

# Quick Reference Guide: Diversity Analysis

## Quick Start (Copy-Paste Ready)

```
# 1. Install packages (run once)
install.packages(c("vegan", "iNEXT", "betapart", "dplyr", "tidyR"))

# 2. Load Libraries
library(vegan)
library(iNEXT)
library(betapart)
library(dplyr)
library(tidyR)

# 3. Set working directory and Load data
setwd("path/to/your/data") # CHANGE THIS
master_taxa <- read.csv("masterTaxaGenus.csv")
benthics <- read.csv("stationBenthicstTESTSITE.csv")
station_info <- read.csv("stationInfoBenSampsTESTSITE.csv")

# 4. Run the main script
source("diversity_analysis.R")
```

## Understanding the Three Diversity Metrics

### Visual Concept

```
Stream System with 5 Sites (riffles)
=====
Site 1: Species A, B, C, D (4 species)
Site 2: Species A, B, E, F (4 species)
Site 3: Species C, D, G, H (4 species)
Site 4: Species E, F, I, J (4 species)
Site 5: Species A, C, E, G (4 species)

ALPHA ( $\alpha$ ): Average diversity within sites
=  $(4 + 4 + 4 + 4 + 4) / 5 = 4$  species per site

GAMMA ( $\gamma$ ): Total diversity across all sites
= 10 unique species total (A, B, C, D, E, F, G, H, I, J)

BETA ( $\beta$ ): How much communities differ
=  $\gamma / \alpha = 10 / 4 = 2.5$ 
= "Species composition changes 2.5 times across sites"
```

## The Three Beta Diversity Methods Compared

Method	Data Used	Range	Best For
Hill Numbers ( $\beta = \gamma/\alpha$ )	Abundance	1 to $\infty$	Understanding how many distinct communities exist
Jaccard	Presence/Absence	0 to 1	Comparing which species are present
Bray-Curtis	Abundance	0 to 1	Detecting abundance-driven differences

## Example Output Interpretation

### Scenario 1: Low Beta Diversity (Homogeneous Community)

```
Year: 2015
gamma_q0 = 25 (total species)
mean_alpha_q0 = 20 (avg species per site)
beta_q0 = 1.25 (25/20)

mean_jaccard = 0.15 (low dissimilarity)
mean_braycurtis = 0.20 (low dissimilarity)
```

**Interpretation:** Sites are very similar! Most species are found at most sites. Low environmental heterogeneity or high dispersal among sites.

### Scenario 2: High Beta Diversity (Heterogeneous Community)

```
Year: 2015
gamma_q0 = 50 (total species)
mean_alpha_q0 = 12 (avg species per site)
beta_q0 = 4.17 (50/12)

mean_jaccard = 0.75 (high dissimilarity)
mean_braycurtis = 0.80 (high dissimilarity)
```

**Interpretation:** Sites are very different! Species composition changes dramatically. Suggests strong environmental gradients, habitat heterogeneity, or dispersal barriers.

### Scenario 3: Rare Species Dominate Pattern ( $q$ values differ)

```
Year: 2015
beta_q0 = 3.5 (many rare species drive turnover)
beta_q1 = 2.1 (intermediate)
beta_q2 = 1.4 (common species are similar across sites)
```

**Interpretation:** Many rare species are site-specific, but common/dominant species are shared across sites. This is typical in streams with good overall habitat quality but local microhabitat variation.

## Formulas Reference

### Hill Numbers

$q = 0$ : Species Richness  
= Number of species

$q = 1$ : Hill-Shannon  
=  $\exp(-\sum(\pi_i \times \ln(\pi_i)))$   
where  $\pi_i$  = proportion of species i

$q = 2$ : Hill-Simpson  
=  $1 / \sum(\pi_i^2)$

### Beta Diversity

Hill approach:

$$\beta = \gamma / \alpha$$

Where:

$\gamma$  = total diversity across all sites

$\alpha$  = average diversity within sites

$\beta$  = "effective number of communities"

### Jaccard Index

$$\text{Jaccard dissimilarity} = (b + c) / (a + b + c)$$

Where:

a = species present at both sites

b = species only at site 1

c = species only at site 2

### Bray-Curtis

$$\text{Bray-Curtis} = \sum |x_{ij} - x_{ik}| / \sum (x_{ij} + x_{ik})$$

Where:

$x_{ij}$  = abundance of species i at site j

$x_{ik}$  = abundance of species i at site k

## Common Patterns in Stream Systems

### Headwaters → Downstream Gradient

Typical pattern:

- High beta diversity ( $\beta = 3-5$ )
- High Bray-Curtis dissimilarity (0.6-0.8)
- $q=2$  beta lower than  $q=0$  (common species track river continuum)

## Disturbed vs. Reference Sites

Disturbed systems:

- Lower alpha diversity
- Lower gamma diversity
- Sometimes higher beta (patchy degradation)

Reference systems:

- Higher alpha diversity
- Higher gamma diversity
- Moderate beta (natural heterogeneity)

## Seasonal Variation

Spring vs. Fall:

- Often different species pools (gamma)
- May have similar beta patterns
- Life cycle phenology drives differences

## Quick Diagnostics

### Is my analysis working correctly?

- ✓ **Check 1:** Is  $\text{gamma} \geq \text{alpha}$  for all years? - If not, check your grouping and calculations
- ✓ **Check 2:** Is  $\text{beta} \geq 1$  for Hill numbers? - Beta cannot be less than 1 mathematically
- ✓ **Check 3:** Are Jaccard and Bray-Curtis between 0 and 1? - If not, check for negative abundances or data errors
- ✓ **Check 4:** Do sample sizes make sense? - Check `n_comparisons` in output tables - For N sites, expect  $N(N-1)/2$  pairwise comparisons

## Red Flags in Your Data

⚠ **Warning signs:** - Beta = 1 exactly (suggests only one site or duplicate sites) - Gamma = Alpha (suggests only one site analyzed) - All dissimilarities = 0 (all sites identical - check data) - All dissimilarities = 1 (no shared species - unusual!)

