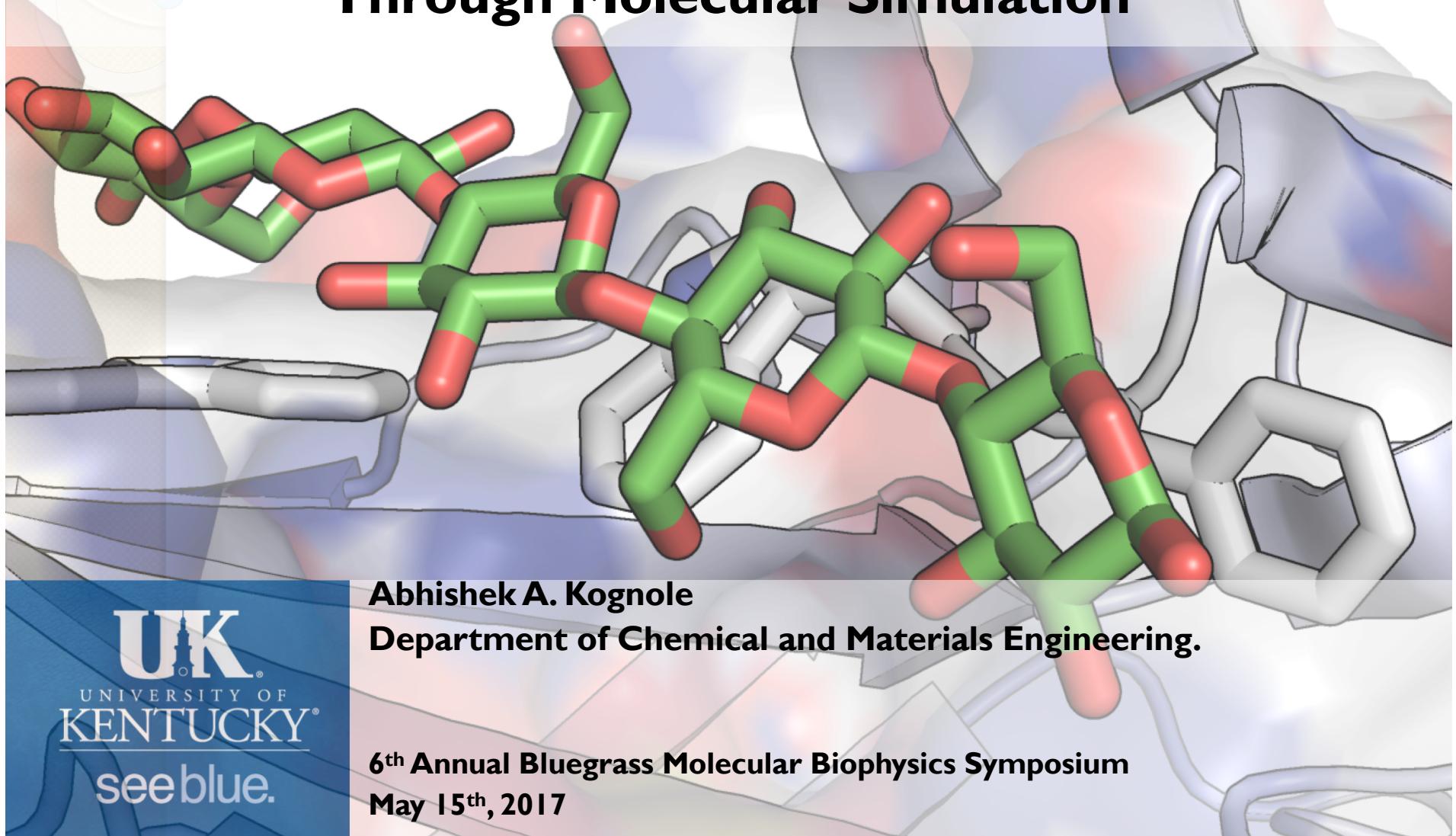


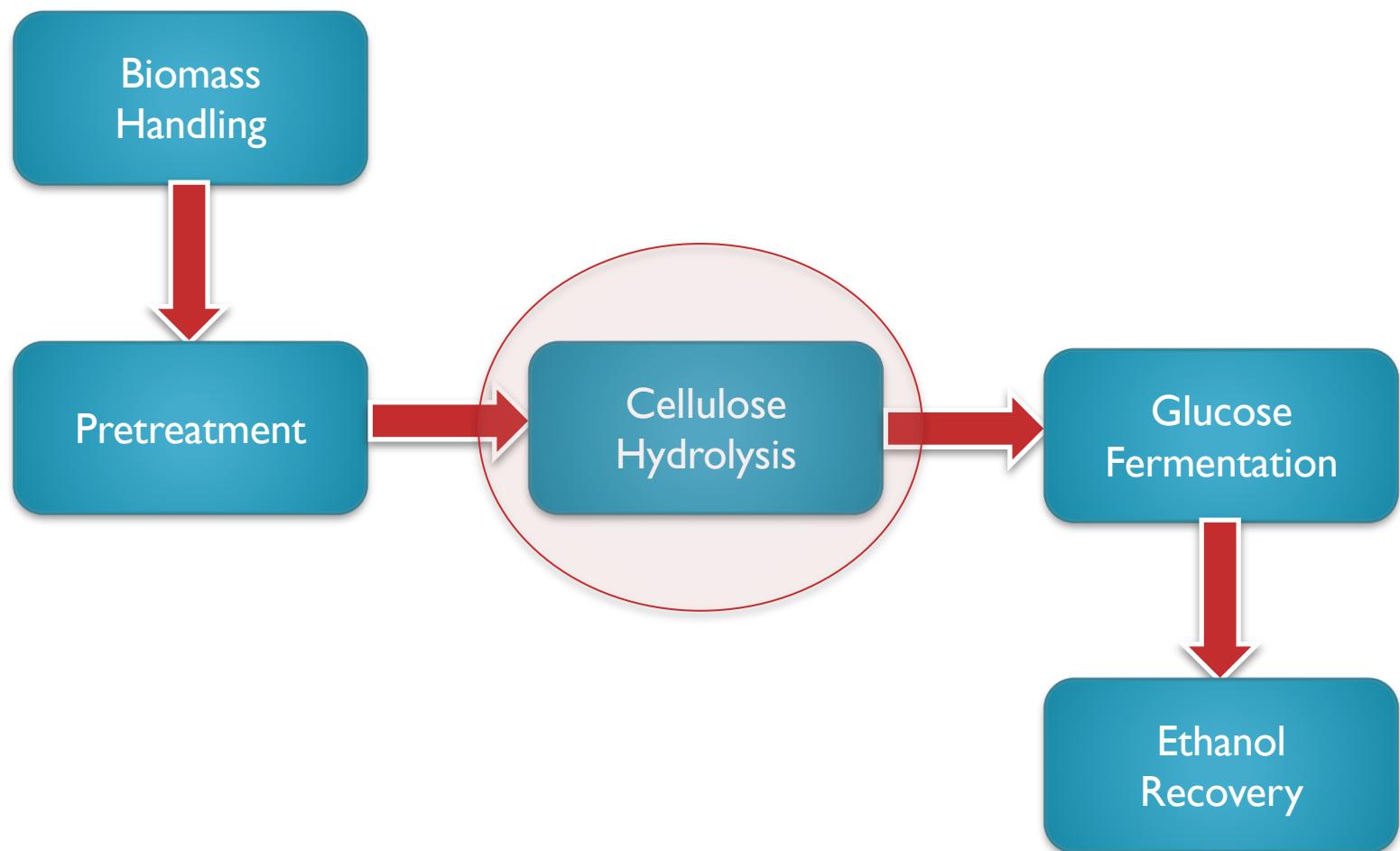
Understanding Substrate Recognition Mechanisms of Type B Carbohydrate Binding Modules (CBMs) Through Molecular Simulation



Abhishek A. Kognole
Department of Chemical and Materials Engineering.

6th Annual Bluegrass Molecular Biophysics Symposium
May 15th, 2017

Introduction: Biochemical biomass conversion



Background

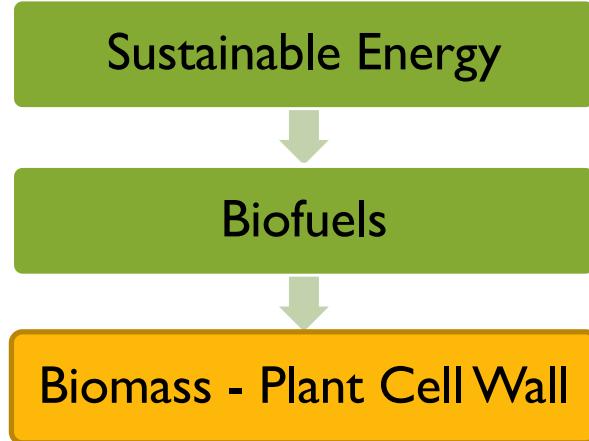
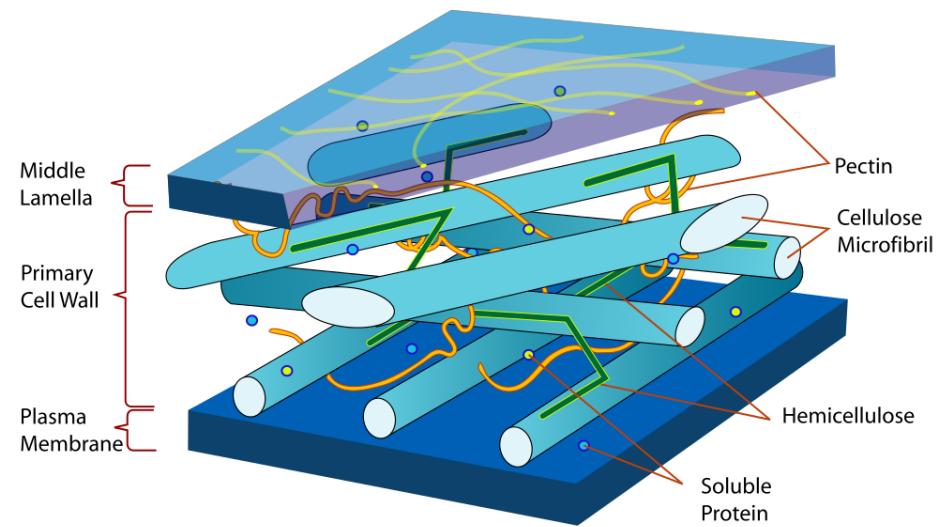
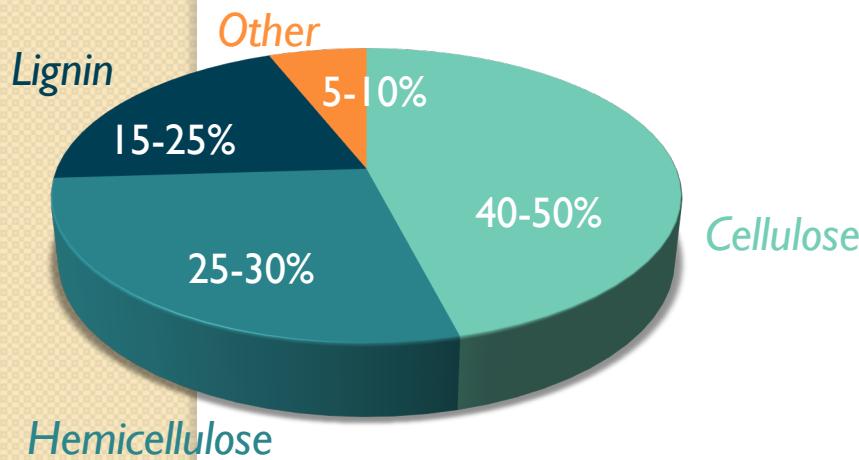
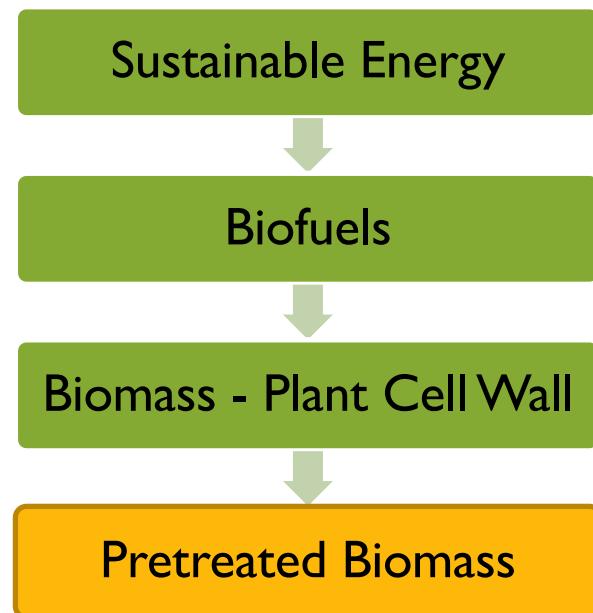


Image: Singh et al., *Renewable and Sustainable Energy Reviews*, 2014.

