

A Ready To Use Web-Application Providing a Personalized Biopsy Schedule for Men With Low-Risk PCa Under Active Surveillance*

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Abstract

Background: Prostate cancer active surveillance (AS) patients undergo repeat biopsies. Active treatment is advised when biopsy Gleason grade group ≥ 2 (*upgrading*). Many patients never experience upgrading, yet undergo biopsies frequently. Personalized biopsy decisions based on upgrading-risk may

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reduce patient burden.

Objective: Develop a risk prediction model and web-application to assist patients/doctors in personalized biopsy decisions.

Design, Setting, and Participants: Model development: world's largest AS study PRIAS, 7813 patients, 1134 experienced upgrading; External validation: largest five cohorts of Movember Foundation's GAP3 database ($> 20,000$ patients, 27 centers worldwide); Data: repeat prostate-specific antigen (PSA) and biopsy Gleason grade.

Outcome Measurements, and Statistical Analysis: A Bayesian joint model fitted to the PRIAS dataset. This model was validated in GAP3 cohorts using risk prediction error, calibration, area under ROC (AUC). Model and personalized biopsy schedules based on predicted risks were implemented in a web-application.

Results and Limitations: Cause-specific cumulative upgrading-risk at year five of follow-up: 35% in PRIAS, at most 50% in GAP3 cohorts. PRIAS based model: PSA velocity was a stronger predictor of upgrading (Hazard Ratio: 2.47, 95%CI: 1.93–2.99) than PSA value (Hazard Ratio: 0.99, 95%CI: 0.89–1.11). Validation: Moderate AUC (0.55–0.75) in PRIAS and GAP3 cohorts. Moderate prediction error (0.1–0.3) in GAP3 cohorts where impact of PSA value and velocity on upgrading-risk was similar to PRIAS, but large (0.3–0.45) otherwise. Recalibration advised for external cohorts.

Conclusions: We successfully developed and validated a model for predicting upgrading-risk, and providing risk-based personalized biopsy decisions, in prostate cancer AS. The model made available via a web-application enables shared decision making of biopsy schedules by comparing fixed and personalized schedules on total biopsies and expected time delay in detecting upgrading.

Patient Summary: Personalized prostate biopsies are a novel alternative to fixed one-size-fits-all schedules. The underlying statistical models are made available through a user-friendly web-application and may help to reduce unnecessary prostate biopsies while maintaining cancer control.

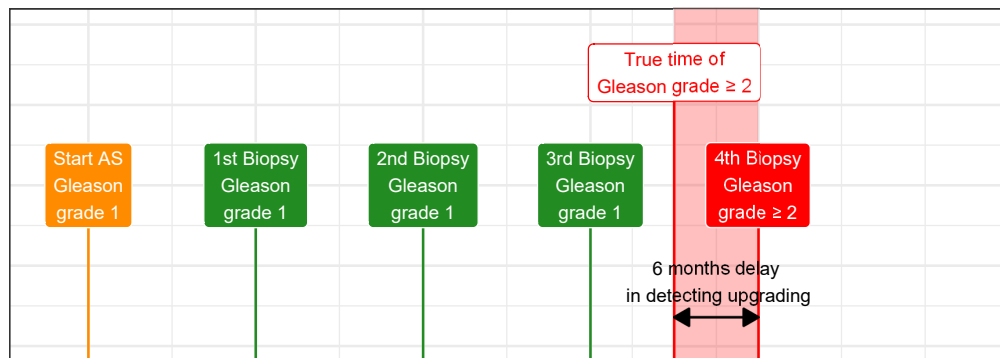
Keywords: Active Surveillance, Biopsies, Personalized Medicine, Prostate Cancer, Shared Decision Making

1. Introduction

Patients with low- and very low-risk screening-detected localized prostate cancer are usually recommended active surveillance (AS) instead of immediate radical treatment [1]. In AS, cancer progression is routinely monitored via prostate-specific antigen (PSA), digital rectal examination, and repeat biopsies. Among these, the strongest indicator of cancer-related outcomes is the biopsy Gleason grade group [2]. When the Gleason grade group increases from group 1 (Gleason 3+3) to 2 (Gleason 3+4) or higher, called *upgrading* [3], patients are commonly advised curative treatment [4].

In most AS protocols, biopsies are scheduled periodically. Consequently,

A Biopsy every year



B Biopsy every 2 years

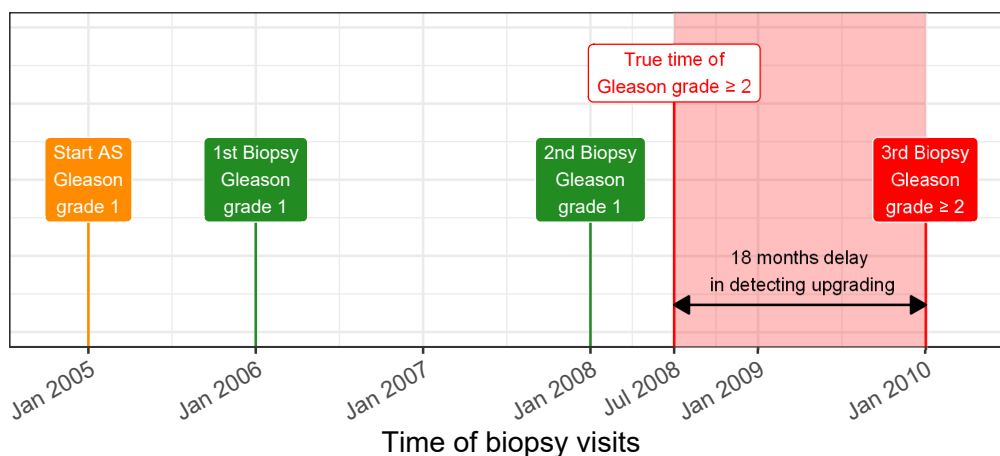


Figure 1: **Trade-off between the number of biopsies and time delay in detecting upgrading (Increase in Gleason grade group from 1 to 2 or higher):** The true time of upgrading for the patient in this figure is July 2008. When biopsies are scheduled annually (**Panel A**), upgrading is detected in January 2009 with a time delay of six months, and a total of four biopsies are scheduled. When biopsies are scheduled biennially (**Panel B**), upgrading is detected in January 2010 with a time delay of 18 months, and a total of three biopsies are scheduled. Since biopsies are conducted periodically, the time of upgrading is observed as an interval. For example, between Jan 2008–Jan 2009 in **Panel A** and between Jan 2008–Jan 2010 in **Panel B**. The phrase ‘Gleason grade group’ is shortened to ‘Gleason grade’ for brevity.

