



# Network analysis of DNA Repair phenotype using database of nano-biomimetic based single cell assay

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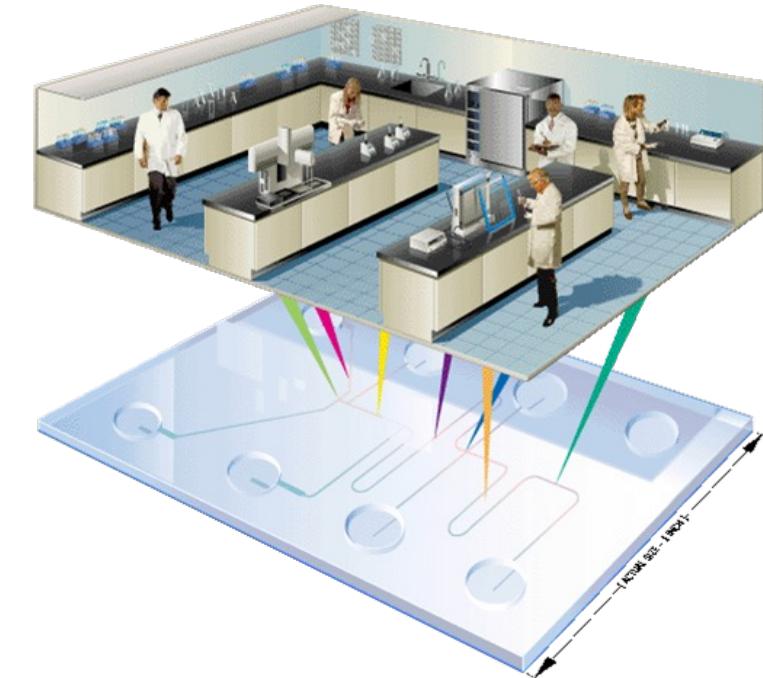
# BIOLOGY IN SINGLE-CELL RESOLUTION

**How and why  
technology enable  
high-throughput  
bio-assays in single  
cell resolution?**

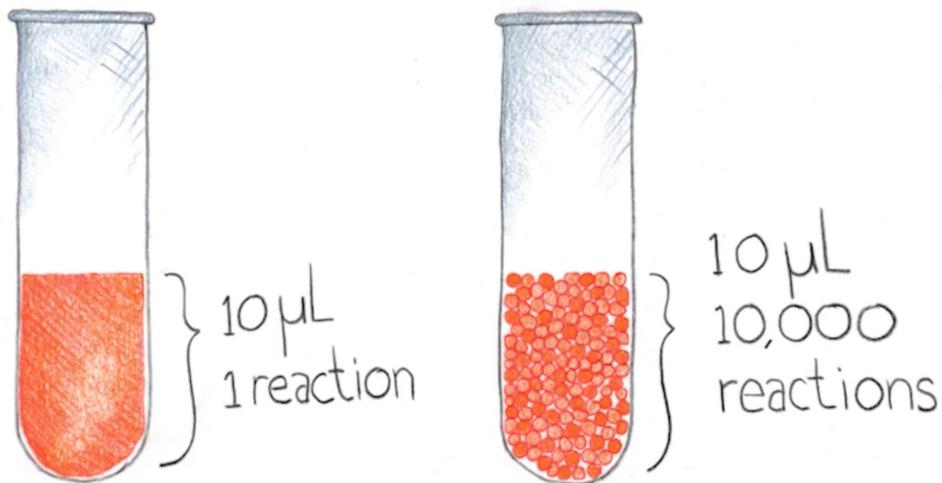
# $\mu$ -TAS concept: miniaturized Total Analysis System

- If the device in question had characteristic dimensions on the microscale.
- A system that could automatically carry out all the functions required for analysis.

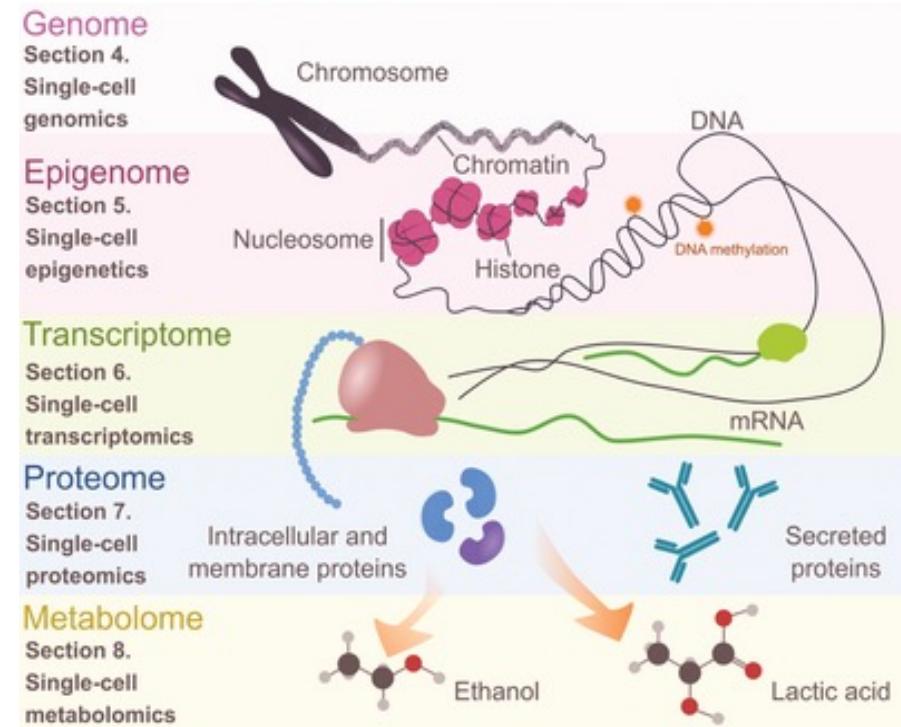
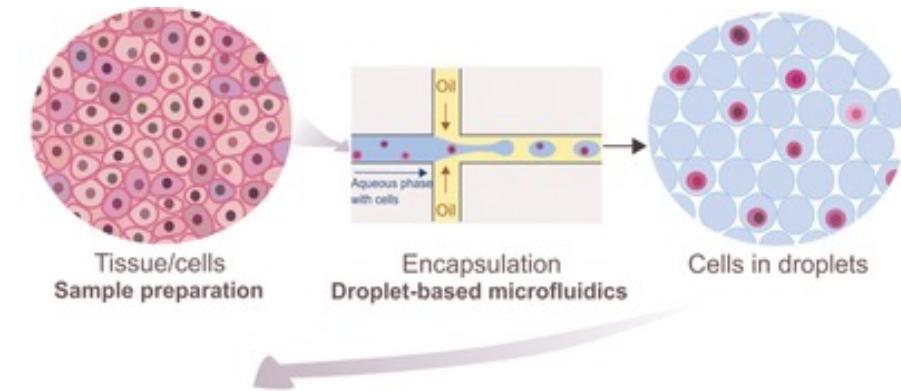
- Sampling
- Transport of the sample
- Any sample preparation steps
  - Ex. chemical reactions, separations, etc.
- Detection



# Single-Cell Analysis Using Droplet Microfluidics

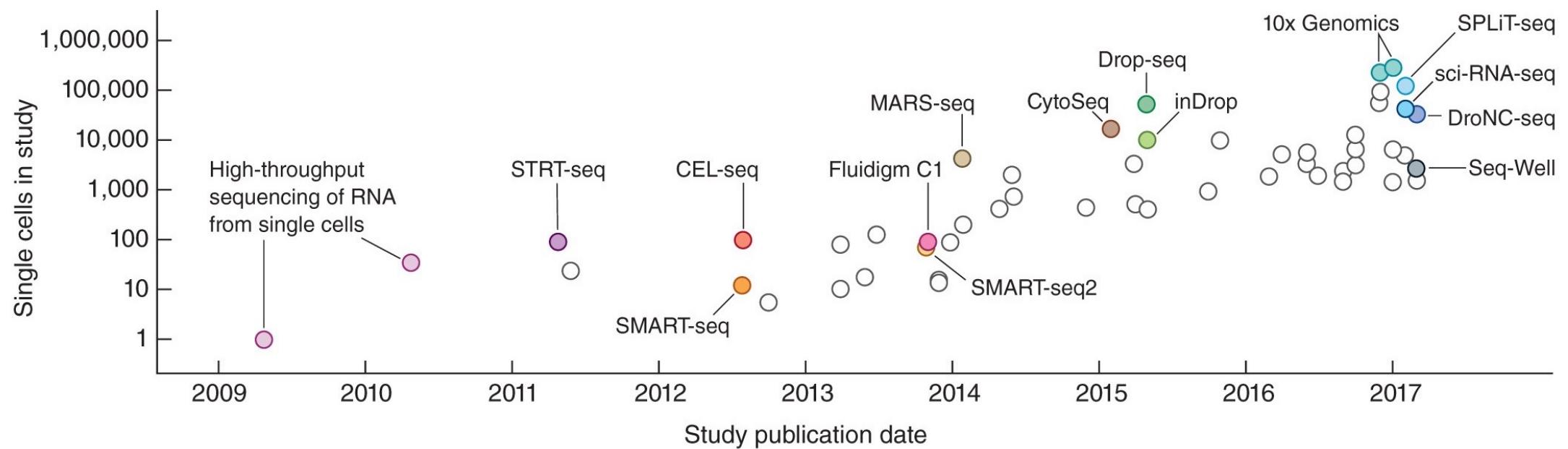


[Macosko, E. et al. \(2015\)](#)



[Matuła, K., et.al \(2020\) | Review](#)

# Scaling of scRNA-seq experiments



# SINGLE CELL DNA-REPAIR MEASUREMENT

**NGS & droplet  
microfluidic  
platforms  
enable high  
throughput  
measurement  
of biochemical  
phenotypes in  
single cells.**

# Nucleic Acids Research

Published online 14 April 2020

*Nucleic Acids Research*, 2020, Vol. 48, No. 10 e59  
doi: 10.1093/nar/gkaa240

## Simultaneous measurement of biochemical phenotypes and gene expression in single cells

Amanda L. Richer<sup>1,2</sup>, Kent A. Riemonyd<sup>③</sup>, Lakotah Hardie<sup>1</sup> and Jay R. Hesselberth<sup>①,2,3,\*</sup>

<sup>1</sup>Department of Biochemistry and Molecular Genetics, Aurora, CO 80045, USA, <sup>2</sup>Molecular Biology Program and  
<sup>3</sup>RNA Bioscience Initiative, University of Colorado School of Medicine, Aurora, CO 80045, USA

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## Method

Experimental molecular assay

## Dataset

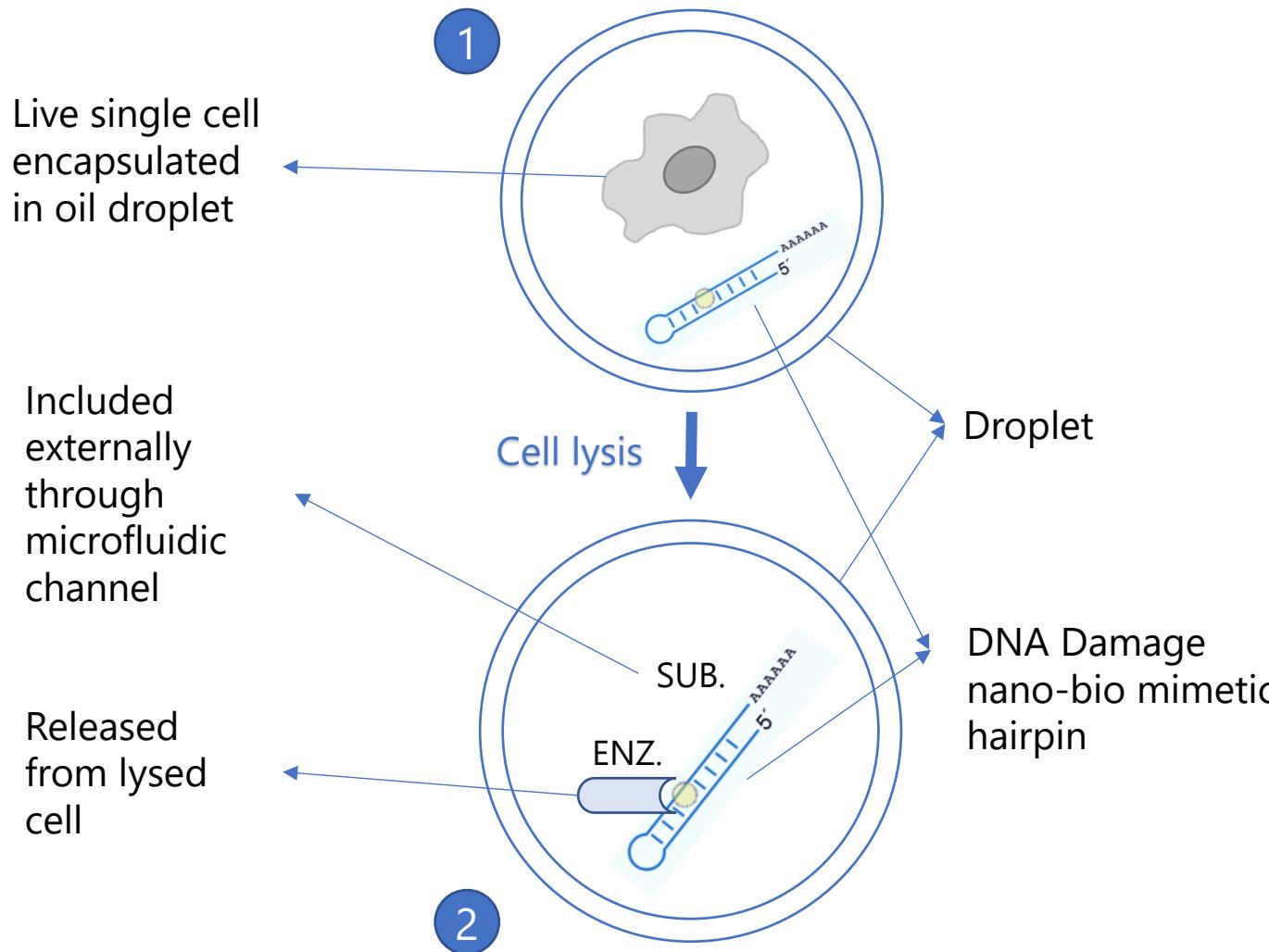


Several single-cell RNA-seq experiments

## Proof of concept

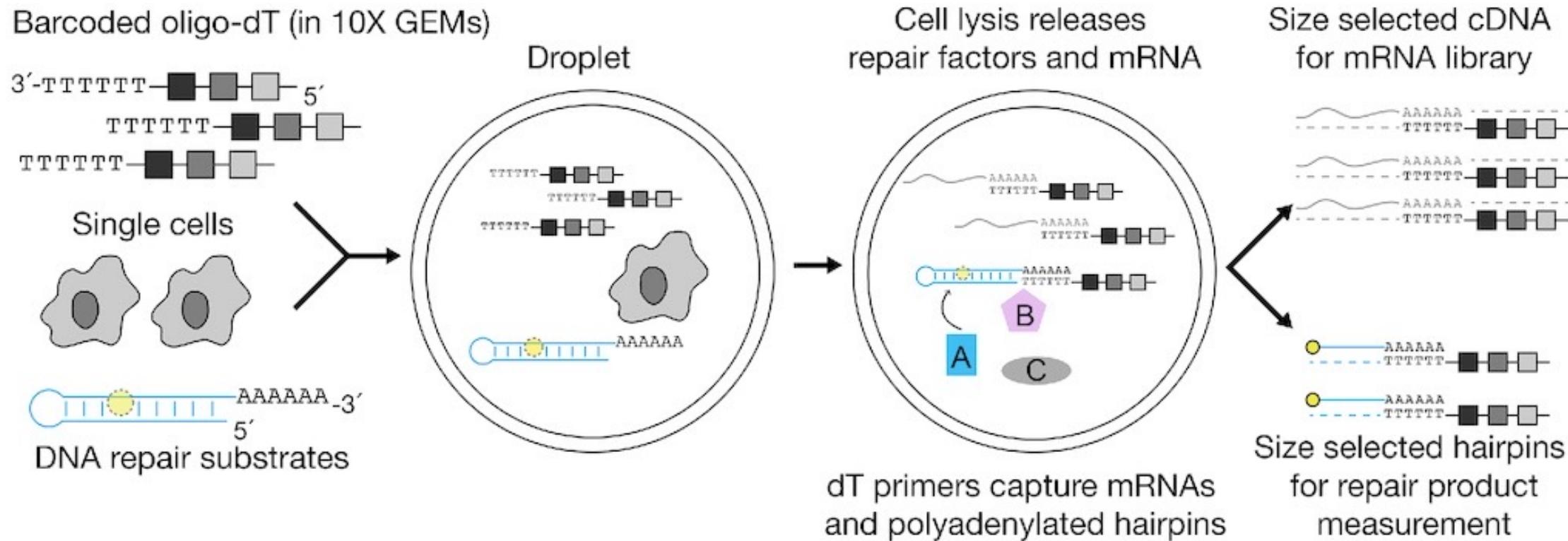
Nucleic-based ***nano-biomimetic*** probes are powerful paradigm to design novel molecular assays

# Mimic DNA repair enzyme-substrate reaction inside droplet



- An external synthetic hairpin with a single lesion damage at certain position included into droplets using the same channel which cell loaded to the microfluidic chip
- This hairpin *mimic* substrate of DNA repair enzymes which released from the cell during in-droplet lysis
- Overall, this protocol made it possible to simply measure amount of enzymatic activity (i.e., number of strand incisions) alongside with mRNA abundance in single cell resolution

# Measuring DNA-repair enzyme activity in single-cell resolution



# Mixing and time series experiment

- KO cells were identified if counts at the repair site (position 44 for ribonucleotide and position 45 for uracil)
- After the emulsion was created, the sample was separated into 3 tubes and incubated for 15, 30, or 60 min at 37 °C prior to reverse transcription at 53 °C.
- 800-1,500 cells were captured at each timepoint.
- DNA repair measurements determine *cell types* in a cell mixing experiment.
- Authors showed it fails to use UNG and RNASEH2C mRNA expression to determine cell types, but estimated repair activity clearly assign cell-types.

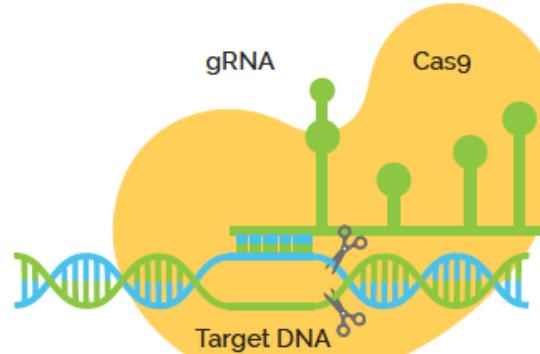
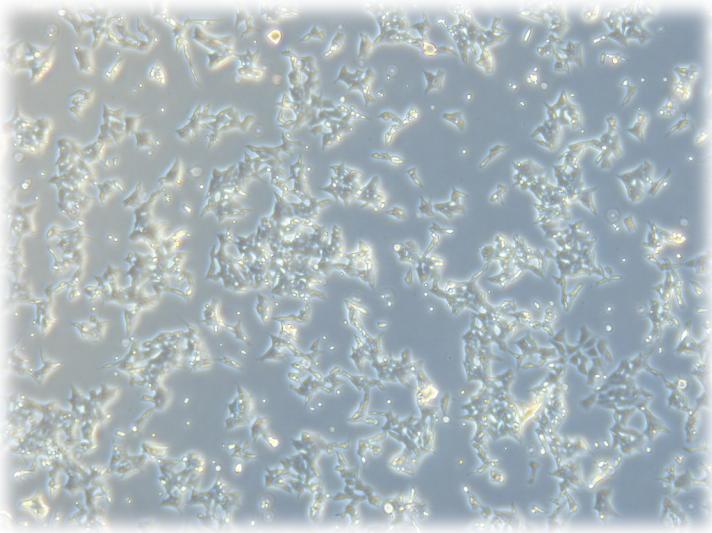
What else we can interpret from this experiment?

