**Deciphering complex traits using deep combinatorial genetic analysis**

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**Abstract**

Many traits are controlled by complex biological systems encoded by multiple genes. An accurate mechanistic understanding of these systems may require mapping complex genotype-to-trait relationships, which in turn requires perturbing genes in many different combinations and observing the impact. Here we describe a method to efficiently engineer and phenotype many multi-gene variant combinations within a targeted gene set, enabling a deep combinatorial genetic analysis (DCGA). We use this method to generate and profile each of ~6,000 combinations of knockouts amongst 16 yeast ABC transporters for resistance to each of 16 drugs. The resulting genotype-to-resistance landscapes revealed many complex drug-dependent genetic interactions. For example, we identified a quadruple knockout (*snq2*∆ *yor1*∆ *ybt1*∆ *ycf1*∆) which conferred resistance to fluconazole and ketoconazole, and found that the addition of *pdr5*∆ yielded a quintuple mutant with high sensitivity to azoles. To better understand the mechanistic basis for the many complex interactions, we hypothesized and computationally reconstructed a non-linear system model, which related genotype to phenotype with high overall accuracy, and guided further experimental characterization of the azole-resistant quadruple knockout. These results motivate the future use of DCGA to discover unexpected high-order genotype-to-trait relationships and exploit them to model complex biological systems.

**Introduction**

Extensive functional interdependency and redundancy in many biological systems results in traits which cannot be straightforwardly understood by observing the individual effects of sequence variants1–4. Genes often encode gene products which form interdependent pathways and protein complexes, such that combinatorial genetic perturbations can yield surprising phenotypes. This phenomenon defines genetic interaction. Observing the phenotypic effects of genes varied in combination, i.e., performing a combinatorial genetic analysis (CGA), can uncover functional dependencies between genes, and can be used to reconstruct large-scale maps of gene co-function5. The ability of CGAs to better understand gene function has been amply demonstrated by comprehensive two-gene interaction maps in yeast via growth profiling of >23 million different double knockout combinations5. The number of genes yielding a phenotype under standard growth conditions climbs from ~2,0006 to ~5,000 when genes are perturbed in pairs5. Furthermore, the resulting genetic interaction map enabled genes to be clustered into functional groups5,7, while smaller-scale CGAs have been used to better predict the genetic basis of drug resistance8, and to inform order-of-action in biological pathways9.

CGA of many biological traits has shown that additional genetic interactions can arise from the simultaneous perturbation of more than two genes. In yeast, triple mutant interactions (for which a triple mutant phenotype cannot be simply explained by the component single and double mutant phenotypes) are likely to vastly outnumber two-gene interactions10. [Fritz stopped here] Such triple mutant interactions can arise when perturbing both simple and complex biological systems11. Many individual examples of more complex multi-variant effects have also been reported, including those arising from >5, 7, and >20 combined variants12–14.

In *E. coli*, it has been suggested that unexpected multi-variant effects can open up many indirect mutational paths which lead to antibiotic resistance15. In vertebrates, complex multi-variant effects have been found to mediate medically-relevant traits such as myeloid malignancies16,17, and their discovery can be practically useful – for example, the induction of pluripotent stem cells is achieved by the simultaneous exogenous expression of four factors18.

Despite their estimate prevalence and contribution towards understanding and manipulating gene and system functions, systematic analysis of complex multi-variant effects are lacking because of major experimental challenges in generating and profiling the potentially vast number of strain variants required to perform a more exhaustive or ‘deep’ combinatorial genetic analysis (DCGA). While genome-wide multi-variant surveys can quickly grow to require an impossible number of strains, this can be constrained by using prior knowledge to rationally prioritize and target a reasonably-sized system of genes for analysis. Curently-available methods, however, make even a targeted DCGA inefficient or infeasible. A major bottleneck remains in the large-scale engineering and profiling of many multi-variant strains, partly because there is a limited availability of usable selection markers20. Methods have been designed to make modifications at multiple loci simultaneously (multiplex automated genome engineering – MAGE)21,22, but do not address the challenge of isolating and profiling the large number of strains required to perform a DCGA. While extensions of MAGE are being developed for in-depth combinatorial genetic and phenotypic characterization in *E. coli* 23,24, these are not yet suitable for accurate profiling.

We first develop and describe an ‘engineered population profiling’ strategy to enable a DCGA of a targeted system of genes. We then demonstrate a DCGA by profiling each of ~5,000 knockout combinations amongst 16 yeast ABC transporters involved in multi-drug resistance for sensitivity to each of 16 compounds, revealing many complex drug-dependent genetic interactions. We then leverage these complex interactions to hypothesize and computationally reconstruct a formal system model of ABC-mediated drug resistance, which was used to understand and further explore the mechanistic basis of a quadruple knockout (*snq2∆yor1∆ybt1∆ycf1∆*) that conferred unexpectedly high *PDR5*-dependent resistance to fluconazole and ketoconazole. The described strategy presents a framework for DCGA of other gene sets, and demonstrates that the revealed complex genotype-to-phenotype relationships can generate more complete and accurate maps of gene and system function in order to better understand complex traits.

**Results**

**Engineered Population Profiling: A Scheme for Generating Combinatorial Mutants**

A straightforward method to introduce complex genetic variation is through a cross between two outbred individuals, so that all progeny inherit a random variant at each position that differs between the parents8. Genotying and profiling these progeny for traits such as gene expression25 or small molecule resistance26 then allows statistical association of the resulting variants to the measured phenotypes. However, such approaches have been traditionally used at a large scale with natural isolates rather than engineered strains26, presenting several limitations. For example, many yeast genes known to be important through knockout studies, such as ABC transporters27, have limited known natural variants. Furthermore, the use of diverse parents differing at hundreds of thousands of positions often results in associations of many linked variants at a single locus to a trait, making it difficult to recover causal variants. A large number of positions varying between parents may also result in a prohibitive or impossible number of individuals required for statistical reconstruction of complex variant-to-phenotype associations, and moreover each term in any recovered complex association will be of unknown causality. To extend cross-based approaches beyond natural strains, we propose a population engineering strategy where one or few individuals containing all the desired variants are created using molecular tools, and are then segregated randomly over a population using one or more controlled crosses.

To enable population engineering, not only must the desired variation be introduced into progeny strains, but these progeny must then be individually identified, genotyped, and phenotyped at a large scale. This can be achieved by the use of various molecular markers. In the strategy described here, we first transform a parental strain with a complex pool of random DNA barcodes28, so that barcode sequence can be used to identify individual parents. Each progeny, resulting from a mating and sporulation event between two individual strains, will then inherit at most one unique barcode from one of its parents, acting as an ‘individual identifier’. The presence of an individual identifier allows for the use of other molecular tools to perform large-scale individual genotyping and phenotyping. Isolating a strain, sequencing its identifier barcode, and performing PCR-based genotyping, for example, associates the identifer barcode with a genotype, thereafter allowing for a ‘barcode-to-genotype lookup’. A second barcode (or combination of barcodes) can be added to uniquely tag PCR products from each individual, so that instead of isolating and sequencing each strain individually, many barcodes can be mapped to their genotype *en masse* by pooling many strains and computationally de-multiplexing the PCR products arising from each individual after a single round of high-throughput sequencing29. The individual identifier barcode also allows for straightforward growth-based phenotyping, as relative strain abundance can be measured over time in a competitive liquid pool using high-throughput barcode sequencing30. Ultimately, the combination of a cross-based approach with molecular engineering tools can result in a large population of genotyped and phenotyped multi-variant individuals, which can be stored as a liquid pool and repeatedly characterized for growth under multiple conditions.

**The Barcoder Pool: A Universal Tool for Population Engineering**

A key step to the proposed population engineering scheme is the introduction of a unique molecular barcode into the genome of each individual. For this, we adapted the Barcoder28 strategy to create a large pool of uniquely-identifiable individual parents. To introduce a high diversity of unique barcodes into a ‘wild-type’ parental strain (RY0148), we used a Cre-mediated recombination strategy. First, we modified RY0148 by the addition of Loxp and Lox2272 recombination sites into the HO locus. Then, we transformed this modifed strain with a complex plasmid pool containing DNA tags flanked by Loxp/Lox2272 recombination sites (Fig S1). Finally, we induced the expression of Cre to allow for recombination and integration of the plasmid-borne barcodes into the HO locus (Fig S1). Because uniquely-identifiable individuals are required for many potential population engineering strategies, this ‘universal barcoder pool’ can be used to engineer a barcoded population when mated with any multi-mutant yeast strain containing the appropriate selection markers (Fig. S1, Methods).

**Engineering a Population of High-Order Combinatorial ABC Transporter Knockouts**

After creating a universal parental barcoder pool, we attempted a cross-based population engineering approach using a previously-created strain with knockouts at all 16 ATP Binding Cassette transporters involved in yeast multidrug resistance (ABC-16)20. The choice of ABC transporters as the pilot gene set was motivated by several factors. First, because the ABC-16 strain does not show major fitness defects in the absence of drugs, it is reasonable to expect that most or all progeny containing a subset of these 16 knockouts would be viable, avoiding the systematic exclusion of certain genotype combinations in the initial population. Second, given the unexpected drug sensitivity profile of the ABC-16 strain compared to individual knockouts, it is suggested that unexpected multi-knockout effects within the ABC transporters could mediate both drug sensitivity and resistance, with some two-knockout ‘compensatory’ interactions mediating resistance having already been found20,31,32. As ABC transporters are part of a large gene family with over 10,000 members across all three domains of life33, similar ‘compensatory’ genetic interactions have been found in their mammalian homologs. For example, a compensatory activation of ABCC3 has been observed upon disruption of ABCC2 in rats34 and in humans in the context of Dubin-Johnson syndrome35. Similarly, mouse ABCG5 and ABCG8 are activated in response to disruption of ABCG2 (a protein involved in breast cancer xenobiotic resistance)36. Ultimately, a DCGA of ABC transporters provides the opportunity to genetically dissect a functionally-important and conserved gene family which mediates functions such as multidrug resistance, disease progression, and basic cellular homeostasis in many organisms37,38.

To begin engineering the ABC transporter knockout population, we first mated the universal barcoder pool with ABC-16, as we had previously engineered this strain to contain the appropriate markers to perform mating, sporulation, and barcoded haploid selection (Methods). Using automated colony picking, ~5,000 MAT**a** and ~5,000 MAT**α** segregants containing random knockouts at 16 genes were isolated into a collection of 384-well plates.