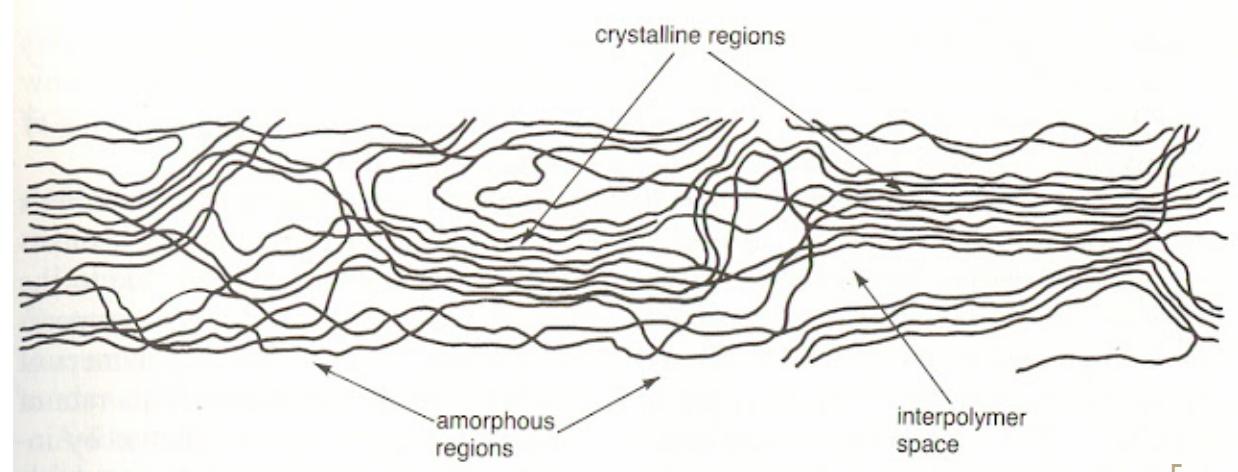


Background



Cellulose morphologies after pretreatment:

- Cello-oligosaccharides
- Amorphous regions
 - Separate insoluble polysaccharides
 - Convoluted insoluble polysaccharides
 - Partially decrystallized polysaccharides
- Crystalline regions



Background

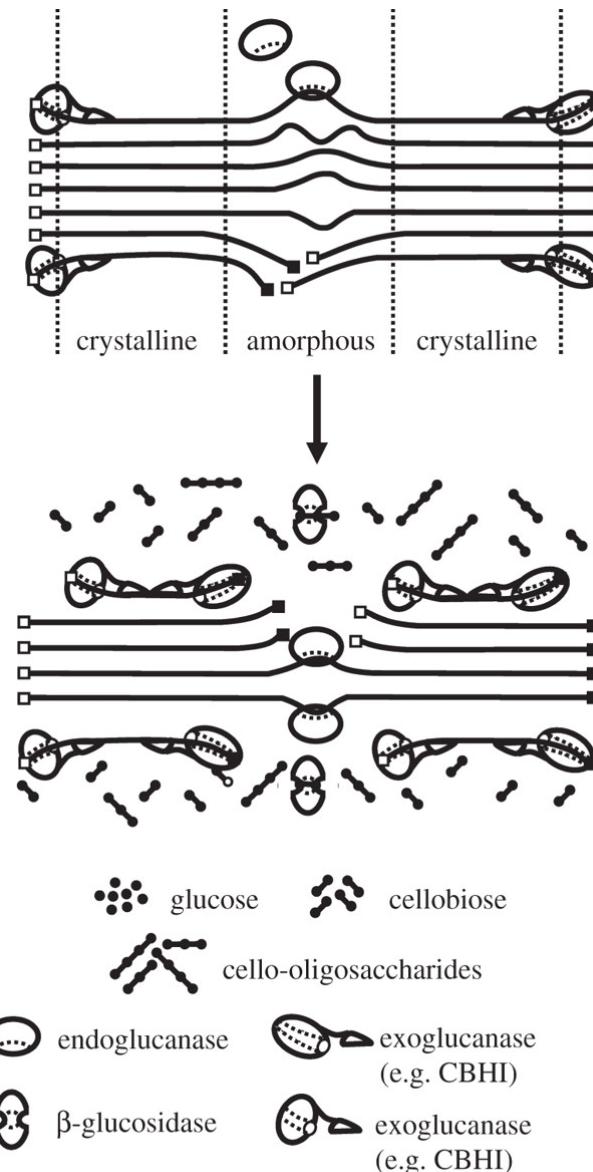
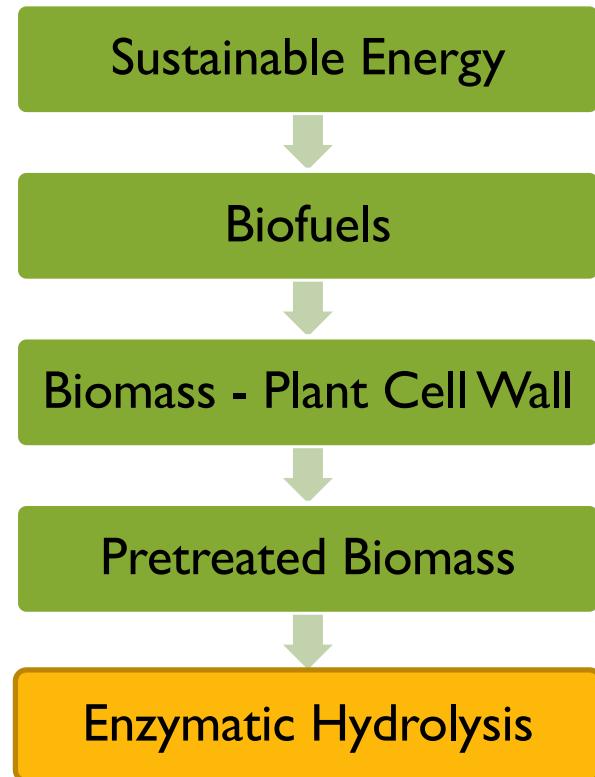
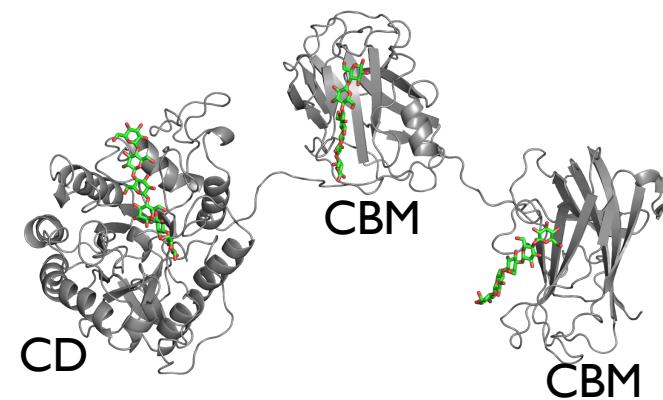
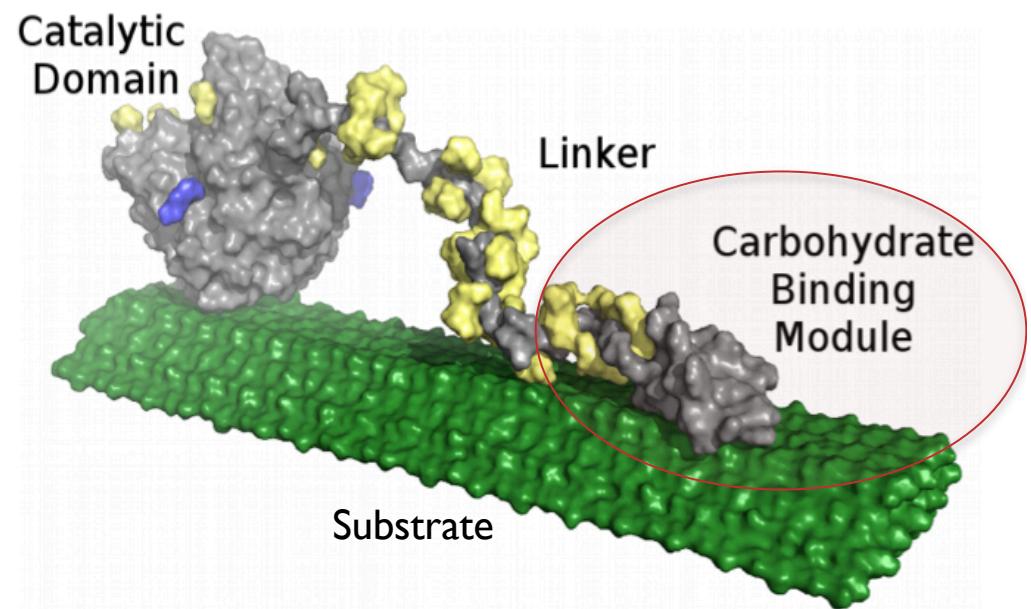
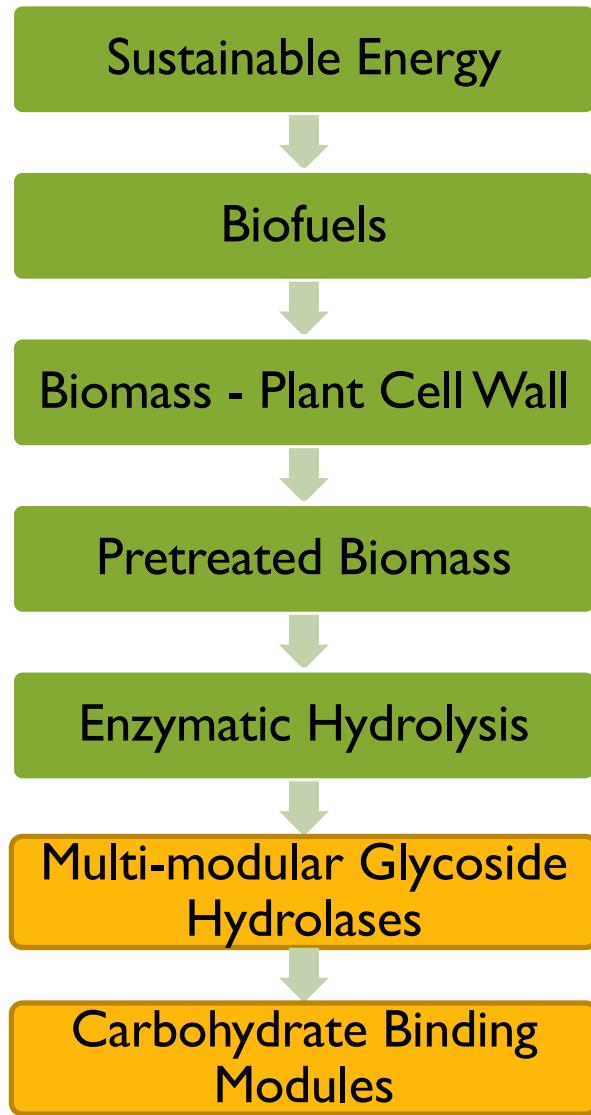
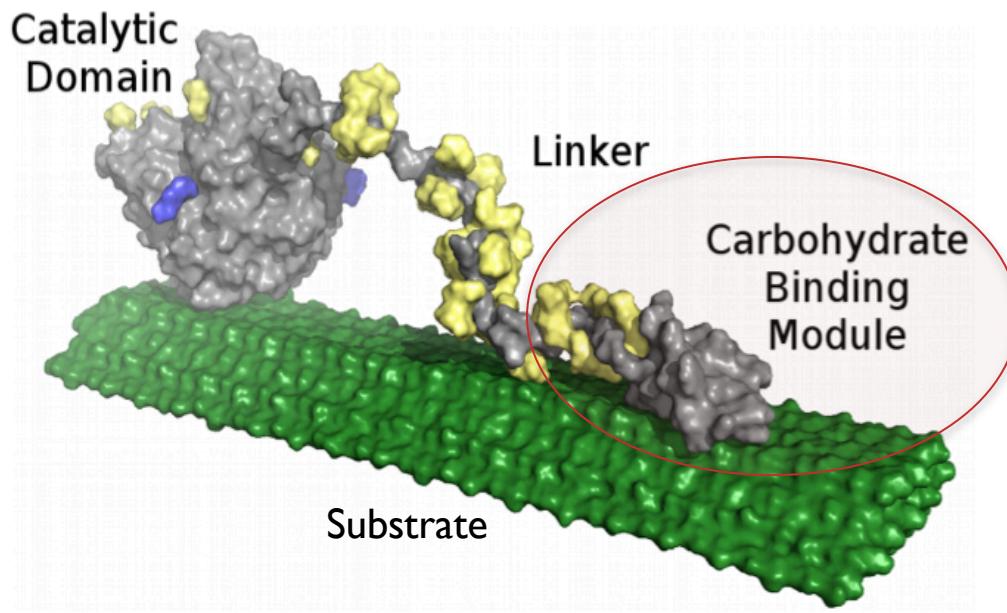