Differential expression analysis

Pathway enrichment analysis

Gene regulatory network analysis

# Assess HAP1 cell line with Knock-out genes



Disease	Lineage
Engineered	Engineered Blood
Disease Subtype	Lineage Subtype
Chronic Myelogenous Leukemia (CML)	CML
Source	Gender
Horizon	TM Male
Discovery	

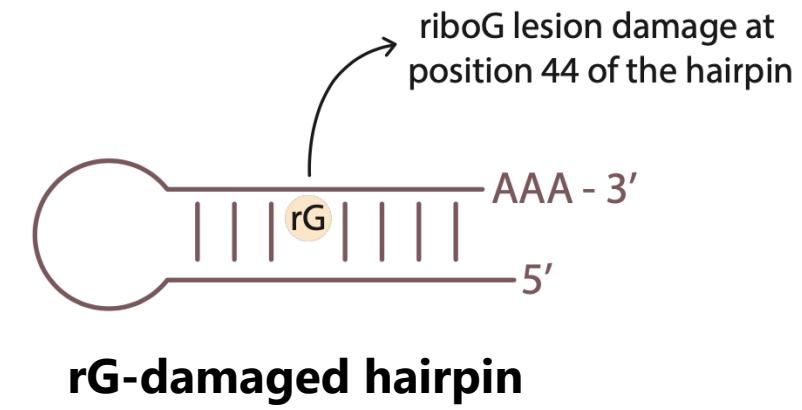
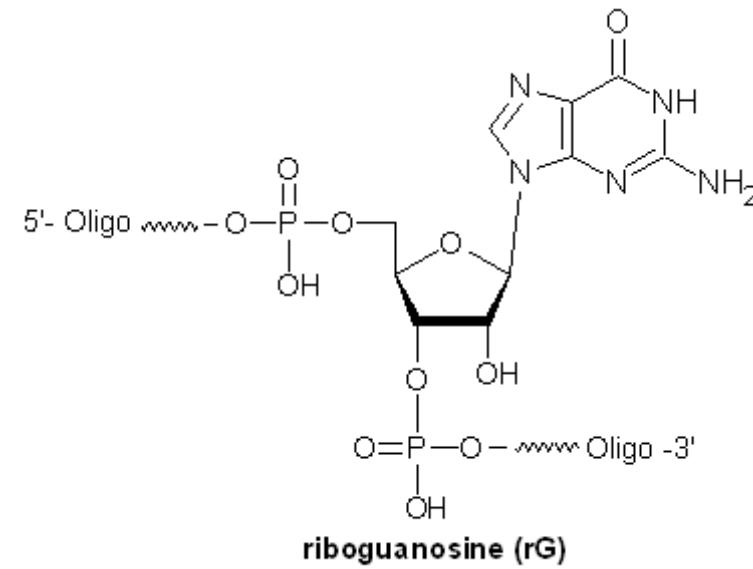
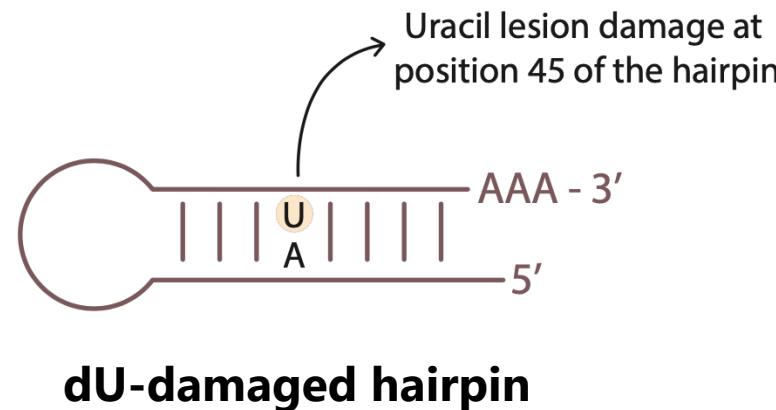
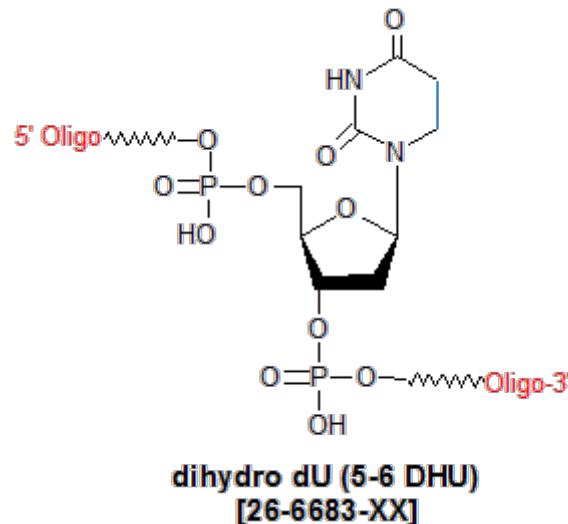
## **RNASEH2C** KO

Ribonuclease H2 Subunit C  
• 164 amino acids

## **UNG** KO

Uracil DNA Glycosylase  
• 313 amino acids

## Synthetic Hairpins as the mimic of DNA repair enzyme substrate



## RNASEH2C

Ribonuclease H2 Subunit C

### INTRODUCTION

### Single cell DNA-repair measurement

#### RNAseH2C - Ribonuclease H2 Subunit C

- Non catalytic subunit of RNase H2, an endonuclease that specifically **degrades the RNA of RNA:DNA hybrids** and mediates the excision of single ribonucleotides from DNA:RNA duplexes.
- Participates in DNA replication, possibly by mediating the removal of lagging-strand Okazaki fragment RNA primers.
- Ribonucleotides are incorporated into DNA by the replicative DNA polymerases at frequencies of about 2 per kb which makes them by far the **most abundant form of potential DNA damage in the cell**.
- Their removal is essential for restoring a stable intact chromosome.

## UNG

Uracil DNA Glycosylase

### INTRODUCTION

### Single cell DNA-repair measurement

#### UNG - Uracil DNA Glycosylase

- Belongs to the uracil-DNA glycosylase (UDG) superfamily
- Excises uracil residues from the DNA which can arise as a result of misincorporation of **dUMP** residues by DNA polymerase or due to deamination of cytosine;
- UNG is the major **uracil-DNA glycosylase** in mammalian cells and is involved in both
  - Error-free base excision repair of genomic uracil
  - Mutagenic uracil-processing at the antibody genes.
- The regulation of UNG in these different processes is currently *not well understood*. 

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# UNG - Uracil DNA Glycosylase

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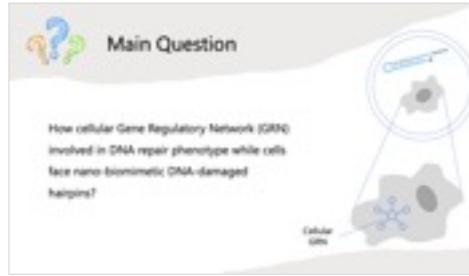
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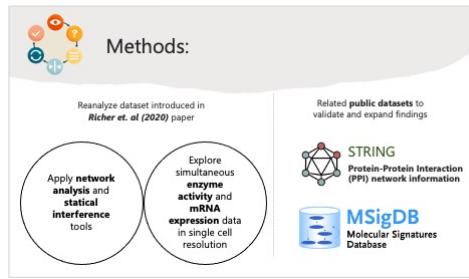
# Thesis Overview



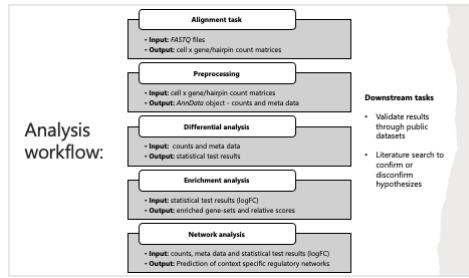
Main Question:



Methods:



Workflows:



Results:

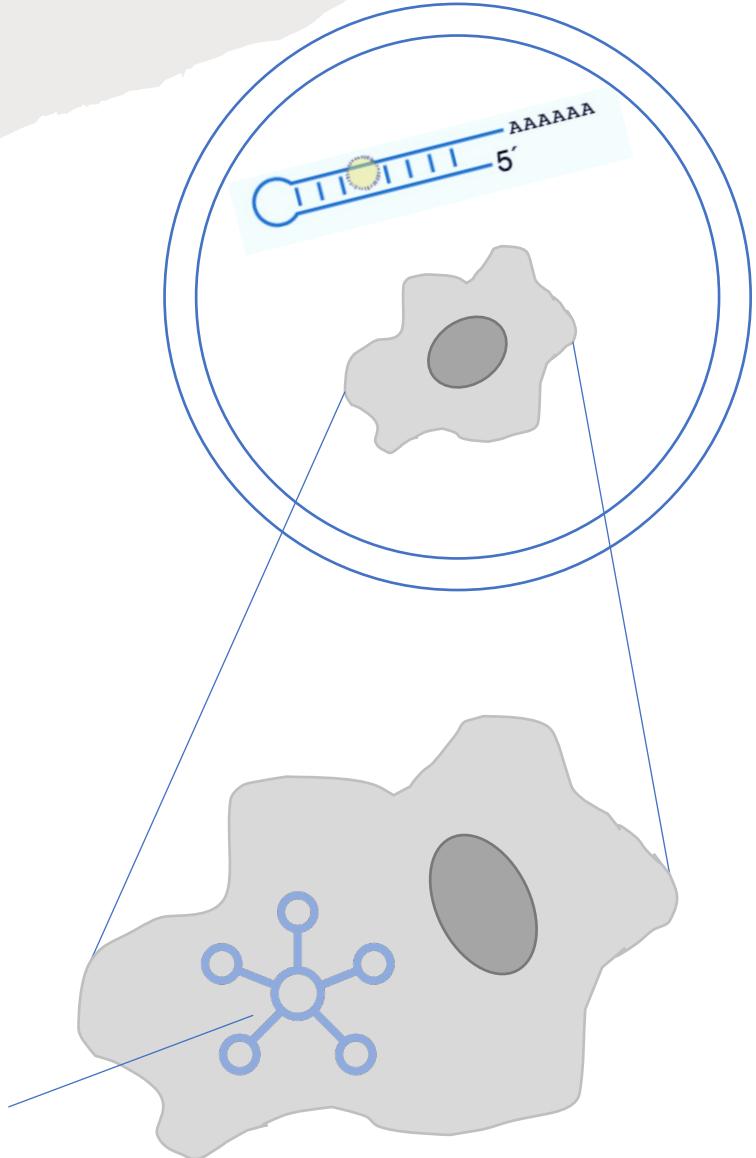


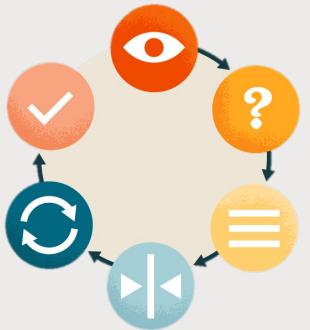


# Main Question

How cellular Gene Regulatory Network (GRN)  
involved in DNA repair phenotype while cells  
face nano-biomimetic DNA-damaged  
hairpins?

Cellular  
GRN





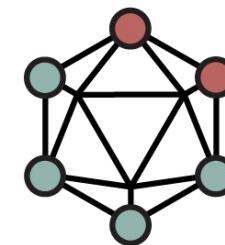
# Methods:

Reanalyze dataset introduced in  
***Richer et. al (2020)*** paper

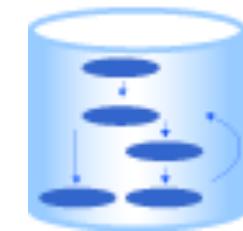
Apply **network analysis** and  
**statical interference**  
tools

Explore simultaneous  
**enzyme activity** and  
**mRNA expression** data  
in single cell  
resolution

Related **public datasets** to validate and expand findings

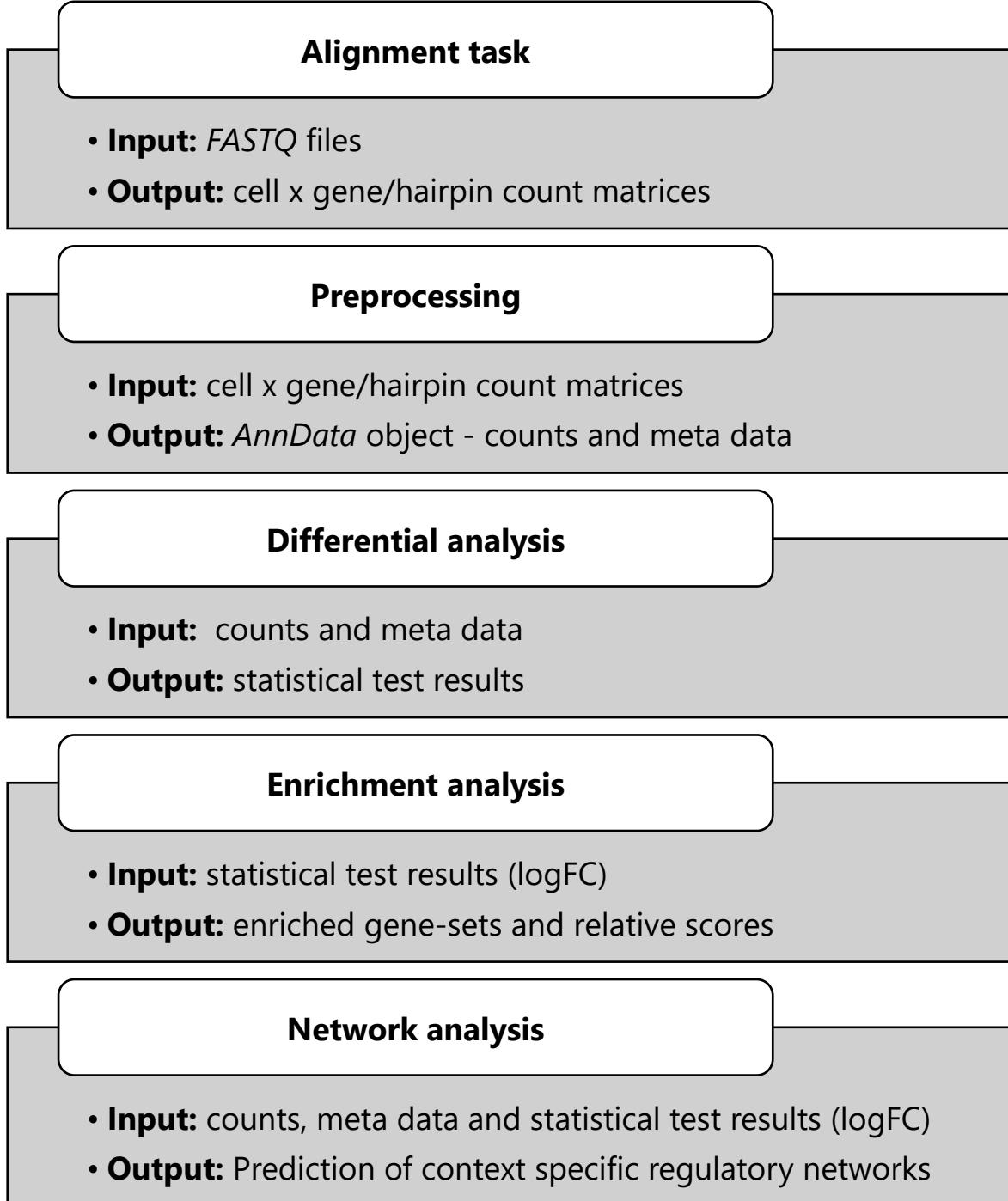


**STRING**  
Protein-Protein Interaction  
(PPI) network information



**MSigDB**  
Molecular Signatures Database

# Analysis workflow:



## Downstream tasks

- Validate results through public datasets
- Literature search to confirm or disconfirm hypothesizes

# Results:

## Basic Analysis

NETWORK ANALYSIS OF DNA REPAIR PHENOTYPE |

- Alignment task
- Preprocessing
- Differential analysis
- Enrichment analysis
- Network analysis

22

## Comparison Analysis

NETWORK ANALYSIS OF DNA REPAIR PHENOTYPE |

- Alignment task
- Preprocessing
- Differential analysis
- Enrichment analysis
- Network analysis

28

## Network and Graph Analysis

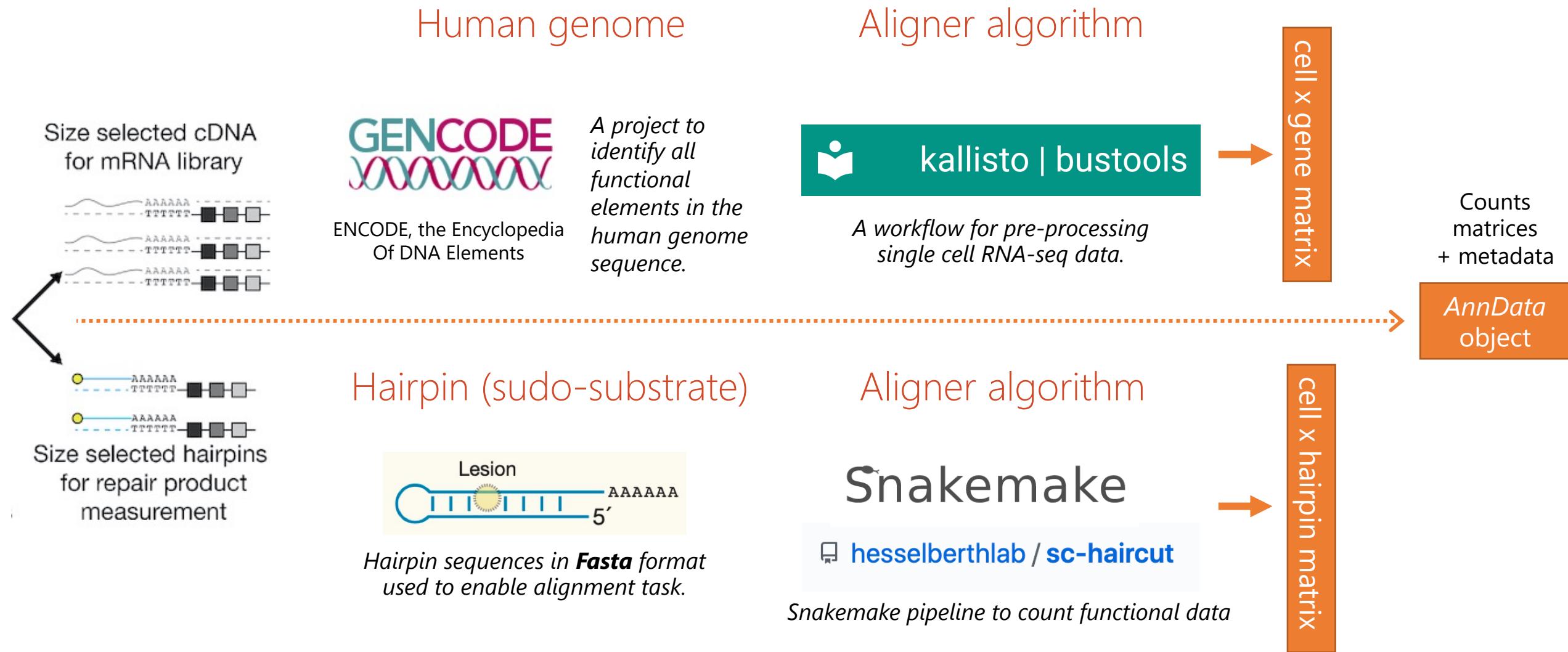
NETWORK ANALYSIS OF DNA REPAIR PHENOTYPE |

- Alignment task
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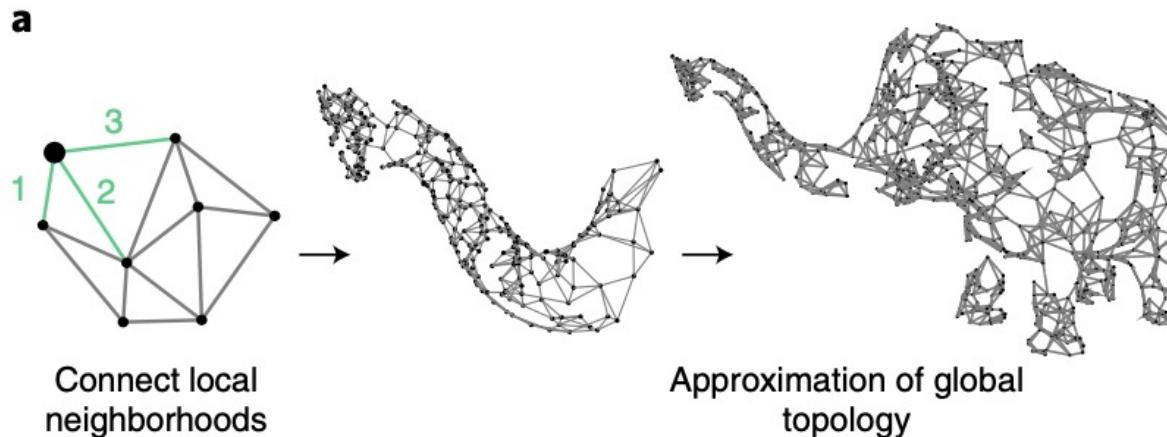
36

# Basic Analysis

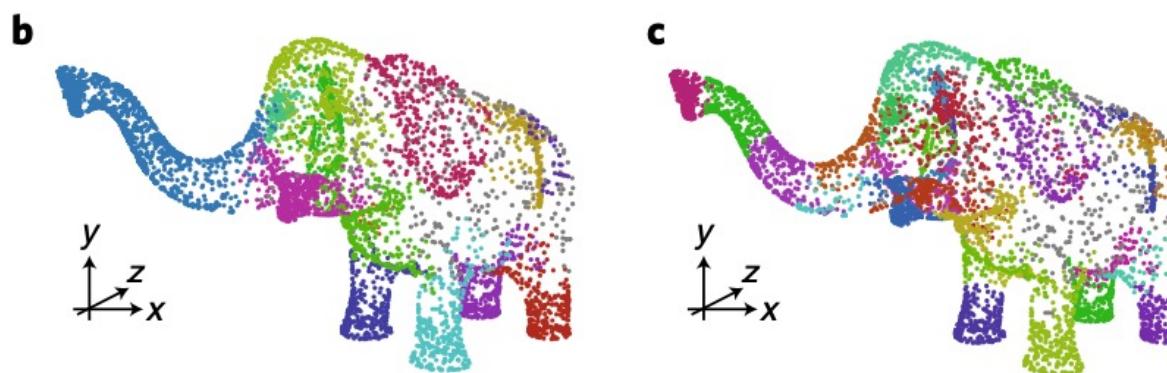
- Alignment task
- Preprocessing
- Differential analysis
- Enrichment analysis
- Network analysis



# Approximating and partitioning complex manifolds



**a**, Complex, curved surfaces can be well approximated by neighborhood graphs. A simple graph connects each point with its  $k$  closest neighbors (kNN graph). As more points and regions are measured, the complex structure of the object can be revealed.