To validate the mating and selection strategies, we pooled the MAT**a** and MAT**α** collections and subjected them to cell sorting, confirming haploidy of the overall pool (Fig. S2D), and furthermore we tested that samples from each pool do not exhibit any growth in the selection conditions of the opposite mating type.

**Genotyping and Barcode Identification for the Engineered Population**

After creating a large collection of barcoded multi-knockout progeny, we aimed to genotype each strain and link this genotype to an individual DNA barcode identifier *en masse*. By adapting our previously-developed row-column-plate PCR (RCP-PCR) strategy29, we amplified strain-specific barcodes and deletion-specific products for each individual. In the ABC-16 strain, each knockout is replaced by a barcode identifying the deleted gene, allowing the use of a single primer set for identification of all gene deletions present in a strain when its amplification products are sequenced (Fig. 1). RCP-PCR introduces additional sequences indicating the plate, row, and column of origin for each amplification product (Methods; Fig. 1), allowing for pooling of PCR products obtained from each well, coupled with high-throughput sequencing to computationally reconstruct a genotype-to-barcode map at a population-wide level (Methods). To validate and calibrate the genotypes and associations determined by high-throughput-sequencing, multiple replicates of 40 individually genotyped ‘gold standard’ strains, as well as two additional control strains with known genotypes, were added to the collection at defined positions (Methods; Data S2). Using data from calibration strains, we estimated an overall genotyping accuracy of 93.2% (Fig. S2A, Methods). An independent method relying on the distribution of knockouts in the pool estimated a similar overall accuracy of 93.8% (Fig. S2C, Methods). Based on the genotyping data, all genes were either unlinked or weakly linked except for *BPT1* and *YBT1* (Fig. S2B; r = 0.49). This moderate linkage was expected as these genes are both on chromosome XII, and are separated by 70.1kb. Surprisingly, three gene pairs – *YOR1*-*YCF1*, *YOR1*-*BPT1*, and *SNQ2*-*PDR5* – had weak but significant negative apparent linkage (-0.04 ≥ r ≥ -0.08), suggesting a weak negative genetic interaction between them (Fig. S2B). This weak interaction may have arisen by some genotype-dependent selection during the sporulation, haploid selection, or automated colony picking steps, and demonstrates the power of a large engineered population to detect subtle associations. Overall, usable genotyping data linked to at least one unique DNA barcode was obtained for 6,709 progeny with 5,095 unique genotypes.

**Phenotyping the Engineered Population for Diverse Drug Resistance Traits**

After establishing a genotype and barcode identity for many individuals, we profiled their resistance to a collection of drugs. Arrayed collections were transferred to two liquid pools separated by mating type (MAT**a** and MAT**α**). These pools were grown in each of 16 different anticancer and antifungal drugs (Data S3), as well as a solvent control. Using high-throughput strain barcode sequencing30, strain frequency was measured at five time points (corresponding to 0, 5, 10, 15, and 20 generations of overall pool growth, Fig. 1) and a drug resistance score was computed for 3,221 MAT**a** and 3,592 MAT**α** strains by comparing esimated growth in each drug to the solvent control (Methods, Fig. 1, Data S5). Strains which were initially absent in the solvent control, as well as strains which showed a considerable drug-independent growth defect, were excluded from analysis (Data S5).

To evaluate sequencing complexity and technical reproducibility, we measured correlation between the resistance scores generated by the UP and DN barcode tags. Resistance scores for 8 drugs were highly correlated (r > 0.9 for both MAT**a** and MAT**α** populations), and moderately correlated (minimum r > 0.75) for 4 other drugs (Fig. S3 A-C). For the remaining 4 drugs, resistance was not highly correlated between UP and DN tags (Fig S3). The expected difference between UP and DN tag estimates were found to be similar in all samples, but a lack of strain-to-strain resistance in some drugs drove their poor tag correlations, and suggests that the 16 ABC transporters tested did not play a major role in mediating resistance to 4 of the tested compounds (Fig. S3C).

**Grouped Profiles Recreate Consistent Drug Resistance Relationships**

To explore the multi-knockout data, we first aimed to identify the subset of ABC transporters which are relevant for resistance to each drug. Given the diverse population, we tested

for knockout effects aggregated over all genetic backgrounds in the population by using a linear model with only single-gene terms to uncover ‘marginal’ associations of each knockout to drug resistance (Methods). While prior literature had reported on the effects of *snq2∆*, *pdr5∆*,and *yor1∆* for the tested drugs, this approach found previously-unreported associations involving deletion of the vacuolar ABC transporters (*ycf1∆*, *ybt1∆*, and *bpt1∆*) in many of the drugs tested (Fig. S4, Data S6). The latter three knockouts had comparably small marginal effects, which can indicate either a minor role towards resistance to the tested drugs and/or a greater dependence of their effects on other knockouts. From the marginal associations which were found in both the MAT**a** and MAT**α** pools (Data S6), 16 out of 21 previously-known single knockout phenotypes were reproduced (Fig. S4; Data S7).

After identification of *snq2∆*, *pdr5∆*, *yor1∆*, *ycf1∆*, *ybt1∆*, and *bpt1∆* as commonly-relevant for resistance to the tested drugs, we grouped strains by their genotype at these 6 transporters, allowing for knockouts at any of the other 10 loci within each group. While not all drug resistance profiles involved these 6 transporters, this 64-genotype grouping allowed for the establishment of a common knockout sensitivity profile for all drugs, which allowed for a comparison of their effects even when tested on two independent populations (Fig S5). Under drugs such as camptothecin and tamoxifen, the MAT**a** and MAT**α** populations had strikingly similar profiles (Fig. 2A), and high reproducibility was often observed between different populations grown in the same drug (r ≥ 0.9 for 11 of 16 drugs, Fig. 2A, Data S7). These profiles, however, did not make readily-apparent the main transporters involved in each drug, nor did they directly show potentially-differing knockout effects in diverse genetic backgrounds. To allow for a broad ‘fitness landscape’ overview and a better exploration of all possible trajectories towards a six-knockout state, we developed a radial representation centered on the wildtype, with concentric extensions showing the effects of cumulative knockouts (Fig. 2C). Reflecting the high profile reproducibility, these radial path structures were visually consistent for independent populations tested on the same compound, while differing greatly between unrelated compounds (Fig. 2D, S7). Furthermore, these radial overviews visually highlighted the main ABC transporters mediating resistance to each drug, and made apparent which multi-knockout genotypes mediated both drug resistance and sensitivity.

**DCGA Reveals High-Order Combinatorial Drug Resistance Effects**

We represented the grouped profiles by a linear fitness landscape, which allowed a more detailed evaluation of transporter knockout effects in the context of any other 32 knockout combinations (Fig. 3A). We evaluated the ability of these fitness landscapes to reproducibly capture previously-known surprising multi-knockout traits. Specifically, we investigated the previously-reported *SNQ2*-mediated benomyl sensitivity, and ‘compensatory activation’ of Snq2 when *PDR5* and/or *YOR1* are disrupted31. In both the MAT**a** and MAT**α** pools, we confidently recapitulated the *snq2∆* sensitivity (Fig. 3A, S8, *p* = 9.2e-24 MAT**a**, 6.3e-44 MAT**α**; Wilcoxon rank sum test), and consistent with compensatory activation, we found the *pdr5∆* and *pdr5∆yor1∆* strains to exhibit increased resistance compared to the wild type (Fig. 3A, S8, *p* = 3.8e-33 MAT**a**, 4.0e-54 MAT**α** for *pdr5∆* and *p* = 2.0e-32 MAT**a**, 1.4e-53 MAT**α** for *pdr5∆yor1∆*). Additionally, this map revealed that the effects of *yor1∆* were background dependent – *yor1∆* was not found to have a significant effect relative to the six-gene wildtype group (*p* = 0.69 MAT**a**, *p* = 0.33 MAT**α**), to increase fitness relative to *pdr5∆* as *yor1∆pdr5∆* (*p* = 2.0e-32 MAT**a**, *p* = 1.57e-37 MAT**α**), and to decrease fitness relative to *snq2∆* as *yor1∆snq2∆* (*p* = 2.6e-09 in MAT**a** and *p* = 1.3e-05 in MAT**α**; Fig. 3A, S8). The *yor1∆snq2∆* phenotype is novel and suggests that Yor1 can serve a secondary role in benomyl resistance, for example by providing some efflux in the absence of Snq2.

After validating the ability of the fitness landscapes to capture previously-known single- and multi-knockout phenotypes in benomyl, we explored surprising genetic relationships under all tested drugs. In the most straightforward cases, sensitive multi-knockout groups were found where removal of a subset of the underlying genes had much less apparent consequences. These cases are illustrated, for example, by the *snq2∆pdr5∆* group under camptothecin (Fig. S8), and by the *snq2∆pdr5∆ybt1∆* and *snq2∆pdr5∆yor1∆* groups under mitoxantrone (Fig. 3A, S8). Broadly, these sensitivity patterns suggest that multiple transporters are able efflux a given drug, conferring redundancy when overall resistance is measured. In several other cases, the fitness landscapes involved complex multi-knockout patterns mediating both drug resistance and sensitivity. In many compounds, multiple paths led to ‘peaks’ conferring more resistance than the wild type state (Fig. S8). Under fluconazole, ketoconazole, and itraconazole, a quadruple deletion – *snq2∆ybt1∆ycf1∆yor1∆* – led to resistance, whereas *pdr5∆snq2∆ybt1∆ycf1∆yor1∆* was comparably as sensitive as the *pdr5∆* group (Fig. 3A, S8). Furthermore, combinations of one or two knockouts within these four genes resulted in suble or absent effects. These findings extend a previously-reported compensatory activation of *PDR5*31, but suggest that the effect is more apparent with further knockouts in *YCF1* and *YBT1* in addition to *SNQ2* and *PDR5*.

Because each group contains a heterogeneous population of individuals with additional background knockouts, we could further visualize the underlying distribution within each group. In the context of fluconazole resistance (Fig. 3B), this revealed that variance in resistance within the *pdr5∆* groups is much less than groups containing *PDR5* (e.g. ; *p* = 2.8e-07, 2.5e-26, Bartlett Test). The lack of variability in the *pdr5∆* groups may be explained by their unconditionally high fluconazole sensitivity, whereas the variability in the *PDR5* groups may reflect still-unidentified and more complex ABC transporter knockout effects in these genetic backgrounds.

