

# Package ‘hpgltools’

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**Type** Package

**Title** A pile of (hopefully) useful R functions

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**Description** This is a set of functions I have been using in my various analyses in the El-Sayed laboratory. The set of tasks included herein run a spectrum from preprocessing count-tables from RNAseq-like data, through differential expression analyses, to post-processing tasks like gene ontology enrichment. Along the way, these function seek to make plotting analyses consistent, provide multiple entry-points to the various tools, and handle corner cases which are not flexibly handled by the packages this is based upon.

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**Suggests** ade4, affy, AnnotationDbi, AnnotationForge, AnnotationHub, BiocGenerics, BiocInstaller, Biostrings, biomaRt, Category, clusterProfiler, corpcor, corrplot, data.table, DBI, DESeq2, DESeq, devtools, directlabels, dplyr, doParallel, DOSE, EDASeq, edgeR, ffpe, fission, genbankr, genefilter, genomeIntervals, GenomeInfoDb, GenomicFeatures, genoPlotR, GenomicRanges, ggden-dro, ggrepel, GO.db, googleVis, goseq, GOstats, gplots, graph, gProfileR, GSEABase, gtools, gridExtra, hash, Heatplus, Hmisc, igraph, inflection, IRanges, jsonlite, KEGGgraph, KEGGREST, knitcitations, lattice, limma, matrixStats, motifRG, multtest, mygene, openxlsx, OrganismDbi, pander, pasilla, pathview, plyr, preprocessCore, qvalue, RamiGO, RColorBrewer, ReactomePA, readr, rentrez, reshape2, RCurl, rGADEM, Rgraphviz, rmarkdown, RMySQL, robustbase, RUVSeq, reshape, rjson, robust, Rsamtools, rtracklayer, S4Vectors, scales, seqinr, seqLogo, statmod, stringi, stringr, survJamda, sva, taxize, testthat, topGO, variancePartition, xtable, XVector

**Imports** Biobase, knitr, ggplot2, magrittr, methods

**VignetteBuilder** knitr

**RoxygenNote** 5.0.1

**R topics documented:**

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all\_ontology\_searches *Perform ontology searches given the results of a differential expression analysis.*

---

**Description**

This takes a set of differential expression results, extracts a subset of up/down expressed genes; passes them to goseq, clusterProfiler, topGO, GOstats, and gProfiler; collects the outputs; and returns them in a (hopefully) consistent fashion. It attempts to handle the differing required annotation/GOid inputs required for each tool and/or provide supported species in ways which the various tools expect.

**Usage**

```
all_ontology_searches(de_out, gene_lengths = NULL, goids = NULL, n = NULL,
  z = NULL, fc = NULL, p = NULL, overwrite = FALSE,
  species = "unsupported", orgdb = "org.Dm.eg.db",
  goid_map = "reference/go/id2go.map", gff_file = NULL, gff_type = "gene",
  do_goseq = TRUE, do_cluster = TRUE, do_topgo = TRUE,
  do_gostats = TRUE, do_gprofiler = TRUE, do_trees = FALSE, ...)
```

**Arguments**

de_out	List of topTables comprising limma/deseq/edger outputs.
gene_lengths	Data frame of gene lengths for goseq.
goids	Data frame of goids and genes.
n	Number of genes at the top/bottom of the fold-changes to define 'significant.'
z	Number of standard deviations from the mean fold-change used to define 'significant.'
fc	Log fold-change used to define 'significant'.
p	Maximum pvalue to define 'significant.'
overwrite	Overwrite existing excel results file?
species	Supported organism used by the tools.
goid_map	Mapping file used by topGO, if it does not exist then goids_df creates it.
gff_file	gff file containing the annotations used by gff2genetable from clusterprofiler.
gff_type	Column to use from the gff file for the universe of genes.
do_goseq	Perform simple_goseq()?
do_cluster	Perform simple_clusterprofiler()?
do_topgo	Perform simple_topgo()?
do_gostats	Perform simple_gostats()?
do_gprofiler	Perform simple_gprofiler()?
do_trees	make topGO trees from the data?

**Value**

a list of up/down ontology results from goseq/clusterprofiler/topgo/gostats, and associated trees.

