Personalized Biopsy Schedules Based on Risk of Gleason Upgrading for Low-Risk Prostate Cancer Active Surveillance Patients*

Anirudh Tomer, MSc^{a,*}, Daan Nieboer, MSc^{b,c}, Monique J. Roobol, PhD^c, Anders Bjartell, MD, PhD^d, Ewout W. Steyerberg, PhD^{b,e}, Dimitris Rizopoulos, PhD^a, Movember Foundation's Global Action Plan Prostate Cancer Active Surveillance (GAP3) consortium^f

Abstract

Objective: To develop a model and methodology for predicting the risk of Gleason *upgrading* in prostate cancer active surveillance (AS) patients, and using the predicted risks to create risk-based *personalized* biopsy schedules as an alternative to one-size-fits-all schedules (e.g., annually). Furthermore,

^aDepartment of Biostatistics, Erasmus University Medical Center, Rotterdam, the Netherlands

^bDepartment of Public Health, Erasmus University Medical Center, Rotterdam, the Netherlands

^cDepartment of Urology, Erasmus University Medical Center, Rotterdam, the Netherlands ^dDepartment of Urology, Skåne University Hospital, Malmö, Sweden

^eDepartment of Biomedical Data Sciences, Leiden University Medical Center, Leiden, the Netherlands

^fThe Movember Foundation's Global Action Plan Prostate Cancer Active Surveillance (GAP3) consortium members presented in Appendix A

^{*}Word count: 3109

^{*}Corresponding author (Anirudh Tomer): Erasmus MC, kamer flex Na-2823, PO Box 2040, 3000 CA Rotterdam, the Netherlands. Tel: $+31\ 10\ 70\ 43393$

Email addresses: a.tomer@erasmusmc.nl (Anirudh Tomer, MSc),

d.nieboer@erasmusmc.nl (Daan Nieboer, MSc), m.roobol@erasmusmc.nl (Monique J. Roobol, PhD), anders.bjartell@med.lu.se (Anders Bjartell, MD, PhD),

 $[\]begin{tabular}{ll} \textbf{e.w.steyerberg@lumc.nl} & (Ewout~W.~Steyerberg,~PhD), \begin{tabular}{ll} \textbf{d.rizopoulos@erasmusmc.nl} \\ & (Dimitris~Rizopoulos,~PhD) \end{tabular}$

to assist patients and doctors in making shared decisions of biopsy schedules, by providing them quantitative estimates of the *burden* and *benefit* of opting for personalized versus any other schedule in AS. Last, to externally validate our model and implement it along with personalized schedules in a ready to use web-application.

Materials and Methods: We used longitudinal prostate-specific antigen (PSA) measurements, timing and results of previous biopsies, and age at baseline from the world's largest AS study, Prostate Cancer Research International Active Surveillance or PRIAS (7813 patients, 1134 experienced upgrading). We fitted a Bayesian joint model for time-to-event and longitudinal data to the PRIAS dataset. We then externally validated our model in the largest six AS cohorts of the Movember Foundation's Global Action Plan (GAP3) database (> 20,000 patients, 27 centers worldwide), covering nearly 73% of all GAP3 patients. We used the predicted upgrading-risks from the validated models to schedule biopsies whenever a patient's risk of upgrading was above a certain threshold. To assist patients in choice of this threshold to compare the resulting schedule with currently practiced schedules, we provided them the timing and the total number of biopsies (burden) planned, and the predicted time delay in detecting upgrading (shorter is better) for each schedule.

Results: The cause-specific cumulative upgrading-risk at year five of follow-up was 35% in PRIAS, and at most 50% in GAP3 cohorts. In the PRIAS based model, PSA velocity was a stronger predictor of upgrading

(Hazard Ratio: 2.47, 95%CI: 1.93–2.99) than PSA value (Hazard Ratio: 0.99, 95%CI: 0.89–1.11). Our model had a moderate area under the receiver operating characteristic curve (0.6–0.7) in validation cohorts. The prediction error was moderate (0.1–0.2) in GAP3 cohorts where the impact of PSA value and velocity on upgrading-risk was similar to PRIAS, but large (0.2–0.3) otherwise. Our model required recalibration of baseline upgrading-risk in validation cohorts. We used predicted upgrading-risk from the validated model to create personalized biopsy schedules for real AS patients and implemented them in a web-application (http://tiny.cc/biopsy).

Conclusions: We successfully developed and validated a model for predicting upgrading-risk, and providing risk-based personalized biopsy decisions, in prostate cancer AS. Personalized prostate biopsies are a novel alternative to fixed one-size-fits-all schedules that may help to reduce unnecessary prostate biopsies while maintaining cancer control. The model and schedules made available via a web-application enable shared decision making of biopsy schedules by comparing fixed and personalized schedules on total biopsies and expected time delay in detecting upgrading.

Keywords: Active Surveillance, Biopsies, Personalized Medicine, Prostate Cancer, Shared Decision Making

1. Introduction

- Patients with low- and very low-risk screening-detected localized prostate
- cancer are usually recommended active surveillance (AS) instead of immedi-
- 4 ate radical treatment [1]. In AS, cancer progression is routinely monitored

via prostate-specific antigen (PSA), digital rectal examination (DRE), repeat biopsies, and recently magnetic resonance imaging (MRI). Among these, the strongest indicator of cancer-related outcomes is the biopsy Gleason grade group [2]. When it increases from group 1 (Gleason 3+3) to 2 (Gleason 3+4) or higher, it is called *upgrading* [3]. Upgrading is a key endpoint in AS upon

which patients are commonly advised curative treatment [4].

In AS, biopsies are always conducted with a time gap between them.

Consequently, upgrading is always detected with a time delay (Figure 1) that

cannot be measured directly. For detecting upgrading timely, many patients

are prescribed fixed and frequent biopsies, most often annually [5]. Such one
size-fits-all schedules lead to unnecessary biopsies in slow/non-progressing

patients. Biopsies are invasive, may be painful, and are prone to medical

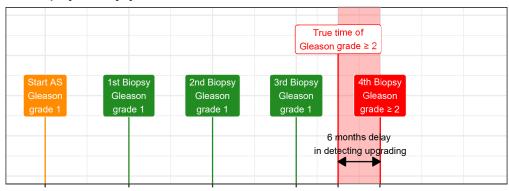
complications such as bleeding and septicemia[6]. Thus, biopsy burden and

patient non-compliance to frequent biopsies [7] have raised concerns regarding

the optimal biopsy schedule [8, 9] in AS.

