BNFO 591: High Performance Computing Assignment IV



Programs in Fortran and Python:

- 1. Adjacency matrix of Titin protein
 - 2. Finding an exact match
 - Partial exact match
 Appendix

Authors:
Hasan Alkhairo
Skyler Kuhn
Alexandrea Stylianou

PARTS 1, 2 and 3

```
program readFile
implicit none
character(len=1),dimension(0:34349):: TitinSeglist
character(len=34350) :: Titin string
character(len=60) :: a
character(len=60) :: aa
character(len=60),dimension(0:10000000):: filelist
character(len=60),dimension(1:573):: sequencelist
character (len=20) :: aaUni
character (len=1) :: tempchar
character(len=2) :: tempkmer
character(len=2), DIMENSION(0:399) :: uni kmer
character(len=300):: b! index length of the sequence array
character(len=1),dimension(0:273) :: probe_seq
character(len=274) :: string seg, match window, match window2
character(len=300),dimension(0:1) :: WPh_list! inialtizing an array to put the sequences aa i
character(len=300),dimension(1:1) :: WPh_seq_list
integer :: n=0, i, j, k, counter1=0, w,x, counter2,y,
t=0,m,match count,z,exactAlignCount,partialAlignCount,u,threshcounter, user
integer :: t1,t2,count_rate
real :: wall_clock_time
integer, DIMENSION(0:19) :: AA count
integer, DIMENSION(0:399) :: kmer count
logical inFile,nextFile
inFile =.TRUE.
!logical nextFile
nextFile = .TRUE.
```

! we are using this for the header of the matrix aaUni = "ACDEFGHIKLMNPQRSTVWY"

