Binding Affinity Prediction of Protein-Ligand complexes using Machine Learning

MSc Project

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Biological Background

What are proteins and ligands?

- **Proteins:** Complex molecules that are work-horses (machines) of a living organism.
- Ligands: Molecules that bind to (receptor) proteins.
- Proteins and ligands bind together to form protein-ligand complexes.







Biological Background

Protein-Ligand complexes

- Ligands bind to proteins at "cavity" like locations called pockets.
- The pockets and the ligands are complementary in shape.

Drugs

- Drugs are ligand molecules that bind to proteins.
- They cause a therapeutic effect after binding to the proteins.







Biological Background

Protein-Binding Affinity

- Assume a dynamic system in which protein P and ligand L are binding and unbinding continuously.
- Let [P] be the concentration of the protein and [L] be the concentration of the ligand. Let [PL] be the concentration of the protein ligand complex.
- Binding affinity can be quantified by using [P], [L] and [PL] (at equilibrium).

BindingAffinity =
$$\frac{[P][L]}{[PL]}$$



Problem Definition

- Determining if a potential drug (ligand) can bind to a target protein is very expensive [3].
- The project tries to reduce the drug discovery costs by eliminating bad leads.
- Problem definition: Predict protein-ligand binding affinity using "In-Silico" methods.



Figure: Project Overview

Problem Definition

There are various problems in the protein-ligand domain. The following figure shows the classification tree.

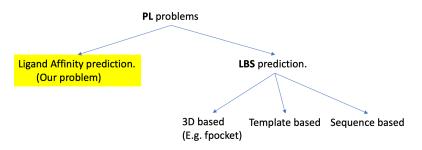


Figure: Protein-Ligand problem classifcation.



Problem Definition

- The input data to the ML model is extracted from a database called PDB Data bank.
- fpocket and RDKit were used to extract the features of proteins and ligands.
- The input features contain information about the 3D structures of the proteins and the ligands.

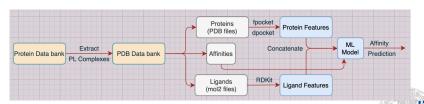


Figure: Data Input Overview.

Feature Extraction

Protein Features:

- fpocket is an LBS prediction algorithm used to predict ligand binding pockets.
- There can be multiple binding pockets for a PL complex.
- Using *dpocket*, 55 descriptors were obtained for every (potentially) binding pocket as real values.

Ligand Features:

 Using RDKit. Chem. Descriptors module, 402 features were extracted as real values.

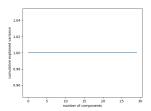
Concatenation:

- The (concatinated) input feature space to the model was
- R⁴⁵⁷.

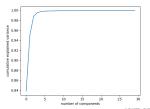
 It was less than R⁴⁵⁷ if feature selection is done before model. training.

Data Preprocessing

- Data points containing NaN (Not a number) values were removed from the data.
- PCA (Principle Component Analysis) was used to find the variance contribution of the features.
- Feature IPC was log scaled for numerical safety during training.



(a) With original Ligand feature IPC.



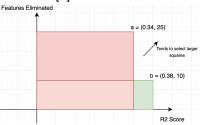
(b) With log scaled Ligand feature IPC.



Feature Selection

Feature selection strategies:

- Manual: 121 ligand descriptors + all protein descriptors.
- Output Correlation: Features with the best Pearson or Spearman correlation w.r.t the affinity score (output) were selected.
- **Genetic Algorithms**: Genetic algorithms with a population score function "**R**2 score * Features Eliminated" was used to select the best features [4]:

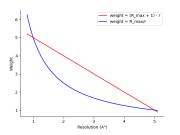




Dealing with measurement resolution

- In PDB bind databank, each complex has a corresponding measurement resolution (Å units).
- The structural detail of the 3D image is inversely proportional to the measurement resolution.
- The weighting of each data point was done according to hyperbolic formulae (or) linear formulae:

$$W_{\text{Hyperbolic}} = \frac{\max R_{1...n}}{R_i} \quad W_{\text{Linear}} = (\max R_{1...n} + 1) - R_i$$

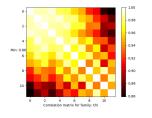






Feature Family Correlations

- Features can be divided into families.
- Important ones are AUTOCORR2d_, Chi, EState_VSA, PEOE_VSA, SMR_VSA, SlogP_VSA, VSA_EState, and fr_.
- Within Chi and AUTOCORR2d_, the features are correlated.
- ML models need to take into account this issue.



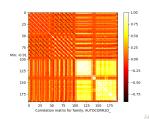


Figure: Correlation Heat Map.

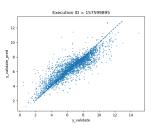
Machine Learning models

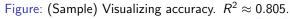


Testing strategy

For every execution, we report the **random seed** used in it. This random seed can be used as a script argument (Execution ID) to reproduce the exact results. To determine the result quality we use:

- R^2 score (Coefficient of determination): $R^2 \in (-\infty, 1.0]$ where 1.0 is the best score.
- **Visualization:** The model's prediction is visualized as a 2D scatter plot. The best plot is y = x line which corresponds to the best R^2 score of 1.0.







Results

Results:

- 2,3,4
- 0



Discussion

Notable points:

- Best models: Linear Regression and Random Forest Regression.
- RF uses correlated features to make itself more robust.
- RF can deal with both discrete and real valued features.

Limitations:

- Linear regression assumes data linearity.
- RF has heavy reliance on ligand features.
- Both models were black box models.
- Testing results were sometimes better than validation results.
 This is because test data < validation data. But the

difference is minimal.

Discussion

Further work:

 A new weighting strategy: Weighting a pocket descriptor based on the overlap between the pocket and the ligand. For example,

$$W_{\mathrm{Total}} = W_{\mathrm{Hyperbolic}} * W_{\mathrm{Overlap}}$$

- Improvement of feature selection: Build 1 model per family of features. Use the best feature as a family surrogate.
- A more explainable model can be built.



Q & A



References

- Du, Li, Xia, Ai, Liang, Sang, Ji and Liu; Insights into Protein–Ligand Interactions: Mechanisms, Models, and Methods (2016)
- Le Guilloux, Schmidtke, and Tuffery; Fpocket: An open source platform for ligand pocket detection(2009)
- DiMasi, Grabowski and Hansen; nnovation in the pharmaceutical industry: New estimates of R & D costs (2016)
- John H. Holland. Genetic Algorithms. (1960)
- Is rotation forest the best classifier for problems with continuous features? A. Bagnall, M. Flynn, J. Large, J. Line, A. Bostrom, and G. Cawley (2020)

Appendix

• Binding affinity between a protein and a ligand is quantified by the K_d , K_i and IC_{50} . Here K_d refers to the dissociation constant, K_i to inhibition constant, and IC_{50} to inhibitory concentration 50%.