Image: Lynd et al., *Microbiology and Molecular Biology Reviews*, 2002.

Carbohydrate Binding Modules (CBMs)



Carbohydrate Binding Modules (CBMs)



Functions of CBM:

1. Maintain proximity to substrate
2. Target specific regions
3. Disrupt surface crystallinity

Other Applications:

Bioprocessing

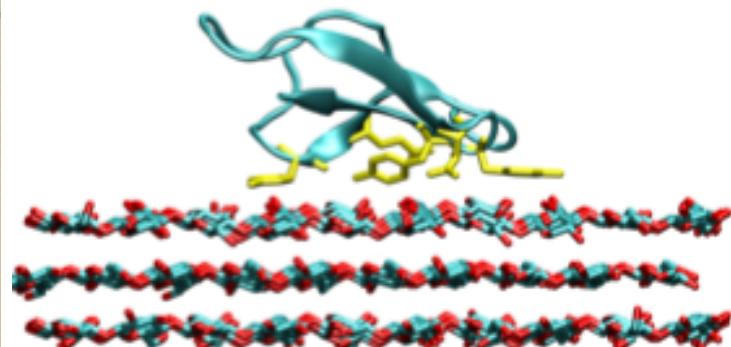
Cell immobilization

Protein Engineering

Different Substrates – Different Types

Type A

(crystalline polysaccharides)



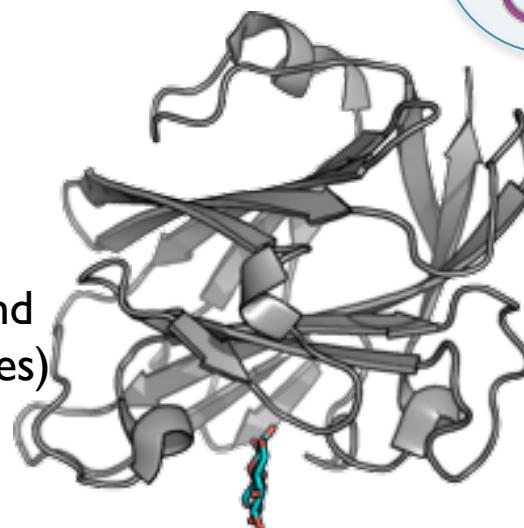
Type B

(single glycan chains)



Type C

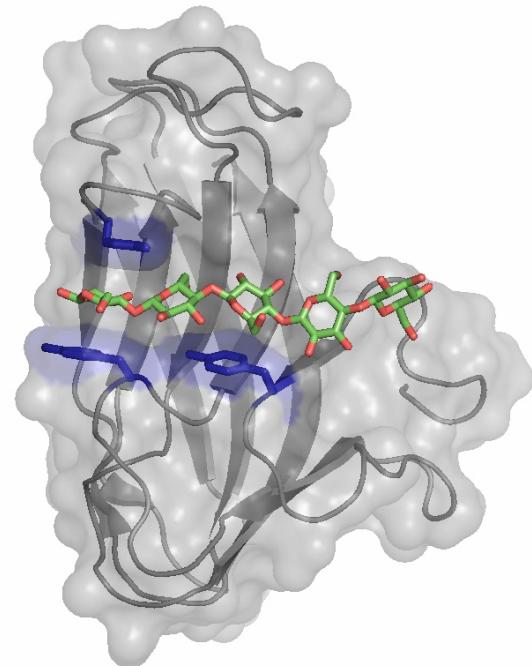
(glycan chain termini and
mono-, di-, tri-saccharides)



Boraston et al., *Biochem. J.*, 2004
Gilbert et al., *Curr. Opin. Struct. Biol.*, 2013

Type B Carbohydrate Binding Modules

- Common characteristics:
- Binding site in the form of groove or cleft
- β - sandwich protein fold
- Target on single glycan chains
 - Soluble oligosaccharides
 - Cellotetraose, cellopentaose, cellohexaose etc.
 - Non-crystalline/amorphous polysaccharides
 - Individual insoluble chains (longer than oligomers)
 - Convoluted insoluble chains
 - Partially decrystallized chains



Nomenclature of CBMs

Abbreviation of CBM from certain family is **CBM#**, where # is its family number

Types

Based on functional activity

3 types as A, B and C

Boraston et al., *Biochem. J.*, 2004

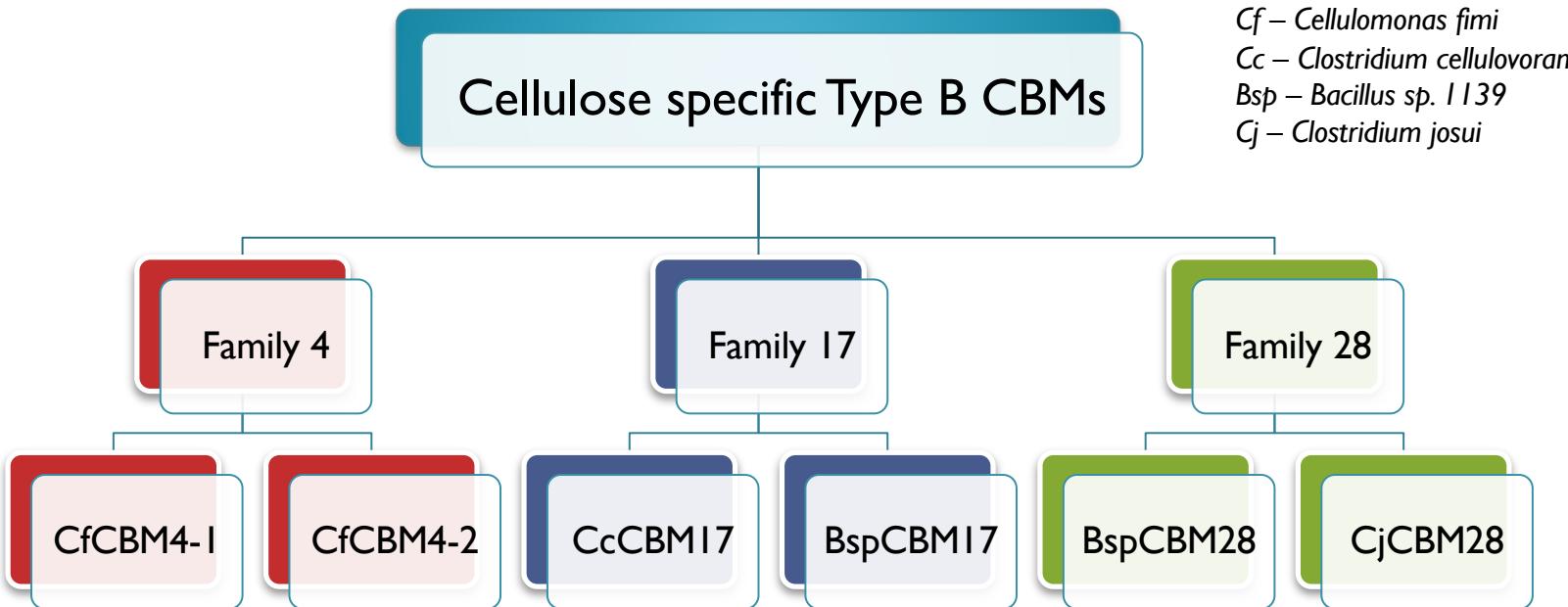
Families

Based on peptide sequence homology

Currently **81** families in database available

<http://www.cazy.org/>





Cf – *Cellulomonas fimi*
Cc – *Clostridium cellulovorans*
Bsp – *Bacillus sp.* 1139
Cj – *Clostridium josui*



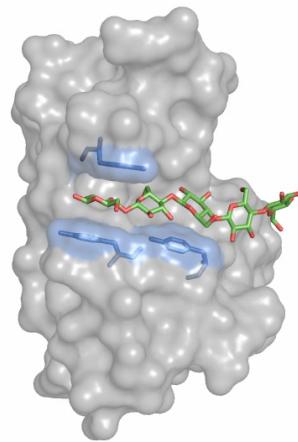
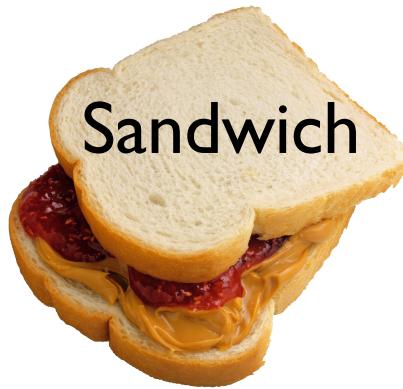
Three parts of the story...

