PROJECT REPORT

Investigating the molecular cause of hypermobile Ehlers-Danlos syndrome

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1 Introduction

1.1 Background

- Ehlers-Danlos syndromes are a group of heritable connective tissue disorders that can be classified into multiple subtypes
- current classification describes 13 subtypes [1]
- · most common form: hypermobile EDS (hEDS) only subtype with unknown molecular cause
- · diagnosis based on clinical presentation
- identify molecular cause for: diagnosis, understanding, potential treatment [2]
- current state: ongoing study with aim to do this by analysing genes of 1000 affected individuals, results not expected before 2025 [3]
- many studies investigated genes but no clear molecular cause with connection to connective tissue established yet [4]
- · until then use data known
- problem with existing data: criteria change in 2017, earlier diagnosed people might not classify anymore [5] + there is a unclear overlap with Hypermobility Spectrum Disorder (HSD) and terms often used interchangeable and diagnoses often grouped together (unclear whether there is a difference) [5, 6]
- we know the clinical representation and how other eds types work (kind of)
- TODO: include information of what is affected, extracellular matrix, collagen, connective tissue

1.2 Aim

- aim: investigate moleculare cause of hypermobile EDS
- · studying differentially expressed genes in hEDS patients
- · which biological processes and pathways are affected
- similarities to pathways/processes/function affected by other EDS genes
- TODO: what do we want to do in the big picture: find candidate genes? is this to ambitious?

2 Methods

2.1 Analysis of Differentially Expressed Genes and Network Creation

- data accessible at NCBI GEO database with the accession number GSE218012 [2]
- analysis with DeSeq2 in R based on the analysis exported from GEO2R [7] to identily up-regulated and down-regulated genes
- |log2FoldChange| > 0.5, pValue < 0.05, pValue adjusted with Benjamini and Hochberg False Discovery Rate
- use Cytoscape to create network [8]
- query differentially expressed genes from string db [9] (confidence cut off 0.4)
- query eds genes related to other eds types additionally: 21 genes retrieved from Disease Ontology with Disease Ontology ID 13359 [10], queried from string, again with a medium confidence cut off of 0.4

 load data about differential expression into network, for EDS genes and differentially expressed genes

2.2 Enrichment analysis and clustering

- · exploratory enrichment analysis on whole network
- · use R-package clusterProfiler [11] for enrichment
- · large network, thus cluster before to get better insights for specific parts
- · two different cluster methods used with different resulting cluster structure
- create subnetworks for clusters with more than 15 nodes [TODO: why this threshold? to focus
 on more relevant because many in MCODE between 10 and 15 nodes, for community clustering
 smaller ones probably found by MCODE]

2.2.1 MCODE

- MCODE finds "densely connected regions in large protein-protein interaction networks that may represent molecular complexes" [12]
- · results in smaller clusters
- · suited to analyse molecular function
- use default parameters
- analysis of molecular function with clusterProfiler with GO over-representation analysis
- · do we see clusters with other molecular functions than expected
- · are genes clustered together with other eds genes
- · are clusters generelly mostly upregulated or downregulated

2.2.2 Community Clustering

- · small cluster not helpful for biological processes and pathways
- · therefore use second clustering method
- community clustering with GLay, "more suitable for functional interpretation" [13]

3 Results

3.1 Differentially Expressed Genes and Network creation

- 908 differentially expressed genes with chosen thresholds section 2.1
- around half are upregulated, around half downregulated [TODO: get exact numbers]
- String was able to query 828 of them [TODO: provide table of not queried in supplementary material
 or at least github?]
- after querying additional genes that are known to be related to other eds types: resulting network with 847 nodes and 6129 edges
- position of known eds genes in network is mostly central (checked degree, clustering coefficient, betweenness centrality and closeness centrality)

