

Malaria Parasite Prediction Project Report

November 2019

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Abstract

The aim of this project is to address the development of computer assisted malaria parasite prediction and classification using Machine learning approaches. We have applied a number of approaches (Traditional and Deep Learning) and Analysed their predictions. In the first approach, we have focussed on reducing noise by applying Gaussian Blurring on Grayscale images. We detected blob in the image by applying global thresholding and then passing it to the Decision Tree Model for prediction. In the second approach, we have applied Convolutional Neural Network on the thresholded images for making prediction. Also, we tried to improve by applying the VGG Pre-Trained Model for Image Classification.

1. Problem Statement

Given an image of a cell, the task is to predict whether it is parasitized or not. The images are publicly accessible from the following link:

https://www.kaggle.com/c/parasitedetection-iiitb2019/data

2. Dataset Description

- The dataset given comprises of two types of images: Uninfected and Parasitized
- Total images present in the dataset for training and validation purpose: 22046
- Total images present in the dataset for testing purpose: 1587
- There are 11023 infected and 11023 un-infected images in the training dataset.

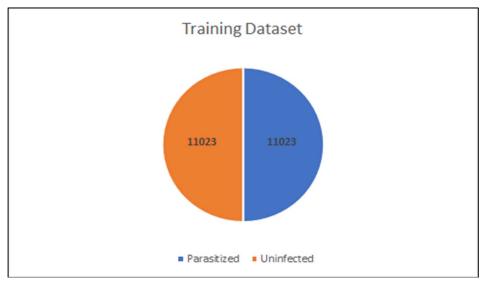


Figure 1: Distribution of Images in Training Dataset

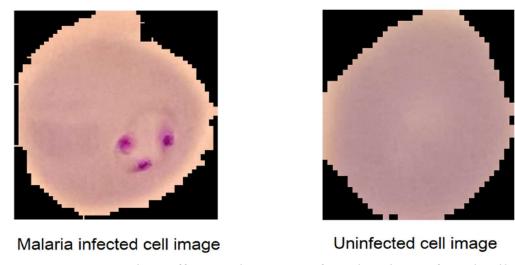


Figure 2: The Difference between Infected and Uninfected cells

3. Exploratory Data Analysis

Following are the manual observations from the dataset:

- Infected cells have a blob present in them which is having a higher intensity purple colour.
- Images are of different dimensions.



Figure 3: Images may have different dimensions

4. Pre-Processing and Feature Extraction

We divided all images in chunks of 100 images and then stored them in separate folders. For each folder, a CSV file is created with Image name, its features (black_stain = True or False) and Label. It is done to quickly load images in order to pre-process them while working on Jupyter Notebook locally.

```
divi...
           Ð
                      Save
                                  Open ▼
 1#! /bin/bash
 2 cd ./Parasitized/;
 3 i=0;
 4 for f in *;
       d=dir_$(printf %03d $((i/
  100+1)));
       mkdir -p $d;
       mv "$f" $d;
8
9
       i=$[$i+1];
10 done
Tab Width: 8 ▼
                  Ln 1, Col 1
```