**b**, The elephant graph (in a) is clustered using the **Leiden clustering algorithm** (resolution  $r = 0.5$ ). The resulting clusters are shown as colors on the 3D model (top) and t-SNE embedding (bottom) of the data.

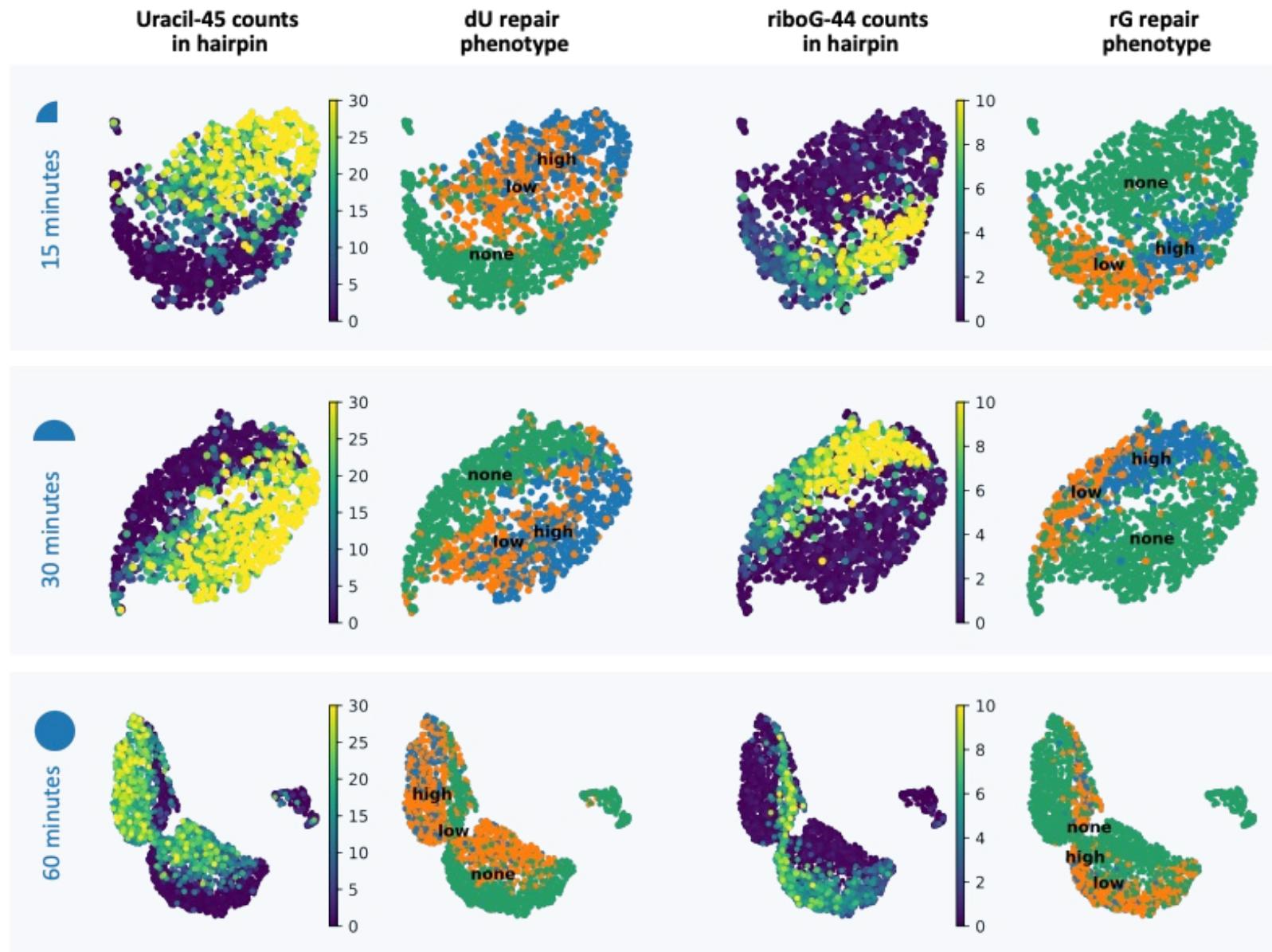
**c**, Clustering resolution is arbitrary. Similar to b, the plots show clustering with increased resolution ( $r = 3$ ). The clusters are smaller but capture equally valid anatomical elements.

## UMAP plots

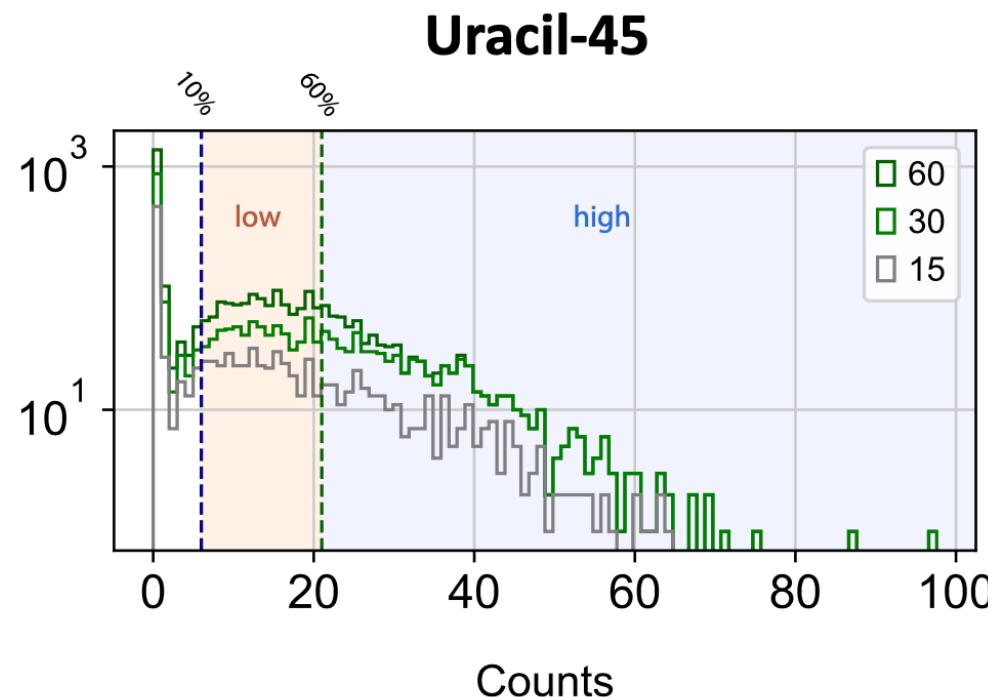
counts of hairpins  
with lesion damage  
(left)

label group of cells  
for repair  
phenotype  
(right)

## Preprocessing

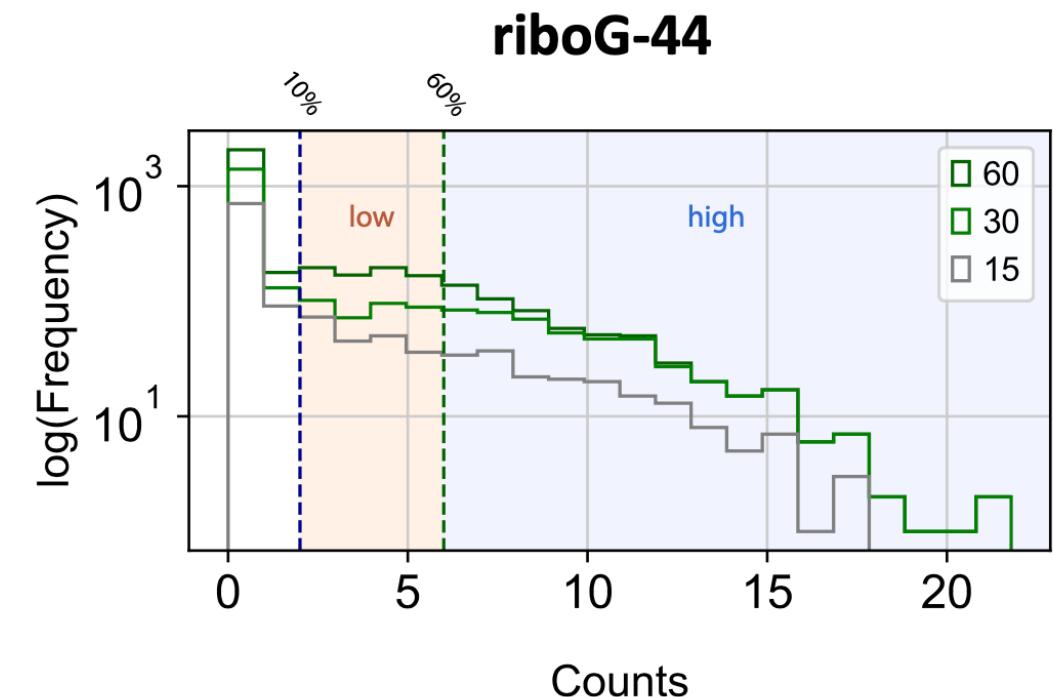


# Define binary label for repair phenotypes



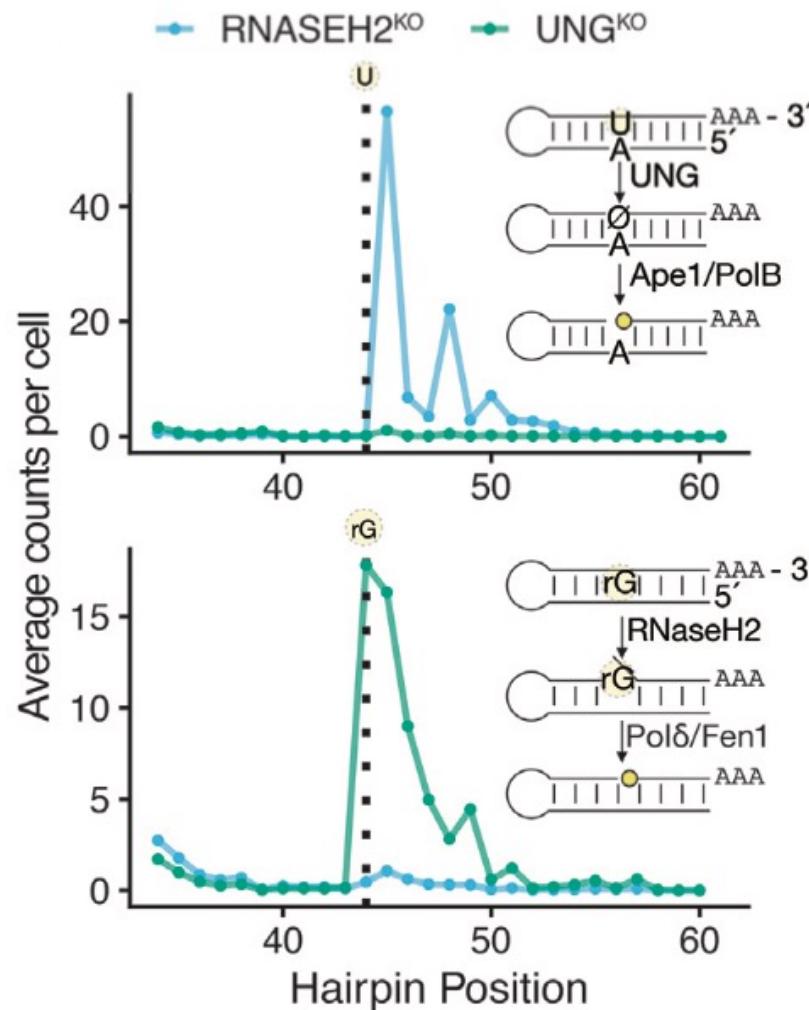
Total cells: 4865

none 3134  
low 1072  
high 659



none 2267  
low 1617  
high 981

# What are labels representing?



## Uracil-44 count:

high	High <b>dU</b> count, high <b>dU</b> repair phenotype
low	Low <b>dU</b> count, low <b>dU</b> repair phenotype
none	<b>UNG<sup>KO</sup></b> cells

} **RNASEH2C<sup>KO</sup>** cells  
fail to incise  
ribonucleotide  
damage

## riboG-45 count:

high	High <b>rG</b> count, high <b>rG</b> repair phenotype
low	Low <b>rG</b> count, low <b>rG</b> repair phenotype
none	<b>RNASEH2C<sup>KO</sup></b> cells

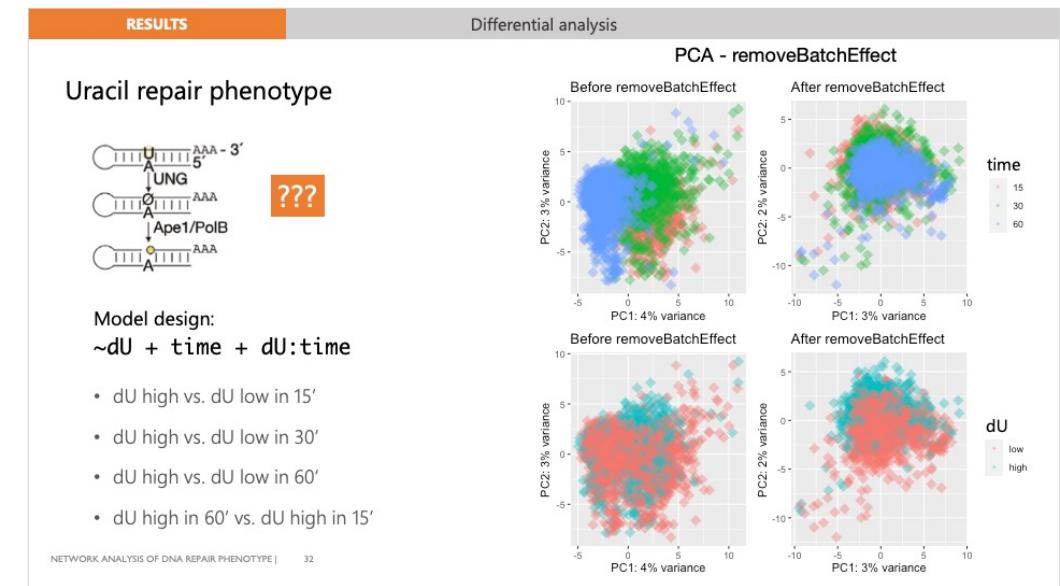
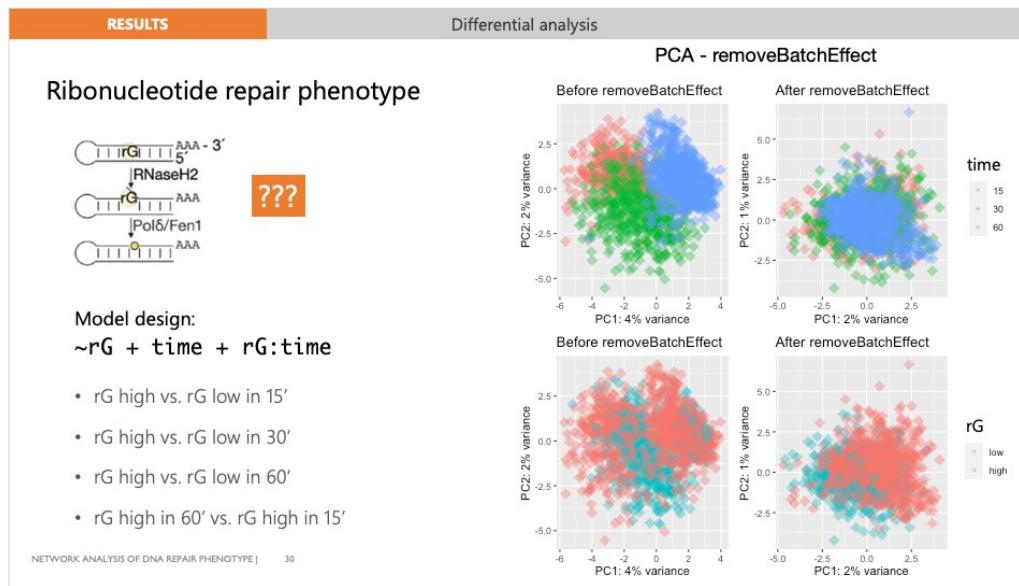
} **UNG<sup>KO</sup>** cells  
fail to incise  
uracil damage

# Comparison Analysis

- Alignment task
- Preprocessing
- **Differential analysis**
- **Enrichment analysis**
- Network analysis

# The model formula and design matrices

- We aim to test and report multiple comparisons in our dataset:



Variables:

- dU (High / Low / None)
- rG (High / Low / None)
- time (15, 30, 60)