11 upgrading is always detected with a time delay (Figure 1). For detecting
 12 upgrading timely, many AS programs schedule fixed and frequent biopsies
 13 (e.g., annually) for all patients [5, 6]. However, this leads to many unnec-
 14 essary biopsies in slow/non-progressing patients. Biopsies are invasive, may
 15 be painful, and are prone to medical complications such as bleeding and
 16 septicemia[7]. Thus, biopsy burden and patient non-compliance to frequent
 17 biopsies [8] has raised concerns regarding the optimal biopsy schedule [9, 10].
 18 To this end, some cohorts have started using magnetic resonance imaging
 19 (MRI) for deciding biopsies. Although, due to currently limited AS data,
 20 MRI's value is not clear. Others have proposed infrequent schedules such
 21 as biennial biopsies as an alternative [9, 11]. However, fundamental dif-
 22 ferences exist in baseline upgrading-risk across cohorts [9]. Thus, biennial
 23 biopsies may still lead to five unnecessary biopsies over ten years (current
 24 study period of large AS programs) for many slow/non-progressing patients.
 25 A promising alternative to fixed and frequent biopsies is personalized biopsy
 26 schedules based on the patient-specific upgrading-risk (Figure 2).

27 The first challenge in creating personalized biopsy schedules is developing
 28 a statistical model to consolidate accumulated patient data (e.g., PSA, pre-
 29 vious biopsy results) into estimates for upgrading-risk. Existing calculators
 30 for upgrading-risk [12, 13] use only the latest PSA measurement of a patient.
 31 In contrast, more information is captured by considering all repeated mea-
 32 surements of PSA, previous biopsy results, and baseline characteristics of a
 33 patient. To this end, a suitable model is the joint model for time-to-event and
 34 longitudinal data [14, 15, 16]. A joint model predicts the upgrading-risk in a
 35 personalized manner. A subsequent challenge, however, is translating risks

A Should a biopsy be conducted at current visit?

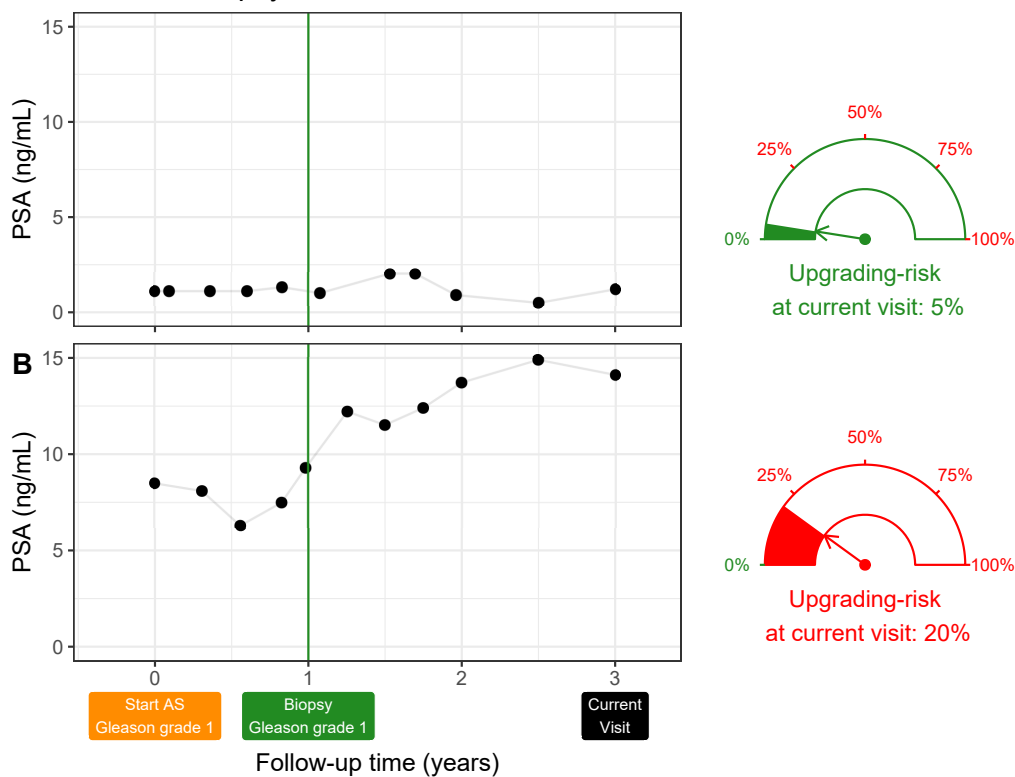


Figure 2: **Motivation for personalized upgrading-risk based decisions of biopsy:** Patient A (**Panel A**) and B (**Panel B**) had their latest biopsy at year one of follow-up (green vertical line). Patient A's prostate-specific antigen (PSA) profile remained stable until his current visit at year three, whereas patient B's profile has shown a rise. Consequently, patient B's upgrading-risk at the current visit (year three) is higher than that of patient A. This makes patient B a more suitable candidate for biopsy than Patient A. Risk estimates in this figure are only illustrative.

into clinical decisions. For example, a 10% upgrading-risk can be perceived high/low depending upon the patient’s age. Patients may also weigh risks of upgrading with the potential *consequences* of another biopsy. Two relevant *consequences* of biopsies (Figure 1) are the timing and the total number of biopsies (burden), and the time delay in detecting upgrading (smaller is beneficial). The relative importance of these *consequences* can vary between the patients, and also over the follow-up period for the same patient.