**Examples**

```
## Not run:
many_comparisons = limma_pairwise(expt=an_expt)
tables = many_comparisons$limma
this_takes_forever = limma_ontology(tables, gene_lengths=lengthdb,
  goids=goids_df, z=1.5, gff_file='length_db.gff')

## End(Not run)
```

---

all_pairwise	<i>Perform limma, DESeq2, EdgeR pairwise analyses.</i>
--------------	--------------------------------------------------------

---

## Description

This takes an expt object, collects the set of all possible pairwise comparisons, sets up experimental models appropriate for the differential expression analyses, and performs them.

## Usage

```
all_pairwise(input, conditions = NULL, batches = NULL, model_cond = TRUE,
             model_batch = TRUE, model_intercept = TRUE, extra_contrasts = NULL,
             alt_model = NULL, libsize = NULL, annot_df = NULL, ...)
```

## Arguments

input	Dataframe/vector or expt class containing count tables, normalization state, etc.
conditions	Factor of conditions in the experiment.
batches	Factor of batches in the experiment.
model_cond	Include condition in the model? This is likely always true.
model_batch	Include batch in the model? This may be true/false/"sva" or other methods supported by get_model_adjust().
model_intercept	Use an intercept model instead of cell means?
extra_contrasts	Optional extra contrasts beyond the pairwise comparisons. This can be pretty neat, let's say one has conditions A,B,C,D,E and wants to do (C/B)/A and (E/D)/A or (E/D)/(C/B) then use this with a string like: "c_vs_b_ctrla = (C-B)-A, e_vs_d_ctrla = (E-D)-A, de_vs_cb = (E-D)-(C-B)".
alt_model	Alternate model to use rather than just condition/batch.
libsize	Library size of the original data to help voom().
annot_df	Annotations to add to the result tables.
...	Picks up extra arguments into arglist, currently only passed to write_limma().

## Value

A list of limma, deseq, edger results.

## Examples

```
## Not run:
finished_comparison = eBayes(limma_output)
data_list = write_limma(finished_comparison, workbook="excel/limma_output.xls")

## End(Not run)
```



---

backup_file	<i>Make a backup of an existing file with n revisions, like VMS!</i>
-------------	----------------------------------------------------------------------

---

**Description**

Sometimes I just want to kick myself for overwriting important files and then I remember using VMS and wish modern computers were a little more like it.

**Usage**

```
backup_file(backup_file, backups = 4)
```

**Arguments**

backup_file	Filename to backup.
backups	How many revisions?

---

basic_pairwise	<i>The simplest possible differential expression method.</i>
----------------	--------------------------------------------------------------

---

**Description**

Perform a pairwise comparison among conditions which takes nothing into account. It `_only_` takes the conditions, a mean value/variance among them, divides by condition, and returns the result. No fancy normalizations, no statistical models, no nothing. It should be the very worst method possible. But, it should also provide a baseline to compare the other tools against, they should all do better than this, always.

**Usage**

```
basic_pairwise(input, design = NULL, force = FALSE, ...)
```

**Arguments**

input	Count table by sample.
design	Data frame of samples and conditions.
force	Force as input non-normalized data?
...	Extra options passed to arglist.

**Value**

Df of pseudo-logFC, p-values, numerators, and denominators.

**See Also****limma DESeq2 edgeR****Examples**

```
## Not run:
stupid_de <- basic_pairwise(expt)

## End(Not run)
```

---

batch_counts	<i>Perform different batch corrections using limma, sva, ruvg, and cbcbs-SEQ.</i>
--------------	-----------------------------------------------------------------------------------

---

**Description**

I found this note which is the clearest explanation of what happens with batch effect data: <https://support.bioconductor.org/p/7>

Just to be clear, there's an important difference between removing a batch effect and modelling a batch effect. Including the batch in your design formula will model the batch effect in the regression step, which means that the raw data are not modified (so the batch effect is not removed), but instead the regression will estimate the size of the batch effect and subtract it out when performing all other tests. In addition, the model's residual degrees of freedom will be reduced appropriately to reflect the fact that some degrees of freedom were "spent" modelling the batch effects. This is the preferred approach for any method that is capable of using it (this includes DESeq2). You would only remove the batch effect (e.g. using limma's `removeBatchEffect` function) if you were going to do some kind of downstream analysis that can't model the batch effects, such as training a classifier. I don't have experience with ComBat, but I would expect that you run it on log-transformed CPM values, while DESeq2 expects raw counts as input. I couldn't tell you how to properly use the two methods together.