Figure 4: Python Code Snippet for dividing images into chunks of 100

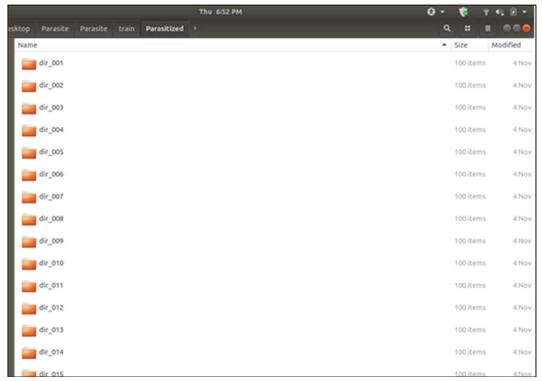


Figure 5: Directories, each having 100 images

	A	В	С
1	Name	black_stain	Label
2	C100P61ThinF_IMG_20150918_144823_cell_160.png	True	1
3	C101P62ThinF_IMG_20150918_151006_cell_70.png	True	1
4	C101P62ThinF_IMG_20150918_151006_cell_76.png	True	1
5	C101P62ThinF_IMG_20150918_151006_cell_63.png	True	1
6	C100P61ThinF_IMG_20150918_145938_cell_174.png	True	1
7	C101P62ThinF_IMG_20150918_151006_cell_72.png	True	1
8	C101P62ThinF_IMG_20150918_151149_cell_76.png	True	1
9	C101P62ThinF_IMG_20150918_151149_cell_72.png	True	1
10	C100P61ThinF_IMG_20150918_144348_cell_143.png	True	1
11	C101P62ThinF_IMG_20150918_151149_cell_82.png	True	1
12	C101P62ThinF_IMG_20150918_151006_cell_65.png	True	1
13	C100P61ThinF_IMG_20150918_144104_cell_168.png	True	1
14	C101P62ThinF_IMG_20150918_151239_cell_92.png	True	1
15	C101P62ThinF_IMG_20150918_151239_cell_75.png	True	1
16	C100P61ThinF_IMG_20150918_144348_cell_141.png	True	1
17	C100P61ThinF_IMG_20150918_144104_cell_162.png	True	1
18	C101P62ThinF_IMG_20150918_151239_cell_82.png	True	1
19	C100P61ThinF_IMG_20150918_144104_cell_170.png	True	1
20	C100P61ThinF_IMG_20150918_144823_cell_159.png	True	1
21	C100P61ThinF_IMG_20150918_144348_cell_139.png	True	1
22	C101P62ThinF_IMG_20150918_151239_cell_85.png	True	1
23	C101P62ThinF_IMG_20150918_151239_cell_100.png	True	1
24	C100P61ThinF_IMG_20150918_144348_cell_138.png	True	1
25	C100P61ThinF_IMG_20150918_145422_cell_167.png	True	1
26	C100P61ThinF_IMG_20150918_145609_cell_150.png	True	1
27	C101P62ThinF_IMG_20150918_151239_cell_90.png	True	1

Figure 6: CSV File containing 100 rows for each folder

Applied traditional Image Processing techniques-

1st Approach for Pre-processing:

• Applied Gaussian Blurring: Used it to remove the noise from the image.

```
# smoothing
blur = cv2.GaussianBlur(img, (5, 5), 0)
```

Figure 7: Code Snippet for Gaussian Blurring



Figure 8 : Image after applying Gaussian Blur

• Converted the Images to Grayscale: Inherent complexity of gray level images is lower than that of coloured images. Also, we are interested in the intensity of the images, and Grayscale images are used for measuring the intensity of light in images.

```
gray img = cv2.cvtColor(blur, cv2.COLOR BGR2GRAY)
```

Figure 9: Code Snippet for Gray-Scaling



Figure 10: Image after Gray-Scaling

• **Applied Global Thresholding:** For separating out the black stain from the image as shown in the figure below.

```
ret, th1 = cv2.threshold(gray img, 127, 255, cv2.THRESH BINARY)
```

Figure 11: Code Snippet for Global Thresholding

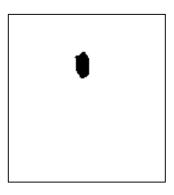


Figure 12: Image after Thresholding

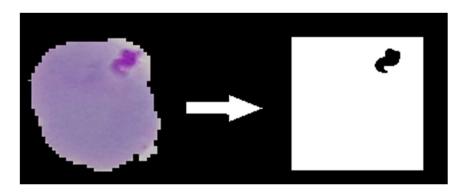


Figure 13: Pre-processing the images

• Create histogram of Black and White intensities for images, based on which we append the images with a black stain into list 'black_stain_list'. We use this as a feature that we can pass to the Decision Tree Model as explained earlier.

```
(n, bins, pathces) = plt.hist(th1.ravel(), bins=255)
just_fname = ntpath.basename(img_filename)
if(n[0] > 0.0):
    fname_list.append(just_fname)
    black_stain_list.append(True)
else:
    fname_list.append(just_fname)
    black_stain_list.append(False)
```

Figure 14: Create Histogram and Bins

• We **train Decision Tree Classifier** on these images and the stain as a feature to make predictions for parasitized or non-parasitized Malaria Cells.