Role of binding site architecture in oligomeric substrate recognition

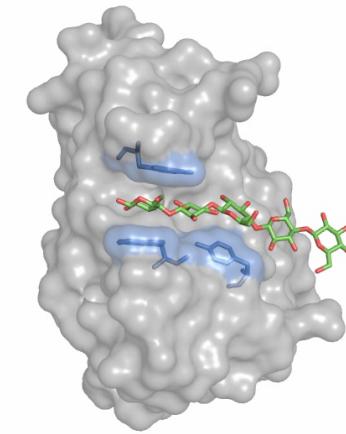
Bi-directional ligand binding in Type B CBMs

Non-crystalline substrate recognition with high- and low-affinity binding sites

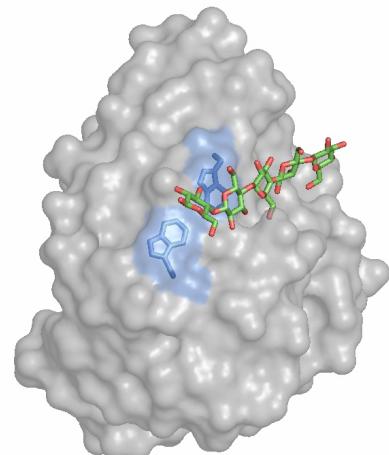
Role of binding site architecture in oligomeric recognition



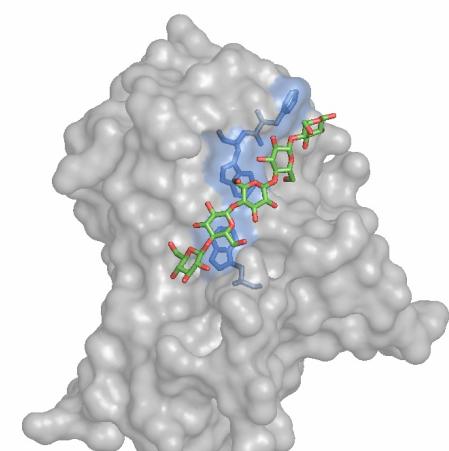
CfCBM4-1



CfCBM4-2



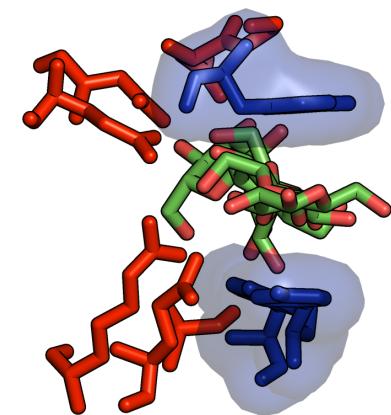
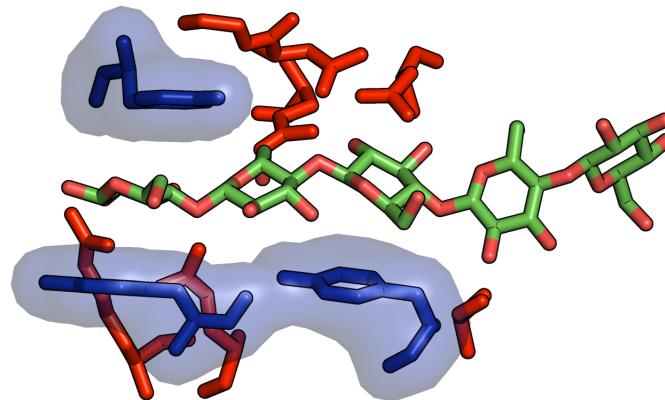
CcCBM17



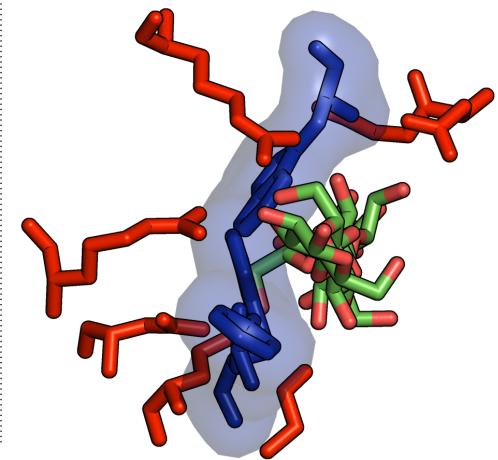
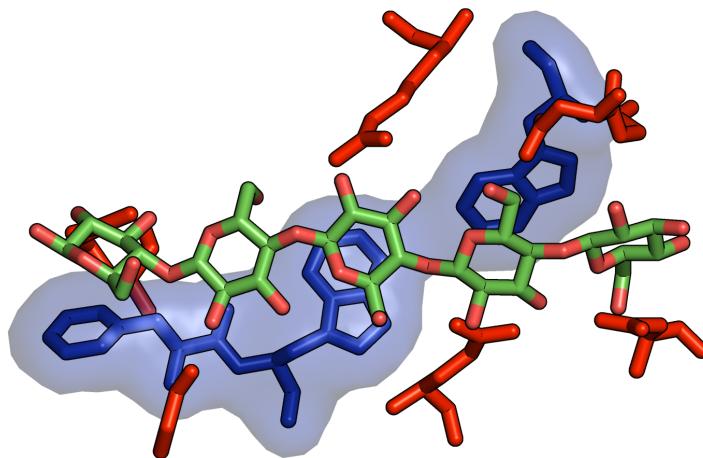
CjCBM28

Differences in binding site topology and hydrogen bonding patterns

Sandwich Platform



Twisted Platform

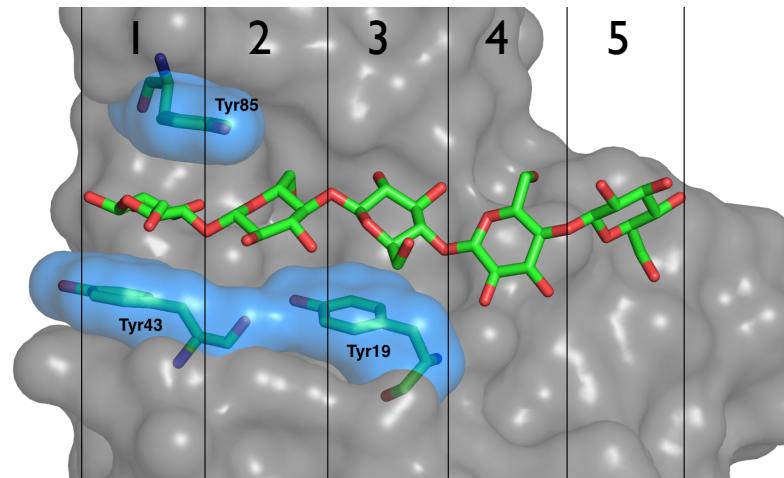


Ligand Binding Affinity

	ΔG (kcal/mol) of Cellooligosaccharide binding to	Experimental (ITC)	Computational (FEP/HREMD)
Sandwich Platform	CfCBM4-1	- 5.24 ± 0.9 ⁽¹⁾	- 4.5 ± 1.3 ⁽⁵⁾
	CfCBM4-2	- 5.80 ± 0.005 ⁽²⁾	- 5.4 ± 1.3
Twisted Platform	CcCBM17	- 5.8 ± 0.025 ⁽³⁾	- 6.9 ± 0.9
	CjCBM28	- 7.7 ± 0.6 ⁽⁴⁾	- 6.3 ± 0.7

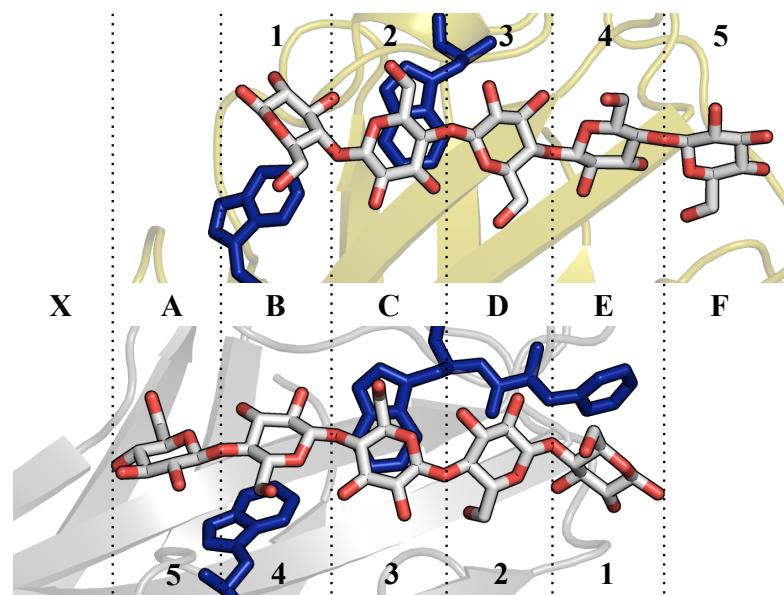
1. Tomme P, Creagh AL, Kilburn DG, and Haynes CA (1996) *Biochemistry*, **35**, 13885-13894.
2. Brun E, Johnson PE, Creagh AL, Tomme P, Webster P, Haynes CA, McIntosh LP (2000) *Biochemistry*, **39**(10), 2445-2458.
3. Notenboom V, Boraston AB, Chiu P, Freelove ACJ, Kilburn DG, Rose DR (2001) *J. Mol. Biol.*, **314**, 797-806.
4. Araki Y, Karita S, Tanaka A, Kondo M, and Goto M (2009) *Biosci. Biotechnol. Biochem.*, **73**(5), 1028-1032.
5. Kognole and Payne (2015) *Glycobiology*, **25**(10), 1100.