!This string will also be used to count unique frequencies as well as 2mer frequencies

```
open(unit=99,file="Titin.txt", status="old") !first file
open(unit=44,file="WPh Probe.txt",status="old") !second file
!!!!!!!!!!!!!! user input
print*,"Please enter a threshold percentage in decimal form"
read(*,*)user
!!!!!!!!!!!!! read in the first file
do while(inFile)
      read(unit=99,FMT=*) a
      if (a=="B") then !checking for non sequence lines. End of file
            inFile= .false.
      endif
      !print *, a
      filelist(n) = a
      n = n + 1 #keep track of index for filelist
end do
do 666 i=1,573,1
                  ! need this loop to remove the header
      !print *, filelist(i)
      sequencelist(i) = filelist(i) !adding just the sequences to this new list
666 continue
do while(nextFile) !same procedure as above, but with the other file
      read(unit=44,FMT=*) b
      if (b=="B") then !used same logic as before
            nextFile=.false.
      endif
      !print*,b
      WPh list(t) = b
      t = t + 1
end do
do 777 j=1,1,1 #simple loop to remove header. May not have been necessary
      WPh_seq_list(j) = WPh_list(j) !same logic to remove the header
      do 444 k=1,274,1 #In this loop we convert the list of sub sequences into one string
```

```
probe_seq(counter1) = WPh_seq_list(j)(k:k) !move probe sequence into another
array
             string_seq(k:k) = WPh_seq_list(j)(k:k) !move probe sequence into a string
             counter1 = counter1 + 1
      444 continue
777 continue
!print*,string_seq !print string, this works
!do 309 j=0,274
      print*,probe_seq(j) !print array, this works
!309 continue
!Making sure everything is in the new sequence array
counter1 = 0
do 667 j=1, 573,1
      do 668 k=1,60,1
             TitinSeqlist(counter1) = sequencelist(j)(k:k)
             !print *, sequencelist(i)
            counter1 = counter1 + 1
      668 continue
667 continue
do 9000 j=0,34349
      z = j + 1!first index of array is zero, first index of a string is 1, therefore we update j
      Titin string(z:z) = TitinSeglist(j)
      !print*,TitinSeglist(j)
9000 continue
print*, Titin string
! Exact Alignment Count
exactAlignCount=0
do 1313 j=1, 34350 #do the length of titin. J will serve as the first index when we slice the
titin string
      match_window = Titin_string(j:j+273)! we do 273 bc slice is inclusive in fortran
      !match window in the first iteration of the loop will be the first 274 AA in titin.
      if (match_window == string_seq) then !string_seq is the entire probe seq
             exactAlignCount = exactAlignCount +1 !if perfect match add 1 to counter
             !print*,"Its a match"
      Endif
1313 continue
```

```
!print*, exactAlignCount
!!!!!!!!!!!!! partial match
! Partial Alignment Count
user = 274 * user !multiply the sequence length by the user input variable
partialAlignCount=0
call system_clock(t1,count_rate) !start timing for partial alignment
do 1314 j=1, 34350 !This approach requires a nested loop. First loop will grab the
match window from titin and then we loop over that in another nested loop
       match_window2 = Titin_string(j:j+273)
       threshcounter = 0
             do 1315 u=1, 274 !In this loop we count how many AA matches are in the titin
match window to the probe seq
              if (match_window2(u:u) == string_seq(u:u)) then
             threshcounter = threshcounter + 1 !we keep counting an AA matches
              if (threshcounter > user) then !Once we meet the threshold for the given
percentage then we exit the loop and add one to the partialAlignCount
                     partialAlignCount = partialAlignCount + 1
                     EXIT
              endif
             endif
       1315 continue
1314 continue
call system_clock(t2) !end timing
print*, partialAlignCount
!!!!!!!!!AA frequencies in titin
do 899 x=1.20
       tempchar = aaUni(x:x) !One AA at a time. Already sorted
       counter2 = 0
       do 669 w=0, 34349 iterate the length of titin. Notice that tempchar will be the same AA
for this nested loop. This is how we count all A and C and so one.
              !print *, TitinSeglist(w)
              if (TitinSeqlist(w) == tempchar) then !Compare current AA to tempchar
              counter2 = counter2 + 1
             endif
       669 continue
       AA_count(x) = counter2 append that count. We know order of counts because of aaUni
899 continue
```

```
!!!!!!!!!!!!!!!!!!!!!Create unique mers using a sorted string of AA
counter2 = 0
!These loops will loop through aaUni = "ACDEFGHIKLMNPQRSTVWY" twice and create 2mers.
!We needed a list of unique 2mers before we could count them in titin.
do 788 x=1,20 !Index for first letter in string
       do 799 y=1,20 !index for second letter in string
              uni_kmer(counter2) = aaUni(x:x) // aaUni(y:y) !add unique 2mer into list
              !This will look like "AA", "AC", "AD"......"CA", "CC", "CD"....
              counter2 = counter2 + 1
       799 continue
788 continue
do 678 x=0,399 !Count 2mers in titin using unique 2mer array created above
       counter2 = 0
       do 567 y=0, 34349
              tempkmer = TitinSeqlist(y) // TitinSeqlist(y+1)
              if (uni kmer(x) == tempkmer) then
              counter2 = counter2 + 1
              endif
       567 continue
       kmer_count(x) = counter2
       !print *, uni_kmer(x), counter2
678 continue
!print *, "A
                     С
                                                                               G
                                    D
                                                  Ε
                                                                               Т
Н
              Κ
                            M
                                   Ν
                                           Р
                                                  O
                                                         R
                                                                                      V
     Υ"
W
counter2 = 1
do 9654 x=0, 399, 20 !Used to print and write a formatted table
       print *, aaUni(counter2:counter2), kmer count(x:x+19) !this is working
       counter2 = counter2 + 1
9654 continue
wall_clock_time = real(t2-t1)/real(count_rate)
print *, wall_clock_time
end program readFile
```

Appendix:

- i. Python Code
- ii . Output of Matrix To a File
- iii.Histogram

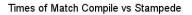
i:Python for part 1

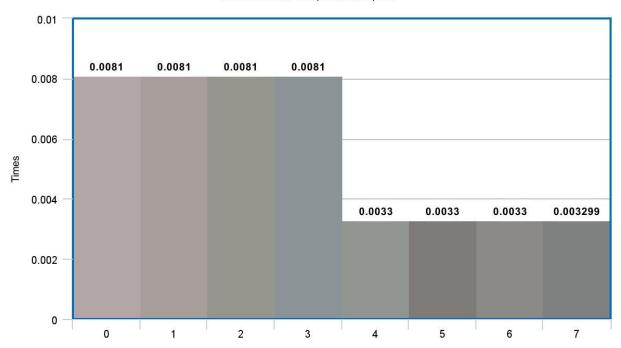
```
fh = open("titin",'r')
full seq = ""
freq_dict = {}
kmer_dict = {}
for x in fh:
  if ">" not in x:
      x = x.strip()
      full_seq += x
for x in full_seq:
  if x in freq_dict: freq_dict[x] += 1
   else: freq_dict[x] = 1
AA letters = []
for k, v in sorted(freq dict.items()):
  AA_letters.append(k)
for x in range(len(full_seq)):
   if len(full_seq[x:x+2]) == 2:
       if full seq[x:x+2] in kmer dict:
           kmer_dict[full_seq[x:x+2]] += 1
       else:
           kmer_dict[full_seq[x:x+2]] = 1
new_dict = {}
kmer_tuple = []
for k, v in sorted(kmer dict.items()):
   kmer tuple.append((k,v))
for x in AA_letters: print x, "\t",
print "\n"
for x in AA_letters:
  print x,
  for y in kmer tuple:
       if x == y[0][0]:
          print y[1],
  print "\n"
print kmer tuple
print freq_dict
```

ii.Matrix

	A	,		C			D		E	F	G		Н	I K	L	М	N	P Q	R S	-
A	٧	129		24					46				180	83		56	151	60	110	153
C		30	194	1		25	36	68	6	21	14	16	65	16	3	30	10	35	32	48
D	44	131	30	21		82	14	92	36	223	22	103	104	116	11	47	110	55	73	153
			158																	
Е	136	200	296		34	119	181	328	88	200	38	226		181	45	165	296	48	127	144
F	72	38	105		3	48	11	83		39			98			19		20	163	50
G		84		20				197		194	24	94	207	134		50	157	82	107	219
н	150		129			30	89	40		19		44		50			18			28
т.		102		23		90		188	38	80	31	120	210	140	23	69	107	67	120	190
	235		189																	
K	158	212	321	46		240		266	63	142	36	167	222	176	34	101	270	50	128	135
L	215	88	150		17	130	42	282	39	65	42	113	234	107		56	113	100	129	162
M		34				18		28			10		45	26		10	18			36
N	50	85	28			59		91				80	98	83		34		18	44	88
P	48	160	131	18	16	95	62	291	64	173	22	185	205	148	18	33	410	47	74	174
0	125	51	242	14		51		99	50	50	10	66	76	71	15	40	41	19	38	51
V	60																			
R	60	99	249		32	102	48	141	58	42		127		128	30	63	115	39	58	84
S	143	206	201		72	176	47	201	95	174	28	129		151	40	63	147		104	180
Т		181		69		105			89	167		154	207	198	29	95		56		180
V	166	183	276	46	80	100		289	62	146	48	181	273	196	30	133	304	95	124	278
W	379	14	240	3	26	16		38	48	10	11	13	43	32	11	16	3	17	14	45
			36																	
Y	122	38	86	3	10	60	28	89	40	51	21	99	85	44	11	15	32	48	52	65

iii.Histogram





We notice that the times are constant, no matter what the match percentage is. This is because we have a nested do loop with counters that are evaluated against thresholds. Nested do loops function in quadratic time (n*m).