

# Class 10: Structural Bioinformatics (pt. 1)

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## The PDB Database

The [Protein Data Bank \(PDB\)](#) is the main repository of biomolecular structure data. We will see what is in it.

```
stats <- read.csv("pdb_stats.csv", row.names = 1)
stats
```

	X.ray	EM	NMR	Integrative	Multiple.methods	Neutron
Protein (only)	178795	21825	12773	343	226	84
Protein/Oligosaccharide	10363	3564	34	8	11	1
Protein/NA	9106	6335	287	24	7	0
Nucleic acid (only)	3132	221	1566	3	15	3
Other	175	25	33	4	0	0
Oligosaccharide (only)	11	0	6	0	1	0
	Other	Total				
Protein (only)	32	214078				
Protein/Oligosaccharide	0	13981				
Protein/NA	0	15759				
Nucleic acid (only)	1	4941				
Other	0	237				
Oligosaccharide (only)	4	22				

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
n.sums <- colSums(stats)
n <- n.sums/n.sums["Total"]
round(n, digits=2)
```

	X.ray	EM	NMR	Integrative
	0.81	0.13	0.06	0.00
Multiple.methods		Neutron	Other	Total
	0.00	0.00	0.00	1.00

```
round((n["X.ray"] + n["EM"]), digits=2)
```

```
X.ray
0.94
```

94% of structures solved by X-ray and EM.

What is the total number of entries in the PDB?

```
total <- n.sums["Total"]
total
```

```
Total
249018
```

Q2: What proportion of structures in the PDB are protein?