## File Output Guide

### `alpha_diversity_results.csv`

Columns:

- SiteID: Unique sample identifier
- q0\_richness: Number of species (Hill  $q=0$ )
- q1\_shannon: Shannon diversity (Hill  $q=1$ )
- q2\_simpson: Simpson diversity (Hill  $q=2$ )
- total\_abundance: Total individuals counted
- StationID: Station identifier
- BenSampID: Sample identifier

- Year: Collection year
- Season: Collection season

Use for:

- Plotting diversity over time
- Comparing sites
- Identifying high/low diversity samples

### [diversity\\_summary\\_by\\_year.csv](#)

Columns:

- Year: Collection year
- gamma\_q0, gamma\_q1, gamma\_q2: Regional diversity
- mean\_alpha\_q0, mean\_alpha\_q1, mean\_alpha\_q2: Avg site diversity
- beta\_q0, beta\_q1, beta\_q2: Beta diversity (Hill numbers)
- mean\_jaccard: Average Jaccard dissimilarity
- mean\_braycurtis: Average Bray-Curtis dissimilarity

Use for:

- Annual trends in diversity
- Comparing beta diversity metrics
- Summary statistics for reports

## Visualization Ideas

### Plot Alpha Diversity Over Time

```
library(ggplot2)

ggplot(alpha_diversity, aes(x = Year, y = q0_richness)) +
  geom_point() +
  geom_smooth(method = "lm") +
  labs(title = "Species Richness Over Time",
       y = "Alpha Diversity (q=0)",
       x = "Year")
```

### Plot Beta Diversity Comparison

```
beta_long <- diversity_summary %>%
  select(Year, beta_q0, beta_q1, beta_q2) %>%
  pivot_longer(cols = starts_with("beta"),
               names_to = "metric",
               values_to = "beta_diversity")

ggplot(beta_long, aes(x = Year, y = beta_diversity, color = metric)) +
  geom_line() +
  geom_point() +
  labs(title = "Beta Diversity by q Value",
       y = "Beta Diversity",
       x = "Year")
```

## Compare Dissimilarity Indices

```
dissim_long <- diversity_summary %>%
  select(Year, mean_jaccard, mean_braycurtis) %>%
  pivot_longer(cols = c(mean_jaccard, mean_braycurtis),
               names_to = "index",
               values_to = "dissimilarity")

ggplot(dissim_long, aes(x = Year, y = dissimilarity, color = index)) +
  geom_line() +
  geom_point() +
  labs(title = "Community Dissimilarity Over Time",
       y = "Mean Dissimilarity",
       x = "Year")
```

## Statistical Testing Ideas

### Test for temporal trends

```
# Linear model for alpha diversity
model_alpha <- lm(q0_richness ~ Year, data = alpha_diversity)
summary(model_alpha)

# Linear model for beta diversity
model_beta <- lm(beta_q0 ~ Year, data = diversity_summary)
summary(model_beta)
```

### Compare seasons

```
# t-test comparing Spring vs Fall
spring_data <- alpha_diversity %>% filter(Season == "Spring")
fall_data <- alpha_diversity %>% filter(Season == "Fall")

t.test(spring_data$q0_richness, fall_data$q0_richness)
```

### PERMANOVA for community composition

```
# Test if years differ in community composition
adonis2(species_matrix ~ Year,
        data = site_metadata,
        method = "bray")
```

## Troubleshooting Checklist

Problem: Script crashes when loading data - [ ] Are all three CSV files in working directory?  
- [ ] Check file names match exactly (case-sensitive) - [ ] Try: `list.files()` to see what files R sees

Problem: No results for certain years - [ ] Check date format in station\_info file - [ ] Verify: `unique(site_metadata$Year)` - [ ] Look for NA values: `sum(is.na(station_info$Year))`

Problem: Alpha diversity values seem wrong - [ ] Check Individuals column for negative values - [ ] Verify species names in FinalID column - [ ] Look for duplicate taxa:  
table(benthics\$FinalID)

Problem: Beta diversity > 10 (seems too high) - [ ] Check if you have very few individuals per site - [ ] Verify gamma calculation is working - [ ] May indicate poor sampling or data quality issues

## Citation Template

If using these methods in a publication:

We calculated alpha, beta, and gamma diversity using Hill numbers ( $q = 0, 1, 2$ ) following Chao et al. (2014). Beta diversity was calculated as  $\beta = \gamma/\alpha$  (Jost 2007). We also calculated pairwise dissimilarity using Jaccard (presence/absence) and Bray-Curtis (abundance-based) indices (Anderson et al. 2011). All analyses were performed in R v4.x using the vegan (Dixon 2003), iNEXT (Hsieh et al. 2016), and betapart (Baselga & Orme 2012) packages.