

Potentially Prebiotic Synthesis of Aminoacyl-RNA via a Bridging Phosphoramidate-Ester Intermediate

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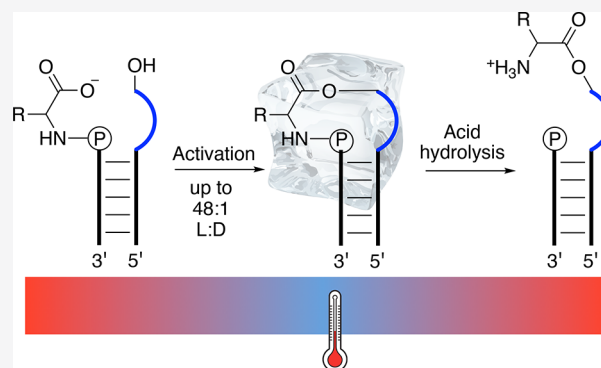


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ABSTRACT: Translation according to the genetic code is made possible by selectivity both in aminoacylation of tRNA and in anticodon/codon recognition. In extant biology, tRNAs are selectively aminoacylated by enzymes using high-energy intermediates, but how this might have been achieved prior to the advent of protein synthesis has been a largely unanswered question in prebiotic chemistry. We have now elucidated a novel, prebiotically plausible stereoselective aminoacyl-RNA synthesis, which starts from RNA-amino acid phosphoramidates and proceeds via phosphoramidate-ester intermediates that subsequently undergo conversion to aminoacyl-esters by mild acid hydrolysis. The chemistry avoids the intermediacy of high-energy mixed carboxy-phosphate anhydrides and is greatly favored under eutectic conditions, which also potentially allow for the requisite pH fluctuation through the variable solubility of CO₂ in solid/liquid water.



INTRODUCTION

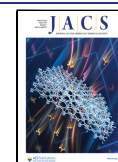
Ribosomal peptide synthesis relies upon aminoacyl-tRNAs as intermediates in the translation of messenger-RNAs into the corresponding genetically coded protein products. Highly specific enzymatic aminoacylation of the 2',3'-diol of tRNAs using aminoacyl-adenylates is crucial to this process. In the absence of enzymes, which bind them tightly and protectively, carbon dioxide promotes the reversible conversion of aminoacyl-adenylates to amino acid *N*-carboxyanhydrides (NCAs), and the latter are subsequently polymerized to random peptides in aqueous solution.^{1,2} Other aminoacyl-RNA mixed anhydrides **1** have been used as substrates for non-enzymatic aminoacyl-RNA synthesis in prebiotic model studies,^{3,4} but they are similarly susceptible to carbon dioxide-promoted destruction. Conceptually, there are two fundamentally different chemical pathways for the formation of mixed anhydrides—via either carboxylate activation or phosphate activation (the latter of which is used by modern day biology). In the former, the phosphate reacts (reversibly) with an activated carboxylate (e.g., NCA, 5(4*H*)-oxazolone, or other species) giving a mixed anhydride **1**.^{5–7} Conversely, in the phosphate activation pathway, a native amino acid reacts with an activated phosphate (e.g., imidoyl phosphate) giving a mixed anhydride **1**.⁷ Carboxylate activation processes^{5,6} are inherently problematic under a carbon dioxide-containing atmosphere in that the formation of NCAs and hence random non-coded peptides cannot be avoided.

Prebiotic activation chemistry is also needed to enable the formation of phosphodiester from alcohols and the corresponding phosphomonoesters. Thus, for example, 5'-phos-