CfCBM4-I



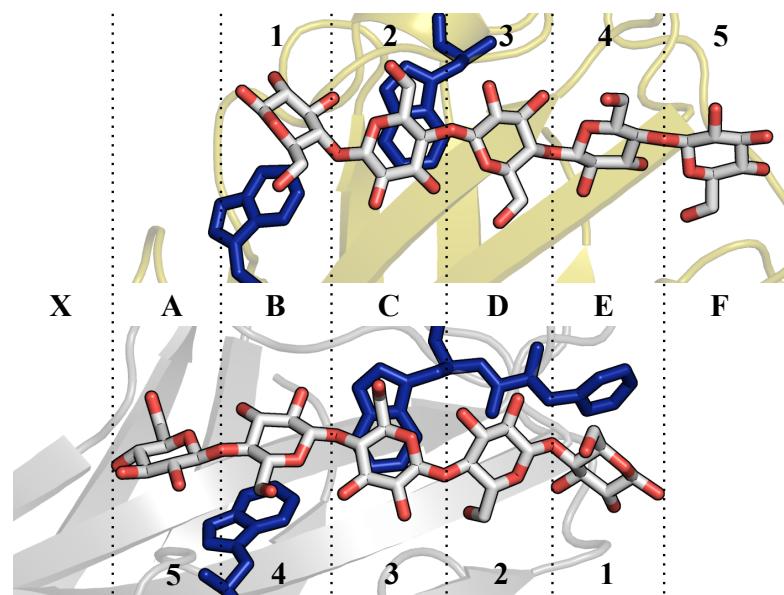
Highest affinity oligomer
- Cellopentaose

CcCBM17



Highest affinity oligomer
- Cellohexaose

CjCBM28



Extra sites available for
Celloheptaose binding

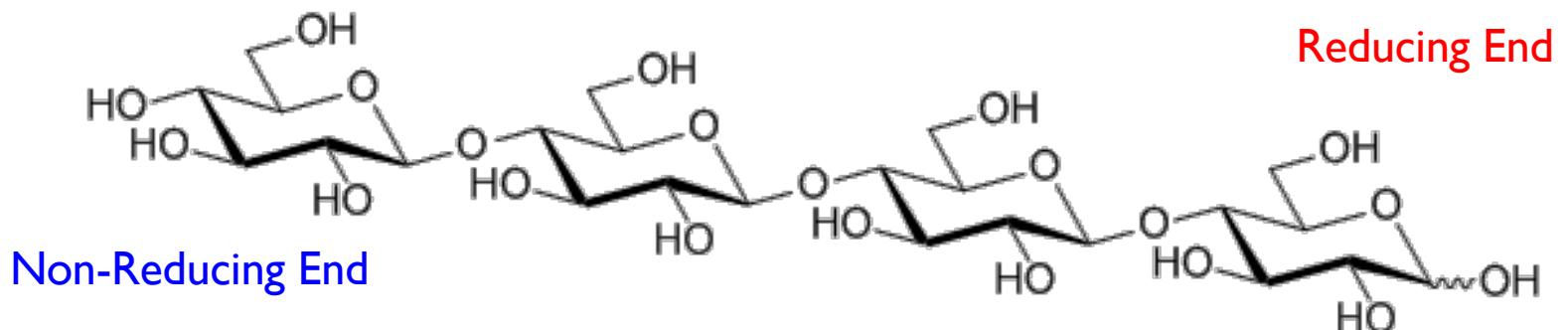
Conclusions – Sandwich vs Twisted

Open topology of twisted platform necessitates tighter binding of cello-oligomer as compared to closed sandwich platform.

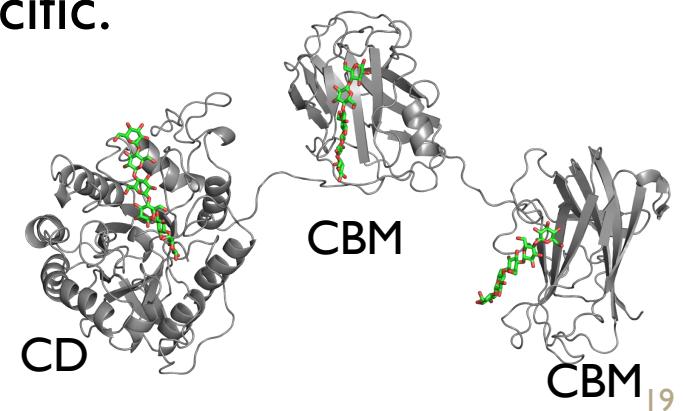
Higher number of and well distributed hydrogen bonding partners along the twisted platform contribute significantly to favorable free energy change.

The twisted binding site may extend further to accommodate longer cello-oligomers and, ultimately, insoluble polysaccharides.

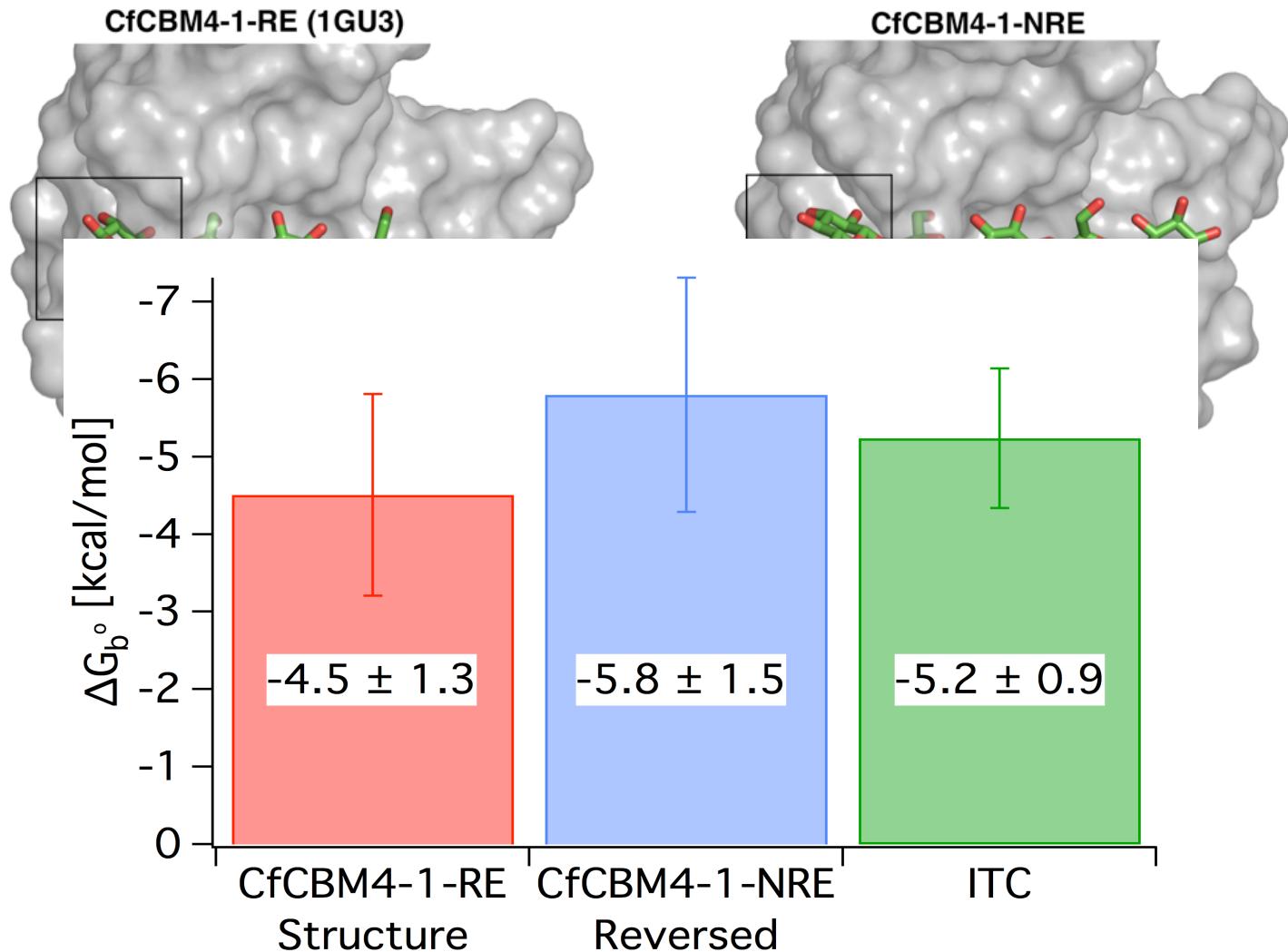
Bi-directional binding in Type B CBMs



- Glucose is a reducing sugar. The polysaccharides of glucose have one reducing end and one non-reducing end.
- Catalytic domains of glycoside hydrolases are either reducing end specific or non-reducing end specific.
- What about the non-catalytic CBMs?
Are they specific too?