```
n.proteins <- stats$Total[1]
round(n.proteins/total, digits=2)
```

```
Total
0.86
```

86% of structures are protein.

## Using Molstar

We will use the main [Molstar viewer online](#)

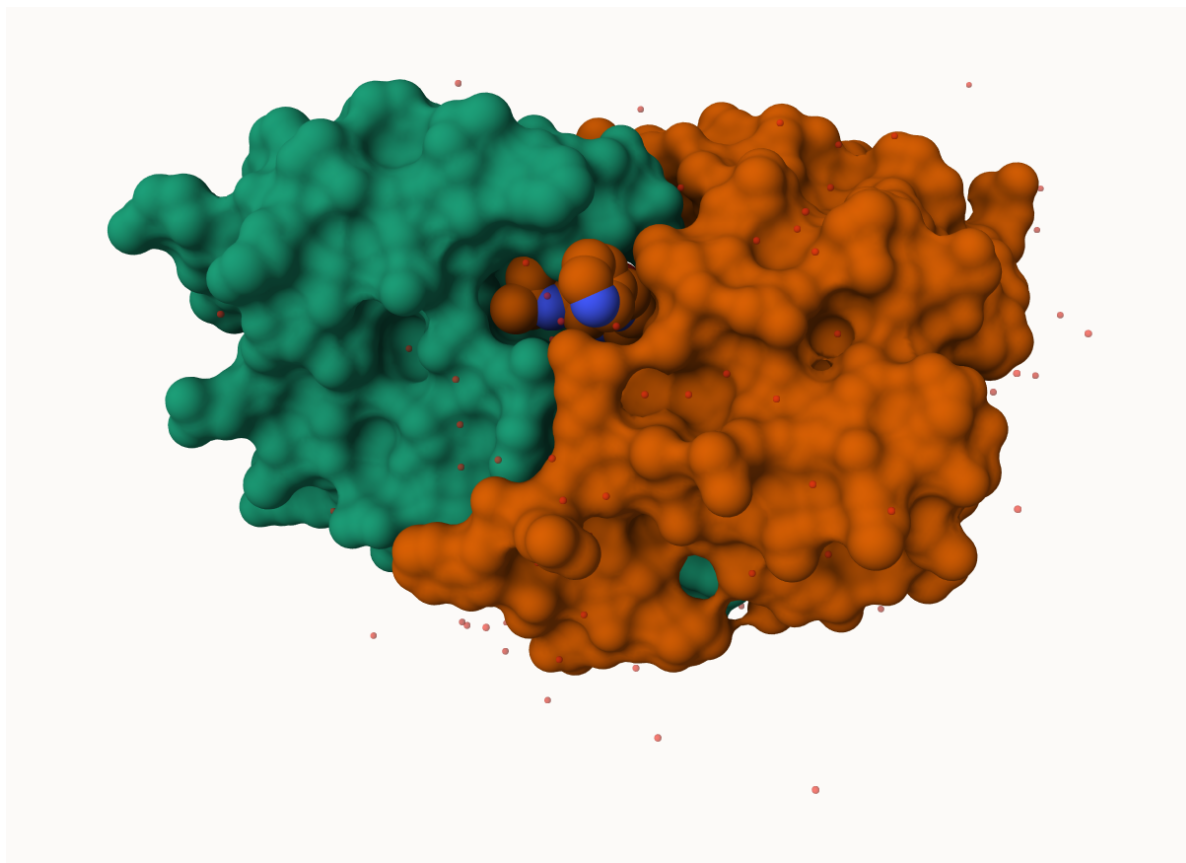


Figure 1: First view of HIV-Pr dimer with bound inhibitor

Q. Generate and insert an image of the HIV-Pr cartoon representation colored by secondary structure, showing the inhibitor (ligand) in ball and stick.

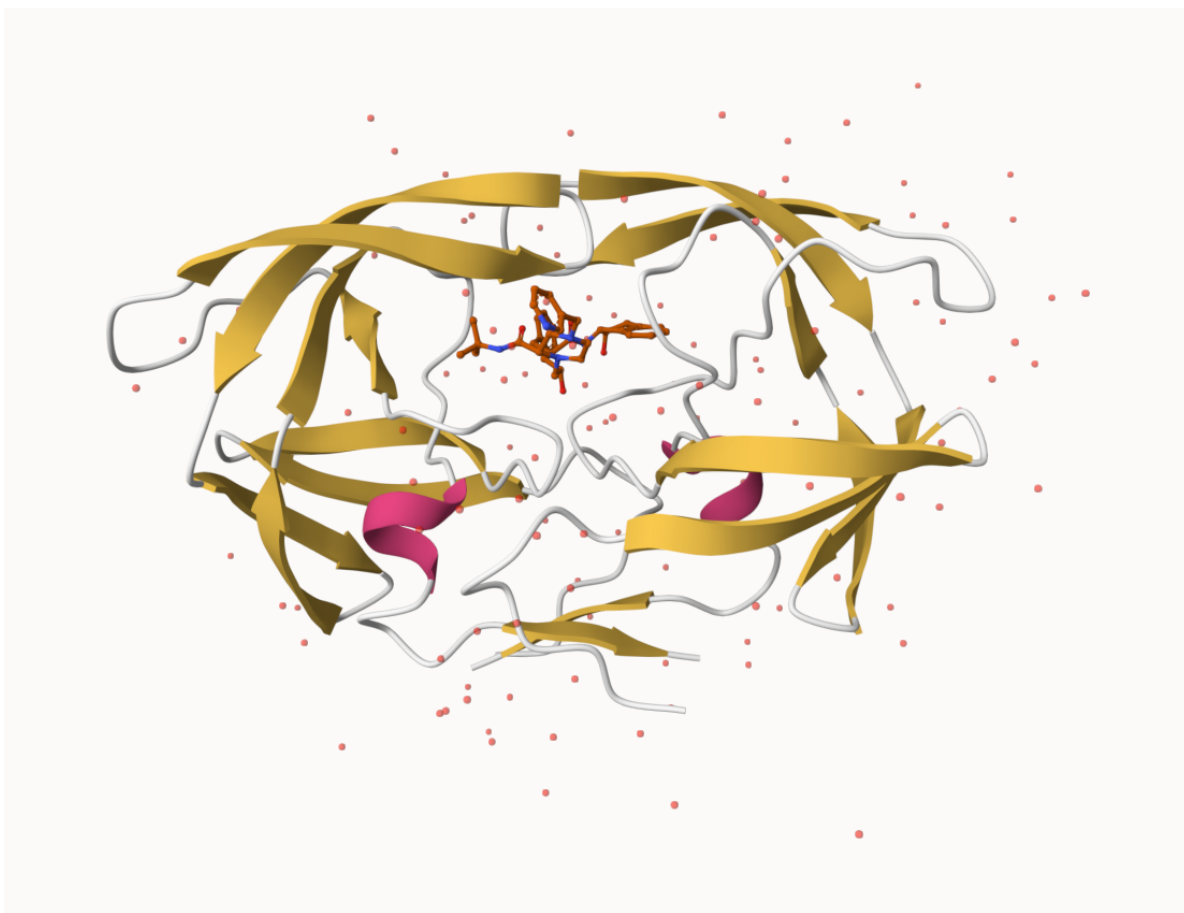


Figure 2: HIV-Pr colored by secondary structure.