Apart from the confirmatory biopsy at year one of AS [7], opinions and practice regarding the timing of remaining biopsies lack a consensus [10]. Some support periodical one-size-fits-all biopsy schedules instead of schedules based on clinical data and MRI results [11, 5]. Others suggest infrequent periodical schedules (e.g., biennially) [8, 12]. In contrast, many AS programs utilize patients' observed PSA, DRE, previous biopsy Gleason grade, and lately, MRI results to decide biopsies [13, 4, 10]. However, each of these approaches has limitations. For example, one-size-fits-all schedules can lead to many unnecessary biopsies because of differences in baseline upgrading-risk across cohorts [8]. Whereas, since observed data has measurement error

A Biopsy every year



B Biopsy every 2 years

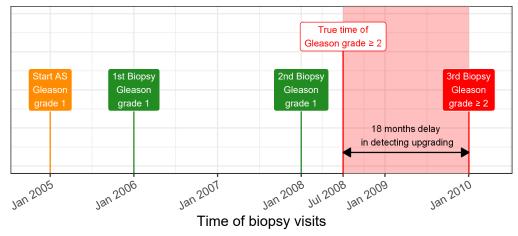


Figure 1: Trade-off between the timing and number of biopsies (burden) and time delay in detecting Gleason upgrading (shorter is better): The true time of Gleason upgrading (increase in Gleason grade group from group 1 to 2 or higher) for the patient in this figure is July 2008. When biopsies are scheduled annually (Panel A), upgrading is detected in January 2009 with a time delay of six months, and a total of four biopsies are scheduled. When biopsies are scheduled biennially (Panel B), upgrading is detected in January 2010 with a time delay of 18 months, and a total of three biopsies are scheduled. Since biopsies are conducted periodically, the time of upgrading is observed as an interval. For example, between Jan 2008–Jan 2009 in Panel A and between Jan 2008–Jan 2010 in Panel B. The phrase 'Gleason grade group' is shortened to 'Gleason grade' for brevity.

(e.g., PSA fluctuations), a flaw of using it directly is that it may lead to poor decisions. Also, typically such decisions rely only on the latest data point and ignore previous repeated measurements. A novel alternative that counters these drawbacks is first processing patient data via a statistical model, and subsequently using model predicted upgrading-risks to create *personalized* biopsy schedules [10] (Figure 2). While, upgrading-risk calculators are not new [14, 15, 16, 17], not all are personalized either. Besides, they do not specify how risk predictions can be exploited to create a schedule.

The work is motivated by the problem of scheduling biopsies in AS, and our goal is two-fold. First, we want to assist practitioners in using clinical data in biopsy decisions in a statistically sound manner. To this end, we plan to develop a robust, generalizable statistical model that provides reliable individual upgrading-risk in AS. Subsequently, we will employ these predictions to derive risk-based personalized biopsy schedules. Our second goal is to enable shared decision making of a biopsy schedule. We intend to achieve this by allowing patients and doctors to compare the burden and benefit (Figure 1) of opting for personalized schedules versus periodical schedules versus schedules based on clinical data. Specifically, we propose timing and number of planned biopsies (more/frequent are burdensome), and the expected time delay in detecting upgrading (shorter is beneficial) for any given schedule. In this whole process, we want to capture the maximum possible information from the available data. Thus, we will exploit all repeated measurements of patients, previous biopsy results, baseline characteristics, and keep our model flexible to accommodate novel biomarkers in the future. To fit this model, we will use data of the world's largest AS study, Prostate Cancer

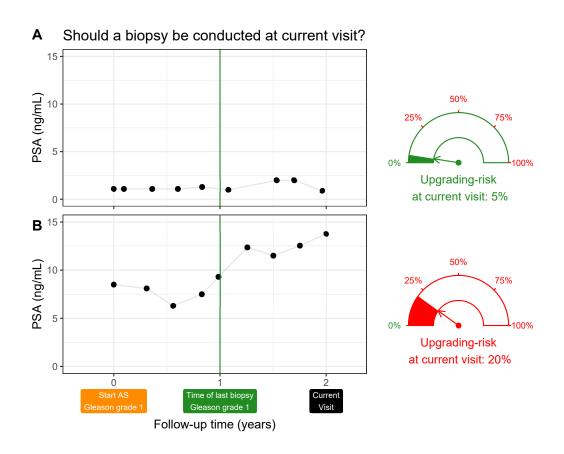


Figure 2: Motivation for personalized upgrading-risk based biopsy decisions: To combine patients' complete longitudinal data and results from previous biopsies into estimates for risk of Gleason upgrading and using these estimates to schedule biopsies. For example, Patient A (Panel A) and B (Panel B) had their latest biopsy at year one of follow-up (green vertical line). Patient A's prostate-specific antigen (PSA) profile remained stable until his current visit at year two, whereas patient B's profile has shown a rise. Consequently, patient B's upgrading-risk at the current visit (year two) is higher than that of patient A. This makes patient B a more suitable candidate for biopsy than Patient A. Risk estimates in this figure are only illustrative.

Research International Active Surveillance (PRIAS). To evaluate our model, we will externally validate it in the largest six AS cohorts from the Movember Foundation's Global Action Plan (GAP3) database [18]. Last, we aim to implement the validated model and methodology in a web-application.

59 2. Patients and Methods

60 2.1. Study Cohort

For developing a statistical model to predict upgrading-risk, we used the world's largest AS dataset, Prostate Cancer International Active Surveillance or PRIAS [4], dated April 2019 (Table 1). In PRIAS, biopsies were scheduled at year one, four, seven, ten, and additional yearly biopsies were scheduled when PSA doubling time was between zero and ten years. We selected all 7813 patients who had Gleason grade group 1 at inclusion in AS. Our primary event of interest is an increase in this Gleason grade group observed upon repeat biopsy, called *upgrading* (1134 patients). Upgrading is a trigger for treatment advice in PRIAS. Also, 2250 patients were provided treatment based on their PSA, the number of biopsy cores with cancer, or anxiety/other reasons. However, our reasons for focusing solely on upgrading are that upgrading is strongly associated with cancer-related outcomes, and other treatment triggers vary between cohorts [10]. For externally validating our model's predictions, we selected the following largest (by the number of repeated measurements) six cohorts from Movember Foundation's GAP3 database [18] version 3.1, covering nearly 73% of the GAP3 patients: the University of Toronto AS (Toronto), Johns Hopkins AS (Hopkins), Memorial Sloan Kettering Cancer Center AS (MSKCC),

King's College London AS (KCL), Michigan Urological Surgery Improvement
Collaborative AS (MUSIC), and University of California San Francisco AS
(UCSF, version 3.2). Only patients with a Gleason grade group 1 at the time
of inclusion in these cohorts were selected. Summary statistics are presented
in Supplementary A.2.