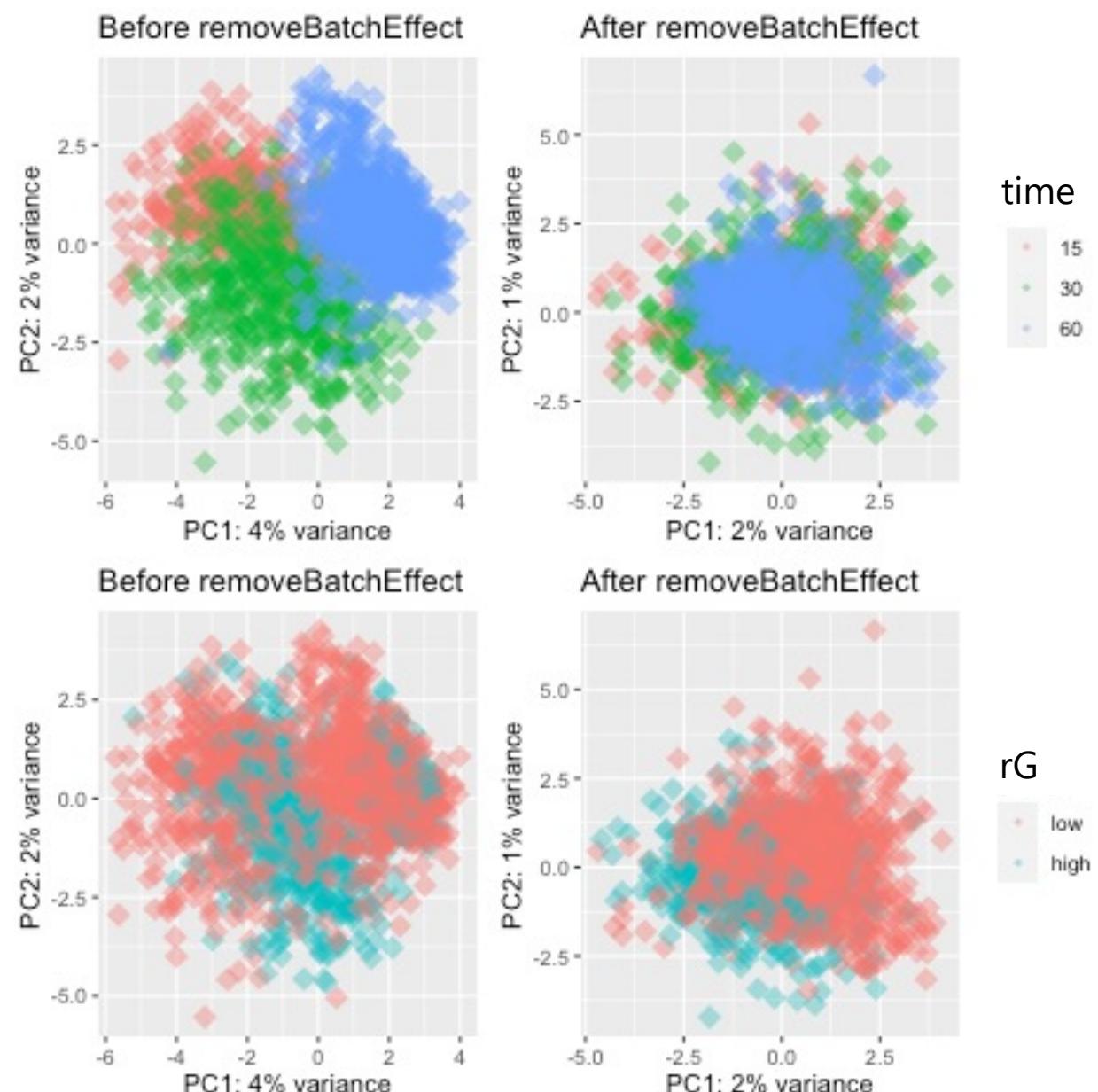
## PCA - removeBatchEffect

## Ribonucleotide repair phenotype

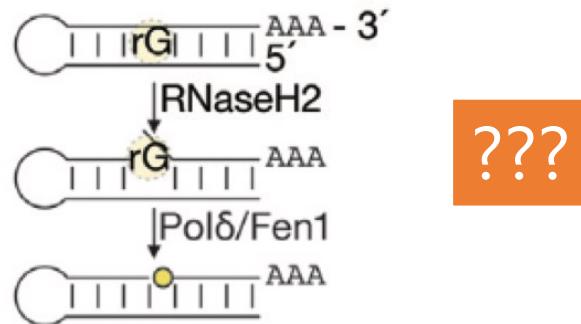


Model design:  
 $\sim \text{rG} + \text{time} + \text{rG:time}$

- rG high vs. rG low in 15'
- rG high vs. rG low in 30'
- rG high vs. rG low in 60'
- rG high in 60' vs. rG high in 15'

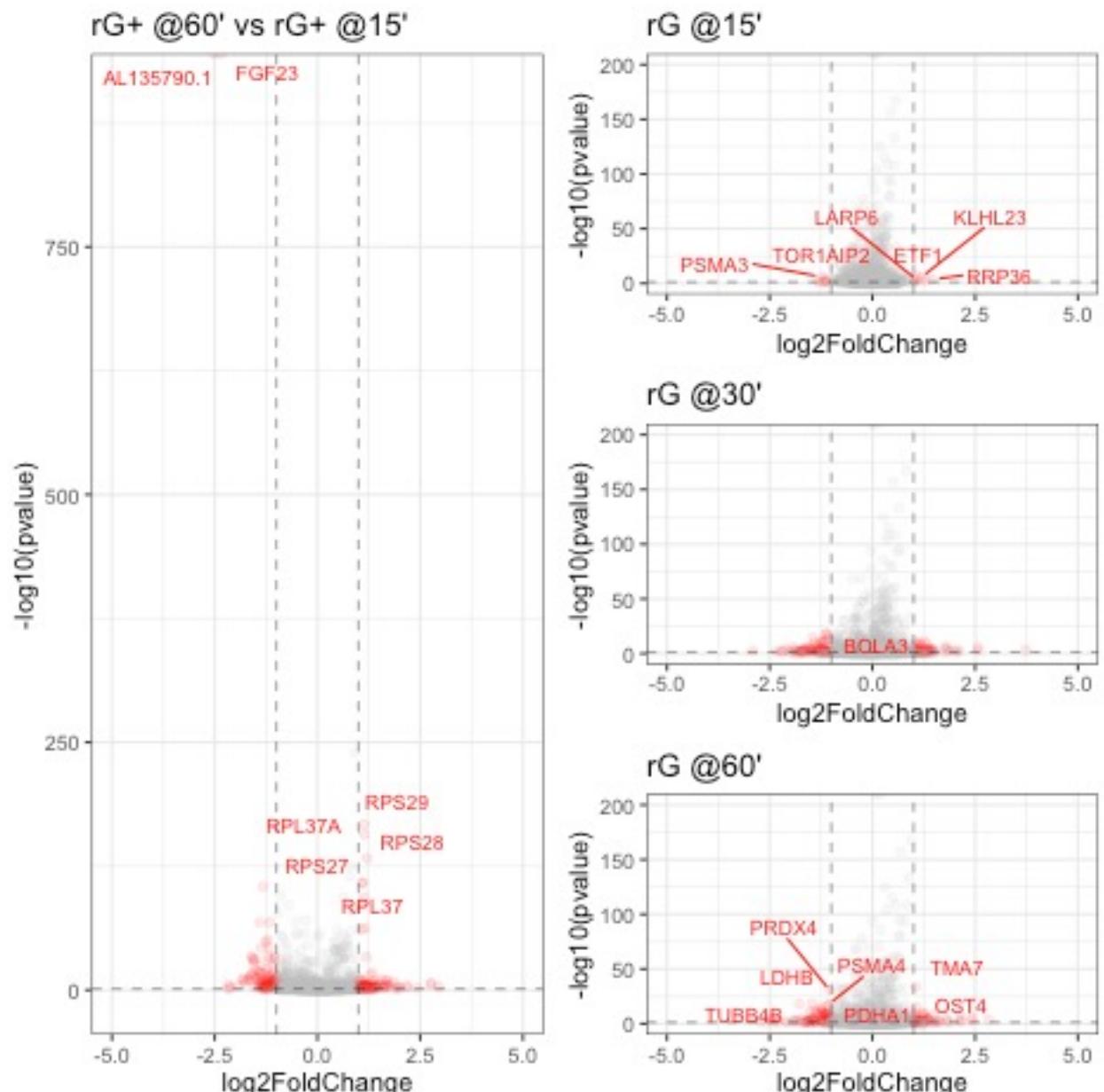


## Ribonucleotide repair phenotype

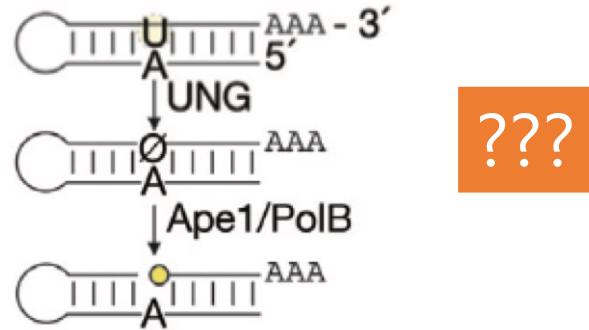


Model design:  
 $\sim \text{rG} + \text{time} + \text{rG:time}$

- rG high vs. rG low in 15'
- rG high vs. rG low in 30'
- rG high vs. rG low in 60'
- rG high in 60' vs. rG high in 15'

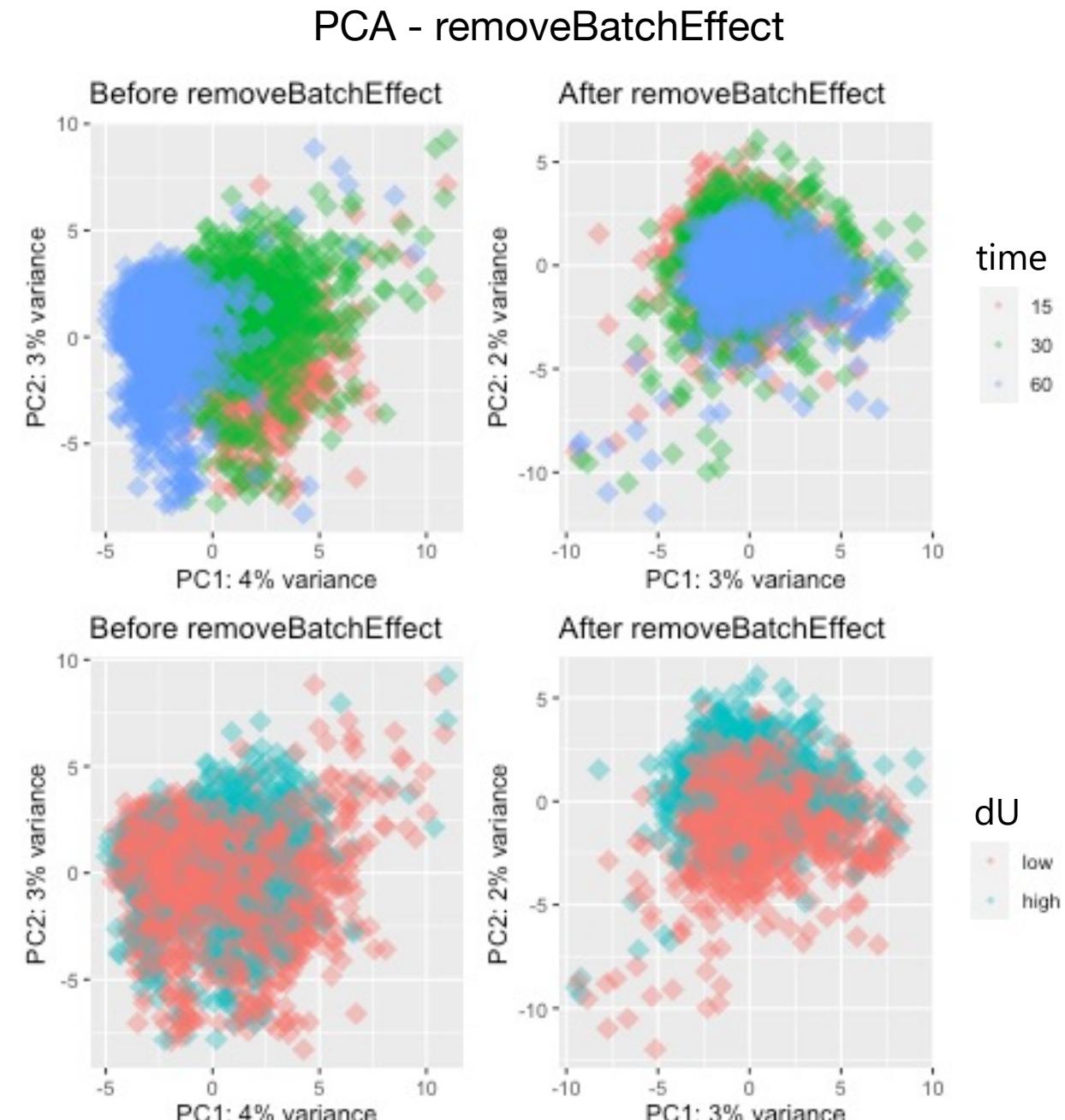


# Uracil repair phenotype

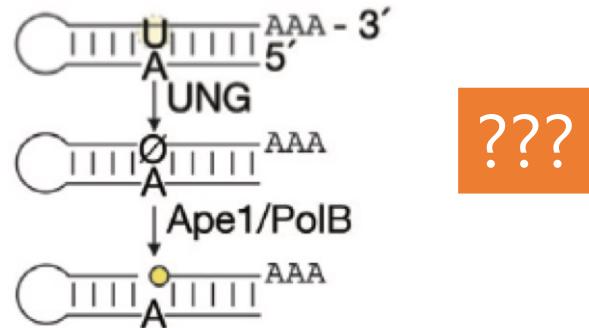


Model design:  
 $\sim dU + \text{time} + dU:\text{time}$

- dU high vs. dU low in 15'
- dU high vs. dU low in 30'
- dU high vs. dU low in 60'
- dU high in 60' vs. dU high in 15'

