 for some traditional and novel measures. *Epidemiology (Cambridge,*  
455 *Mass)* 2010;21(1):128.
- 456 [23] Rizopoulos D, Molenberghs G, Lesaffre EM. Dynamic predictions with  
457 time-dependent covariates in survival analysis using joint modeling and  
458 landmarking. *Biometrical Journal* 2017;59(6):1261–76.
- 459 [24] Turnbull BW. The empirical distribution function with arbitrarily  
460 grouped, censored and truncated data. *Journal of the Royal Statisti-*  
461 *cal Society Series B (Methodological)* 1976;38(3):290–5.
- 462 [25] Ankerst DP, Xia J, Thompson Jr IM, Hoeffler J, Newcomb LF, Brooks  
463 JD, et al. Precision medicine in active surveillance for prostate can-  
464 cer: development of the canary–early detection research network active  
465 surveillance biopsy risk calculator. *European urology* 2015;68(6):1083–8.
- 466 [26] Tomer A, Rizopoulos D, Nieboer D, Drost FJ, Roobol MJ, Steyerberg  
467 EW. Personalized decision making for biopsies in prostate cancer active  
468 surveillance programs. *Medical Decision Making* 2019;39(5):499–508.

- 469 [27] Balasubramanian R, Lagakos SW. Estimation of a failure time distribu-  
470 tion based on imperfect diagnostic tests. *Biometrika* 2003;90(1):171–82.