Choice of predictors:. In our model, we utilized all repeated PSA measurements, timing and previous biopsy Gleason grade, and age at inclusion in AS. Other predictors such as prostate volume, MRI results may also be important. While MRI is already used for targeting biopsies, but currently, there are arguments both for and against its use in deciding the time of biopsies [13, 11, 19]. MRI is still a recent addition in most AS protocols, and thus, repeated MRI data is very sparsely available in both PRIAS and GAP3 databases to make a stable prediction model. Prostate volume data is also sparsely available, especially in validation cohorts. Based on these reasons, we did not include them in our model. However, the model we propose next is extendable to include MRI and other novel biomarkers in the future.a

95 2.2. Statistical Model

Modeling an AS dataset such as PRIAS, posed certain challenges. First,
PSA was measured longitudinally, and over follow-ups, it did not always increase linearly. Also, PSA was available only until a patient observed upgrading. Hence, we need to accommodate the within-patient correlation for PSA,
the association between the Gleason grades and PSA profiles of a patient,
and handle missing PSA measurements after a patient experienced upgrading. Second, since the PRIAS biopsy schedule uses PSA, a patient's observed

Table 1: **Summary of the PRIAS dataset as of April 2019**. The primary event of interest is upgrading, that is, increase in Gleason grade group from group 1 [2] to 2 or higher. IQR: interquartile range, PSA: prostate-specific antigen. Study protocol URL: https://www.prias-project.org

Characteristic	Value
Total centers	> 100
Total patients	7813
Upgrading (primary event)	1134
Treatment	2250
Watchful waiting	334
Loss to follow-up	249
Death (unrelated to prostate cancer)	95
Death (related to prostate cancer)	2
Median age at diagnosis (years)	66 (IQR: 61-71)
Median maximum follow-up per patient (years)	1.8 (IQR: 0.9-4.0)
Total PSA measurements	67578
Median number of PSA measurements per patient	6 (IQR: 4–12)
Median PSA value (ng/mL)	5.7 (IQR: 4.1–7.7)
Total biopsies	15686
Median number of biopsies per patient	2 (IQR: 1–2)

time of upgrading also depends on their PSA. That is, while estimating the
effect of PSA on the upgrading-risk, we have to adjust for the effect of PSA on
the observed times of upgrading. Third, many patients obtained treatment
and watchful waiting. Since we considered events other than upgrading as
censoring, the model needs to account for patients' reasons for treatment or
watchful waiting (e.g., age, treatment based on observed data). A model that
handles these challenges in a statistically sound manner is the joint model
for time-to-event and longitudinal data [20, 14, 21].

Our joint model consisted of two sub-models. Namely, a linear mixed-111 effects sub-model [22] for longitudinally measured PSA (log-transformed), and a relative-risk sub-model (similar to the Cox model) for the interval-113 censored time of upgrading. Patient age was used in both sub-models. 114 Whereas, results and timing of the previous negative biopsies were used 115 only in the risk sub-model (Panel C, Figure 3). To account for PSA fluctuations [23], we assumed t-distributed measurement errors for PSA. The 117 correlation between PSA measurements of a patient was established using 118 patient-specific random-effects. We fitted a unique curve to the PSA mea-119 surements of each patient (Panel A, Figure 3). Subsequently, we calculated the mathematical derivative of the patient's fitted PSA profile (Equation 2, Supplementary A), to obtain his follow-up time specific instantaneous PSA 122 velocity (Panel B, Figure 3). This instantaneous velocity is a stronger predictor of upgrading than the widely used average PSA velocity [24]. We modeled the impact of PSA on upgrading-risk by employing fitted PSA value and instantaneous velocity as predictors in the risk sub-model. However, to adjust these effects for the PSA dependent PRIAS biopsy schedule, we used

a full likelihood method for parameter estimation. This approach also accommodates watchful waiting and treatment protocols that are also based
on patient data. Specifically, the parameters of our two sub-models were
estimated jointly under the Bayesian paradigm (Supplementary A) using the
R package JMbayes [25].

2.3. Risk Prediction and Model Validation

Our model provides predictions for upgrading-risk over the entire future follow-up period of a patient (Panel C, Figure 3). However, we recommend 135 using predictions after year one. This is because most AS programs recom-136 mend a confirmatory biopsy at year one, especially to detect patients who 137 may be misdiagnosed as low-grade at inclusion in AS. The model also automatically updates risk-predictions over follow-up as more patient data becomes available (Figure 5, Supplementary B). We validated our model inter-140 nally in the PRIAS cohort, and externally in the largest six GAP3 database 141 cohorts. We employed calibration plots [26, 27] and follow-up time-dependent 142 mean absolute risk prediction error or MAPE [28] to graphically and quantitatively evaluate our model's risk prediction accuracy, respectively. We assessed our model's ability to discriminate between patients who experi-145 ence/do not experience upgrading via the time-dependent area under the 146 receiver operating characteristic curve or AUC [28]. 147

The aforementioned *time-dependent* AUC and MAPE [28] are temporal extensions of their standard versions [27] in a longitudinal setting. Specifically, at every six months of follow-up, we calculated a unique AUC and MAPE for predicting upgrading-risk in the subsequent one year (Supplementary B.1). For emulating a realistic situation, we calculated the AUC and