We aimed to model and fomally capture the many striking and reproducible multi-knockout drug resistance phenotypes evident within the engineered population. For this, we used a log-linear model which extends the multiplicative expectation of combined genetic effects39 to incorporate potential interactions (denoted by ε) of arbitrary complexity (see Methods). We treated the search for complex genetic interactions as a feature prioritization and selection problem, testing for the presence of up to to 4-gene interactions within a total space of 2,516 coefficients (See Methods; Fig. 3C). We found 23 2-way, 12 3-way, and 8 4-way interactions which were reproducible between the MAT**a** and MAT**α** pools at a stringent Bonferroni corrected *p*-value cutoff of 0.05 (Data S6). To verify generalizability of the modelled genetic interactions, we tested the ability of models made from pooled resistance data of one mating to predict knockout phenotypes of the opposite mating type (Fig. S9), finding similar population-wide predictive power. This formalized approach captured many of the striking phenotypes found by examination of the fitness landscapes. For example, *yor1∆* was found to have no main effect under benomyl, to have a positive genetic interaction with *pdr5∆*, and to have a negative genetic interaction with *snq2∆* (Fig. 3C, Data S6). In camptothecin, *pdr5∆* and *snq2∆* each had a minor effect, as well as a strong negative interaction between them (Fig. 3C, Data S6). Similarly, the surprising phenotype in mitoxantrone was modelled as a combination of a small marginal effect of *snq2∆*, a negative interaction , and further triple negative interactions upon *ybt1∆* and *yor1∆* as and (Fig. 3C, Data S6). This genetic interaction pattern suggests not only that these four genes efflux mitoxantrone in parallel, but also a differential efflux capacity between them –it may be hypothesized, for example, that Snq2has the highest efflux activity because it is the only one that shows a knockout defect by itself. A similar ‘parallel resistance’ phenotype was observed for *pdr5∆snq2∆yor1∆* in cisplatin (Fig. 3C, Data S6). Interestingly, the multi-knockout resistance phenotype in fluconazole was modelled (in addition to one- and two- gene effects involving *pdr5∆*, *yor1∆*, and *ybt1∆*) as the combination of three positive three-gene interactions (, , ). The dependence of these multi-knockout effects on the presence of *PDR5* was then further modelled as three additional negative four-way interactions(, , ).

**Detailed Validation of Complex Genetic Inhibition Model**

Through analysis of the grouped fitness landscapes and formally-determined genetic interactions, we modelled a set of transporter-transporter relationships. These relationships were highly drug-dependent, and were derived from a combination of evidence from two, three, and four knockout phenotypes (Fig. 4A, Data S8). We modelled all phenotypes as arising either from parallel clearance of a drug by two or more transporters, and/or the antagonism of an ABC transporter by the presence of another (Fig. 4A). These models suggest an extensive role for *YBT1* and *YCF1* in many drugs – *YBT1* is hypothesized to work in parallel with *SNQ2*, *PDR5*, or *YOR1* in the context of mitoxantrone, itraconazole, and methotrexate resistance, and to antagonize *PDR5* in the context of fluconazole resistance (Fig. 4A). *YCF1* is modelled to antagonize *SNQ2* in bisantrene resistance, to antagonize *PDR5* in fluconazole and ketoconazole, and to work in parallel with *SNQ2* in the context of benomyl resistance (Fig. 4A). The absence of these phenotypes in prior literature may be expected given the absent or subtle single-knockout effects of *YBT1* and *YCF1* in many drugs. We note that *BPT1* has been excluded from these models, despite its knockout seeming to confer resistance to several compounds, especially when combined with other genetic disruptions (Fig ??). However, the genetic effects of *bpt1∆* in the engineered population are inconsistent a role in effluxing any of the drugs tested, or antagonizing any other ABC transporter studied. The *bpt1∆* profiles may involve complex genetic interactions of ABC transporters with other systems such as those mediating membrane permeability40, but this was not further explored. Interactions of *bpt1∆* and other knockouts with genes other than the studied ABC transporters may also be responsible for the higher-than-wildtype resistance of the ABC-16 strain to some compounds20,40.

To validate the knockout-to-phenotype associations revealed by the engineered population profiles, we sought to generate individual strains containing all knockout combinations of *pdr5∆, snq2∆, yor1∆, ybt1∆,* and *ycf1∆* in a wildtype background. These strains were generated by mating a 7-deletion strain in the collection with a wildtype and performing a series of genotyping and selection steps (Fig 4B, Methods). Fluconazole resistance of these individual strains correlated well with the pooled resistance, both when measured as the expected concentration to cause 50% inhibition (r = 0.92, Fig 4C - D), and as the individual growth at a concentration identical to the pool (r = 0.9, Fig. S10). Congruent with the pool data, *snq2∆yor1∆ybt1∆ycf1∆* had the highest resistance, and only minor resistance effects were seen for strains with one and two knockout combinations amongst these four genes. Given the ‘parallel repression’ model of Pdr5 by the presence of *SNQ2*, *YOR1*, *YBT1*, and *YCF1*, we explored two potential mechanisms of inhibition. Inhibition of Pdr5p by *SNQ2* and *YOR1* has been previously reported, and thought to be mediated by transcriptional activation by Pdr131, although the mechanism underlying this compensatory transcriptional activation is unknown. To investigate whether *PDR5* mRNA abundance was elevated in the *snq2∆yor1∆ybt1∆ycf1∆* compared to the wild type and *snq2∆yor1∆*, we performed qRT-PCR in both the solvent control and fluconazole. In both the solvent control and fluconazole, *PDR5* transcript abundance was elevated in the *snq2∆yor1∆ybt1∆ycf1∆* strain relative to the wild type strain (*p* = 0.029 in DMSO; *p* = 0.019 in fluconazole; Fig 4D). Surprisingly, we did not find evidence of increased *PDR5* transcript levels in the *snq2∆yor1∆* strain (*p* = 0.83 in DMSO; *p* = 0.35 in fluconazole; Fig. 4D), suggesting strain-specific differenced in the dynamics of the compensatory activation phenomenon.

Given the lack of evidence for transcriptional induction in the *snq2∆yor1∆ strain*, but modest improvements in growth in the grouped data (median log2-resistance of -0.17 vs -0.28, *p* = 1.5e-14 in MAT**a** and 1.4e-32 in MAT**α**), we explored alternative mechanisms of repression. A previous study investigating an analogous compensatory activation of Snq2 by deletion of *PDR5* and *YOR1* found unchanged protein abundance and localization upon knockout of these two genes, and suggested that physical interaction between these two proteins may lead to their mutual repression32. Consistent with this model, homodimers of Pdr5p, Snq2p, and Yor1p have been detected using the mDHFR protein complementation assay (PCA)41,42, and a Pdr5p homodimer has also been detected using the membrane yeast-two-hybrid (MYTH)32 assay. An investigation of Pdr5p structure using single-particle electron microscopy furthermore finds a four-lobed homodimeric conformation, with an asymetric conformation in each lobe supporting a functional dependence between the two units which may be disrupted by interactions with its paralogues43. Consistent with an interaction-based inhibition model, a Pdr5-Snq2 interaction has been reported in both MYTH and PCA32,41. However, this interaction-based inhibition model would also predict a previously-unreported Pdr5-Yor1 heterodimer, which was re-tested here using both MYTH and PCA. While PCA did not find evidence for this interaction (Fig. S11), it was found using MYTH (Fig. 4F, S12). All previously-known MYTH and PCA interactions amongst Pdr5, Snq2, and Yor1 were also reproduced here (Fig. 4F, S11, S12). These protein-protein interaction patterns may also underlie other patterns of mutual repression between *YOR1, SNQ2*, and *PDR5* suggested in drugs other than fluconazole (Fig. 4A). Taken together, these experiments support a mixed inhibition model where all four genes appear tosuppress *PDR5* expression, while *SNQ2* and *YOR1* potentially inhibit the formation of an active homodimeric form.

**Discussion**

The lack of tools to efficiently engineer many combinations of gene variants and characterize their impact has resulted in a limited genetic understanding of many complex traits. We present a method to straightforwardly generate and profile a large population of multi-mutant individuals, and demonstrate that their characterization in diverse environments can reveal previously-unknown gene functions and an expanded model of gene-gene relationships, even in a within highly-studied gene family. Within the 16 ABC transporters studied here, knockout phenotypes were highly dependent on both the drug being tested and the genetic background, motivating similar exploration of other gene groups, and the further use of this population for studying the ABC-transporter-mediated drug clearance of other compounds over single-knockout approaches.

The development of a ‘universal barcoder pool’ enabled the use of a cross-based method to efficiently introduce mutations into a population of uniquely-identifiable cells, and for a straightforward adaptation of this method for use with other multi-gene deletion yeast strains. In other model organisms such as *C. elegans*, methods to introduce targeted gene knockouts44 or loss-of-function mutations45 may enable analogous strategies. The cross-based approach permits flexibility in the distribution of mutations between the two parents, permitting a similar strategy even if introduction of all mutations into a single individual would cause considerable defects or lethality. To further avoid such potential ‘dead ends’, it is also possible to extend the cross-based strategy using a more complex series of matings between several multi-mutant parents.

To extend engineered population profiling beyond cross-based methods, development of new tools may allow the direct introduction of multi-allele diversity into a population using molecular tools. While such ‘direct’ engineering may simplify cross-based approaches, it importantly allows for population engineering of non-mating model systems, such as human cell lines. The ability to work with cell lines would enable, for example, analogous studies of human ABC transporter function. However, the direct introduction of variation into many strains at multiple loci at intermediate frequency is more challenging than engineering one or a few parental strains, and this is therefore an area of future development24.

The engineered ABC-16 population is by far the largest collection of genotyped multi-mutants amongst these genes. However, the majority of knockout combinations amongst the 16 ABC transporters (~92%) remain to be generated and characterized. A major bottleneck to achieving the scalability required to generate the remaining strains was the requirement that a separate PCR reaction must be peformed on each isolated individual in order to link genotype to a DNA barcode. A method to perform genotyping and DNA barcode mapping in a pooled (i.e. ‘population-wide’) will likely permit the engineering of populations which are orders of magnitude larger than what was demonstrated here, allowing a better representation of all possible multi-knockout strains, and the exploration of even more combinatorially complex interactions involving multiple alleles at multiple genes4.

