

# Group1\_finalprojectRmd

## Set Working Directory

## Load Packages

```
library(tidyverse)
library(readxl)
library(broom)
library(cowplot)
library(phyloseq)
library(vegan)
set.seed(7)
```

#Import Data

```
library(readr)
biographical <- read_delim("~/Group1_Bio201_FinalProject/raw_data/biographical.txt",
  "\t", escape_double = FALSE, trim_ws = TRUE)
```

```
## Parsed with column specification:
## cols(
##   Participant_ID = col_character(),
##   Use_Data = col_character(),
##   Sex = col_character(),
##   Age = col_double(),
##   Race_ethnicity = col_character(),
##   Weight_kg = col_double(),
##   Height_meters = col_double(),
##   BMI = col_double()
## )
```

```
View(biographical)
```

```
library(readr)
phylum_avg_shared <- read_delim("~/Group1_Bio201_FinalProject/raw_data/phylum_avg_shared.txt",
  "\t", escape_double = FALSE, trim_ws = TRUE)
```

```
## Parsed with column specification:
## cols(
##   .default = col_double(),
##   Participant_ID = col_character(),
##   Semester = col_character(),
##   Study_week = col_character()
## )
```

```
## See spec(...) for full column specifications.
```

```
View(phylum_avg_shared)

library(readr)
all_data_wkly <- read_delim("~/Group1_Bio201_FinalProject/raw_data/Bigzipfile/all_data_wkly.txt",
  "\t", escape_double = FALSE, trim_ws = TRUE)

## Parsed with column specification:
## cols(
##   Participant_ID = col_character(),
##   Study_week = col_character(),
##   Semester = col_character(),
##   Supplement_consumed = col_character(),
##   Quantity_compliant = col_character(),
##   Frequency = col_character(),
##   pH_median = col_double(),
##   pH_mean = col_double(),
##   Bristol_median = col_logical(),
##   Bristol_mean = col_logical(),
##   Blood_glucose_median = col_double(),
##   Blood_glucose_mean = col_double(),
##   Acetate_median = col_double(),
##   Acetate_mean = col_double(),
##   Butyrate_median = col_double(),
##   Butyrate_mean = col_double(),
##   Propionate_median = col_double(),
##   Propionate_mean = col_double()
## )

## Warning: 755 parsing failures.
##   row      col      expected actual
## 1066 Bristol_median 1/0/T/F/TRUE/FALSE      4 '~/Group1_Bio201_FinalProject/raw_data/Bigzipfile/all_
## 1066 Bristol_mean   1/0/T/F/TRUE/FALSE      4 '~/Group1_Bio201_FinalProject/raw_data/Bigzipfile/all_
## 1067 Bristol_median 1/0/T/F/TRUE/FALSE      5 '~/Group1_Bio201_FinalProject/raw_data/Bigzipfile/all_
## 1067 Bristol_mean   1/0/T/F/TRUE/FALSE      5 '~/Group1_Bio201_FinalProject/raw_data/Bigzipfile/all_
## 1068 Bristol_median 1/0/T/F/TRUE/FALSE      5 '~/Group1_Bio201_FinalProject/raw_data/Bigzipfile/all_
## ....
## See problems(...) for more details.
```

```
View(all_data_wkly)
```

#Tidy Data ##Butyrate and Propionate df

```
c_all_data_wkly <- all_data_wkly %>%
  rename_all(tolower) %>%
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, butyrate_mean, propionate_mean)
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes") #curated all data

#wide dfs
but_wide <- all_data_wkly %>%
  rename_all(tolower) %>%
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, butyrate_mean, propionate_mean)
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes") %>% #keep only weeks 1 and 3
```

```

spread(study_week, butyrate_mean) %>%
mutate(delta_butyrate = week3 - week1) #calculate difference in butyrate

pro_wide <- all_data_wkly %>%
  rename_all(tolower) %>%
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, propionate_mean) %>%
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes") %>% #keep only weeks 1 and 3
  spread(study_week, propionate_mean) %>%
  mutate(delta_propionate = week3 - week1) #calculate difference in propionate

but_pro_wide <- inner_join(x = but_wide, y = pro_wide,
  by = c("participant_id",
        "semester", "supplement_consumed", "frequency")) %>%
  rename(but_wk1 = week1.x, but_wk3 = week3.x, pro_wk1 = week1.y, pro_wk3 = week3.y) %>%
  select(-starts_with("quantity_compliant")) #all measurements by participant ID

#Save Data Frames:
write_delim(c_all_data_wkly, path = "curated_data/c_all_data_wkly.txt",
  delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(but_wide, path = "curated_data/but_wide.txt",
  delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(pro_wide, path = "curated_data/pro_wide.txt",
  delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(but_pro_wide, path = "curated_data/but_pro_wide.txt",
  delim = "\t", col_names = TRUE, quote = FALSE)

```

##BMI df

```

BMI_data <- biographical %>%
  rename_all(tolower) %>%
  select(participant_id, sex, age, race_ethnicity, weight_kg, height_meters) %>%
  drop_na(weight_kg, height_meters) %>%
  mutate(BMI = weight_kg/(height_meters)^2) #calculate all BMIs; n = 252

underweight_BMI <- BMI_data %>%
  filter(BMI < 18.5) #subset by CDC category: underweight - n = 12

healthy_BMI <- BMI_data %>%
  filter(BMI >= 18.5, BMI < 25) #subset by CDC category: healthy - n = 186

overweight_BMI <- BMI_data %>%
  filter(BMI >= 25, BMI < 30) #subset by CDC category: overweight - n = 39

obese_BMI <- BMI_data %>%
  filter(BMI >= 30) #subset by CDC category: obese - n = 15

#total n = 252

#Save Data Frames
write_delim(BMI_data, path = "curated_data/BMI_data.txt",
  delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(underweight_BMI, path = "curated_data/underweight_BMI.txt",
  delim = "\t", col_names = TRUE, quote = FALSE)

```

```
write_delim(healthy_BMI, path = "curated_data/healthy_BMI.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(overweight_BMI, path = "curated_data/overweight_BMI.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(obese_BMI, path = "curated_data/obese_BMI.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
```

##Phylum df

```
#Creating Processed Phylum Data Frame:
phylum_processed_df <- phylum_avg_shared %>%
  rename_all(tolower) %>%
  filter(study_week == "week1" | study_week == "week3") %>%
  select("participant_id", "semester", "study_week", "firmicutes", "bacteroidetes <bacteroidetes>") %>%
  rename(., "bacteroidetes" = "bacteroidetes <bacteroidetes>") %>%
  mutate(., f_b_ratio = firmicutes/bacteroidetes)

#Checking Sample Sizes for Consistency by Week:
phylum_processed_df %>%
  filter(study_week == "week1") %>%
  summarise(sample_size = n()) # n = 508
```

```
## # A tibble: 1 x 1
##   sample_size
##   <int>
## 1       508
```

```
phylum_processed_df %>%
  filter(study_week == "week3") %>%
  summarise(sample_size = n()) # n = 480. 508-480 = 28 inconsistencies.
```

```
## # A tibble: 1 x 1
##   sample_size
##   <int>
## 1       480
```

```
#Split by Weeks and Recombine so only samples with both weeks present:
phylum_week1 <- phylum_processed_df %>%
  filter(study_week == "week1")

phylum_week3 <- phylum_processed_df %>%
  filter(study_week == "week3")
phylum_week3
```

```
## # A tibble: 480 x 6
##   participant_id semester study_week firmicutes bacteroidetes f_b_ratio
##   <chr>          <chr>    <chr>      <dbl>         <dbl>      <dbl>
## 1 U042          Fall2015 week3      13515         16756      0.807
## 2 U044          Fall2015 week3      15632         7587       2.06
## 3 U046          Fall2015 week3       7980.        17707      0.451
## 4 U048          Fall2015 week3      12427        22233      0.559
```

```
## 5 U050      Fall2015 week3      12079      9706      1.24
## 6 U052      Fall2015 week3      5324      16858     0.316
## 7 U054      Fall2015 week3     12668.     14542.     0.871
## 8 U057      Fall2015 week3      8265      16255     0.508
## 9 U059      Fall2015 week3      8742      14673     0.596
## 10 U062     Fall2015 week3      6117.     21792.     0.281
## # ... with 470 more rows
```

```
phylum_processed_df_both_weeks_present <- inner_join(phylum_week1, phylum_week3, by = (c("participant_id", "semester", "study_week")))

#Final Data Frames:
phylum_processed_df
```

```
## # A tibble: 988 x 6
##   participant_id semester study_week firmicutes bacteroidetes f_b_ratio
##   <chr>          <chr>    <chr>      <dbl>      <dbl>      <dbl>
## 1 U042          Fall2015 week1      14119      18149     0.778
## 2 U042          Fall2015 week3      13515      16756     0.807
## 3 U044          Fall2015 week3      15632       7587     2.06
## 4 U046          Fall2015 week3       7980.     17707     0.451
## 5 U048          Fall2015 week3      12427     22233     0.559
## 6 U050          Fall2015 week3      12079      9706      1.24
## 7 U051          Fall2015 week1     19352      4827      4.01
## 8 U052          Fall2015 week1      4944     13166     0.376
## 9 U052          Fall2015 week3      5324     16858     0.316
## 10 U053         Fall2015 week1      7061     22655     0.312
## # ... with 978 more rows
```

```
phylum_processed_df_both_weeks_present
```

```
## # A tibble: 451 x 11
##   participant_id semester.x study_week.x firmicutes.x bacteroidetes.x
##   <chr>          <chr>    <chr>      <dbl>      <dbl>
## 1 U042          Fall2015 week1      14119      18149
## 2 U052          Fall2015 week1      4944     13166
## 3 U054          Fall2015 week1     11479     20357
## 4 U062          Fall2015 week1      8366     25134.
## 5 U063          Fall2015 week1     16913.     10266
## 6 U064          Fall2015 week1     10583     10990
## 7 U066          Fall2015 week1      9209     14273
## 8 U067          Fall2015 week1     19163.     16210.
## 9 U068          Fall2015 week1      6034     20364
## 10 U070         Fall2015 week1     11963     13446
## # ... with 441 more rows, and 6 more variables: f_b_ratio.x <dbl>,
## #   semester.y <chr>, study_week.y <chr>, firmicutes.y <dbl>,
## #   bacteroidetes.y <dbl>, f_b_ratio.y <dbl>
```

```
phylum_processed_df_both_weeks_present_final <- phylum_processed_df_both_weeks_present %>%
  rename(firmicutes_wk1 = firmicutes.x, bacteroidetes_wk1 = bacteroidetes.x, fb_ratio_wk1 =
    f_b_ratio.x, firmicutes_wk3 = firmicutes.y, bacteroidetes_wk3 = bacteroidetes.y, fb_ratio_wk3 =
    f_b_ratio.y, semester = semester.x) %>%
  select(-starts_with("study_week"), -starts_with("semester.y"))
```

```
View(phylum_processed_df_both_weeks_present_final)

#Save data frames:
write_delim(phylum_processed_df, path = "curated_data/phylum_processed_df.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(phylum_processed_df_both_weeks_present, path = "curated_data/phylum_processed_df_both_weeks_present.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(phylum_processed_df_both_weeks_present_final, path = "curated_data/phylum_processed_df_both_weeks_present_final.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
```

##Joined BMI and scfas

```
BMI_but_pro <- inner_join(x = BMI_data, y = but_pro_wide,
                        by = c("participant_id")) #n = 178

underweight_but_pro <- inner_join(x= underweight_BMI, y = but_pro_wide,
                                by = c("participant_id")) #n = 10

healthy_but_pro <- inner_join(x= healthy_BMI, y = but_pro_wide,
                             by = c("participant_id")) #n = 133

overweight_but_pro <- inner_join(x= overweight_BMI, y = but_pro_wide,
                                by = c("participant_id")) #n = 25

obese_but_pro <- inner_join(x= obese_BMI, y = but_pro_wide,
                           by = c("participant_id")) #n = 10

#total n = 178

#Save Data Frames
write_delim(BMI_but_pro, path = "curated_data/BMI_but_pro.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(underweight_but_pro, path = "curated_data/underweight_but_pro.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(healthy_but_pro, path = "curated_data/healthy_but_pro.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(overweight_but_pro, path = "curated_data/overweight_but_pro.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(obese_but_pro, path = "curated_data/obese_but_pro.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
```

##Joined BMI, scfas, and phylums

```
BMI_phylum_scfa <- inner_join(x = BMI_but_pro, y = phylum_processed_df_both_weeks_present_final,
                              by = c("participant_id")) #n = 139

underweight_phylum_scfa <- inner_join(x = underweight_but_pro, y = phylum_processed_df_both_weeks_present_final,
                                       by = c("participant_id")) #n = 9

healthy_phylum_scfa <- inner_join(x = healthy_but_pro, y = phylum_processed_df_both_weeks_present_final,
                                   by = c("participant_id")) #n = 105

overweight_phylum_scfa <- inner_join(x = overweight_but_pro, y = phylum_processed_df_both_weeks_present_final,
                                       by = c("participant_id")) #n = 20
```

```
obese_phylum_scfa <- inner_join(x = obese_but_pro, y = phylum_processed_df_both_weeks_present_final,
                                by = c("participant_id")) #n = 5
```

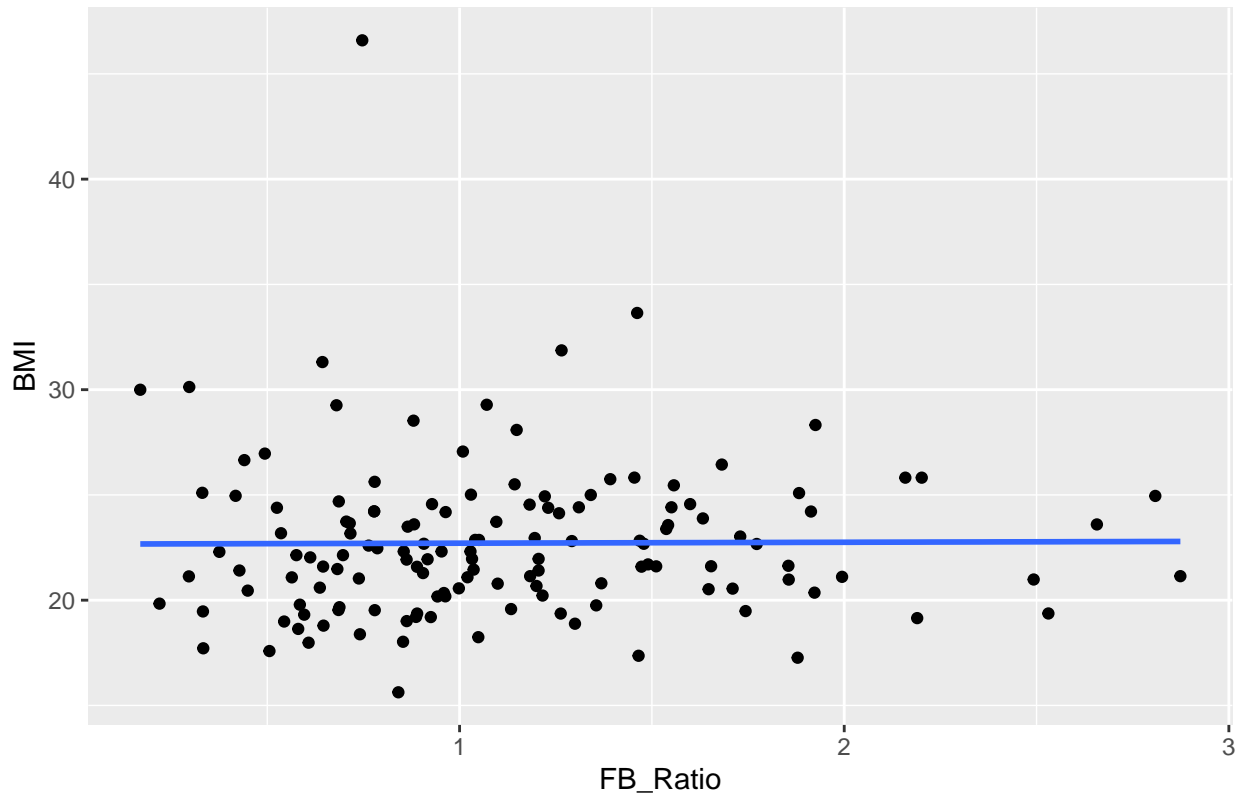
```
#Save Data Frames
```

```
write_delim(BMI_phylum_scfa, path = "curated_data/BMI_phylum_scfa.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(underweight_phylum_scfa, path = "curated_data/underweight_phylum_scfa.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(healthy_phylum_scfa, path = "curated_data/healthy_phylum_scfa.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(overweight_phylum_scfa, path = "curated_data/overweight_phylum_scfa.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
write_delim(obese_phylum_scfa, path = "curated_data/obese_phylum_scfa.txt",
            delim = "\t", col_names = TRUE, quote = FALSE)
```

```
#Analysis ##Broad (All BMIs) ###F:B and BMI
```

```
lm_Broad_BMI_plot_wk1<-BMI_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = BMI)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
              se = FALSE) + #hides confidence interval around line
  xlab("FB_Ratio") +
  ylab("BMI") + labs(title = "Broad BMI and F:B Ratio Correlation wk1")
print(lm_Broad_BMI_plot_wk1)
```

## Broad BMI and F:B Ratio Correlation wk1



```
lm_Broad_BMI_wk1<-BMI_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ BMI, data = .) #test relationship
summary(lm_Broad_BMI_wk1)
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ BMI, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.95021 -0.40410 -0.09109  0.34665  1.76222
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.0918175  0.2875401   3.797 0.000219 ***
## BMI           0.0009424  0.0124923   0.075 0.939977
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5505 on 137 degrees of freedom
## Multiple R-squared:  4.154e-05, Adjusted R-squared:  -0.007257
## F-statistic: 0.005691 on 1 and 137 DF, p-value: 0.94
```

```
lm_Broad_BMI_plot_wk3<-BMI_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
```

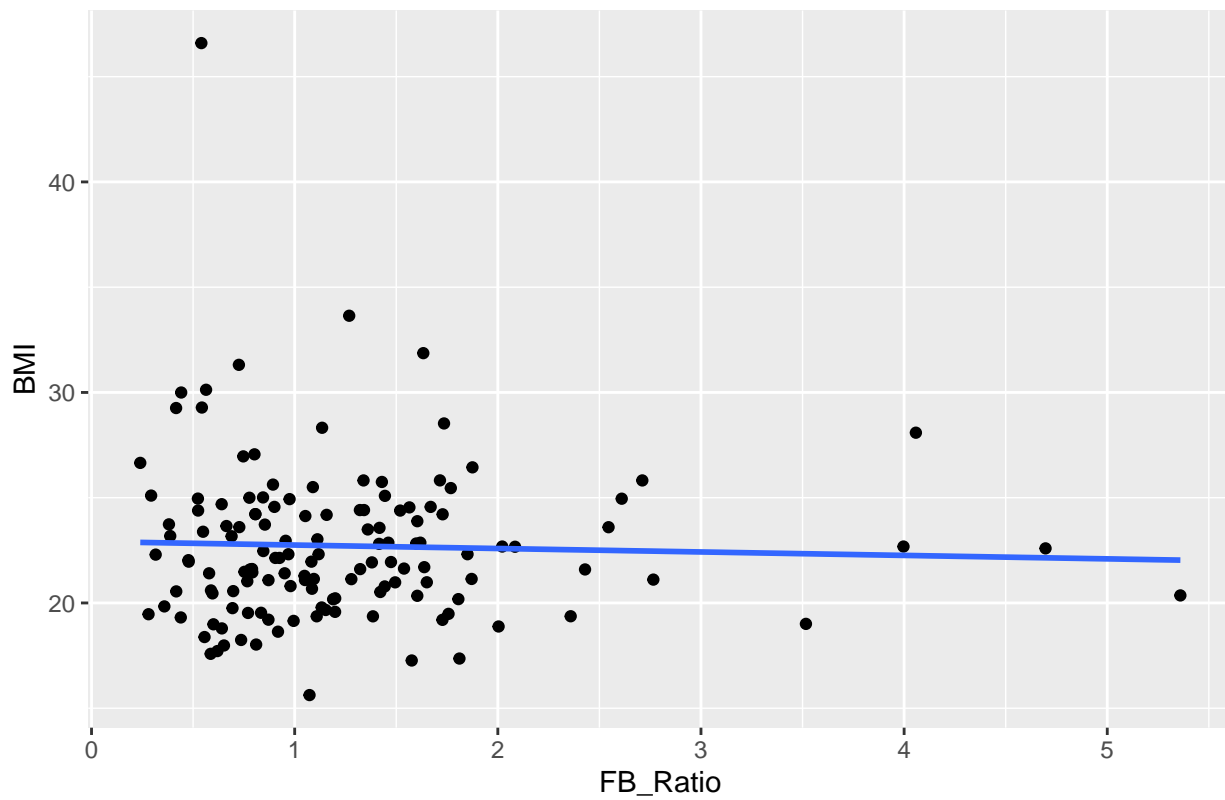


```

    y = BMI)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
    se = FALSE) + #hides confidence interval around line
  xlab("FB_Ratio") +
  ylab("BMI") + labs(title = "Broad BMI and F:B Ratio Correlation wk3")
print(lm_Broad_BMI_plot_wk3)

```

Broad BMI and F:B Ratio Correlation wk3



```

lm_Broad_wk3_BMI<-BMI_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ BMI, data = .) #test relationship
summary(lm_Broad_wk3_BMI)

```

```

##
## Call:
## lm(formula = fb_ratio_wk3 ~ BMI, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.9814 -0.4982 -0.1734  0.2963  4.1048
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.412211   0.426001   3.315  0.00117 **
## BMI         -0.007715   0.018508  -0.417  0.67745
## ---

```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.8157 on 137 degrees of freedom
## Multiple R-squared:  0.001267,    Adjusted R-squared:  -0.006023
## F-statistic: 0.1737 on 1 and 137 DF,  p-value: 0.6775
```

```
save_plot(filename = "figures/lm_Broad_BMI_plot_wk1.pdf",
          plot = lm_Broad_BMI_plot_wk1,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)

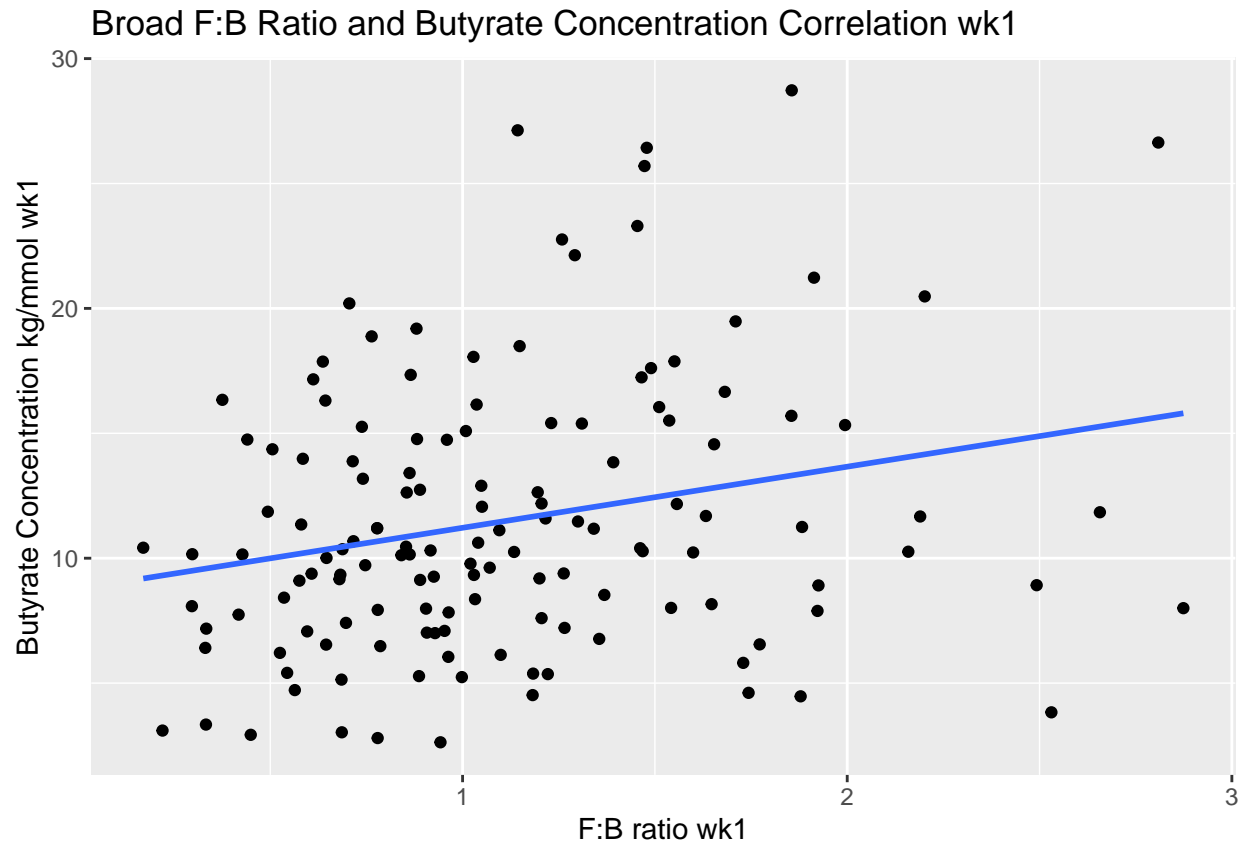
save_plot(filename = "figures/lm_Broad_BMI_plot_wk3.pdf",
          plot = lm_Broad_BMI_plot_wk3,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the BMI of all the participants is .94, which is not statistically significant. The  $R^2$  value was also insignificant being very small at  $R^2 = -.0073$ . There is no significant correlation between the week 1 firmicutes - bacteroidetes ratio and BMI.

Similarly, the week 3 F - B ratio and BMI also had an insignificant p-value and  $R^2$  value at  $p = .67$  and  $R^2 = -.006$  respectively. This also represents no significant correlation.

###F:B and Butyrate

```
lm_wk1_but_Broad_plot<-BMI_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = but_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("F:B ratio wk1") +
  ylab("Butyrate Concentration kg/mmol wk1") + labs(title = "Broad F:B Ratio and Butyrate Concentration")
print(lm_wk1_but_Broad_plot)
```



```
lm_wk1_but_Broad<-BMI_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ but_wk1, data = .) #test relationship
summary(lm_wk1_but_Broad)
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ but_wk1, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.91708 -0.38098 -0.07662  0.25754  1.84617
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.832191   0.105412   7.895 8.39e-13 ***
## but_wk1      0.024450   0.008282   2.952 0.00371 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5338 on 137 degrees of freedom
## Multiple R-squared:  0.05981,    Adjusted R-squared:  0.05295
## F-statistic: 8.716 on 1 and 137 DF,  p-value: 0.003713
```

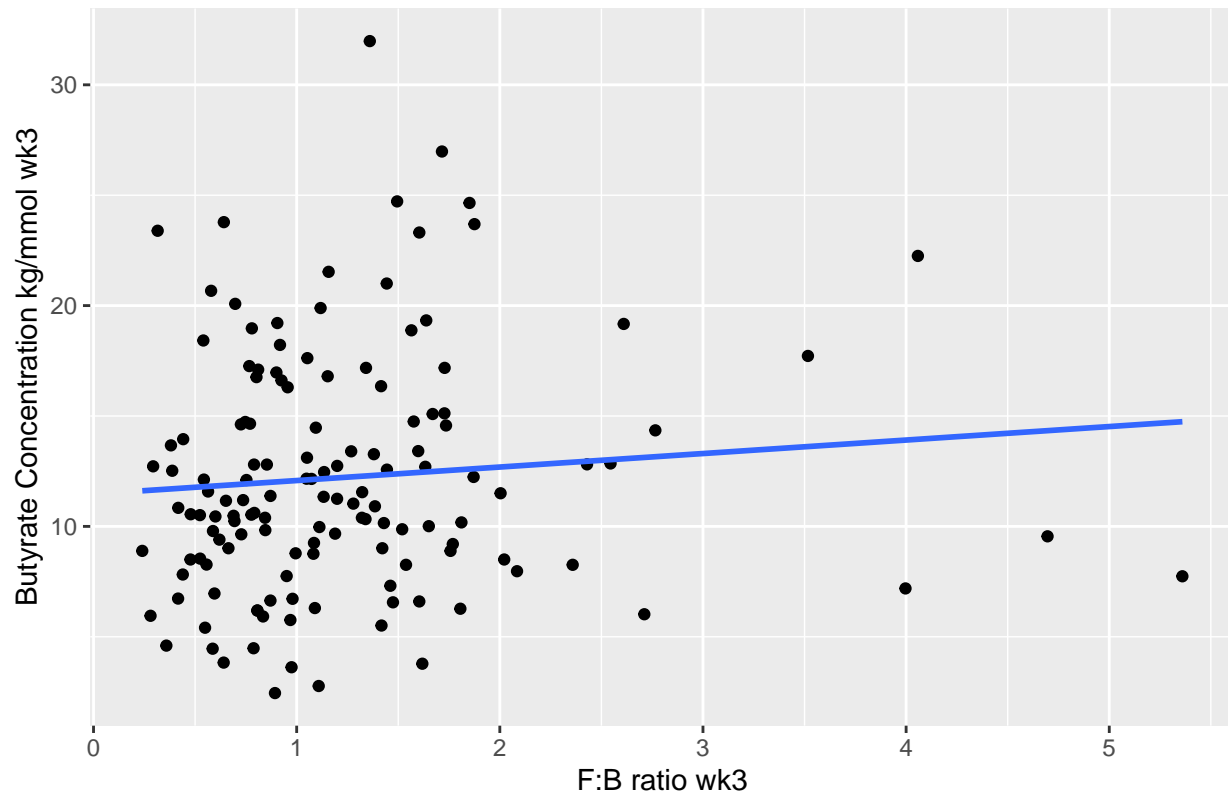
```
lm_wk3_but_Broad_plot<-BMI_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
```

```

    y = but_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
    se = FALSE) + #hides confidence interval around line
  xlab("F:B ratio wk3") +
  ylab("Butyrate Concentration kg/mmol wk3") + labs(title = "Broad F:B Ratio and Butyrate Concentration")
print(lm_wk3_but_Broad_plot)

```

Broad F:B Ratio and Butyrate Concentration Correlation wk3



```

lm_wk3_but_Broad<-BMI_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ but_wk3, data = .) #test relationship
summary(lm_wk3_but_Broad)

```

```

##
## Call:
## lm(formula = fb_ratio_wk3 ~ but_wk3, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -1.0730 -0.5200 -0.1801  0.2977  4.1839
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.07089    0.16967   6.311 3.58e-09 ***
## but_wk3      0.01359    0.01269   1.071  0.286
## ---

```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.8128 on 137 degrees of freedom
## Multiple R-squared:  0.008309,    Adjusted R-squared:  0.00107
## F-statistic: 1.148 on 1 and 137 DF,  p-value: 0.2859
```

```
save_plot(filename = "figures/lm_wk1_but_Broad_plot.pdf",
          plot = lm_wk1_but_Broad_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.2)

save_plot(filename = "figures/lm_wk3_but_Broad_plot.pdf",
          plot = lm_wk3_but_Broad_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.2)
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 Butyrate concentration of all the participants is .0037, which is statistically significant. However, the  $R^2$  value is insignificant being very small at  $R^2=0.0053$ , which represents too much variation in the data. There is no significant correlation between the week 1 firmicutes - bacteroidetes ratio and Butyrate Concentration.

The week 3 F - B ratio and week 3 Butyrate concentration of all the participants has a insignificant p-value and  $R^2$  value at  $p=.29$  and  $R^2 = 0.0011$  respectively. This also represents no significant correlation.