The goal of this work is to develop a robust, generalizable model that gives reliable estimates for individual upgrading-risk, and to create personalized biopsy schedules based on this risk. To facilitate shared decision making of biopsy schedules, we also aim to provide quantitative estimates of *consequences* of opting for a personalized versus the standard fixed schedule. For developing our model, we will use the world’s largest AS dataset PRIAS. Subsequently, we want to externally validate our model in the largest five AS cohorts from the Movember Foundation’s GAP3 database [17]. Last, we intend to implement our model and methodology in a web-application.

2. Patients and Methods

2.1. Study Cohort

For developing a statistical model to predict upgrading-risk, we used the world’s largest AS dataset, Prostate Cancer International Active Surveillance or PRIAS [4] (Table 1). In PRIAS, PSA was measured quarterly for the first two years of follow-up and semiannually thereafter. Biopsies were scheduled at year one, four, seven, and ten of follow-up. Additional yearly biopsies were scheduled when PSA doubling time was between zero and ten years.

Table 1: **Summary of the PRIAS dataset.** The primary event of interest is upgrading, that is, increase in Gleason grade group from group 1 [2] to 2 or higher. IQR: interquartile range, PSA: prostate-specific antigen. Study protocol URL: <https://www.prias-project.org>

Characteristic	Value
Total centers	> 100
Total patients	7813
Upgrading (primary event)	1134
Treatment	2250
Watchful waiting	334
Loss to follow-up	249
Death (unrelated to prostate cancer)	95
Death (related to prostate cancer)	2
Median age at diagnosis (years)	66 (IQR: 61–71)
Median follow-up period per patient (years)	1.8 (IQR: 0.9–4.0)
Total PSA measurements	67578
Median number of PSA measurements per patient	6 (IQR: 4–12)
Median PSA value (ng/mL)	5.7 (IQR: 4.1–7.7)
Total biopsies	15686
Median number of biopsies per patient	2 (IQR: 1–2)

60 We selected all 7813 patients who had Gleason grade group 1 at the time
 61 of inclusion in PRIAS. Our primary event of interest is an increase in this
 62 Gleason grade group upon repeat biopsy, called *upgrading* (1134 patients).
 63 Upgrading is a trigger for treatment advice in PRIAS. Although, 2250 pa-
 64 tients were provided treatment based on their PSA, or number of biopsy cores
 65 with cancer, or anxiety/other reasons. Our reasons for focusing solely on up-
 66 grading are, namely, upgrading is strongly associated with cancer-related
 67 outcomes, and other triggers for treatment vary between cohorts [5].

68 For model validation, we selected the largest five cohorts from Movember
 69 Foundation’s GAP3 database [17]. These were, namely, the University of
 70 Toronto AS (Toronto), Johns Hopkins AS (Hopkins), Memorial Sloan Ket-
 71 tering Cancer Center AS (MSKCC), King’s College London AS (KCL), and
 72 Michigan Urological Surgery Improvement Collaborative AS (MUSIC). Only
 73 patients with a Gleason grade group 1 at the time of inclusion in these cohorts
 74 were selected (Supplementary A.2).

75 2.2. Statistical Model

76 For developing an upgrading-risk prediction model, the available data in
 77 the PRIAS cohort was patient age at inclusion in AS, longitudinally measured
 78 PSA, timing of repeat biopsies and corresponding Gleason grades, and ob-
 79 served time of upgrading. Analysis of this data required modeling the within-
 80 patient correlation for PSA, the association between the Gleason grades and
 81 PSA profiles of a patient, and handling missing PSA measurements after a
 82 patient experienced upgrading. In such situations, a commonly used model
 83 is the joint model for time-to-event and longitudinal data [14, 15, 16].

84 Our joint model consisted of two sub-models. First, a linear mixed sub-

85 model [18] for longitudinally measured PSA (log-transformed). Second, a
 86 relative-risk sub-model (similar to the Cox model) for obtaining the cause-
 87 specific upgrading-risk. Patient age was included as a predictor in both sub-
 88 models. In the PSA sub-model, we fitted a curve to the PSA measurements
 89 (Panel A, Figure 3). Subsequently, we calculate the mathematical deriva-
 90 tive of the fitted PSA profile over time to obtain his PSA velocity. This
 91 instantaneous velocity varies over follow-up (Panel B, Figure 3), and hence
 92 it is more precise than the widely employed constant PSA velocity [19]. We
 93 modeled the impact of PSA on upgrading-risk by including fitted PSA value
 94 and velocity as predictors in the relative-risk model. Also, the time of the
 95 latest negative biopsy was utilized in the relative-risk sub-model (Panel C,
 96 Figure 3). The parameters of the two sub-models were estimated jointly
 97 (Supplementary A) using the R package **JMbayes** [20].