A major element in the demonstrated engineered population profiling approach was the use of a strain-specific molecular barcode. Use of the appropriate selection conditions coupled with sequencing of molecular barcodes alows for many traits which can be measured and linked to a specific genotype *en masse*. For example, drug efflux dynamics may be studied by incubating cells with fluorescence-conjugated drugs and measuring efflux-mediated reduction in fluoresence using a combination of cell sorting and barcode sequencing at various time points. Similar sorting strategies can convert many fluorescence-based assays into a barcode sequencing output, such as those using a reporter to study the effects of knockouts on the activation of signalling pathways, phosphorylation state, epigenetic modifications, or protein abundance46. In addition to DNA-based molecular barcodes, methods to genotype large populations of cells after imaging are being developed, which would allow high-content characterization of multi-knockout strains47. Thus, with the appropriate design, multi-knockout strains obtained by population engineering can be characterized for many phenotypes at a large scale.

We envision that the profiling of engineered populations will permit a DCGA of biological systems in multiple organisms. Future molecular tools will allow an expansion of the demonstrated approach in scale, allelic variety, and complexity of phenotypic characterization. Ultimately, this will allow for a much greater understanding of many gene functions and gene-gene relationships in the context of a living system.

**Materials and Methods**

**Yeast Strains**

RY0622/GM512 (Green Monster MAT**a**):

*MAT***a** *adp1Δ snq2Δ ycf1Δ pdr15Δ yor1Δ vmr1Δ pdr11Δ nft1Δ bpt1Δ ybt1Δ pdr18Δ yol075cΔ aus1Δ pdr5Δ pdr10Δ pdr12Δ can1Δ::GMToolkit*-**a** (*CMVpr-rtTA KanMX4 STE2pr-Sp-his5*) *his3Δ1 leu2Δ0 ura3Δ0 met15Δ0*

RY0146 (Toolkit-a strain):

*MAT****a*** *lyp1Δ his3Δ1 leu2Δ0 ura3Δ0 met15Δ0 can1Δ::GMToolkit-a (CMVpr-rtTA KanMX4 STE2pr-Sp-his5)*

RY0148 (Barcoder Strain MAT**α**):

*MAT****α*** *lyp1Δ his3Δ1 leu2Δ0 ura3Δ0 met15Δ0 can1Δ::GMToolkit-α (CMVpr-rtTA NatMX4 STE3pr-LEU2) ho*∆*::LoxP UP-tag HphMX4 DN-tag Lox2272*

**Media**

**SC (SC-His, SC-Leu, SC-Ura)**

**YPD (+HygroB, +Clonnat, +G418)**

**Creating the Barcoder Plasmid**

We added a barcoder locus flanked by LoxP and Lox2272 into a pSH47 plasmid backbone expressing GAL1pr-CRE. This barcoder locus consisted of a random 25bp DNA sequence (‘UP tag’) in between two common primer regions (US1 and US2), followed by a HphMX4 cassette, and another random 25bp DNA sequence (‘DN tag’) in between two common primer regions (DS1 and DS2).

First, a barcoded HphMX4 construct was created. HphMX4 was amplified from a pIS420 plasmid using the STEP1F and STEP1R primers containing HphMX4 homology and US2/DS1 overhangs (Data S1). The PCR program used for this step was 98°C for 30sec; 25 cycles of 98°C for 10sec, 59°C for 10sec, 72°C for 60sec; 72°C for 5min; 4°C forever. These PCR products were purified using a Qiagen Qiaspin kit and confirmed using 2% gel electrophoresis. To the resulting purified products, the STEP2F and STEP2R primers were used to add the random barcodes and US1/DS2 regions with the following PCR program: 98°C for 30sec; 25 cycles of 98°C for 10sec, 68°C for 10sec, 72°C for 60sec; 72°C for 5min; 4°C forever. These resulting products were again purified using a Qiagen Qiaspin kit and ~1.5-1.6kb products were confirmed using 2% gel electrophoresis. To add LoxP/Lox2272 sites, PCR was performed with the STEP2 products using the SacI-LoxP-HphMX4-Barcode-F / SacI-Lox2272-HphMX4-Barcode-R primers. The PCR program used for this step was: 98°C for 30sec; 26 cycles of 98°C for 15sec, 64°C for 20sec, 72°C for 65sec; 72°C for 5min; 4°C forever. The resulting PCR products were purified using a Qiagen Qiaspin Kit, and ~1950bp products were confirmed using 2% gel electrophoresis. Two PCR reactions were performed on the resulting products to confirm correct synthesis. The first PCR reaction was performed with the SacI Reamp F/US2 primer pairs, and the second was performed using DS1/SacI Reamp R primer pairs. The PCR program used for both of these reactions was: 98°C for 30sec; 25 cycles of 98°C for 10sec, 59°C for 15sec, 72°C for 30sec; 72°C for 5min; 4°C forever. Expected sizes (~132bp, 137bp) were confirmed using 4% gel electrophoresis. All above PCR reactions were performed using High Fidelity Phusion Master Mix (NEB).

To prepare for cloning of the barcoder locus, pSH47 was digested with SacI using 100μl of 250ng/μl pSH47, 100μl NEB Buffer 4, 10μl BSA, 10μl SacI-HF in 1ml sterile water. 100μl of this mixture was incubated at 37°C for two hours, and inactivated by incubation at 65°C for 20min. Digest products were purified using a Qiagen Qiaspin kit, and confirmed using 0.8% gel electrophoresis.

**Generating a Barcoder Strain**

A linear URA3 cassette flanked by LoxP and Lox2272 sites and homology to the HO gene was amplified from purified pIS418 with the 5'HO-LoxP-URA and URA-Lox2272-3'HO primers using the following PCR program: 98°C for 30sec; 25 cycles of 98°C for 10sec, 60°C for 10sec, 72°C for 70sec; 72°C for 5min; 4°C forever. This PCR reaction was performed using High Fidelity Phusion Master Mix (NEB) and was purified using Qiagen Qiaspin. This cassette was integrated into the HO locus of the RY0148 strain through transformation to serve as the ‘landing pad’ for barcode integration using an EZ transformation kit. Transformants selected for growth in SC –Ura plates, and were later verified to exhibit no growth in 5-FOA. A transformant was selected to confirm HO locus integration using three PCR reactions with the following primer pairs: 5'HO-URAreamp + midURA-5'; 5'HO-URAreamp + midURA-3'; 5'HO-URAreamp + 3'HO-URAreamp. All PCR reactions were performed using High Fidelity Phusion Master Mix (NEB) with the following program: 98°C for 30sec; 25 cycles of 98°C for 10sec, 50°C for 10sec, 72°C for 70sec; 72°C for 5min; 4°C forever. Expected PCR product size was confirmed using 2% gel electrophoresis.

The HO::LoxP-URA3-Lox2272 integrant strain was then transformed with a mixture of digested pSH47 and purified PCR products to enable in-yeast-assembly48. Transformation was carried out using a previously established protocol49, with a ~1:6 mixture of digested pSH47:HphMX4 barcode cassette (~12μg digested pSH47 and 15μg cassette). Transformants were grown at 30°C in YPG +HygroB plates for 3 days, allowing both selection of successful transformants and Gal1p-Cre induction. These cells were then scraped and grown overnight in 5-FOA plates to select against non-recombinant strains, and strains containing the recombined barcoder plasmid.

Twenty colonies were tested for barcode integration using PCR and Sanger sequencing. Lysates were made by mixing a sample of each colony with 2μl Sterile DNA Free Water, 2μl 0.2M pH 7.4 Sodium Phosphate Buffer, 0.5 μl 5U/μl zymoresearch zymolyase and incubated at 37°C for 25min and 95°C for 10 min, and stopped by adding 125μl of sterile DNA-free Water. To each lyzed colonly, two sets of primer pairs to verify the strain barcode-specific UP and DN tag - US2 and a sequence complementary to 5’ of the HO gene (5’HO); DS1 and a sequence complementary to the 3’ of the HO gene (3’HO), using the following program: 98°C for 30sec; 25 cycles of 98°C for 10sec, 59°C for 15sec, 72°C for 30sec; 72°C for 5min; 4°C forever. PCR reactions were performed using High Fidelity Phusion Master Mix (NEB) and analyzed using 4% gel electrophoresis to verify the presence of 263bp and 251bp bands. EXOSAP purification was performed on the PCR products by adding 10μl EXOSAP mix (0.025μl ExoI (0.5U), 0.1μl Antarctic Phosphatase (0.5U), 3.5μl 10X Antarctic Phosphatase Buffer, 6.375μl dH2O) to 25μl of PCR products and incubating at 37°C for 30min; 80°C for 20min, then diluting with 35μl of DNA-free H2O to stop the reaction. Diluted EXOSAP products were Sanger sequenced with the 5’HO seq and 3’HO seq primers to confirm the correct barcode construct.

**Creating A ‘Gold Standard’ Genotyped Set**

To create a ‘Gold Standard’ genotyped set, 40 progeny strains (19 MAT**a** and 21 MAT**α**) were subject to individual strain genotyping. For these 40 strains, and for an RY0148 isolate, the strain-specific UP and DN tags were also PCR amplified using two sets of primers and subject to Sanger sequencing as above.

To genotype each strain at the 16 ABC transporter loci, two PCR reactions were performed for each locus - one to determine the presence of a GFP integration cassette, and another to determine the presence of the wild type gene, as previously described20. For the cassette confirmation reactions, locus–specific PCR primers from the 5′ flanking sequences of each gene were paired with a common primer complementary to the *GFP* cassette (Data S2). Gene presence confirmation primers were designed individually for each gene (Data S2). PCR reactions were performed with a Platinum HiFi mix using the following program: 94°C for 2min; 34 cycles of 94°C for 30sec, 55°C for 30sec, 68°C for 60sec; 68°C for 10min; 4°C forever. PCR products were analyzed using gel electrophoresis.

**Generating Barcoded Random Knockout Progeny**

Mating, sporulation, and haploid selection was performed between the RY0622 ‘Green Monster’ strain (MAT**a**) and the RY0148 barcoder strain (MAT**α**) as previously described20, selecting for MAT**a** and MAT**α** progeny separately. Using colony plating, sporulation efficiency was estimated at 24% - 1080 colonies grew in SC, 140 colonies grew in SC –His (MAT**a** haploid selection), and 120 colonies grew in SC –Leu (MAT**α** haploid selection). The two pools were then grown in YPD +HygroB to select for barcoded haploids. The SC –Leu pool was further grown in SC –Ura to select against barcoder strain parents that may have escaped diploid selection. Using a QPix colony picker, 5,461 MAT**a**  and 5,461 MAT**α** colonies were picked onto 384 well plates. In addition, 299 known positions in both the MAT**a** and MAT**α** arrayed collections consisted of known strains – either one of 40 ‘Gold Standard’ genotyped strains, RY0148, or RY0622 – to act as genotyping controls (Data S2).