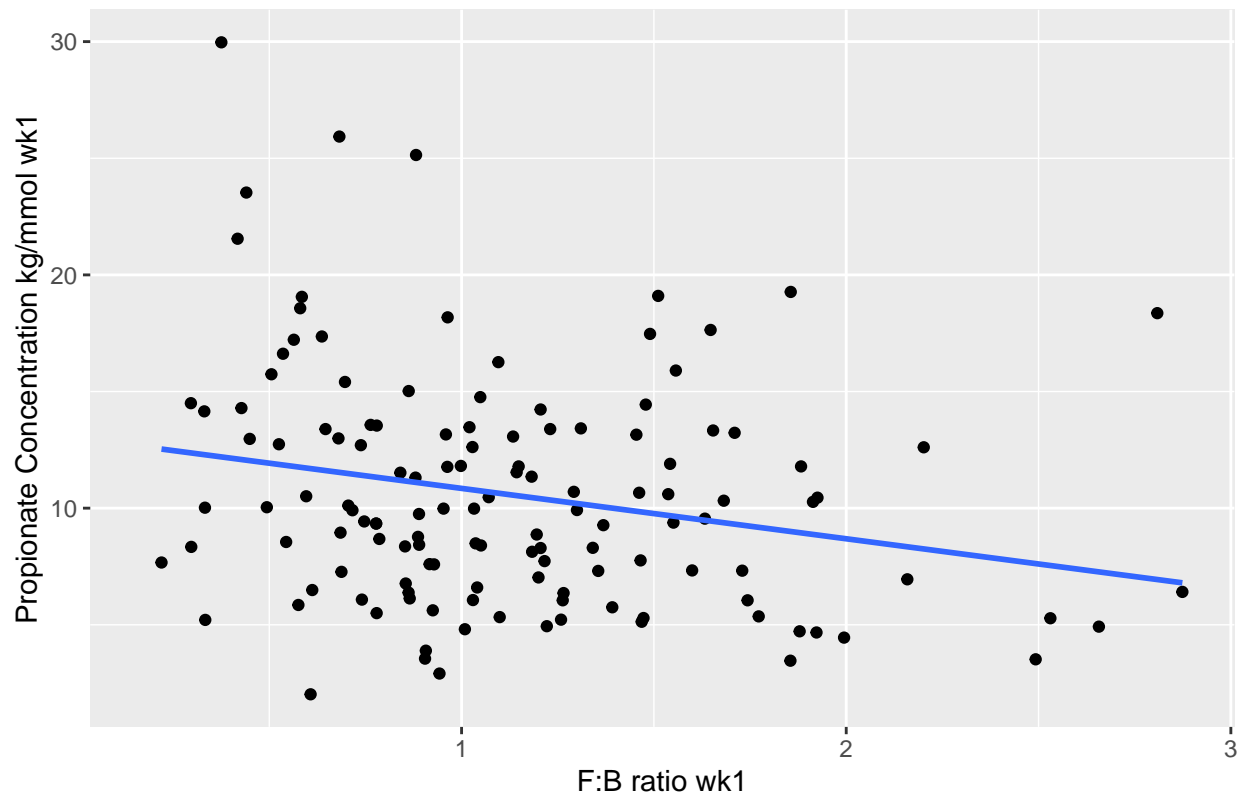
###F:B and Propionate

```
lm_wk1_pro_Broad_plot<-BMI_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = pro_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("F:B ratio wk1") +
  ylab("Propionate Concentration kg/mmol wk1") + labs(title = "Broad F:B Ratio and Propionate Concentration")
print(lm_wk1_pro_Broad_plot)
```

```
## Warning: Removed 6 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 6 rows containing missing values (geom_point).
```

## Broad F:B Ratio and Propionate Concentration Correlation wk1



```
lm_wk1_pro_Broad<-BMI_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ pro_wk1, data = .) #test relationship
summary(lm_wk1_pro_Broad)
```

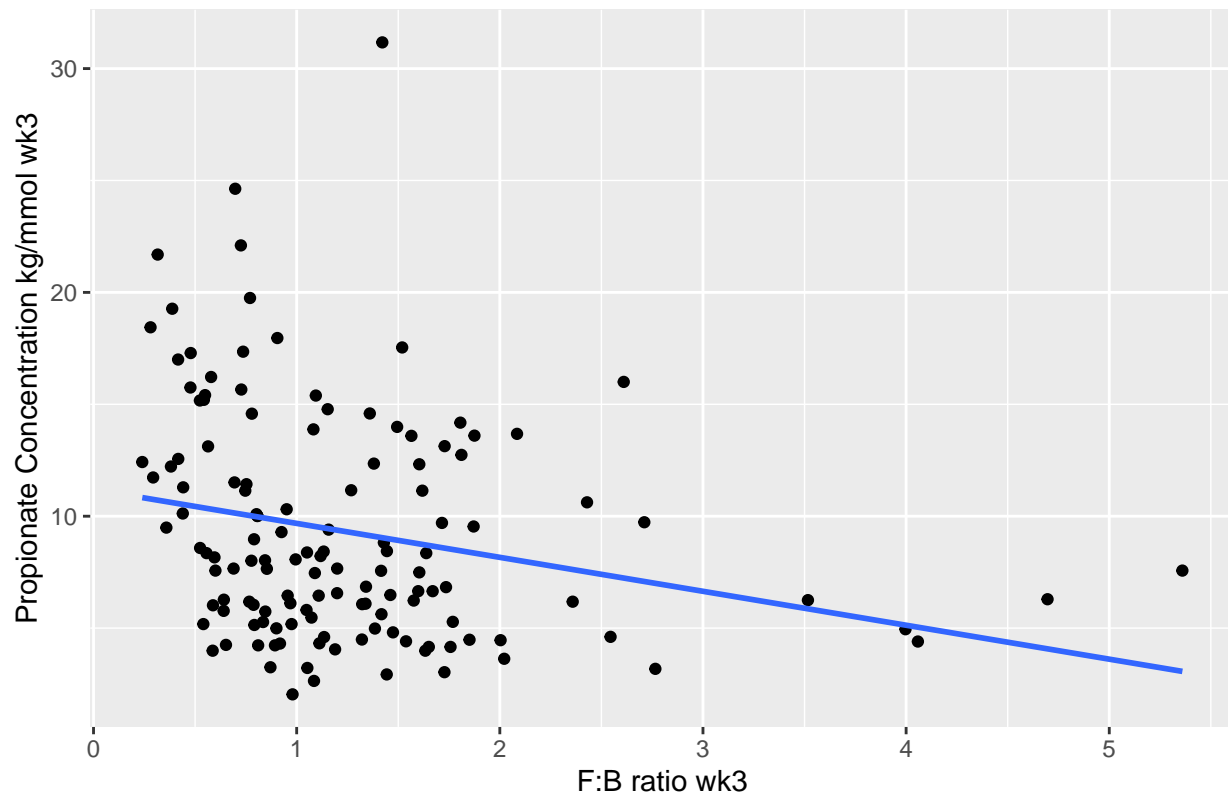
```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ pro_wk1, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.97952 -0.37792 -0.09592  0.26827  1.88128
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.394523   0.107700   12.95  < 2e-16 ***
## pro_wk1      -0.025437   0.009216   -2.76  0.00661 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5282 on 131 degrees of freedom
## (6 observations deleted due to missingness)
## Multiple R-squared:  0.05496,    Adjusted R-squared:  0.04774
## F-statistic: 7.618 on 1 and 131 DF,  p-value: 0.006607
```

```
lm_wk3_pro_Broad_plot<-BMI_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = pro_wk3))+
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("F:B ratio wk3") +
  ylab("Propionate Concentration kg/mmol wk3") + labs(title = "Broad F:B Ratio and Propionate Concentration wk3")
print(lm_wk3_pro_Broad_plot)
```

```
## Warning: Removed 4 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 4 rows containing missing values (geom_point).
```

### Broad F:B Ratio and Propionate Concentration Correlation wk3



```
lm_wk3_pro_Broad<-BMI_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ pro_wk3, data = .) #test relationship
summary(lm_wk3_pro_Broad)
```

```
##
## Call:
## lm(formula = fb_ratio_wk3 ~ pro_wk3, data = .)
##
## Residuals:
```

##	Min	1Q	Median	3Q	Max
----	-----	----	--------	----	-----

```
## -0.8853 -0.5050 -0.2312  0.2681  4.0435
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.61489    0.14364  11.243 < 2e-16 ***
## pro_wk3      -0.03942    0.01356  -2.908  0.00426 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.7994 on 133 degrees of freedom
## (4 observations deleted due to missingness)
## Multiple R-squared:  0.05978,    Adjusted R-squared:  0.05271
## F-statistic: 8.457 on 1 and 133 DF,  p-value: 0.004263
```

```
save_plot(filename = "figures/lm_wk1_pro_Broad_plot.pdf",
          plot = lm_wk1_pro_Broad_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)
```

```
## Warning: Removed 6 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 6 rows containing missing values (geom_point).
```

```
save_plot(filename = "figures/lm_wk3_pro_Broad_plot.pdf",
          plot = lm_wk3_pro_Broad_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)
```

```
## Warning: Removed 4 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 4 rows containing missing values (geom_point).
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 Propionate concentration of all the participants is .0066, which is statistically significant. However, the  $R^2$  value is insignificant being very small at  $R^2=0.0048$ , which represents too much variation in the data. There is no significant correlation between the week 1 firmicutes - bacteroidetes ratio and Propionate concentration.

Similarly, The week 3 F - B ratio and week 3 Propionate concentration of all the participants has a significant p-value and insignificant  $R^2$  value at  $p=.0043$  and  $R^2 = .053$  respectively. This also represents no significant correlation.

##Underweight ###F:B and BMI

```
lm_under_BMI_wk1<-underweight_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ BMI, data = .) #test relationship
summary(lm_under_BMI_wk1)
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ BMI, data = .)
##
## Residuals:
```

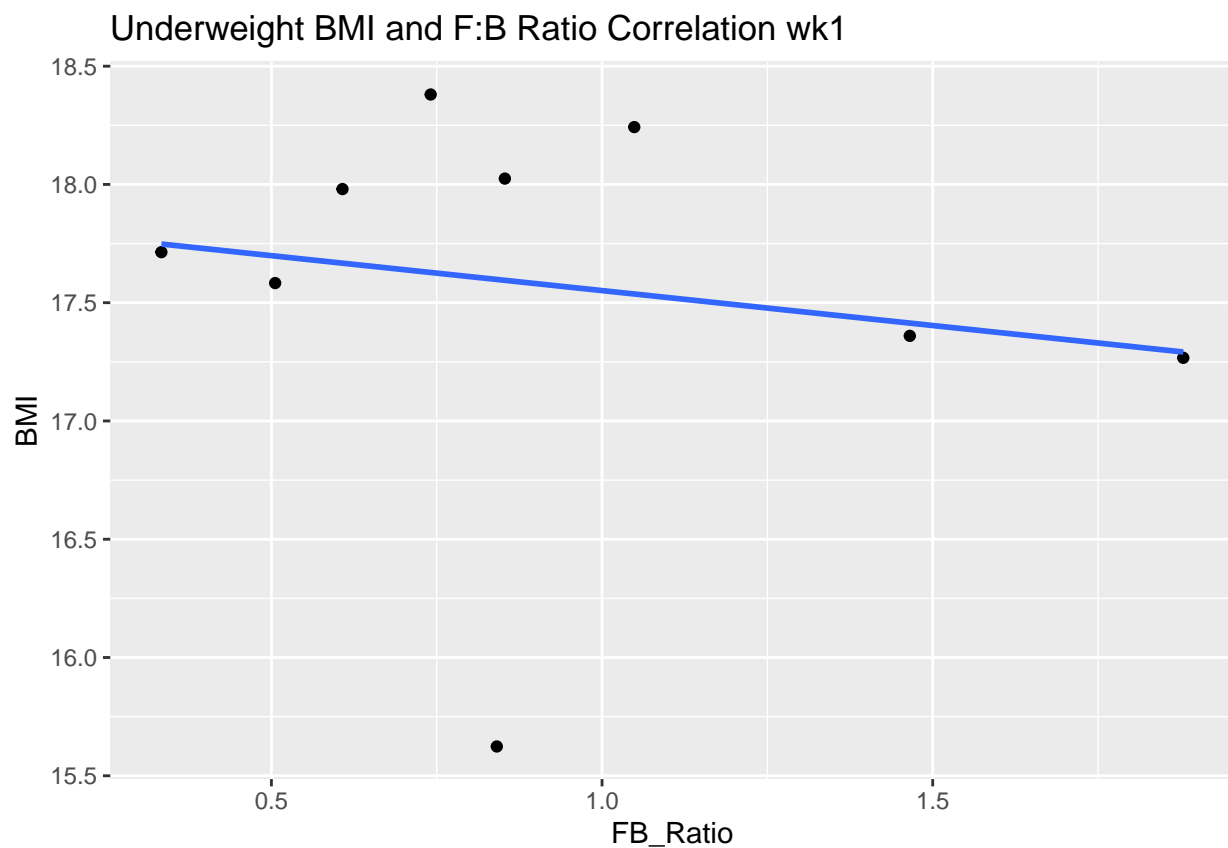


```
##      Min      1Q   Median      3Q      Max
## -0.57151 -0.27914 -0.09574  0.19793  0.92787
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)   2.7263     3.8631   0.706   0.503
## BMI          -0.1028     0.2196  -0.468   0.654
##
## Residual standard error: 0.5116 on 7 degrees of freedom
## Multiple R-squared:  0.03036,    Adjusted R-squared:  -0.1082
## F-statistic: 0.2192 on 1 and 7 DF,  p-value: 0.6539
```

```
#p-value = 0.6539
```

```
#R-squared = -0.1082
```

```
lm_under_BMI_plot_wk1<-underweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = BMI)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("FB_Ratio") +
  ylab("BMI") + labs(title = "Underweight BMI and F:B Ratio Correlation wk1")
print(lm_under_BMI_plot_wk1)
```

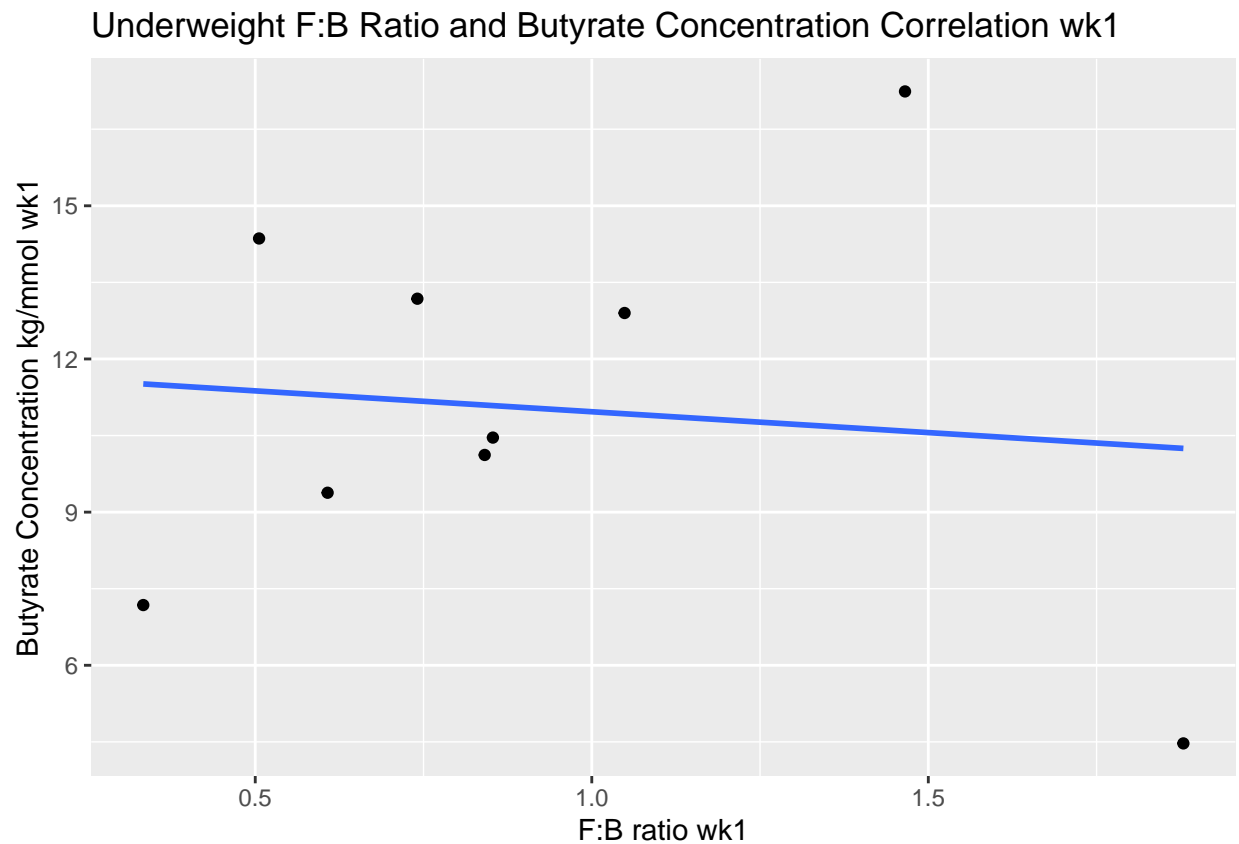


```
save_plot(filename = "figures/lm_under_BMI_plot_wk1.pdf",
          plot = lm_under_BMI_plot_wk1,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the BMI of “underweight” participants was 0.6539, which is not statistically significant. Furthermore, the R value was extremely low (-0.1082), and therefore insignificant. There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and BMI among participants whose BMI falls within the CDC’s “underweight” category.

###F:B and Butyrate

```
lm_wk1_but_under_plot<-underweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
            y = but_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
            se = FALSE) + #hides confidence interval around line
  xlab("F:B ratio wk1") +
  ylab("Butyrate Concentration kg/mmol wk1") + labs(title = "Underweight F:B Ratio and Butyrate Concentration wk1")
print(lm_wk1_but_under_plot)
```

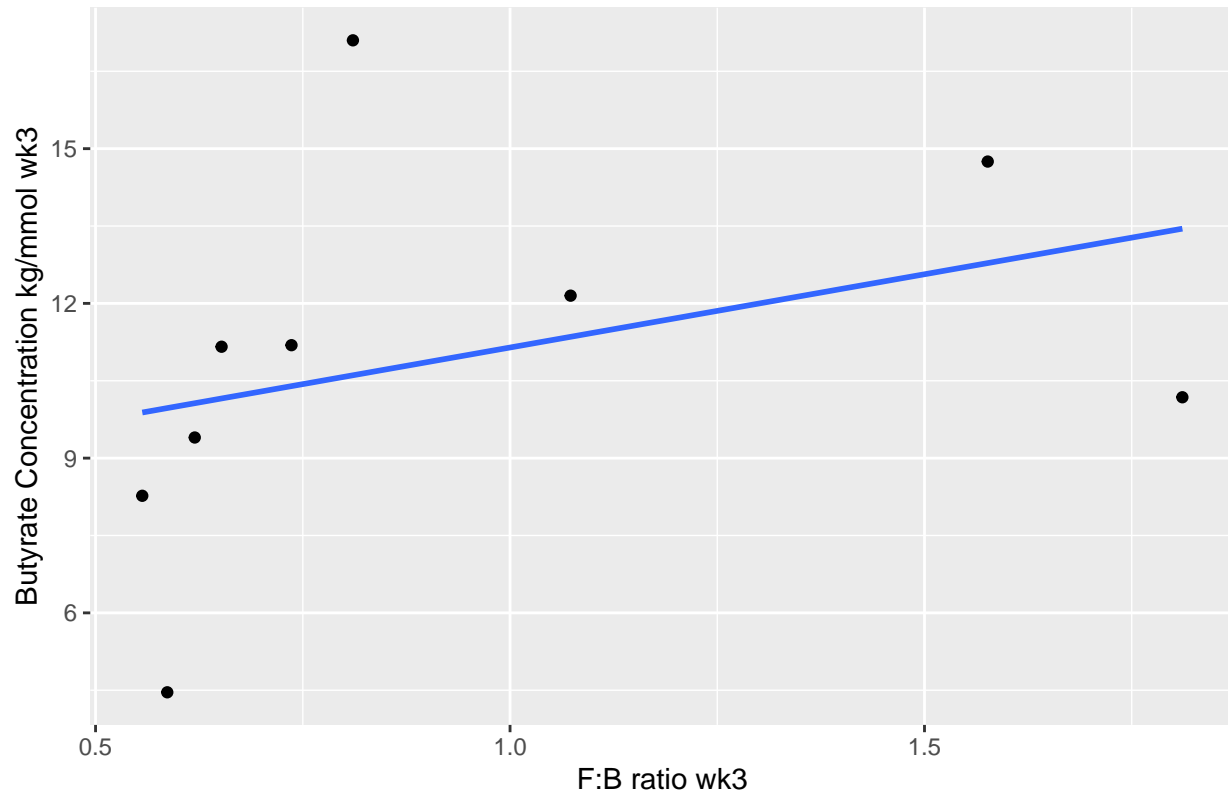


```
lm_wk1_but_under<-underweight_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ but_wk1, data = .) #test relationship
summary(lm_wk1_but_under)
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ but_wk1, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.63556 -0.33327 -0.09032  0.15344  0.87472
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.06200    0.54926   1.933  0.0944 .
## but_wk1      -0.01292    0.04728  -0.273  0.7924
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5168 on 7 degrees of freedom
## Multiple R-squared:  0.01056,    Adjusted R-squared:  -0.1308
## F-statistic: 0.07474 on 1 and 7 DF,  p-value: 0.7924
```

```
lm_wk3_but_under_plot<-underweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = but_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("F:B ratio wk3") +
  ylab("Butyrate Concentration kg/mmol wk3") + labs(title = "Underweight F:B Ratio and Butyrate Concentration")
print(lm_wk3_but_under_plot)
```

## Underweight F:B Ratio and Butyrate Concentration Correlation wk3



```
lm_wk3_but_under<-underweight_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ but_wk3, data = .) #test relationship
summary(lm_wk3_but_under)
```

```
##
## Call:
## lm(formula = fb_ratio_wk3 ~ but_wk3, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.40413 -0.25710 -0.20974  0.08336  0.91089
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.43773    0.51248   0.854   0.421
## but_wk3      0.04543    0.04461   1.018   0.342
##
## Residual standard error: 0.4595 on 7 degrees of freedom
## Multiple R-squared:  0.129, Adjusted R-squared:  0.004608
## F-statistic: 1.037 on 1 and 7 DF, p-value: 0.3424
```

```
save_plot(filename = "figures/lm_wk1_but_under_plot.pdf",
  plot = lm_wk1_but_under_plot,
  nrow = 1, ncol = 1,
  base_aspect_ratio = 2.1)
```

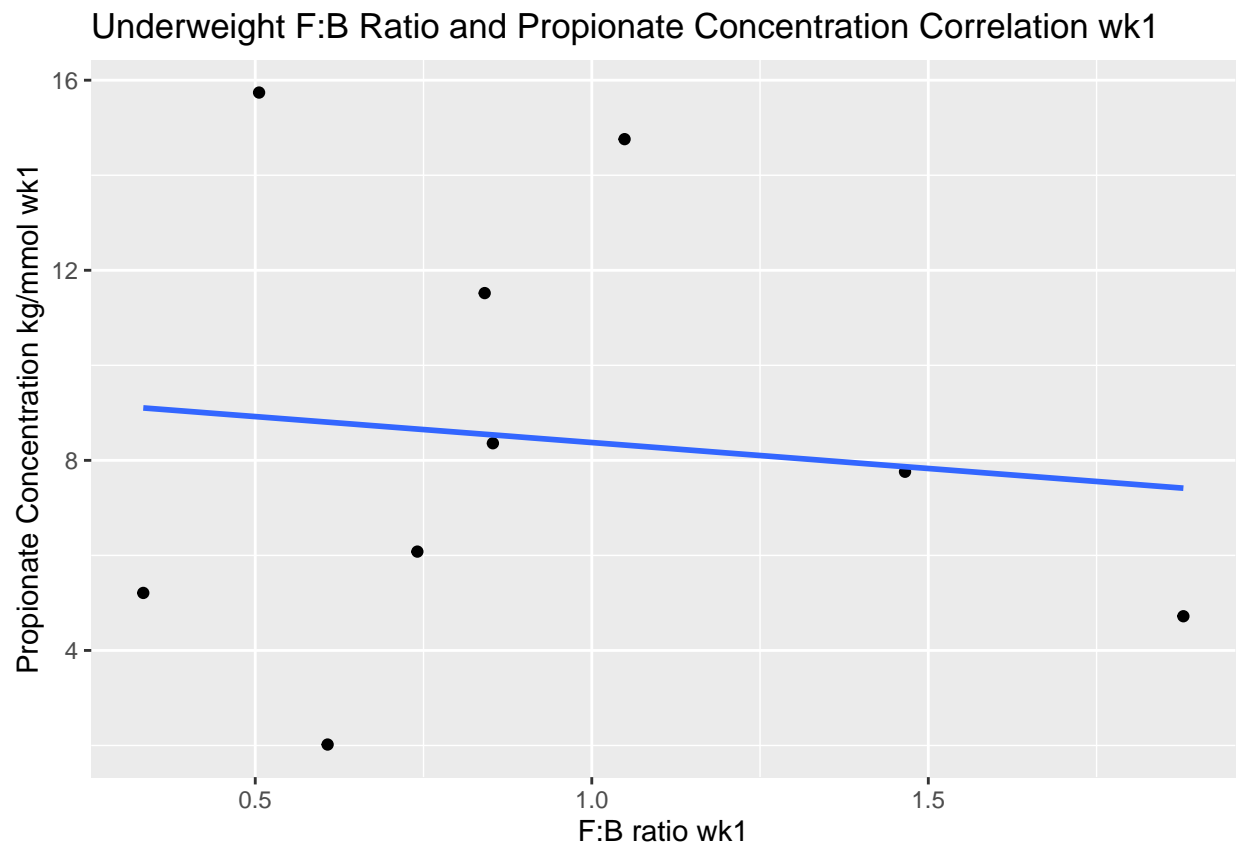
```
save_plot(filename = "figures/lm_wk3_but_under_plot.pdf",
          plot = lm_wk3_but_under_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 Butyrate concentration of the underweight participants is .79, which is not statistically significant. However, the  $R^2$  value is insignificant being very small at  $R^2 = -.1308$ , which represents too much variation in the data. There is no significant correlation between the week 1 firmicutes - bacteroidetes ratio and Butyrate Concentration.

The week 3 F:B ratio and week 3 Butyrate concentration of the underweight participants has a insignificant p-value and  $R^2$  value at  $p = .3424$  and  $R^2 = 0.004608$  respectively. This also represents no significant correlation.

###F:B and Propionate

```
lm_wk1_pro_under_plot<-underweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = pro_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("F:B ratio wk1") +
  ylab("Propionate Concentration kg/mmol wk1") + labs(title = "Underweight F:B Ratio and Propionate Correlation wk1")
print(lm_wk1_pro_under_plot)
```



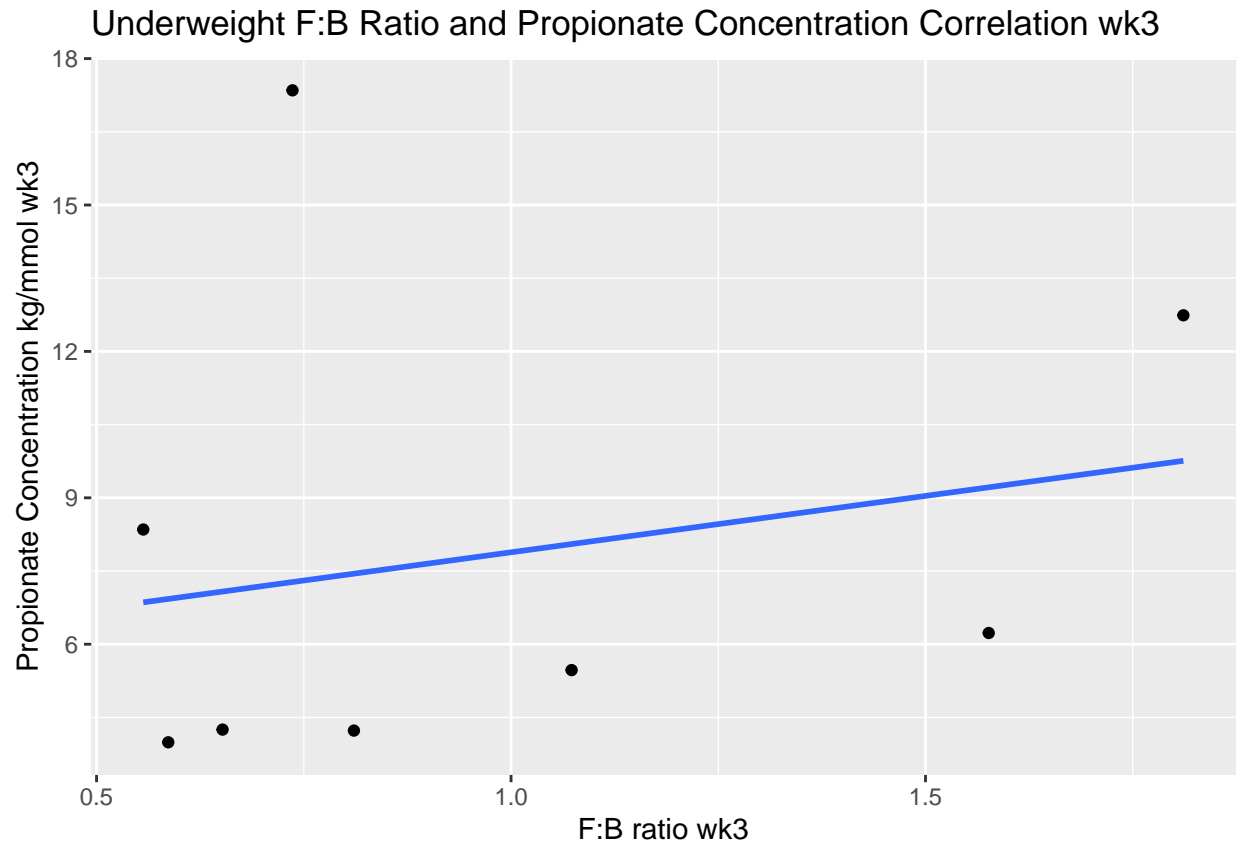
```
lm_wk1_pro_under<-underweight_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ pro_wk1, data = .) #test relationship
summary(lm_wk1_pro_under)
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ pro_wk1, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.62432 -0.32751 -0.06757  0.20391  0.91518
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.01969    0.37301   2.734  0.0292 *
## pro_wk1      -0.01185    0.03910  -0.303  0.7707
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5162 on 7 degrees of freedom
## Multiple R-squared:  0.01295,    Adjusted R-squared:  -0.1281
## F-statistic: 0.09181 on 1 and 7 DF,  p-value: 0.7707
```

```
lm_wk3_pro_under_plot<-underweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = pro_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("F:B ratio wk3") +
  ylab("Propionate Concentration kg/mmol wk3") + labs(title = "Underweight F:B Ratio and Propionate Cor")
print(lm_wk3_pro_under_plot)
```

```
## Warning: Removed 1 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 1 rows containing missing values (geom_point).
```



```
lm_wk3_pro_under<-underweight_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ pro_wk3, data = .) #test relationship
summary(lm_wk3_pro_under)
```

```
##
## Call:
## lm(formula = fb_ratio_wk3 ~ pro_wk3, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.4521 -0.3348 -0.1638  0.2721  0.7259
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.80012    0.35362   2.263  0.0643 .
## pro_wk3      0.02238    0.03912   0.572  0.5880
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5005 on 6 degrees of freedom
## (1 observation deleted due to missingness)
## Multiple R-squared:  0.05172,    Adjusted R-squared:  -0.1063
## F-statistic: 0.3273 on 1 and 6 DF,  p-value: 0.588
```

```

save_plot(filename = "figures/lm_wk1_pro_under_plot.pdf",
          plot = lm_wk1_pro_under_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)

save_plot(filename = "figures/lm_wk3_pro_under_plot.pdf",
          plot = lm_wk3_pro_under_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)

```

```
## Warning: Removed 1 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 1 rows containing missing values (geom_point).
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 Propionate concentration of all the participants is .7707, which is not statistically significant. Additionally, the  $R^2$  value is insignificant being very small at  $R^2 = -0.01281$ , which represents too much variation in the data. There is no significant correlation between the week 1 firmicutes - bacteroidetes ratio and Propionate concentration for underweight participants.

Similarly, The week 3 F - B ratio and week 3 Propionate concentration for underweight participants has a significant p-value and insignificant  $R^2$  value at  $p = 0.588$  and  $R^2 = -0.1063$  respectively. This also represents no significant correlation.

```
##Healthy Weight ###Wk1 F:B and BMI
```

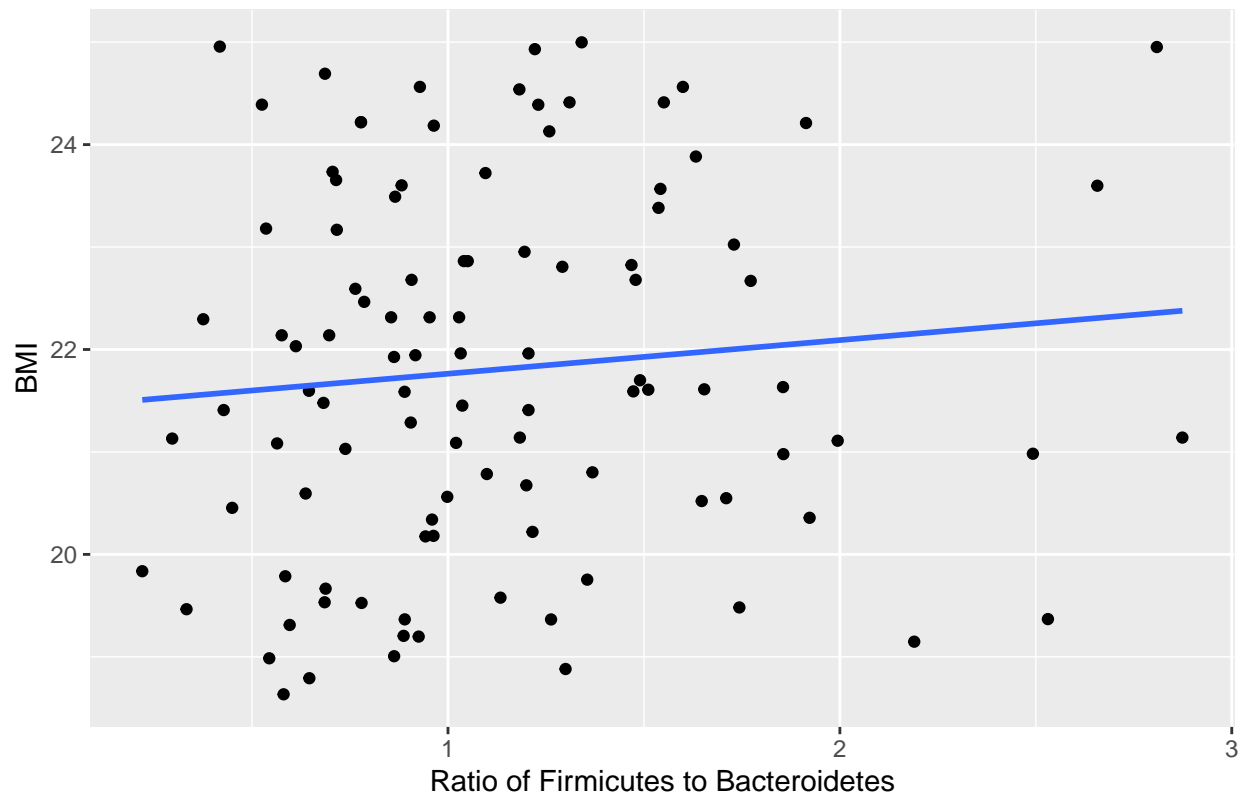
```

#PLOT
hwk1fb_BMI_plot<-healthy_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = BMI)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Ratio of Firmicutes to Bacteroidetes by healthy BMI",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "BMI")
print(hwk1fb_BMI_plot)

```



## Ratio of Firmicutes to Bacteroidetes by healthy BMI



```
#TEST
hwk1fb_BMI_test <- healthy_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ BMI, data = .) #test relationship
summary(hwk1fb_BMI_test) #view results
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ BMI, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.8493 -0.3908 -0.1168  0.3073  1.7647
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)   0.45952    0.65672   0.700   0.486
## BMI           0.03073    0.03002   1.024   0.308
##
## Residual standard error: 0.5471 on 103 degrees of freedom
## Multiple R-squared:  0.01008,    Adjusted R-squared:  0.000465
## F-statistic: 1.048 on 1 and 103 DF,  p-value: 0.3083
```