3.2 Enrichment analysis and clustering

3.2.1 MCODE

- 3 clusters with more than 15 nodes (first with 66 nodes & 1953 connections, second with 44 nodes & 686 connections, third with 16 nodes & 114 connections)
- first and largest cluster upregulated only
- · second one as well
- 3rd MCODE cluster, shown in figure 1, is mostly upgregulated with 2 downregulated nodes, some not relevantly differentially expressed, also only 9/16 are not known eds genes, 8 are
- the eds genes are all below the threshold of |log2FoldChange|>0.5, 1 of them is 1 of the two downregulated genes
- quite interesting to see genes closely related to other eds genes, COL21A1 is also strongly upregulated (log2FoldChange > 2)
- all known EDS genes with ADAMTS2 as exception have a high Closeness Centrality
- · regarding the enrichment of this cluster: not surprising to see extracellular

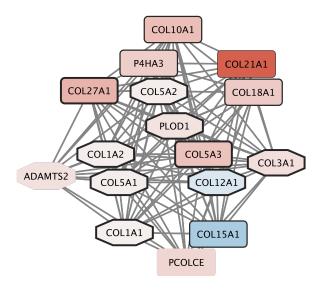


Figure 1: MCODE cluster 3 [TODO: add legend for shape and colour]

3.2.2 Community Clustering

- 6 clusters with more than 15 nodes
- 3 very small (18, 29, 29 nodes), two medium sized (76 and 105 nodes) and one very large cluster (363 nodes)
- · especially medium sized clusters highly interconnected
- biggest one mix of up-regulated and down-regulated genes, contains all 21 genes known to cause other EDS types
- both medium sized clusters mostly upregulated \rightarrow interesting!

4 Discussion

References

- [1] Malfait F, Francomano C, Byers P, Belmont J, Berglund B, Black J, et al. The 2017 international classification of the Ehlers–Danlos syndromes. American Journal of Medical Genetics Part C: Seminars in Medical Genetics. 2017 3;175:8-26.
- [2] Ritelli M, Colombi M. Molecular Genetics and Pathogenesis of Ehlers-Danlos Syndrome and Related Connective Tissue Disorders. Genes. 2020 5;11.
- [3] HEDGE (Hypermobile Ehlers-Danlos Genetic Evaluation) Study The Ehlers Danlos Society;. Available from: https://www.ehlers-danlos.com/hedge/.
- [4] Caliogna L, Guerrieri V, Annunziata S, Bina V, Brancato AM, Castelli A, et al. Biomarkers for Ehlers-Danlos Syndromes: There Is a Role? International Journal of Molecular Sciences 2021, Vol 22, Page 10149. 2021 9;22:10149.
- [5] Gensemer C, Burks R, Kautz S, Judge DP, Lavallee M, Norris RA. Hypermobile Ehlers-Danlos syndromes: Complex phenotypes, challenging diagnoses, and poorly understood causes. Developmental dynamics: an official publication of the American Association of Anatomists. 2021 3;250:318.
- [6] Ritelli M, Chiarelli N, Cinquina V, Zoppi N, Bertini V, Venturini M, et al. RNA-Seq of Dermal Fibroblasts from Patients with Hypermobile Ehlers-Danlos Syndrome and Hypermobility Spectrum Disorders Supports Their Categorization as a Single Entity with Involvement of Extracellular Matrix Degrading and Proinflammatory Pathomechanisms. Cells. 2022 12;11.
- [7] Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. Genome Biology. 2014 12;15:1-21.
- [8] Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome research. 2003 11;13:2498-504.
- [9] Szklarczyk D, Kirsch R, Koutrouli M, Nastou K, Mehryary F, Hachilif R, et al. The STRING database in 2023: protein-protein association networks and functional enrichment analyses for any sequenced genome of interest. Nucleic acids research. 2023 1;51:D638-46.
- [10] Schriml LM, Mitraka E, Munro J, Tauber B, Schor M, Nickle L, et al. Human Disease Ontology 2018 update: Classification, content and workflow expansion. Nucleic Acids Research. 2019 1;47:D955-62.
- [11] Wu T, Hu E, Xu S, Chen M, Guo P, Dai Z, et al. clusterProfiler 4.0: A universal enrichment tool for interpreting omics data. Innovation. 2021 8;2.
- [12] Bader GD, Hogue CWV. An automated method for finding molecular complexes in large protein interaction networks. BMC Bioinformatics. 2003 1;4:1-27.

[13] Su G, Kuchinsky A, Morris JH, States DJ, Meng F. GLay: community structure analysis of biological networks. Bioinformatics. 2010 12;26:3135-7.