**Pooled Strain Genotyping**

A previously-developed Row-Column-Plate (RCP)-PCR protocol29 was adapted in order to perform *en-masse* genotyping of the random knockout progeny using high throughput sequencing. This protocol first uniquely tags PCR products originating from the same well on a given plate, by the use of a 5’ tag encoding the well row (R) in forward primers, a 5’ tag encoding the well column (C) in the reverse primers29. Additionally, these primers contain a linker sequence (PS1 or PS2) which primes a second reaction encoding the plate of origin (Data S2).

For each well in the collection, lysates were made on a new set of plates. 4μl of overnight yeast culture was mixed with 8μl 0.2M sodium phosphate buffer (pH 7.4), 4μl DNA free dH2O, 0.05μl 5U/μl Zymoresearch zymolyase and incubated at 37°C for 35 minutes. 64μl DNA free dH2O was added to each well to stop the reaction.

Four ‘Row-Column’ PCR reactions were performed on the lysates with the following primer pairs: PS1+R+U1 and PS2+C+U2 to amplify DNA barcodes encoding the UP tags for each gene deletion; PS1+R+D1 and PS2+C+D2 to amplify the deletion-specific DN tags; PS1+R+US1 and PS2+C+US2 to amplify the strain-specific UP tag; PS1+R+DS1 and PS2+C+DS2 to amplify the strain-specific DN tag (Data S2). PCR reactions were performed with 2μl of lysed colonies using a Hydrocycler with the following program: 95°C for 5min; 23 cycles of 95°C for 60sec, 57°C for 35sec, 72°C for 45sec; 72°C for 2min; 4°C forever. Row-Column PCR products from each plate were pooled and size was verified on a 4% agarose gel. PCR products from each plate were pooled and 260μl was purified using a Qiagen Qiaquik Spin kit. DNA yield was quantified using a Nanoquant. From the resulting products from each plate, Illumina adapters containing plate tags were added using an additional PCR reaction as previously described29. A pair of PXX\_PE1.0 and PYY\_PE2.0 primers (Data S2) were added to 3-6μl pooled products (calibrated to ~150ng) from each plate to encode the plate of origin, and were amplified using the following PCR program: 98°C for 30sec; 15 cycles of 98°C for 10sec, 59°C for 15sec, 72°C for 40sec; 72°C for 2min; 4°C forever. All PCR reactions above were performed using High Fidelity Phusion Master Mix (NEB).

Expected product size from the plate tags was confirmed on 4% agarose gel. PCR products were purified using a Qiagen Qiaquik Spin kit. qPCR was performed on all plate tag PCR products using a light cycler and KAPA Illumina sequencing quantification kit. qPCR results were used to pool approximately equal amounts of all samples, and 100μl of this multiplexed sample were run on a 4% gel. Products of the desired size (260-290bp) were isolated from each lane, and purified using a Qiagen gel purify kit and another qPCR was run on the purified sample.

**Analysis of Pooled Strain Genotyping Data**

Pooled strain genotyping PCR products were sequenced using an Illumina HiSeq, and the reads were demultiplexed into individual samples corresponding to a plate and well of origin using a Perl script.

For each sample, a genotype calling pipeline determined the strain-specific tag sequences and genotype from the reads. The parameters of this pipeline were trained based on known reference strains. Cross-validated accuracy for each gene is reported in Fig. S2A.

UP or DN tag identity and a corresponding genotype was successfully determined for 7,195 samples. For 7,030 samples, the UP or DN tag was unique, and for 165 samples, both the UP and DN tag sequences were redundant with another sample where the called genotype was isogenic or highly similar (≤2 differences), indicating the presence of a single strain in multiple wells. When processing the sequencing data, a single strain was randomly chosen to represent each unique UP and DN tag sequence.

**Examining Putative Wild-Type Pool Strains**

For 73 MAT**a** and 131 MAT**α** strains, pooled sequencing analysis had called the genotype as wild-type. Many of these strains were isolated and tested for the presence of one or more gene knockout cassettes by growth in SC –Ura. Out of 96 MAT**α** strains, 74 exhibited no detectable growth in SC –Ura, and likely arose from remaining barcoder parents which had escaped a previous SC –Ura selection step. The genotypes for the these 74 strains were kept as is, while the other 23 strains, as well as 46 untested strains were discarded from the analysis. Out of 45 MAT**a** strains, all exhibited growth in SC-Ura. Individual genotyping was performed for these MAT**a** strains, and was successful for 40 of 45 strains, confirming the lack of true wild types. These strains had their stated genotype was corrected (Data S2). The 5 unsucessfully genotyped strains, as well as 28 additional strains were discarded from analysis. When calculating linkage and distribution of gene knockouts (Fig. S2), the wild-type MAT**α** strains were excluded from analysis as they were likely parental strains rather than progeny arising from mating.

**Estimating Genotyping Accuracy by Knockout Distribution**

To lend independent support to the genotyping accuracy determined by gold standard strains, an alternate method based on the distribution of knockouts in the population was used. Since *en masse* genotyping associates barcode sequences with ABC transporter knockouts, the absence of a given barcode implies either a wild-type genotype at that locus or a failure in amplification, sequencing, or calling. Conversely, cases where a wild-type is called as a mutant are expected to be comparably rare. Excess wild-type calls lead to a reduction in the average number of knockouts in the pool, and can be used to estimate genotyping accuracy. The average number of knockouts in the pool was 7.0, lower than the 8 expected with perfect genotyping. If there are no wild-type to mutant miscalls, this number is most likely with an asymmetric genotyping accuracy of 93.8%, compared to the 93.2% estimated by comparison to gold standards (Fig. S2C).

**Indiviual Liquid Growth Profiling**

To measure individual strain growth, the OD600 nm of a 0.0625 OD600 nm starting culture was measured in the appropriate medium every 15 mins using a GENios microplate reader (Tecan).

**Drug Testing for Growth Inhibition**

The effects of 16 different drugs on strain growth were tested to find a concentration which inhibits wild type growth by approximately 20% (Data S3). All drugs used were dissolved in 2% DMSO, which was used as a solvent control. Growth was determined by the Average\_G metric50, which represents the average generation time.

**Population Growth Profiling by High-Throughput Sequencing**

Progeny with at least one mapped strain-specific barcode (Data S2) were combined into two separate liquid YPD + glycerol pools separated by mating type, and kept at −80°C. Samples from the original YPD + glycerol pool were thawn and added to the appropriate drug or solvent containing medium at a final concentration of 0.0625 OD600 nm in 10ml. For the solvent control, a 0 generation sample was immediately harvested for sequencing. After growth to approximately 2 OD600 nm, a sample was taken from each drug for sequencing and cells were resuspended in fresh medium to a final concentration of 0.0625 OD600 nm. This process was repeated until 4 generations of samples were collected. Collected samples corresponded approximately to 5, 10, 15, and 20 generations of growth. Harvested samples were subject to genomic DNA extraction using a YeaStar™ Genomic DNA Kit, quantified using a Qubit® 2.0 fluorometer, and diluted to a final concentration of 20ng/μl. Approximately 350ng of isolated DNA was extracted from each sample and added to 20μL of 2x Platinum PCR SuperMix High Fidelity, 1μL of 10μM F primer, and 1μl of 10μM R primer. F and R primer pairs were PXX+US1/ PYY+US2 and PXX+DS1/PYY+DS2 for the strain-specific UP and DN tag, respectively. PXX and PYY correspond to sequences containing plate-specific Illumina sequencing adapters, as well as tags which were used to demultiplex the samples (See Data S2). PCR products amplified using the following PCR program: 98°C for 30sec; 24 cycles of 98°C for 10sec, 60°C for 10sec, 72°C for 1min; 72°C for 5min; 4°C forever.

PCR products were subject to gel electrophoresis, and ~210bp bands were isolated, subject to gel purification, and eluted in 60μl tris buffer. DNA yield was quantified in duplicate using a KAPA qPCR assay kit, at 1,000-fold, 10,000-fold, and 100,000-fold dilutions to find a concentration within standard curve range. Samples were pooled to yield approximately equal amounts of DNA, and subject to sequencing using an Illumina NextSeq 500 Mid Output kit.

**Sequence Data Processing**

Paired-end Illumina sequencing data were first de-multiplexed using a custom Python script which searches for an exact match to the tag regions of the PXX and PYY primers within each pair of reads. For each strain in each de-multiplexed sample (corresponding to a combination of mating type, timepoint, and drug), strain identification is attempted. To perform this identification, a search is performed for all barcodes matching the sample mating type. If an exact match is not found, up to two ungapped mismatches are permitted to assign a putative strain identity, which is then accepted if there are at least 2 additional mismatches separating this identity with the next closest match (e.g. if 2 mismatches are present with the closest match, then the next closest match must have 4 or more mismatches). This process was performed for both the forward and reverse reads (corresponding to the UP and DN tags) for each strain, and potential cases where the putative strain identity differed between tags were discarded.

All samples with less than 200,000 reads were discarded from the analysis. Additionally, if a sample was discarded for one mating type, the corresponding sample for the opposite mating type was also discarded (e.g. if ‘miconazole t=15 MAT**a**’ was discarded due to lack of coverage, ‘miconazole t=15 MAT**α**’ would also be discarded regardless of coverage).

**Defining a Resistance Metric**

From the sequence data processing, a count was assigned for each strain in a pool under drug sequenced at time (. If both an UP and DN tag for a given strain were successfully linked to a genotype, the UP and DN counts in a given sample were averaged to yield . Otherwise the available tag was used as . The counts in each sample were then converted to a frequency by division with the total count for all strains in that sample:

The frequency of each strain was then converted into a ‘virtual area under the growth curve’ () by first multiplying the frequency at each time point by the expected overall pool growth at that time (, since is defined by the number of generations), then taking the integral over all measured timepoints to (the total number of pool generations measured). Frequencies between measured timepoints were linearly interpolated.

The drug resistance score of a strain is defined as the ratio of its total growth in a drug compared to its growth in the solvent control , per pool generation:

**Finding Complex Genetic Interactions using a Linear Model**

The multiplicative model of genetic interactions39 was applied to the metric. In this model, the expected resistance of a double knockout strain in a given drug () is the product of the resistances of the corresponding single knockout strains:

To use this model in an additive rather than multiplicative form, the log of the resistance metric was taken , so that:

We defined a two-gene interaction term as the deviation of the observed double mutant log-fitness from this expectation, rather than the traditional linear difference from a multiplicative estimate.