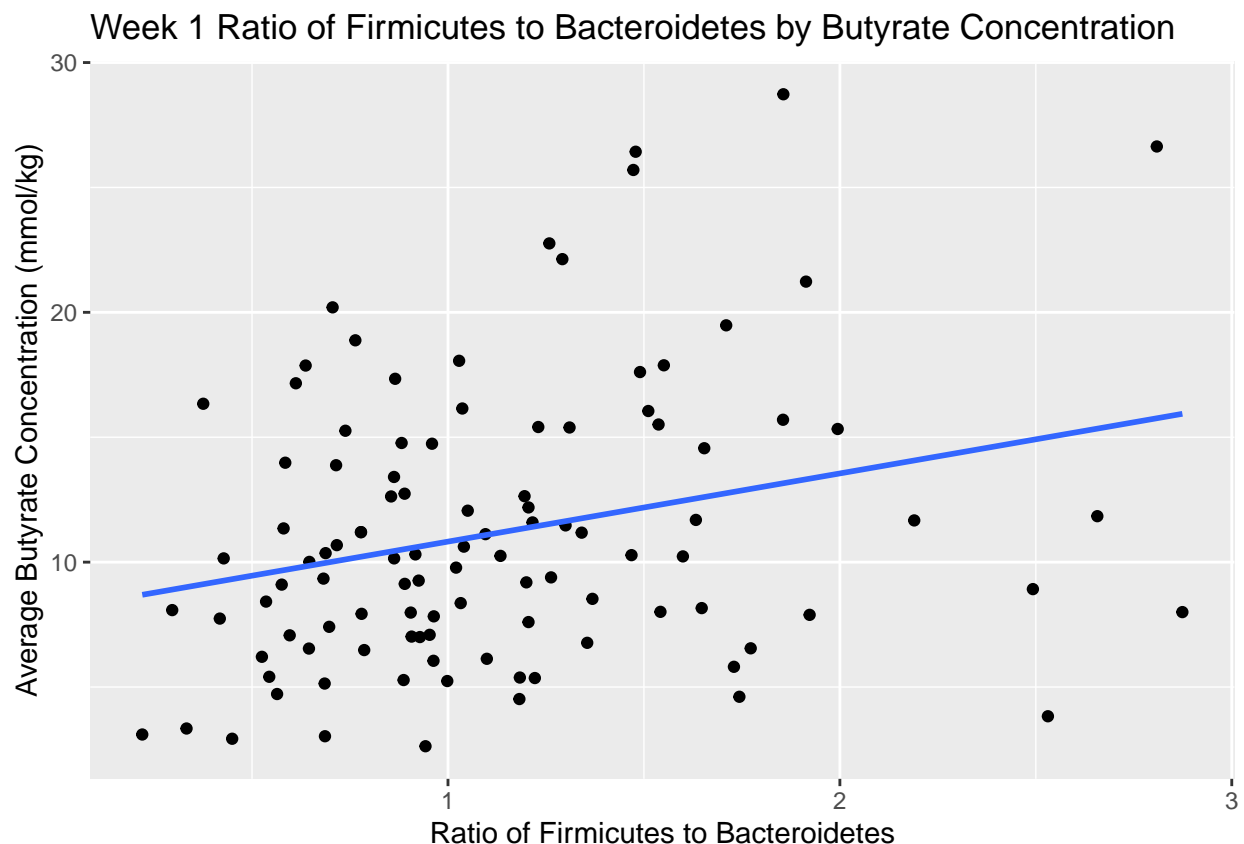
```
#p-value = 0.3083
#Adjusted R-squared = 0.000465
```

```
#Save Plot
save_plot(filename = "figures/hwk1fb_BMI_plot.pdf",
          plot = hwk1fb_BMI_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the BMI of “healthy” participants was 0.3083, which is not statistically significant. Furthermore, the R value was an insignificant value close to zero (0.000465). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and BMI among participants whose BMI falls within the CDC’s “Healthy” category.

###Wk1 F:B and Wk1 Butyrate

```
#PLOT
hwk1fb_wk1but_plot<-healthy_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
            y = but_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
            se = FALSE) + #hides confidence interval around line
  labs(title = "Week 1 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Butyrate Concentration (mmol/kg)")
print(hwk1fb_wk1but_plot)
```



```

#TEST
hwk1fb_wk1but_test <- healthy_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ but_wk1, data = .) #test relationship
summary(hwk1fb_wk1but_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk1 ~ but_wk1, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.89047 -0.36217 -0.07759  0.24425  1.82806
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.834786   0.116413   7.171 1.17e-10 ***
## but_wk1      0.026389   0.009333   2.828  0.00564 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5297 on 103 degrees of freedom
## Multiple R-squared:  0.07203,    Adjusted R-squared:  0.06302
## F-statistic: 7.995 on 1 and 103 DF,  p-value: 0.005637

#p-value = 0.005637
#Adjusted R-squared = 0.06302

#Save Plot
save_plot(filename = "figures/hwk1fb_wk1but_plot.pdf",
  plot = hwk1fb_wk1but_plot,
  nrow = 1, ncol = 1,
  base_aspect_ratio = 2.1)

```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 average butyrate concentration among “healthy” participants was 0.005637, which falls within a 95% confidence interval. However, the R value was an insignificant value close to zero (0.06302). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and the week 1 average butyrate concentration among participants whose BMI falls within the CDC’s “Healthy” category.

###Wk1 F:B and Wk1 Propionate

```

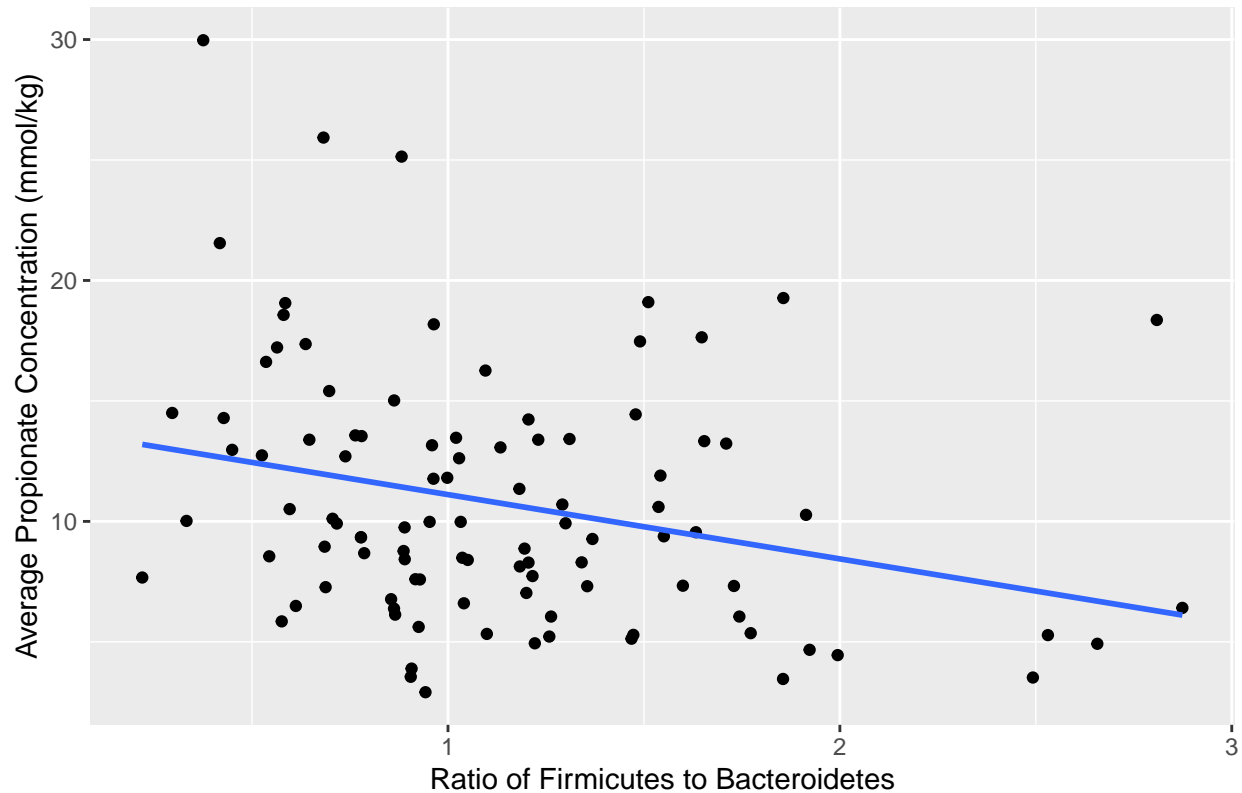
#PLOT
hwk1fb_wk1pro_plot<-healthy_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
    y = pro_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
    se = FALSE) + #hides confidence interval around line
  labs(title = "Week 1 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration",
    x = "Ratio of Firmicutes to Bacteroidetes",
    y = "Average Propionate Concentration (mmol/kg)")
print(hwk1fb_wk1pro_plot)

```

```
## Warning: Removed 4 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 4 rows containing missing values (geom_point).
```

### Week 1 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration



```
#TEST
hwk1fb_wk1pro_test <- healthy_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ pro_wk1, data = .) #test relationship
summary(hwk1fb_wk1pro_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk1 ~ pro_wk1, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -1.0026 -0.3938 -0.1033  0.1928  1.8976
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.44580    0.12032  12.017  < 2e-16 ***
## pro_wk1      -0.02912    0.01008  -2.889  0.00475 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5235 on 99 degrees of freedom
```

```
## (4 observations deleted due to missingness)
## Multiple R-squared:  0.07773,    Adjusted R-squared:  0.06841
## F-statistic: 8.344 on 1 and 99 DF,  p-value: 0.004754
```

```
#p-value = 0.004754
#Adjusted R-squared = 0.06841
```

```
#Save Plot
save_plot(filename = "figures/hwk1fb_wk1pro_plot.pdf",
          plot = hwk1fb_wk1pro_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)
```

```
## Warning: Removed 4 rows containing non-finite values (stat_smooth).
```

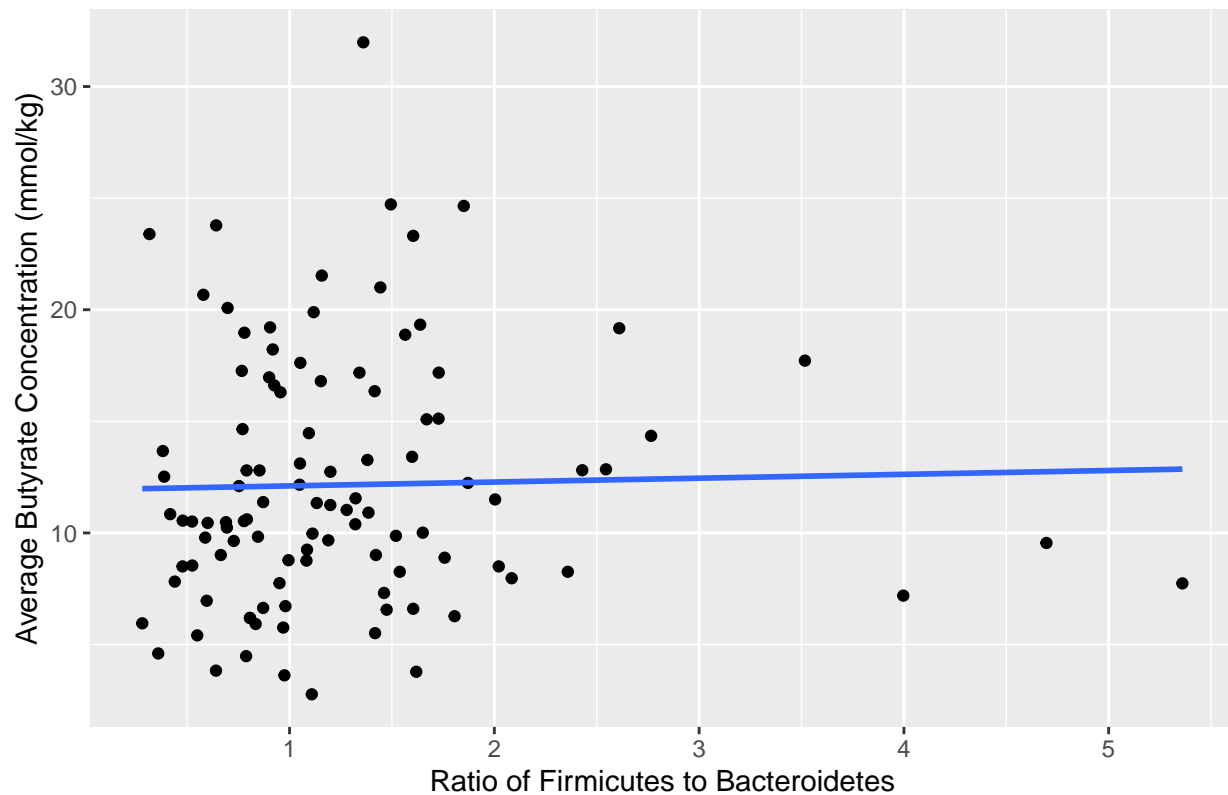
```
## Warning: Removed 4 rows containing missing values (geom_point).
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 average propionate concentration among “healthy” participants was 0.004754, which falls within a 95% confidence interval. However, the R value was an insignificant value close to zero (0.06841). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and the week 1 average propionate concentration among participants whose BMI falls within the CDC’s “Healthy” category.

### Wk3 F:B and Wk3 Butyrate

```
#PLOT
hwk3fb_wk3but_plot<-healthy_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = but_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 3 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Butyrate Concentration (mmol/kg)")
print(hwk3fb_wk3but_plot)
```

### Week 3 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration



```
#TEST
hwk3fb_wk3but_test <- healthy_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ but_wk3, data = .) #test relationship
summary(hwk3fb_wk3but_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk3 ~ but_wk3, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.9964 -0.5083 -0.1810  0.2699  4.1076
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.222715    0.195979   6.239 9.89e-09 ***
## but_wk3      0.003825    0.014673   0.261  0.795
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.8328 on 103 degrees of freedom
## Multiple R-squared:  0.0006594, Adjusted R-squared: -0.009043
## F-statistic: 0.06797 on 1 and 103 DF, p-value: 0.7948
```

```

#p-value = 0.7948
#Adjusted R-squared = -0.009043

#Save Plot
save_plot(filename = "figures/hwk3fb_wk3but_plot.pdf",
          plot = hwk3fb_wk3but_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)

```

The p-value for the linear correlation of the week 3 firmicutes to bacteroidetes ratio and the week 3 average butyrate concentration among “healthy” participants was 0.7948, which is not statistically significant. Furthermore, the R value was an insignificant value close to zero (-0.009043). There does not appear to be a statistically significant linear relationship between the week 3 firmicutes to bacteroidetes ratio and the week 3 average butyrate concentration among participants whose BMI falls within the CDC’s “Healthy” category.

###Wk3 F:B and Wk3 Propionate

```

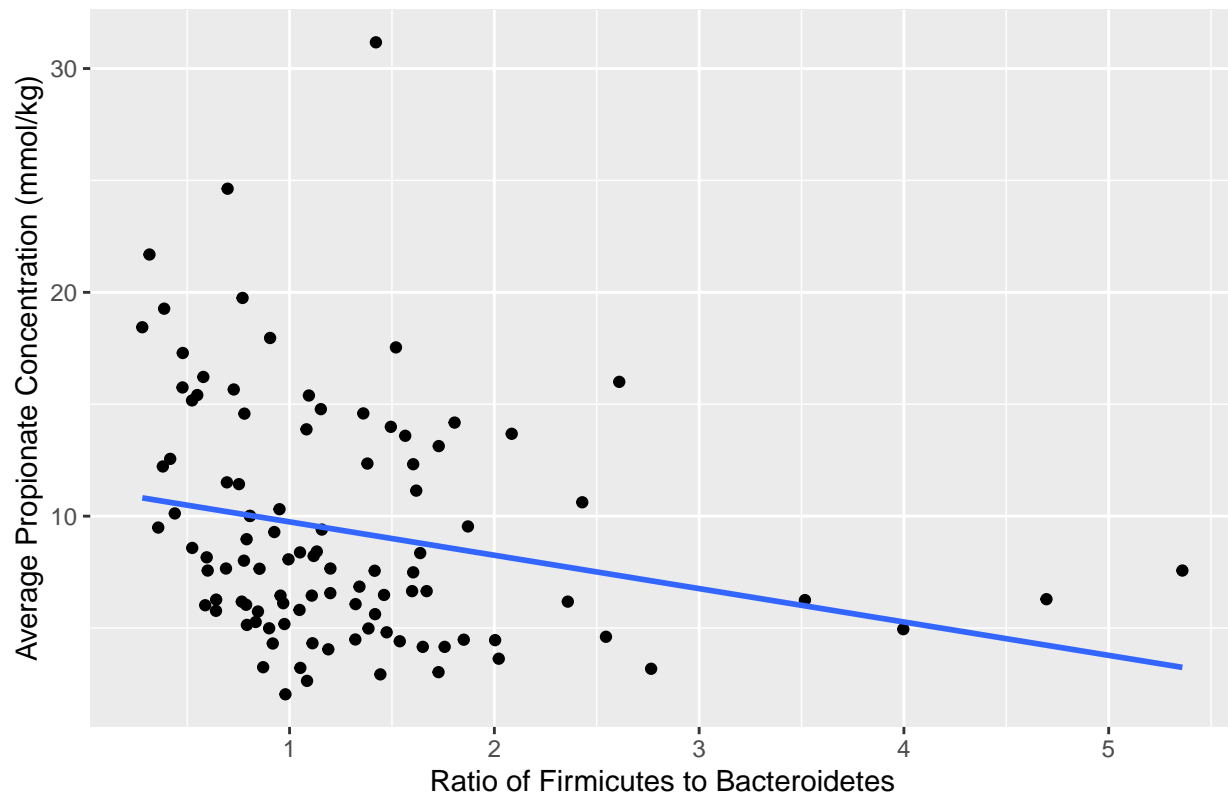
#PLOT
hwk3fb_wk3pro_plot<-healthy_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = pro_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 3 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Propionate Concentration (mmol/kg)")
print(hwk3fb_wk3pro_plot)

```

## Warning: Removed 3 rows containing non-finite values (stat\_smooth).

## Warning: Removed 3 rows containing missing values (geom\_point).

### Week 3 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration



```
#TEST
hwk3fb_wk3pro_test <- healthy_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ pro_wk3, data = .) #test relationship
summary(hwk3fb_wk3pro_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk3 ~ pro_wk3, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.9143 -0.5030 -0.1932  0.2727  4.0151
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.62858    0.16498   9.871  <2e-16 ***
## pro_wk3      -0.03748    0.01540  -2.433   0.0167 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.8185 on 100 degrees of freedom
## (3 observations deleted due to missingness)
## Multiple R-squared:  0.05589,    Adjusted R-squared:  0.04645
## F-statistic:  5.92 on 1 and 100 DF,  p-value: 0.01675
```



```

#p-value = 0.01675
#Adjusted R-squared = 0.04645

#Save Plot
save_plot(filename = "figures/hwk3fb_wk3pro_plot.pdf",
          plot = hwk3fb_wk3pro_plot,
          nrow = 1, ncol = 1,
          base_aspect_ratio = 2.1)

```

```
## Warning: Removed 3 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 3 rows containing missing values (geom_point).
```

The p-value for the linear correlation of the week 3 firmicutes to bacteroidetes ratio and the week 3 average propionate concentration among “healthy” participants was 0.01675, which falls within a 95% confidence interval. However, the R value was an insignificant value close to zero (0.04645). There does not appear to be a statistically significant linear relationship between the week 3 firmicutes to bacteroidetes ratio and the week 3 average propionate concentration among participants whose BMI falls within the CDC’s “Healthy” category.

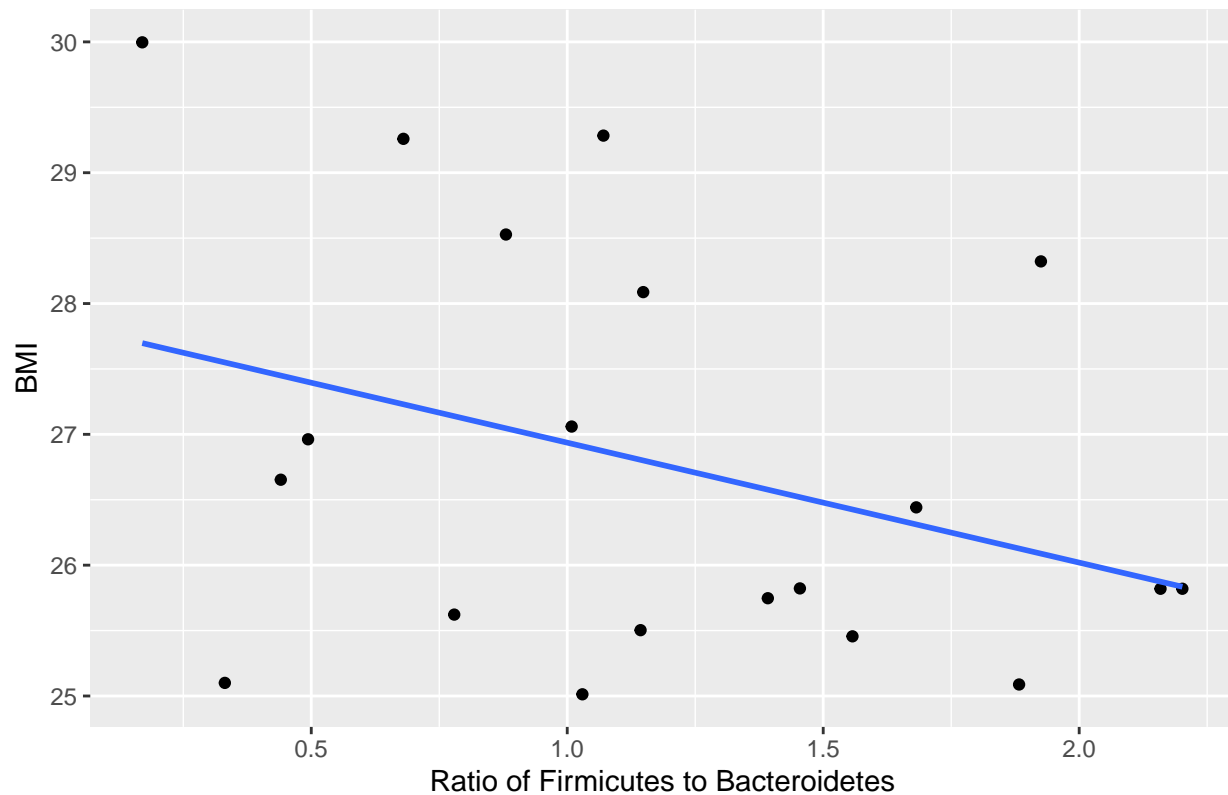
```
##Overweight ##Wk1 F:B and BMI
```

```

#PLOT
ovwk1fb_BMI_plot<-overweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = BMI)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
              se = FALSE) + #hides confidence interval around line
  labs(title = "Ratio of Firmicutes to Bacteroidetes by Overweight BMI",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "BMI")
print(ovwk1fb_BMI_plot)

```

## Ratio of Firmicutes to Bacteroidetes by Overweight BMI



```
#TEST
ovwk1fb_BMI_test <- overweight_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ BMI, data = .) #test relationship
summary(ovwk1fb_BMI_test) #view results
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ BMI, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -1.06352 -0.41901  0.01222  0.29048  0.95898
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  4.73184    2.25636   2.097  0.0504 .
## BMI         -0.13295    0.08412  -1.581  0.1314
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5797 on 18 degrees of freedom
## Multiple R-squared:  0.1219, Adjusted R-squared:  0.07309
## F-statistic: 2.498 on 1 and 18 DF,  p-value: 0.1314
```

```
#p-value = 0.1314
#Adjusted R-squared = 0.07309
```

```
#Save Plot
```

```
save_plot(filename = "figures/ovwk1fb_BMI_plot.pdf", plot = ovwk1fb_BMI_plot, nrow = 1, ncol = 1, base.s
```

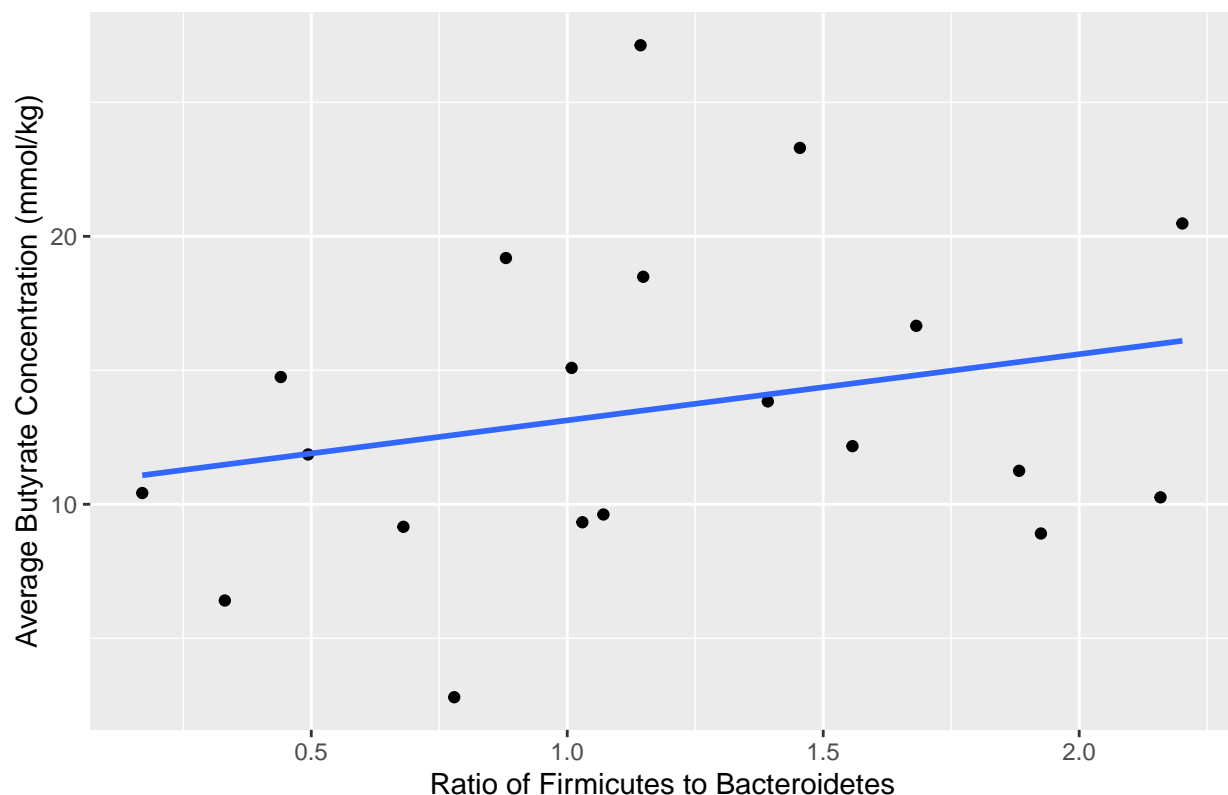
The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the BMI of “overweight” participants was 0.1314, which is not statistically significant. Furthermore, the R value was an insignificant value close to zero (0.07309). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and BMI among participants whose BMI falls within the CDC’s “Overweight” category.

```
###Wk1 F:B and Wk1 Butyrate
```

```
#PLOT
```

```
ovwk1fb_wk1but_plot<-overweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = but_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 1 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Butyrate Concentration (mmol/kg)")
print(ovwk1fb_wk1but_plot)
```

Week 1 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration



```

#TEST
ovwk1fb_wk1but_test <- overweight_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ but_wk1, data = .) #test relationship
summary(ovwk1fb_wk1but_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk1 ~ but_wk1, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.92175 -0.39333 -0.07635  0.42356  1.07132
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)   0.82642    0.34136   2.421  0.0263 *
## but_wk1       0.02545    0.02316   1.099  0.2863
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5989 on 18 degrees of freedom
## Multiple R-squared:  0.06286,    Adjusted R-squared:  0.0108
## F-statistic: 1.207 on 1 and 18 DF,  p-value: 0.2863

#p-value = 0.2863
#Adjusted R-squared = 0.0108

#Save Plot
save_plot(filename = "figures/ovwk1fb_wk1but_plot.pdf", plot = ovwk1fb_wk1but_plot, nrow = 1, ncol = 1,

```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 average butyrate concentration among “overweight” participants was 0.2863, which falls within a 95% confidence interval. However, the R value was an insignificant value close to zero (0.0108). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and the week 1 average butyrate concentration among participants whose BMI falls within the CDC’s “Overweight” category.

### Wk1 F:B and Wk1 Propionate

```

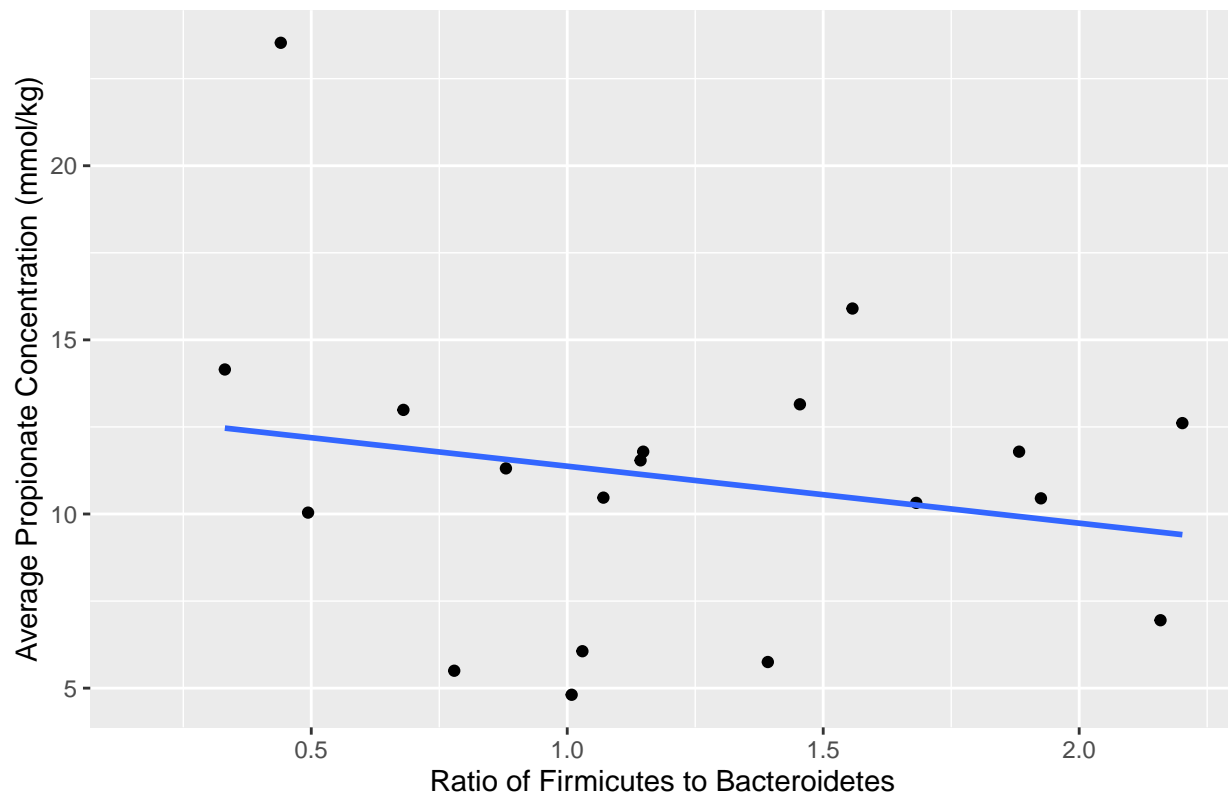
#PLOT
ovwk1fb_wk1pro_plot<-overweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = pro_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 1 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Propionate Concentration (mmol/kg)")
print(ovwk1fb_wk1pro_plot)

```

```
## Warning: Removed 1 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 1 rows containing missing values (geom_point).
```

### Week 1 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration



```
#TEST
ovwk1fb_wk1pro_test <- overweight_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ pro_wk1, data = .) #test relationship
summary(ovwk1fb_wk1pro_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk1 ~ pro_wk1, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.80543 -0.41150 -0.06605  0.45390  1.02211
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.53069    0.36471   4.197 0.000605 ***
## pro_wk1      -0.02785    0.03092  -0.901 0.380231
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5723 on 17 degrees of freedom
## (1 observation deleted due to missingness)
## Multiple R-squared:  0.04557,    Adjusted R-squared:  -0.01058
## F-statistic: 0.8116 on 1 and 17 DF,  p-value: 0.3802
```

```

#p-value = 0.3802
#Adjusted R-squared = -0.01058

#Save Plot
save_plot(filename = "figures/ovwk1fb_wk1pro_plot.pdf", plot = ovwk1fb_wk1pro_plot, nrow = 1, ncol = 1,

## Warning: Removed 1 rows containing non-finite values (stat_smooth).

## Warning: Removed 1 rows containing missing values (geom_point).

```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 average propionate concentration among “overweight” participants was 0.3802, which falls within a 95% confidence interval. However, the R value was an insignificant value close to zero (-0.01058). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and the week 1 average propionate concentration among participants whose BMI falls within the CDC’s “Overweight” category.