**Usage**

```
batch_counts(count_table, design, batch = TRUE, batch1 = "batch",
             batch2 = NULL, noscale = TRUE, ...)
```

**Arguments**

count_table	Matrix of (pseudo)counts.
design	Model matrix defining the experimental conditions/batches/etc.
batch	String describing the method to try to remove the batch effect (or FALSE to leave it alone, TRUE uses limma).
batch1	Column in the design table describing the presumed covariant to remove.
batch2	Column in the design table describing the second covariant to remove (only used by limma at the moment).
noscale	Used for combatmod, when true it removes the scaling parameter from the invocation of the modified combat.
...	More options for you!

**Value**

The 'batch corrected' count table and new library size. Please remember that the library size which comes out of this may not be what you want for voom/limma and would therefore lead to spurious differential expression values.

**See Also**

**limma edgeR RUVSeq sva cbcSEQ**

**Examples**

```
## Not run:
limma_batch <- batch_counts(table, design, batch1='batch', batch2='strain')
sva_batch <- batch_counts(table, design, batch='sva')

## End(Not run)
```

---

bioc\_all

---

*Grab a copy of all bioconductor packages and install them by type*


---

**Description**

This uses jsonlite to get a copy of all bioconductor packages by name and then iterates through them with BiocInstaller to install all of them. It performs a sleep between each installation in an attempt to avoid being obnoxious. As a result, it will of a necessity take forever.

**Usage**

```
bioc_all(release = "3.4", mirror = "bioc.ism.ac.jp", base = "packages",
  type = "software", suppress_updates = TRUE, suppress_auto = TRUE,
  force = FALSE)
```

**Arguments**

release	Bioconductor release to use, should probably be adjusted to automatically find it.
mirror	Bioconductor mirror to use.
base	Base directory on the mirror to download from.
type	Type in the tree to use (software, annotation, or experiment)
force	Install if already installed?
suppressUpdates	For BiocLite(), don't update?
suppressAutoUpdate	For BiocLite(), don't update?

**Value**

a number of packages installed

---

biomart_orthologs	<i>Use biomart to get orthologs between supported species.</i>
-------------------	----------------------------------------------------------------

---

### Description

Biomart's function getLDS is incredibly powerful, but it makes me think very polite people are going to start knocking on my door, and it fails weirdly pretty much always. This function attempts to alleviate some of that frustration.

### Usage

```
biomart_orthologs(gene_ids, first_species = "hsapiens",
  second_species = "mmusculus", host = "dec2015.archive.ensembl.org",
  trymart = "ENSEMBL_MART_ENSEMBL")
```

### Arguments

gene_ids	List of gene IDs to translate.
first_species	Linnean species name for one species.
second_species	Linnean species name for the second species.
host	Ensembl server to query.
trymart	Assumed mart name to use.

### Value

Df of orthologs.

---

cbcb_batch_effect	<i>A function suggested by Hector Corrada Bravo and Kwame Okrah for batch removal</i>
-------------------	---------------------------------------------------------------------------------------

---

### Description

During a lab meeting, the following function was suggested as a quick and dirty batch removal tool

### Usage

```
cbcb_batch_effect(normalized_counts, model)
```

### Arguments

normalized_counts	Data frame of log2cpm counts.
model	Balanced experimental model containing condition and batch factors.

**Value**

Dataframe of residuals after subtracting batch from the model.

**See Also**

[voom lmFit](#)

**Examples**

```
## Not run:
newdata <- cbc_b_batch_effect(counts, expt_model)

## End(Not run)
```

---

cbcb_filter_counts	<i>Filter low-count genes from a data set using cpm data and a threshold.</i>
--------------------	-------------------------------------------------------------------------------

---

**Description**

This was a function written by Kwame Okrah and perhaps also Laura Dillon to remove low-count genes. It drops genes based on a cpm threshold and number of samples.

**Usage**

```
cbcb_filter_counts(count_table, threshold = 2, min_samples = 2)
```

**Arguments**

count_table	Data frame of (pseudo)counts by sample.
threshold	Lower threshold of counts for each gene.
min_samples	Minimum number of samples.

**Value**

Dataframe of counts without the low-count genes.