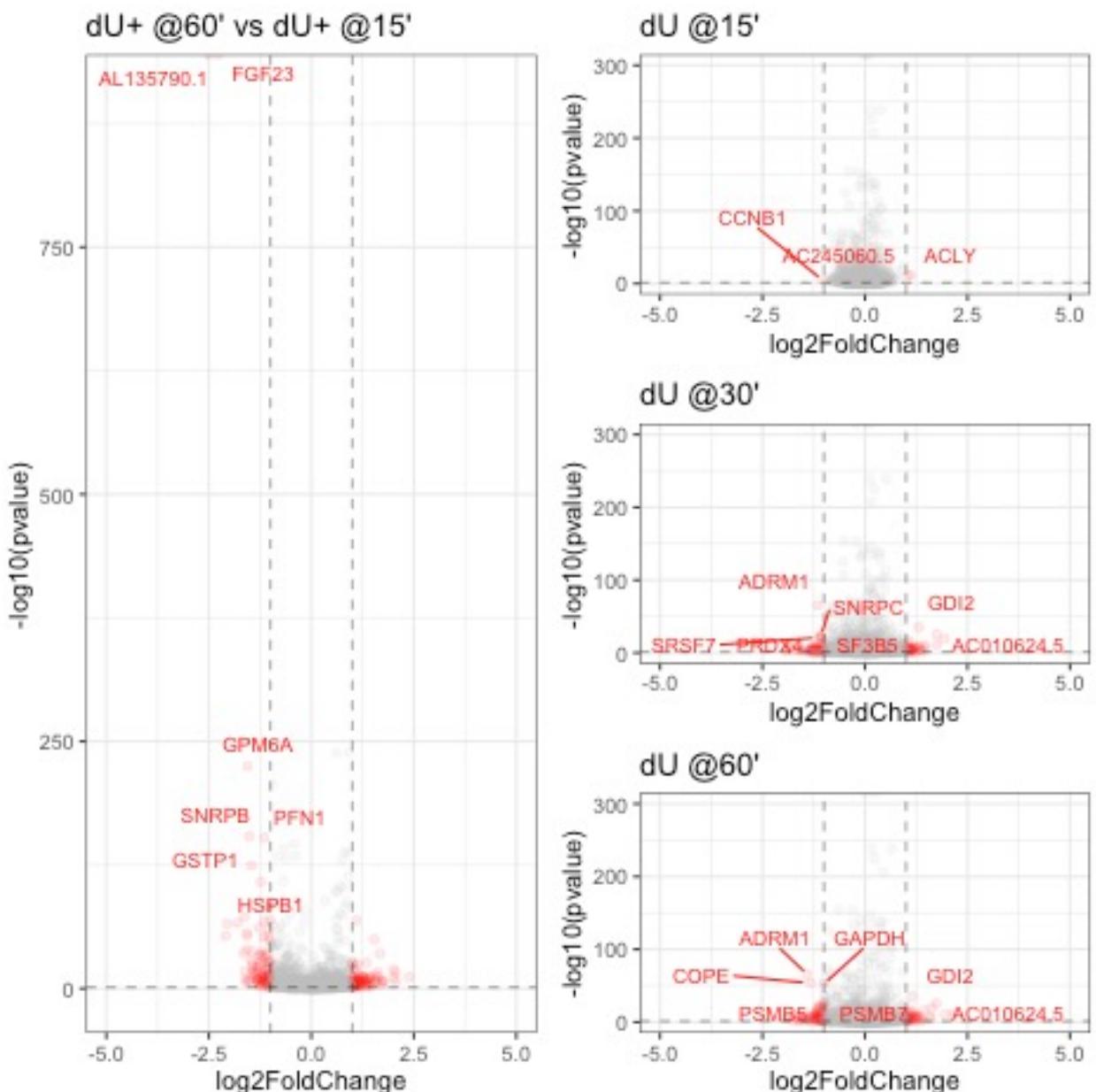


# Uracil repair phenotype



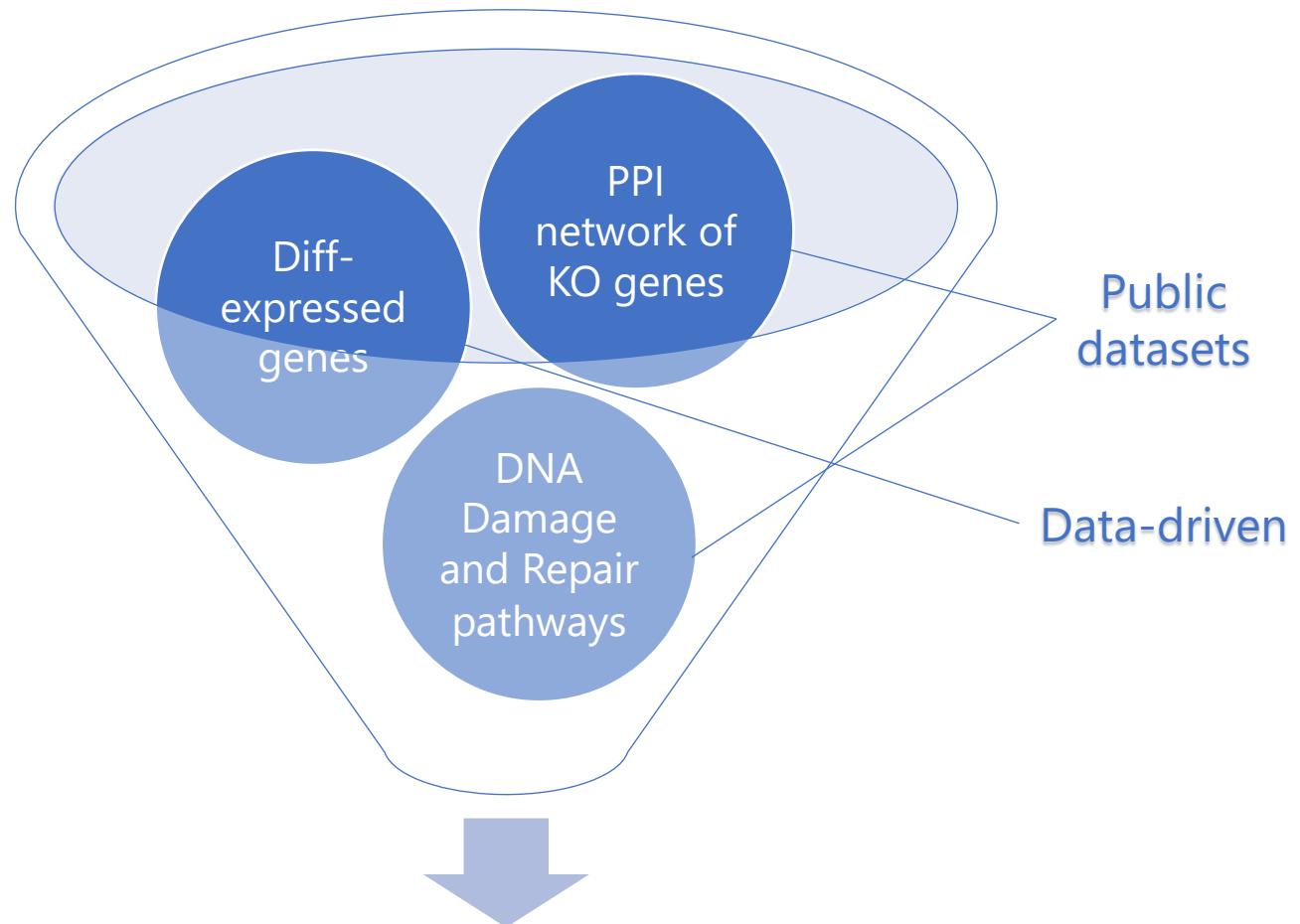
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- dU high vs. dU low in 60'
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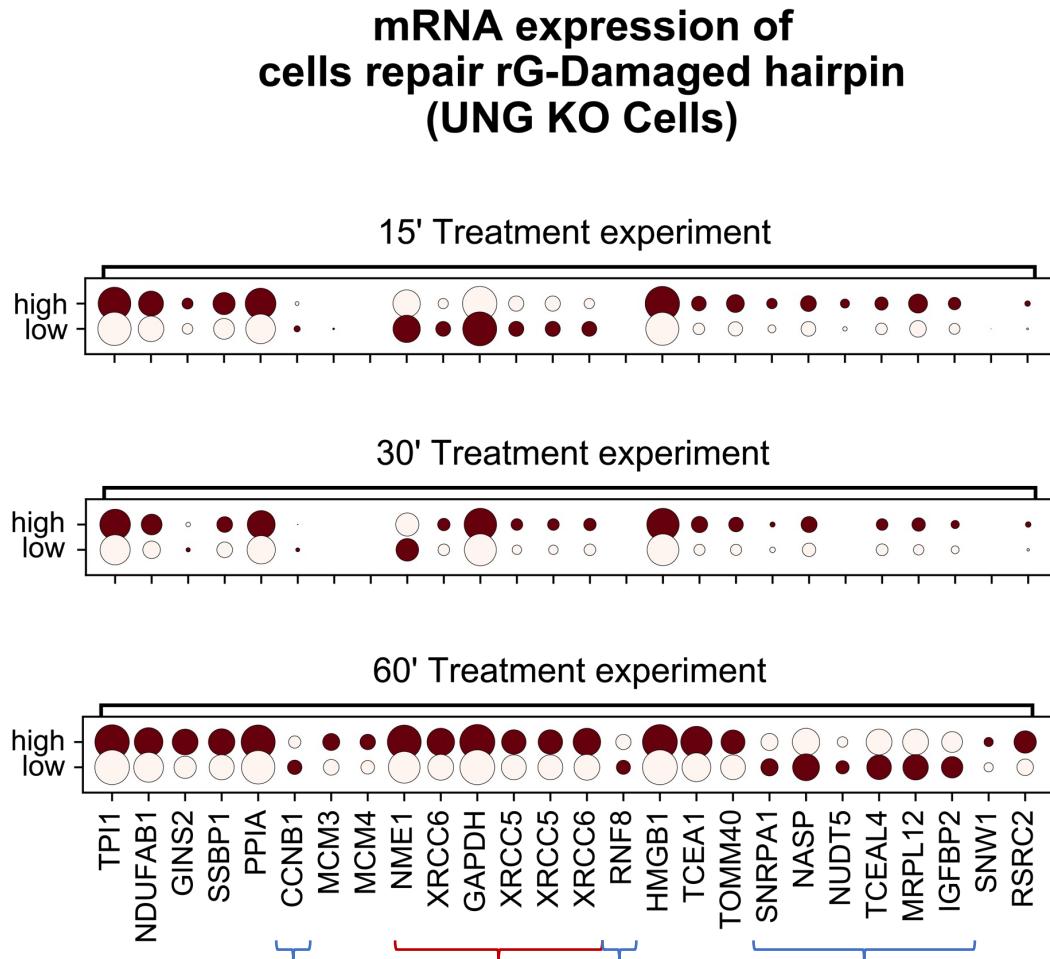
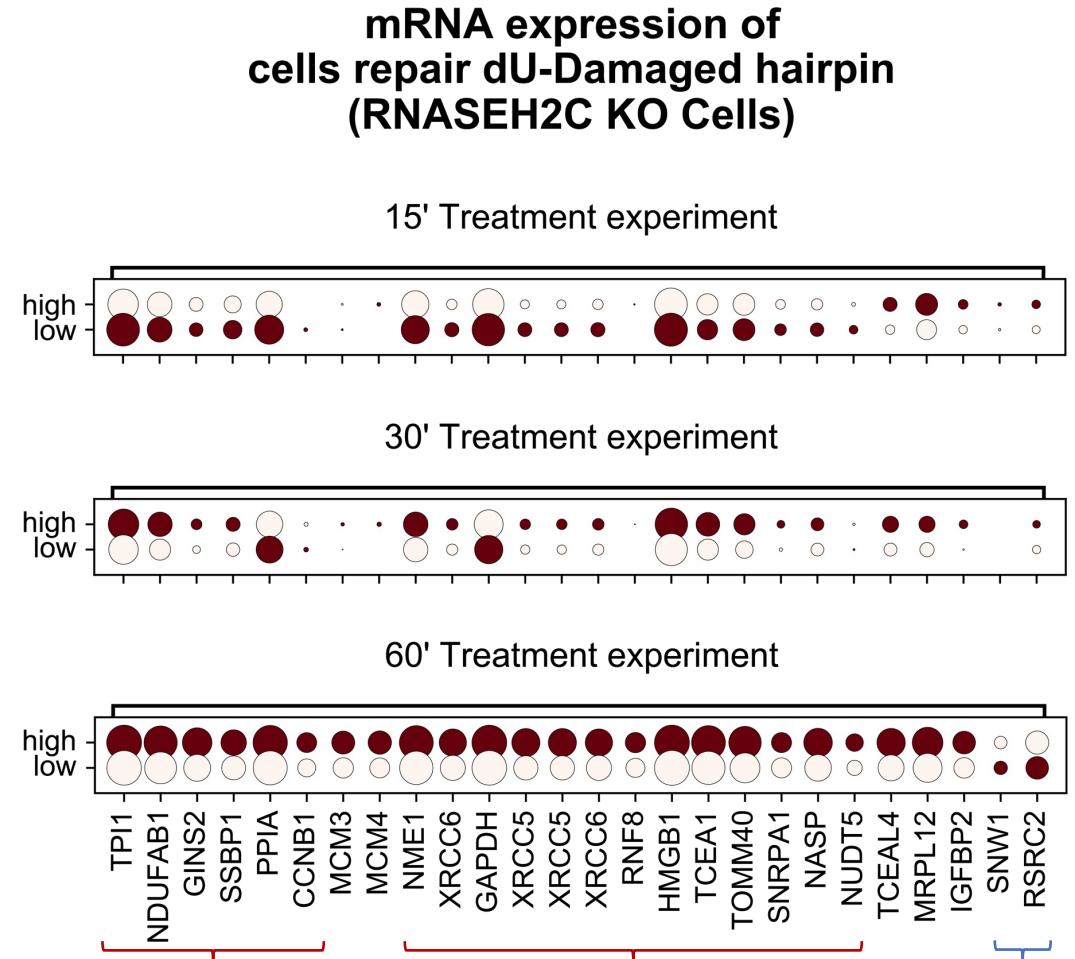


Find genes with expression alteration over time

Long list of investigated genes



Manually select altered genes

**rG Repair Phenotype****dU Repair Phenotype**

# Network and Graph Analysis

- Alignment task
- Preprocessing
- Differential analysis
- Enrichment analysis
- **Network analysis**

# GRN - Gene Regulatory Networks

## pySCENIC

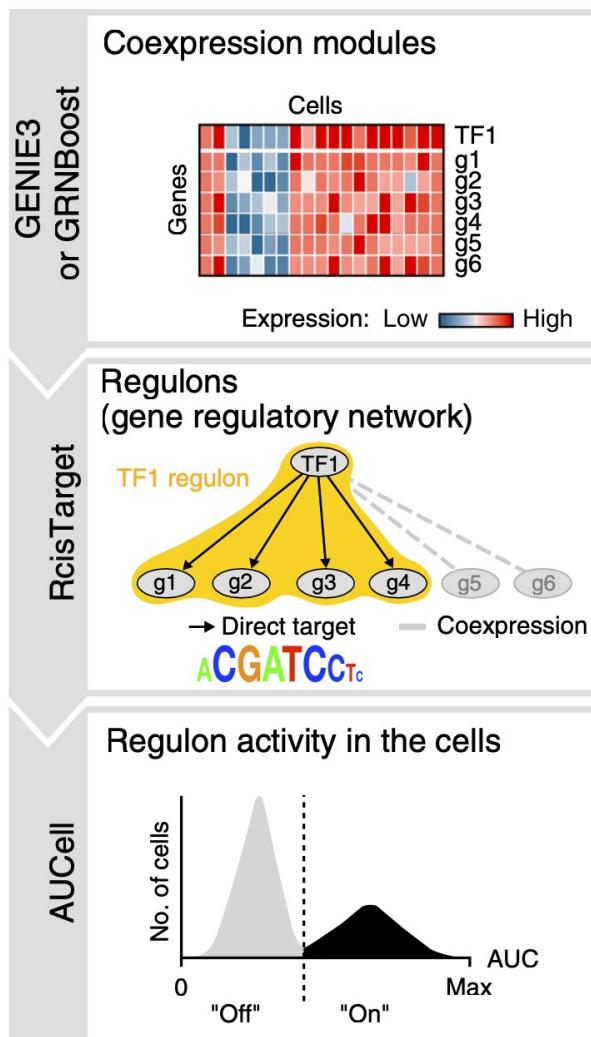
A lightning-fast python implementation of the SCENIC pipeline (**S**ingle-Cell **r**egulatory Network Inference and **C**lustering)

Enables  
biologists to infer  
from scRNA-seq  
data

Transcription factors (TFs)  
Gene Regulatory Networks (GRNs)  
Cell types

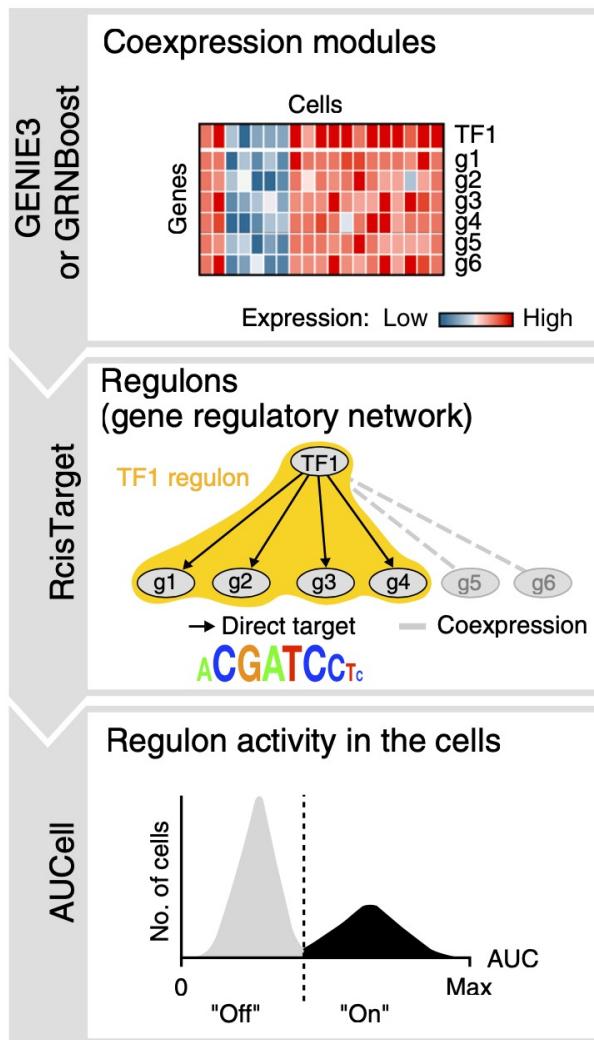


# pySCENIC workflow

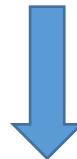


1. Sets of genes that are coexpressed with TFs are identified using **GENIE3**
2. Since the **GENIE3** modules are only based on coexpression, they may include many **false positives** and indirect targets.  $\Rightarrow$  To identify putative direct-binding targets, each coexpression module is subjected to *cis*-regulatory motif analysis using **RcisTarget**.
3. Estimate AUC score as regulons activity representation among cells.

# GRN analysis results



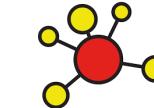
(24,484,108) co-expression profile found



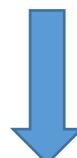
(4,652,523) TF/target interaction found

**Columns:** TF      Target      importance

*igraph*



I aim to create weighted and directed graph toward further analysis (finding context specific master regulators)



(4,865) Cells in 3 time points and 4 unique conditions considered.

(268) Regulons' functionality for each cells evaluated.

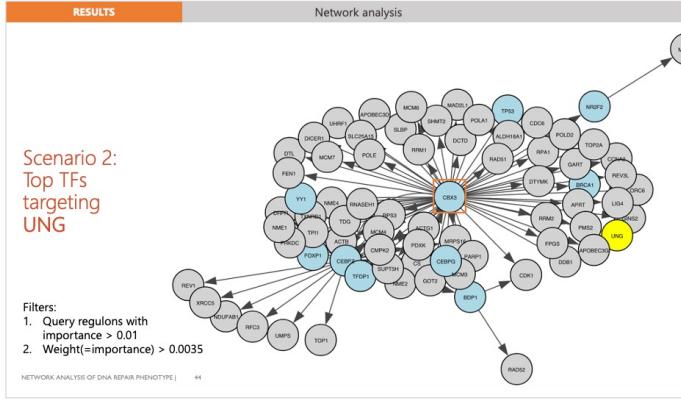
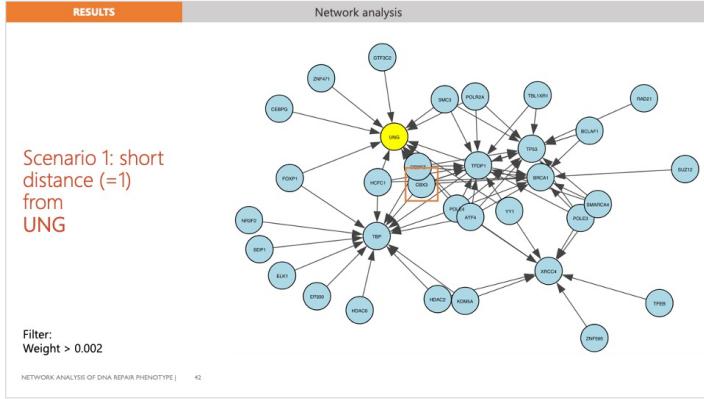
# Build and analyze context-specific networks

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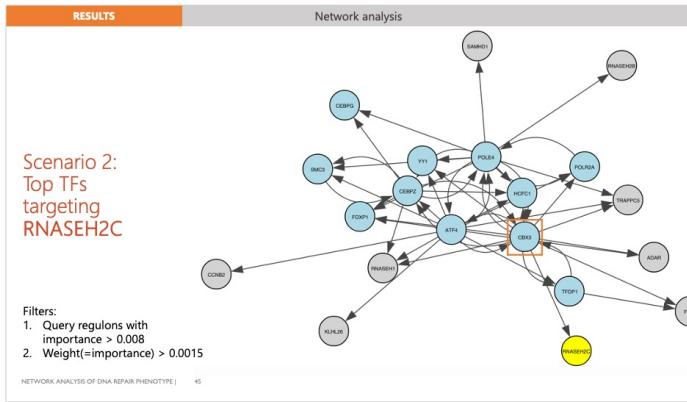
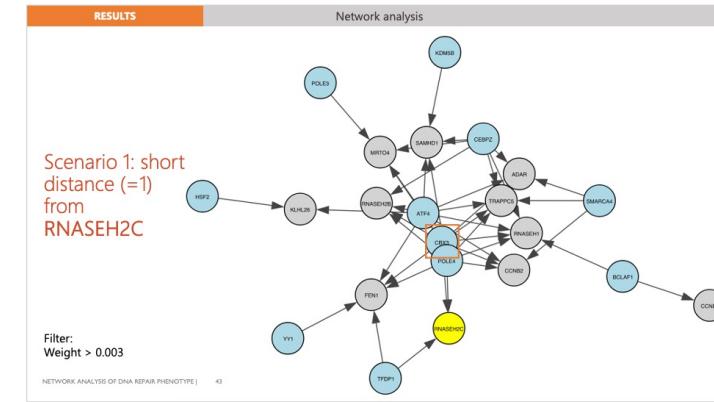
1. Make the large context-specific GRN network
  - Number of vertices in the graph: 18,292
  - Number of edges in the graph 4,652,523
2. Create sub-networks contain nodes from PPI network of KO genes
  1. UNG
    - Number of vertices in the graph: 447
    - Number of edges in the graph 48,703
  2. RNASEH2C
    - Number of vertices in the graph: 281
    - Number of edges in the graph 3,482