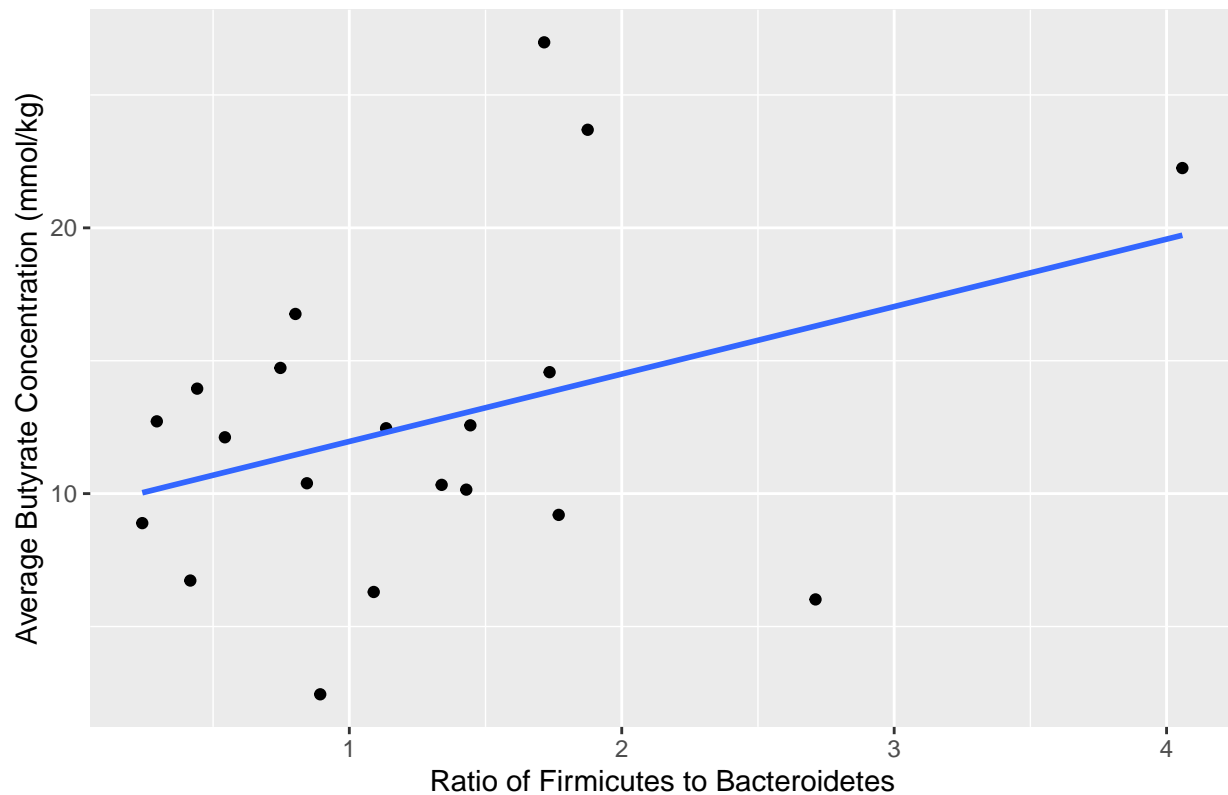
###Wk3 F:B and Wk3 Butyrate

```

#PLOT
ovwk3fb_wk3but_plot<-overweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = but_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 3 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Butyrate Concentration (mmol/kg)")
print(ovwk3fb_wk3but_plot)

```

### Week 3 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration



```
#TEST
ovwk3fb_wk3but_test <- overweight_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ but_wk3, data = .) #test relationship
summary(ovwk3fb_wk3but_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk3 ~ but_wk3, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.98589 -0.65940 -0.07489  0.21897  2.24361
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)   0.56514    0.45450   1.243   0.230
## but_wk3       0.05615    0.03247   1.729   0.101
##
## Residual standard error: 0.8666 on 18 degrees of freedom
## Multiple R-squared:  0.1425, Adjusted R-squared:  0.09483
## F-statistic: 2.991 on 1 and 18 DF,  p-value: 0.1009

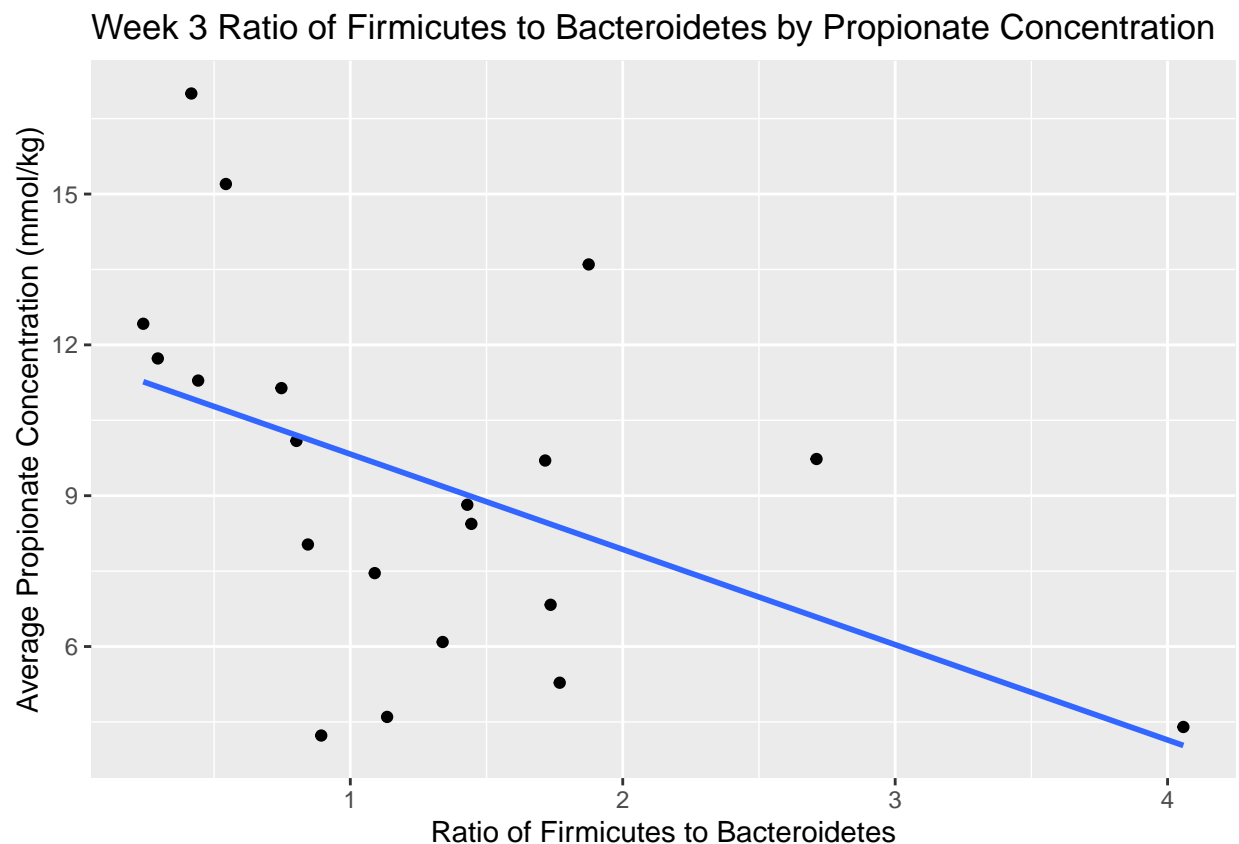
#p-value = 0.1009
#Adjusted R-squared = 0.09483
```

```
#Save Plot
save_plot(filename = "figures/ovwk3fb_wk3but_plot.pdf", plot = ovwk3fb_wk3but_plot, nrow = 1, ncol = 1,
```

The p-value for the linear correlation of the week 3 firmicutes to bacteroidetes ratio and the week 3 average butyrate concentration among “overweight” participants was 0.1009, which is not statistically significant. Furthermore, the R value was an insignificant value close to zero (0.09483). There does not appear to be a statistically significant linear relationship between the week 3 firmicutes to bacteroidetes ratio and the week 3 average butyrate concentration among participants whose BMI falls within the CDC’s “Overweight” category.

###Wk3 F:B and Wk3 Propionate

```
#PLOT
ovwk3fb_wk3pro_plot<-overweight_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = pro_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 3 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Propionate Concentration (mmol/kg)")
print(ovwk3fb_wk3pro_plot)
```





```

#TEST
ovwk3fb_wk3pro_test <- overweight_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ pro_wk3, data = .) #test relationship
summary(ovwk3fb_wk3pro_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk3 ~ pro_wk3, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.9969 -0.5880 -0.1631  0.1110  2.1885
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)   2.40195     0.52000   4.619 0.000213 ***
## pro_wk3       -0.12101     0.05229  -2.314 0.032668 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.8215 on 18 degrees of freedom
## Multiple R-squared:  0.2293, Adjusted R-squared:  0.1865
## F-statistic: 5.356 on 1 and 18 DF,  p-value: 0.03267

#p-value = 0.03267
#Adjusted R-squared = 0.1865

#Save Plot
save_plot(filename = "figures/ovwk3fb_wk3pro_plot.pdf", plot = ovwk3fb_wk3pro_plot, nrow = 1, ncol = 1,

```

The p-value for the linear correlation of the week 3 firmicutes to bacteroidetes ratio and the week 3 average propionate concentration among “healthy” participants was 0.03267, which technically supports the alternative hypothesis. However, the R value was an insignificant value (0.1865). There does not appear to be a statistically significant linear relationship between the week 3 firmicutes to bacteroidetes ratio and the week 3 average propionate concentration among participants whose BMI falls within the CDC’s “Overweight” category.

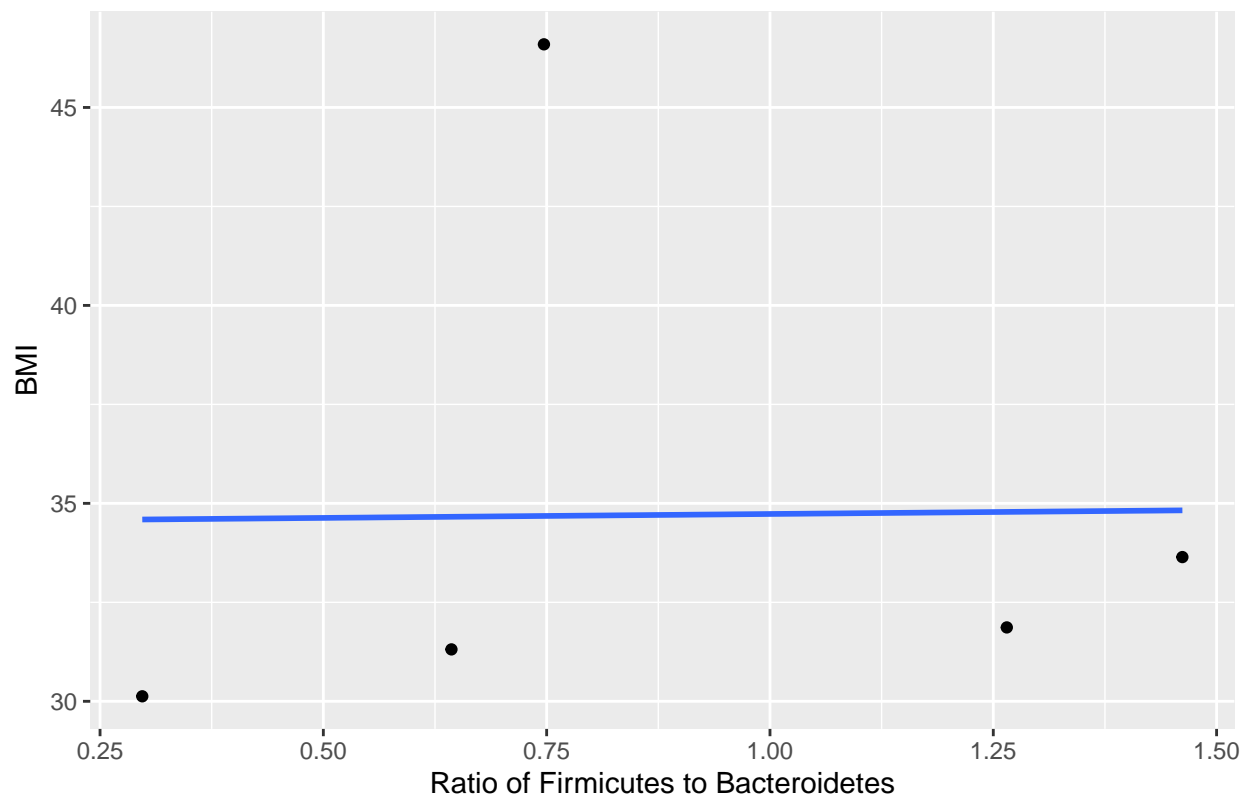
##Obese ###Wk1 F:B and BMI

```

#PLOT
obwk1fb_BMI_plot<-obese_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = BMI)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Ratio of Firmicutes to Bacteroidetes by Obese BMI",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "BMI")
print(obwk1fb_BMI_plot)

```

## Ratio of Firmicutes to Bacteroidetes by Obese BMI



```
#TEST
obwk1fb_BMI_test <- obese_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ BMI, data = .) #test relationship
summary(obwk1fb_BMI_test) #view results
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ BMI, data = .)
##
## Residuals:
##      1      2      3      4      5
## -0.2362 -0.1475  0.5798 -0.5811  0.3850
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.8491209  1.4261369   0.595   0.594
## BMI          0.0009733  0.0404790   0.024   0.982
##
## Residual standard error: 0.5476 on 3 degrees of freedom
## Multiple R-squared:  0.0001927, Adjusted R-squared:  -0.3331
## F-statistic: 0.0005781 on 1 and 3 DF, p-value: 0.9823
```

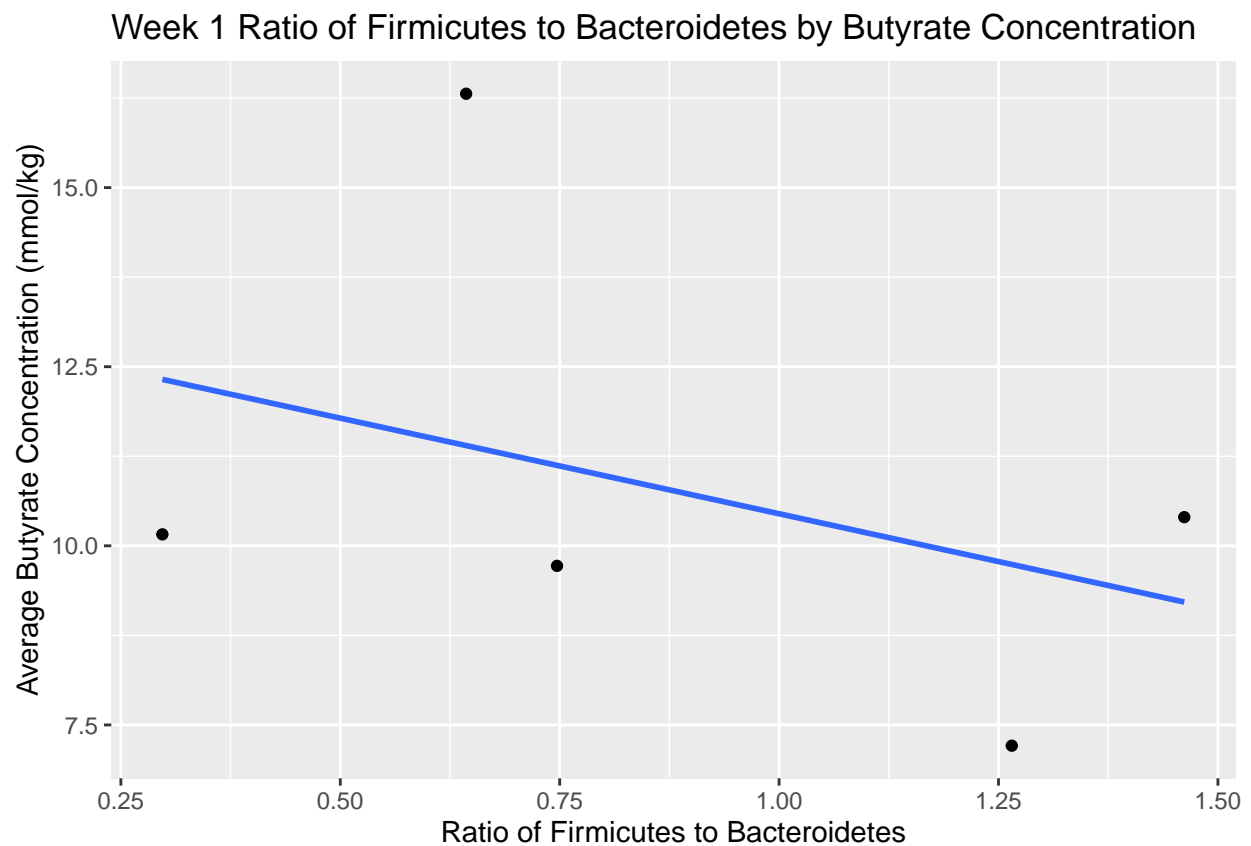
```
#p-value = 0.9823
#Adjusted R-squared = 0.3331
```

```
#Save Plot
save_plot(filename = "figures/obwk1fb_BMI_plot.pdf", plot = obwk1fb_BMI_plot, nrow = 1, ncol = 1, base_
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the BMI of “obese” participants was 0.9823, which is not statistically significant. Furthermore, the R value was an insignificant value (0.3331). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and BMI among participants whose BMI falls within the CDC’s “Obese” category.

###Wk1 F:B and Wk1 Butyrate

```
#PLOT
obwk1fb_wk1but_plot<-obese_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = but_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 1 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Butyrate Concentration (mmol/kg)")
print(obwk1fb_wk1but_plot)
```



```
#TEST
obwk1fb_wk1but_test <- obese_phylum_scfa %>%
```

```
lm(fb_ratio_wk1 ~ but_wk1, data = .) #test relationship
summary(obwk1fb_wk1but_test) #view results
```

```
##
## Call:
## lm(formula = fb_ratio_wk1 ~ but_wk1, data = .)
##
## Residuals:
##      1      2      3      4      5
## 0.05688 -0.19146  0.55956 -0.61765  0.19267
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.45749    0.84471   1.725   0.183
## but_wk1      -0.05340    0.07562  -0.706   0.531
##
## Residual standard error: 0.5072 on 3 degrees of freedom
## Multiple R-squared:  0.1425, Adjusted R-squared:  -0.1433
## F-statistic: 0.4986 on 1 and 3 DF,  p-value: 0.531
```

```
#p-value = 0.531
```

```
#Adjusted R-squared = -0.1433
```

```
#Save Plot
```

```
save_plot(filename = "figures/obwk1fb_wk1but_plot.pdf", plot = obwk1fb_wk1but_plot, nrow = 1, ncol = 1,
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 average butyrate concentration among “obese” participants was 0.531, which falls within a 95% confidence interval. However, the R value was an insignificant value (-0.1433). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and the week 1 average butyrate concentration among participants whose BMI falls within the CDC’s “Obese” category.

### Wk1 F:B and Wk1 Propionate

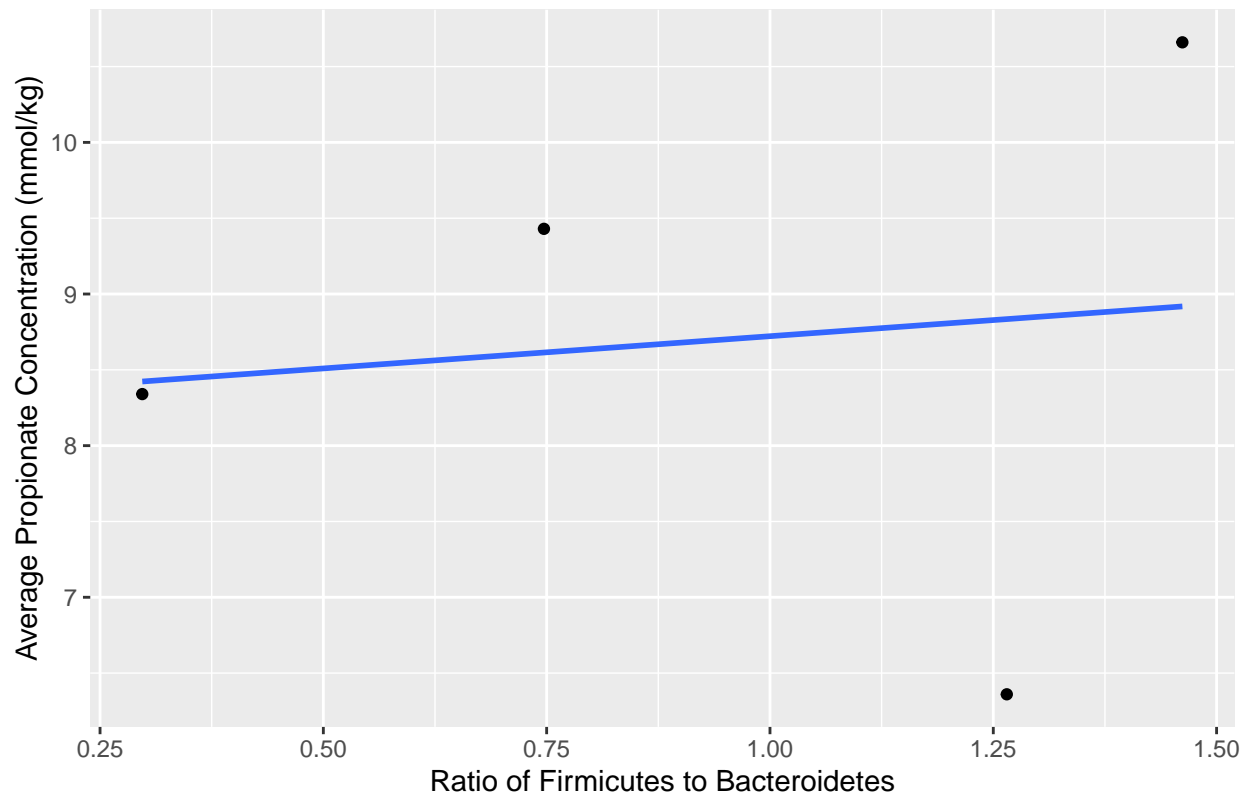
```
#PLOT
```

```
obwk1fb_wk1pro_plot<-obese_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk1,
             y = pro_wk1)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 1 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Propionate Concentration (mmol/kg)")
print(obwk1fb_wk1pro_plot)
```

```
## Warning: Removed 1 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 1 rows containing missing values (geom_point).
```

## Week 1 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration



```
#TEST
obwk1fb_wk1pro_test <- obese_phylum_scfa %>%
  lm(fb_ratio_wk1 ~ pro_wk1, data = .) #test relationship
summary(obwk1fb_wk1pro_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk1 ~ pro_wk1, data = .)
##
## Residuals:
##      2      3      4      5
## -0.2216  0.4497 -0.6329  0.4049
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.63585    1.78708   0.356   0.756
## pro_wk1      0.03529    0.20216   0.175   0.877
##
## Residual standard error: 0.6386 on 2 degrees of freedom
## (1 observation deleted due to missingness)
## Multiple R-squared:  0.01501,    Adjusted R-squared:  -0.4775
## F-statistic: 0.03047 on 1 and 2 DF,  p-value: 0.8775
```

```
#p-value = 0.8775
#Adjusted R-squared = -0.4775
```

```
#Save Plot
save_plot(filename = "figures/obwk1fb_wk1pro_plot.pdf", plot = obwk1fb_wk1pro_plot, nrow = 1, ncol = 1,
```

```
## Warning: Removed 1 rows containing non-finite values (stat_smooth).
```

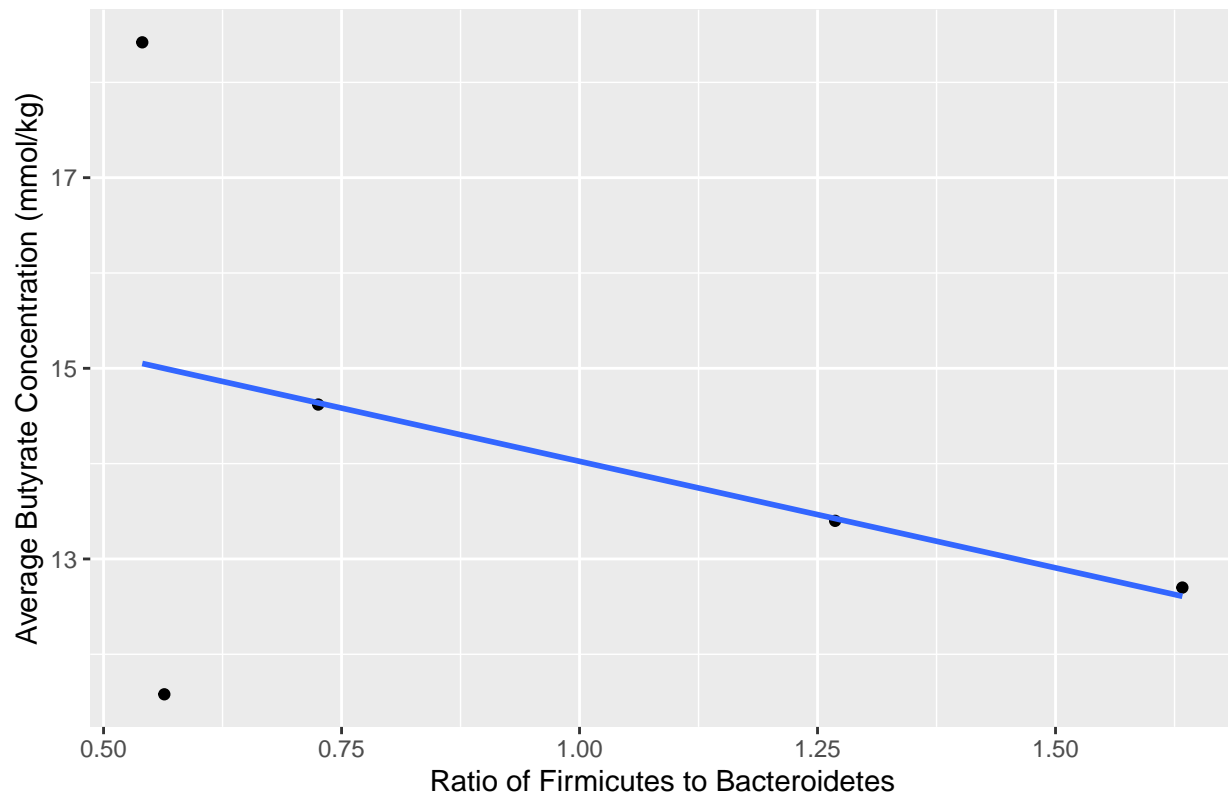
```
## Warning: Removed 1 rows containing missing values (geom_point).
```

The p-value for the linear correlation of the week 1 firmicutes to bacteroidetes ratio and the week 1 average propionate concentration among “obese” participants was 0.8775, which falls within a 95% confidence interval. However, the R value was an insignificant value (-0.4775). There does not appear to be a statistically significant linear relationship between the week 1 firmicutes to bacteroidetes ratio and the week 1 average propionate concentration among participants whose BMI falls within the CDC’s “Obese” category.

###Wk3 F:B and Wk3 Butyrate

```
#PLOT
obwk3fb_wk3but_plot<-obese_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = but_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 3 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Butyrate Concentration (mmol/kg)")
print(obwk3fb_wk3but_plot)
```

### Week 3 Ratio of Firmicutes to Bacteroidetes by Butyrate Concentration



```
#TEST
obwk3fb_wk3but_test <- obese_phylum_scfa %>%
  lm(fb_ratio_wk3 ~ but_wk3, data = .) #test relationship
summary(obwk3fb_wk3but_test) #view results

##
## Call:
## lm(formula = fb_ratio_wk3 ~ but_wk3, data = .)
##
## Residuals:
##      1      2      3      4      5
## -0.18497 -0.08322  0.26603 -0.57585  0.57801
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  2.01291    1.38645   1.452   0.242
## but_wk3     -0.07541    0.09669  -0.780   0.492
##
## Residual standard error: 0.5091 on 3 degrees of freedom
## Multiple R-squared:  0.1686, Adjusted R-squared:  -0.1086
## F-statistic: 0.6082 on 1 and 3 DF,  p-value: 0.4923

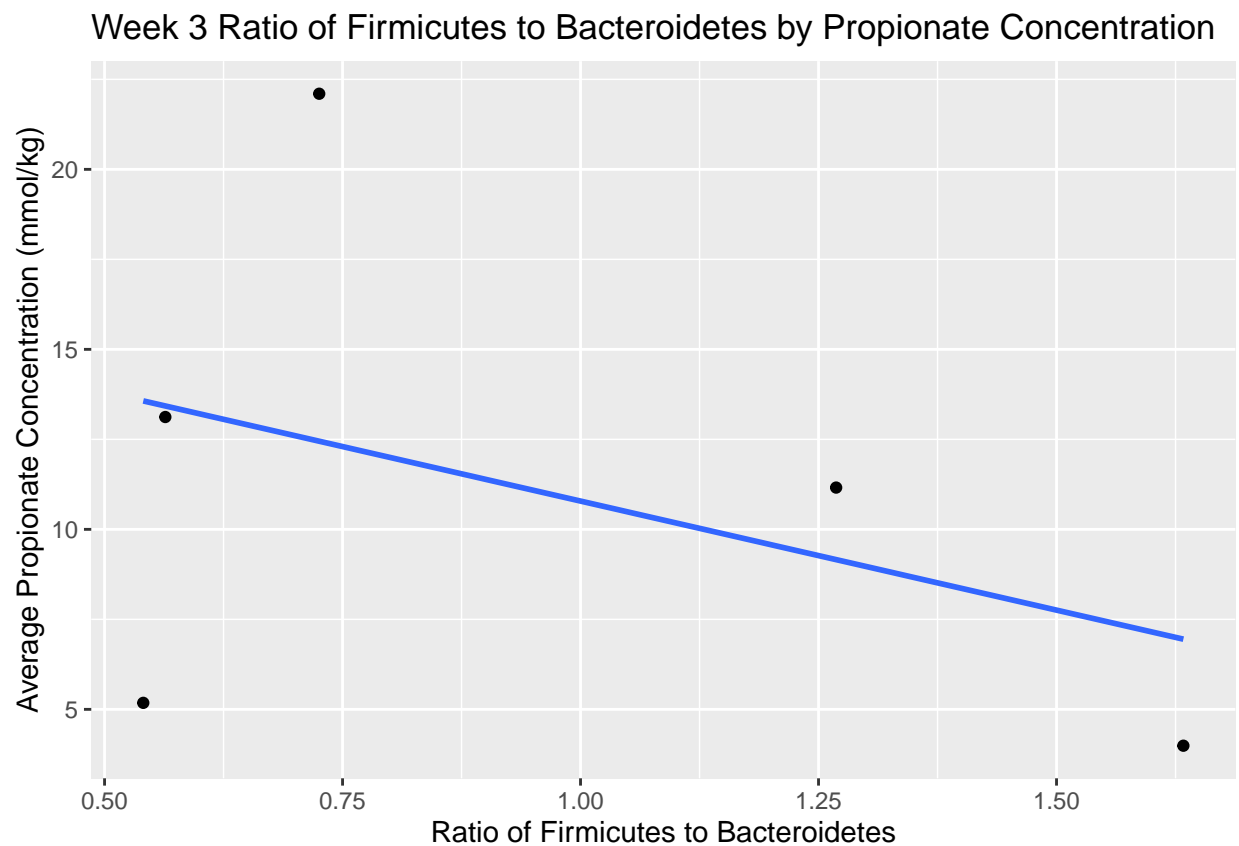
#p-value = 0.4923
#Adjusted R-squared = -0.1086
```

```
#Save Plot
save_plot(filename = "figures/obwk3fb_wk3but_plot.pdf", plot = obwk3fb_wk3but_plot, nrow = 1, ncol = 1,
```

The p-value for the linear correlation of the week 3 firmicutes to bacteroidetes ratio and the week 3 average butyrate concentration among “obese” participants was 0.4923, which is not statistically significant. Furthermore, the R value was an insignificant value close to zero (-0.1086). There does not appear to be a statistically significant linear relationship between the week 3 firmicutes to bacteroidetes ratio and the week 3 average butyrate concentration among participants whose BMI falls within the CDC’s “Obese” category.

###Wk3 F:B and Wk3 Propionate

```
#PLOT
obwk3fb_wk3pro_plot<-obese_phylum_scfa %>%
  ggplot(aes(x = fb_ratio_wk3,
             y = pro_wk3)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  labs(title = "Week 3 Ratio of Firmicutes to Bacteroidetes by Propionate Concentration",
       x = "Ratio of Firmicutes to Bacteroidetes",
       y = "Average Propionate Concentration (mmol/kg)")
print(obwk3fb_wk3pro_plot)
```



```
#TEST
obwk3fb_wk3pro_test <- obese_phylum_scfa %>%
```



```
lm(fb_ratio_wk3 ~ pro_wk3, data = .) #test relationship
summary(obwk3fb_wk3pro_test) #view results
```

```
##
## Call:
## lm(formula = fb_ratio_wk3 ~ pro_wk3, data = .)
##
## Residuals:
##      1      2      3      4      5
## 0.07465 -0.56512  0.32348 -0.32846  0.49544
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.24509    0.45287   2.749  0.0708 .
## pro_wk3      -0.02689    0.03520  -0.764  0.5005
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.5109 on 3 degrees of freedom
## Multiple R-squared:  0.1629, Adjusted R-squared:  -0.1162
## F-statistic: 0.5837 on 1 and 3 DF,  p-value: 0.5005
```

```
#p-value = 0.5005
```

```
#Adjusted R-squared = -0.1162
```

```
#Save Plot
```

```
save_plot(filename = "figures/obwk3fb_wk3pro_plot.pdf", plot = obwk3fb_wk3pro_plot, nrow = 1, ncol = 1,
```

The p-value for the linear correlation of the week 3 firmicutes to bacteroidetes ratio and the week 3 average propionate concentration among “obese” participants was 0.5005, which falls within a 95% confidence interval. However, the R value was an insignificant value close to zero (-0.1162). There does not appear to be a statistically significant linear relationship between the week 3 firmicutes to bacteroidetes ratio and the week 3 average propionate concentration among participants whose BMI falls within the CDC’s “Obese” category.

#Final Conclusions Based on the linear correlation calculations and plots above, it does not appear that any of the above tested variables have a statistically significant relationship. It is possible that we failed to detect a significant relationship, because of the many limitations of this project. For one, the only type of relationship tested were linear correlations, but it is possible that these variables are related in other ways. Additionally, our analysis used weekly average measurements for scfa concentrations (mainly due to time constraints) rather than using individual scfa measurements. Finally, there was limited BMI data which narrowed our sample sizes quite a bit (participants were especially low in the underweight and obese BMI categories).