**Examples**

```
## Not run:
filtered_table <- cbc_b_filter_counts(count_table)

## End(Not run)
```

---

check\_clusterprofiler *Make sure that clusterProfiler is ready to run.*

---

### Description

Many of our ontology searches are using non-supported organisms. These need to have a geneTable.rda file in place which maps the gene IDs to GO IDs. This function checks for that file and attempts to set it up if it is not found.

### Usage

```
check_clusterprofiler(gff = "test.gff", goids_df = NULL)
```

### Arguments

gff	Ggff file containing annotation data (gene lengths).
goids_df	Data frame of gene IDs and GO ontologies 1:1, other columns are ignored.

### Value

GO2EG data structure created, probably don't save this, it is entirely too big.

### Examples

```
## Not run:
go2eg <- check_clusterprofiler(gff, goids_df)
rm(go2eg)

## End(Not run)
```

---

choose\_dataset *Choose a suitable data set for Edger/DESeq*

---

### Description

The \_pairwise family of functions all demand data in specific formats. This tries to make that consistent.

### Usage

```
choose_dataset(input, force = FALSE, ...)
```

### Arguments

input	Expt input.
force	Force non-standard data
...	More options for future expansion

**Value**

List the data, conditions, and batches in the data.

---

choose_model	<i>Try out a few experimental models and return a likely working option.</i>
--------------	------------------------------------------------------------------------------

---

**Description**

The `_pairwise` family of functions all demand an experimental model. This tries to choose a consistent and useful model for all for them. This does not try to do multi-factor, interacting, nor dependent variable models, if you want those do them yourself and pass them off as `alt_model`.

**Usage**

```
choose_model(conditions, batches, model_batch = TRUE, model_cond = TRUE,  
             model_intercept = TRUE, alt_model = NULL, alt_string = NULL,  
             intercept = 0, reverse = FALSE)
```

**Arguments**

<code>conditions</code>	Factor of conditions in the putative model.
<code>batches</code>	Factor of batches in the putative model.
<code>model_batch</code>	Try to include batch in the model?
<code>model_cond</code>	Try to include condition in the model? (Yes!)
<code>model_intercept</code>	Use an intercept model instead of cell-means?
<code>alt_model</code>	Use your own model.
<code>alt_string</code>	String describing an alternate model.

**Value**

List including a model matrix and strings describing cell-means and intercept models.

---

choose\_orgdb

*Load the appropriate orgDb environment for a given species.*


---

### Description

Ok, so these are a bit more complex than I realized. The heirarchy as I now understand it (probably wrong) is that orgdb objects provide ID mappings among the various DBs. txdb objects provide the actual annotation information, and organismdbs acquire both (but only exist for a few species). Let's face it, I will never remember that the yeast orgdb is 'org.Sc.sgd.something'. This function is intended to make that process easier. Feed it a species name which makes sense: 'homo\_sapiens' and it will assume you mean orgdb.whatever and load that into your environment. This should also make a reasonable attempt at installing the appropriate orgdb if it is not already in your R library tree.

### Usage

```
choose_orgdb(species = "saccharomyces_cerevisiae")
```

### Arguments

species                      Human readable species name

### Value

orgdb object for the relevant species, or an error if I don't have a mapping for it.

### Examples

```
object <- choose_orgdb("homo_sapiens")
```

---

choose\_txdb

*Load the appropriate TxDb environment for a given species.*


---

### Description

Ok, so these are a bit more complex than I realized. The heirarchy as I now understand it (probably wrong) is that orgdb objects provide ID mappings among the various DBs. txdb objects provide the actual annotation information, and organismdbs acquire both (but only exist for a few species). Let's face it, I will never remember that the yeast orgdb is 'org.Sc.sgd.something'. This function is intended to make that process easier. Feed it a species name which makes sense: 'homo\_sapiens' and it will assume you mean orgdb.whatever and load that into your environment. This should also make a reasonable attempt at installing the appropriate orgdb if it is not already in your R library tree.

### Usage

```
choose_txdb(species = "saccharomyces_cerevisiae")
```



**Arguments**

species            Human readable species name

**Value**

orgdb object for the relevant species, or an error if I don't have a mapping for it.