98 *2.3. Model Validation*

99 We validated our PRIAS based risk prediction model internally in the
 100 PRIAS cohort, and externally using the largest five GAP3 database cohorts
 101 (Section 2.1 and Supplementary A.2). We assessed our model’s ability to
 102 discriminate between patients who experience/do not experience upgrading
 103 via the area under the receiver operating characteristic curve or AUC [21].
 104 We employed calibration plots [22, 23] and mean absolute risk prediction
 105 error [21] to graphically and quantitatively evaluate our model’s risk predic-
 106 tion accuracy. Since AS studies are longitudinal, both AUC and prediction
 107 error vary over follow-up (Supplementary B.1). Lastly, to resolve any po-
 108 tential model miscalibration in validation cohorts, we aimed to recalibrate
 109 our model’s baseline hazard of upgrading (Supplementary A), individually

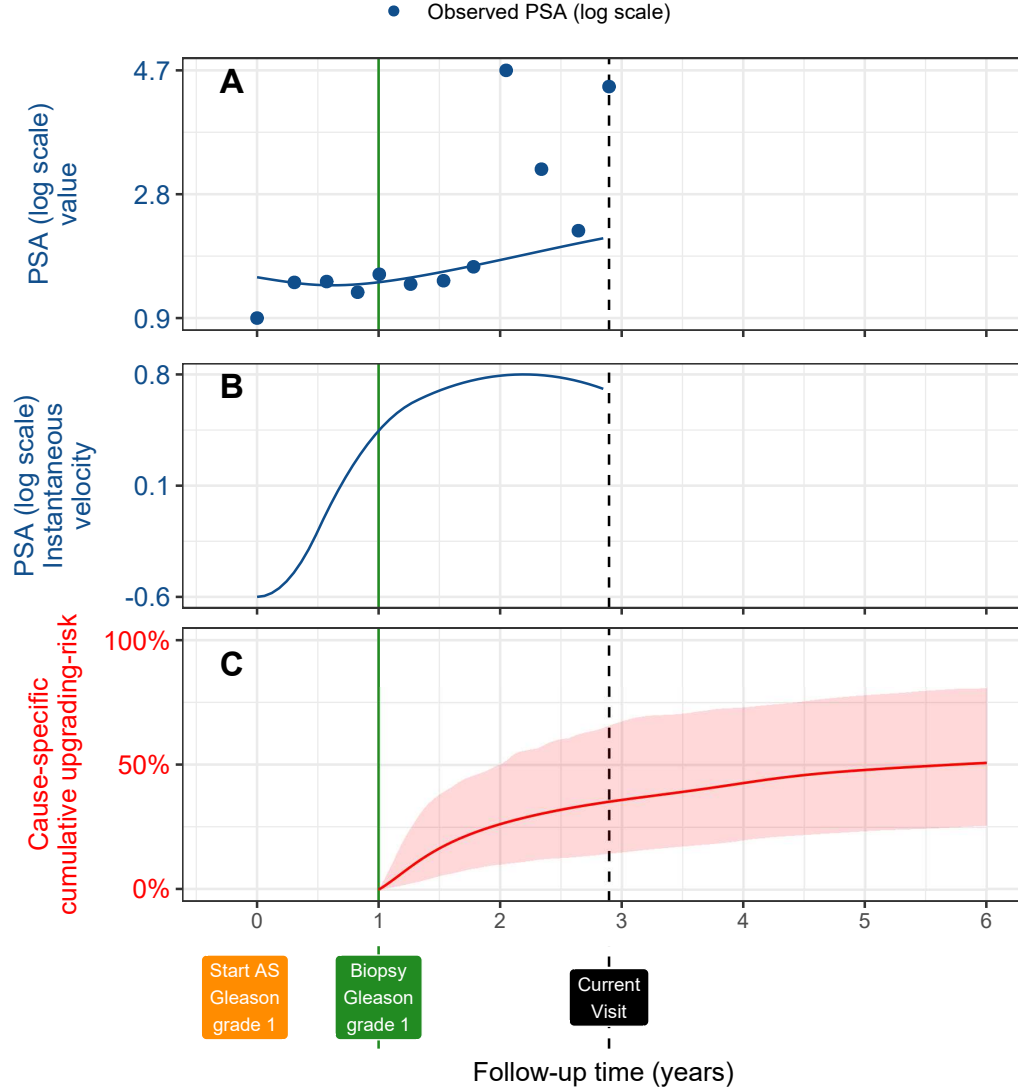


Figure 3: **Illustration of the joint model on a real PRIAS patient.** **Panel A:** Observed PSA (blue dots) and fitted PSA (solid blue line), log-transformed. **Panel B:** Estimated instantaneous velocity of PSA (log-transformed). **Panel C:** Predicted cause-specific cumulative upgrading-risk (95% credible interval shaded). Upgrading is defined as an increase in the Gleason grade group from group 1 [2] to 2 or higher. This upgrading-risk is calculated starting from the time of the latest negative biopsy (vertical green line at year 1 of follow-up). The joint model estimated it by combining the fitted PSA (log scale) value and instantaneous velocity, and time of the latest negative biopsy. Black dashed line at year 3 denotes the time of current visit.

110 for each cohort.

111 3. Results

112 The cause-specific cumulative upgrading-risk at year five of follow-up was
 113 35% in PRIAS, and at most 50% in the five validation cohorts (Panel B,
 114 Figure 4). That is, many patients may not require any biopsy in the first
 115 five years of AS.

116 In the joint model fitted to the PRIAS dataset, the adjusted hazard ratio
 117 of upgrading for an increase in patient age from 61 to 71 years (25-th to
 118 75-th percentile) was 1.45 (95%CI: 1.30–1.63). For an increase in fitted PSA
 119 value (log scale) from 2.36 to 3.07 (25-th to 75-th percentile), the adjusted
 120 hazard ratio was 0.99 (95%CI: 0.89–1.11). In contrast to PSA value, instan-
 121 taneous PSA velocity was a stronger predictor of upgrading-risk, because
 122 an increase in velocity from -0.09 to 0.31 (25-th to 75-th percentile) had a
 123 hazard ratio of 2.47 (95%CI: 1.93–2.99). The impact of PSA value and veloc-
 124 ity on upgrading-risk varied between cohorts (Table 6, Supplementary A.2).
 125 Detailed results are in Supplementary A.2.