#Supplemental: Pre-Work This analysis was completed prior to analyzing BMI data and was not presented in our final presentation. These tests investigate potential relationships between butyrate and propionate and were eventually subsetted by specific supplements. We also tried subsetting for only when butyrate increased and only when propionate decreased. We hypothesized that an increase in butyrate would be correlated with a decrease in propionate. #Broad ### Tidy Data #### Week 1 - Week 3 Change All

```
but_wide <- all_data_wkly %>%
```

```
  rename_all(tolower) %>%
```

```
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, buty:
```

```
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes") %>% #keep only wee
```

```

spread(study_week, butyrate_mean) %>%
mutate(delta_butyrate = week1 - week3) #calculate difference in butyrate

pro_wide <- all_data_wkly %>%
  rename_all(tolower) %>%
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, propionate_mean) %>%
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes") %>% #keep only weeks 1 and 3
  spread(study_week, propionate_mean) %>%
  mutate(delta_propionate = week1 - week3) #calculate difference in propionate

delta_but_pro_wide <- inner_join(x = but_wide, y = pro_wide,
                                by = c("participant_id",
                                      "semester", "supplement_consumed", "frequency")) %>%
  select(-starts_with("week"), -starts_with("quantity_compliant"))

incbut_broad_delta_wide <- delta_but_pro_wide %>%
  filter(delta_butyrate > 0) #subset for only increased butyrate

decpro_broad_delta_wide <- delta_but_pro_wide %>%
  filter(delta_propionate < 0) #subset for only increased butyrate

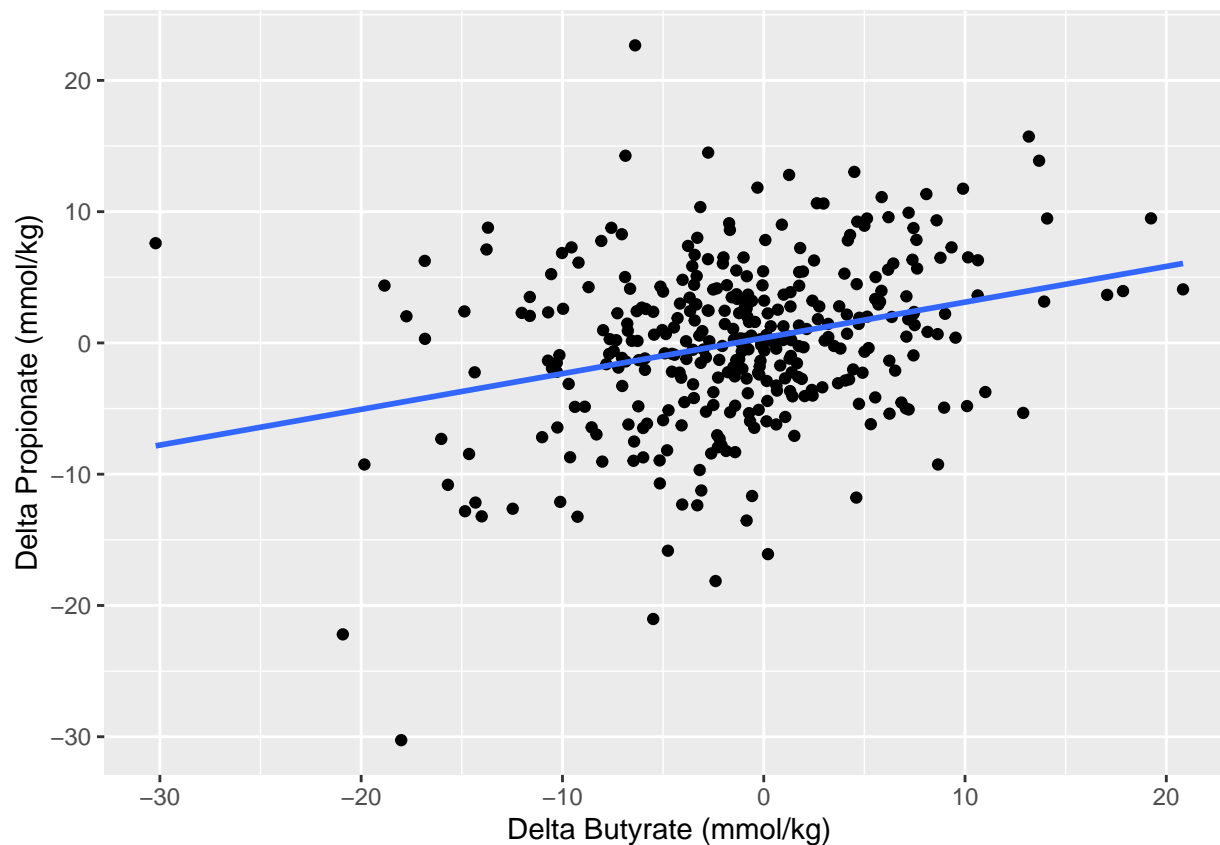
###Analysis #####Plot

linear_correlation_plot<-delta_but_pro_wide %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
              se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(linear_correlation_plot)

```

```
## Warning: Removed 130 rows containing non-finite values (stat_smooth).
```

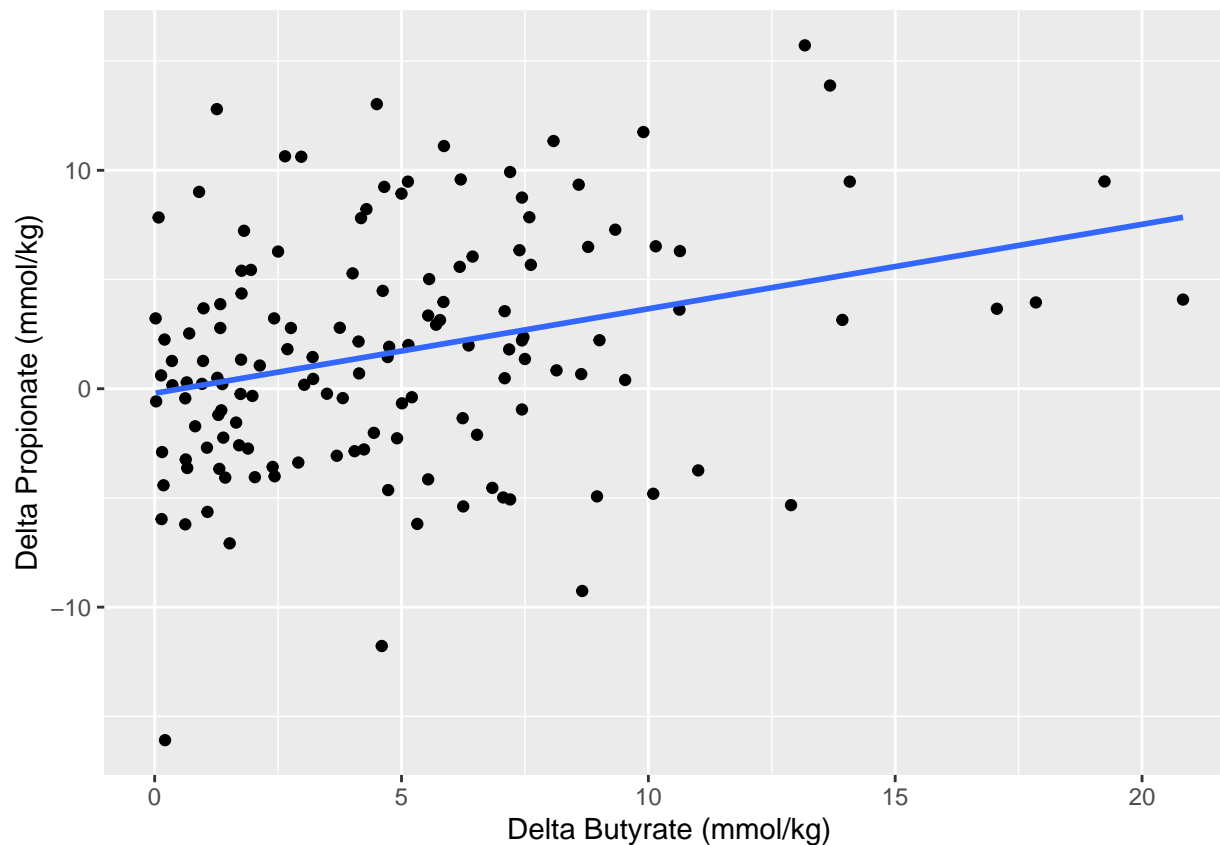
```
## Warning: Removed 130 rows containing missing values (geom_point).
```



```
incbut_broad_plot<-incbut_broad_delta_wide %>%  
  ggplot(aes(x = delta_butyrate,  
             y = delta_propionate)) +  
  geom_point() + #puts data points to match x and y coordinates  
  geom_smooth(method = "lm", #used to create a linear best fit line  
             se = FALSE) + #hides confidence interval around line  
  xlab("Delta Butyrate (mmol/kg)") +  
  ylab("Delta Propionate (mmol/kg)")  
print(incbut_broad_plot)
```

```
## Warning: Removed 17 rows containing non-finite values (stat_smooth).
```

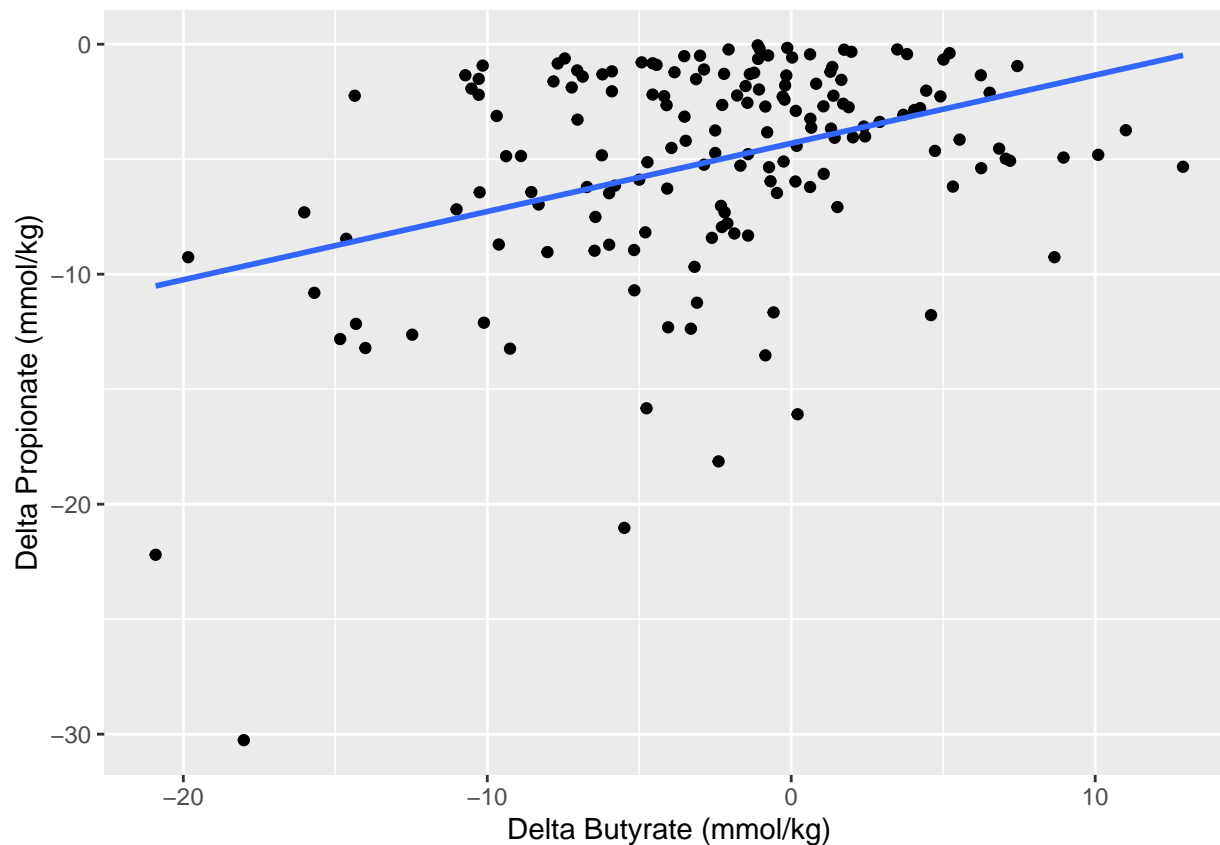
```
## Warning: Removed 17 rows containing missing values (geom_point).
```



```
decpro_broad_plot<-decpro_broad_delta_wide %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(decpro_broad_plot)
```

```
## Warning: Removed 2 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 2 rows containing missing values (geom_point).
```



#### Assumptions

```
nrow(delta_but_pro_wide) # sample size
```

```
## [1] 481
```

```
shapiro.test(delta_but_pro_wide$delta_butyrate) #call column of df with values = vector
```

```
##
## Shapiro-Wilk normality test
##
## data: delta_but_pro_wide$delta_butyrate
## W = 0.98552, p-value = 0.0006516
```

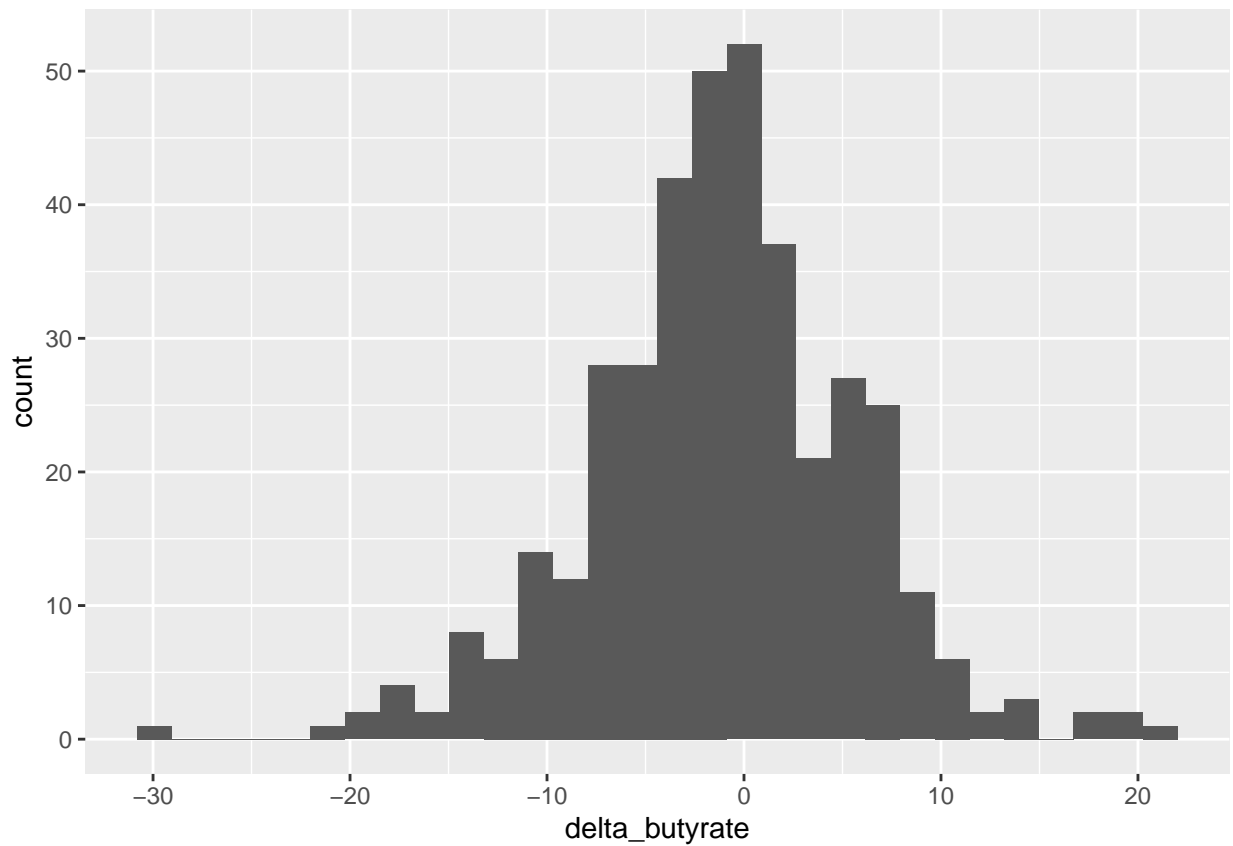
```
shapiro.test(delta_but_pro_wide$delta_propionate) #call column of df with values = vector
```

```
##
## Shapiro-Wilk normality test
##
## data: delta_but_pro_wide$delta_propionate
## W = 0.97482, p-value = 7.065e-06
```

```
ggplot(delta_but_pro_wide, aes(x=delta_butyrate)) +
  geom_histogram() #histograms show the number of data points (count) at each value
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

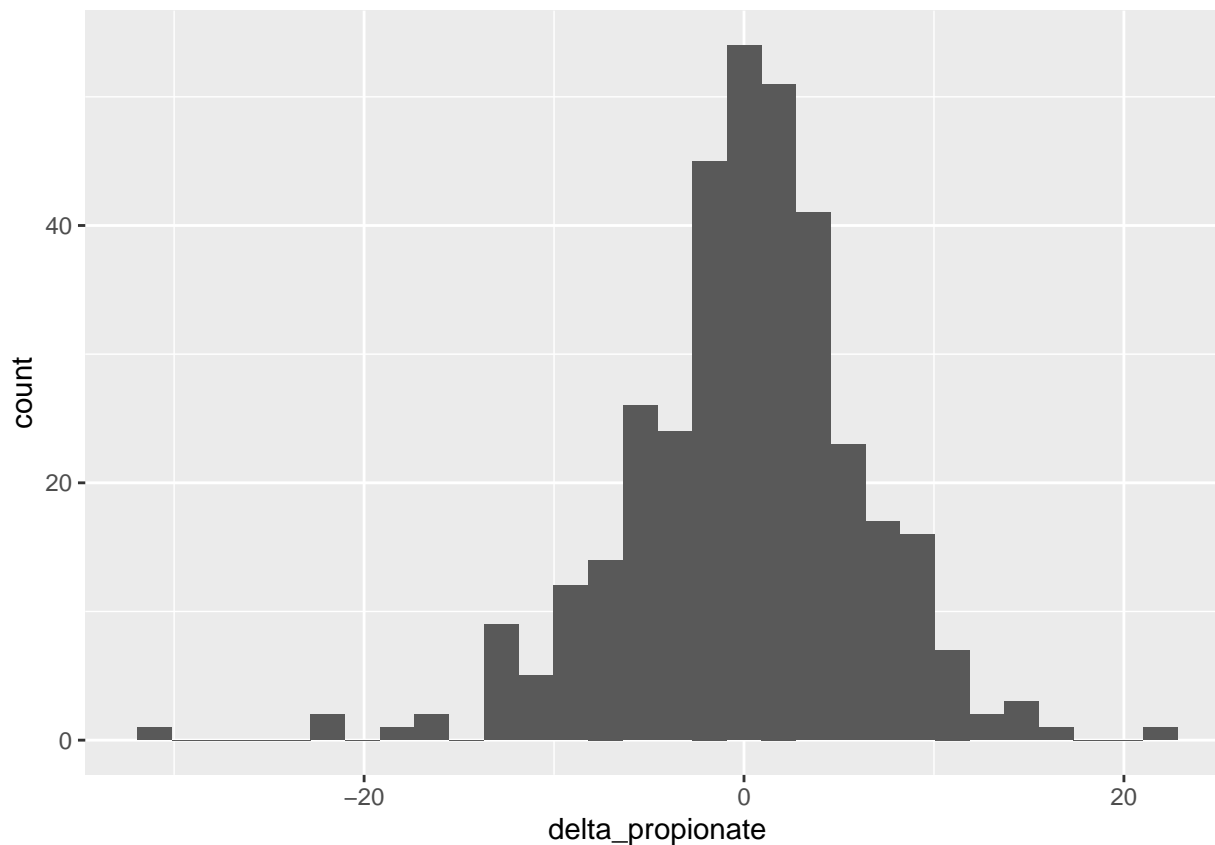
```
## Warning: Removed 94 rows containing non-finite values (stat_bin).
```



```
ggplot(delta_but_pro_wide, aes(x=delta_propionate)) +  
  geom_histogram() #histograms show the number of data points (count) at each value
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

```
## Warning: Removed 124 rows containing non-finite values (stat_bin).
```



####Test

```
all_data_lm <- delta_but_pro_wide %>%
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship
summary(all_data_lm)
```

```
##
## Call:
## lm(formula = delta_butyrate ~ delta_propionate, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -31.3254  -3.6260   0.3011   3.8289  20.8232
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)   -1.27816    0.34647  -3.689 0.000261 ***
## delta_propionate  0.31494    0.05505   5.721 2.28e-08 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 6.491 on 349 degrees of freedom
## (130 observations deleted due to missingness)
## Multiple R-squared:  0.08575,    Adjusted R-squared:  0.08313
## F-statistic: 32.73 on 1 and 349 DF,  p-value: 2.277e-08
```

*#The p-value is low at 2.8e-8 indicating significance, but the R2 value is .08 which is low meaning tha*

```
incbut_broad_test <- incbut_broad_delta_wide %>%  
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship  
summary(incbut_broad_test) #view results
```

```
##  
## Call:  
## lm(formula = delta_butyrate ~ delta_propionate, data = .)  
##  
## Residuals:  
##      Min       1Q   Median       3Q      Max   
## -6.2408 -2.9809 -0.8744  2.1977 15.3592   
##  
## Coefficients:  
##              Estimate Std. Error t value Pr(>|t|)      
## (Intercept)      4.5485     0.3534  12.871 < 2e-16 ***  
## delta_propionate  0.2261     0.0624   3.623  0.00041 ***  
## ---  
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1  
##  
## Residual standard error: 3.974 on 137 degrees of freedom  
## (17 observations deleted due to missingness)  
## Multiple R-squared:  0.08742,    Adjusted R-squared:  0.08076   
## F-statistic: 13.12 on 1 and 137 DF,  p-value: 0.0004098
```

```
#p-value = 0.01729  
#Adjusted R-squared = 0.02205
```

```
decpro_broad_test <- decpro_broad_delta_wide %>%  
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship  
summary(decpro_broad_test) #view results
```

```
##  
## Call:  
## lm(formula = delta_butyrate ~ delta_propionate, data = .)  
##  
## Residuals:  
##      Min       1Q   Median       3Q      Max   
## -15.2399 -3.4067  0.1421  3.1362 15.5741   
##  
## Coefficients:  
##              Estimate Std. Error t value Pr(>|t|)      
## (Intercept)     -0.08565     0.64929  -0.132   0.895      
## delta_propionate  0.48752     0.09403   5.185 6.49e-07 ***  
## ---  
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1  
##  
## Residual standard error: 5.597 on 159 degrees of freedom  
## (2 observations deleted due to missingness)  
## Multiple R-squared:  0.1446, Adjusted R-squared:  0.1392   
## F-statistic: 26.88 on 1 and 159 DF,  p-value: 6.491e-07
```



```
#p-value = 0.05918
#Adjusted R-squared = 0.01359
```

```
#By Supplement ##BRMPS ###Tidy Data
```

```
BRMPS_delta_wide <- delta_but_pro_wide %>%
  filter(supplement_consumed == "BRMPS", frequency == "1xdaily") #subset for BRMPS 1x
```

```
inc_BRMPS_delta_wide <- delta_but_pro_wide %>%
  filter(supplement_consumed == "BRMPS", frequency == "1xdaily", delta_butyrate > 0) #subset for only i
```

```
dec_BRMPS_delta_wide <- delta_but_pro_wide %>%
  filter(supplement_consumed == "BRMPS", frequency == "1xdaily", delta_propionate < 0) #subset for only
```

```
###Analysis #####Assumptions
```

```
# check sample size
BRMPS_delta_wide %>%
  summarise(sample_size = n()) #use summarise() and n() to get counts of ppl
```

```
## # A tibble: 1 x 1
##   sample_size
##       <int>
## 1         66
```

```
#n=66
```

```
inc_BRMPS_delta_wide %>%
  summarise(sample_size = n()) #use summarise() and n() to get counts of ppl
```

```
## # A tibble: 1 x 1
##   sample_size
##       <int>
## 1         25
```

```
#n=39
```

```
dec_BRMPS_delta_wide %>%
  summarise(sample_size = n()) #use summarise() and n() to get counts of ppl
```

```
## # A tibble: 1 x 1
##   sample_size
##       <int>
## 1         26
```

```
#n=38
```

```
# normality
shapiro.test(BRMPS_delta_wide$delta_butyrate) #p-value = 0.5985
```

```
##
## Shapiro-Wilk normality test
##
## data: BRMPS_delta_wide$delta_butyrate
## W = 0.98444, p-value = 0.5985
```

```
shapiro.test(BRMPS_delta_wide$delta_propionate) #p-value = 0.8763
```

```
##
## Shapiro-Wilk normality test
##
## data: BRMPS_delta_wide$delta_propionate
## W = 0.98977, p-value = 0.8763
```

```
shapiro.test(inc_BRMPS_delta_wide$delta_butyrate) #p-value = 0.00022222
```

```
##
## Shapiro-Wilk normality test
##
## data: inc_BRMPS_delta_wide$delta_butyrate
## W = 0.88399, p-value = 0.008361
```

```
shapiro.test(inc_BRMPS_delta_wide$delta_propionate) #p-value = 0.8143
```

```
##
## Shapiro-Wilk normality test
##
## data: inc_BRMPS_delta_wide$delta_propionate
## W = 0.9799, p-value = 0.8831
```

```
shapiro.test(dec_BRMPS_delta_wide$delta_butyrate) #p-value = 0.7484
```

```
##
## Shapiro-Wilk normality test
##
## data: dec_BRMPS_delta_wide$delta_butyrate
## W = 0.97632, p-value = 0.7878
```

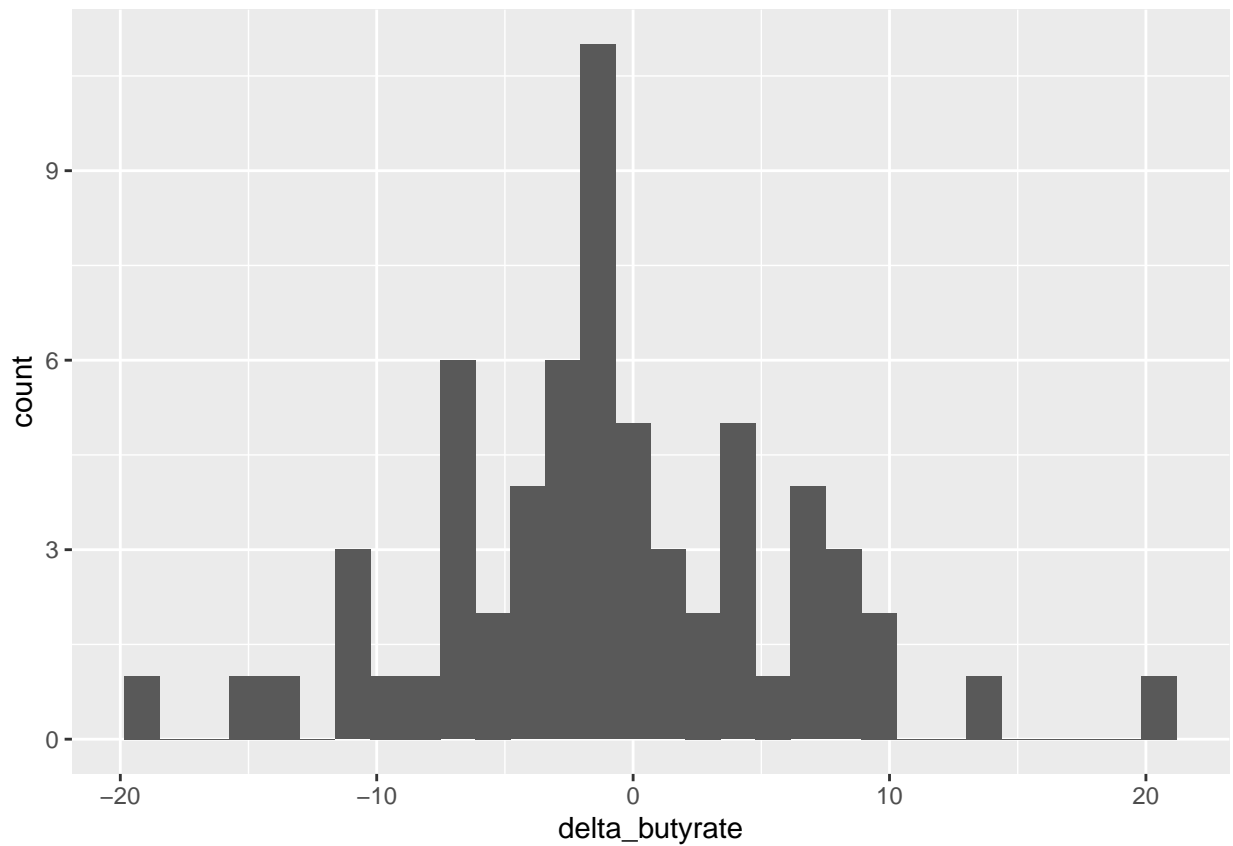
```
shapiro.test(dec_BRMPS_delta_wide$delta_propionate) #p-value = 0.002383
```

```
##
## Shapiro-Wilk normality test
##
## data: dec_BRMPS_delta_wide$delta_propionate
## W = 0.91161, p-value = 0.0287
```

```
# histogram
ggplot(BRMPS_delta_wide, aes(x=delta_butyrate)) +
  geom_histogram() #histogram for delta butyrate = very normal
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

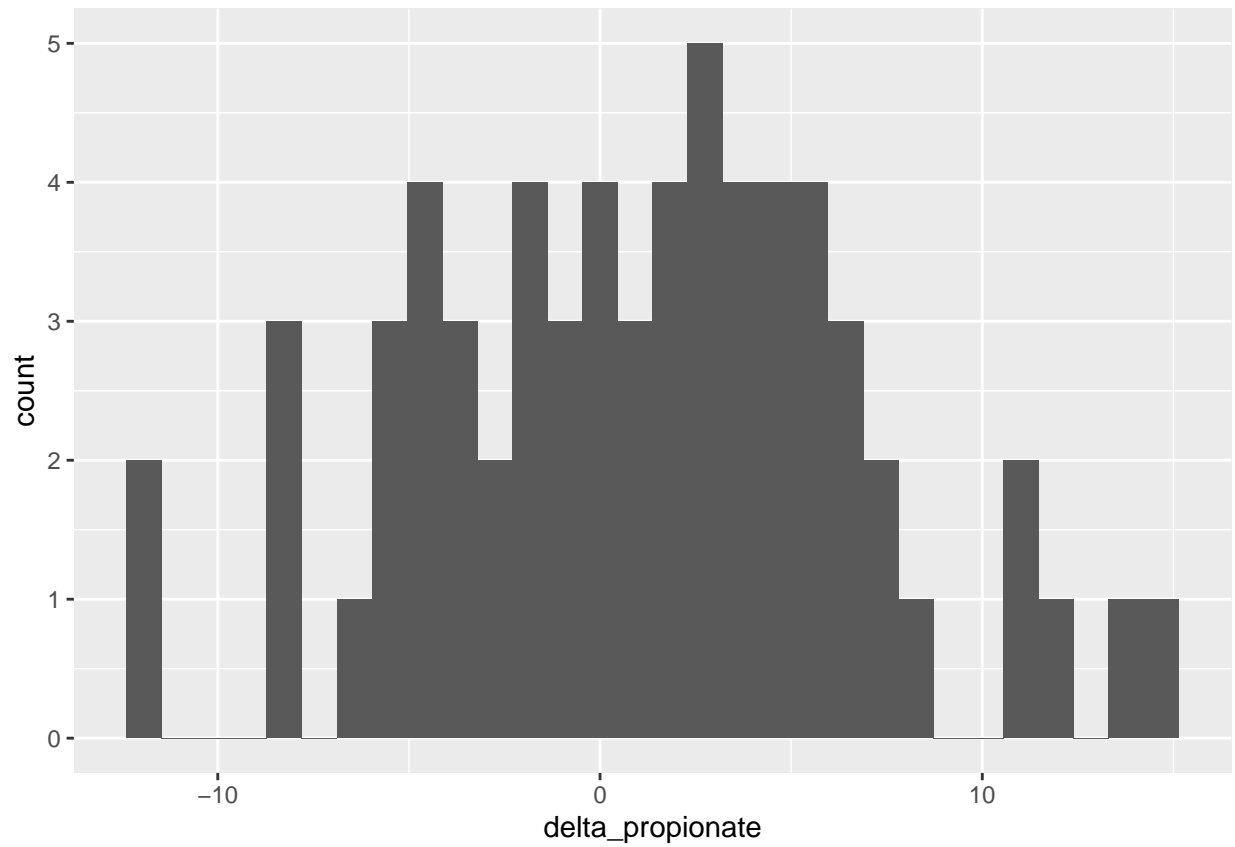
```
## Warning: Removed 2 rows containing non-finite values (stat_bin).
```



```
ggplot(BRMPS_delta_wide, aes(x=delta_propionate)) +  
  geom_histogram() #histogram for delta propionate = roughly normal
```

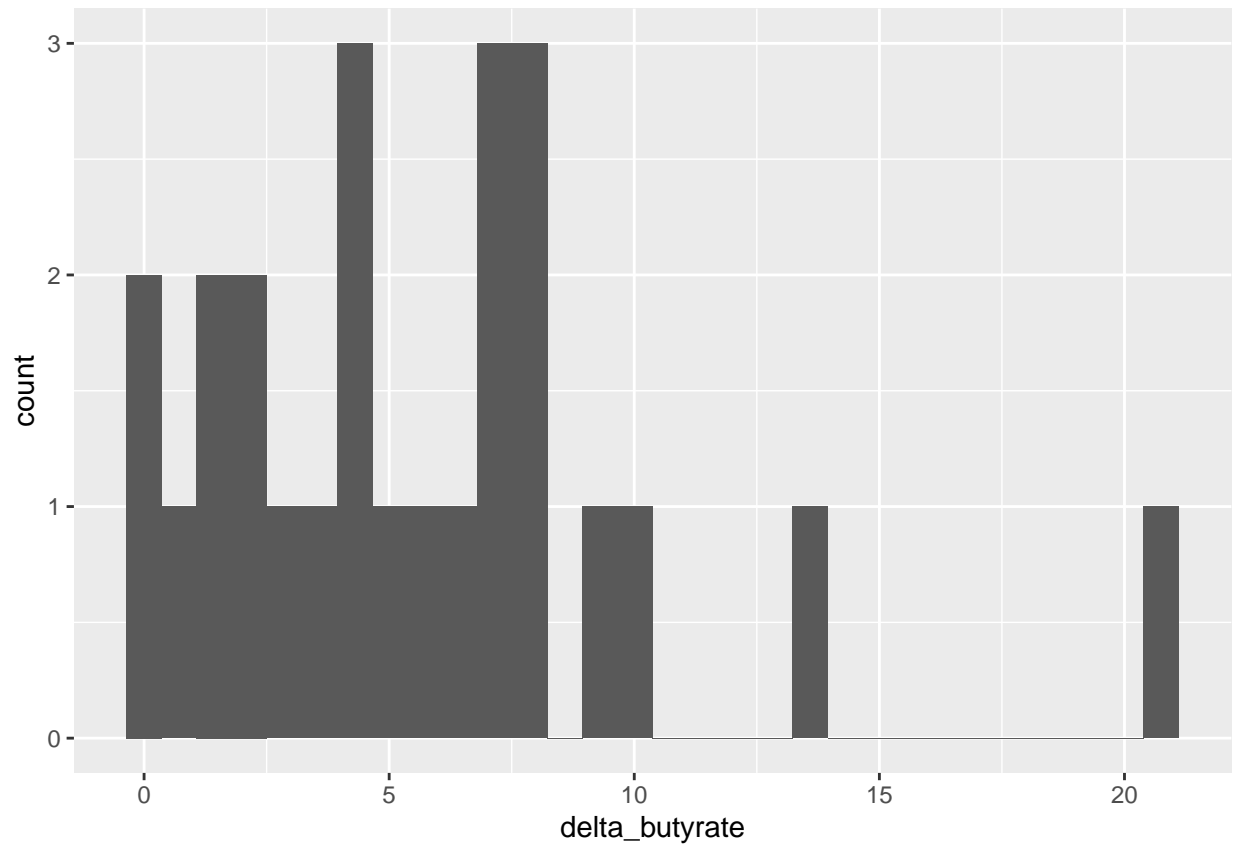
```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

```
## Warning: Removed 2 rows containing non-finite values (stat_bin).
```