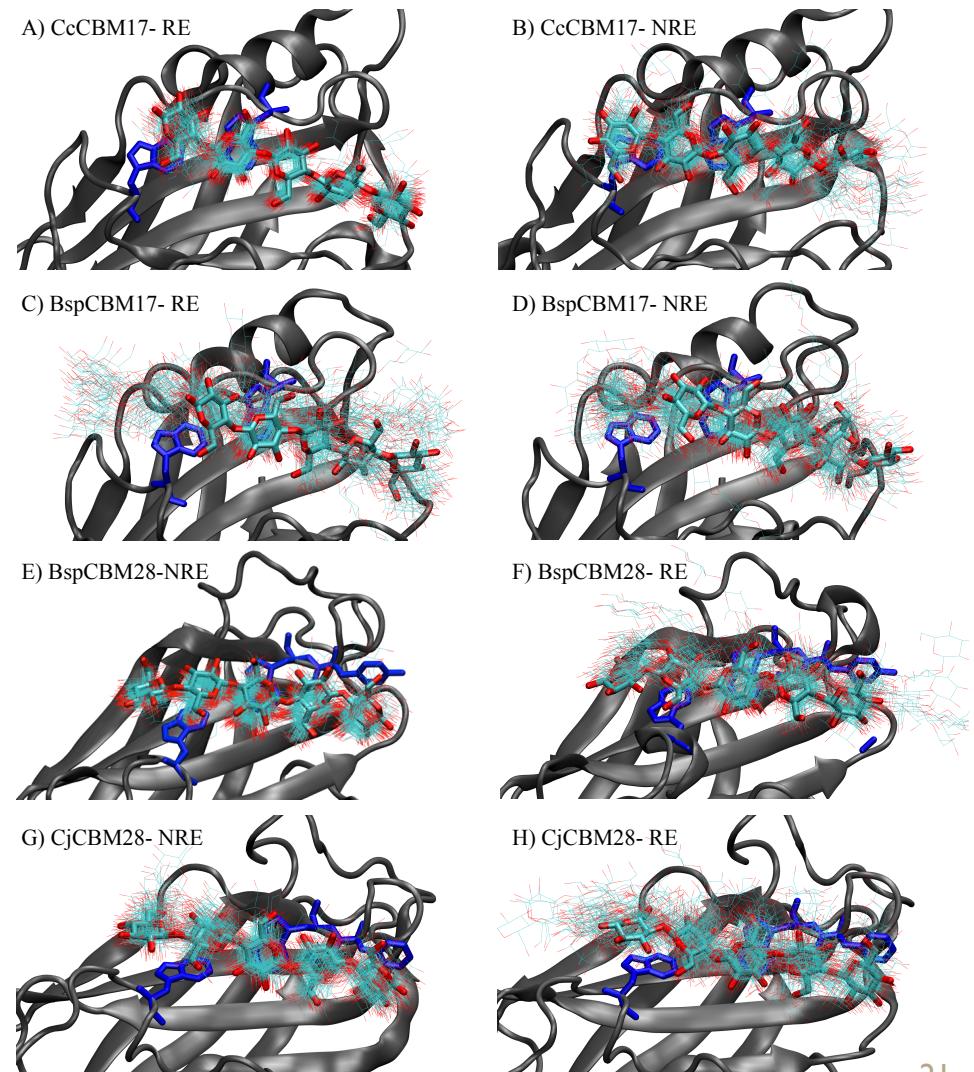


Bi-directional cello-oligomer binding in Family 4 CBMs



Bi-directional cello-oligomer binding extends to Family 17 and 28 CBMs

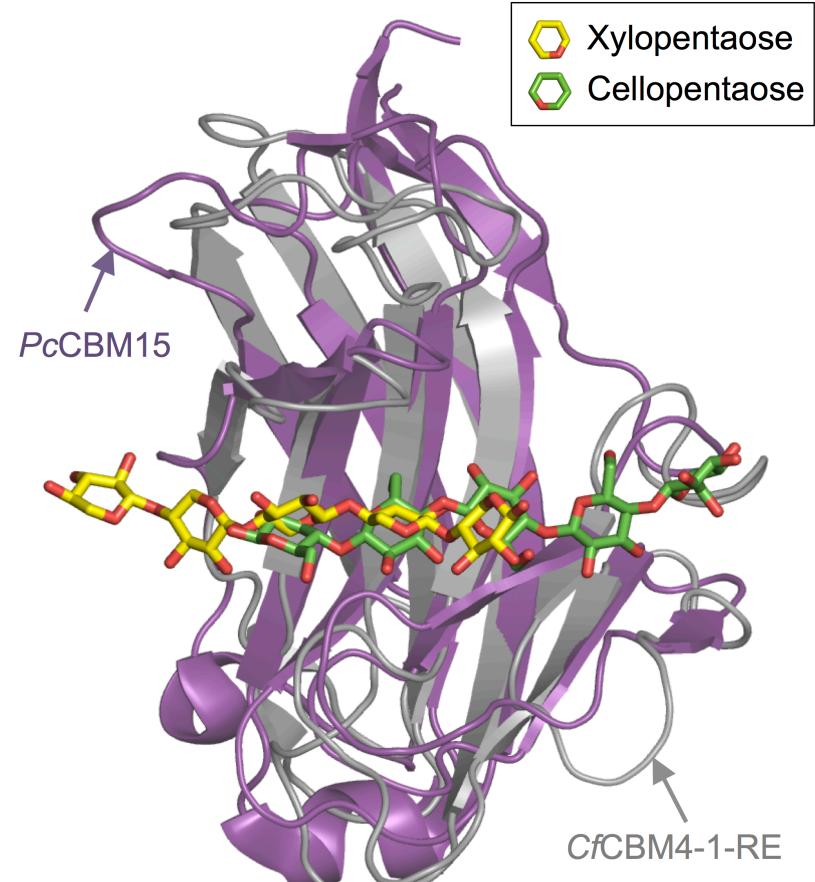
	<i>CfCBM4-1</i>
	<i>CfCBM4-2</i>
	<i>CcCBM17</i>
	<i>BspCBM17</i>
	<i>BspCBM28</i>
	<i>CjCBM28</i>



Note – ‘ β - sandwich’ is a type of protein fold, not same as a binding site architecture.

General to β -sandwich CBMs?

- 29 of the 69 CBM families demonstrate the β -sandwich protein fold
- 10 of these 29 families have glycan bound structures available (34 structures in total)
 - 22 structures bind the ligand in the **same** direction as IGU3
 - 12 structures bind the ligand in the **opposite** direction of IGU3



Gray: CfCBM4-I (IGU3)

Purple: *Pseudomonas cellulosa* CBM15 (IGNY)

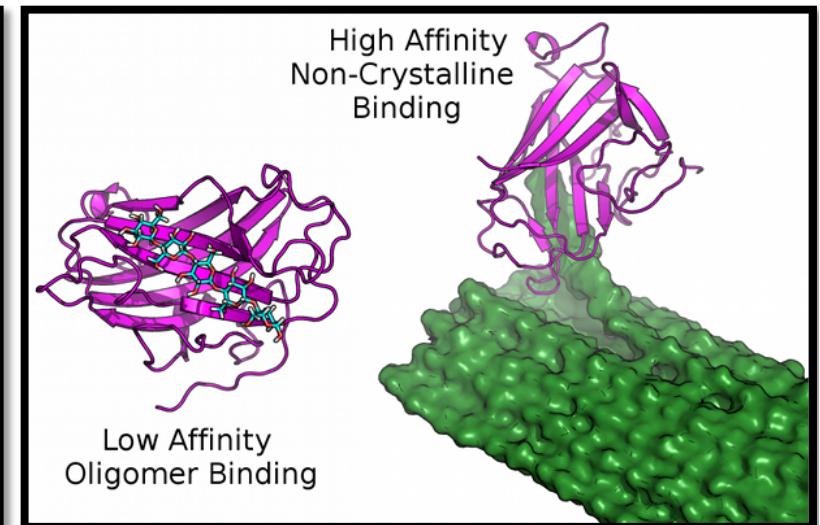
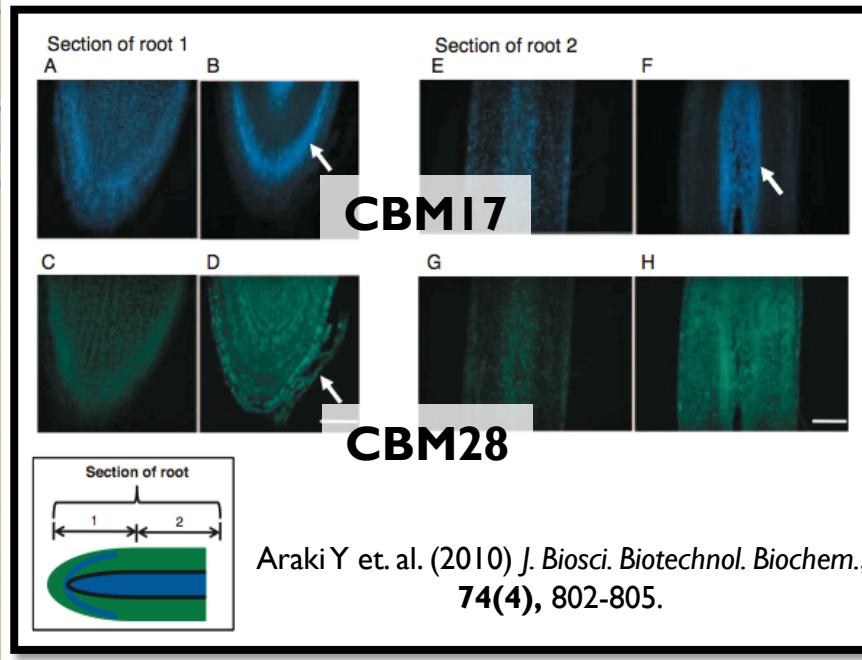
Conclusions – Bi-directionality

Cello-oligomers are recognized by family 4 CBMs in either orientation and there is no thermodynamic preference for reducing end.

We confirm that the bi-directional binding of cello-oligomers extends to twisted platform of family 17 and family 28 CBMs.

Bi-directional binding phenomenon may not be limited to only cellulose specific Type B CBMs, potentially generalizes to all β -sandwich CBMs.

Non-crystalline substrate recognition



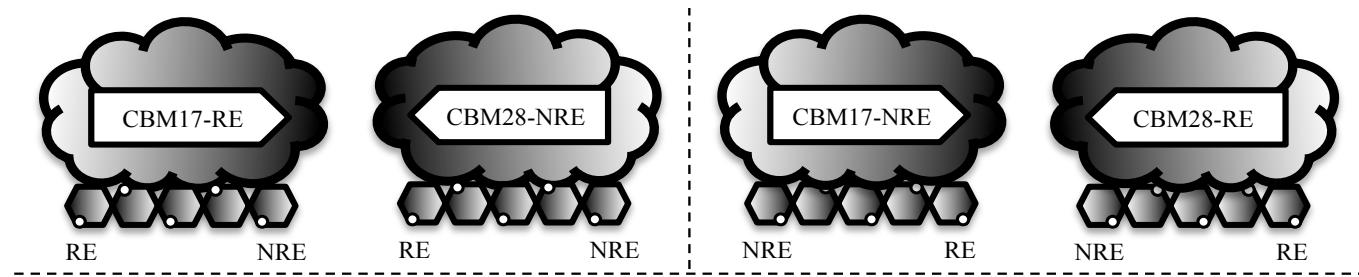
Hypothesis

TABLE I
Adsorption parameters for the binding of CcCBM17 and BspCBM28 to Avicel™ and RC in 50 mM potassium phosphate, pH 7.0, at 25 °C
 Errors represent the standard deviations of four binding experiments.

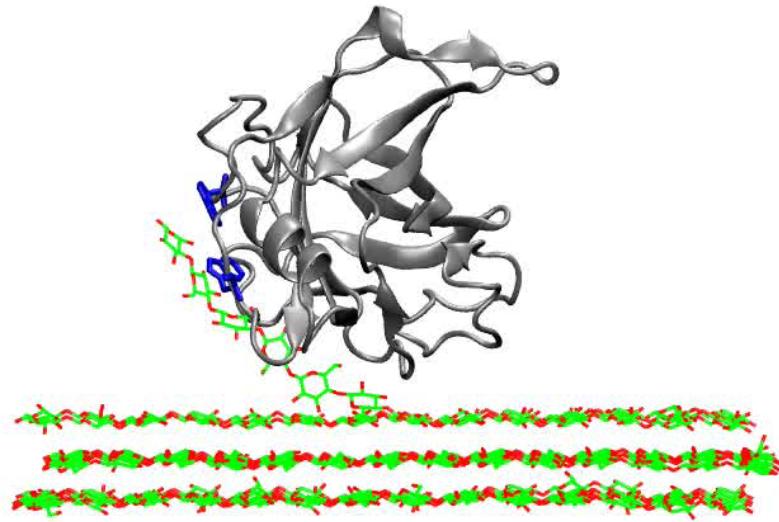
	Site 1			Site 2		
	K_{a1} $\times 10^5 \text{ M}^{-1}$	ΔG_1 kcal/mol	$[N_1]_o^a$ $\mu\text{mol/g}$	K_{a2} $\times 10^5 \text{ M}^{-1}$	ΔG_2 kcal/mol	$[N_2]_o^a$ $\mu\text{mol/g}$
<i>CcCBM17</i>						
Avicel	8.70 (± 4.20)	-8.04 (± 0.45)	0.26 (± 0.06)	0.07 (± 0.02)	-5.31 (± 0.37)	5.01 (± 0.88)
RC	11.30 (± 1.40)	-8.41 (± 0.32)	8.57 (± 0.52)	0.18 (± 0.05)	-5.88 (± 0.36)	15.92 (± 1.26)
<i>BspCBM28</i>						
Avicel	4.20 (± 1.30)	-7.72 (± 0.38)	0.08 (± 0.02)	0.20 (± 0.05)	-5.95 (± 0.36)	0.79 (± 0.05)
RC	9.90 (± 2.30)	-8.28 (± 0.35)	3.72 (± 0.36)	0.21 (± 0.07)	-5.93 (± 0.38)	6.84 (± 0.54)

^a $[N_x]_o$, binding capacity, density of binding sites per gram of cellulose. Boraston AB et. al. (2003) *J. Biol. Chem.*, **278(8)**, 6120-6127.