126 The follow-up time-dependent mean absolute risk prediction error; time-
 127 dependent AUC; and calibration plot of our model in different validation
 128 cohorts are shown in Panel B, Figure 8, Supplementary B; Panel A, Fig-
 129 ure 4; and Panel B, Figure 4, respectively. In all cohorts, AUC was moderate
 130 (0.55 to 0.75). Mean absolute prediction error was large (0.3 to 0.45) in those
 131 cohorts where the impact of PSA value and velocity on upgrading-risk was
 132 different from PRIAS (e.g., MUSIC cohort, Table 6, Supplementary A.2),
 133 and moderate (0.1 to 0.3) otherwise. To resolve issues in calibration-at-large

(Panel B, Figure 4), we recalibrated the baseline hazard of upgrading in all cohorts (Figure 6, Supplementary B). We compared risk predictions from the recalibrated models with predictions from separately fitted joint models to each cohort (Figure 7, Supplementary B). The difference in predictions was lowest in Johns Hopkins cohort (impact of PSA similar to PRIAS, Table 5, Supplementary A.2). Comprehensive validation results are in Supplementary B.

3.1. *Personalized Biopsy Schedules*

We utilized the fitted joint model to create upgrading-risk based personalized biopsy schedules. To this end, given a new patient’s accumulated PSA measurements (Panel A, Figure 3) and biopsy results, we first predicted his cause-specific cumulative upgrading-risk at his current as well as future PSA follow-up visits (Panel A, Figure 5). These PSA visits occur every six months in PRIAS. Subsequently, we scheduled personalized biopsies on those future follow-up visits of a patient, where his conditional cumulative upgrading-risk was more than a certain threshold (Supplementary C), for example, 10% risk. We maintained a minimum gap of one year between consecutive biopsies (PRIAS recommendation). Example personalized schedules based on 5% and 10% risk thresholds are shown in Panel B, Figure 5, and in Figure 9–11, Supplementary C. Both the risk predictions and resulting personalized schedules were dynamic because they were updated as more follow-up data became available over follow-up (Figure 5, Supplementary B).

The choice of the risk threshold in the personalized schedule dictates the timing and the total number of biopsies, and the expected time delay (Figure 1) in detecting upgrading. We estimated the time delay for both

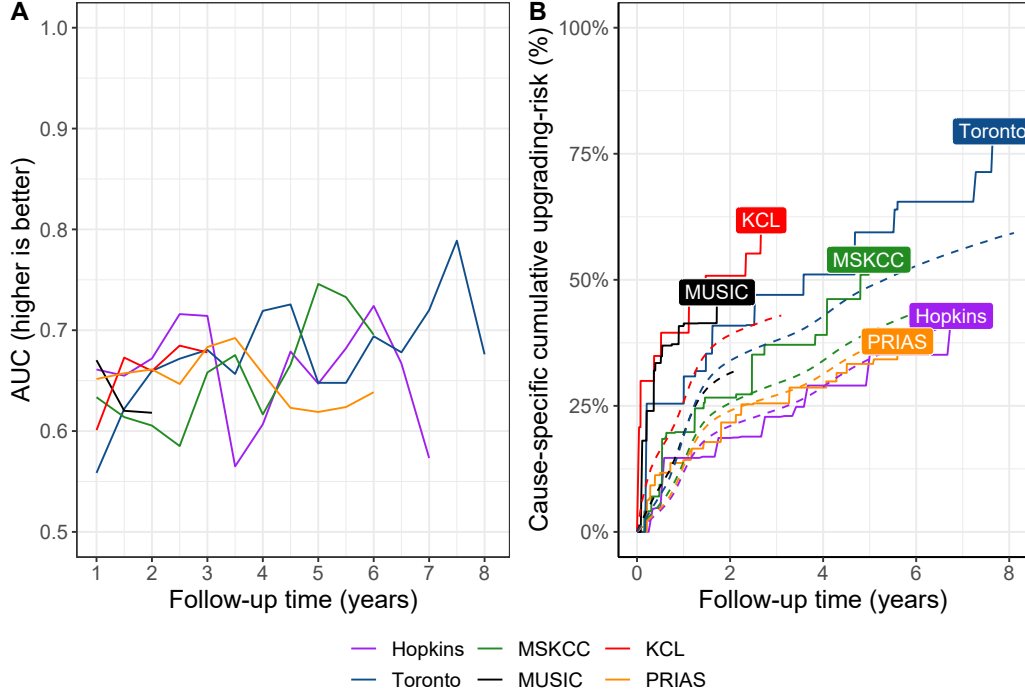


Figure 4: **Model Validation Results.** **Panel A:** time dependent area under the receiver operating characteristic curve or AUC (measure of discrimination). **Panel B:** calibration-at-large indicates model miscalibration. This is because solid lines depicting the non-parametric estimate of the cause-specific cumulative upgrading-risk [24], and dashed lines showing the average cause-specific cumulative upgrading-risk obtained using the joint model fitted to the PRIAS dataset, are not overlapping. Same plot after recalibration is shown in Figure 6, Supplementary B. Full names of Cohorts are *PRIAS*: Prostate Cancer International Active Surveillance, *Toronto*: University of Toronto Active Surveillance, *Hopkins*: Johns Hopkins Active Surveillance, *MSKCC*: Memorial Sloan Kettering Cancer Center Active Surveillance, *KCL*: King's College London Active Surveillance, *MUSIC*: Michigan Urological Surgery Improvement Collaborative Active Surveillance.