# Different scenarios to explore KO subnetworks

UNG Sub-net



RNASEH2C Sub-net

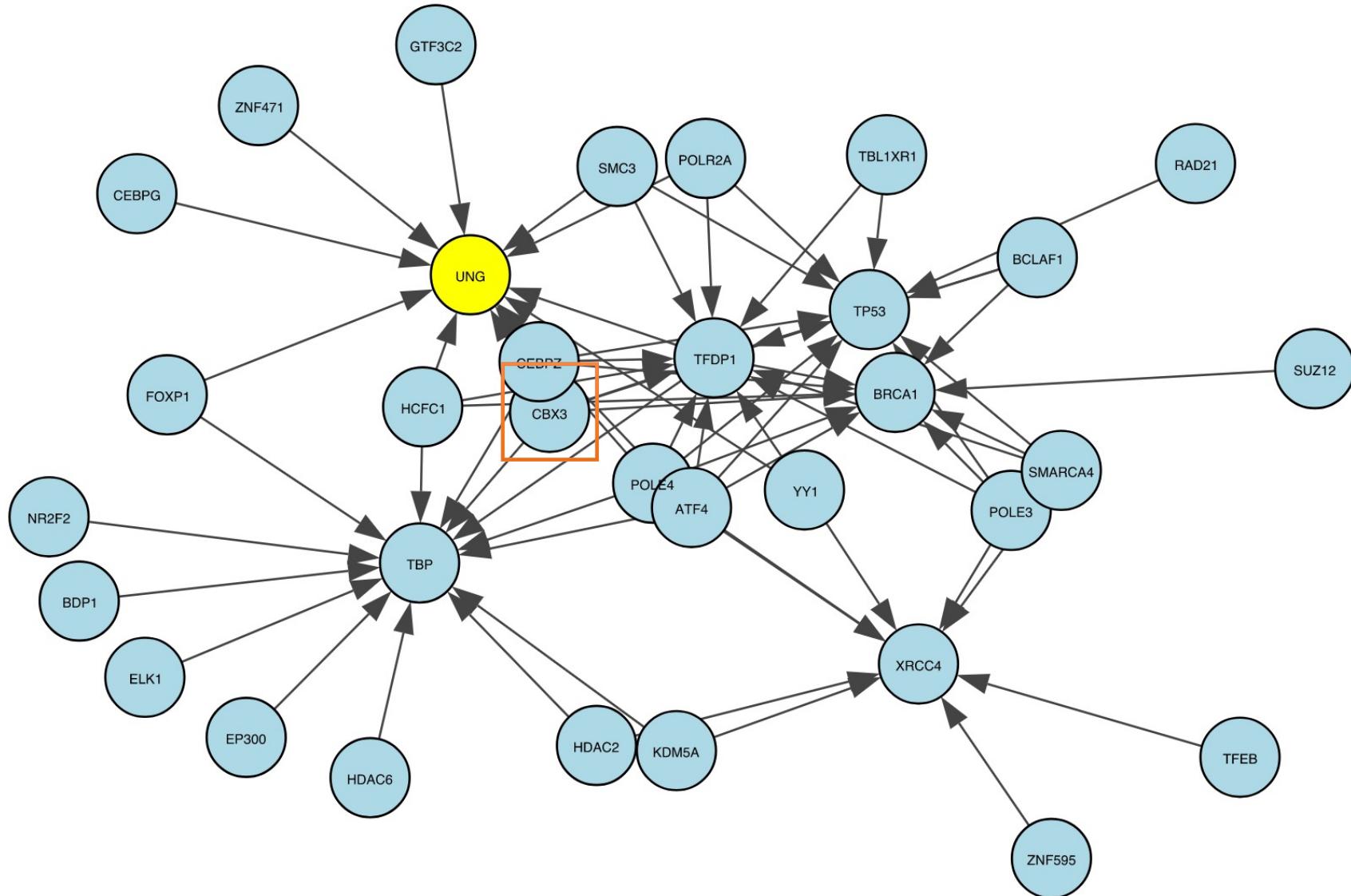


Candidate a regulon with dynamic activity over time



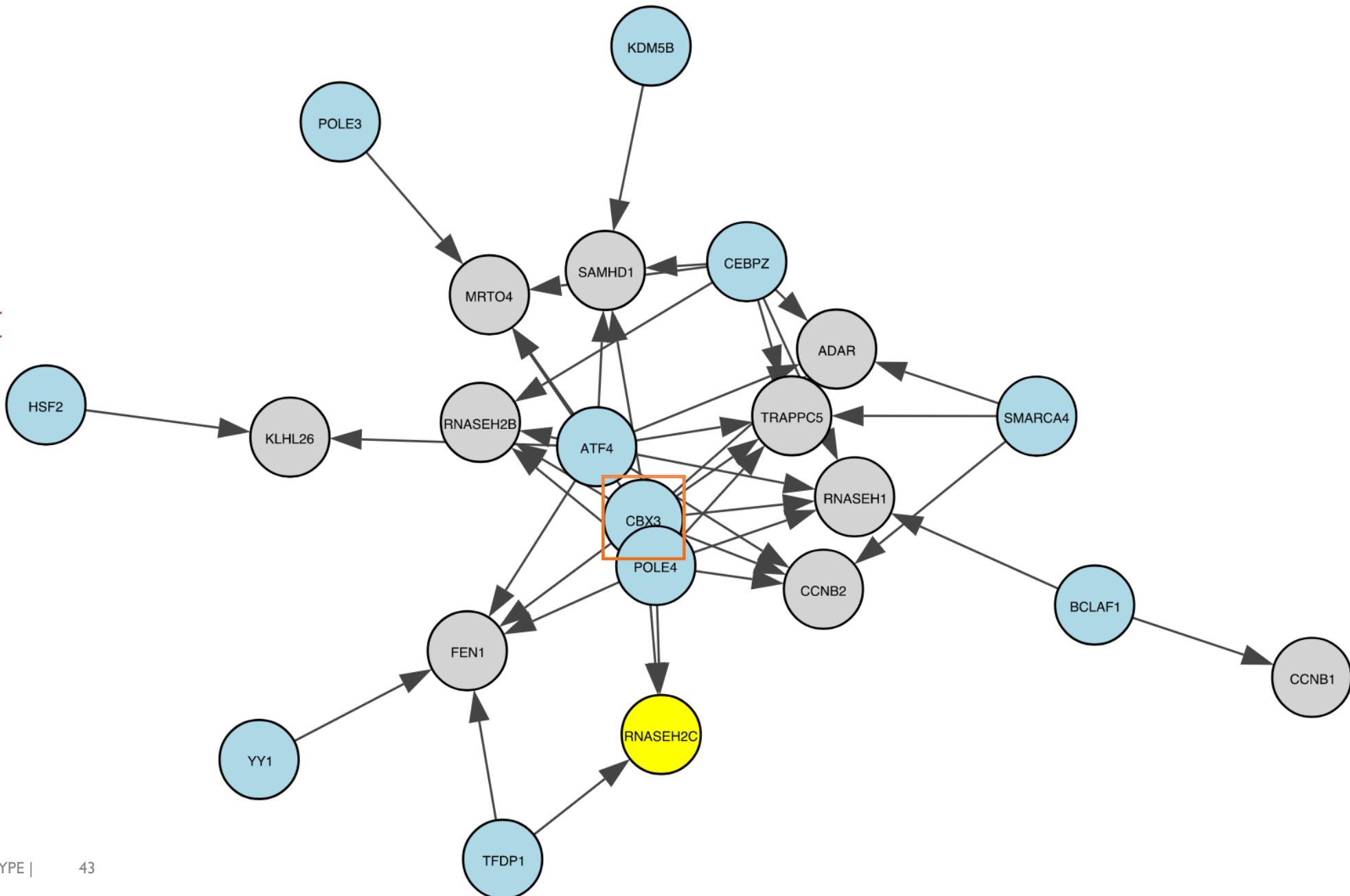
Scenario 1: short  
distance (=1)  
from  
UNG

Filter:  
Weight > 0.002



Scenario 1: short  
distance (=1)  
from  
**RNASEH2C**

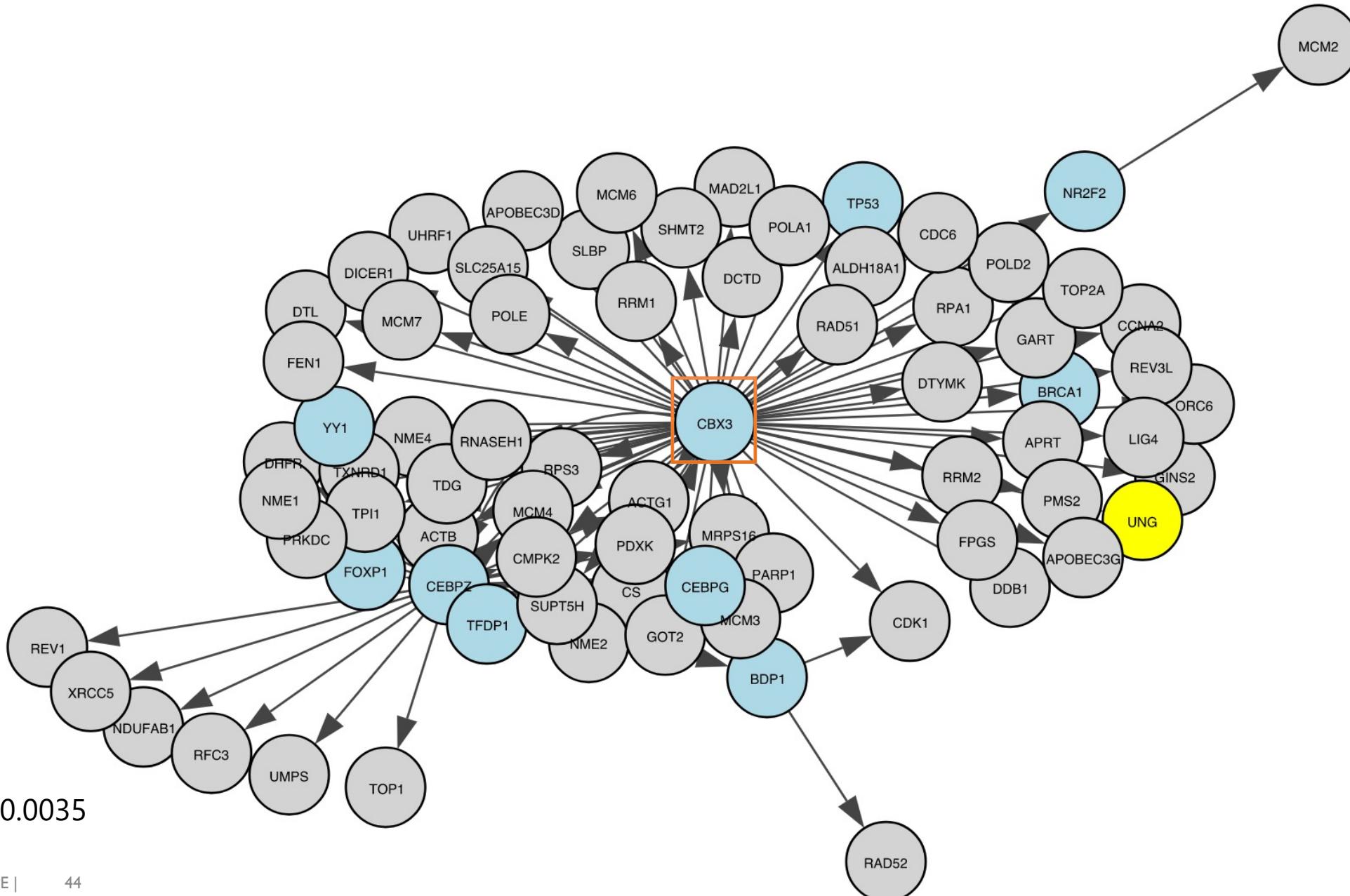
Filter:  
Weight > 0.003



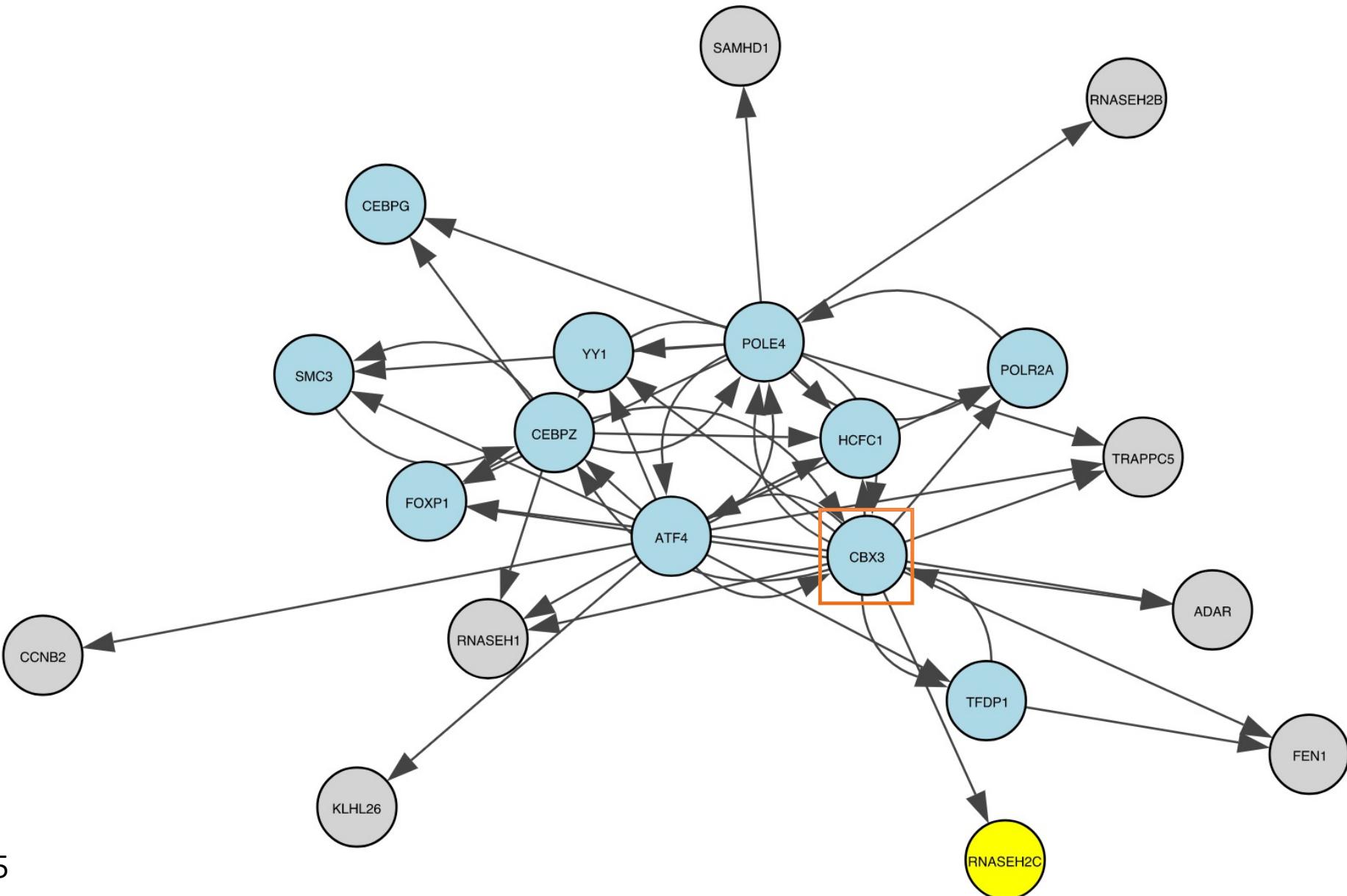
## Scenario 2: Top TFs targeting UNG

Filters:

1. Query regulons with importance > 0.01
2. Weight(=importance) > 0.0035



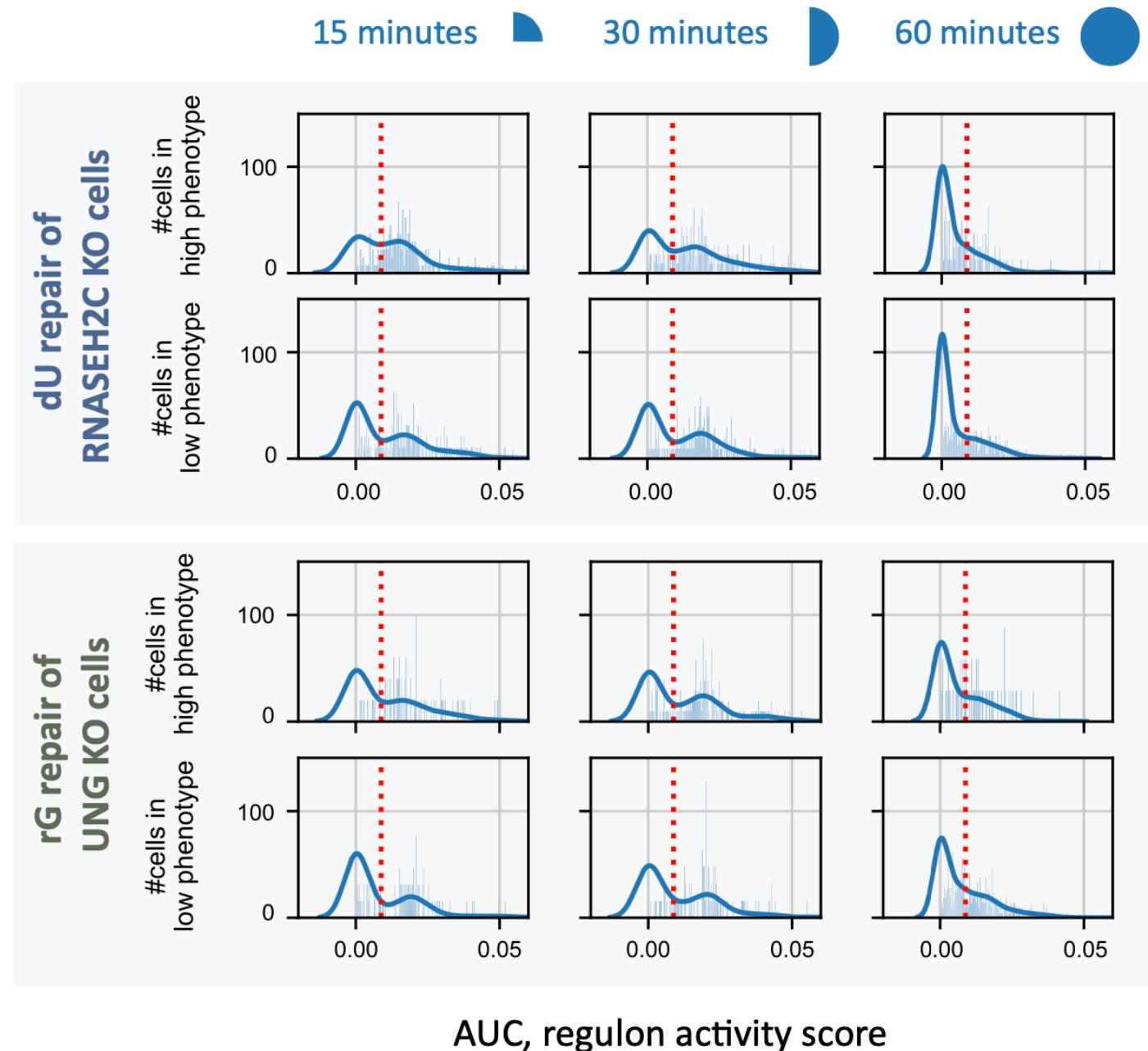
## Scenario 2: Top TFs targeting RNASEH2C



### Filters:

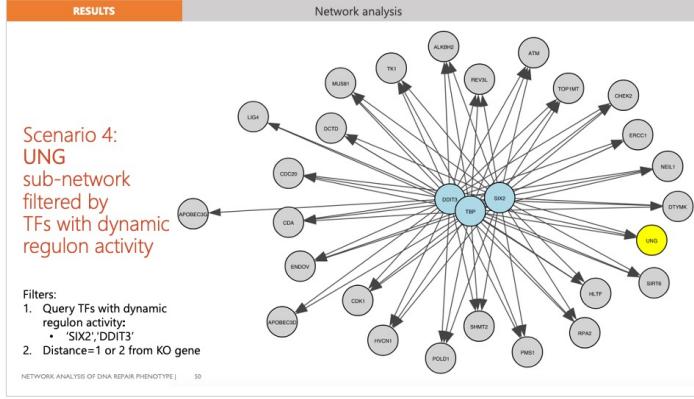
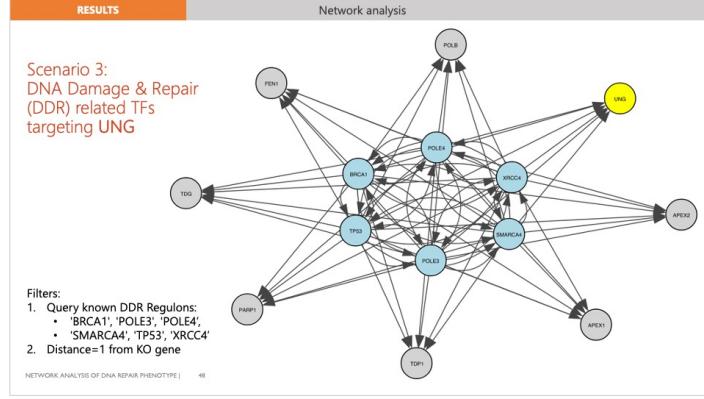
1. Query regulons with importance > 0.008
2. Weight(=importance) > 0.0015

CBX3;  
Chromobox  
protein  
homolog 3

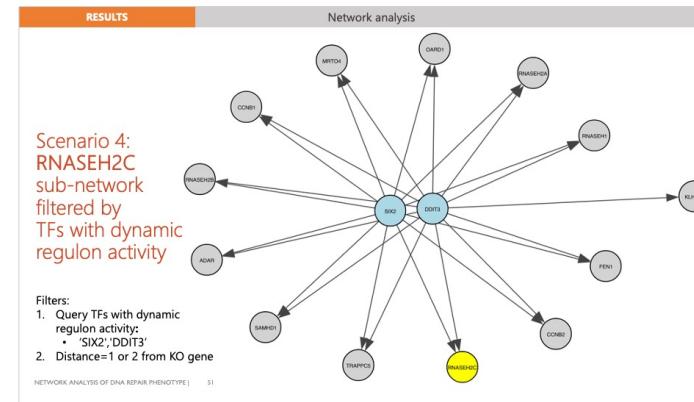
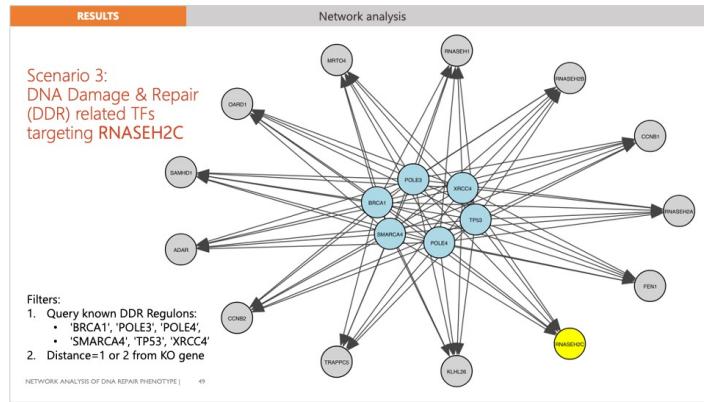


# Different scenarios to explore KO subnetworks

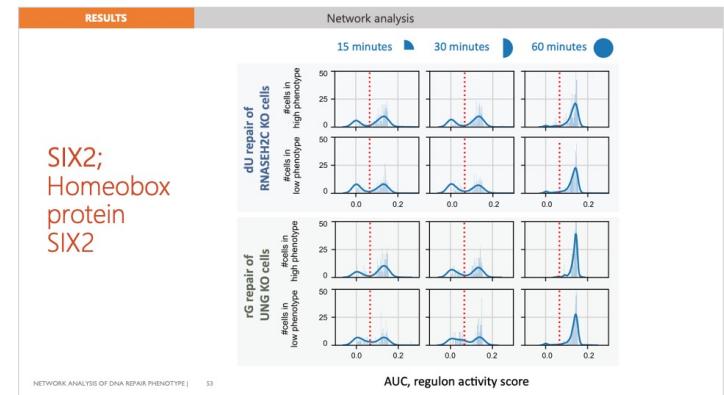
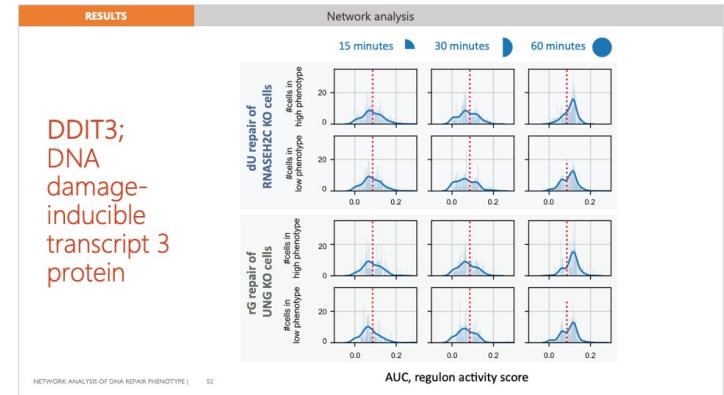
UNG Sub-net