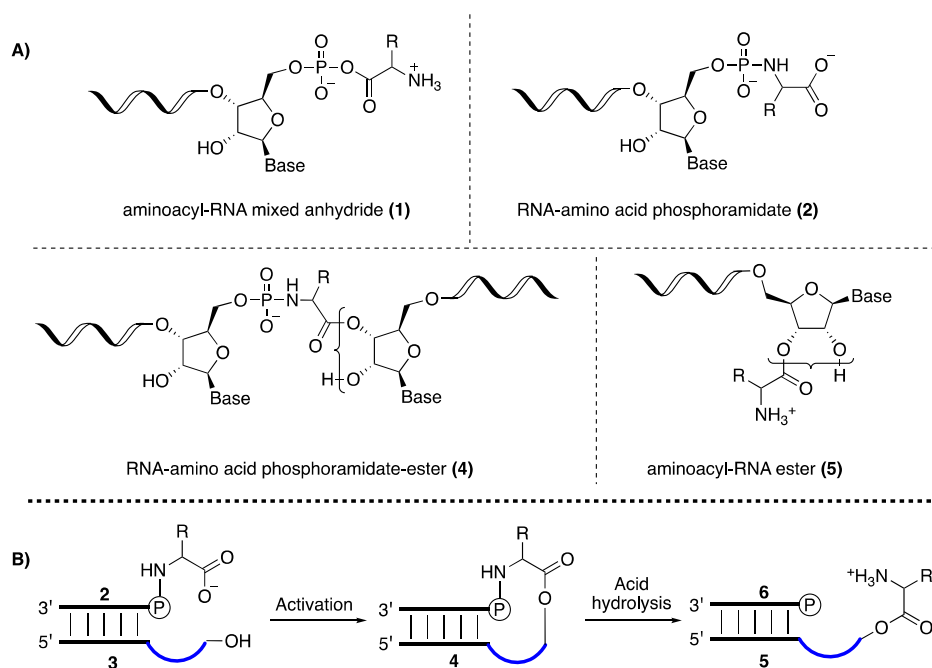
phorimidazolides are used for prebiotic RNA ligation or monomer extension chemistry.^{8,9} Seeking to unify the various building block activation chemistries, we were intrigued by the work of Weber and Orgel et al. showing that amino acids can react with 5'-phosphorimidazolides through their amino groups generating amino acid phosphoramidates **2** (Scheme 1).¹⁰ By using adenine nucleotide 2'/3'-aminoacyl-esters and 5'-phosphorimidazolides, bridged phosphoramidate-esters could be formed by templating with poly(U).¹¹ The Richert group has built upon these earlier results and showed that RNA-peptide phosphoramidates can couple to 3'-amino-2',3'-dideoxy-terminated oligonucleotides to give bridging peptide phosphoramidate-amides on an RNA template.¹² However, the reaction of RNA-amino acid phosphoramidates **2** (Scheme 1) with the 2',3'-diol of a canonical oligonucleotide **3** to generate phosphoramidate-esters **4** is inherently more interesting as the products are potentially hydrolyzable to aminoacyl-esters **5** and 5'-phosphoryl-RNAs **6** under mildly acidic conditions.¹³ Synthesis of aminoacyl-esters in this way would bypass the unstable mixed anhydrides **1** employed in other strategies¹⁴ and might have been more easily achievable before the advent of enzymes.

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Scheme 1. (A) Chemical Structures of RNA-Amino Acid Hybrid Compounds; (B) Reaction Scheme for the Formation of Aminoacyl-RNA Esters^a



^aRNA-amino acid phosphoramidate-ester (middle structure, 4) is formed from RNA phosphoramidate (2, left) and ester acceptor RNA (3) under activation conditions. The phosphoramidate bond can then be cleaved under mild acid conditions generating aminoacyl-RNA (5, right).

We have previously postulated that loops could provide chemoselectivity in aminoacylation and avoid the necessity of extra strands of RNA.¹⁴ tRNAs have 5'-phosphate groups and a short 3'-overhang at the end of the acceptor stem and accordingly we wondered if phosphoramidate-ester 4 formation would be possible in such a stem-overhang configuration. We chose UUCA as an overhang sequence as it shows strong sequence similarity to the CCA overhang of modern tRNAs and was a successful sequence identified in our previous work.¹⁴

RESULTS

A chemically synthesized RNA-amino acid phosphoramidate (2-Gly, 3'-AGCGAp-Gly-OH, 100 μ M) was mixed with ester acceptor RNA strand 3 (3'-ACCUUUCGCU, 100 μ M) and activation buffer (made up of water-soluble carbodiimide EDC and imidazole in buffered aqueous solution). While EDC is prebiotically implausible, we use it here as others have done previously as a model-activating agent, allowing the formation of an acyl imidazolide.¹² The reaction was incubated at room temperature overnight and afterward analyzed by HPLC (Figure S1). In comparison to controls containing 5'-phosphoryl-RNA 6 in place of 2-Gly (Figure S17), an extra peak could be observed in the HPLC trace of the reaction products consistent with the formation of phosphoramidate-ester 4-Gly. After purification by HPLC, this product was shown by MALDI-TOF mass spectroscopy to indeed have the mass of 4-Gly (3'AGCGAp-Gly-ACCUUUCGCU, calculated MW 4767.7; found, 4768.0 [M + H]). This peak also disappeared on treatment with alkali, consistent with the presence of an ester linkage in the product.¹⁵