2nd Approach for Pre-processing:

• Applied SIFT to extract features

```
def get_sift(images, name='sift'):
   img_path_list = list(images)
    # SIFT descriptor for 1 image
    def get_image_sift(image, vector_size=15):
        alg = cv2.xfeatures2d.SIFT_create()
       kps = alg.detect(image, None)
        kps = sorted(kps, key=lambda x: -x.response)[:vector_size]
        # Making descriptor of same size
        # Descriptor vector size is 128
        needed_size = (vector_size * 128)
        if len(kps) == 0:
           return np.zeros(needed_size)
        kps, dsc = alg.compute(image, kps)
        dsc = dsc.flatten()
        if dsc.size < needed size:</pre>
            dsc = np.concatenate([dsc, np.zeros(needed_size - dsc.size)])
       return dsc
    # SIFT descriptor for all images
    features = []
    for img in img_path_list:
       image_arr = cv2.imread(img)
       dsc = get_image_sift(image_arr)
       {\tt features.append(dsc)}
    result = np.array(features)
    return result
```

Figure 15: Code Snippet for SIFT

Applied PCA on the features extracted from SIFT.

```
pca = PCA(n_components=5)
pca_sift_train = pca.fit_transform(norm_sift_train)
pca_sift_val = pca.transform(norm_sift_val)
```

Figure 16: PCA Applied on Feature Matrix Obtained by SIFT

5. Model Building

We first split the dataset into train and test sets and then apply the following models:

• We have applied **Decision Trees** on the images obtained after applying the approach described under section 5.1 (Thresholded images). The reason for applying decision trees is that we have only one feature and accuracy obtained is also considerably good.

```
1 from sklearn.tree import DecisionTreeClassifier
1 model = DecisionTreeClassifier()
1 model.fit(feature_data,labeled_data)
DecisionTreeClassifier(class_weight=None, criterion='gini', max_depth=None, max_features=None, max_leaf_nodes=None, min_impurity_decrease=0.0, min_impurity_split=None, min_samples_leaf=1, min_samples_split=2, min_weight_fraction_leaf=0.0, presort=False, random_state=None, splitter='best')
```

Figure 17: Code Snippet for Decision Tree Model

 We have applied CNN Model (Deep Learning Approach) on the original dataset (after resizing all the images to same dimensions).

```
1 model = Sequential()
2
3 model.add(Conv2D(16, (3,3), input_shape = (img_width, img_height, 3), activation='relu'))
4 model.add(MaxPool2D(2,2))
5 model.add(Dropout(0.2))
6
7 model.add(Conv2D(32, (3,3), activation='relu'))
8 model.add(MaxPool2D(2,2))
9 model.add(Dropout(0.3))
10
11 model.add(Flatten())
12 model.add(Dense(64, activation='relu'))
13 model.add(Dropout(0.5))
14
15 model.add(Dense(1, activation='sigmoid'))
```

Figure 18: Code Snippet for CNN Model

Model: "sequential"		
Layer (type)	Output Shape	Param #
conv2d (Conv2D)	(None, 62, 62, 16)	448
max_pooling2d (MaxPooling2D)	(None, 31, 31, 16)	0
dropout (Dropout)	(None, 31, 31, 16)	0
conv2d_1 (Conv2D)	(None, 29, 29, 32)	4640
max_pooling2d_1 (MaxPooling2	(None, 14, 14, 32)	0
dropout_1 (Dropout)	(None, 14, 14, 32)	0
flatten (Flatten)	(None, 6272)	0
dense (Dense)	(None, 64)	401472
dropout_2 (Dropout)	(None, 64)	0
dense_1 (Dense)	(None, 1)	65
Total params: 406,625 Trainable params: 406,625 Non-trainable params: 0		

Figure 19: CNN Model

• We have applied VGG Model (Pre-Trained Model) on the dataset. VGG has been shown to perform well on complex datasets like the ImageNet dataset which has over 1000 categories. Since ours is a much simpler dataset, we decided to apply VGG on our dataset.

```
1 from keras.applications.vgg16 import VGG16
2 model = VGG16(input_shape=(64, 64, 3), classes=1, weights=None)
```