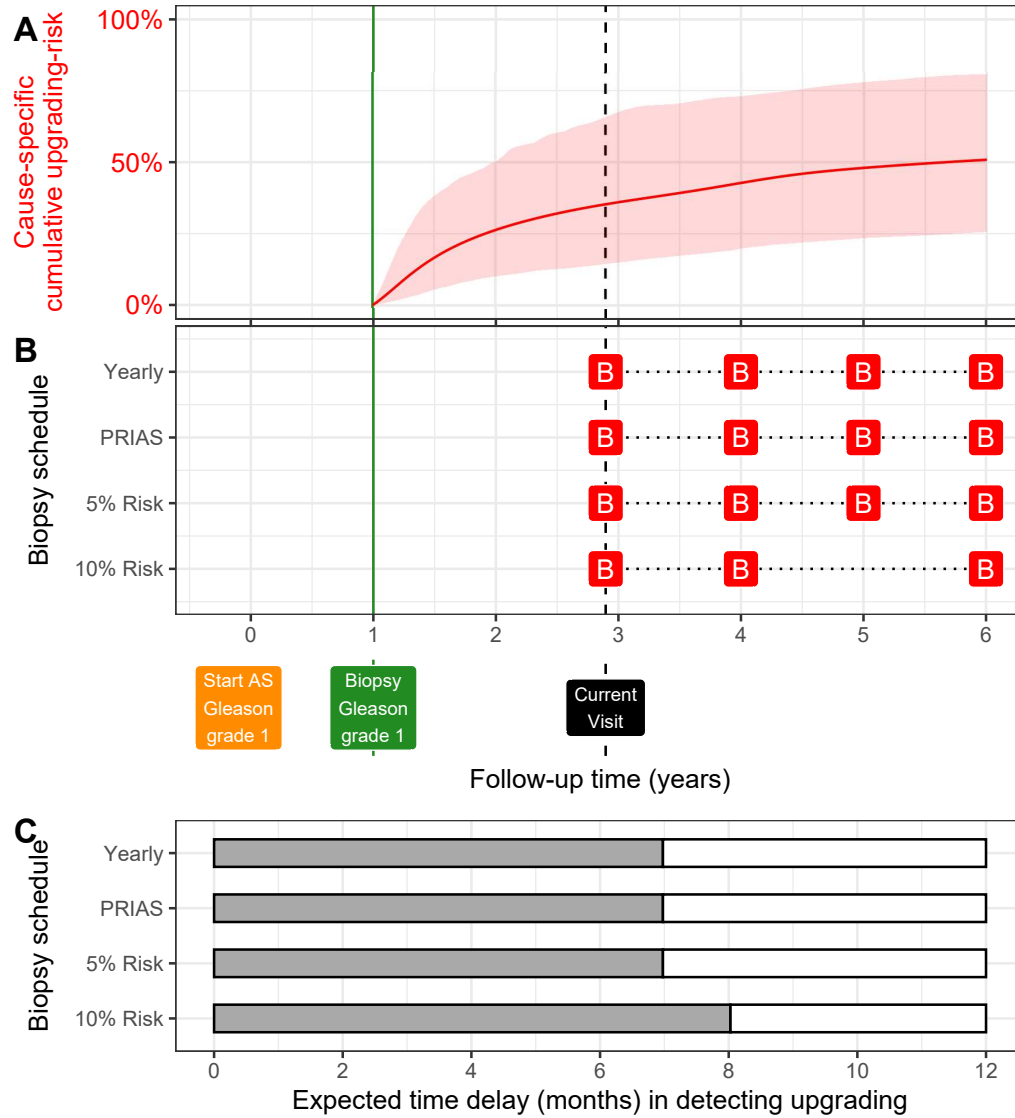


Figure 5: **Illustration of personalized and fixed schedules of biopsies.** Due to a lack of space, the PSA profile of this patient is shown in Figure 3. **Panel A:** Predicted cumulative upgrading-risk (95% credible interval shaded). **Panel B:** Personalized and fixed schedules of biopsies, with a red 'B' indicating a scheduled biopsy. The green vertical line at year 1 denotes the time of the latest negative biopsy. Black dashed line at year 3 denotes the time of the current visit. **Panel C:** Expected time delay in detecting upgrading (months) for different schedules. A compulsory biopsy was scheduled at year six (maximum biopsy scheduling time in PRIAS, Supplementary C) in all schedules for a meaningful comparison between them.

personalized and fixed schedules (Panel C in Figure 5 and Figure 9–11, Supplementary C). Since we estimated the time delay in a personalized manner as well, patients/doctors can compare personalized schedules based on different risk thresholds, with fixed schedules, before making a choice.

3.2. *Web-Application*

We implemented our model and personalized schedules in a user-friendly web-application https://emcbiostatistics.shinyapps.io/prias_biopsy_recommender/. Currently, the web-application supports PRIAS and the five validation cohorts. Patient data can be entered manually and in Microsoft Excel format. Predictions for upgrading-risk are available for a currently limited, cohort-specific, follow-up period (Table 7, Supplementary C). The web-application visualizes the timing of biopsies, and expected time delay in detecting upgrading, for personalized schedules based on 5%, 10%, and 15% risk threshold; annual biopsies; biennial biopsies; and PRIAS schedule.

4. Discussion

We successfully developed and externally validated a model for predicting upgrading-risk [3], and providing risk-based personalized biopsy decisions, in prostate cancer AS. Our work has four novel features over earlier risk calculators [15, 25]. First, our model was fitted to the world’s largest AS dataset PRIAS and externally validated in the largest five cohorts of the Movember Foundation’s GAP3 database [17]. Second, the model predicts a patient’s current and future upgrading-risk in a dynamic and personalized manner. Third, we use the risks to make a personalized schedule, and

also calculate expected time delay in detecting upgrading (less is beneficial) if that schedule is followed. Thus, patients/doctors can compare schedules before making a choice. Fourth, we implemented our methodology in a user-friendly web-application (https://emcbiostatistics.shinyapps.io/prias_biopsy_recommender/) for PRIAS and validated cohorts.