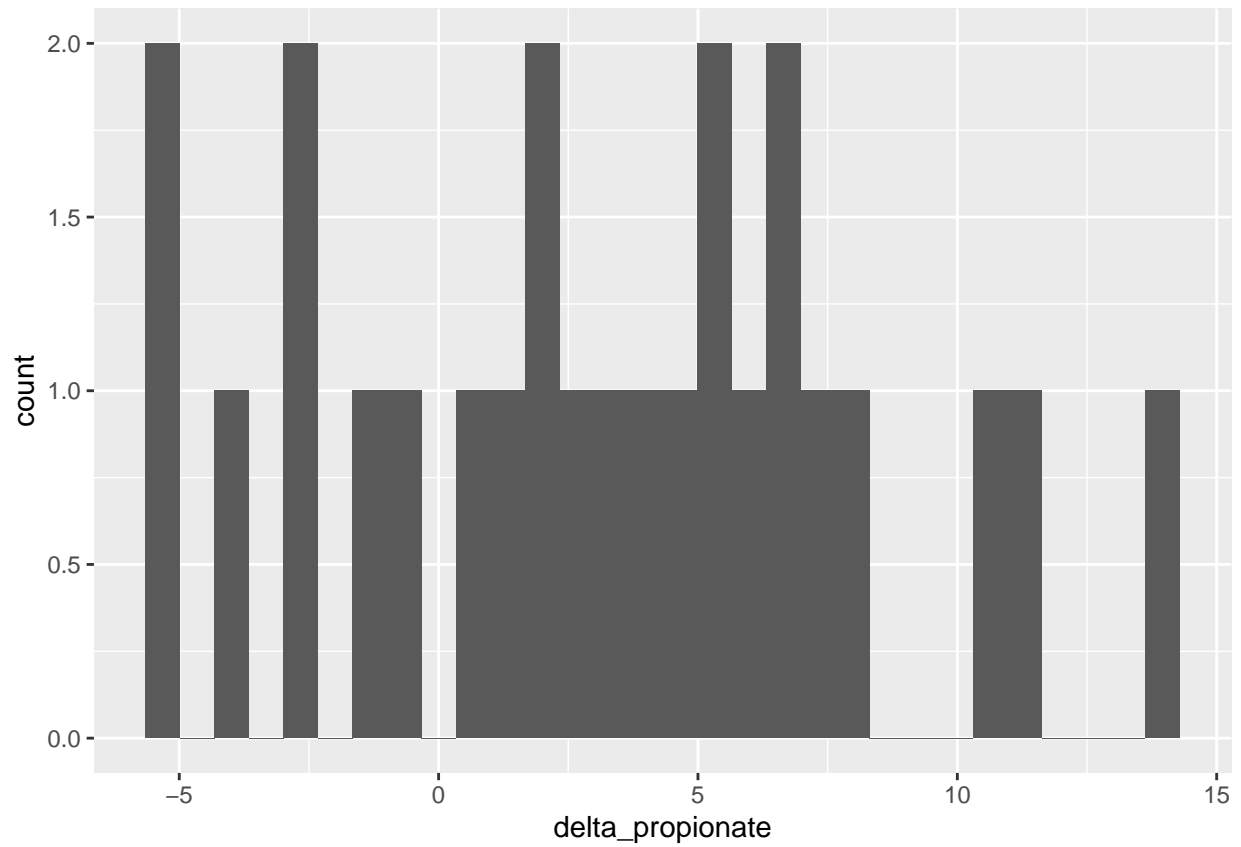
```
ggplot(inc_BRMPS_delta_wide, aes(x=delta_butyrate)) +  
  geom_histogram() #histogram for delta butyrate = very skewed right
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



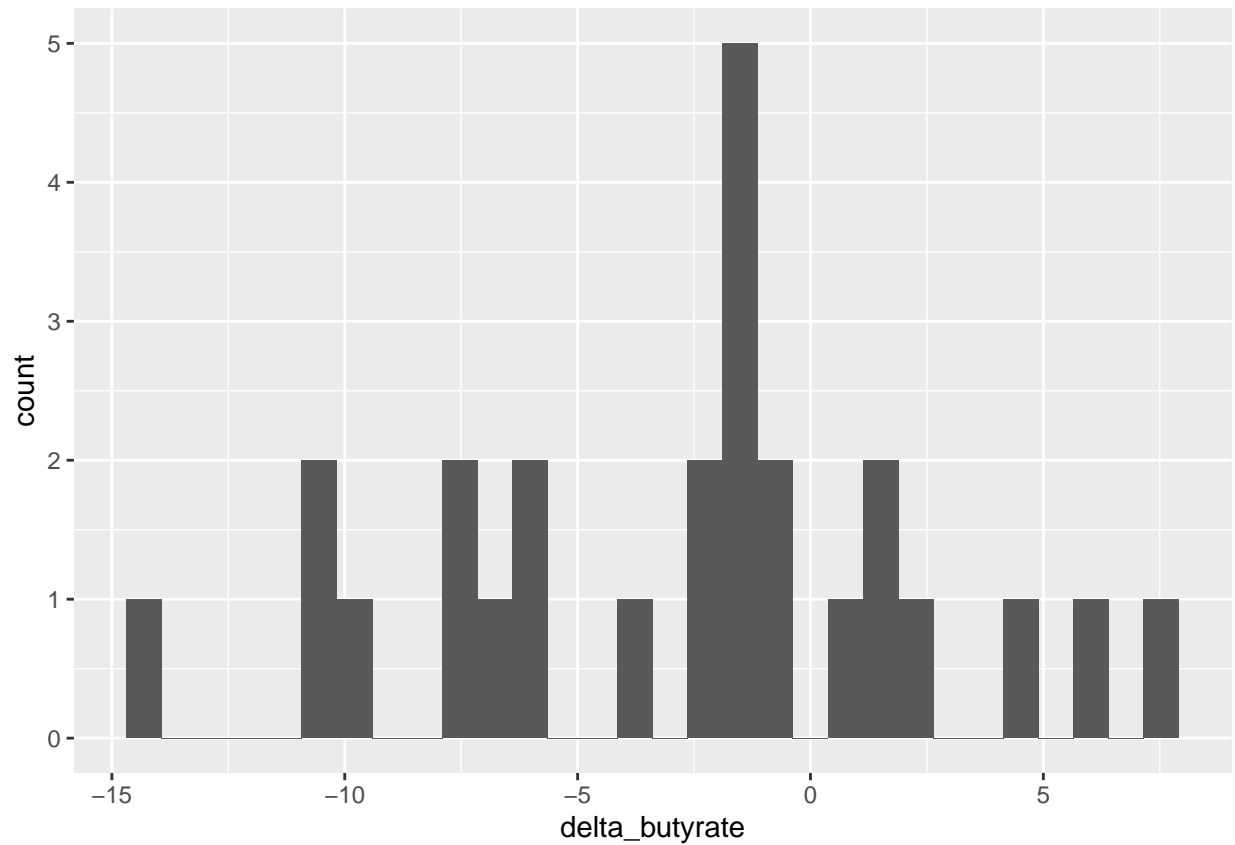
```
ggplot(inc_BRMPS_delta_wide, aes(x=delta_propionate)) +  
  geom_histogram() #histogram for delta propionate = roughly normal
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



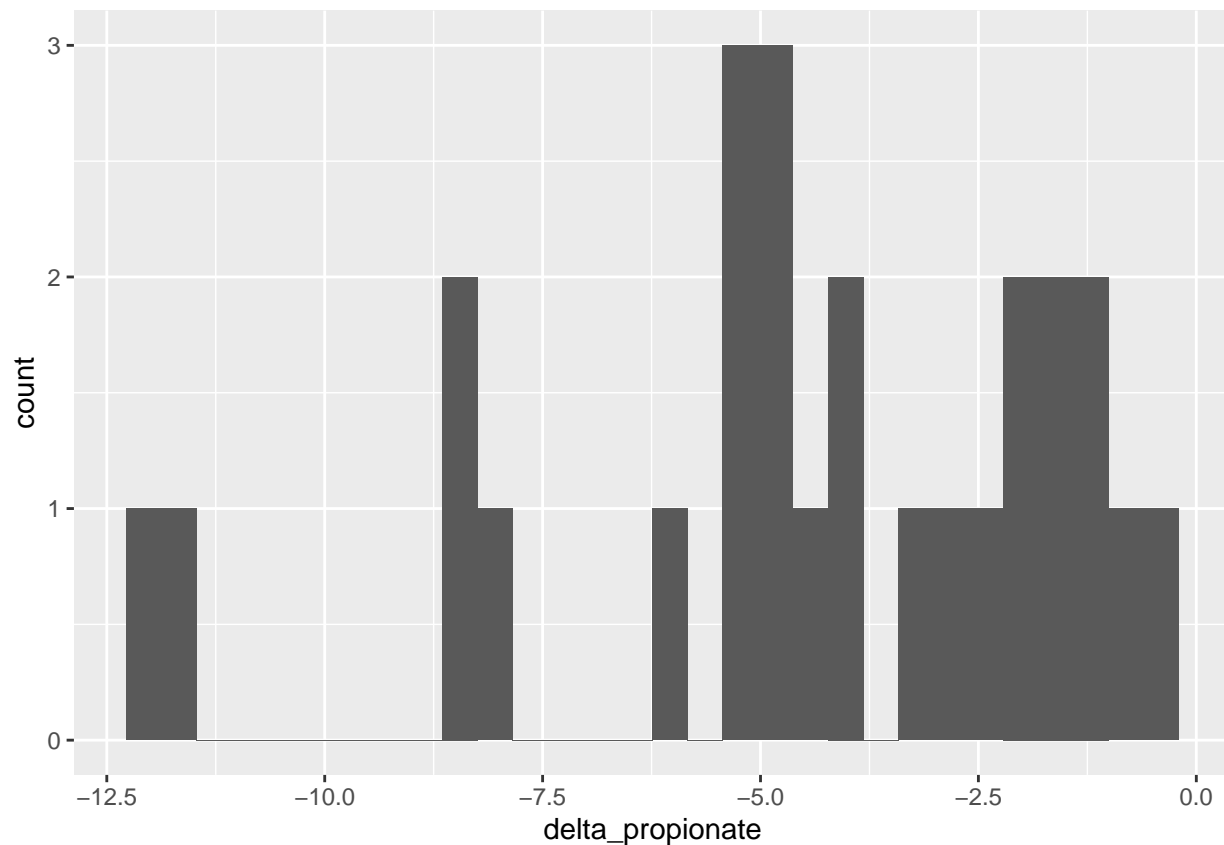
```
ggplot(dec_BRMPs_delta_wide, aes(x=delta_butyrate)) +  
  geom_histogram() #histogram for delta butyrate = roughly normal
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



```
ggplot(dec_BRMPs_delta_wide, aes(x=delta_propionate)) +  
  geom_histogram() #histogram for delta propionate = skewed left
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



```
# variances
var.test(x = BRMPS_delta_wide$delta_butyrate,
         y = BRMPS_delta_wide$delta_propionate,
         alternative = "two.sided") # p-value = 0.1289
```

```
##
## F test to compare two variances
##
## data: BRMPS_delta_wide$delta_butyrate and BRMPS_delta_wide$delta_propionate
## F = 1.47, num df = 63, denom df = 63, p-value = 0.1289
## alternative hypothesis: true ratio of variances is not equal to 1
## 95 percent confidence interval:
## 0.8930754 2.4196868
## sample estimates:
## ratio of variances
## 1.470021
```

```
var.test(x = inc_BRMPS_delta_wide$delta_butyrate,
         y = inc_BRMPS_delta_wide$delta_propionate,
         alternative = "two.sided") # p-value = 0.1072
```

```
##
## F test to compare two variances
##
## data: inc_BRMPS_delta_wide$delta_butyrate and inc_BRMPS_delta_wide$delta_propionate
```



```
## F = 0.83061, num df = 24, denom df = 24, p-value = 0.653
## alternative hypothesis: true ratio of variances is not equal to 1
## 95 percent confidence interval:
## 0.366024 1.884884
## sample estimates:
## ratio of variances
## 0.83061

var.test(x = dec_BRMPS_delta_wide$delta_butyrate,
         y = dec_BRMPS_delta_wide$delta_propionate,
         alternative = "two.sided") # p-value = 2.42e-05 UNEQUAL VARIANCES

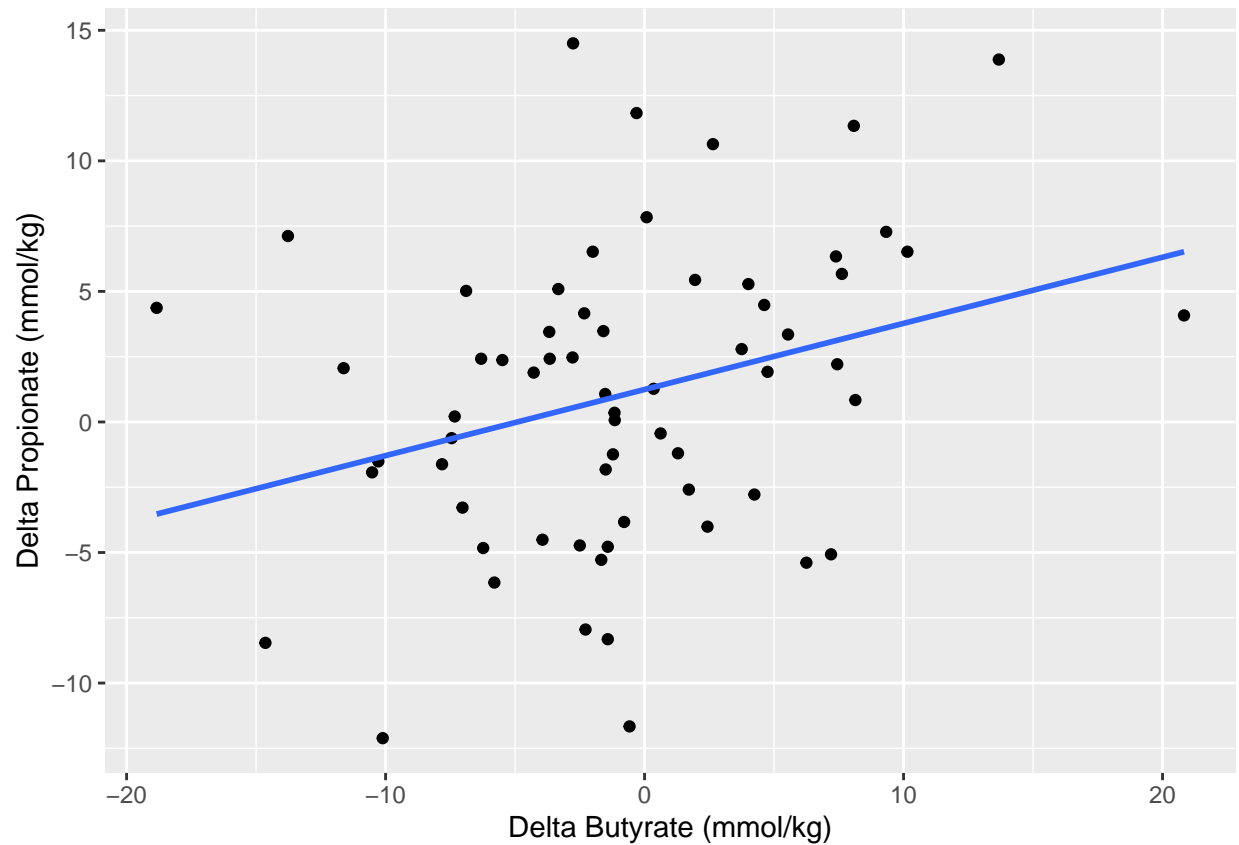
##
## F test to compare two variances
##
## data: dec_BRMPS_delta_wide$delta_butyrate and dec_BRMPS_delta_wide$delta_propionate
## F = 2.9015, num df = 25, denom df = 25, p-value = 0.009921
## alternative hypothesis: true ratio of variances is not equal to 1
## 95 percent confidence interval:
## 1.300965 6.471320
## sample estimates:
## ratio of variances
## 2.901544

####Plot

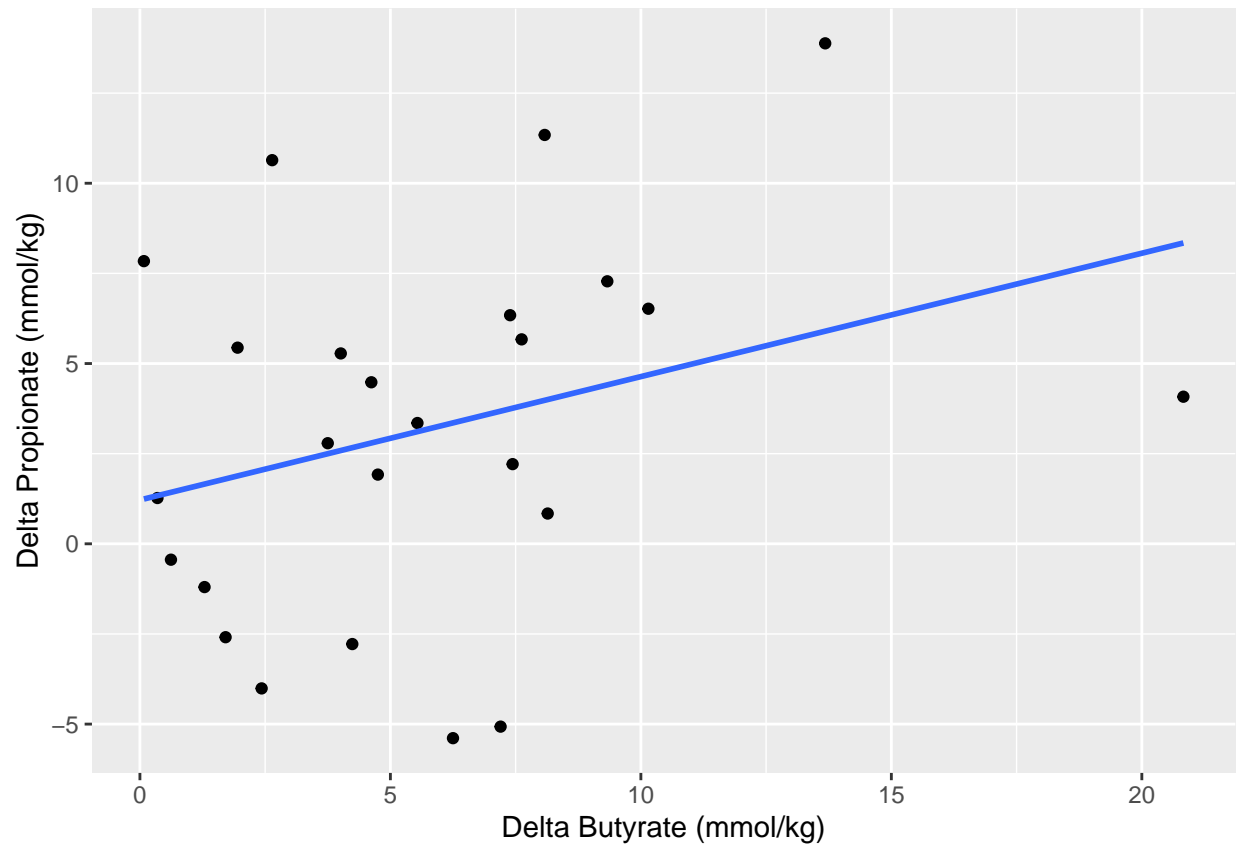
BRMPS_plot<-BRMPS_delta_wide %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(BRMPS_plot)

## Warning: Removed 2 rows containing non-finite values (stat_smooth).

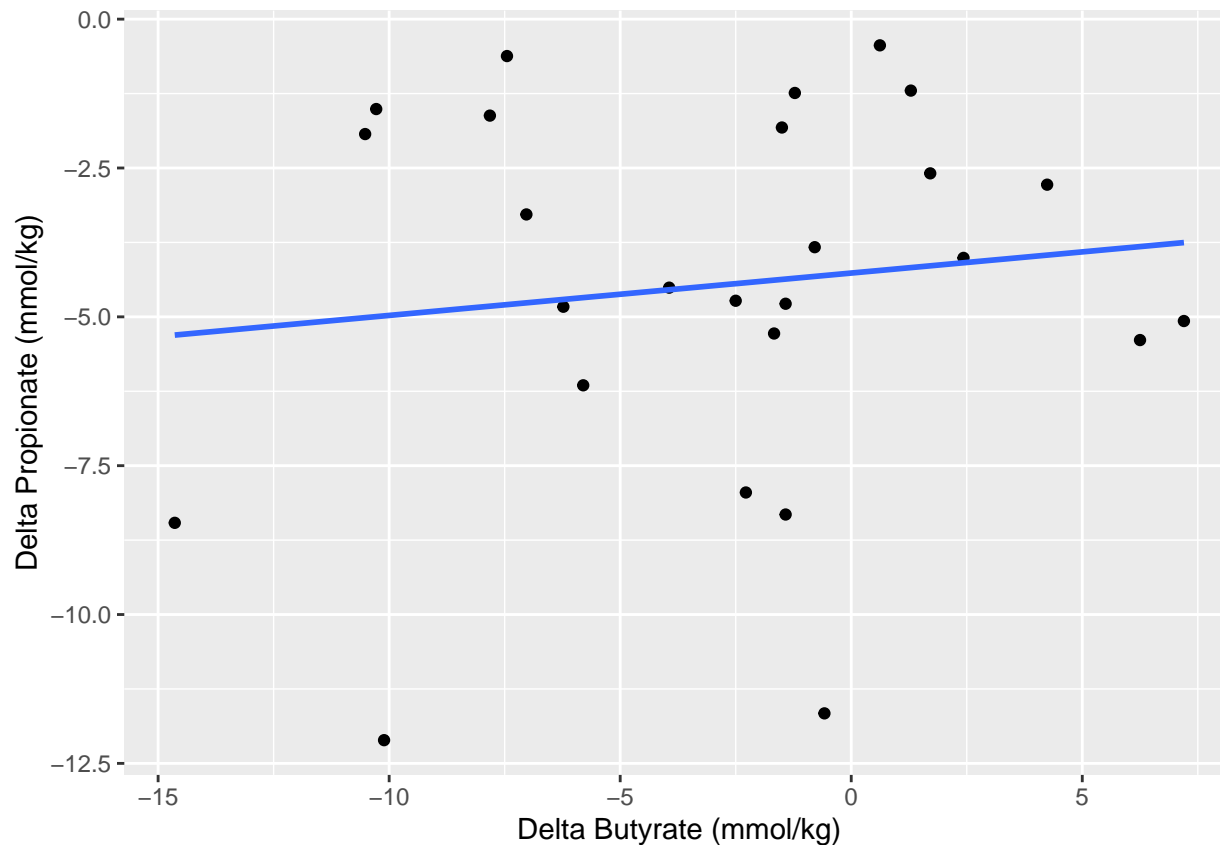
## Warning: Removed 2 rows containing missing values (geom_point).
```



```
inc_BRMPS_plot<-inc_BRMPS_delta_wide %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(inc_BRMPS_plot)
```



```
dec_BRMPs_plot<-dec_BRMPs_delta_wide %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(dec_BRMPs_plot)
```



####Test

```
BRMPS_test <- BRMPS_delta_wide %>%
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship
summary(BRMPS_test) #view results
```

```
##
## Call:
## lm(formula = delta_butyrate ~ delta_propionate, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -19.2410  -3.8181   0.2266   4.3802  20.5369
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    -1.2259     0.8479  -1.446   0.1533
## delta_propionate  0.3723     0.1465   2.540   0.0136 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 6.675 on 62 degrees of freedom
## (2 observations deleted due to missingness)
## Multiple R-squared:  0.09428,    Adjusted R-squared:  0.07967
## F-statistic: 6.454 on 1 and 62 DF,  p-value: 0.01359
```

```
#p-value = 0.01359
#Adjusted R-squared = 0.07967
```

```
inc_BRMPS_test <- inc_BRMPS_delta_wide %>%
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship
summary(inc_BRMPS_test) #view results
```

```
##
## Call:
## lm(formula = delta_butyrate ~ delta_propionate, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -7.0060 -2.4114 -0.2698  2.4032 14.8128
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      4.8576      1.0681   4.548 0.000144 ***
## delta_propionate  0.2842      0.1806   1.574 0.129098
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.499 on 23 degrees of freedom
## Multiple R-squared:  0.09726,    Adjusted R-squared:  0.05801
## F-statistic: 2.478 on 1 and 23 DF,  p-value: 0.1291
```

```
#p-value = 0.9439
#Adjusted R-squared = -0.02689
```

```
dec_BRMPS_test <- dec_BRMPS_delta_wide %>%
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship
summary(dec_BRMPS_test) #view results
```

```
##
## Call:
## lm(formula = delta_butyrate ~ delta_propionate, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -10.990  -4.169   1.102   3.235  10.150
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)     -1.9037      1.8751  -1.015   0.320
## delta_propionate  0.2064      0.3451   0.598   0.555
##
## Residual standard error: 5.445 on 24 degrees of freedom
## Multiple R-squared:  0.01468,    Adjusted R-squared: -0.02638
## F-statistic: 0.3576 on 1 and 24 DF,  p-value: 0.5555
```

```
#p-value = 0.1572
#Adjusted R-squared = 0.02853
```

```
##LOODAT ###Tidy Data
```

```
but_wide_LOODAT <- all_data_wkly %>%  
  rename_all(tolower) %>%  
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, butyrate_mean)  
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes", frequency == "1xdaily")  
  spread(study_week, butyrate_mean) %>%  
  mutate(delta_butyrate = week3 - week1) #calculate difference in butyrate  
but_wide_LOODAT
```

```
## # A tibble: 16 x 8  
##   participant_id semester supplement_cons~ frequency quantity_compli~  
##   <chr>          <chr>      <chr>          <chr>      <chr>  
## 1 U700          Winter2~ LOODAT          1xdaily    yes  
## 2 U701          Winter2~ LOODAT          1xdaily    yes  
## 3 U706          Winter2~ LOODAT          1xdaily    yes  
## 4 U707          Winter2~ LOODAT          1xdaily    yes  
## 5 U708          Winter2~ LOODAT          1xdaily    yes  
## 6 U710          Winter2~ LOODAT          1xdaily    yes  
## 7 U711          Winter2~ LOODAT          1xdaily    yes  
## 8 U712          Winter2~ LOODAT          1xdaily    yes  
## 9 U713          Winter2~ LOODAT          1xdaily    yes  
## 10 U715         Winter2~ LOODAT          1xdaily    yes  
## 11 U716         Winter2~ LOODAT          1xdaily    yes  
## 12 U717         Winter2~ LOODAT          1xdaily    yes  
## 13 U719         Winter2~ LOODAT          1xdaily    yes  
## 14 U720         Winter2~ LOODAT          1xdaily    yes  
## 15 U721         Winter2~ LOODAT          1xdaily    yes  
## 16 U723         Winter2~ LOODAT          1xdaily    yes  
## # ... with 3 more variables: week1 <dbl>, week3 <dbl>,  
## #   delta_butyrate <dbl>
```

```
pro_wide_LOODAT <- all_data_wkly %>%  
  rename_all(tolower) %>%  
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, propionate_mean)  
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes", frequency == "1xdaily")  
  spread(study_week, propionate_mean) %>%  
  mutate(delta_propionate = week3 - week1) #calculate difference in propionate  
pro_wide_LOODAT
```

```
## # A tibble: 16 x 8  
##   participant_id semester supplement_cons~ frequency quantity_compli~  
##   <chr>          <chr>      <chr>          <chr>      <chr>  
## 1 U700          Winter2~ LOODAT          1xdaily    yes  
## 2 U701          Winter2~ LOODAT          1xdaily    yes  
## 3 U706          Winter2~ LOODAT          1xdaily    yes  
## 4 U707          Winter2~ LOODAT          1xdaily    yes  
## 5 U708          Winter2~ LOODAT          1xdaily    yes  
## 6 U710          Winter2~ LOODAT          1xdaily    yes  
## 7 U711          Winter2~ LOODAT          1xdaily    yes  
## 8 U712          Winter2~ LOODAT          1xdaily    yes  
## 9 U713          Winter2~ LOODAT          1xdaily    yes  
## 10 U715         Winter2~ LOODAT          1xdaily    yes
```

```
## 11 U716      Winter2~ LOODAT      1xdaily  yes
## 12 U717      Winter2~ LOODAT      1xdaily  yes
## 13 U719      Winter2~ LOODAT      1xdaily  yes
## 14 U720      Winter2~ LOODAT      1xdaily  yes
## 15 U721      Winter2~ LOODAT      1xdaily  yes
## 16 U723      Winter2~ LOODAT      1xdaily  yes
## # ... with 3 more variables: week1 <dbl>, week3 <dbl>,
## #   delta_propionate <dbl>
```

```
delta_but_pro_wide_LOODAT <- inner_join(x = but_wide_LOODAT, y = pro_wide_LOODAT,
  by = c("participant_id",
        "semester", "supplement_consumed")) %>%
  select(-starts_with("week"))
delta_but_pro_wide_LOODAT
```

```
## # A tibble: 16 x 9
##   participant_id semester supplement_cons~ frequency.x quantity_compli~
##   <chr>          <chr>      <chr>          <chr>      <chr>
## 1 U700      Winter2~ LOODAT      1xdaily  yes
## 2 U701      Winter2~ LOODAT      1xdaily  yes
## 3 U706      Winter2~ LOODAT      1xdaily  yes
## 4 U707      Winter2~ LOODAT      1xdaily  yes
## 5 U708      Winter2~ LOODAT      1xdaily  yes
## 6 U710      Winter2~ LOODAT      1xdaily  yes
## 7 U711      Winter2~ LOODAT      1xdaily  yes
## 8 U712      Winter2~ LOODAT      1xdaily  yes
## 9 U713      Winter2~ LOODAT      1xdaily  yes
## 10 U715     Winter2~ LOODAT      1xdaily  yes
## 11 U716     Winter2~ LOODAT      1xdaily  yes
## 12 U717     Winter2~ LOODAT      1xdaily  yes
## 13 U719     Winter2~ LOODAT      1xdaily  yes
## 14 U720     Winter2~ LOODAT      1xdaily  yes
## 15 U721     Winter2~ LOODAT      1xdaily  yes
## 16 U723     Winter2~ LOODAT      1xdaily  yes
## # ... with 4 more variables: delta_butyrate <dbl>, frequency.y <chr>,
## #   quantity_compliant.y <chr>, delta_propionate <dbl>
```

###Analysis #####Assumptions

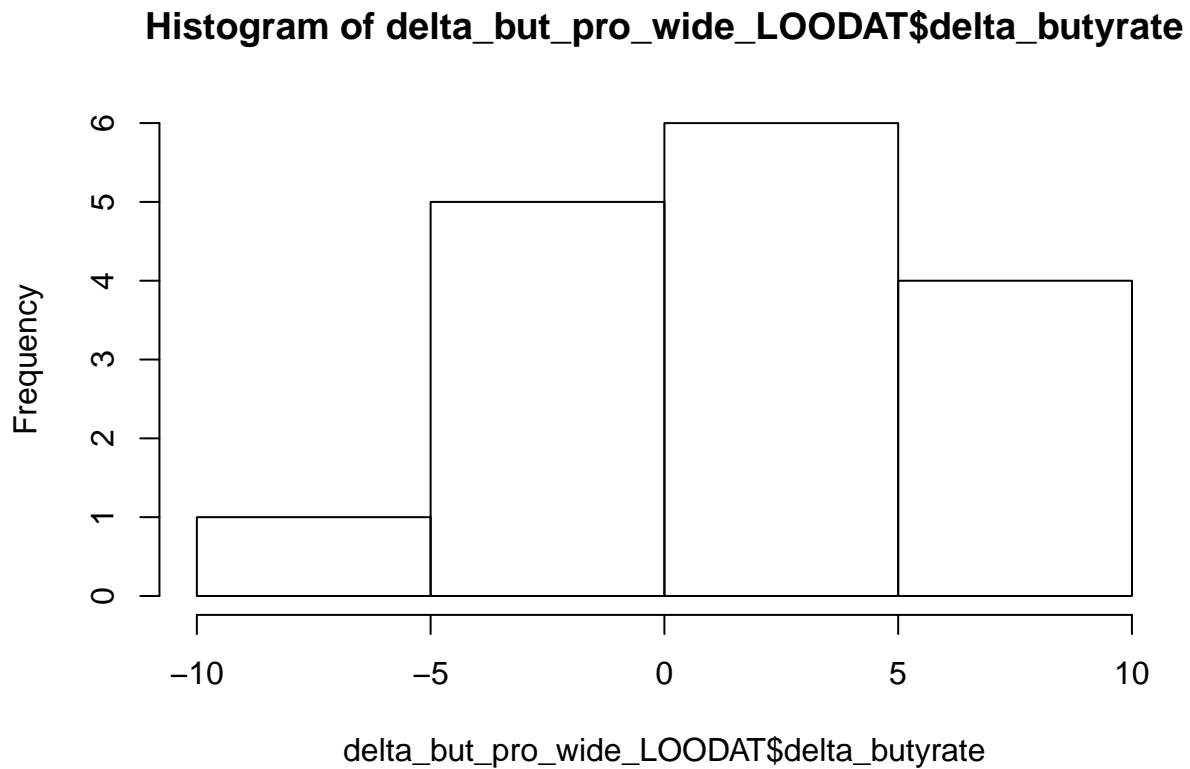
```
#Sample Size
delta_but_pro_wide_LOODAT %>%
  summarise(sample_size = n()) #n=16
```

```
## # A tibble: 1 x 1
##   sample_size
##   <int>
## 1         16
```

```
#Normality
#Butyrate:
shapiro.test(delta_but_pro_wide_LOODAT$delta_butyrate)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data: delta_but_pro_wide_LOODAT$delta_butyrate  
## W = 0.9732, p-value = 0.8874
```

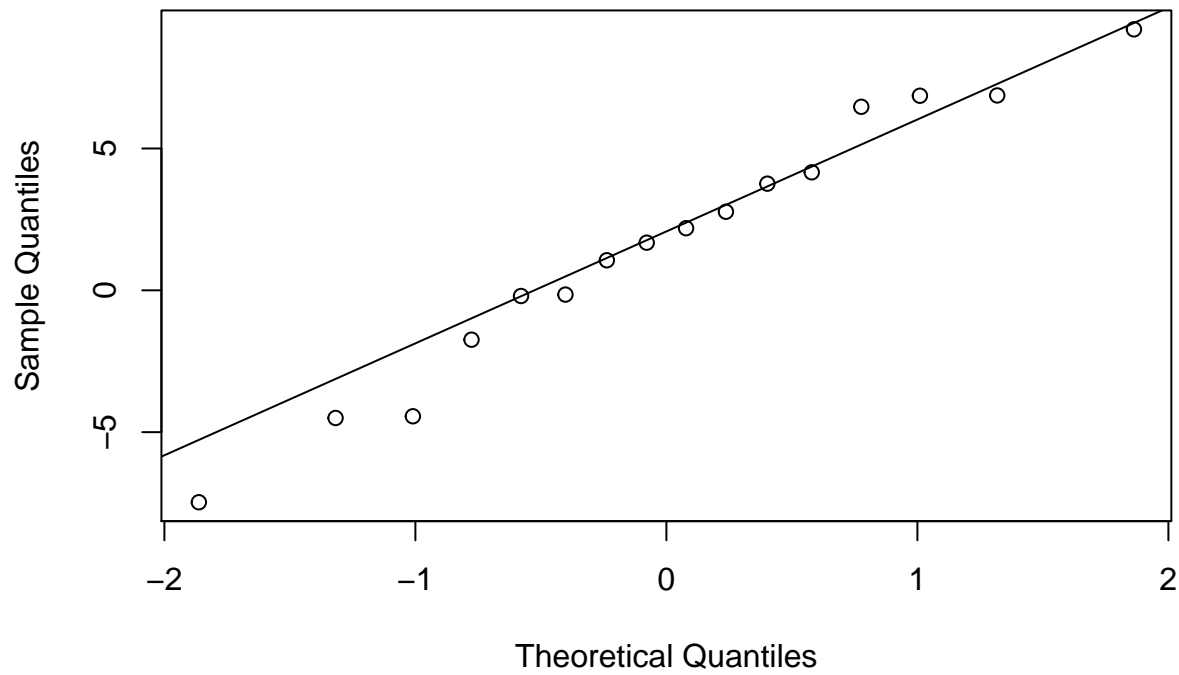
```
hist(delta_but_pro_wide_LOODAT$delta_butyrate)
```



```
qqnorm(delta_but_pro_wide_LOODAT$delta_butyrate)  
qqline(delta_but_pro_wide_LOODAT$delta_butyrate) #p=0.8874. Normality supported.
```