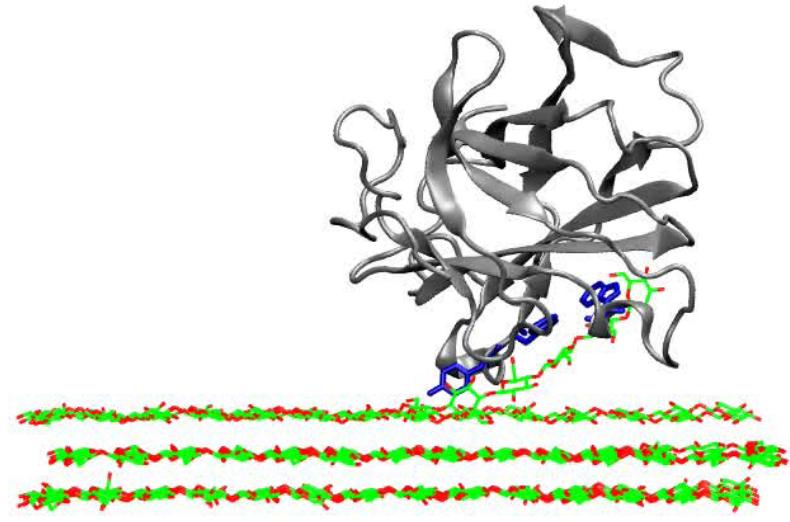
Modeling of CBMs on non-crystalline cellulose



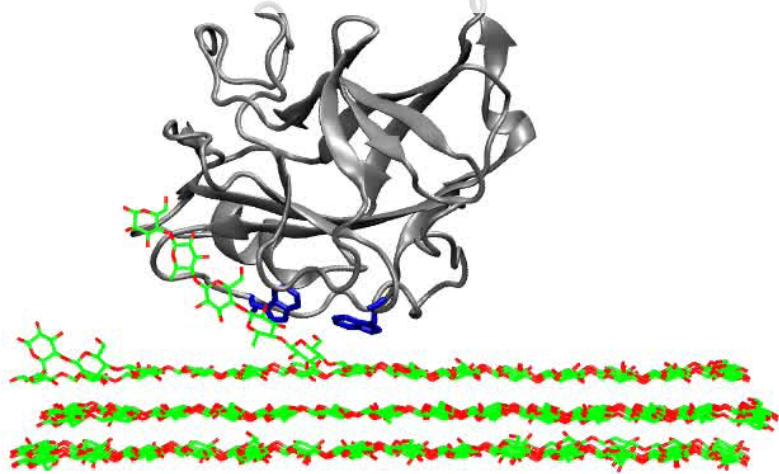
CcCBM17-Forward



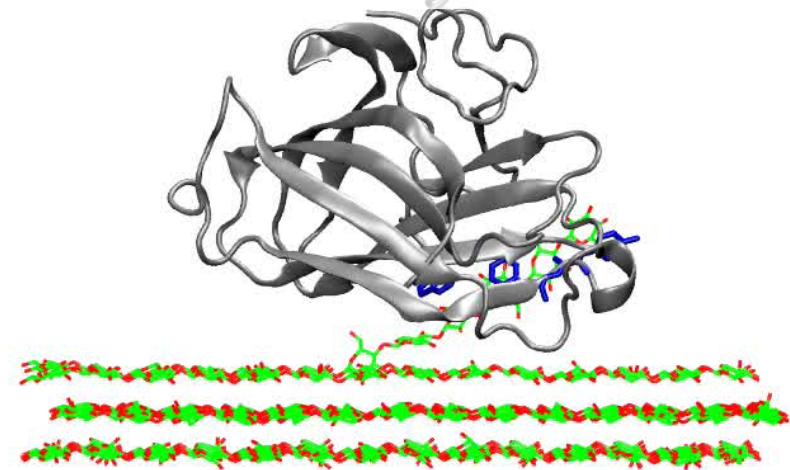
BspCBM28-Forward



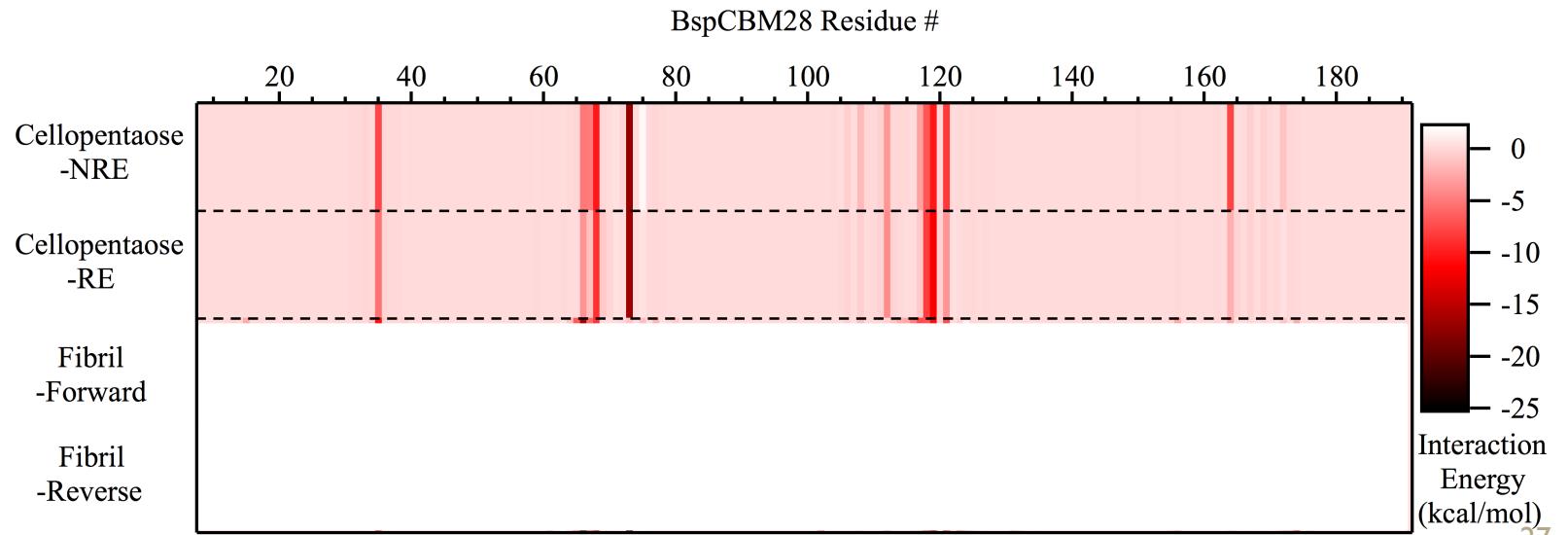
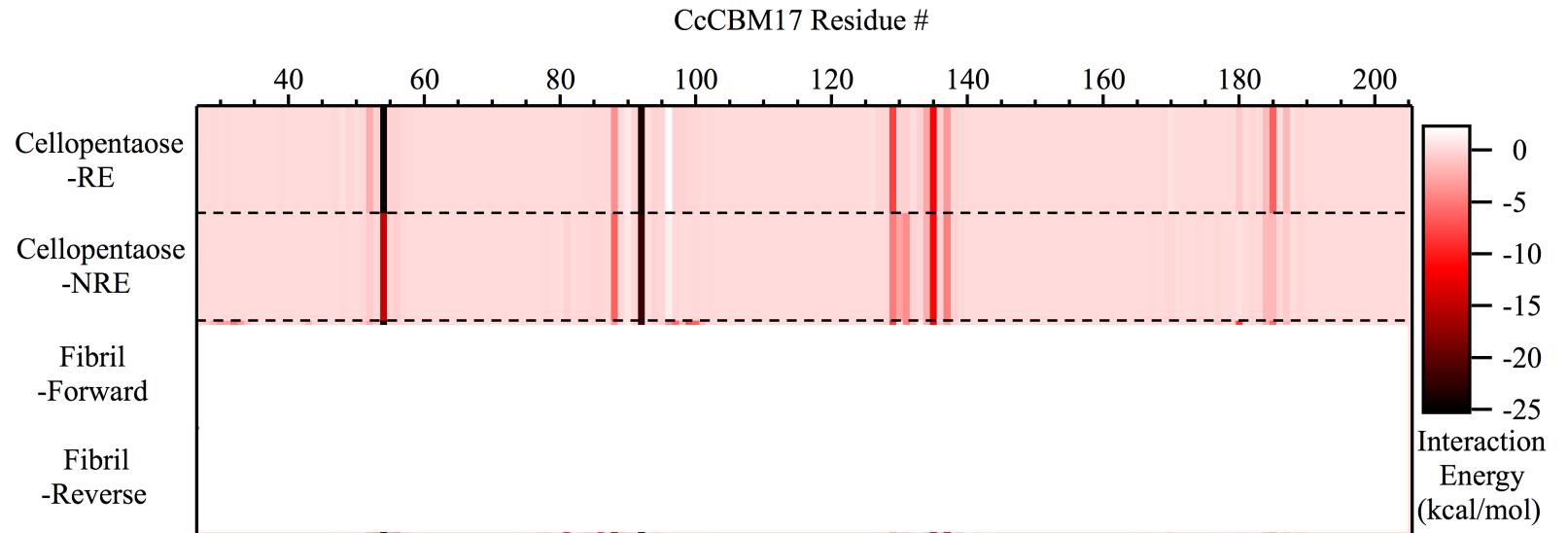
CcCBM17-Reverse



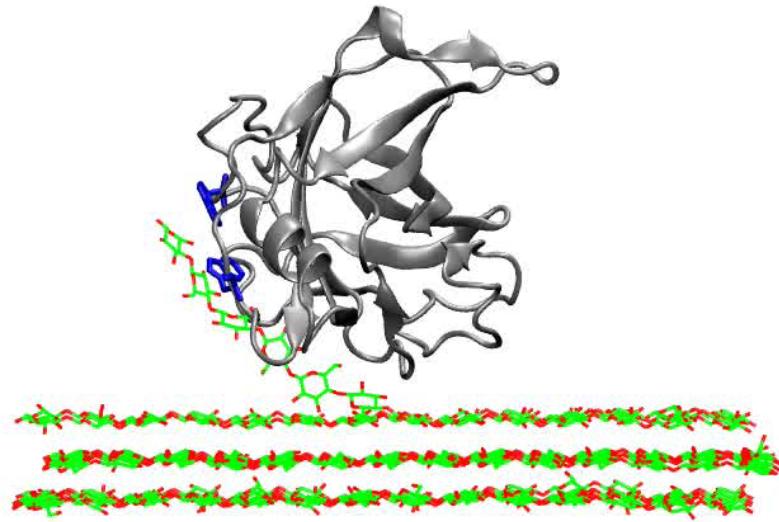
BspCBM28-Reverse



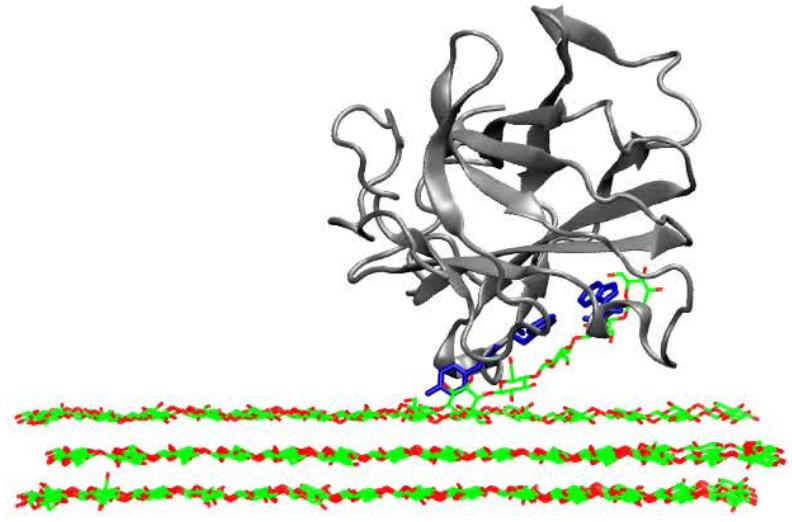
Total substrate interaction per CBM residue