**Examples**

```
object <- choose_txdb("homo_sapiens")
```

---

circos_arc	<i>Write arcs between chromosomes in circos.</i>
------------	--------------------------------------------------

---

**Description**

Ok, so when I said I only do 1 chromosome images, I lied. This function tries to make writing arcs between chromosomes easier. It too works in 3 stages, It writes out a data file using cfgout as a basename and the data from df in the circos arc format into circos/data/bob\_arc.txt It then writes out a configuration plot stanza in circos/conf/bob\_arc.conf and finally adds an include to circos/bob.conf

**Usage**

```
circos_arc(df, cfgout = "circos/conf/default.conf", first_col = "chr1",
  second_col = "chr2", color = "blue", radius = 0.75, thickness = 3)
```

**Arguments**

df	Dataframe with starts/ends and the floating point information.
cfgout	Master configuration file to write.
first_col	Name of the first chromosome.
second_col	Name of the second chromosome.
color	Color of the chromosomes.
radius	Outer radius at which to add the arcs.
thickness	Integer thickness of the arcs.

**Details**

In its current implementation, this only understands two chromosomes. A minimal amount of logic and data organization will address this weakness.

**Value**

The file to which the arc configuration information was written.

---

circos_heatmap	<i>Write tiles of arbitrary heat-mappable data in circos.</i>
----------------	---------------------------------------------------------------

---

### Description

This function tries to make the writing circos heatmaps easier. Like `circos_plus_minus()` and `circos_hist()` it works in 3 stages, It writes out a data file using `cfgout` as a basename and the data from `df` in the circos histogram format into `circos/data/bob_heatmap.txt` It then writes out a configuration plot stanza in `circos/conf/bob_heatmap.conf` and finally adds an include to `circos/bob.conf`

### Usage

```
circos_heatmap(df, annot_df, cfgout = "circos/conf/default.conf",
               colname = "logFC", chr = "chr1", colors = NULL, outer = 0.9,
               width = 0.08, spacing = 0)
```

### Arguments

<code>df</code>	Dataframe with starts/ends and the floating point information.
<code>annot_df</code>	Annotation data frame with starts/ends.
<code>cfgout</code>	Master configuration file to write.
<code>colname</code>	Name of the column with the data of interest.
<code>chr</code>	Name of the chromosome (This currently assumes a bacterial chromosome).
<code>colors</code>	Colors of the heat map.
<code>outer</code>	Floating point radius of the circle into which to place the heatmap.
<code>width</code>	Width of each tile in the heatmap.
<code>spacing</code>	Radial distance between outer, inner, and inner to whatever follows.

### Value

Radius after adding the histogram and the spacing.

---

circos_hist	<i>Write histograms of arbitrary floating point data in circos.</i>
-------------	---------------------------------------------------------------------

---

### Description

This function tries to make the writing of histogram data in circos easier. Like `circos_plus_minus()` it works in 3 stages, It writes out a data file using `cfgout` as a basename and the data from `df` in the circos histogram format into `circos/data/bob_hist.txt` It then writes out a configuration plot stanza in `circos/conf/bob_hist.conf` and finally adds an include to `circos/bob.conf`

**Usage**

```
circos_hist(df, annot_df, cfgout = "circos/conf/default.conf",
            colname = "logFC", chr = "chr1", color = "blue", fill_color = "blue",
            outer = 0.9, width = 0.08, spacing = 0)
```

**Arguments**

df	Dataframe with starts/ends and the floating point information.
annot_df	Annotation data frame containing starts/ends.
cfgout	Master configuration file to write.
colname	Name of the column with the data of interest.
chr	Name of the chromosome (This currently assumes a bacterial chromosome).
color	Color of the plotted data.
fill_color	Guess!
outer	Floating point radius of the circle into which to place the data.
width	Radial width of each tile.
spacing	Distance between outer, inner, and inner to whatever follows.

**Value**

Radius after adding the histogram and the spacing.

---

circos_ideogram	<i>Create the description of chromosome markings.</i>
-----------------	-------------------------------------------------------

---

**Description**

This function writes ideogram files for circos.

**Usage**

```
circos_ideogram(name = "default", conf_dir = "circos/conf",
               band_url = NULL)
```

**Arguments**

name	Name of the configuration file to which to add the ideogram.
conf_dir	Where does the configuration live?
band_url	Provide a url for making these imagemaps?

**Value**

The file to which the ideogram configuration was written.