Encouraged by these findings, we further synthesized two different RNA-amino acid phosphoramidates (2-L-Ala, 3'AGCGAp-L-Ala-OH and 2-D-Ala, 3'AGCGAp-D-Ala-OH)

to explore their comparable reactivity and the stereoselectivity of ester formation. Based on the ³¹P NMR spectra (Figure S19), the yields of formation of 2-L-Ala and 2-D-Ala from their parent 5'-phosphoryl-RNAs were similar, suggesting that there is no stereoselectivity in the conventional synthesis of phosphoramidates of single-stranded RNA. After HPLC purification, each of the RNA-amino acid phosphoramidates was combined with 10-mer RNA ester acceptor 3 (3'-ACCUUUCGCU, 100 μ M) in activation buffer and again new HPLC peaks were evident after the reaction. As shown in Figures S2 and S3, the L- and D-alanine phosphoramidate-esters 4-L-Ala and 4-D-Ala were formed after 18 h, their identity was confirmed by alkaline hydrolysis and the former additionally by the MALDI-TOF mass spectra (4-L-Ala: 3'AGCGAp-L-Ala-ACCUUUCGCU, calculated MW 4781.7; found, 4782.2 [M + H]). Interestingly, the yield of 4-D-Ala was significantly lower than that of 4-L-Ala (9.5:1, quantified as an average of the ratios across three replicates using an internal standard). It is known that aminoacyl-imidazolides are key to the aminoacylation of one of the hydroxyl groups of a diol in water,¹⁶ presumed due to the protonation of the imidazole leaving group mediated through the second hydroxyl group,² and we suspect that acyl-imidazolides are similarly crucial to the chemistry we have discovered as we observed no phosphoramidate-ester 4-L-Ala product when using activation buffer in the absence of imidazole (Figure S14). Other non-base-sensitive peaks also appeared independent of the presence of imidazole. It has been reported that EDC (albeit at much higher concentrations) reacts with RNA to form cyclic structures or modify nucleobases, and this could account for some of the other base insensitive peaks we observe (Figures S1–S18).^{17,18}

With these results in hand, we next wanted to scope out the chemoselectivity and stereoselectivity for different amino acids.