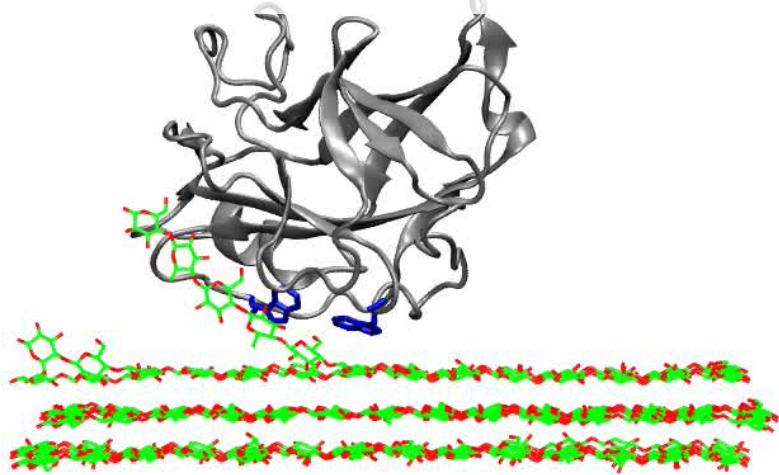
CcCBM17-Forward



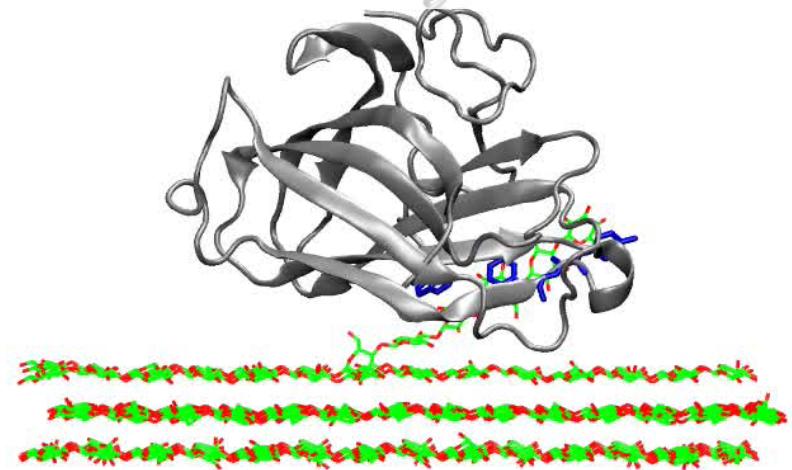
BspCBM28-Forward



CcCBM17-Reverse



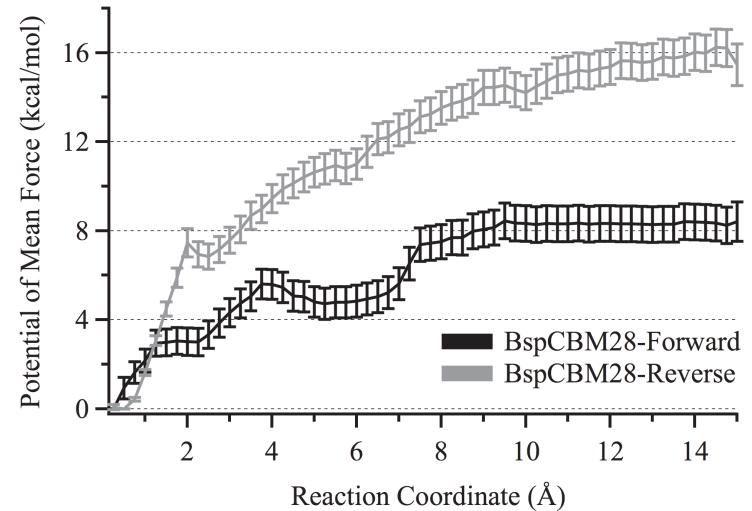
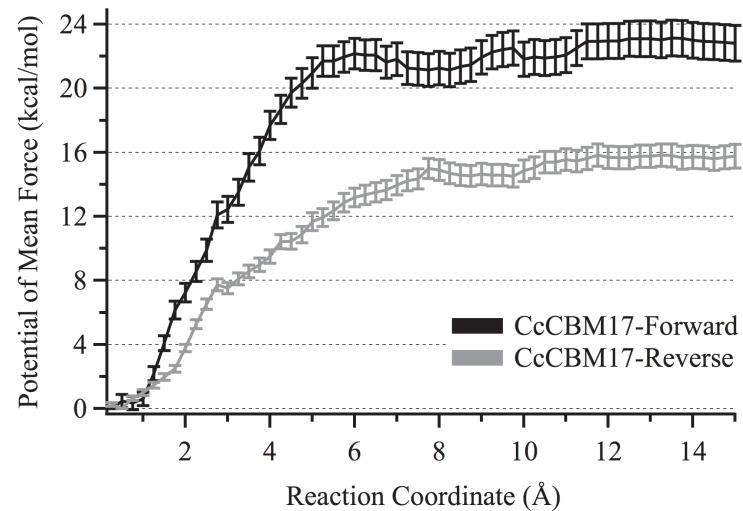
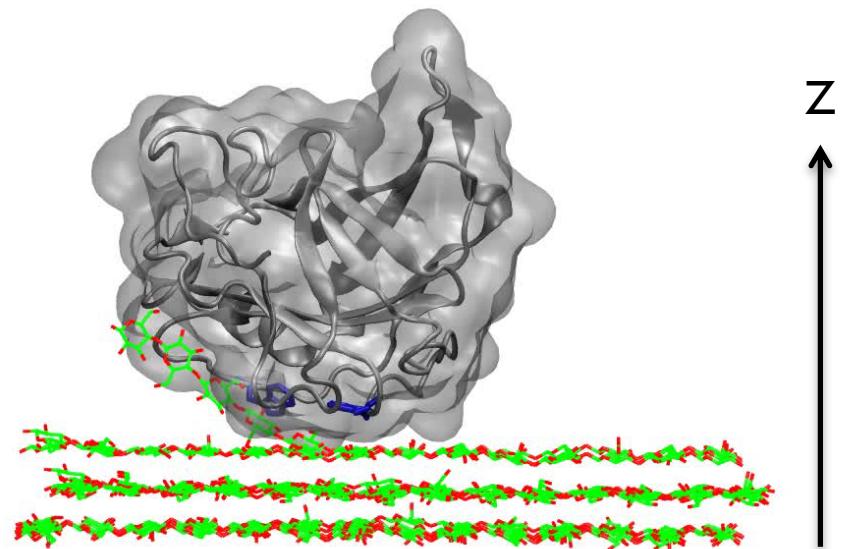
BspCBM28-Reverse



Umbrella sampling

Calculation of potential of mean force (PMF) to pull the CBM away from cellulose microfibril.

- Reaction coordinate is distance between projections of CBM and substrate on z-axis
- 30 windows of 0.5 Å each



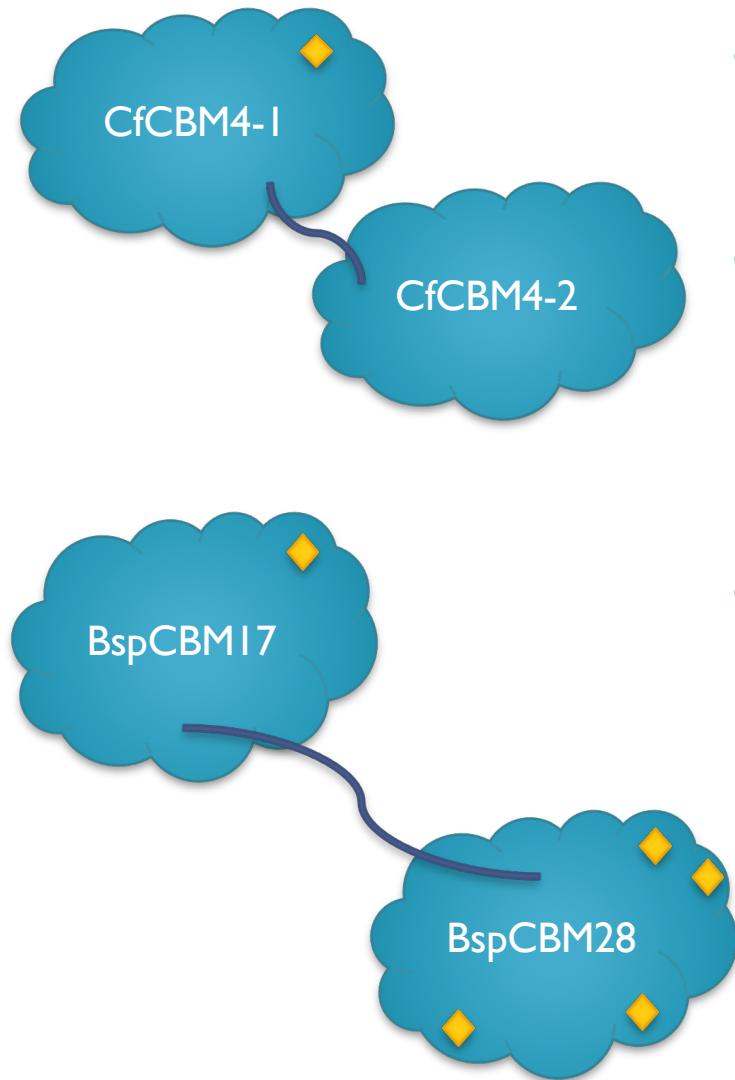
Conclusion – Non-crystalline binding

Non-crystalline recognition by Type B CBMs involves significant interactions of additional domains that are not involved in oligomeric recognition.

The high- and low-affinity binding sites for family 17 and 28 CBMs correspond to a range of non-crystalline substrate with increasing affinity as substrate approaches crystallinity.

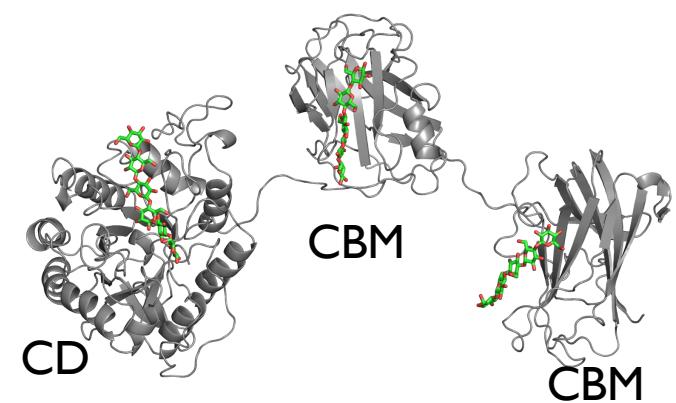
CBMs can have preferential affinity for certain substrate morphologies based on favorable binding orientations which, in turn, could result in uncompetitive binding.

Future work



Tandem CBMs

- Associate the individual recognition mechanisms and binding affinities with evolution of tandem CBM systems.
- Investigate the dynamics of tandem systems to uncover the effects of protein-protein networking and linker chain lengths on additive/co-operative binding.
- Understand mechanism of feeding the substrate to catalytic domain (CD).



Summary

For faster hydrolysis, the oligomeric and non-crystalline cellulose which is a significant part of pretreated biomass can be targeted specifically by harnessing the abilities of Type B CBMs to do so.

Results from bi-directional binding studies suggest how Type B CBMs may have evolved certain mechanisms to increase the frequency of recognizing the substrate.

Characterization of protein-carbohydrate/protein-protein interactions through molecular simulations holds huge potential to uncover crucial insights that may have been left unexplored. They have significant implications in fields like structure based drug discovery as well.

Acknowledgements

Advisor - Dr. Christina M. Payne

Payne lab members –

Dr. Suvamay Jana

Yue Yu

Dr. Inacrist Geronimo

Ingvild Isaksen

Japheth Gado

István Tamás

Committee Members –

Dr. Tate T. H. Tsang

Dr. Brad Berron

Dr. Matthew Gentry

Funding Source

Division of Chemistry
Award # 1404849



Computational Resources

CCS XSEDE

Center for
Computational Sciences

Extreme Science and Engineering
Discovery Environment

Thank You! see blue.
in everything we do.