Q4. Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

This is a simplified view for visibility.

Q5. There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have?

H2O 308

Q6. Generate and insert an image showing catalytic ASP 25 residue as ball and stick, and the all-important active site water molecule as space-fill.

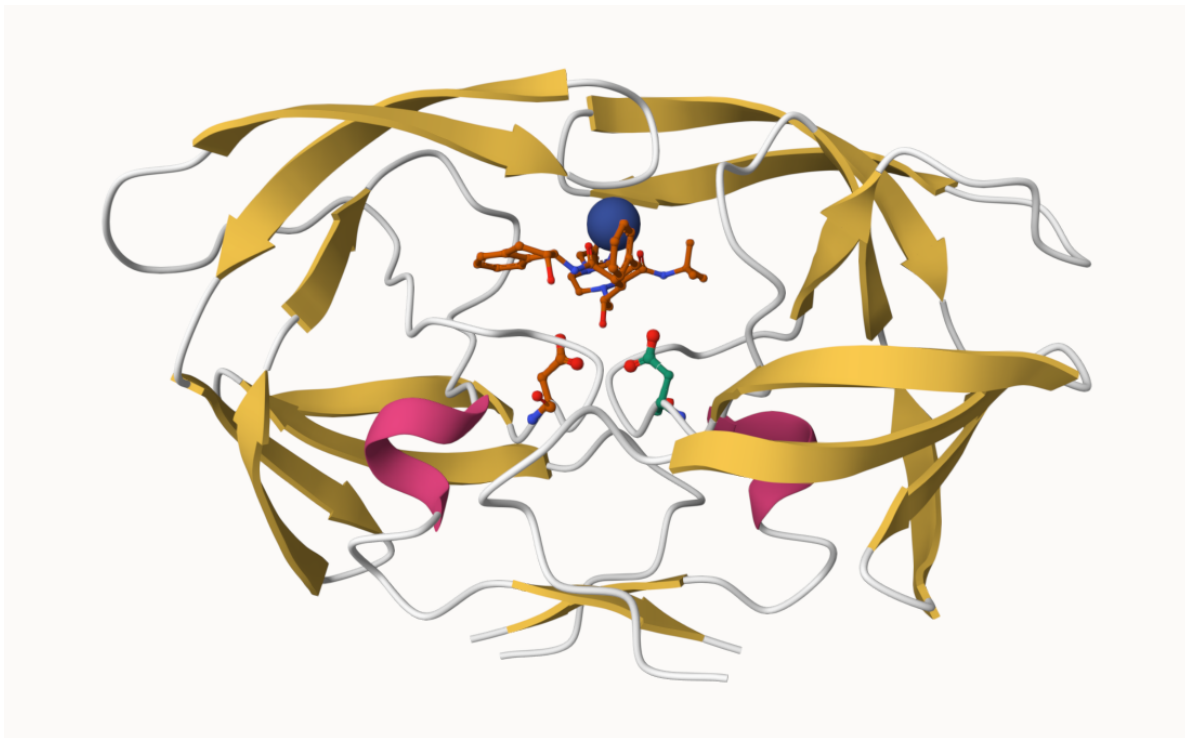


Figure 3: HIV-Pr with ASP 25 and active site H2O 308 highlighted.

## The Bio3D Package for structural bioinformatics