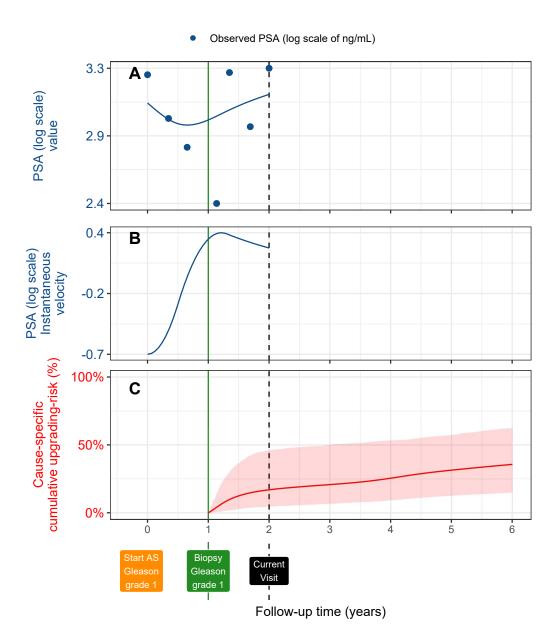


Figure 3: Illustration of the joint model on a real PRIAS patient. Panel A: Observed PSA (blue dots) and fitted PSA (solid blue line), log-transformed from ng/mL. Panel B: Estimated instantaneous velocity of PSA (log-transformed). Panel C: Predicted cause-specific cumulative upgrading-risk (95% credible interval shaded). Upgrading is defined as an increase in the Gleason grade group from group 1 [2] to 2 or higher. This upgrading-risk is calculated starting from the time of the latest negative biopsy (vertical green line at year one of follow-up). The joint model estimated it by combining the fitted PSA (log scale) value and instantaneous velocity, and time of the latest negative biopsy. Black dashed line at year two denotes the time of current visit.

MAPE at each follow-up using only the validation data available until that follow-up. Last, to resolve any potential model miscalibration in validation cohorts, we aimed to recalibrate our model's baseline hazard of upgrading (Supplementary B.1), individually for each cohort.

3. Results

The cause-specific cumulative upgrading-risk at year five of follow-up was 158 35% in PRIAS and at most 50% in validation cohorts (Panel B, Figure 4). 159 In the fitted PRIAS model, the adjusted hazard ratio (aHR) of upgrading for an increase in patient age from 61 to 71 years (25-th to 75-th percentile) was 161 1.45 (95%CI: 1.30–1.63). For an increase in fitted PSA value from 2.36 to 3.07 162 (25-th to 75-th percentile, log scale), the aHR was 0.99 (95%CI: 0.89-1.11). 163 The strongest predictor of upgrading-risk was instantaneous PSA velocity, with an increase from -0.09 to 0.31 (25-th to 75-th percentile), giving an aHR of 2.47 (95%CI: 1.93–2.99). The aHR for PSA value and velocity was 166 different in each GAP3 cohort (Supplementary Table 8). 167 The time-dependent AUC, calibration plot, and time-dependent MAPE of 168 our model are shown in Figure 4, and Supplementary Figure 8. In all cohorts, time-dependent AUC was moderate (0.6 to 0.7) over the whole follow-up period. Time-dependent MAPE was moderate (0.1 to 0.2) in those cohorts 171 where the impact of PSA on upgrading-risk was similar to PRIAS (e.g., Hopkins cohort, Supplementary Table 8), and large (0.2 to 0.3) otherwise. 173 Our model was miscalibrated for validation cohorts (Panel B, Figure 4). Recalibrating the baseline hazard of upgrading in validation cohorts resolved this issue (Supplementary Figure 6). We compared risk predictions from the recalibrated models, with predictions from separately fitted cohort-specific joint models (Supplementary Figure 7). The difference in predictions was lowest in the Johns Hopkins cohort (impact of PSA on upgrading-risk similar to PRIAS). Comprehensive results are in Supplementary A.2 and B.

$_{181}$ 3.1. Personalized Biopsy Schedules

We employed the PRIAS based fitted model to create personalized biopsy 182 schedules for real PRIAS patients. Particularly, first using the model and patient's observed data, we predicted his cumulative upgrading-risk (Figure 5) 184 on all of his future follow-up visits (biannually in PRIAS). Subsequently, 185 we planned biopsies on those future visits where his conditional cumulative upgrading-risk was more than a certain threshold (see Supplementary C for mathematical details). The choice of this threshold dictates the timing of biopsies in a risk-based personalized schedule. For example, personalized 189 schedules based on 5% and 10% risk thresholds are shown in Figure 5, and 190 in Supplementary Figure 10–12. 191

To facilitate the choice of a risk-threshold, and for comparing the consequences of opting for a risk-based schedule versus any other schedule (e.g., annual, PRIAS), we predict expected time delay in detecting upgrading if a schedule is followed. This delay can be predicted for any schedule. For example, in Panel C of Figure 5, the annual schedule has the least expected delay. In contrast, a personalized schedule based on a 10% risk threshold has a slightly larger expected delay, but it also schedules much fewer biopsies. An important aspect of this delay is that it is personalized as well. This is because it is estimated using all available clinical data of the patient (see Supplementary C). That is, even if two different patients are prescribed the

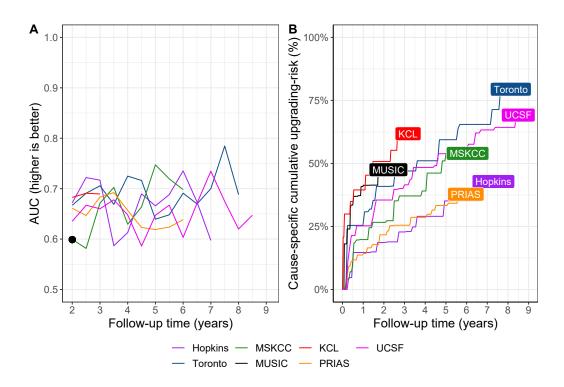


Figure 4: Model Validation Results. Panel A: time-dependent area under the receiver operating characteristic curve or AUC (measure of discrimination). Panel B: calibration-at-large indicates model miscalibration. This is because solid lines depicting the non-parameteric estimate of the cause-specific cumulative upgrading-risk [29], and dashed lines showing the average cause-specific cumulative upgrading-risk obtained using the joint model fitted to the PRIAS dataset, are not overlapping. Recalibrating the baseline hazard of upgrading resolved this issue (Supplementary Figure 6). Full names of Cohorts are PRIAS: Prostate Cancer International Active Surveillance, Toronto: University of Toronto Active Surveillance, Hopkins: Johns Hopkins Active Surveillance, MSKCC: Memorial Sloan Kettering Cancer Center Active Surveillance, KCL: King's College London Active Surveillance, MUSIC: Michigan Urological Surgery Improvement Collaborative Active Surveillance, UCSF: University of California San Francisco AS.

same biopsy schedule, their expected delays will be different. While the timing and the total number of planned biopsies denote the burden of a schedule,
a shorter expected time delay in detecting upgrading can be a benefit. These
two, along with other measures such as a patient's comorbidities, anxiety,
etc., can help to make an informed decision of biopsy.