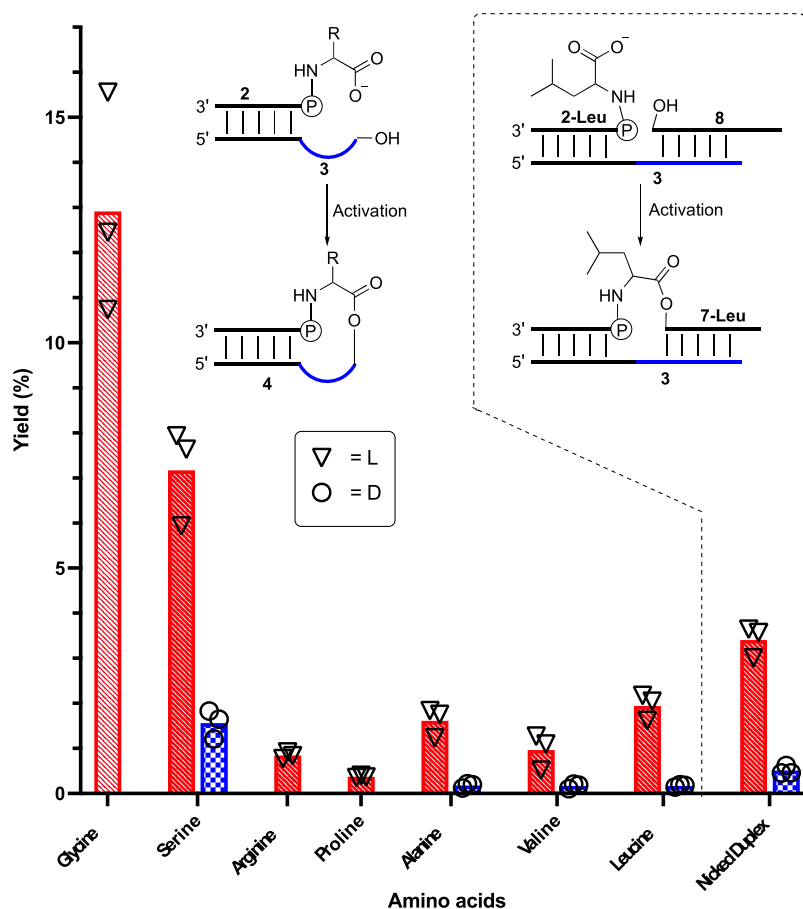


Figure 1. Room-temperature yields for formation of phosphoramidate-esters 4 via a nicked loop and 7-Leu via a nicked duplex. Bars are mean values based on three replicates. Each replicate is represented by a circular/triangular data point. Red hashed bars represent L-amino acid stereochemistry and blue checkered bars D- amino acid (where appropriate).

Using the same oligonucleotide sequence as before, we isolated phosphoramidates 2 of leucine, valine, arginine, proline, and serine covering a range of prebiotically plausible early amino acids.¹⁹ On individually submitting all 5 RNA-L-amino acid phosphoramidates to ester-forming conditions in activation buffer with 10-mer RNA ester acceptor 3 (3'-ACCUUUCG-CU, 100 μ M), peaks for all phosphoramidate-ester products could be identified (correct MALDI-TOF masses and susceptibility to alkaline hydrolysis) (Figures S4–S11, Table S1). However, the yields varied according to the amino acid side chain (Figure 1). Comparable reactions using D-amino acid phosphoramidates all afforded significantly lower yields of phosphoramidate-esters compared to the corresponding L-amino acid phosphoramidates (Table S3). Thus, it appears that with the esterification chemistry described herein, there is a general relative stereochemical correlation between L-amino acids and RNA based on D-ribose.

In some cases, HPLC peaks corresponding to the phosphoramidate-ester products 4 were observed with “shoulders” or as double peaks (e.g., Figure S4). In all cases, we found that both peaks decreased in intensity upon alkaline hydrolysis although not always at the same rate. These observations suggest the formation of mixtures of 2'- and 3'-linked phosphoramidate-esters 4. Similar aminoacyl-migration is known with aminoacyl-tRNAs²⁰ and so we have not concerned ourselves further with this aspect of regioselectivity.

High stereoselectivity and at least moderate chemoselectivity for specific amino acids would be features essential for the

development of genetic encoding of biologically useful peptides before the emergence of enzymatic aminoacylation. We have previously established that variations in RNA sequence are associated with different yields in another aminoacyl-transfer system.¹⁴

It is possible that in sequences that contain a palindromic (or partially palindromic) sequence, the esterification could occur through a nicked-duplex type configuration rather than by way of a stem with a folded back overhang. Alternatively, complementary sequences of RNA may bind to the overhang sequence, producing a nicked-duplex. To study this, we decided to impose a nicked-duplex configuration in our system by including an additional oligonucleotide with a sequence complementary to the loop region (Figure 1). The reaction of amide 2-L-Leu, template 3, and complement 3'-AAG-GUAAU 8 in activation buffer produced a new alkali sensitive peak with a long HPLC retention time, while no trace of the previously observed phosphoramidate-ester product 4-L-Leu could be detected. This new peak was isolated and shown to have a mass consistent with the 13-mer phosphoramidate-ester 7-L-Leu (3'-AGCGAp-Leu-AAGGUAAU, calculated MW 4323.7; found, 4324.2 [M + H]) and to be alkali sensitive. On repeating the experiment using the D-leucine phosphoramidate 2-D-Leu, we saw a greatly reduced intensity peak posited, on the basis of alkaline sensitivity, to be due to the D-Leu-13-mer 7-D-Leu (L/D 7:1 compared to L/D 12:1 for leucine nicked-loop esterification, Figures S12 and S13). This

result suggests that any sequences that allow esterification via a nicked-duplex configuration will have reduced L/D selectivity.