## Normal Q-Q Plot

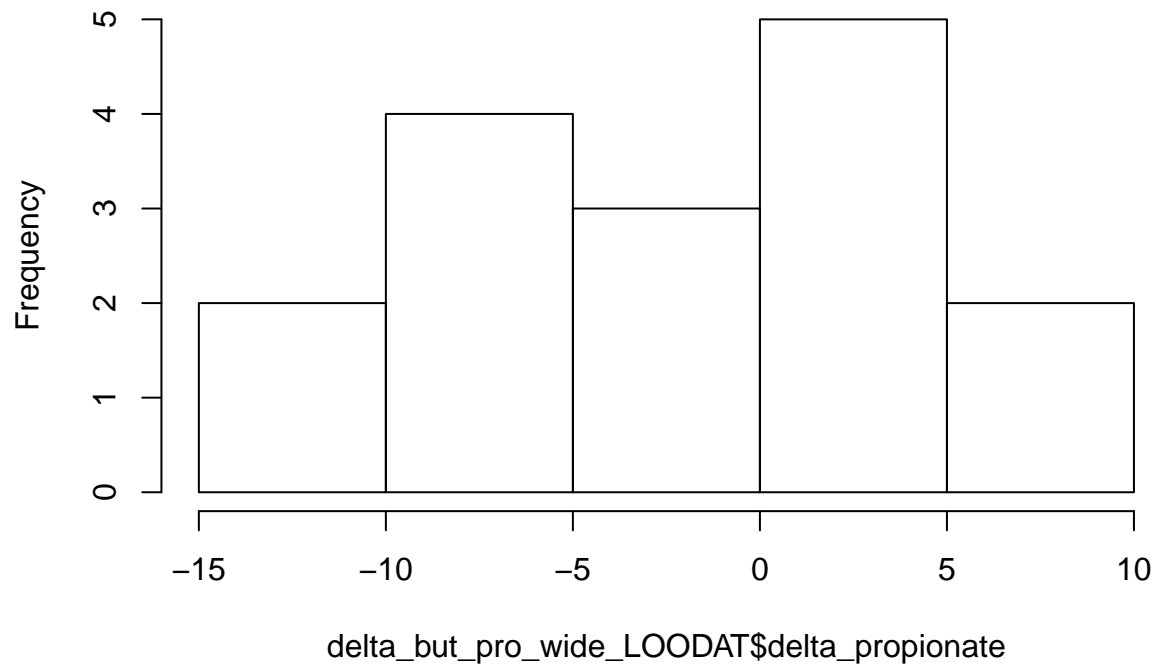


```
#Propionate:  
shapiro.test(delta_but_pro_wide_LOODAT$delta_propionate)
```

```
##  
##  Shapiro-Wilk normality test  
##  
## data:  delta_but_pro_wide_LOODAT$delta_propionate  
## W = 0.97347, p-value = 0.8912
```

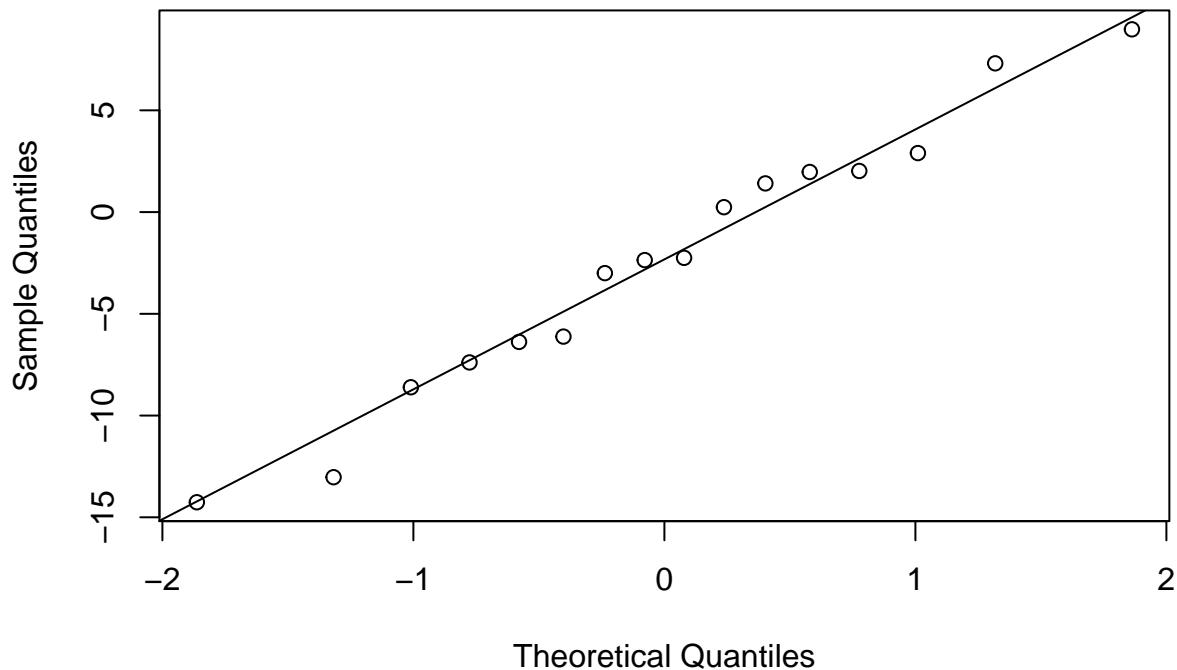
```
hist(delta_but_pro_wide_LOODAT$delta_propionate)
```

**Histogram of delta\_but\_pro\_wide\_LOODAT\$delta\_propionate**



```
qqnorm(delta_but_pro_wide_LOODAT$delta_propionate)
qqline(delta_but_pro_wide_LOODAT$delta_propionate) #p=0.8912. Normality supported
```

## Normal Q-Q Plot



```
#Equality of Variances
```

```
var.test(delta_but_pro_wide_LOODAT$delta_butyrate, delta_but_pro_wide_LOODAT$delta_propionate, alternat
```

```
##
## F test to compare two variances
##
## data: delta_but_pro_wide_LOODAT$delta_butyrate and delta_but_pro_wide_LOODAT$delta_propionate
## F = 0.49078, num df = 15, denom df = 15, p-value = 0.1796
## alternative hypothesis: true ratio of variances is not equal to 1
## 90 percent confidence interval:
##  0.2041992 1.1795684
## sample estimates:
## ratio of variances
##      0.490782
```

```
var.test(delta_but_pro_wide_LOODAT$delta_butyrate, delta_but_pro_wide_LOODAT$delta_propionate, alternat
```

```
##
## F test to compare two variances
##
## data: delta_but_pro_wide_LOODAT$delta_butyrate and delta_but_pro_wide_LOODAT$delta_propionate
## F = 0.49078, num df = 15, denom df = 15, p-value = 0.9102
## alternative hypothesis: true ratio of variances is greater than 1
## 90 percent confidence interval:
##  0.248848      Inf
```

```

## sample estimates:
## ratio of variances
##          0.490782

var.test(delta_but_pro_wide_LOODAT$delta_butyrate, delta_but_pro_wide_LOODAT$delta_propionate, alternat

##
## F test to compare two variances
##
## data:  delta_but_pro_wide_LOODAT$delta_butyrate and delta_but_pro_wide_LOODAT$delta_propionate
## F = 0.49078, num df = 15, denom df = 15, p-value = 0.08982
## alternative hypothesis: true ratio of variances is less than 1
## 90 percent confidence interval:
##  0.0000000 0.9679278
## sample estimates:
## ratio of variances
##          0.490782

#-----
#Assumptions selecting for butyrate increase:
delta_but_pro_wide_LOODAT_dbup <- delta_but_pro_wide_LOODAT %>%
  filter(delta_butyrate > 0) %>%
  summarise(sample_size = n()) #n=10
delta_but_pro_wide_LOODAT_dbup

## # A tibble: 1 x 1
##   sample_size
##       <int>
## 1         10

delta_but_pro_wide_LOODAT_dbup <- delta_but_pro_wide_LOODAT %>%
  filter(delta_butyrate > 0)

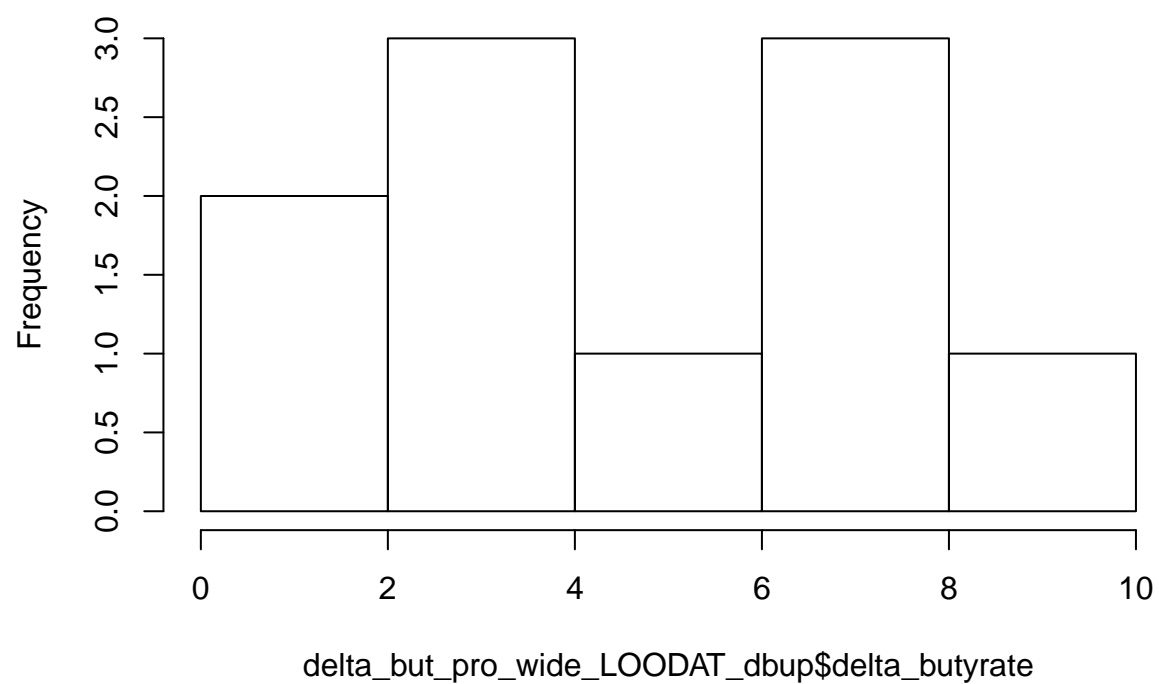
#Normality
#Butyrate:
shapiro.test(delta_but_pro_wide_LOODAT_dbup$delta_butyrate)

##
## Shapiro-Wilk normality test
##
## data:  delta_but_pro_wide_LOODAT_dbup$delta_butyrate
## W = 0.93429, p-value = 0.4914

hist(delta_but_pro_wide_LOODAT_dbup$delta_butyrate)

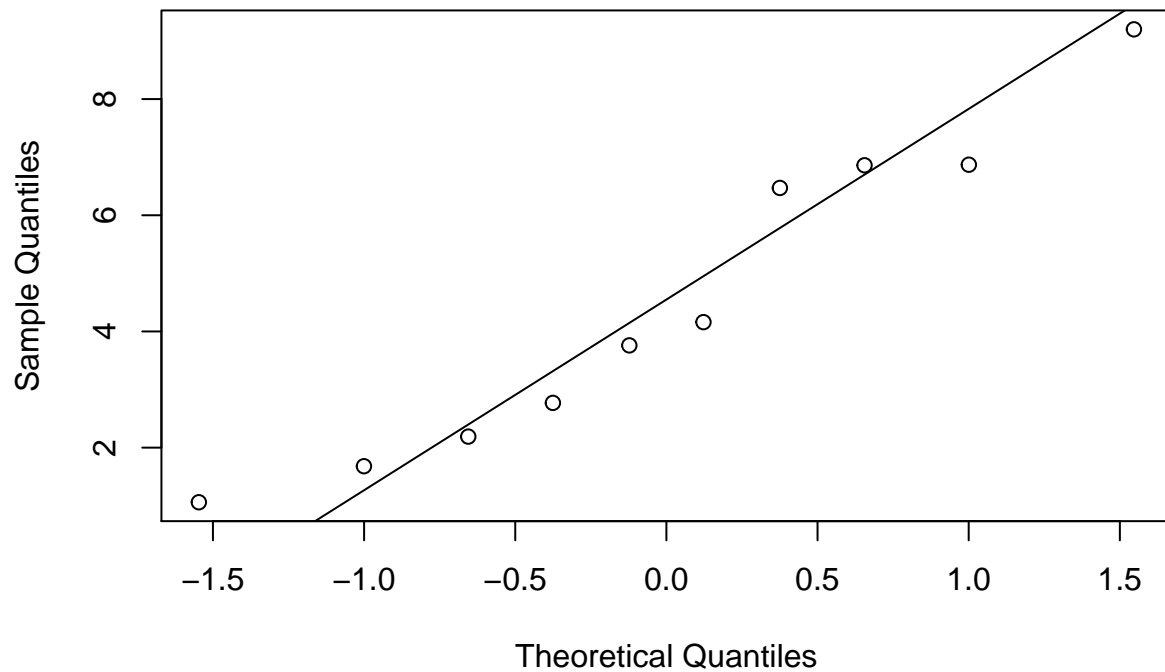
```

**Histogram of delta\_but\_pro\_wide\_LOODAT\_dbup\$delta\_butyrate**



```
qqnorm(delta_but_pro_wide_LOODAT_dbup$delta_butyrate)
qqline(delta_but_pro_wide_LOODAT_dbup$delta_butyrate) #p=0.4914. Normality supported.
```

### Normal Q-Q Plot

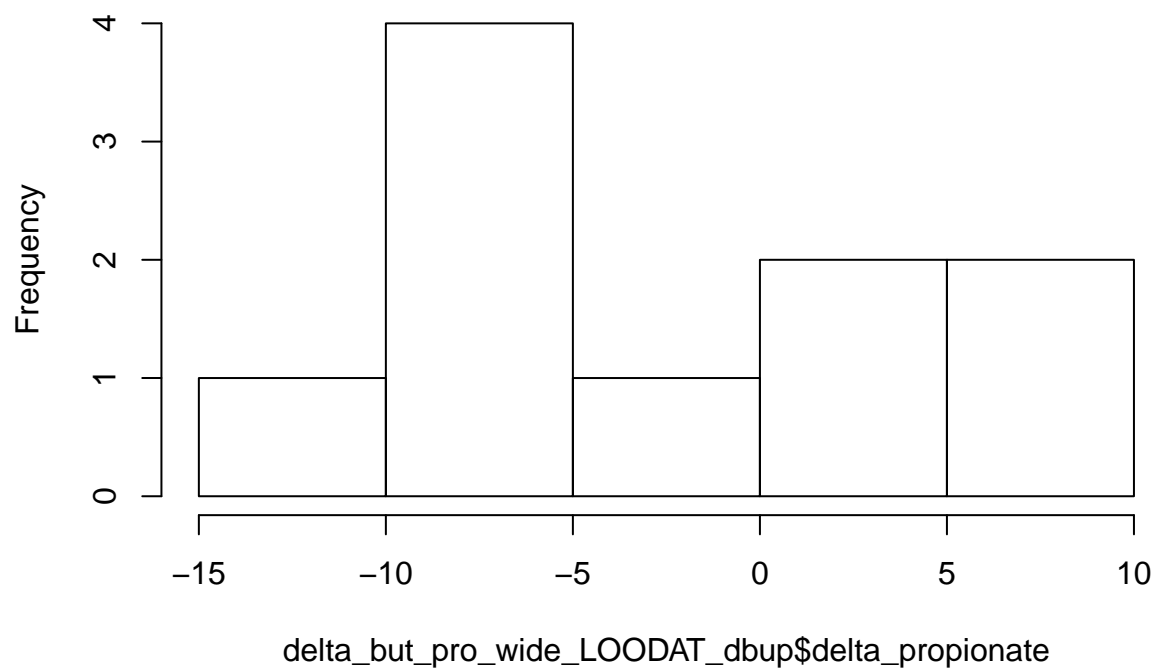


```
#Propionate:  
shapiro.test(delta_but_pro_wide_LOODAT_dbup$delta_propionate)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data: delta_but_pro_wide_LOODAT_dbup$delta_propionate  
## W = 0.95511, p-value = 0.729
```

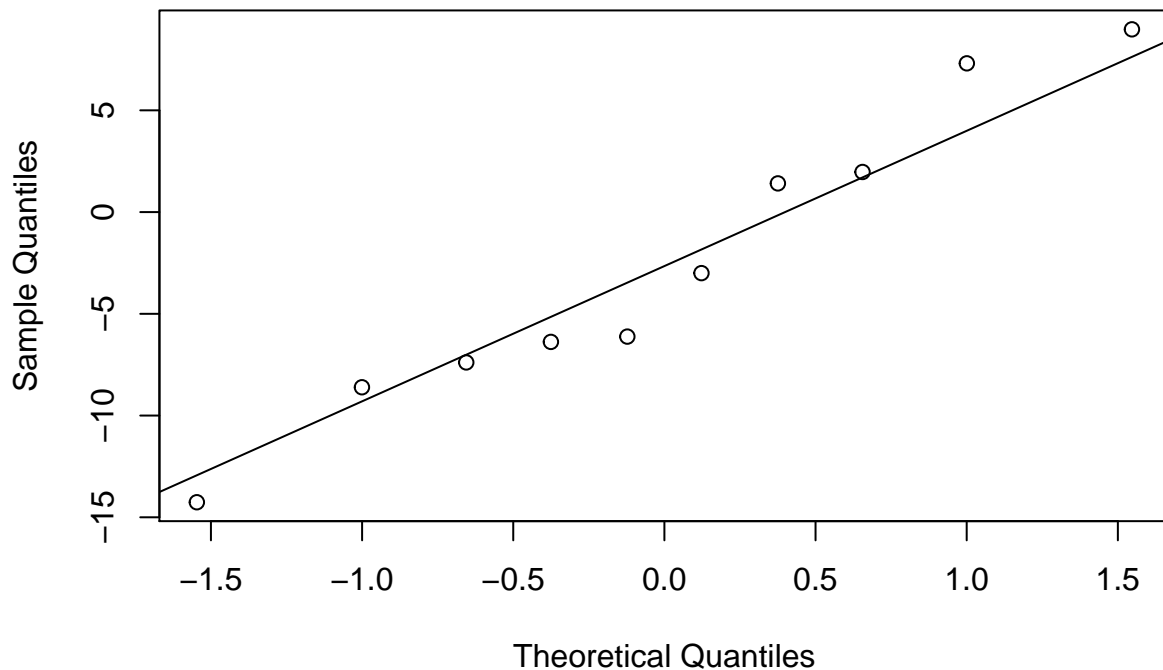
```
hist(delta_but_pro_wide_LOODAT_dbup$delta_propionate)
```

Histogram of delta\_but\_pro\_wide\_LOODAT\_dbup\$delta\_propionate



```
qqnorm(delta_but_pro_wide_LOODAT_dbup$delta_propionate)
qqline(delta_but_pro_wide_LOODAT_dbup$delta_propionate) #p=0.729. Normality supported
```

## Normal Q-Q Plot



*#Equality of Variances*

```
var.test(delta_but_pro_wide_LOODAT_dbup$delta_butyrate, delta_but_pro_wide_LOODAT_dbup$delta_propionate
```

```
##
## F test to compare two variances
##
## data: delta_but_pro_wide_LOODAT_dbup$delta_butyrate and delta_but_pro_wide_LOODAT_dbup$delta_propionate
## F = 0.13425, num df = 9, denom df = 9, p-value = 0.006273
## alternative hypothesis: true ratio of variances is not equal to 1
## 90 percent confidence interval:
## 0.04223278 0.42677754
## sample estimates:
## ratio of variances
## 0.1342535
```

```
var.test(delta_but_pro_wide_LOODAT_dbup$delta_butyrate, delta_but_pro_wide_LOODAT_dbup$delta_propionate
```

```
##
## F test to compare two variances
##
## data: delta_but_pro_wide_LOODAT_dbup$delta_butyrate and delta_but_pro_wide_LOODAT_dbup$delta_propionate
## F = 0.13425, num df = 9, denom df = 9, p-value = 0.9969
## alternative hypothesis: true ratio of variances is greater than 1
## 90 percent confidence interval:
## 0.05501425 Inf
```



```

## sample estimates:
## ratio of variances
##      0.1342535

var.test(delta_but_pro_wide_LOODAT_dbup$delta_butyrate, delta_but_pro_wide_LOODAT_dbup$delta_propionate

##
## F test to compare two variances
##
## data: delta_but_pro_wide_LOODAT_dbup$delta_butyrate and delta_but_pro_wide_LOODAT_dbup$delta_propionate
## F = 0.13425, num df = 9, denom df = 9, p-value = 0.003136
## alternative hypothesis: true ratio of variances is less than 1
## 90 percent confidence interval:
##  0.0000000 0.3276243
## sample estimates:
## ratio of variances
##      0.1342535

#-----
#Assumptions selecting for propionate Conditions:
delta_but_pro_wide_LOODAT_pdown <- delta_but_pro_wide_LOODAT %>%
  filter(delta_propionate < 0) %>%
  summarise(sample_size = n()) #n=9
delta_but_pro_wide_LOODAT_pdown

## # A tibble: 1 x 1
##   sample_size
##   <int>
## 1         9

delta_but_pro_wide_LOODAT_pdown <- delta_but_pro_wide_LOODAT %>%
  filter(delta_propionate < 0)

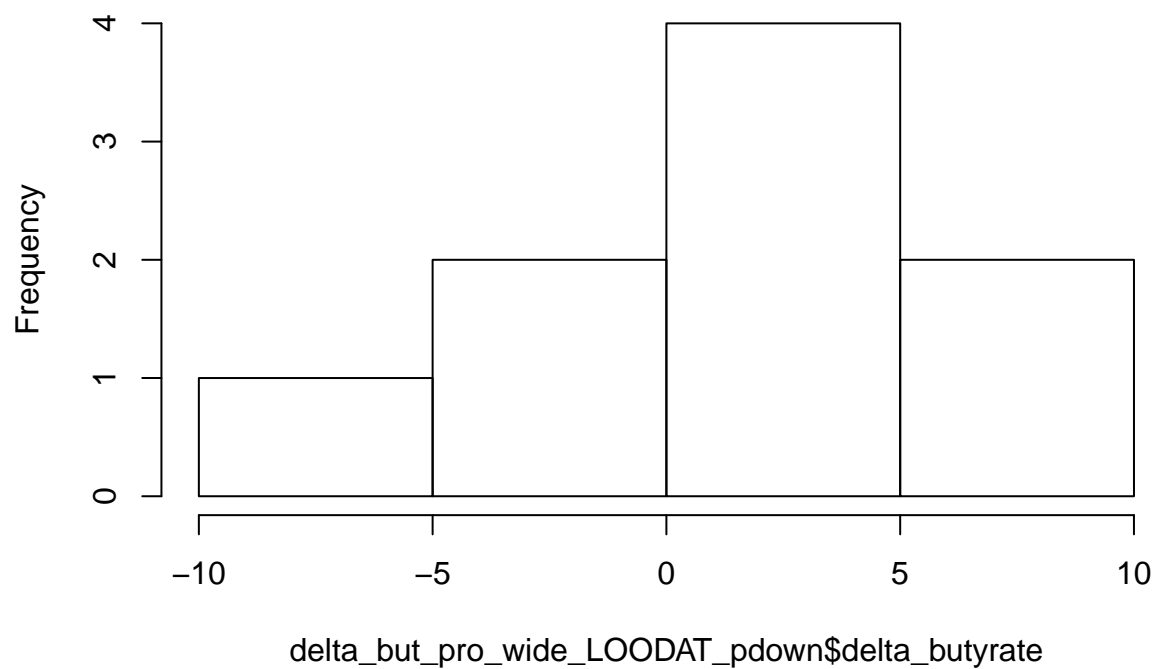
#Normality
#Butyrate:
shapiro.test(delta_but_pro_wide_LOODAT_pdown$delta_butyrate)

##
## Shapiro-Wilk normality test
##
## data: delta_but_pro_wide_LOODAT_pdown$delta_butyrate
## W = 0.96189, p-value = 0.8178

hist(delta_but_pro_wide_LOODAT_pdown$delta_butyrate)

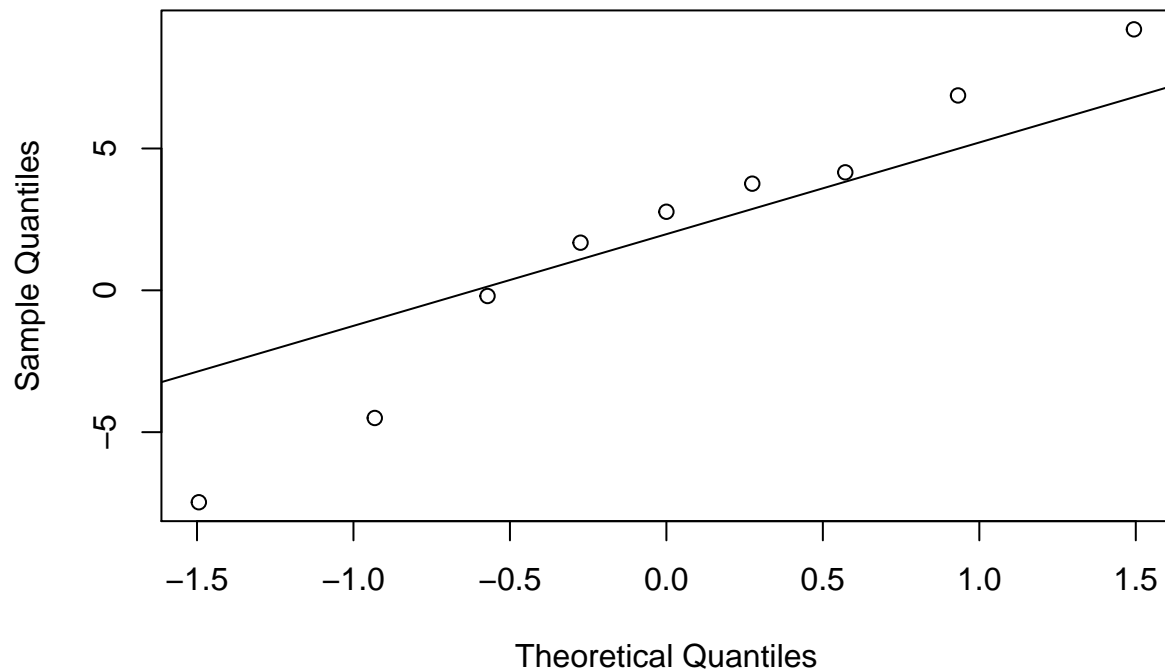
```

### Histogram of delta\_but\_pro\_wide\_LOODAT\_pdown\$delta\_butyrate



```
qqnorm(delta_but_pro_wide_LOODAT_pdown$delta_butyrate)
qqline(delta_but_pro_wide_LOODAT_pdown$delta_butyrate) #p=0.8178. Normality supported.
```

## Normal Q-Q Plot

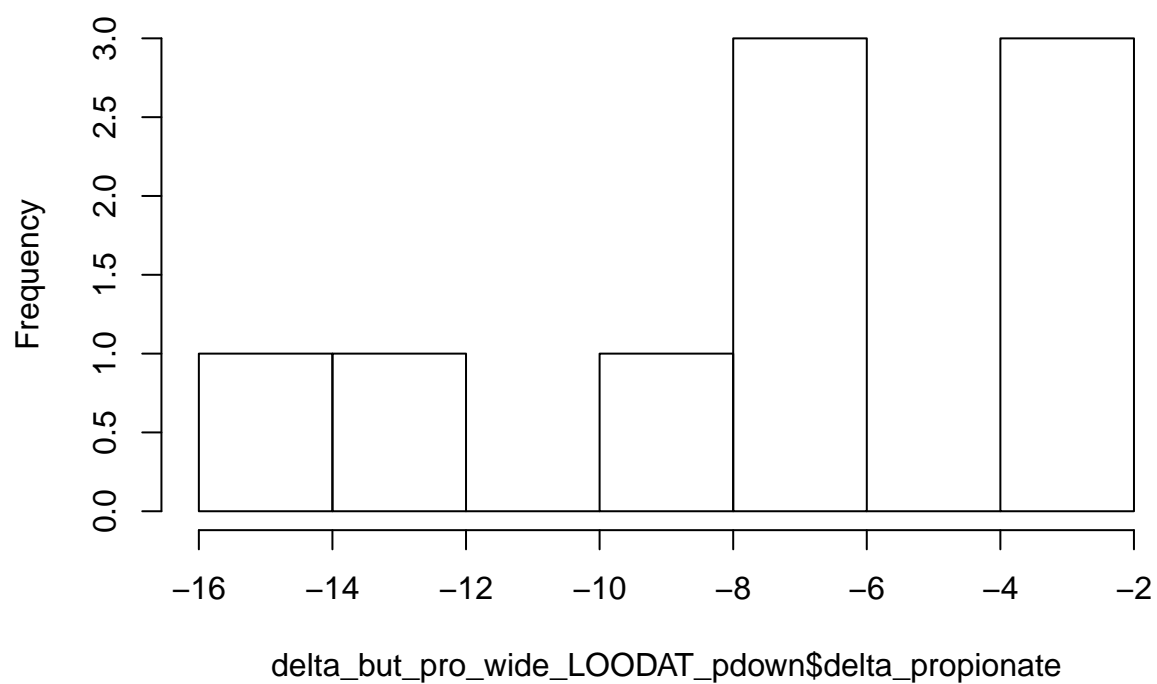


```
#Propionate:  
shapiro.test(delta_but_pro_wide_LOODAT_pdown$delta_propionate)
```

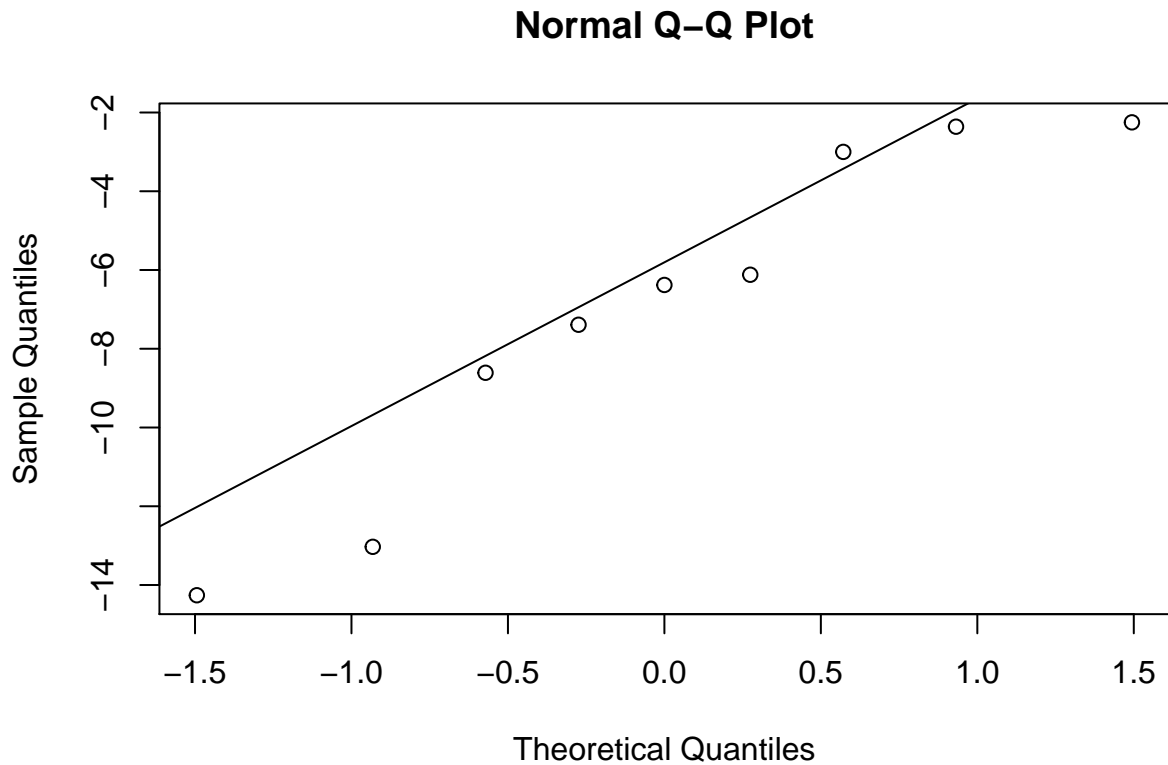
```
##  
##  Shapiro-Wilk normality test  
##  
## data:  delta_but_pro_wide_LOODAT_pdown$delta_propionate  
## W = 0.90401, p-value = 0.2762
```

```
hist(delta_but_pro_wide_LOODAT_pdown$delta_propionate)
```

Histogram of delta\_but\_pro\_wide\_LOODAT\_pdown\$delta\_propionate



```
qqnorm(delta_but_pro_wide_LOODAT_pdown$delta_propionate)
qqline(delta_but_pro_wide_LOODAT_pdown$delta_propionate) #p=0.2762. Normality supported
```