207 3.2. Web-Application

We implemented the PRIAS based model and recalibrated models for GAP3 cohorts, and personalized schedules in a user-friendly web-application https://emcbiostatistics.shinyapps.io/prias_biopsy_recommender/.
This application also works on mobile devices. Patient data can be entered in Microsoft Excel format. The maximum follow-up time up to which predictions can be obtained depends on each cohort (Supplementary Table 9).
The web-application supports personalized, annual, and PRIAS schedules. For personalized schedules, users can control the choice of risk-threshold and compare the risk-based schedule's timing of biopsies, and expected time delay in detecting upgrading, with annual and PRIAS schedule before making a decision.

19 4. Discussion

We successfully developed and externally validated a statistical model for predicting upgrading-risk [3] in prostate cancer AS, and providing risk-based personalized biopsy decisions. Our work has four novel features over earlier risk calculators [14, 15]. First, our model was fitted to the world's largest AS dataset PRIAS and externally validated in the largest six cohorts of the Movember Foundation's GAP3 database [18]. Second, the model predicts a

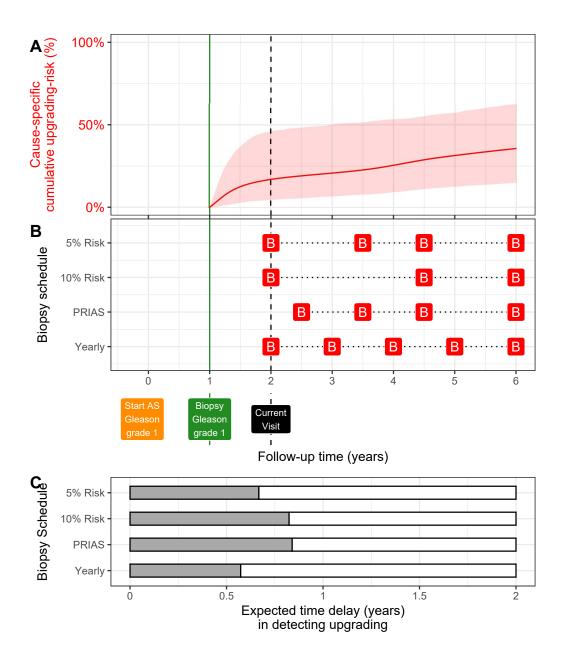


Figure 5: Illustration of personalized and fixed schedules of biopsies for patient from Figure 3. Panel A: Predicted cumulative upgrading-risk (95% credible interval shaded). Panel B: Different biopsy schedules with a red 'B' indicating a future biopsy. Risk: 5% and Risk: 10% are personalized schedules in which a biopsy is planned whenever the conditional cause-specific cumulative upgrading-risk is above 5% or 10% risk, respectively. Green vertical line at year one is the time of the latest negative biopsy. Black dashed line at year two denotes the time of the current visit. Panel C: Expected time delay in detecting upgrading (years) if patient progresses before year six. A compulsory biopsy was scheduled at year six (maximum biopsy scheduling time in PRIAS, Supplementary C) in all schedules for a meaningful comparison between them.

patient's current and future upgrading-risk in a personalized manner. Third, using the predicted risks, we created personalized biopsy schedules and also calculated the expected time delay in detecting upgrading (less is beneficial) if that schedule was followed. Thus, patients/doctors can compare schedules before making a choice. Fourth, we implemented our methodology in a user-friendly web-application (https://emcbiostatistics.shinyapps.io/prias_biopsy_recommender/) for both PRIAS and validated cohorts.

Our model and methods can be useful for numerous patients from PRIAS 233 and validated GAP3 cohorts. The model utilizes all repeated PSA measurements, results of previous biopsies, and baseline characteristics of a patient. Although we could not include MRI and PSA volume because of sparsely 236 available data in both PRIAS and GAP3 databases, our model is extendable to include them in the near future. The current discrimination ability of our model, exhibited by the time-dependent AUC, was between 0.6 and 0.7 over-follow. While this is moderate, it is also so because unlike the standard AUC [27] the time-dependent AUC is more conservative as it utilizes only the validation data available until the time at which it is calculated. The same holds for the time-dependent MAPE (mean absolute prediction error), although it varied much more between cohorts than AUC. In cohorts where the effect size for the impact of PSA value and velocity on upgrading-risk 245 was similar to that for PRIAS (e.g., Hopkins cohort), MAPE was moderate. 246 Otherwise, MAPE was large (e.g., KCL and MUSIC cohorts). We required 247 recalibration of our model's baseline hazard of upgrading for all validation cohorts. The PRIAS and validation cohorts cover nearly 73% of all GAP3 patients.

The clinical implications of our work are as follows. First, the cause-251 specific cumulative upgrading-risk at year five of follow-up was at most 50% 252 in all cohorts (Panel B, Figure 4). That is, many patients may not re-253 quire some of the biopsies planned in the first five years of AS. Given the non-compliance and burden of frequent biopsies [7], the availability of our 255 methodology as a web-application may encourage patients/doctors to con-256 sider upgrading-risk based personalized schedules instead. An additional ad-257 vantage of personalized schedules is that they update as more patient data 258 becomes available over follow-up. We have shown via a simulation study [30] that personalized schedules plan, on average, six fewer biopsies compared to 260 annual schedule and two fewer biopsies than the PRIAS schedule in slow/non-261 progressing AS patients, while maintaining almost the same time delay in 262 detecting upgrading as PRIAS schedule. Personalized schedules with dif-263 ferent risk thresholds indeed have different performance. In this regard, to assist patients/doctors in choosing between fixed schedules and personalized 265 schedules based on different risk thresholds, the web-application provides a 266 patient-specific estimate of the expected time delay in detecting upgrading, 267 for both personalized and fixed schedules. We hope that this will objectively 268 address patient apprehensions regarding adverse outcomes in AS. Last, we note that our web-application should only be used to schedule biopsies after 270 the first compulsory confirmatory biopsy at year one of follow-up. 271

This work has certain limitations. Predictions for upgrading-risk and personalized schedules are available only for a currently limited, cohort-specific, follow-up period (Supplementary Table 9). This problem can be mitigated by refitting the model with new follow-up data in the future. Recently, some

272

cohorts started utilizing MRI to explore the possibility of targeting visible lesions by biopsy. Presently, the GAP3 database has limited MRI follow-277 up data available. As more such data becomes available, the current model 278 can be extended to include MRI based predictors. We scheduled biopsies using cause-specific cumulative upgrading-risk, which ignores competing events 280 such as treatment based on the number of positive biopsy cores. Employing a 281 competing-risk model may lead to improved personalized schedules. Upgrad-282 ing is susceptible to inter-observer variation too. Models which account for 283 this variation [14, 31] will be interesting to investigate further. Even with an enhanced risk prediction model, the methodology for personalized scheduling 285 and calculation of expected time delay (Supplementary C) need not change. 286 Last, our web-application only allows uploading patient data in Microsoft Excel format. Connecting it with patient databases can improve usability. However, a validated upgrading-risk calculator and risk-based biopsy schedules implemented in a web-application is a significant first step towards more informed biopsy decisions in AS.