Our PRIAS based model is useful for a large number of patients from the PRIAS and the following validation cohorts: Johns Hopkins AS (Hopkins), Memorial Sloan Kettering Cancer Center AS, King’s College London AS (KCL), and Michigan Urological Surgery Improvement Collaborative AS (MUSIC). The model had a moderate AUC (0.55–0.75), a measure of discrimination, in all validation cohorts. In contrast, the mean absolute risk prediction error varied much more between cohorts. It was moderate in cohorts where the effect size for impact of PSA value and velocity on upgrading-risk was similar to that for PRIAS (e.g., Hopkins cohort). Otherwise, as in the case of KCL or MUSIC cohorts, the prediction error was large. Also, in cohorts with longer follow-up periods, prediction error improved over time as more follow-up data became available. Both KCL and MUSIC cohorts currently have a small follow-up period. Hence, we expect that prediction error will improve in the future with more data. Last, we required recalibration of our model’s baseline hazard of upgrading, individually for all validation cohorts.

The clinical implications of our work are as follows. First, the cause-specific cumulative upgrading-risk at year five of follow-up was at most 50% in all cohorts (Panel B, Figure 4). That is, many patients may not require any biopsy in the first five years of AS. Given the non-compliance and burden of

207 frequent biopsies [8], the availability of our methodology as a web-application
 208 may encourage patients/doctors to consider upgrading-risk based personal-
 209 ized schedules instead. An additional advantage of these schedules is that
 210 they update as more patient data becomes available over follow-up. Fur-
 211 thermore, to assist patients/doctors in choosing between personalized and
 212 fixed schedules, the web-application provides a patient-specific estimate of
 213 time delay in detecting upgrading, for following both personalized and fixed
 214 schedules. We hope that this will objectively address patient apprehensions
 215 regarding adverse outcomes in AS.

216 This work has certain limitations. Predictions for upgrading-risk and per-
 217 sonalized schedules are available only for a currently limited, cohort-specific,
 218 follow-up period (Table 7, Supplementary C). This problem can be mitigated
 219 by refitting the model with new follow-up data in the future. It is important
 220 to differentiate the instantaneous PSA velocity (predictor for upgrading-risk
 221 in our model), from the currently used and criticized constant PSA veloc-
 222 ity [19]. Instantaneous PSA velocity changes over time and hence is more
 223 precise than constant velocity. Along with PSA, in some cohorts recently,
 224 MRI is also used for deciding biopsies. However, the utility of MRI can
 225 only be determined with more follow-up data in the future. Subsequently,
 226 MRI data can also be added as a predictor in our model. Decisions based
 227 on information combined from multiple sources can yield better results than
 228 based on MRI or PSA alone. We scheduled biopsies using cause-specific
 229 cumulative upgrading-risk. Accounting for competing events, such as treat-
 230 ment based on the number of positive biopsy cores, may lead to improved
 231 personalized biopsy decisions. Although, in this work, we did not consider

such additional triggers for treatment because, unlike upgrading, they differ between cohorts [5]. Upgrading is susceptible to inter-observer variation too. Models which account for this variation [15, 26] will be interesting to investigate further. However, the methodology for personalized scheduling, and for comparison of various schedules need not change.

5. Conclusions

We successfully developed and validated a model for predicting upgrading-risk, and providing risk-based personalized biopsy decisions, in prostate cancer AS. The model made available via a user-friendly web-application (https://emcbiostatistics.shinyapps.io/prias_biopsy_recommender/) enables shared decision making of biopsy schedules by comparing fixed and personalized schedules on total biopsies and expected time delay in detecting upgrading. Novel biomarkers and MRI data can be added as predictors in the model to improve predictions in the future. Recalibration of the baseline hazard of upgrading-risk is advised before using the model in cohorts other than the PRIAS cohort.

Author Contributions

Anirudh Tomer had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

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 256 *Drafting of the manuscript:* Tomer, and Rizopoulos
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 278 **tion Plan Prostate Cancer Active Surveillance (GAP3) consortium**

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375 References

- 376 1. Briganti A, Fossati N, Catto JW, Cornford P, Montorsi F, Mottet N,
 377 Wirth M, Van Poppel H. Active surveillance for low-risk prostate cancer:
 378 the European Association of Urology position in 2018. *European urology*
 379 2018;74(3):357–68.

- 380 2. Epstein JI, Egevad L, Amin MB, Delahunt B, Srigley JR, Humphrey PA.
 381 The 2014 international society of urological pathology (isup) consensus
 382 conference on gleason grading of prostatic carcinoma. *The American*
 383 *journal of surgical pathology* 2016;40(2):244–52.

- 384 3. Bruinsma SM, Roobol MJ, Carroll PR, Klotz L, Pickles T, Moore CM,
 385 Gnanapragasam VJ, Villers A, Rannikko A, Valdagni R, et al. Expert
 386 consensus document: semantics in active surveillance for men with lo-
 387 calized prostate cancerresults of a modified delphi consensus procedure.
 388 *Nature reviews urology* 2017;14(5):312.

- 389 4. Bul M, Zhu X, Valdagni R, Pickles T, Kakehi Y, Rannikko A, Bjartell A,
 390 Van Der Schoot DK, Cornel EB, Conti GN, et al. Active surveillance for
 391 low-risk prostate cancer worldwide: the prias study. *European urology*
 392 2013;63(4):597–603.

- 393 5. Nieboer D, Tomer A, Rizopoulos D, Roobol MJ, Steyerberg EW. Active
 394 surveillance: a review of risk-based, dynamic monitoring. *Translational*
 395 *andrology and urology* 2018;7(1):106–15.

- 396 6. Loeb S, Carter HB, Schwartz M, Fagerlin A, Braithwaite RS, Lepor H.