When modelling an expected triple knockout fitness, all relevant two-gene interaction terms are added:

Similarly, a three gene interaction term is the deviation from this expectation:

This definition can be analogously extended for interactions of arbitrary complexity, and signficant terms denote interactions between the corresponding knockouts. The genotype of each strain is encoded using a binary variable, so that if and only if all the corresponding genes in the term are knocked out.

For each sample, a linear model was trained relating genotype to log resistance , including all terms up to a chosen level of complexity. Additionally, each trained model contained terms stating the plate of origin for each strain as a way to account for a potential batch effect.

To perform marginal association, we fitted an initial linear model with no terms, and performed stepwise feature elimination (eliminating the gene with the highest value at each step) until all included terms had a significance level of . Linear model term significance was tested using the Type III Sums of Squares ANOVA implementation given in the car package in R.

To train models containing terms of up to -way complexity, additional steps were performed. First, an initial set of a genes was chosen by testing whether a knockout of the 16 ABC transporters resulted in a significant population-level difference in log-resistance, first in the context of the overall population, then in the context of all additional gene knockouts (where is the desired level of gene-gene interaction complexity to be fit in the overall model). Significant differences were evaluated using a t-test, requiring a value of , where is the total number of genetic background contexts tested for that gene. A linear model including genes passing this test and all of their way interactions was then fit.

To speed up feature elimination, many terms were first heuristically eliminated using cross-validated LASSO regression implmented using the cv.glmnet function in R (using the default parameters of alpha = 1, nfolds = 10, type.measure = ‘deviance’). The λ which minimizes the mean cross validated error value was then chosen, and all terms with a nonzero coefficient at that λ value were selected for further elimination. These terms were subjected to stepwise elimination (again eliminating the gene with the highest value at each step), at a significance threshold of where is the number of terms in the initial linear model.

**Targeted Mating and Selection to Obtain 32 Knockouts**

The TWAS21230902 strain (*pdr10∆ pdr18∆ pdr5∆ snq2∆ ybt1∆ ycf1∆ yor1∆;* Data S2) was subject to individual strain genotyping, confirming the genotype generated using the RCP-PCR based method. This strain (MAT**α**) was mated with RY0146 (MAT**a**), and subject to sporulation and MAT**a** haploid selection20. Individuals from this cross were arrayed onto a 384 well plate, and individually genotyped at *PDR10* and *PDR18*. Strains with no deletions at these genes were further genotyped at *PDR5, SNQ2, YBT1, YCF1,* and *YOR1.* PCR reactions for individual genotyping of these progeny used the Qiagen Mix with the following program: 95°C for 5min; 34 cycles of 95°C for 30sec, 57°C for 30sec, 72°C for 30sec; 68°C for 10min; 4°C forever. After analysis of genotyping results, one strain of each genotype combination was chosen to create the 32-strain collection. These chosen 32 strains were again individually genotyped at these 5 loci for validation.

**Analysis of Liquid Growth Data**

**MYTH Testing of Protein-Protein Interactions**

PDR5, YOR1, and SNQ2 were cloned into the L2 AMBV MYTH bait vector to add a Cub-LexA-VP16 MYTH tag as previously described25. A previously-cloned artificial MYTH-tagged bait plasmid was retrieved, and acted as a negative interaction control. NubG-PDR5 (PDR5 prey) and NubI-PDR5 (PDR5 positive interaction control) strains were retrieved from a previously constructed genomic prey library25. Previously-constructed Ost1p-NubG (negative interaction control) and Ost1p-NubI (positive interaction control) strains were also retrieved. All prey-bait combinations were obtained using individual transformations and selected for growth in SD –Trp (SD –W)51. Colonies of transformed strains were grown in solid medium for 5 days in SD –W, SD –Trp–Ade–His (SD –WAH), SD –WAH +25μM fluconazole + 2% DMSO, SD –WAH +50μM fluconazole + 2% DMSO, and SD –WAH + 2% DMSO.

**PCA Testing of Protein-Protein Interactions**

*PDR5*, *YOR1*, and *SNQ2* MAT**a** (mDHFR-F[1,2]-NatMX fusions) and MAT**α** (mDHFR-F[3]-HphMX fusions) PCA strains were obtained from a previous genome-wide screen41. Additional strains acting required to recreated positive and negative controls were also obtained from this screen (Fig. S11). Strains were individually mated and diploids were selected on solid YPD supplemented with Hygromycin B and Nourseothricin (YPD +Hyg +Nat). Diploid strains were spotted on solid YPD +Hyg +Nat supplemented with either 2% DMSO, 2% DMSO + 200 μg/mL methotrexate, or 2% DMSO + 200 μg/mL methotrexate + 46.8μM fluconazole. Strains were grown for 72 hours at 30°C.

**Quantitative RT-PCR**

RNA extraction was performed using the QIAGEN RNeasy® kit. 1μg of isolate was treated with DNAse and analyzed usign an Agilent Bioanalyzer to quantify and verify purity. cDNA synthesis was perfomed using a combination of oligo-DT and random hexamer primers. qPCR on these samples was then performed using a SensiFAST™ Real-Time PCR Kit and Ct values were quantified using a CFX machine.

**Availability of Data and Materials**

**Competing Interests**

The authors declare that they have no competing interests.

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**Author Contributions**

N.Y, F.P.R & A.C conceived the experiments. N.Y, M.G, L.M, S.Z & T.F performed experiments. A.C and N.Y analyzed the data. A.C, F.P.R, & N.Y. wrote the paper.

**Additional Data Files**

**Additional Data S1.** List of primers used in this study. Includes the primers used to construct the barcoder strain, perform genotyping, RCP-PCR overhangs, and pool multiplexing primers.

**Additional Data S2.** Genotyping data in the engineered population. Includes a list of control strains used in high-throughput genotyping, initial genotyping results, re-genotyping of putative wild-type strains, and the final set of genotyping data used.

**Additional Data S3.** Drugs used in this study and their concentration in the pooled growth data.

**Additional Data S4.** List of primer pairs used to multiplex pooled growth sequencing data.

**Additional Data S5.** Growth and resistance metrics obtained for all strains in both the MAT**a** and MAT**α** pools.

**Additional Data S6.** Summary of linear modelling results obtained in this study.

**Additional Data S7.** Previously-known drug knockout associationswithin the 16 ABC transporters and 16 drugs studied.

**Additional Data S8.** Functional interpretations of genetic interactions present in the data.

**References**

1. Benfey, P. N. & Mitchell-Olds, T. From Genotype to Phenotype: Systems Biology Meets Natural Variation. *Science (80-. ).* **320,** (2008).

2. Hartwell, L. Robust Interactions. *Science (80-. ).* **303,** (2004).

3. Hartman, J. L., Garvik, B. & Hartwell, L. Principles for the Buffering of Genetic Variation. *Science (80-. ).* **291,** (2001).

4. Civelek, M. & Lusis, A. J. Systems genetics approaches to understand complex traits. *Nat. Rev. Genet.* **15,** 34–48 (2014).

5. Costanzo, M. *et al.* A global genetic interaction network maps a wiring diagram of cellular function. *Science (80-. ).* **353,** (2016).

6. Giaever, G. *et al.* Functional profiling of the Saccharomyces cerevisiae genome. *Nature* **418,** 387–391 (2002).

7. Costanzo, M. *et al.* The genetic landscape of a cell. *Science* **327,** 425–31 (2010).

8. Bloom, J. S., Ehrenreich, I. M., Loo, W. T., Lite, T.-L. V. & Kruglyak, L. Finding the sources of missing heritability in a yeast cross. *Nature* **494,** 234–7 (2013).

9. St Onge, R. P. *et al.* Systematic pathway analysis using high-resolution fitness profiling of combinatorial gene deletions. *Nat. Genet.* **39,** 199–206 (2007).

10. Tong, A. H. Y. *et al.* Global mapping of the yeast genetic interaction network. *Science* **303,** 808–13 (2004).

11. Braberg, H. *et al.* Quantitative analysis of triple-mutant genetic interactions. *Nat. Protoc.* **9,** 1867–81 (2014).

12. Taylor, M. B., Ehrenreich, I. M., Rothstein, R., Hu, T. & Mast, J. Genetic Interactions Involving Five or More Genes Contribute to a Complex Trait in Yeast. *PLoS Genet.* **10,** e1004324 (2014).

13. Beh, C. T., Cool, L., Phillips, J. & Rine, J. Overlapping functions of the yeast oxysterol-binding protein homologues. *Genetics* **157,** 1117–40 (2001).

14. Wieczorke, R. *et al.* Concurrent knock-out of at least 20 transporter genes is required to block uptake of hexoses in Saccharomyces cerevisiae. *FEBS Lett.* **464,** 123–8 (1999).

15. Palmer, A. C. *et al.* Delayed commitment to evolutionary fate in antibiotic resistance fitness landscapes. *Nat. Commun.* **6,** 7385 (2015).

16. Cancer Genome Atlas Research Network *et al.* Genomic and Epigenomic Landscapes of Adult De Novo Acute Myeloid Leukemia. *N. Engl. J. Med.* **368,** 2059–2074 (2013).

17. Heckl, D. *et al.* Generation of mouse models of myeloid malignancy with combinatorial genetic lesions using CRISPR-Cas9 genome editing. *Nat. Biotechnol.* **32,** 941–6 (2014).

18. Takahashi, K. & Yamanaka, S. Induction of Pluripotent Stem Cells from Mouse Embryonic and Adult Fibroblast Cultures by Defined Factors. *Cell* **126,** 663–676 (2006).

19. Weinreich, D. M., Lan, Y., Wylie, C. S. & Heckendorn, R. B. Should evolutionary geneticists worry about higher-order epistasis? *Curr. Opin. Genet. Dev.* **23,** 700–707 (2013).

20. Suzuki, Y. *et al.* Knocking out multigene redundancies via cycles of sexual assortment and fluorescence selection. *Nat. Methods* **8,** 159–64 (2011).

21. Wang, H. H. *et al.* Programming cells by multiplex genome engineering and accelerated evolution. *Nature* **460,** 894–8 (2009).

22. DiCarlo, J. E. *et al.* Yeast oligo-mediated genome engineering (YOGE). *ACS Synth. Biol.* **2,** 741–9 (2013).

23. Zeitoun, R. I. *et al.* Multiplexed tracking of combinatorial genomic mutations in engineered cell populations. *Nat. Biotechnol.* **33,** 631–637 (2015).