5. Conclusions

We successfully developed a statistical model and methodology for predicting upgrading-risk, and providing risk-based personalized biopsy decisions, in prostate cancer AS. We externally validated our model, covering nearly 73% patients from the Movember Foundations' GAP3 database.

The model made available via a user-friendly web-application (https://
emcbiostatistics.shinyapps.io/prias_biopsy_recommender/) enables shared
decision making of biopsy schedules by comparing fixed and personalized

- schedules on total biopsies and expected time delay in detecting upgrading.
- Novel biomarkers and MRI data can be added as predictors in the model to
- 302 improve predictions in the future. Recalibration of baseline upgrading-risk
- 303 is advised for cohorts not validated in this work.

304 Author Contributions

- Anirudh Tomer had full access to all the data in the study and takes
- responsibility for the integrity of the data and the accuracy of the data anal-
- 307 ysis.
- 308 Study concept and design: Tomer, Nieboer, Roobol, Bjartell, and Rizopoulos
- 309 Acquisition of data: Tomer, Nieboer, and Roobol
- 310 Analysis and interpretation of data: Tomer, Nieboer, and Rizopoulos
- 311 Drafting of the manuscript: Tomer, and Rizopoulos
- 312 Critical revision of the manuscript for important intellectual content: Tomer,
- Nieboer, Roobol, Bjartell, Steyerberg, and Rizopoulos
- 314 Statistical analyses: Tomer, Nieboer, Steverberg, and Rizopoulos
- 315 Obtaining funding: Roobol, Steverberg, and Rizopoulos
- 316 Administrative, technical or material support: Nieboer
- 317 Supervision: Roobol, and Rizopoulos
- Other: none

Acknowledgments

- We thank Jozien Helleman from the Department of Urology, Erasmus
- University Medical Center, for coordinating the project. The first and last
- authors would like to acknowledge support by Nederlandse Organisatie voor

Wetenschappelijk Onderzoek (the national research council of the Netherlands) VIDI grant nr. 016.146.301, and Erasmus University Medical Center funding. Part of this work was carried out on the Dutch national einfrastructure with the support of SURF Cooperative. The authors also thank the Erasmus University Medical Center's Cancer Computational Biology Center for giving access to their IT-infrastructure and software that was used for the computations and data analysis in this study.

The PRIAS website is funded by the Prostate Cancer Research Foundation, Rotterdam (SWOP). We would like to thank the PRIAS consortium for enabling this research project. This work was supported by the Movember Foundation. The funder did not play any role in the study design, collection, analysis or interpretation of data, or in the drafting of this paper.

Appendix A. Members of The Movember Foundation's Global Action Plan Prostate Cancer Active Surveillance (GAP3) consortium

Principle Investigators: Bruce Trock (Johns Hopkins University, The
James Buchanan Brady Urological Institute, Baltimore, USA), Behfar Ehdaie
(Memorial Sloan Kettering Cancer Center, New York, USA), Peter Carroll (University of California San Francisco, San Francisco, USA), Christopher Filson (Emory University School of Medicine, Winship Cancer Institute, Atlanta, USA), Jeri Kim / Christopher Logothetis (MD Anderson
Cancer Centre, Houston, USA), Todd Morgan (University of Michigan and
Michigan Urological Surgery Improvement Collaborative (MUSIC), Michigan, USA), Laurence Klotz (University of Toronto, Sunnybrook Health Sciences Centre, Toronto, Ontario, Canada), Tom Pickles (University of British

Columbia, BC Cancer Agency, Vancouver, Canada), Eric Hyndman (University of Calgary, Southern Alberta Institute of Urology, Calgary, Canada), 348 Caroline Moore (University College London & University College London Hospital Trust, London, UK), Vincent Gnanapragasam (University of Cambridge & Cambridge University Hospitals NHS Foundation Trust, Cam-351 bridge, UK), Mieke Van Hemelrijck (King's College London, London, UK 352 & Guy's and St Thomas' NHS Foundation Trust, London, UK), Prokar 353 Dasgupta (Guy's and St Thomas' NHS Foundation Trust, London, UK), Chris Bangma (Erasmus Medical Center, Rotterdam, The Netherlands/representative of Prostate cancer Research International Active Surveillance 356 (PRIAS) consortium), Monique Roobol (Erasmus Medical Center, Rotter-357 dam, The Netherlands/representative of Prostate cancer Research International Active Surveillance (PRIAS) consortium), The PRIAS study group, 359 Arnauld Villers (Lille University Hospital Center, Lille, France), Antti Rannikko (Helsinki University and Helsinki University Hospital, Helsinki, Fin-361 land), Riccardo Valdagni (Department of Oncology and Hemato-oncology, Università degli Studi di Milano, Radiation Oncology 1 and Prostate Cancer Program, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy), Antoinette Perry (University College Dublin, Dublin, Ireland), Jonas Hugosson (Sahlgrenska University Hospital, Göteborg, Sweden), Jose Rubio-Briones 366 (Instituto Valenciano de Oncología, Valencia, Spain), Anders Bjartell (Skåne 367 University Hospital, Malmö, Sweden), Lukas Hefermehl (Kantonsspital Baden, 368 Baden, Switzerland), Lee Lui Shiong (Singapore General Hospital, Singapore, Singapore), Mark Frydenberg (Monash Health; Monash University, Melbourne, Australia), Yoshiyuki Kakehi / Mikio Sugimoto (Kagawa Uni-

```
versity Faculty of Medicine, Kagawa, Japan), Byung Ha Chung (Gangnam
   Severance Hospital, Yonsei University Health System, Seoul, Republic of Ko-
373
   rea)
374
       Pathologist: Theo van der Kwast (Princess Margaret Cancer Centre,
375
   Toronto, Canada).
                        Technology Research Partners: Henk Obbink (Royal
376
   Philips, Eindhoven, the Netherlands), Wim van der Linden (Royal Philips,
377
   Eindhoven, the Netherlands), Tim Hulsen (Royal Philips, Eindhoven, the
378
   Netherlands), Cees de Jonge (Royal Philips, Eindhoven, the Netherlands).
379
       Advisory Regional statisticians: Mike Kattan (Cleveland Clinic, Cleve-
380
   land, Ohio, USA), Ji Xinge (Cleveland Clinic, Cleveland, Ohio, USA), Ken-
381
   neth Muir (University of Manchester, Manchester, UK), Artitaya Lophatananon
382
   (University of Manchester, Manchester, UK), Michael Fahey (Epworth Health-
383
   Care, Melbourne, Australia), Ewout Steverberg (Erasmus Medical Center,
   Rotterdam, The Netherlands), Daan Nieboer (Erasmus Medical Center, Rot-
   terdam, The Netherlands); Liying Zhang (University of Toronto, Sunnybrook
386
   Health Sciences Centre, Toronto, Ontario, Canada)
387
       Executive Regional statisticians: Ewout Steverberg (Erasmus Medical
388
   Center, Rotterdam, The Netherlands), Daan Nieboer (Erasmus Medical Cen-
   ter, Rotterdam, The Netherlands); Kerri Beckmann (King's College London,
   London, UK & Guy's and St Thomas' NHS Foundation Trust, London, UK),
391
   Brian Denton (University of Michigan, Michigan, USA), Andrew Hayen (Uni-
392
   versity of Technology Sydney, Australia), Paul Boutros (Ontario Institute of
393
   Cancer Research, Toronto, Ontario, Canada).
       Clinical Research Partners' IT Experts: Wei Guo (Johns Hopkins Uni-
395
   versity, The James Buchanan Brady Urological Institute, Baltimore, USA),
```