*#Equality of Variances*

```
var.test(delta_but_pro_wide_LOODAT_pdown$delta_butyrate, delta_but_pro_wide_LOODAT_pdown$delta_propiona
```

```
##
## F test to compare two variances
##
## data: delta_but_pro_wide_LOODAT_pdown$delta_butyrate and delta_but_pro_wide_LOODAT_pdown$delta_prop
## F = 1.4404, num df = 8, denom df = 8, p-value = 0.6178
## alternative hypothesis: true ratio of variances is not equal to 1
## 90 percent confidence interval:
## 0.4189646 4.9523882
## sample estimates:
## ratio of variances
## 1.440443
```

```
var.test(delta_but_pro_wide_LOODAT_pdown$delta_butyrate, delta_but_pro_wide_LOODAT_pdown$delta_propiona
```

```
##
## F test to compare two variances
##
## data: delta_but_pro_wide_LOODAT_pdown$delta_butyrate and delta_but_pro_wide_LOODAT_pdown$delta_prop
## F = 1.4404, num df = 8, denom df = 8, p-value = 0.3089
## alternative hypothesis: true ratio of variances is greater than 1
## 90 percent confidence interval:
## 0.5562953 Inf
```

```

## sample estimates:
## ratio of variances
##          1.440443

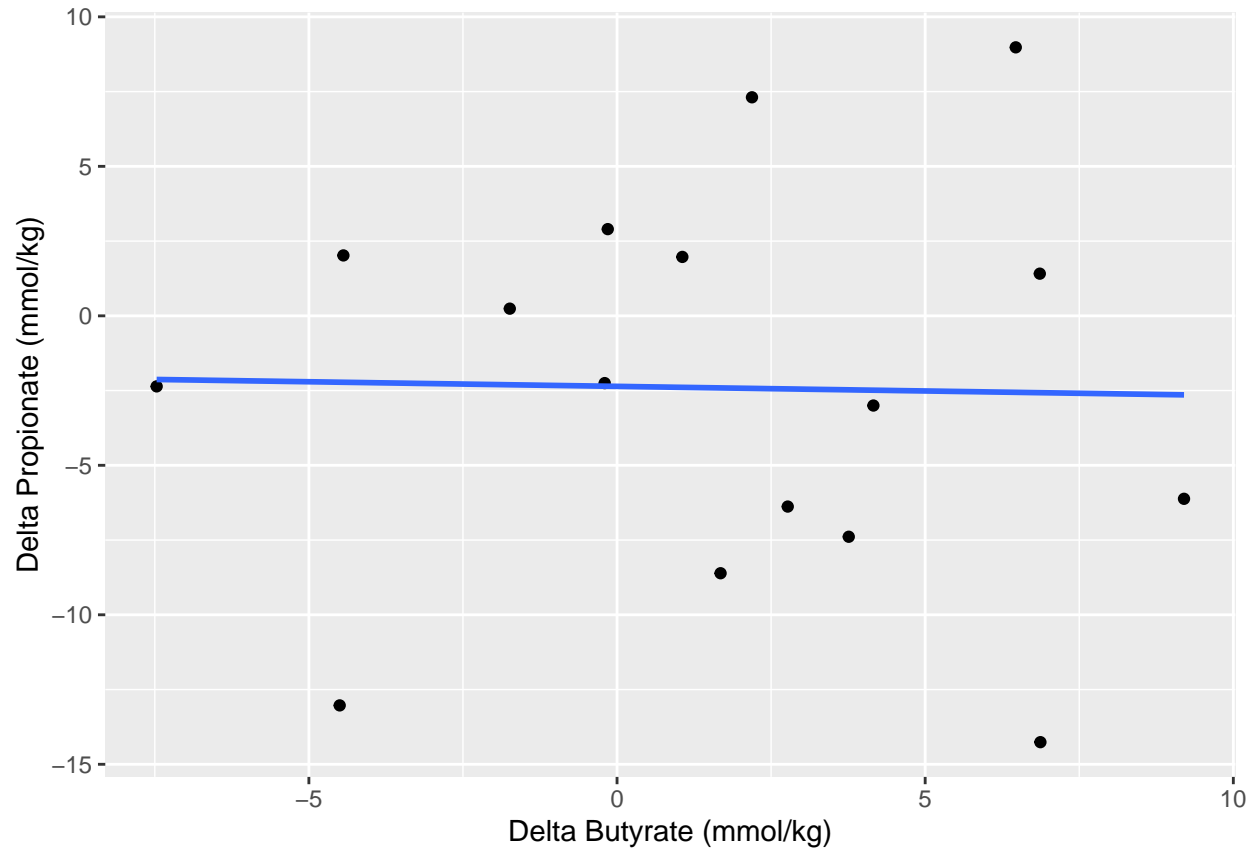
var.test(delta_but_pro_wide_LOODAT_pdown$delta_butyrate, delta_but_pro_wide_LOODAT_pdown$delta_propionate)

##
## F test to compare two variances
##
## data:  delta_but_pro_wide_LOODAT_pdown$delta_butyrate and delta_but_pro_wide_LOODAT_pdown$delta_propionate
## F = 1.4404, num df = 8, denom df = 8, p-value = 0.6911
## alternative hypothesis: true ratio of variances is less than 1
## 90 percent confidence interval:
##  0.000000 3.729809
## sample estimates:
## ratio of variances
##          1.440443

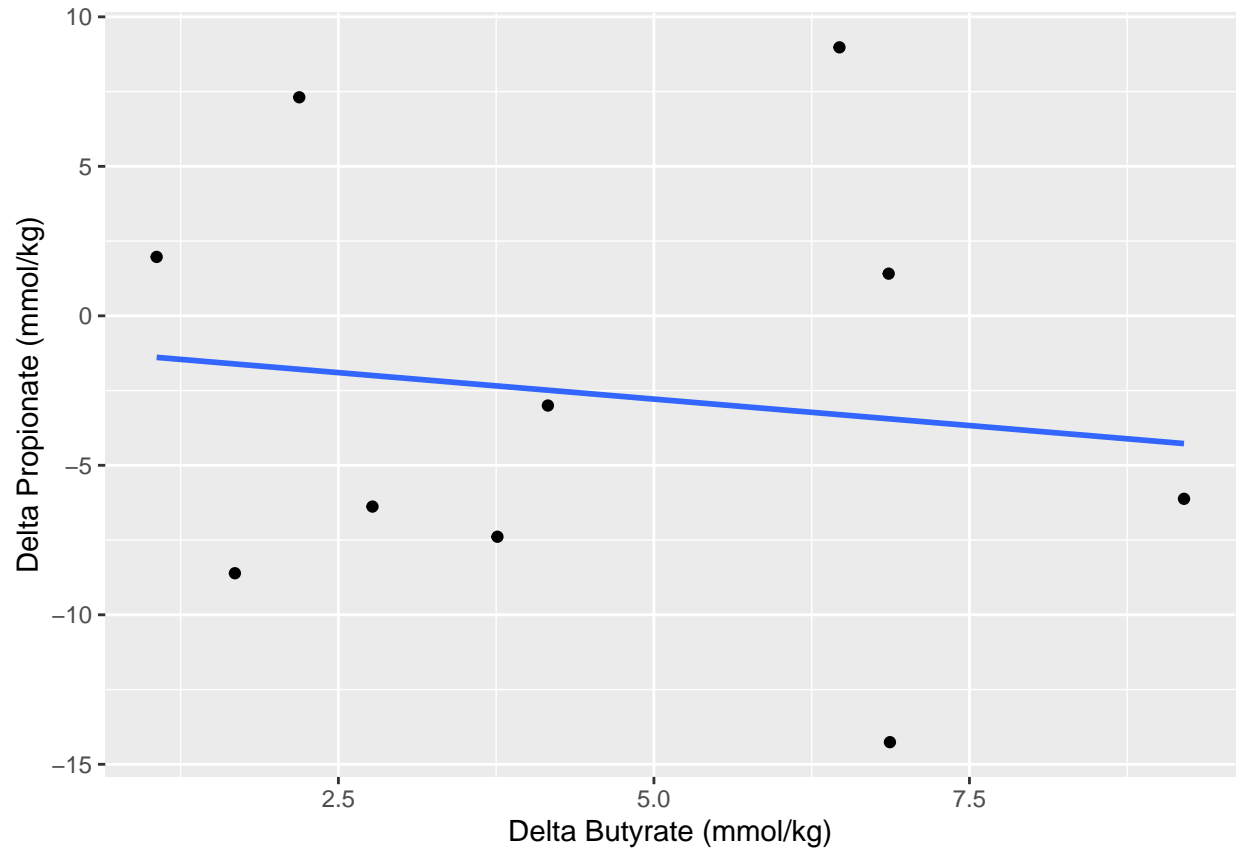
####Plot

LOODAT_correlation_plot <- delta_but_pro_wide_LOODAT %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(LOODAT_correlation_plot)

```

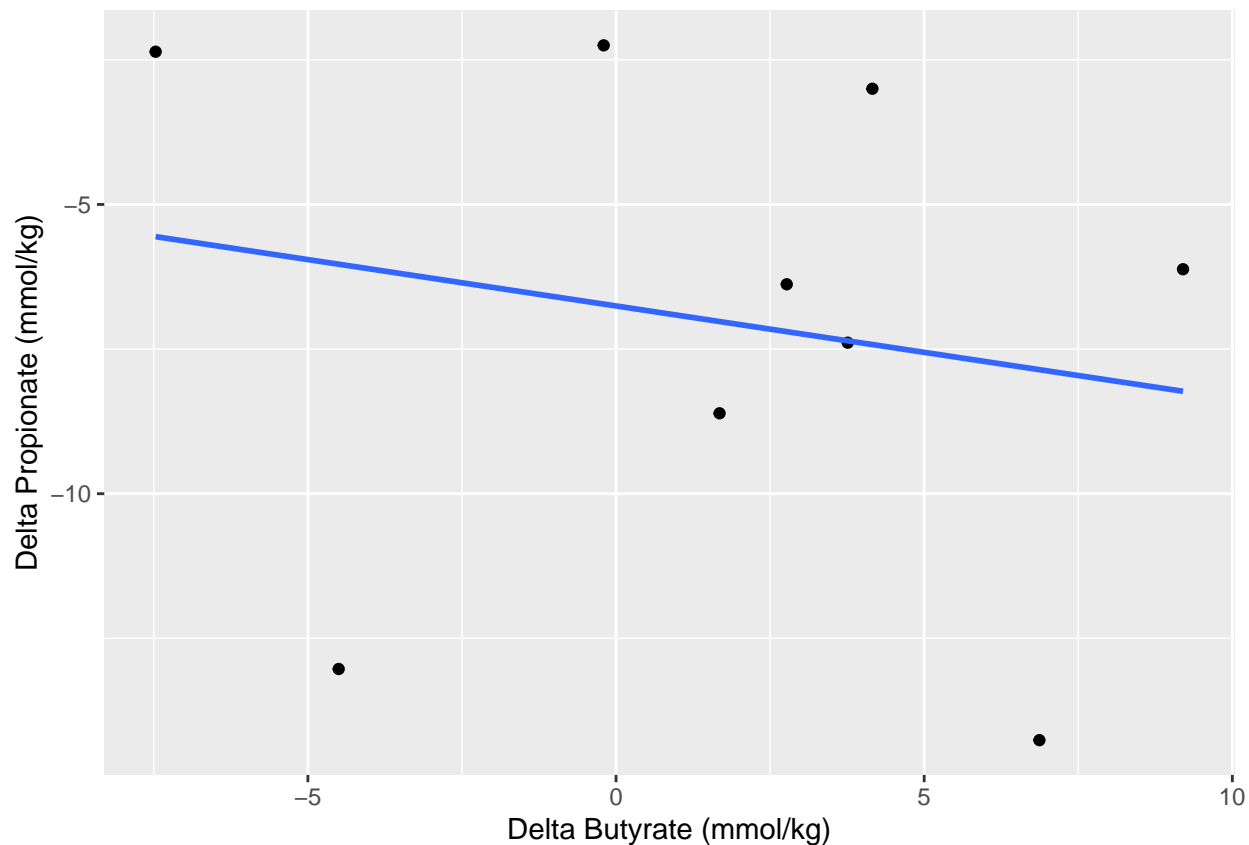


```
#-----
#Plot Assuming butyrate Conditions Met:
LOODAT_correlation_plot_dbup <- delta_but_pro_wide_LOODAT_dbup %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(LOODAT_correlation_plot_dbup)
```



```
#-----
#Plot Assuming propionate Conditions Met:
LOODAT_correlation_plot_pdown <- delta_but_pro_wide_LOODAT_pdown %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(LOODAT_correlation_plot_pdown)
```





####Test

```
LOODAT_test <- delta_but_pro_wide_LOODAT %>%
  lm(delta_propionate ~ delta_butyrate, data = .)
summary(LOODAT_test) #view results
```

```
##
## Call:
## lm(formula = delta_propionate ~ delta_butyrate, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -11.6886  -4.1799  -0.0638   4.2724  11.5391
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  -2.35949    1.82785  -1.291   0.218
## delta_butyrate -0.03085    0.38141  -0.081   0.937
##
## Residual standard error: 6.86 on 14 degrees of freedom
## Multiple R-squared:  0.0004672, Adjusted R-squared:  -0.07093
## F-statistic: 0.006543 on 1 and 14 DF, p-value: 0.9367
```

```
#-----
#Test Assuming Butyrate Condition Met:
LOODAT_test_dbup <- delta_but_pro_wide_LOODAT_dbup %>%
```

```
lm(delta_propionate ~ delta_butyrate, data = .)
summary(LOODAT_test_dbup) #view results
```

```
##
## Call:
## lm(formula = delta_propionate ~ delta_butyrate, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -10.812  -4.879  -1.180   4.481  12.286
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    -1.0149     4.9587  -0.205   0.843
## delta_butyrate -0.3541     0.9568  -0.370   0.721
##
## Residual standard error: 7.769 on 8 degrees of freedom
## Multiple R-squared:  0.01683,    Adjusted R-squared:  -0.1061
## F-statistic: 0.137 on 1 and 8 DF,  p-value: 0.7209
```

```
#-----
#Test Assuming Propionate Condition Met:
LOODAT_test_pdown <- delta_but_pro_wide_LOODAT_pdown %>%
  lm(delta_propionate ~ delta_butyrate, data = .)
summary(LOODAT_test_pdown) #view results
```

```
##
## Call:
## lm(formula = delta_propionate ~ delta_butyrate, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -6.9969 -1.5860   0.8187   3.1969   4.4725
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    -6.7546     1.6283  -4.148   0.0043 **
## delta_butyrate -0.1603     0.3090  -0.519   0.6199
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.588 on 7 degrees of freedom
## Multiple R-squared:  0.03703,    Adjusted R-squared:  -0.1005
## F-statistic: 0.2692 on 1 and 7 DF,  p-value: 0.6199
```

```
##BRMPS+HiMaize
```

```
but_wide_HMB <- all_data_wkly %>%
  rename_all(tolower) %>%
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, butyrate_mean)
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes", frequency == "1xday")
  spread(study_week, butyrate_mean) %>%
```

```

mutate(delta_butyrate = week3 - week1) #calculate difference in butyrate

pro_wide_HMB <- all_data_wkly %>%
  rename_all(tolower) %>%
  select(participant_id, semester, supplement_consumed, frequency, quantity_compliant, study_week, propionate_mean) %>%
  filter(study_week == "week1" | study_week == "week3", quantity_compliant == "yes", frequency == "1xday") %>%
  spread(study_week, propionate_mean) %>%
  mutate(delta_propionate = week3 - week1) #calculate difference in propionate

delta_but_pro_wide_HMB <- inner_join(x = but_wide_HMB, y = pro_wide_HMB,
  by = c("participant_id",
        "semester", "supplement_consumed", "frequency", "quantity_compliant")) %>%
  select(-starts_with("week"))

```

###Tidy Data ###Analysis ###Assumptions

```
nrow(delta_but_pro_wide_HMB) # number of data points
```

```
## [1] 23
```

```
shapiro.test(delta_but_pro_wide_HMB$delta_butyrate) #call column of df with values = vector
```

```
##
##  Shapiro-Wilk normality test
##
## data:  delta_but_pro_wide_HMB$delta_butyrate
## W = 0.94331, p-value = 0.2767
```

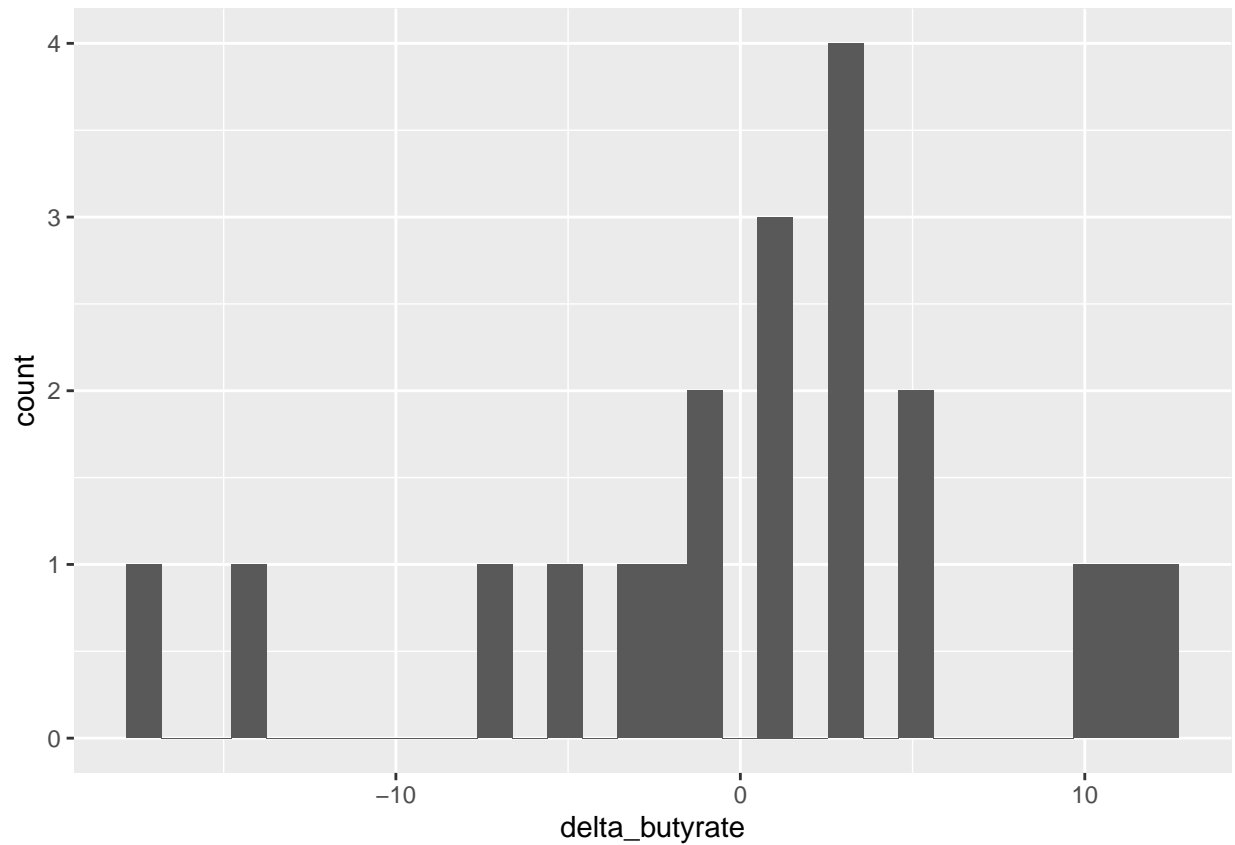
```
shapiro.test(delta_but_pro_wide_HMB$delta_propionate) #call column of df with values = vector
```

```
##
##  Shapiro-Wilk normality test
##
## data:  delta_but_pro_wide_HMB$delta_propionate
## W = 0.88154, p-value = 0.0156
```

```
ggplot(delta_but_pro_wide_HMB, aes(x=delta_butyrate)) +
  geom_histogram() #histograms show the number of data points (count) at each value
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

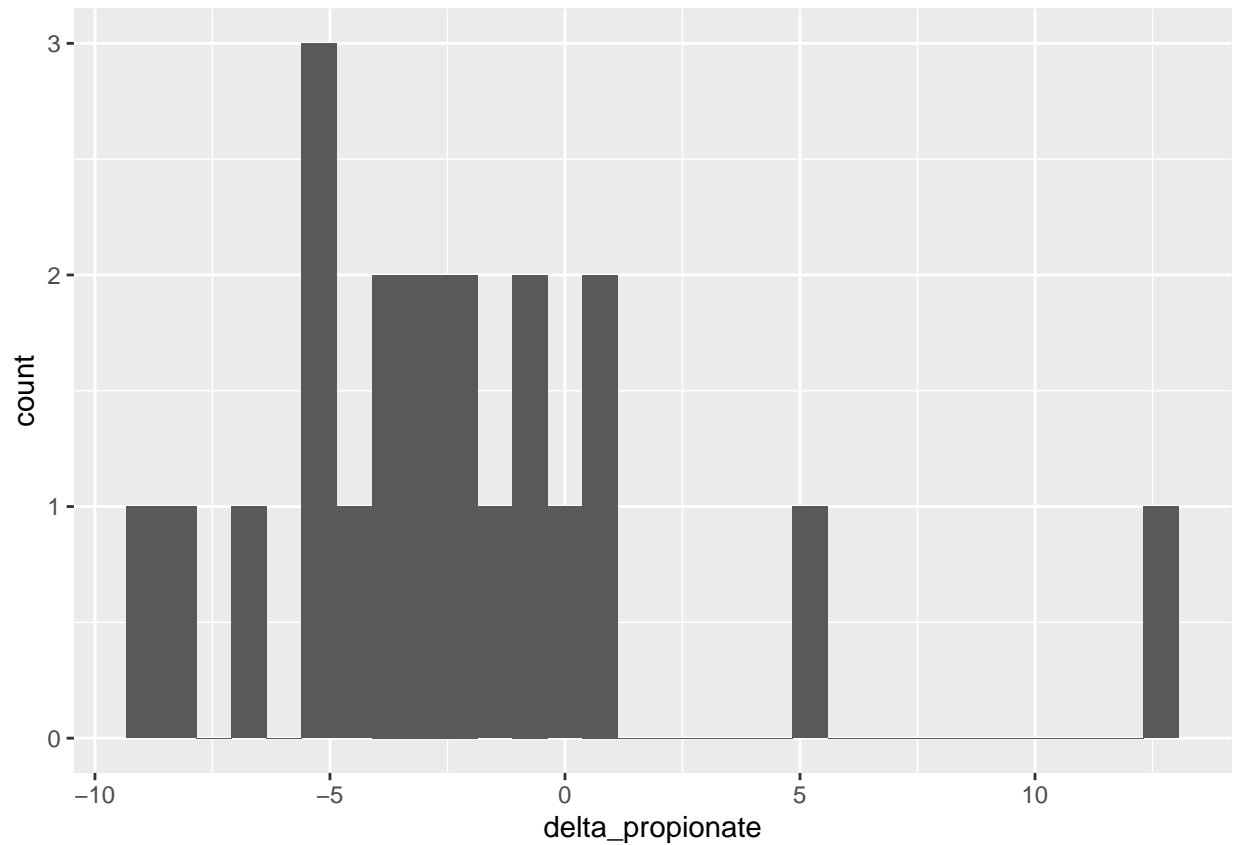
```
## Warning: Removed 3 rows containing non-finite values (stat_bin).
```



```
ggplot(delta_but_pro_wide_HMB, aes(x=delta_propionate)) +  
  geom_histogram() #histograms show the number of data points (count) at each value
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

```
## Warning: Removed 2 rows containing non-finite values (stat_bin).
```

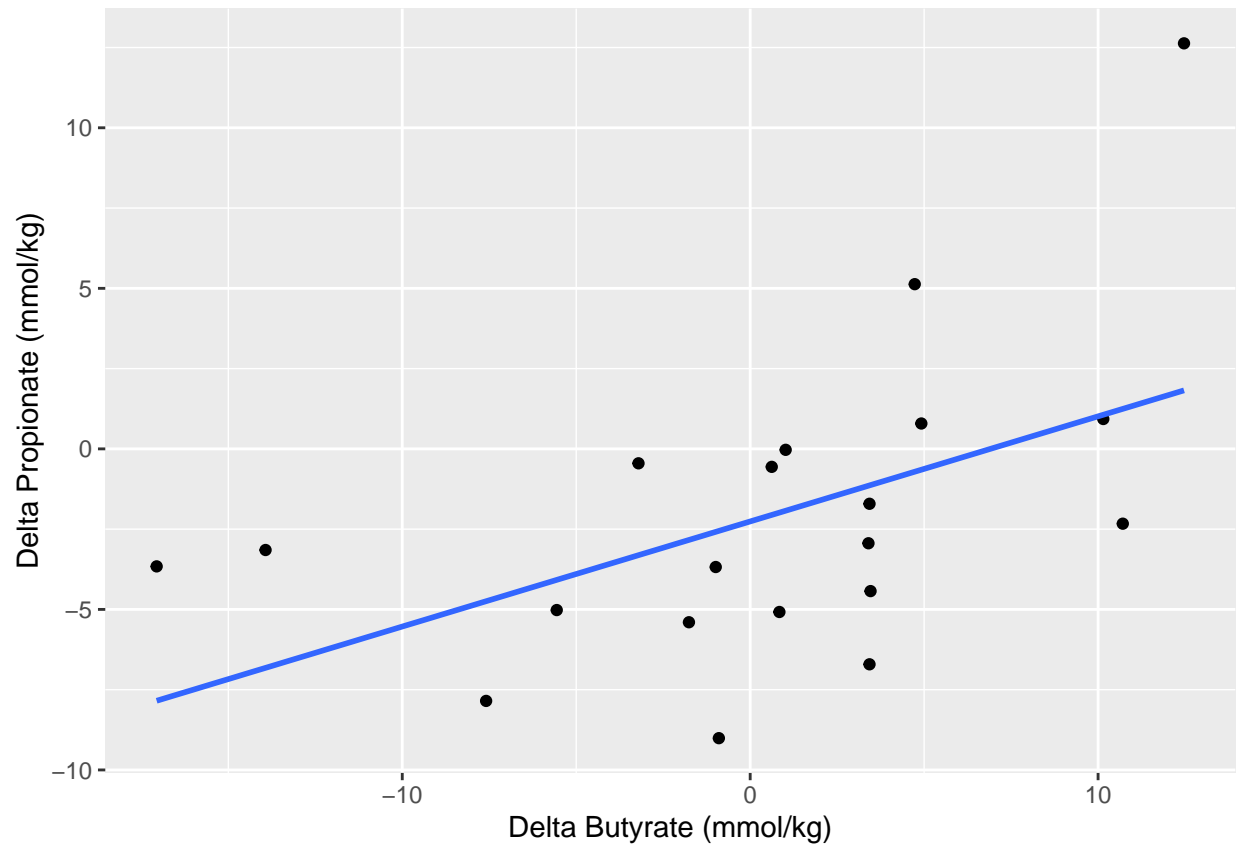


####Plot

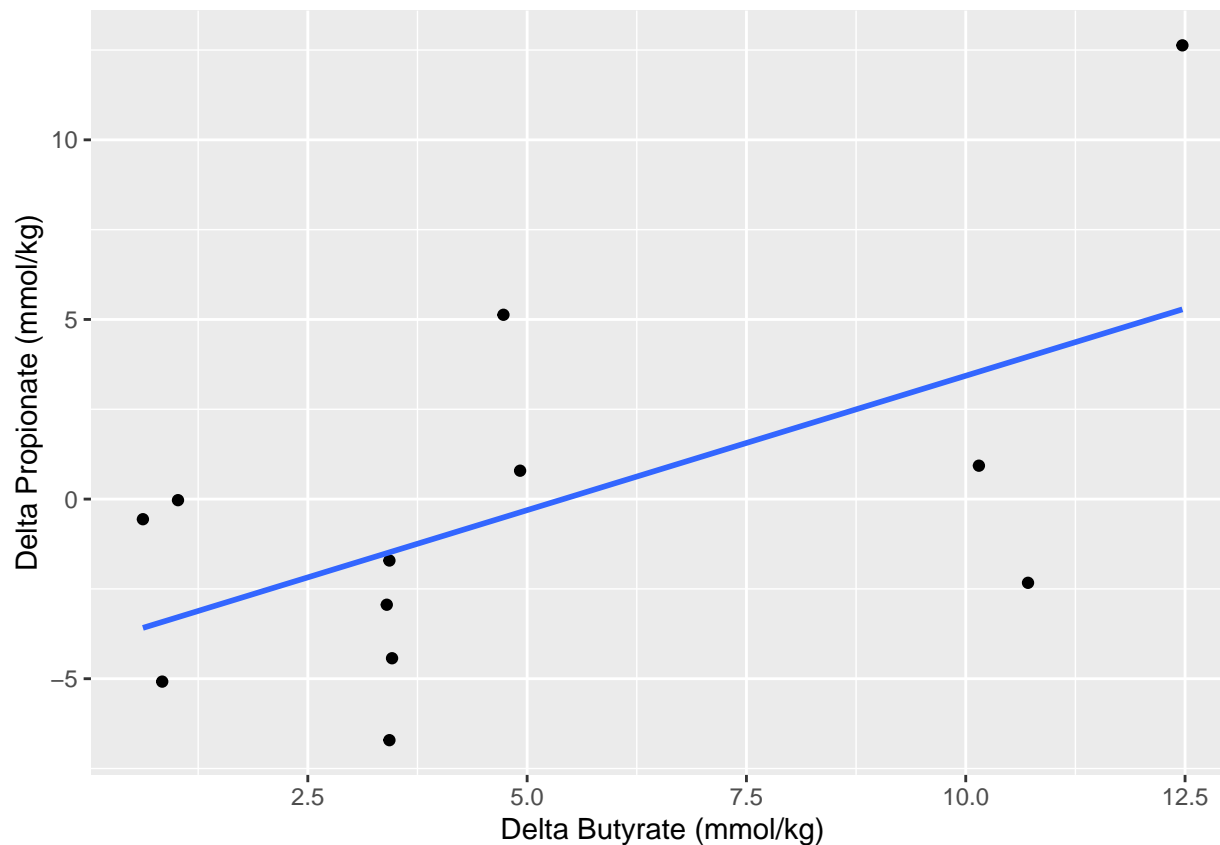
```
linear_correlation_HMB_plot<-delta_but_pro_wide_HMB %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(linear_correlation_HMB_plot)
```

## Warning: Removed 3 rows containing non-finite values (stat\_smooth).

## Warning: Removed 3 rows containing missing values (geom\_point).



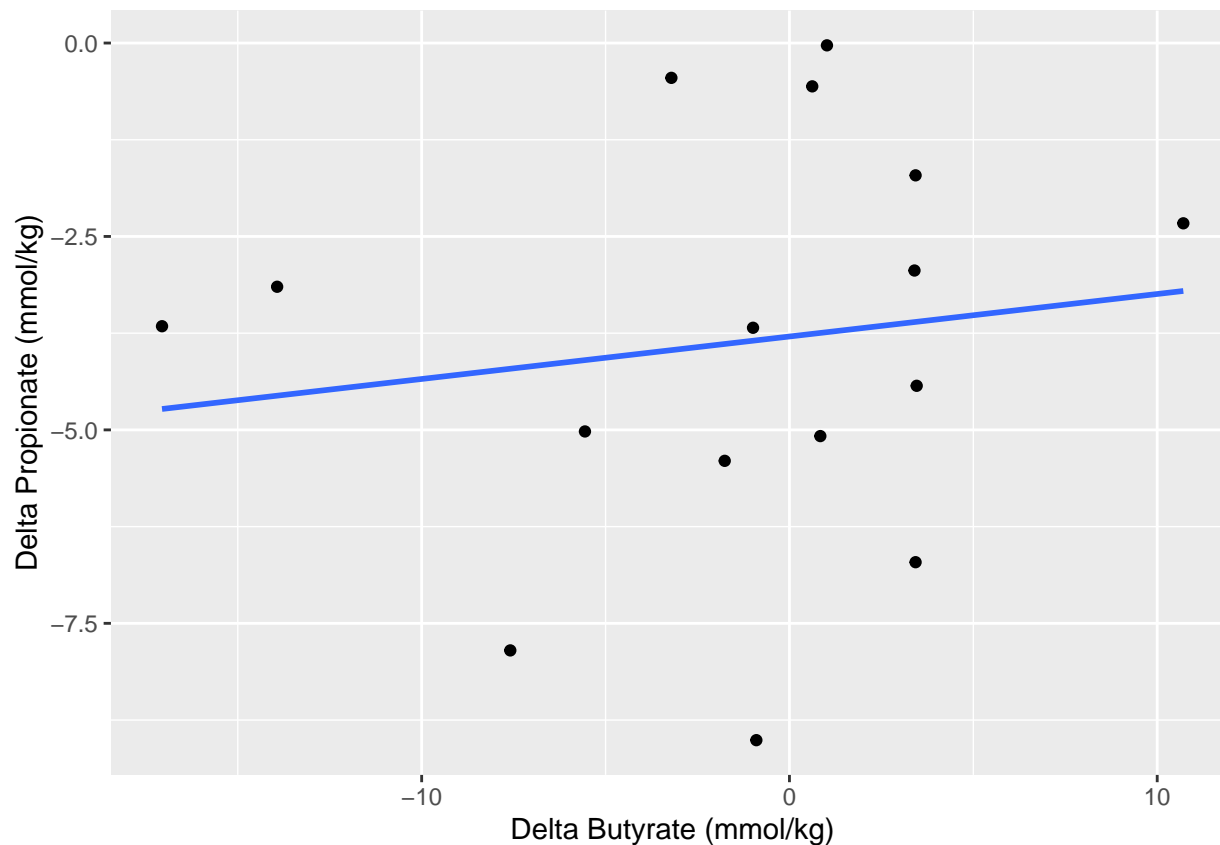
```
lm_HMB_highBut_plot<-delta_but_pro_wide_HMB %>%
  filter(delta_butyrate > "0") %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(lm_HMB_highBut_plot)
```



```
lm_HMB_LowPro_plot<-delta_but_pro_wide_HMB %>%
  filter(delta_propionate < "0") %>%
  ggplot(aes(x = delta_butyrate,
             y = delta_propionate)) +
  geom_point() + #puts data points to match x and y coordinates
  geom_smooth(method = "lm", #used to create a linear best fit line
             se = FALSE) + #hides confidence interval around line
  xlab("Delta Butyrate (mmol/kg)") +
  ylab("Delta Propionate (mmol/kg)")
print(lm_HMB_LowPro_plot)
```

```
## Warning: Removed 1 rows containing non-finite values (stat_smooth).
```

```
## Warning: Removed 1 rows containing missing values (geom_point).
```



####Test

```
HMB_lm <- delta_but_pro_wide_HMB %>%
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship
summary(HMB_lm) #view results
```

```
##
## Call:
## lm(formula = delta_butyrate ~ delta_propionate, data = .)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -16.2628  -1.9142   0.4301   3.7494  10.4611
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)      2.0816     1.6196   1.285  0.2150
## delta_propionate  0.7865     0.3148   2.498  0.0224 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 6.595 on 18 degrees of freedom
## (3 observations deleted due to missingness)
## Multiple R-squared:  0.2575, Adjusted R-squared:  0.2162
## F-statistic: 6.242 on 1 and 18 DF, p-value: 0.02238
```



*# Small .022 p-value indicates significance, and the other R2 is small-ish at .2 representing that the*

```
HMB_lm_HiBut <- delta_but_pro_wide_HMB %>%  
  filter(delta_butyrate > "0") %>%  
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship  
summary(HMB_lm_HiBut) #view results
```

```
##  
## Call:  
## lm(formula = delta_butyrate ~ delta_propionate, data = .)  
##  
## Residuals:  
##      Min       1Q   Median       3Q      Max   
## -4.2210 -2.1401 -0.4485  1.4428  6.6682   
##  
## Coefficients:  
##              Estimate Std. Error t value Pr(>|t|)      
## (Intercept)      5.0938     0.9913   5.139 0.000439 ***  
## delta_propionate  0.4515     0.1999   2.259 0.047472 *    
## ---  
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1  
##  
## Residual standard error: 3.425 on 10 degrees of freedom  
## Multiple R-squared:  0.3378, Adjusted R-squared:  0.2716   
## F-statistic: 5.102 on 1 and 10 DF,  p-value: 0.04747
```

*# Small .047 p-value indicates significance, and the R2 is small-ish at .33 representing that the corr*

```
HMB_lm_LoPro <- delta_but_pro_wide_HMB %>%  
  filter(delta_propionate < "0") %>%  
  lm(delta_butyrate ~ delta_propionate, data = .) #test relationship  
summary(HMB_lm_LoPro) #view results
```

```
##  
## Call:  
## lm(formula = delta_butyrate ~ delta_propionate, data = .)  
##  
## Residuals:  
##      Min       1Q   Median       3Q      Max   
## -15.6378  -3.1755   0.9399   4.2090  11.6175   
##  
## Coefficients:  
##              Estimate Std. Error t value Pr(>|t|)      
## (Intercept)    -0.005734   3.245037  -0.002    0.999   
## delta_propionate 0.387006   0.701894   0.551    0.590   
##  
## Residual standard error: 7.077 on 14 degrees of freedom  
## (1 observation deleted due to missingness)  
## Multiple R-squared:  0.02125, Adjusted R-squared:  -0.04866   
## F-statistic: 0.304 on 1 and 14 DF,  p-value: 0.5901
```

*# Somewhat small .59 p-value indicates possible significance, and the tiny R2 of -.049 represents that*