Serendipitously, on resubmitting a sample to HPLC analysis that had been stored at $-16\text{ }^{\circ}\text{C}$ for a week after the initial reaction at room temperature, we observed greatly increased intensity of the phosphoramidate-ester product peak. However, samples kept at $-70\text{ }^{\circ}\text{C}$ showed no evidence of a similarly enhanced reaction. This suggested that the esterification reaction could take place under eutectic conditions with increased yields. After submitting fresh samples to $-16\text{ }^{\circ}\text{C}$ in activation buffer for 7 days, the L-amino acid phosphoramidate-ester products formed in much higher yields (Figure 2), while formation of D-amino acid phosphoramidate-esters did not improve significantly over the room-temperature yields, resulting in improved L/D selectivity over the room-temperature conditions (Table S3). Yields increased further by 14 days, but the L/D selectivity then decreased, likely due to lower

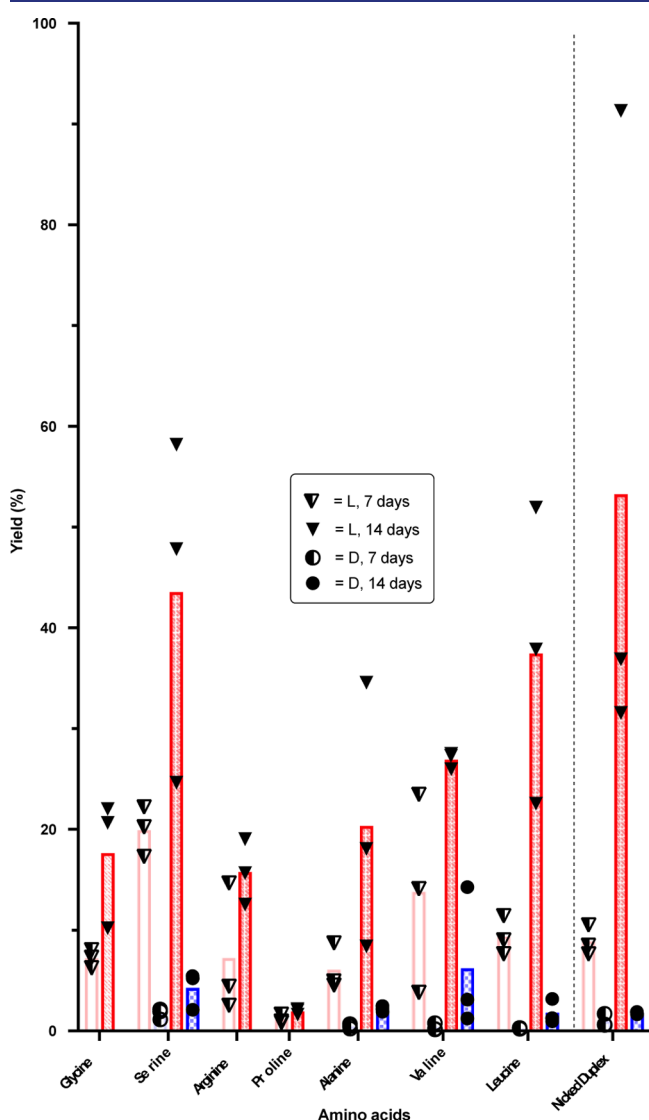


Figure 2. Eutectic yields for formation of phosphoramidate-esters 4 via a nicked loop and 7-Leu via a nicked duplex. Each replicate is represented by a circular/triangular data point. Red hashed bars represent L-amino acid stereochemistry and blue checkered bars D-amino acid (where appropriate). Darker colors indicate longer reaction times.

levels of starting materials available in the “L-reactions” over the second week. As the individual data points of Figure 2 show, there is a large variation between replicates of the eutectic samples despite the high reproducibility of the results from the room-temperature reaction (Figure 1) from the same starting mixture. We suggest that, among other things, this could be due to relatively poor temperature control in a large $-16\text{ }^{\circ}\text{C}$ freezer.