Nicole Benfante (Memorial Sloan Kettering Cancer Center, New York, USA), Janet Cowan (University of California San Francisco, San Francisco, USA), 398 Dattatraya Patil (Emory University School of Medicine, Winship Cancer In-399 stitute, Atlanta, USA), Emily Tolosa (MD Anderson Cancer Centre, Houston, Texas, USA), Tae-Kyung Kim (University of Michigan and Michigan 401 Urological Surgery Improvement Collaborative, Ann Arbor, Michigan, USA), 402 Alexandre Mamedov (University of Toronto, Sunnybrook Health Sciences 403 Centre, Toronto, Ontario, Canada), Vincent LaPointe (University of British 404 Columbia, BC Cancer Agency, Vancouver, Canada), Trafford Crump (University of Calgary, Southern Alberta Institute of Urology, Calgary, Canada), 406 Vasilis Stavrinides (University College London & University College London Hospital Trust, London, UK), Jenna Kimberly-Duffell (University of Cambridge & Cambridge University Hospitals NHS Foundation Trust, Cambridge, UK), Aida Santaolalla (King's College London, London, UK & Guy's and St Thomas' NHS Foundation Trust, London, UK), Daan Nieboer (Eras-411 mus Medical Center, Rotterdam, The Netherlands), Jonathan Olivier (Lille 412 University Hospital Center, Lille, France), Tiziana Rancati (Fondazione IR-CCS Istituto Nazionale dei Tumori di Milano, Milan, Italy), Helén Ahlgren (Sahlgrenska University Hospital, Göteborg, Sweden), Juanma Mascarós (Instituto Valenciano de Oncología, Valencia, Spain), Annica Löfgren (Skåne 416 University Hospital, Malmö, Sweden), Kurt Lehmann (Kantonsspital Baden, 417 Baden, Switzerland), Catherine Han Lin (Monash University and Epworth 418 HealthCare, Melbourne, Australia), Hiromi Hirama (Kagawa University, Kagawa, Japan), Kwang Suk Lee (Yonsei University College of Medicine, Gangnam Severance Hospital, Seoul, Korea).

Research Advisory Committee: Guido Jenster (Erasmus MC, Rotterdam, 422 the Netherlands), Anssi Auvinen (University of Tampere, Tampere, Fin-423 land), Anders Bjartell (Skåne University Hospital, Malmö, Sweden), Ma-424 soom Haider (University of Toronto, Toronto, Canada), Kees van Bochove (The Hyve B.V. Utrecht, Utrecht, the Netherlands), Ballentine Carter (Johns 426 Hopkins University, Baltimore, USA – until 2018). 427 Management team: Sam Gledhill (Movember Foundation, Melbourne, 428 Australia), Mark Buzza / Michelle Kouspou (Movember Foundation, Melbourne, Australia), Chris Bangma (Erasmus Medical Center, Rotterdam, The Netherlands), Monique Roobol (Erasmus Medical Center, Rotterdam, The Netherlands), Sophie Bruinsma / Jozien Helleman (Erasmus Medical Center, Rotterdam, The Netherlands).

434 References

- [1] Briganti A, Fossati N, Catto JW, Cornford P, Montorsi F, Mottet N, et al. Active surveillance for low-risk prostate cancer: the European Association of Urology position in 2018. European urology 2018;74(3):357–68.
- [2] Epstein JI, Egevad L, Amin MB, Delahunt B, Srigley JR, Humphrey PA.

 The 2014 international society of urological pathology (isup) consensus

 conference on gleason grading of prostatic carcinoma. The American

 journal of surgical pathology 2016;40(2):244–52.
- [3] Bruinsma SM, Roobol MJ, Carroll PR, Klotz L, Pickles T, Moore CM, et al. Expert consensus document: semantics in active surveillance for

- men with localized prostate cancer—results of a modified delphi consensus procedure. Nature reviews urology 2017;14(5):312.
- [4] Bul M, Zhu X, Valdagni R, Pickles T, Kakehi Y, Rannikko A, et al.