Figure 20: Code Snippet for VGG Model

Layer (type)	Output Shape	Param #
input 1 (InputLayer)	(None, 64, 64, 3)	0
block1_conv1 (Conv2D)	(None, 64, 64, 64)	1792
block1_conv2 (Conv2D)	(None, 64, 64, 64)	36928
block1_pool (MaxPooling2D)	(None, 32, 32, 64)	0
block2_conv1 (Conv2D)	(None, 32, 32, 128)	73856
block2_conv2 (Conv2D)	(None, 32, 32, 128)	147584
block2_pool (MaxPooling2D)	(None, 16, 16, 128)	0
block3_conv1 (Conv2D)	(None, 16, 16, 256)	295168
block3_conv2 (Conv2D)	(None, 16, 16, 256)	590080
block3_conv3 (Conv2D)	(None, 16, 16, 256)	590080
block3_pool (MaxPooling2D)	(None, 8, 8, 256)	0
block4_conv1 (Conv2D)	(None, 8, 8, 512)	1180160
block4_conv2 (Conv2D)	(None, 8, 8, 512)	2359808
block4_conv3 (Conv2D)	(None, 8, 8, 512)	2359808
block4_pool (MaxPooling2D)	(None, 4, 4, 512)	0
block5_conv1 (Conv2D)	(None, 4, 4, 512)	2359808
block5_conv2 (Conv2D)	(None, 4, 4, 512)	2359808
block5_conv3 (Conv2D)	(None, 4, 4, 512)	2359808
block5_pool (MaxPooling2D)	(None, 2, 2, 512)	0
flatten (Flatten)	(None, 2048)	0
fcl (Dense)	(None, 4096)	8392704
fc2 (Dense)	(None, 4096)	16781312
predictions (Dense)	(None, 1)	4097
Total params: 39,892,801 Trainable params: 39,892,801 Non-trainable params: 0		

Figure 21: VGG Model

6. Observations and Comparison of Models

Approaches	Validation Accuracy
Thresholding + Decision Tree	92.53%
SIFT + Random Forest	89.13%
SIFT + Decision Tree	83.94%
SIFT + Naïve Bayes	82.42%
CNN	94.42%
VGG	50.00%

Results obtained from different approaches:

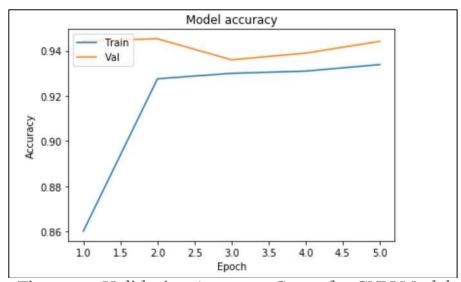


Figure 22: Validation Accuracy Curve for CNN Model

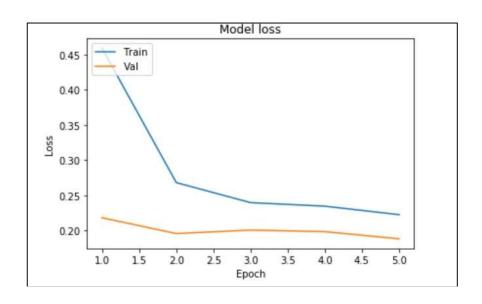


Figure 23: Validation Loss Curve for CNN Model

Figure 24: Snippet for Validation Accuracy and Loss of VGG Model.

Figure 25: Snippet for Validation Accuracy and Loss for CNN Model.

```
Validation Accuracy in 'DT' = 0.8394557823129252

[[1830 347]

[ 361 1872]]

Recall in 'DT' = 0.8406063389986219

Precision in 'DT' = 0.8352350524874487

F1 Score in 'DT' = 0.8379120879120879
```

Figure 26: Validation Accuracy for Decision Tree using SIFT.

```
Validation Accuracy in 'NB' = 0.8242630385487528
[[1685 492]
  [ 283 1950]]
Recall in 'NB' = 0.7740009186954525
Precision in 'NB' = 0.8561991869918699
F1 Score in 'NB' = 0.8130277442702052
```

Figure 27: Validation Accuracy for Naive Bayes using SIFT.

```
Validation Accuracy in 'RF' = 0.8913832199546485
[[2004 173]
  [ 306 1927]]
Recall in 'RF' = 0.9205328433624254
Precision in 'RF' = 0.8675324675324675
F1 Score in 'RF' = 0.8932471584577668
```

Figure 28: Validation Accuracy for Random Forest using SIFT.