---

circos_karyotype	<i>Create the description of (a)chromosome(s) for circos.</i>
------------------	---------------------------------------------------------------

---

### Description

This function tries to save me from having to get the lengths of arcs for bacterial chromosomes manually correct, and writes them as a circos compatible karyotype file. The outfile parameter was chosen to match the configuration directive outlined in `circos_prefix()`, however that will need to be changed in order for this to work in variable conditions. Next time I make one of these graphs I will do that I suspect. In addition, this currently only understands how to write bacterial chromosomes, that will likely be fixed when I am asked to write out a L.major karyotype. These defaults were chosen because I have a chromosome of this length that is correct.

### Usage

```
circos_karyotype(name = "default", conf_dir = "circos/conf",
  length = NULL, chr_name = "chr1", segments = 6, color = "white",
  chr_num = 1, fasta = NULL)
```

### Arguments

name	Name of the chromosome (This currently assumes a bacterial chromosome).
conf_dir	Where to put the circos configuration file(s).
length	Length of the chromosome (the default is mgas5005).
chr_name	Short name of the chromosome.
segments	How many segments to cut the chromosome into?
color	Color segments of the chromosomal arc?
chr_num	Number to record for each chromosome.
fasta	Fasta file to use to create the karyotype.

### Value

The output filename.

---

circos_make	<i>Write a simple makefile for circos.</i>
-------------	--------------------------------------------

---

### Description

I regenerate all my circos pictures with `make(1)`. This is my makefile.

### Usage

```
circos_make(target = "", output = "circos/Makefile",
  circos = "/usr/bin/circos")
```

**Arguments**

target	Default make target.
output	Makefile to write.
circos	Location of circos. I have a copy in home/bin/circos and use that sometimes.

**Value**

a kitten

---

circos_plus_minus	<i>Write tiles of bacterial ontology groups using the categories from microbesonline.org.</i>
-------------------	-----------------------------------------------------------------------------------------------

---

**Description**

This function tries to save me from writing out ontology definitions and likely making mistakes. It uses the start/ends from the gff annotation along with the 1 letter GO-like categories from microbesonline.org. It then writes two data files circos/data/bob\_plus\_go.txt, circos/data/bob\_minus\_go.txt along with two configuration files circos/conf/bob\_minus\_go.conf and circos/conf/bob\_plus\_go.conf and finally adds an include to circos/bob.conf

**Usage**

```
circos_plus_minus(go_table, cfgout = "circos/conf/default.conf",
  chr = "chr1", outer = 1, width = 0.08, spacing = 0)
```

**Arguments**

go_table	Dataframe with starts/ends and categories.
cfgout	Master configuration file to write.
chr	Name of the chromosome.
outer	Floating point radius of the circle into which to place the plus-strand data.
width	Radial width of each tile.
spacing	Radial distance between outer, inner, and inner to whatever follows.

**Value**

Radius after adding the plus/minus information and the spacing between them.

---

circos_prefix	<i>Write the beginning of a circos configuration file.</i>
---------------	------------------------------------------------------------

---

### Description

A few parameters need to be set when starting circos. This sets some of them and gets ready for plot stanzas.

### Usage

```
circos_prefix(name = "mgas", conf_dir = "circos/conf", radius = 1800,
  band_url = NULL)
```

### Arguments

name	Name of the map, called with 'make name'.
conf_dir	Directory containing the circos configuration data.
radius	Size of the image.
band_url	Place to imagemap link.

### Details

In its current implementation, this really assumes that there will be no highlight stanzas and at most 1 link stanza. chromosomes. A minimal amount of logic and data organization will address these weaknesses.

### Value

The master configuration file name.

---

circos_suffix	<i>Write the end of a circos master configuration.</i>
---------------	--------------------------------------------------------

---

### Description

circos configuration files need an ending. This writes it.

### Usage

```
circos_suffix(cfgout = "circos/conf/default.conf")
```

### Arguments

cfgout	Master configuration file to write.
--------	-------------------------------------

**Value**

The filename of the configuration.