 Active surveillance for low-risk prostate cancer worldwide: the prias

 study. European urology 2013;63(4):597–603.
- Loeb S, Carter HB, Schwartz M, Fagerlin A, Braithwaite RS, Lepor H.
 Heterogeneity in active surveillance protocols worldwide. Reviews in urology 2014;16(4):202–3.
- Loeb S, Vellekoop A, Ahmed HU, Catto J, Emberton M, Nam R, et al.
 Systematic review of complications of prostate biopsy. European urology
 2013;64(6):876–92.
- ⁴⁵⁶ [7] Bokhorst LP, Alberts AR, Rannikko A, Valdagni R, Pickles T, Kakehi Y, et al. Compliance rates with the Prostate Cancer Research International Active Surveillance (PRIAS) protocol and disease reclassification in noncompliers. European Urology 2015;68(5):814–21.
- et al. Comparative analysis of biopsy upgrading in four prostate cancer active surveillance cohorts. Annals of internal medicine 2018;168(1):1–9.
- [9] Bratt O, Carlsson S, Holmberg E, Holmberg L, Johansson E, Josefsson A, et al. The study of active monitoring in sweden (sams): a randomized study comparing two different follow-up schedules for active surveillance of low-risk prostate cancer. Scandinavian journal of urology 2013;47(5):347–55.

- [10] Nieboer D, Tomer A, Rizopoulos D, Roobol MJ, Steyerberg EW. Active
 surveillance: a review of risk-based, dynamic monitoring. Translational
 andrology and urology 2018;7(1):106–15.
- [11] Chesnut GT, Vertosick EA, Benfante N, Sjoberg DD, Fainberg J, Lee T, et al. Role of changes in magnetic resonance imaging or clinical stage in evaluation of disease progression for men with prostate cancer on active surveillance. European Urology 2019;.
- ⁴⁷⁵ [12] de Carvalho TM, Heijnsdijk EA, de Koning HJ. Estimating the risks ⁴⁷⁶ and benefits of active surveillance protocols for prostate cancer: a mi-⁴⁷⁷ crosimulation study. BJU international 2017;119(4):560–6.
- 13] Kasivisvanathan V, Giganti F, Emberton M, Moore CM. Magnetic resonance imaging should be used in the active surveillance of patients with localised prostate cancer. European urology 2020;77(3):318.
- ⁴⁸¹ [14] Coley RY, Zeger SL, Mamawala M, Pienta KJ, Carter HB. Prediction of the pathologic gleason score to inform a personalized management program for prostate cancer. European urology 2017;72(1):135–41.
- 484 [15] Ankerst DP, Xia J, Thompson Jr IM, Hoefler J, Newcomb LF, Brooks
 485 JD, et al. Precision medicine in active surveillance for prostate can486 cer: development of the canary—early detection research network active
 487 surveillance biopsy risk calculator. European urology 2015;68(6):1083–8.
- ⁴⁸⁸ [16] Partin AW, Yoo J, Carter HB, Pearson JD, Chan DW, Epstein JI, et al.

 The use of prostate specific antigen, clinical stage and gleason score to

- predict pathological stage in men with localized prostate cancer. The

 Journal of urology 1993;150(1):110-4.
- [17] Makarov DV, Trock BJ, Humphreys EB, Mangold LA, Walsh PC, Epstein JI, et al. Updated nomogram to predict pathologic stage of prostate cancer given prostate-specific antigen level, clinical stage, and biopsy gleason score (partin tables) based on cases from 2000 to 2005. Urology 2007;69(6):1095–101.
- [18] Bruinsma SM, Zhang L, Roobol MJ, Bangma CH, Steyerberg EW,
 Nieboer D, et al. The movember foundation's gap3 cohort: a profile of
 the largest global prostate cancer active surveillance database to date.
 BJU international 2018;121(5):737–44.
- 501 [19] Schoots IG, Petrides N, Giganti F, Bokhorst LP, Rannikko A, Klotz

 L, et al. Magnetic resonance imaging in active surveillance of prostate

 cancer: a systematic review. European urology 2015;67(4):627–36.
- [20] Tomer A, Nieboer D, Roobol MJ, Steyerberg EW, Rizopoulos D. Personalized schedules for surveillance of low-risk prostate cancer patients.

 Biometrics 2019;75(1):153–62.
- [21] Rizopoulos D. Joint Models for Longitudinal and Time-to-Event Data:
 With Applications in R. CRC Press; 2012. ISBN 9781439872864.
- [22] Laird NM, Ware JH, et al. Random-effects models for longitudinal data.
 Biometrics 1982;38(4):963-74.
- [23] Nixon RG, Wener MH, Smith KM, Parson RE, Strobel SA, Brawer
 MK. Biological variation of prostate specific antigen levels in serum:

- an evaluation of day-to-day physiological fluctuations in a well-defined cohort of 24 patients. The Journal of urology 1997;157(6):2183–90.
- ⁵¹⁵ [24] Cooperberg MR, Brooks JD, Faino AV, Newcomb LF, Kearns JT, Car-⁵¹⁶ roll PR, et al. Refined analysis of prostate-specific antigen kinetics to ⁵¹⁷ predict prostate cancer active surveillance outcomes. European urology ⁵¹⁸ 2018;74(2):211–7.
- 519 [25] Rizopoulos D. The R package JMbayes for fitting joint models for lon-520 gitudinal and time-to-event data using MCMC. Journal of Statistical 521 Software 2016;72(7):1–46.
- ⁵²² [26] Royston P, Altman DG. External validation of a cox prognostic model: principles and methods. BMC medical research methodology 2013;13(1):33.
- 525 [27] Steyerberg EW, Vickers AJ, Cook NR, Gerds T, Gonen M, Obuchowski
 N, et al. Assessing the performance of prediction models: a framework
 for some traditional and novel measures. Epidemiology (Cambridge,
 Mass) 2010;21(1):128.
- 529 [28] Rizopoulos D, Molenberghs G, Lesaffre EM. Dynamic predictions with 530 time-dependent covariates in survival analysis using joint modeling and 531 landmarking. Biometrical Journal 2017;59(6):1261–76.
- ⁵³² [29] Turnbull BW. The empirical distribution function with arbitrarily grouped, censored and truncated data. Journal of the Royal Statistical Society Series B (Methodological) 1976;38(3):290–5.

- [30] Tomer A, Rizopoulos D, Nieboer D, Drost FJ, Roobol MJ, Steyerberg
 EW. Personalized decision making for biopsies in prostate cancer active
 surveillance programs. Medical Decision Making 2019;39(5):499–508.
- ⁵³⁸ [31] Balasubramanian R, Lagakos SW. Estimation of a failure time distribu-⁵³⁹ tion based on imperfect diagnostic tests. Biometrika 2003;90(1):171–82.