```
library(bio3d)
hiv <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
hiv
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
```

```
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)  
 Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)  
 Non-protein/nucleic Atoms#: 172 (residues: 128)  
 Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD  
 QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE  
 ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP  
 VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,  
 calpha, remark, call

Q7. How many amino acid residues are there in this pdb object?

198

Q8. Name one of the two non-protein residues?

MK1

Q9. How many protein chains are in this structure?

2

```
head(hiv$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b							
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10							
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62							
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64							
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40							
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87							
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40							
	segid			eletsy	charge															
1	<NA>		N	<NA>																
2	<NA>		C	<NA>																
3	<NA>		C	<NA>																
4	<NA>		O	<NA>																
5	<NA>		C	<NA>																
6	<NA>		C	<NA>																

```
pdbseq(hiv)
```

```
 1  2  3  4  5  6  7  8  9 10 11 12 13 14 15 16 17 18 19 20
"P" "Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K"
21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40
"E" "A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G"
41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
"R" "W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D"
61 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80
"Q" "I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T"
81 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99  1
"P" "V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F" "P"
 2  3  4  5  6  7  8  9 10 11 12 13 14 15 16 17 18 19 20 21
"Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K" "E"
22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41
"A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G" "R"
42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61
"W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D" "Q"
62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80 81
"I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T" "P"
82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99
"V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F"
```

Let's try out the new **bio3dview** package that is not yet on CRAN. We can use the **remotes** package to install any R package from GitHub.

### Quick Viewing of PDBs

```
library(bio3dview)
library(NGLViewerR)
```

```
#sele <- atom.select(hiv, resno=25)

#view.pdb(hiv, backgroundColor = "lightgreen",
#         highlight = sele,
#         highlight.style = "spacefill") |>
# setSpin()
```

## Prediction of Protein Flexibility

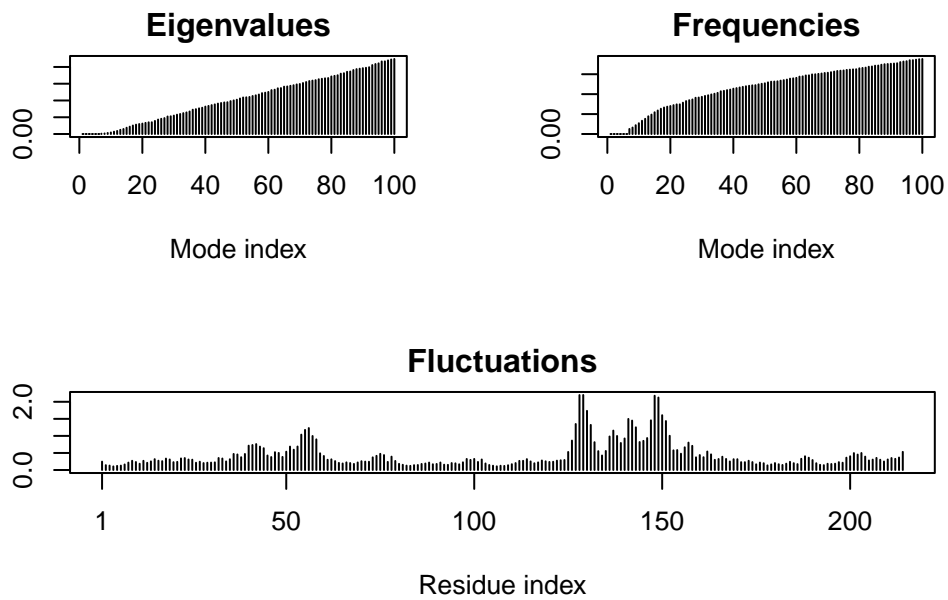
```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
m <- nma(adk)
```

Building Hessian... Done in 0.03 seconds.  
Diagonalizing Hessian... Done in 0.31 seconds.

```
plot(m)
```



Write out our results as a small trajectory movie:

```
#mktrj(m, file="results.pdb")
```

Alternative viewing of flexible parts:



```
#view.nma(m)
```