---

circos_tile	<i>Write tiles of arbitrary categorical point data in circos.</i>
-------------	-------------------------------------------------------------------

---

**Description**

This function tries to make the writing circos tiles easier. Like `circos_plus_minus()` and `circos_hist()` it works in 3 stages, It writes out a data file using `cfgout` as a basename and the data from `df` in the circos histogram format into `circos/data/bob_tile.txt` It then writes out a configuration plot stanza in `circos/conf/bob_tile.conf` and finally adds an include to `circos/bob.conf`

**Usage**

```
circos_tile(df, annot_df, cfgout = "circos/conf/default.conf",
            colname = "logFC", chr = "chr1", colors = NULL, outer = 0.9,
            width = 0.08, spacing = 0)
```

**Arguments**

<code>df</code>	Dataframe with starts/ends and the floating point information.
<code>annot_df</code>	Annotation data frame defining starts/stops.
<code>cfgout</code>	Master configuration file to write.
<code>colname</code>	Name of the column with the data of interest.
<code>chr</code>	Name of the chromosome (This currently assumes a bacterial chromosome)
<code>colors</code>	Colors of the data.
<code>outer</code>	Floating point radius of the circle into which to place the categorical data.
<code>width</code>	Width of each tile.
<code>spacing</code>	Radial distance between outer, inner, and inner to whatever follows.

**Value**

Radius after adding the histogram and the spacing.

---

cluster\_trees

*Take clusterprofile group data and print it on a tree as per topGO.*


---

## Description

TopGO's ontology trees can be very illustrative. This function shoe-horns clusterProfiler data into the format expected by topGO and uses it to make those trees.

## Usage

```
cluster_trees(de_genes, cpdata, goid_map = "reference/go/id2go.map",
  goids_df = NULL, score_limit = 0.2, overwrite = FALSE,
  selector = "topDiffGenes", pval_column = "adj.P.Val")
```

## Arguments

de_genes	List of genes deemed 'interesting'.
cpdata	Data from simple_clusterprofiler().
goid_map	Mapping file of IDs to GO ontologies.
goids_df	Dataframe of mappings used to build goid_map.
score_limit	Scoring limit above which to ignore genes.
overwrite	Overwrite an existing goid mapping file?
selector	Name of a function for applying scores to the trees.
pval_column	Name of the column in the GO table from which to extract scores.

## Value

plots! Trees! oh my!

## See Also

**Ramigo** [showSigOfNodes](#)

## Examples

```
## Not run:
cluster_data <- simple_clusterprofiler(genes, stuff)
ctrees <- cluster_trees(genes, cluster_data)

## End(Not run)
```



---

combine_de_tables	<i>Combine portions of deseq/limma/edger table output.</i>
-------------------	------------------------------------------------------------

---

### Description

This hopefully makes it easy to compare the outputs from limma/DESeq2/EdgeR on a table-by-table basis.

### Usage

```
combine_de_tables(all_pairwise_result, extra_annot = NULL, csv = NULL,
  excel = NULL,
  excel_title = "Table SXXX: Combined Differential Expression of YYY",
  excel_sheet = "combined_DE", keepers = "all", include_basic = TRUE,
  add_plots = TRUE, plot_dim = 6, compare_plots = TRUE)
```

### Arguments

all_pairwise_result	Output from all_pairwise().
extra_annot	Add some annotation information?
excel	Filename for the excel workbook, or null if not printed.
excel_title	Title for the excel sheet(s). If it has the string 'YYY', that will be replaced by the contrast name.
excel_sheet	Name the excel sheet.
keepers	List of reformatted table names to explicitly keep certain contrasts in specific orders and orientations.
include_basic	Include my stupid basic logFC tables?
add_plots	Add plots to the end of the sheets with expression values?
plot_dim	Number of inches squared for the plot if added.

### Value

Table combining limma/edger/deseq outputs.

### See Also

[all\\_pairwise](#)

### Examples

```
## Not run:
pretty = combine_de_tables(big_result, table='t12_vs_t0')

## End(Not run)
```

---

compare_go_searches	<i>Compare the results from different ontology tools</i>
---------------------	----------------------------------------------------------

---

**Description**

Combine the results from goseq, cluster profiler, topgo, and gostats; poke at them with a stick and see what happens. The general idea is to pull the p-value data from each tool and contrast that to the set of all possible ontologies. This allows one to do a correlation coefficient between them. In addition, take the 1-pvalue for each ontology for each tool. Thus for strong p-values the score will be near 1 and so we can sum the scores for all the tools. Since topgo has 4 tools, the total possible is 7 if everything has a p-value equal to 0.