- 397 Heterogeneity in active surveillance protocols worldwide. *Reviews in*
398 *urology* 2014;16(4):202–3.
- 399 7. Loeb S, Vellekoop A, Ahmed HU, Catto J, Emberton M, Nam R, Rosario
400 DJ, Scattoni V, Lotan Y. Systematic review of complications of prostate
401 biopsy. *European urology* 2013;64(6):876–92.
- 402 8. Bokhorst LP, Alberts AR, Rannikko A, Valdagni R, Pickles T, Kakehi Y,
403 Bangma CH, Roobol MJ, PRIAS study group . Compliance rates with
404 the Prostate Cancer Research International Active Surveillance (PRIAS)
405 protocol and disease reclassification in noncompliers. *European Urology*
406 2015;68(5):814–21.
- 407 9. Inoue LY, Lin DW, Newcomb LF, Leonardson AS, Ankerst D, Gulati R,
408 Carter HB, Trock BJ, Carroll PR, Cooperberg MR, et al. Comparative
409 analysis of biopsy upgrading in four prostate cancer active surveillance
410 cohorts. *Annals of internal medicine* 2018;168(1):1–9.
- 411 10. Bratt O, Carlsson S, Holmberg E, Holmberg L, Johansson E, Josefsson
412 A, Nilsson A, Nyberg M, Robinsson D, Sandberg J, et al. The study of
413 active monitoring in sweden (sams): a randomized study comparing two
414 different follow-up schedules for active surveillance of low-risk prostate
415 cancer. *Scandinavian journal of urology* 2013;47(5):347–55.
- 416 11. de Carvalho TM, Heijnsdijk EA, de Koning HJ. Estimating the risks
417 and benefits of active surveillance protocols for prostate cancer: a mi-
418 crosimulation study. *BJU international* 2017;119(4):560–6.

- 419 12. Partin AW, Yoo J, Carter HB, Pearson JD, Chan DW, Epstein JI, Walsh
 420 PC. The use of prostate specific antigen, clinical stage and gleason score
 421 to predict pathological stage in men with localized prostate cancer. *The*
 422 *Journal of urology* 1993;150(1):110–4.
- 423 13. Makarov DV, Trock BJ, Humphreys EB, Mangold LA, Walsh PC, Ep-
 424 stein JI, Partin AW. Updated nomogram to predict pathologic stage of
 425 prostate cancer given prostate-specific antigen level, clinical stage, and
 426 biopsy gleason score (partin tables) based on cases from 2000 to 2005.
 427 *Urology* 2007;69(6):1095–101.
- 428 14. Tomer A, Nieboer D, Roobol MJ, Steyerberg EW, Rizopoulos D. Per-
 429 sonalized schedules for surveillance of low-risk prostate cancer patients.
 430 *Biometrics* 2019;75(1):153–62.
- 431 15. Coley RY, Zeger SL, Mamawala M, Pienta KJ, Carter HB. Prediction
 432 of the pathologic gleason score to inform a personalized management
 433 program for prostate cancer. *European urology* 2017;72(1):135–41.
- 434 16. Rizopoulos D. Joint Models for Longitudinal and Time-to-Event Data:
 435 With Applications in R. CRC Press; 2012. ISBN 9781439872864.
- 436 17. Bruinsma SM, Zhang L, Roobol MJ, Bangma CH, Steyerberg EW,
 437 Nieboer D, Van Hemelrijck M, consortium MFGAPPCASG, Trock B,
 438 Ehdaie B, et al. The movember foundation’s gap3 cohort: a profile of
 439 the largest global prostate cancer active surveillance database to date.
 440 *BJU international* 2018;121(5):737–44.

- 441 18. Laird NM, Ware JH, et al. Random-effects models for longitudinal data.
442 *Biometrics* 1982;38(4):963–74.
- 443 19. Vickers AJ, Savage C, O'Brien MF, Lilja H. Systematic review of pre-
444 treatment prostate-specific antigen velocity and doubling time as pre-
445 dictors for prostate cancer. *Journal of Clinical Oncology* 2009;27(3):398.
- 446 20. Rizopoulos D. The R package JMBayes for fitting joint models for lon-
447 gitudinal and time-to-event data using MCMC. *Journal of Statistical*
448 *Software* 2016;72(7):1–46.
- 449 21. Rizopoulos D, Molenberghs G, Lesaffre EM. Dynamic predictions with
450 time-dependent covariates in survival analysis using joint modeling and
451 landmarking. *Biometrical Journal* 2017;59(6):1261–76.
- 452 22. Royston P, Altman DG. External validation of a cox prognostic
453 model: principles and methods. *BMC medical research methodology*
454 2013;13(1):33.
- 455 23. Steyerberg EW, Vickers AJ, Cook NR, Gerds T, Gonen M, Obuchowski
456 N, Pencina MJ, Kattan MW. Assessing the performance of prediction
457 models: a framework for some traditional and novel measures. *Epidemi-*
458 *ology (Cambridge, Mass)* 2010;21(1):128.
- 459 24. Turnbull BW. The empirical distribution function with arbitrarily
460 grouped, censored and truncated data. *Journal of the Royal Statisti-*
461 *cal Society Series B (Methodological)* 1976;38(3):290–5.
- 462 25. Ankerst DP, Xia J, Thompson Jr IM, Hoefler J, Newcomb LF, Brooks
463 JD, Carroll PR, Ellis WJ, Gleave ME, Lance RS, et al. Precision

- 464 medicine in active surveillance for prostate cancer: development of the
465 canary–early detection research network active surveillance biopsy risk
466 calculator. *European urology* 2015;68(6):1083–8.
- 467 26. Balasubramanian R, Lagakos SW. Estimation of a failure time distribu-
468 tion based on imperfect diagnostic tests. *Biometrika* 2003;90(1):171–82.