We next studied the formation of aminoacyl-RNA esters 5 by mild acid hydrolysis of phosphoramidate-esters 4 (Figure 3). The HPLC-purified phosphoramidate-esters 4 were

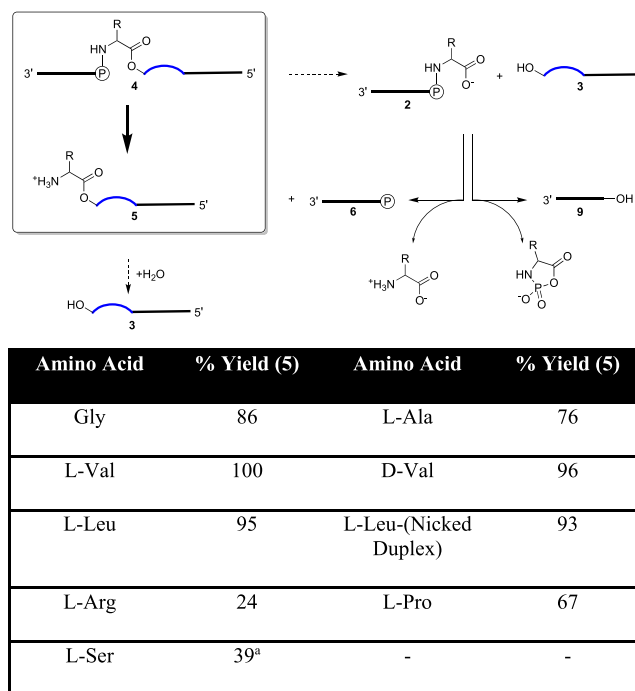


Figure 3. Top—Mechanism of hydrolysis of the phosphoramidate-ester 4 at pH = 3. The majority of 4 undergo phosphoramidate bond hydrolysis giving 5'-phosphoryl-5-mer 6 and the desired aminoacyl-10-mer 5 product, which can then degrade slowly back to template 3. Minority hydrolyses at the ester bond of 4 gives 3 and 5' phosphoramidate-5-mer 2. Phosphoramidate 2 degrades either by direct hydrolysis to 5'-phosphoryl-5-mer 6, or by intramolecular oligonucleotide dephosphorylation to the 5'-hydroxyl-5-mer 9 in an amino acid-specific fashion. Bottom—Yields of formation of aminoacyl-10-mer 5 from phosphoramidate-ester 4 with different amino acids. ^aEstimate due to unknown products of hydrolysis. A discussion of the differing yields of Arg, Pro, and Ser is presented in the ESI.

dissolved in pH = 3 formate buffer (final concentration, 83 mM formate) with an internal HPLC standard and kept at $25\text{ }^{\circ}\text{C}$ overnight. HPLC analysis revealed that all the phosphoramidate-ester 4 was hydrolyzed, giving 5'-phosphoryl-5-mer 6 RNA (3'-AGCGAp), aminoacyl-10-mer 5 (H-amino acid-3'-ACCUUUCGCU), and 10-mer RNA 3 (3'-ACCUUUCGCU), which resulted from subsequent slow ester hydrolysis. The free aminoacyl-ester 5 was produced quantitatively or near quantitatively for glycine, alanine, valine, and leucine (the latter either from the product of the nicked-loop or nicked-duplex chemistry), with little difference observed between L- and D-valine phosphoramidate-esters 4-L-Val and 4-D-Val (the only amino acid for which we could isolate sufficient D-phosphoramidate-ester to study). The identity of 10-mer RNA 3 was confirmed by spiking with an authentic sample,

and the identity of aminoacyl-RNA **5** was confirmed by its susceptibility to alkaline hydrolysis. After such hydrolysis, the aminoacyl-RNA **5** peak disappeared, and the peak corresponding to the 10-mer RNA **3** increased adding further weight to the characterization (Figures S21–S34). To varying degrees with different amino acids, we also identified 5'-hydroxyl-5-mer **9** (e.g., Figure S22). We hypothesize that this likely resulted from hydrolysis of the ester bond of the phosphoramidate-ester **4** (to produce 10-mer RNA **3** and phosphoramidate-5-mer **2**), followed by internal hydrolysis either in a similar manner to that previously hypothesized by the McGuigan group in relation to "Protide" activation,²¹ or by in-line hydrolysis to afford **9**. On submitting all amino acid-phosphoramidate starting materials **2** individually to pH = 3 formate buffer (60 mM final concentration formate), we identified only 2 products—5'-phosphoryl-5-mer **6** and 5'-hydroxyl-5-mer **9**—in varying yields (Table S2), with a trend for increased formation of **9** following the Thorpe–Ingold effect for alanine, valine, and leucine. What is more, we also saw a consistent increase in yield of **6** with D- relative to L-amino acids. In all cases, acidification of phosphoramidate-esters **4** predominantly resulted in hydrolysis of the phosphoramidate over the ester. With the exception of serine (Figure S30), no unassignable peaks were observed for any of acid hydrolyses, implying the conditions maintain the integrity of the RNA.