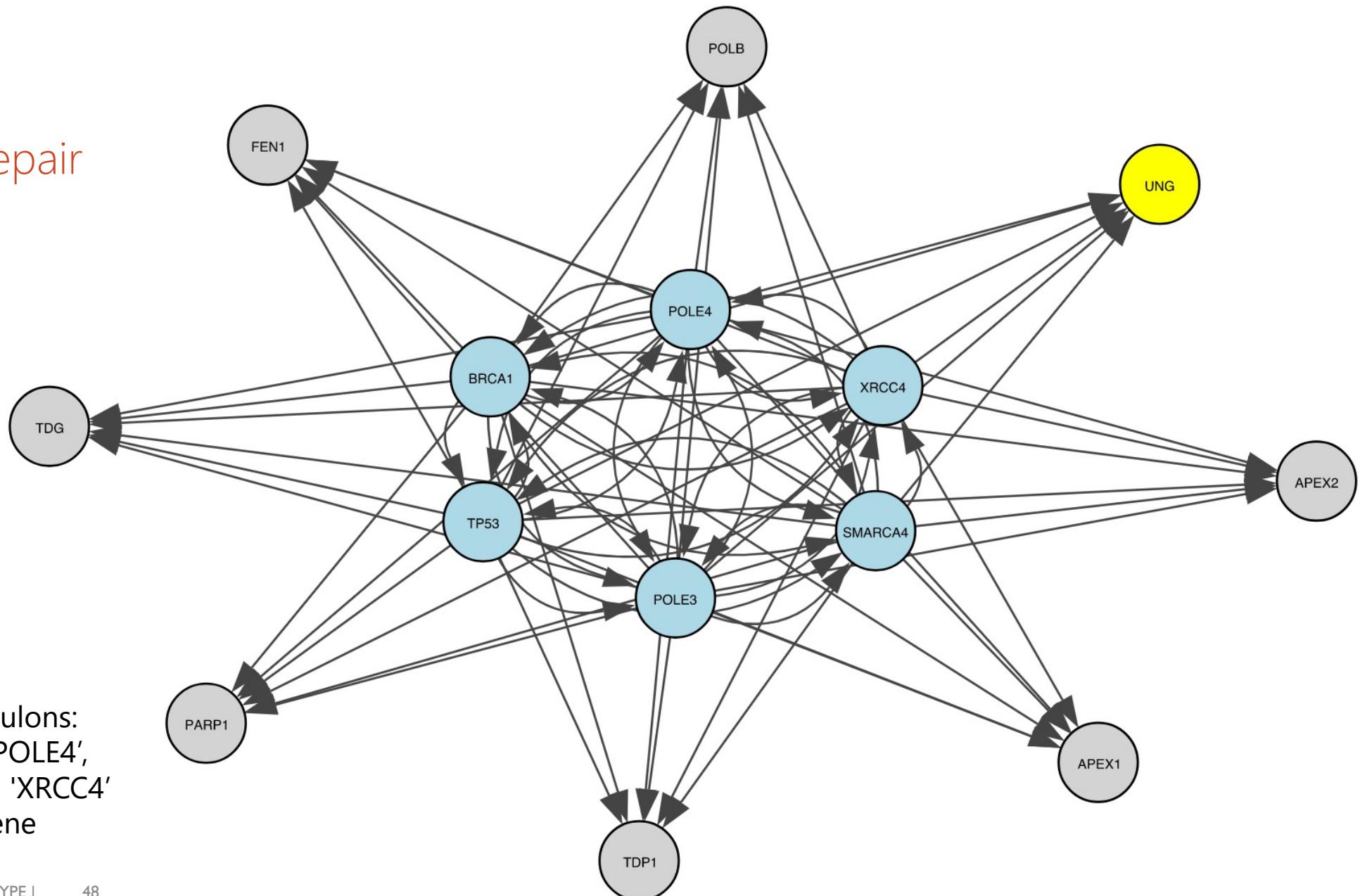
RNASEH2C Sub-net



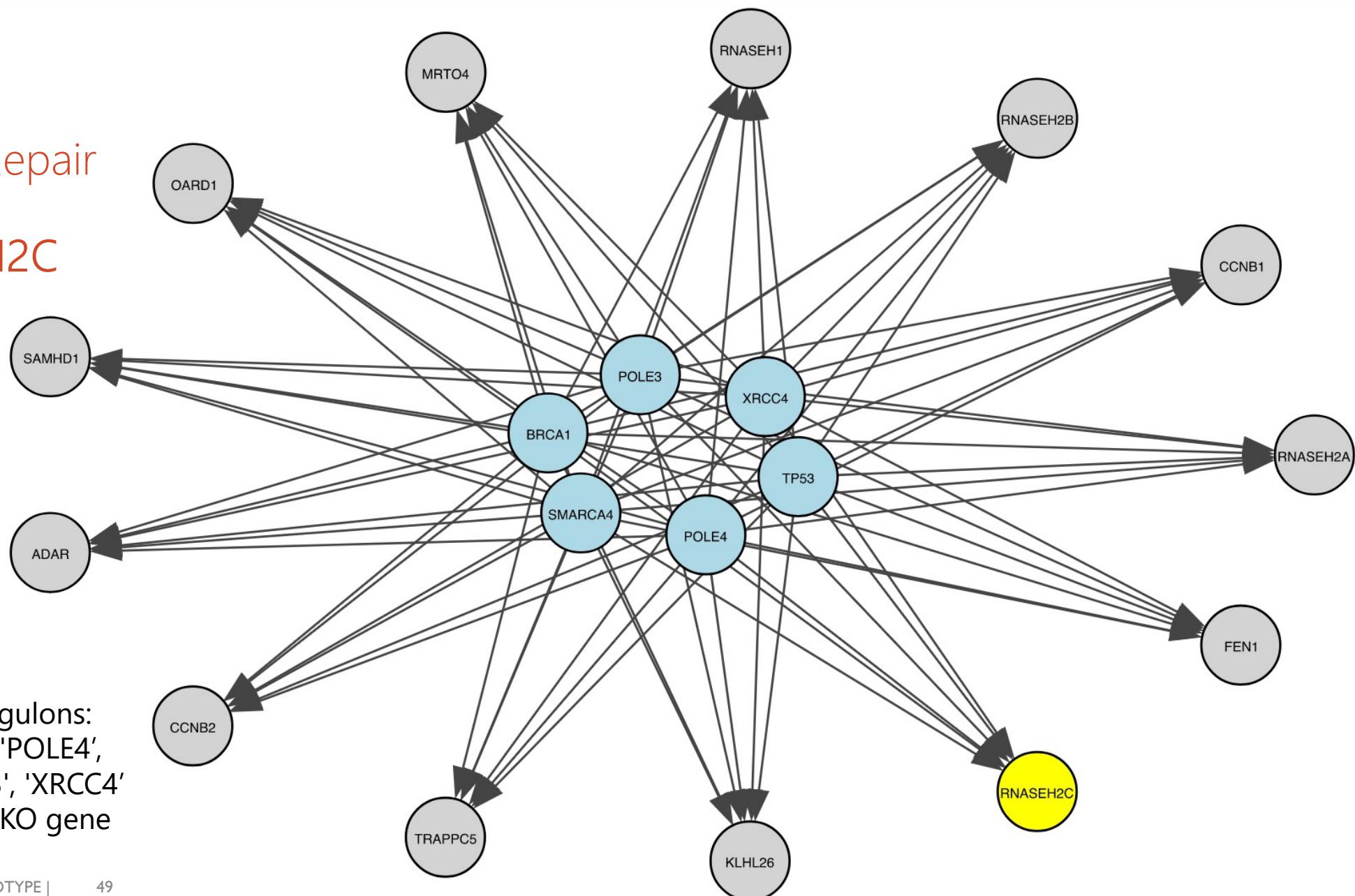
## Candidate regulons with dynamic activity over time



Scenario 3:  
DNA Damage & Repair  
(DDR) related TFs  
targeting UNG



Scenario 3:  
DNA Damage & Repair  
(DDR) related TFs  
targeting RNASEH2C



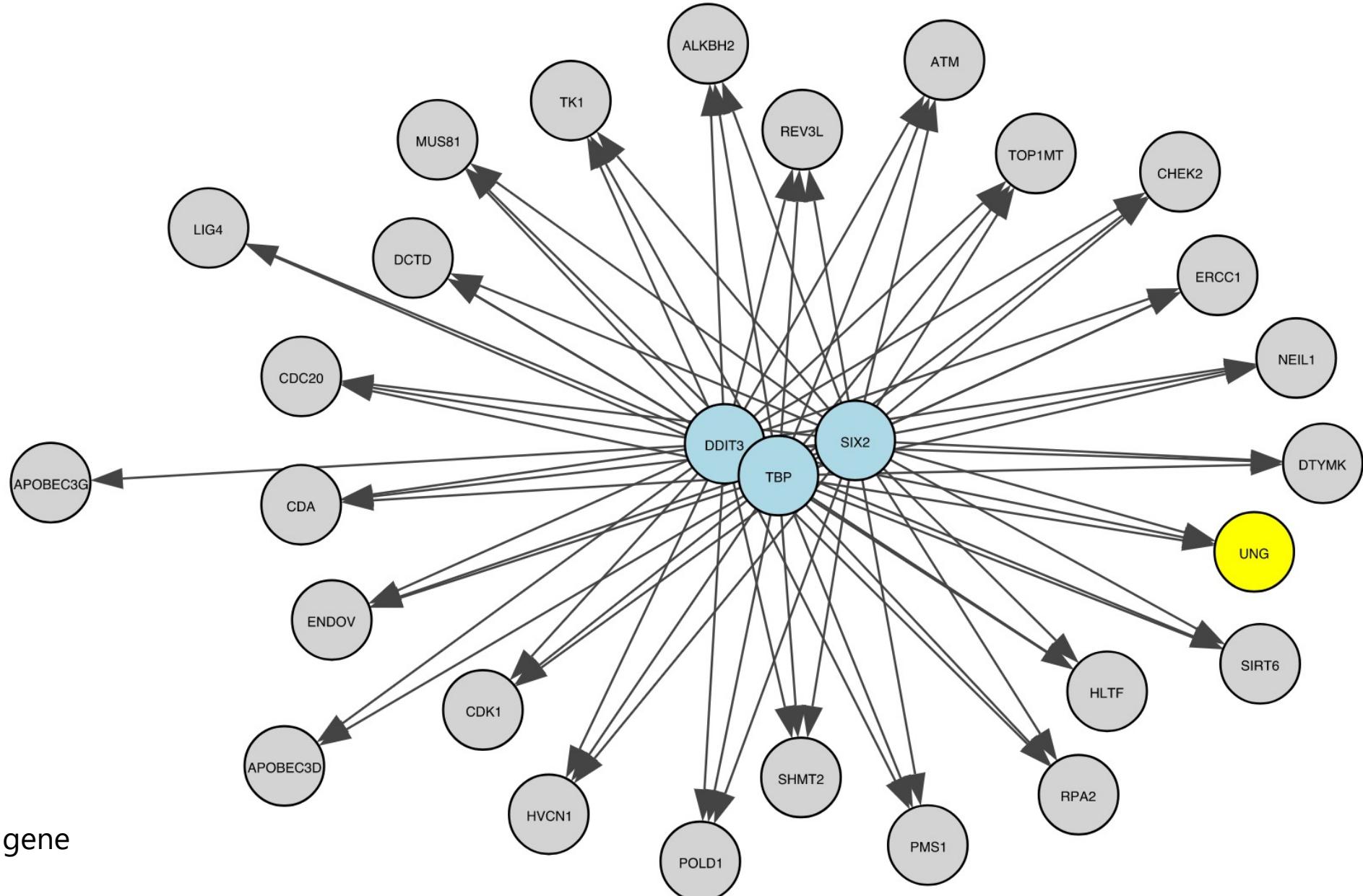
Filters:

1. Query known DDR Regulons:
  - 'BRCA1', 'POLE3', 'POLE4',
  - 'SMARCA4', 'TP53', 'XRCC4'
2. Distance=1 or 2 from KO gene

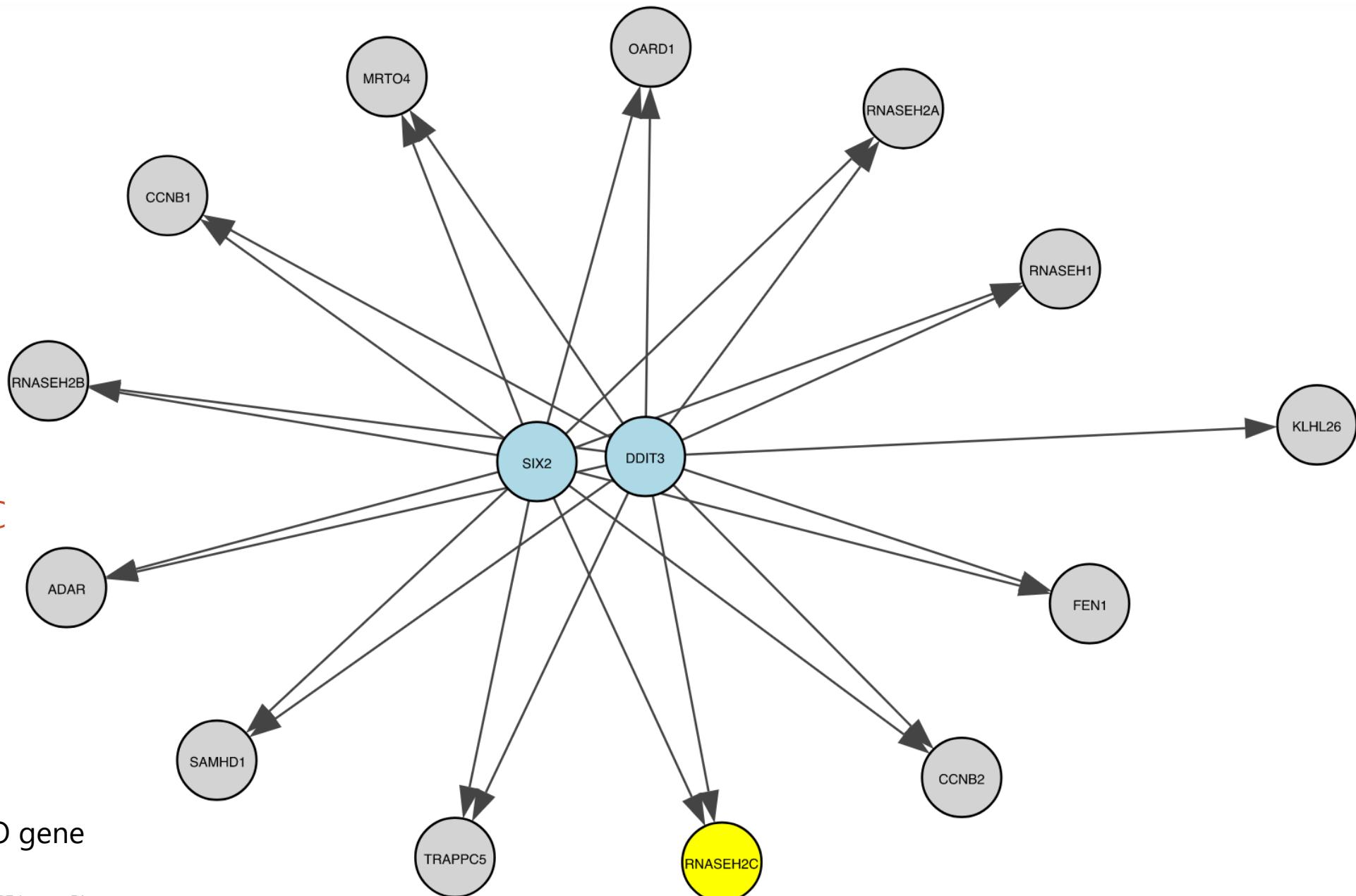
Scenario 4:  
UNG  
sub-network  
filtered by  
TFs with dynamic  
regulon activity

Filters:

1. Query TFs with dynamic regulon activity:
  - 'SIX2', 'DDIT3'
2. Distance=1 or 2 from KO gene



Scenario 4:  
RNASEH2C  
sub-network  
filtered by  
TFs with dynamic  
regulon activity

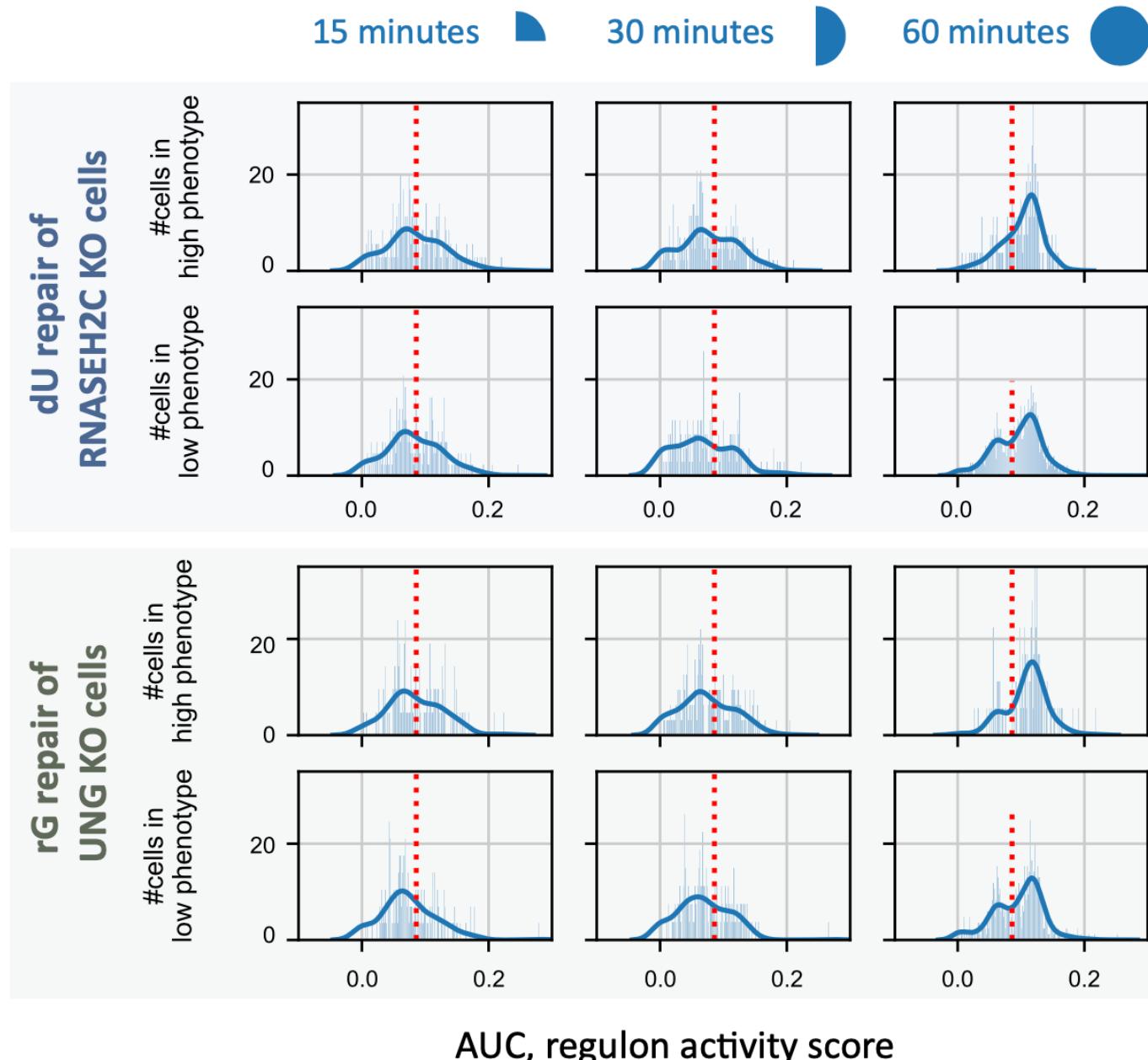


Filters:

1. Query TFs with dynamic regulon activity:
  - 'SIX2', 'DDIT3'
2. Distance=1 or 2 from KO gene

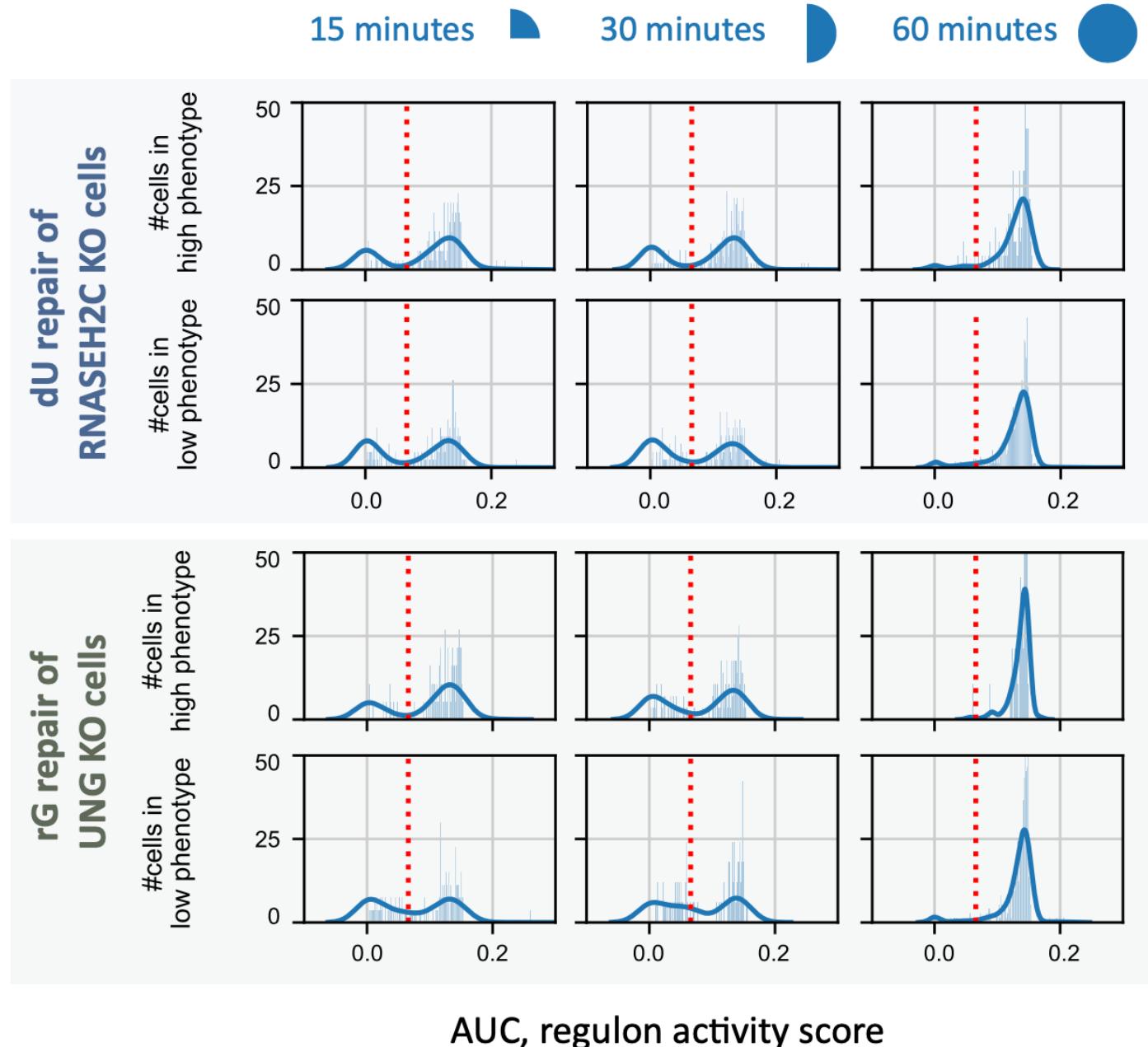
# DDIT3; DNA damage- inducible transcript 3 protein

## Network analysis

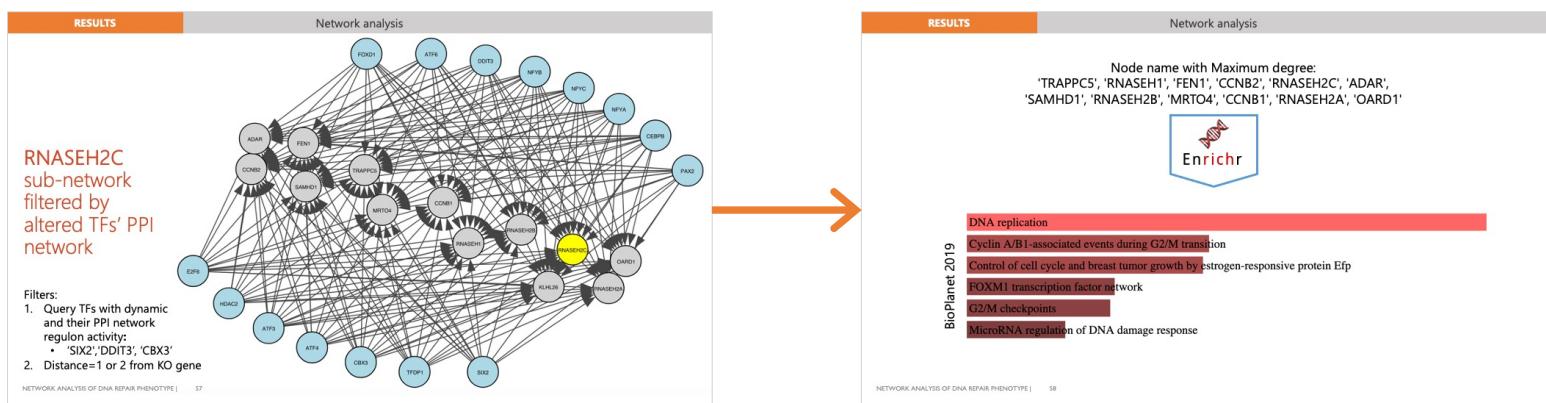
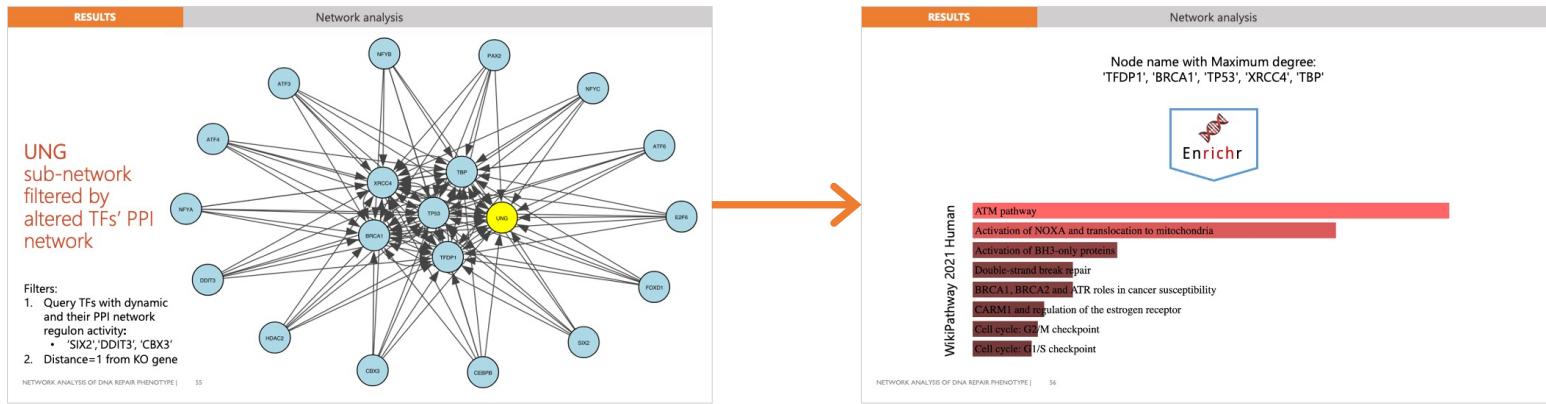


SIX2;  
Homeobox  
protein  
SIX2

## Network analysis



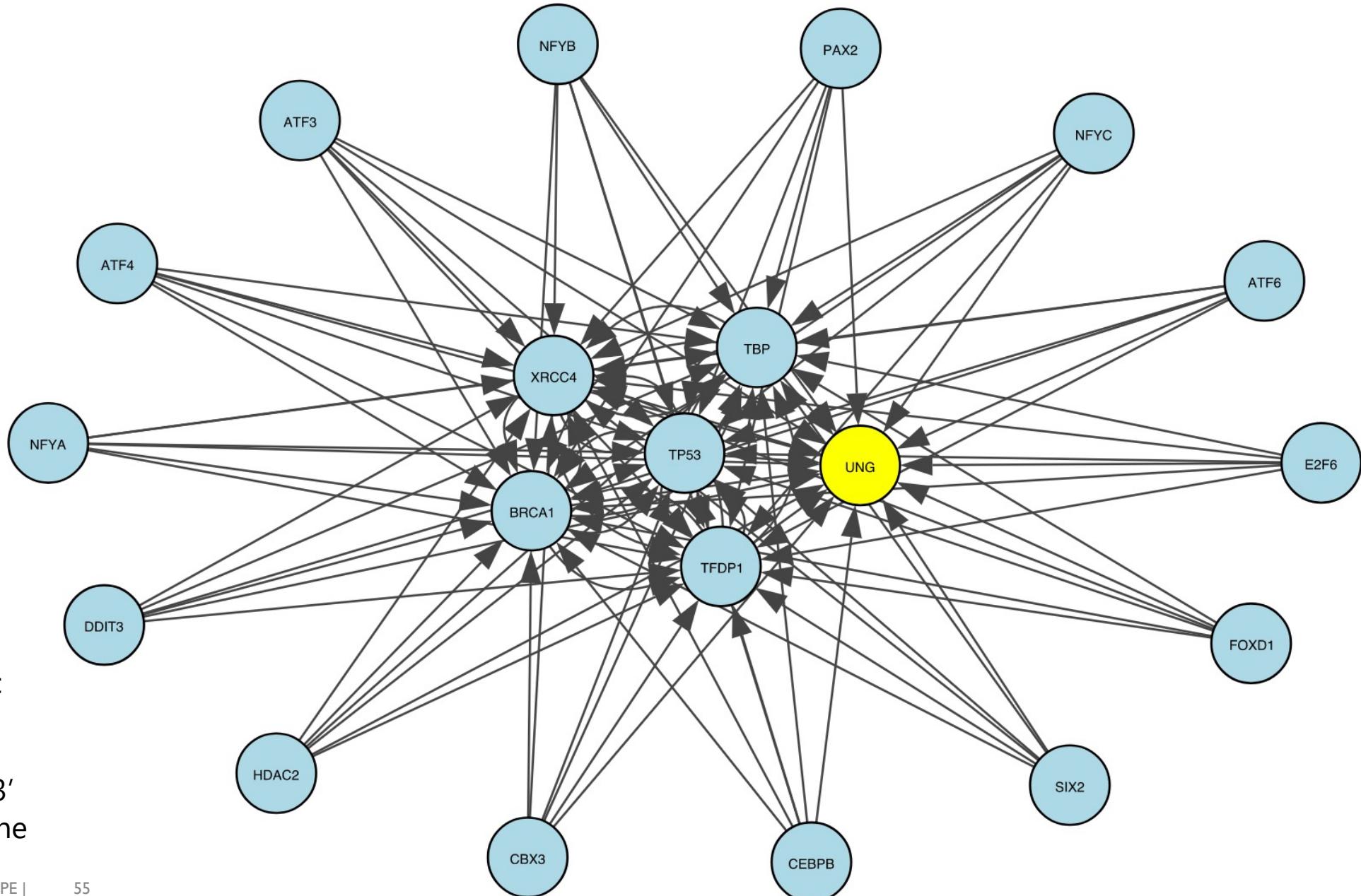
# Filter KO sub-network by altered TFs' PPI network



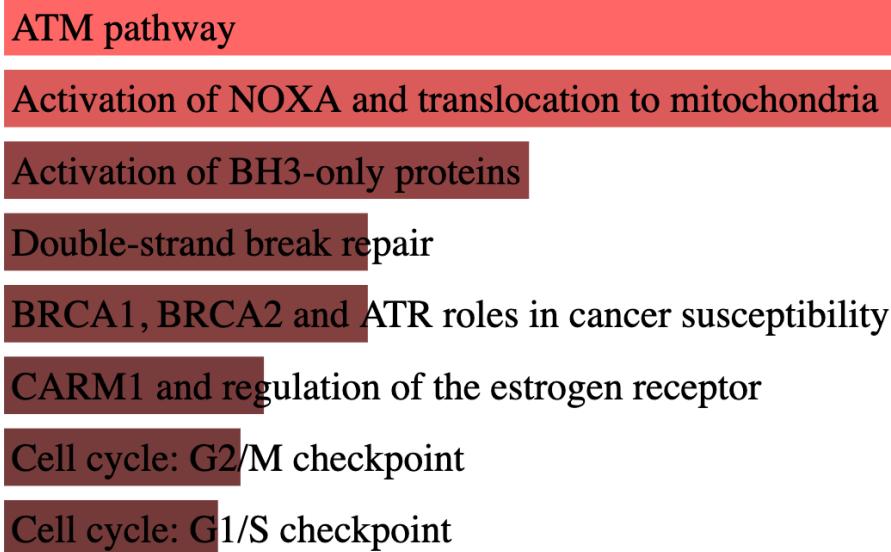
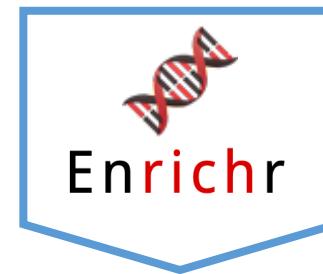
UNG  
sub-network  
filtered by  
altered TFs' PPI  
network

Filters:

1. Query TFs with dynamic and their PPI network regulon activity:
  - 'SIX2', 'DDIT3', 'CBX3'
2. Distance=1 from KO gene



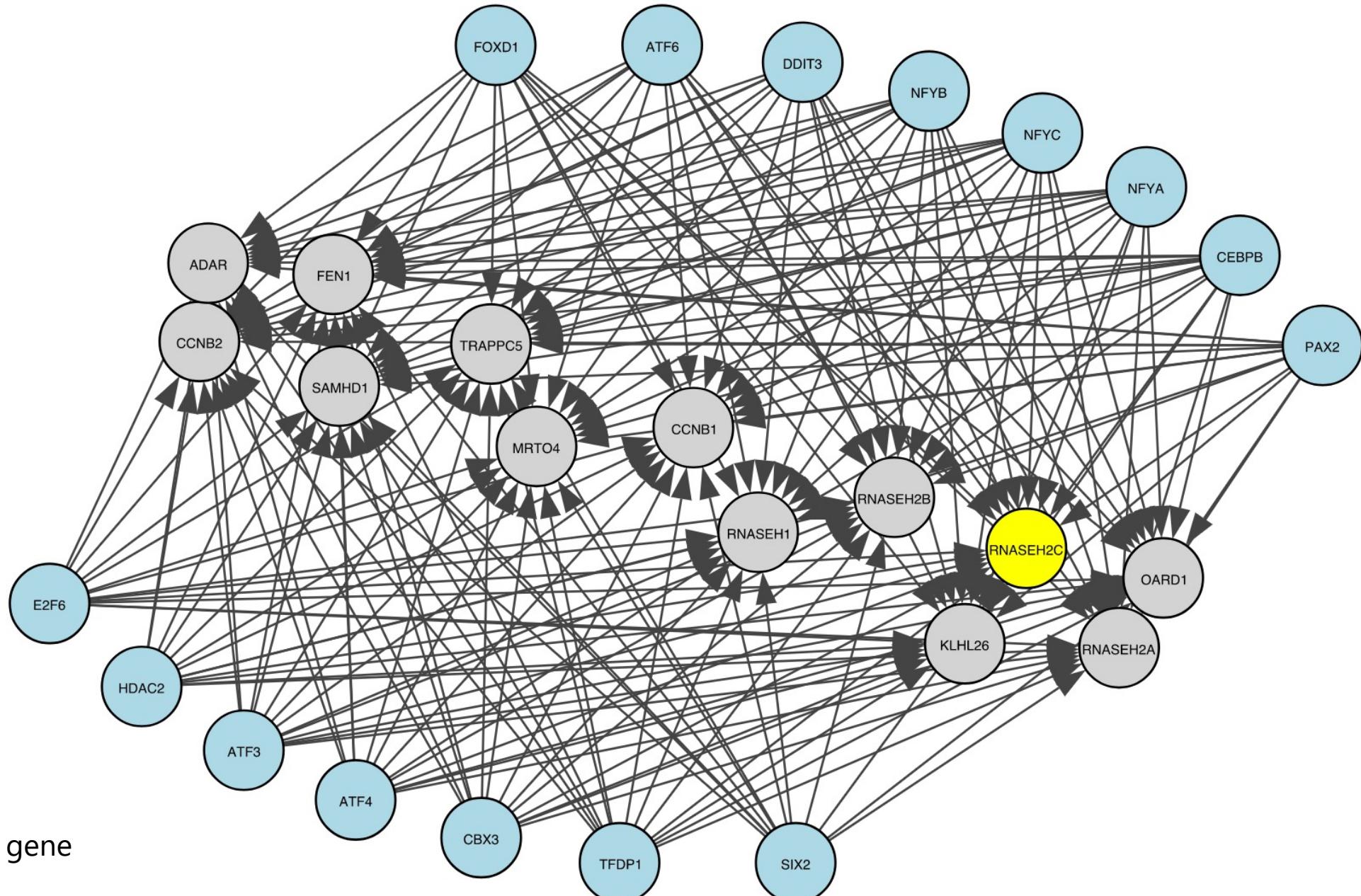
Node name with Maximum degree:  
'TFDP1', 'BRCA1', 'TP53', 'XRCC4', 'TBP'



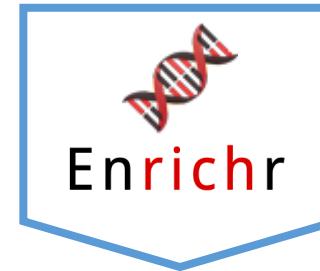
## RNASEH2C sub-network filtered by altered TFs' PPI network

Filters:

1. Query TFs with dynamic and their PPI network regulon activity:
  - 'SIX2', 'DDIT3', 'CBX3'
2. Distance=1 or 2 from KO gene



Node name with Maximum degree:  
'TRAPPC5', 'RNASEH1', 'FEN1', 'CCNB2', 'RNASEH2C', 'ADAR',  
'SAMHD1', 'RNASEH2B', 'MRTO4', 'CCNB1', 'RNASEH2A', 'OARD1'



DNA replication

Cyclin A/B1-associated events during G2/M transition

Control of cell cycle and breast tumor growth by estrogen-responsive protein Efp

FOXM1 transcription factor network

G2/M checkpoints

MicroRNA regulation of DNA damage response

# CONCLUSIONS

- Nano-bio-mimetic DNA damaged hairpins (DNA repair enzyme substrate) induce alterations in cellular gene regulatory network through changing some TF activities, and gene expression over time.
- We observed *CCNB1* over-represented in cells with high dU repair at 60' although it's opposite (over-represented in cells with low phenotype) in rG repair and earlier time repairing dU.
- It suggests potential dynamics of cell cycle due to the presence of DNA damage stimulus.
- RNASEH2C<sup>KO</sup> Cells with high dU-repair might be forbidden to replicate through a cell cycle check point. On the other hand, rG damage might skip the check point.
- Our analysis suggests *SIX2* and *DDIT3* TFs' activity increase by time due to the stimulus.
- CBX3 is a TF with high centrality in the main context-specific network and subnetworks. It seems its activity decrease by time due to the stimulus.