24. Zeitoun, R. I., Pines, G., Grau, W. C. & Gill, R. T. Quantitative Tracking of Combinatorially Engineered Populations with Multiplexed Binary Assemblies. *ACS Synth. Biol.* **6,** 619–627 (2017).

25. Brem, R. B. & Kruglyak, L. The landscape of genetic complexity across 5,700 gene expression traits in yeast. *Proc. Natl. Acad. Sci. U. S. A.* **102,** 1572–7 (2005).

26. Perlstein, E. O., Ruderfer, D. M., Roberts, D. C., Schreiber, S. L. & Kruglyak, L. Genetic basis of individual differences in the response to small-molecule drugs in yeast. *Nat. Genet.* **39,** 496–502 (2007).

27. Lee, A. Y. *et al.* Mapping the cellular response to small molecules using chemogenomic fitness signatures. *Science* **344,** 208–11 (2014).

28. Yan, Z. *et al.* Yeast Barcoders: a chemogenomic application of a universal donor-strain collection carrying bar-code identifiers. *Nat. Methods* **5,** 719–725 (2008).

29. Yachie, N. *et al.* Pooled-matrix protein interaction screens using Barcode Fusion Genetics. *Mol. Syst. Biol.* **12,** 863 (2016).

30. Smith, A. M. *et al.* Quantitative phenotyping via deep barcode sequencing. *Genome Res.* **19,** 1836–42 (2009).

31. Kolaczkowska, A., Kolaczkowski, M., Goffeau, A. & Moye-Rowley, W. S. Compensatory activation of the multidrug transporters Pdr5p, Snq2p, and Yor1p by Pdr1p in Saccharomyces cerevisiae. *FEBS Lett.* **582,** 977–83 (2008).

32. Snider, J. *et al.* Mapping the functional yeast ABC transporter interactome. *Nat. Chem. Biol.* **9,** 565–72 (2013).

33. Kovalchuk, A. & Driessen, A. J. M. Phylogenetic analysis of fungal ABC transporters. *BMC Genomics* **11,** 177 (2010).

34. Donner, M. & Keppler, D. Up-regulation of basolateral multidrug resistance protein 3 (Mrp3) in cholestatic rat liver. *Hepatology* **34,** 351–359 (2001).

35. König, J., Rost, D., Cui, Y. & Keppler, D. Characterization of the human multidrug resistance protein isoform MRP3 localized to the basolateral hepatocyte membrane. *Hepatology* **29,** 1156–1163 (1999).

36. Huls, M. *et al.* The breast cancer resistance protein transporter ABCG2 is expressed in the human kidney proximal tubule apical membrane. *Kidney Int.* **73,** 220–225 (2008).

37. Jungwirth, H. & Kuchler, K. Yeast ABC transporters – a tale of sex, stress, drugs and aging. *FEBS Lett.* **580,** 1131–8 (2006).

38. Dean, M., Rzhetsky, A. & Allikmets, R. The human ATP-binding cassette (ABC) transporter superfamily. *Genome Res.* **11,** 1156–66 (2001).

39. Mani, R., St Onge, R. P., Hartman, J. L., Giaever, G. & Roth, F. P. Defining genetic interaction. *Proc. Natl. Acad. Sci. U. S. A.* **105,** 3461–6 (2008).

40. Khakhina, S. *et al.* Control of Plasma Membrane Permeability by ABC Transporters. *Eukaryot. Cell* **14,** 442–453 (2015).

41. Tarassov, K. *et al.* An in vivo map of the yeast protein interactome. *Science* **320,** 1465–70 (2008).

42. Schlecht, U. *et al.* Multiplex assay for condition-dependent changes in protein-protein interactions. *Proc. Natl. Acad. Sci. U. S. A.* **109,** 9213–8 (2012).

43. Ferreira-Pereira, A. *et al.* Three-dimensional reconstruction of the Saccharomyces cerevisiae multidrug resistance protein Pdr5p. *J. Biol. Chem.* **278,** 11995–9 (2003).

44. C. elegans Deletion Mutant Consortium. Large-Scale Screening for Targeted Knockouts in the Caenorhabditis elegans Genome. *G3 Genes,Genomes,Genetics* **2,** 1415–1425 (2012).

45. Thompson, O. *et al.* The million mutation project: A new approach to genetics in Caenorhabditis elegans. *Genome Res.* **23,** 1749–1762 (2013).

46. Brockmann, M. *et al.* Genetic wiring maps of single-cell protein states reveal an off-switch for GPCR signalling. *Nature* **546,** 307–311 (2017).

47. Emanuel, G., Moffitt, J. R. & Zhuang, X. High-throughput, image-based screening of genetic variant libraries. *bioRxiv* (2017).

48. Gibson, D. G. *et al.* Enzymatic assembly of DNA molecules up to several hundred kilobases. *Nat. Methods* **6,** 343–5 (2009).

49. Gietz, R. D. & Schiestl, R. H. High-efficiency yeast transformation using the LiAc/SS carrier DNA/PEG method. *Nat. Protoc.* **2,** 31–34 (2007).

50. Proctor, M. *et al.* in 239–269 (Humana Press, 2011). doi:10.1007/978-1-61779-173-4\_15

51. Snider, J. *et al.* Detecting interactions with membrane proteins using a membrane two-hybrid assay in yeast. *Nat. Protoc.* **5,** 1281–1293 (2010).

**Figures**

**Figure 1.** Overview of the engineered population profiling process.

We created a barcoded wild-type pool (Fig. S1) to enable construction of an engineered population using any multi-mutant strain. In this study, this pool was mated with a 16 ABC transporter knockout strain (ABC-16). The genotype at 16 ABC transporters is indicated by the squares drawn in each cell (black = knockout, white = wild type). Diploids from this cross were subject to sporulation and barcoded haploids were then selected. Each haploid inherits either a wildtype or knockout allele at these 16 loci. Single colonies were picked and arrayed onto a series of 384-well plates. *En masse* genotyping was performed on this collection using an RCP-PCR29 strategy, which uses a combination of row and column tags to allow identification of PCR products arising from the same well in each plate. An additional PCR reaction adds a plate tag (not shown). High throughput sequencing of pooled RCP-PCR products allows large scale genotyping and identification of a strain-specific DNA barcode for many strains. Strains with a sucessfully determined barcode and genotype are transferred into two liquid pools based on mating type (MAT**a** or MAT**α**), and grown under each of 16 drugs. High throughput sequencing of strain-specific DNA barcodes at t = 0, 5, 10, 15, and 20 generations of growth in each drug reconstructs a drug by strain resistance profile, profiling the engineered population.

**Figure 2.** An exploration and assessment of multi-knockout fitness landscapes within a 6-gene group.

**A** Comparison of MAT**a** and MAT**α** group resistance profiles in camptothecin and tamoxifen. Individuals were grouped by their genotype at *pdr5∆, snq2∆, ybt1∆, bpt1∆, ycf1∆,* and *yor1∆*. The 6-locus genotype of each group is indicated by the legend. Individuals in each group vary at the remaining 10 loci. Each point represents the average log2(resistance) of the indicated group in the MAT**a** pools (x-axis) and MAT**α** pools (y-axis). Profiles for the remaining drugs are shown in Fig. S6.

**B** Distribution of MAT**a**–MAT**α** correlations of the grouped log2(resistance) profiles amongst all drugs tested.

**C** A radial landscape of benomyl resistance. The graph is centered by the 6-gene wildtype group, with outward extensions adding cumulative knockouts. Each section is coloured by the average resistance of the corresponding 6-gene group relative to the 6-gene wild type. Extensions to 1, 2, and 6 total knockouts are shown. Sections are coloured by the mean resistance of each group relative to the 6-gene wildtype. The colour scale extends equally in both directions by the largest observed difference in resistance between the 6-gene wildtype and any other group (blue for increased resistance, orange for decreased resistance). All paths where any additional knockouts do not result in significant changes (Bonferroni-adjusted *p* < 0.05, Mann-Whitney U test) are pruned.

**D** As in B, showing radial fitness landscapes for 10 additional drugs. The remaining 5 drugs are shown in Fig. S7.

**Figure 3.**  Exploration and formalization of surprising multi-gene knockout phenotypes.

**A** A linear landscape of resistance to mitoxantrone, benomyl, and fluconazole in the MAT**a** pools amongst 6-gene groups. The 6-gene genotype of each group is indicated by the legend. Groups are arranged on the x-axis by the number of knockouts (with jitter added to improve clarity), and the y-axis by average drug resistance. Groups separated by single knockouts are connected by lines. Solid lines indicate significant differences in resistance (Bonferroni-adjusted *p* < 0.05, Mann-Whitney U test), otherwise dashed lines are used. Linear landscapes for all pools are drawn in Fig. S8.

**B** Distribution of fluconazole resistance amongst all *ybt1∆,* *yor1∆*, *snq2∆, ycf1∆,* and *pdr5∆* knockout groups in the MAT**a** pool. Group genotype is indicated for each line using the same legend as in A). The resistance distribution as a kernel density function is drawn.The distribution of *pdr5∆* groups (magenta) are paired with their corresponding *PDR5+* equivalent (grey).

**C** A linear model was used to formally determine significant gene knockout and genetic interaction effects mediating resistance to the tested drugs (see Methods). Linear model terms which were significant (Bonferroni adjusted *p* < 0.05) in both MAT**a** and MAT**α** pools of their given drug are coloured according the legend on the left. Other terms are coloured in grey. ε terms represent n-way interactions (see Methods). Coefficents are sorted by term complexity. Term complexity is also indicated by the grey colour scale to the immediate left of the heatmap.

**Figure 4.** Interpreting a complex genetic landscape in fluconazole.

**A** A model of drug-dependent transporter-transporter relationships interpreted from the engineered population profiles. Antagonism indicates genetic evidence that the presence of one transporter negatively affects the ability of another to impart drug resistance. Parallelism indicates genetic evidence that multiple transporters are redundantly responsible for resistance to a drug. Data S8 lists the genetic evidence for each relationship.

**B** The TWAS21230902 MAT**α** strain (*pdr10∆ pdr18∆ pdr5∆ snq2∆ ybt1∆ ycf1∆ yor1∆;* Data S2) was mated with RY0146 (MAT**a**), and subject to sporulation and MAT**a** haploid selection. Individuals from this cross were genotyped, and individual strains containing each of 32 knockout combinations at *pdr5∆*, *snq2∆*, *ybt1∆*, *ycf1∆*, and *yor1∆* were identified and isolated.