DISCUSSION

This work has demonstrated that efficient production of aminoacyl-RNAs **5** can be achieved via stepwise esterification of RNA-amino acid phosphoramidates **2** to proximal oligonucleotide-2',3'-diols **3**, followed by mild acid hydrolysis of the phosphoramidate linkage. The lack of acidic degradation could be attributed to the short length of treatment (17 h), a factor which would have to be incorporated into the prebiotic scenario. The proximity required to enable esterification can be realized in nicked-loop or nicked-duplex configurations. The esterification reaction is stereoselective—being preferred for L- over D-amino acid residues with RNA based on D-ribose—and the stereoselectivity is highest in the nicked-loop configuration. The reaction with one set of oligonucleotide sequences exhibits some chemoselectivity for different amino acid side chains auguring well for higher chemoselectivity when sequence variability is combinatorially explored in future studies. If this chemistry was realized with glutamic or aspartic acid, it is likely that the prebiotic activating agent would also activate their side chains to nucleophilic attack. Whether these amino acids are relevant at the origins of translation would have to take this into account.

Yields of the intermediate phosphoramidate-ester **4** were greatly improved when the reaction was performed under eutectic conditions as was stereoselectivity, but we do not know why. The rates of ester formation and hydrolysis and competing direct EDC hydration are all likely to be slowed at –16 °C but probably differentially. The lower temperatures are also likely to promote a more rigid RNA secondary structure, and this could enhance stereoselectivity. The effects of salt concentrations and pH are well documented in the eutectic phase,^{22,23} but there are also likely to be other effects at play that we have not considered, and while eutectic effects on RNA chemistry have been studied at length,^{24,25} understanding the effect on these conditions in more detail is clearly required.

The second reaction in the sequence occurs under different conditions to the first, and it is tempting to speculate about environmental effects that could cause this change. Carbon dioxide is substantially less soluble in ice than it is in water and this, along with the previously documented partitioning of HCl from brine into ice,²⁶ could contribute to the pH under eutectic conditions being elevated relative to the fully thawed state. The dilution and acidification accompanying melting of icy brines under a carbon dioxide rich atmosphere could both favor the phosphoramidate bond hydrolysis required to generate aminoacyl-esters and enable RNA duplex dissociation.²⁷ Upon refreezing, the pH of the sample would increase as would the concentration of solutes and RNA leading to the readoption of duplex and other structures. If the aminoacyl-esters of two different stem-overhangs were thereby brought into proximity, transpeptidation might ensue. Studies to investigate this possibility with a view to understand the origin of (coded) peptide synthesis are now underway.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/jacs.2c00772>.

Experimental methods including RNA synthesis methods and additional data including HPLC chromatograms, ³¹P NMR spectra, MALDI and LCMS identification, hydrolyses yields of phosphoramidate **2**, HPLC calibration curves, and ratio of yields of formation of L:D phosphoramidate esters (PDF)

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Author Contributions

[†]S.J.R. and Z.L. contributed equally.

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Notes

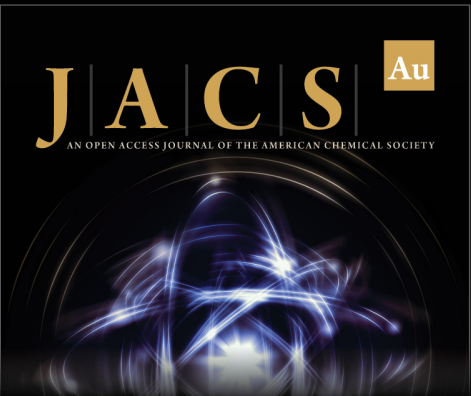
The authors declare no competing financial interest.

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