**C** The resistance of 32 single strains to various concentrations of fluconazole was measured as growth compared to the solvent control (DMSO). Resistance was measured at the concentrations labelled on the x-axis, and linearly interpolated in between. Resistance of individual strains at concentrations lower than 15.6μM are shown in Fig. S10.

**D** Comparing the IC50 of fluconazole derived from single-strain growth experiments to the resistance expected by in the grouped pool data (MAT**a** and MAT**α** log2-resistance were averaged). Strain genotype is indicated by the legend.

**E** Measuring the mRNA expression of *PDR5* in wild-type (RY0566), *snq2∆yor1∆*, and *snq2∆yor1∆ybt1∆ycf1∆* strains. *PDR5* mRNA expression was measured using qRT-PCR and normalized relative to *UBC6*. Values represent the ratio of *PDR5* expression compared to the average in RY0566. Error bars indicate standard deviation. Three replicates were used in each experiment. P-values were calculated using a t-test.

**F** Comparing the modeled *PDR5* repression by *YOR1* and *SNQ2* with with protein-protein interactions found using MYTH and PCA. Interactions were measured in both this study (Fig. S11, S12) and previous studies32,41. Spotting assays show a re-testing of Pdr5-Snq2, Pdr5-Pdr5, and Pdr5-Yor1 interactions using MYTH. NubG-PDR5, NubI-PDR5, Ost1-NubG, and Ost1-NubI strains were each transformed with plasmids containing clones of *PDR5*, *YOR1*, *SNQ2*, or an artificial bait fused to Cub (YOR1-L2, PDR5-L2, SNQ2-L2, Artificial L2 bait). NubI fusions are expected to spontaneously reconstitute ubiquitin with Cub, while NubG fusions are expected to require a protein-protein interaction for reconstitution. Ost1 is a component of the oligosaccharyltransferase complex localized to the endoplasmic reticulum membrane and is not expected to interact with any baits tested. Colonies of transformed strains were spotted on SD –Trp (SD –W) and SD –Trp–Ade–His (SD –WAH). SD –WAH selects for reconstitution of ubiquitin. Spotting assays were also performed in the presence of DMSO and fluconazole (Fig. S12).

**Figure S1.** Creation of a parent barcoder pool.

**A** Engineering of a barcoder pool cassette. An HphMX4 cassette was amplified from pIS420, with overhangs adding the US2 and DS1 sites. A second PCR reaction was performed to add 25 random base pairs for use as UP and DN tags, as well as two constant US1 and DS2 regions. A third PCR reaction then adds LoxP/Lox2272 sites, and homology to the pSH47 SacI site.

**B** Transforming a pool of barcoder parents. RY0148 was modified to add a LoxP-URA3-Lox2272 site and was co-transformed with the barcoder pool cassette and SacI-digested pSH47 to enable reconstitution of a pSH47-based barcoder plasmid construct through in-yeast assembly. Transformants were selected by growth in YPG +Hyg for 3 days to allow for both selection of successful in-yeast assembly products, as well as induction of Cre to enable recombination and replacement of URA3 with the barcoder pool cassette. Loss of URA3 through Cre-enabled recombination is selected by subsequent growth in 5-FOA.

**Figure S2.** Analysis of pool genotyping quality.

**A** Expected genotyping accuracy at the 16 ABC transporters surveyed. Accuracy was estimated by evaluating the performance of the *en masse* genotyping protocol on a set of known reference strains (Methods, Data S2).

**B** Tests of gene linkage within the MAT**a** pools (upper triangle) and MAT**α** pools (lower triangle). The Pearson correlation coefficient of the corresponding genotype pairs are indicated on the right. Pairs without significant correlation (Bonferroni-corrected *p* value ≥ 0.05) are shaded in grey.

**C** Distribution of knockouts in the combined MAT**a** and MAT**α** pools. The observed number of strains with a given number of knockouts are indicated in grey. The expected number of strains with a given number of knockouts at 93.8% genotyping accuracy under a random assortment model are indicated in black.

**D** Distribution of DNA content in the MAT**a** and MAT**α** pools compared to a diploid and haploid reference. DNA content was measured using flow cytometry.

**Figure S3.** Correlation of fitness estimates from UP and DN tag counts.

**A** Correlation of resistance estimates in the MAT**a** pool under each drug obtained using UP tag counts (x-axis) compared to those obtained by DN tag counts (y-axis). Strains missing either a mapped UP or DN tag were excluded.

**B** As in A), using data from the MAT**α** pool.

**C** Distribution of UPtag-DNtag correlation in the pools. The minimum correlation in the MAT**a** and MAT**α** a pool for each drug is taken.

**D** Comparing UP and DN tag correlation with pool variability in resistance (as ). is taken as the minimum from from UP and DN tag estimates.

**Figure S4.** Reproducible marginal gene knockout resistance effects in the pool.

A linear model was used to formally determine significant gene knockout effects mediating resistance to the tested drugs. Linear model terms which were significant (Bonferroni adjusted *p* < 0.05) in both MAT**a** and MAT**α** pools for their given drug are coloured according the legend on the left. Other terms are coloured in grey.

**Figure S5.** Profiles of grouped genotype data.

Strains were grouped by knockout genotypes at *pdr5∆, snq2∆, ybt1∆, ycf1∆,* and *yor1∆*. Each section in the heatmap represents the average log2-resistance of strains with the genotype indicated on the y-axis, grown in the pool indicated on the x-axis. Pools are arranged by hierarchical clustering of the grouped resistance data using complete linkage.

**Figure S6.** Reproducibility of grouped genotype resistance.

Strains were grouped on knockout genotypes at *pdr5∆, snq2∆, ybt1∆, ycf1∆,* and *yor1∆*. Each point represents a group of strains containing the 6-locus genotype indicated by the legend. Strains in each group vary at the remaining 10 loci. Each point represents the median log2-resistance of each group in the MAT**a** (x-axis) and MAT**α** (y-axis) pools.

**Figure S7.** A radial fitness landscape in six additional drugs.

A radial fitness landscape in six drugs showing all multi-knockout paths. Each graph is centered by the 6-gene wildtype group, with outward extensions adding cumulative knockouts. Each section is coloured by the average resistance of its corresponding knockout group relative to the 6-gene wild type. Extensions to 1, 2, and 6 total knockouts are shown. Sections are coloured by the mean resistance of each group relative to the 6-gene wildtype. The colour scale extends equally in both directions by the largest observed difference in log2-resistance between the 6-gene wildtype and any other group (blue for increased resistance, orange for decreased resistance). All paths where any additional knockouts do not result in significant changes (Bonferroni-adjusted *p* < 0.05, Mann-Whitney U test) are pruned.

**Figure S8.** A linear landscape of resistance to 16 drugs.

**A** A linear landscape of resistance to all tested drugs in the MAT**a** pools amongst 6-gene groups. The 6-gene genotype of each group is indicated by the legend. Groups are arranged on the x-axis by the number of knockouts (with jitter added to improve clarity), and the y-axis by average drug resistance. Groups separated by single knockouts are connected by lines. Solid lines indicate significant differences in resistance (Bonferroni-adjusted *p* < 0.05, Mann-Whitney U test), otherwise dashed lines are used.

**B** As in A), with values shown for the MAT**α** pool.

**Figure S9.** Reproducibility of linear models.

For each drug, a linear model was trained to identify significant terms mediating drug resistance (see Methods). Linear model predictions of log2-resistance are indicated on the y-axis and values observed in the pool are indicated on the x-axis. Each plot is labelled by the pool mating type which was used to train the linear model on the y-axis and to assess its predictions on the x-axis.

**Figure S10.** Comparing drug resistance measured from single-strain experiments to the grouped pool data.

Resistance of individual strains containing each of 32 knockout combinations at *pdr5∆*, *snq2∆*, *ybt1∆*, *ycf1∆*, and *yor1∆* was measured and compared to the resistance to the pool data. Pool strains were grouped based on genotype at these 5 loci, median log2-resistance was determined for each group in MAT**a** and MAT**α** pools, and these values were averaged to obtain a single pool value. Strain genotype is indicated by the legend. Growth of individual strains was measured at 1.9, 3.9, 7.8, 15.6, 23.4, 31.2, 35, and 40μm of fluconazole.

**Figure S11.** Measuring all protein-protein interactions between Pdr5, Snq2, and Yor1 using mDHFR PCA.

*PDR5*, *YOR1*, and *SNQ2* MAT**a** (mDHFR-F[1,2]-NatMX fusions) and MAT**α** (mDHFR-F[3]-HphMX fusions) PCA strains were obtained from a previous genome-wide screen41. Strains were individually mated to obtain the indicated diploids. Diploid strains were spotted on YPD containing either DMSO, DMSO + methotrexate (MTX), or DMSO + MTX + 46.8μM fluconazole. MTX selects for successful reconstruction of mDHFR from the F[1,2] and F[3] fragments via a protein-protein interaction. Link-F[1,2]/ Link-F[3] is a diploid strain which tests against interaction of the universal linker regions when fused to the mDHFR fragments. Zip-F[1,2]/ Zip-F[3] is a diploid strain which tests for interaction between two leucine Zipper sequences fused to the mDHFR fragments. Strains were grown for 3 days at 30°C.

**Figure S12.** Measuring protein-protein interactions of Pdr5 with Snq2 and Yor1 using MYTH.

NubG-PDR5, NubI-PDR5, Ost1-NubG, and Ost1-NubI strains were retrieved from a previously constructed genomic prey library25 and were each transformed with plasmids containing clones of *PDR5*, *YOR1*, *SNQ2*, or an artificial bait fused to Cub (YOR1-L2, PDR5-L2, SNQ2-L2, Artificial L2 bait). NubI fusions are expected to spontaneously reconstitute ubiquitin with Cub, while NubG fusions are expected to require a protein-protein interaction for reconstitution. Ost1 is a component of the oligosaccharyltransferase complex localized to the endoplasmic reticulum membrane and is not expected to interact with any baits tested. Colonies of transformed strains were spotted on SD –Trp (SD –W), SD –Trp–Ade–His (SD –WAH), SD –WAH +25μM fluconazole + 2% DMSO, SD –WAH +50μM fluconazole + 2% DMSO, and SD –WAH + 2% DMSO. SD –WAH conditions select for reconstitution of ubiquitin.