**Usage**

```
compare_go_searches(goseq = NULL, cluster = NULL, topgo = NULL,
  gostats = NULL)
```

**Arguments**

goseq	The goseq result from simple_goseq()
cluster	The result from simple_clusterprofiler()
topgo	Guess
gostats	Yep, ditto

**Value**

a summary of the similarities of ontology searches

---

compare_logfc_plots	<i>Compare logFC values from limma and friends</i>
---------------------	----------------------------------------------------

---

**Description**

There are some peculiar discrepancies among these tools, what is up with that?

**Usage**

```
compare_logfc_plots(combined_tables)
```

**Arguments**

combined_tables	The combined tables from limma et al.
-----------------	---------------------------------------

**Value**

Some plots

---

`compare_surrogate_estimates`

*Perform a comparison of the surrogate estimators demonstrated by Jeff Leek.*

---

### Description

This is entirely derivative, but seeks to provide similar estimates for one's own actual data and catch corner cases not taken into account in that document (for example if the estimators don't converge on a surrogate variable). This will attempt each of the surrogate estimators described by Leek: pca, sva supervised, sva unsupervised, ruv supervised, ruv residuals, ruv empirical. Upon completion it will perform the same limma expression analysis and plot the ranked t statistics as well as a correlation plot making use of the extracted estimators against condition/batch/whatever else. Finally, it does the same ranking plot against a linear fitting Leek performed and returns the whole pile of information as a list.

### Usage

```
compare_surrogate_estimates(expt, extra_factors = NULL, do_catplots = FALSE)
```

### Arguments

<code>expt</code>	Experiment containing a design and other information.
<code>extra_factors</code>	Character list of extra factors which may be included in the final plot of the data.
<code>do_catplots</code>	Include the catplots? They don't make a lot of sense yet, so probably no.

### Value

List of the results.

---

<code>compare_tables</code>	<i>See how similar are results from limma/deseq/edger.</i>
-----------------------------	------------------------------------------------------------

---

### Description

limma, DEseq2, and EdgeR all make somewhat different assumptions. and choices about what makes a meaningful set of differentially. expressed genes. This seeks to provide a quick and dirty metric describing the degree to which they (dis)agree.

### Usage

```
compare_tables(limma = NULL, deseq = NULL, edger = NULL, basic = NULL,  
  include_basic = TRUE, annot_df = NULL, ...)
```

Arguments

limma	Data from limma_pairwise().
deseq	Data from deseq2_pairwise().
edger	Data from edger_pairwise().
basic	Data from basic_pairwise().
include_basic	include the basic data?
annot_df	Include annotation data?
...	More options!

Value

Heatmap showing how similar they are along with some correlations between the three players.

See Also

[limma\\_pairwise](#) [edger\\_pairwise](#) [deseq2\\_pairwise](#)

Examples

```
## Not run:
l = limma_pairwise(expt)
d = deseq_pairwise(expt)
e = edger_pairwise(expt)
fun = compare_tables(limma=l, deseq=d, edger=e)

## End(Not run)
```

---

concatenate_runs	<i>Sum the reads/gene for multiple sequencing runs of a single condition/batch.</i>
------------------	-------------------------------------------------------------------------------------

---

Description

On occasion we have multiple technical replicates of a sequencing run. This can use a column in the experimental design to identify those replicates and sum the counts into a single column in the count tables.

Usage

```
concatenate_runs(expt, column = "replicate")
```

Arguments

expt	Experiment class containing the requisite metadata and count tables.
column	Column of the design matrix used to specify which samples are replicates.

**Value**

Expt with the concatenated counts, new design matrix, batches, conditions, etc.

**See Also**

**Biobase**

**Examples**

```
## Not run:
compressed = concatenate_runs(expt)

## End(Not run)
```

---

convert_counts	<i>Perform a cpm/rpkm/whatever transformation of a count table.</i>
----------------	---------------------------------------------------------------------

---

**Description**

I should probably tell it to also handle a simple df/vector/list of gene lengths, but I haven't. `cp_seq_m` is a cpm conversion of the data followed by a rp-ish conversion which normalizes by the number of the given oligo. By default this oligo is 'TA' because it was used for tseq which should be normalized by the number of possible transposition sites by mariner. It could, however, be used to normalize by the number of methionines, for example – if one wanted to do such a thing.

**Usage**

```
convert_counts(data, convert = "raw", ...)
```

**Arguments**

<code>data</code>	Matrix of count data.
<code>convert</code>	Type of conversion to perform: <code>edgecpm/cpm/rpkm/cp_seq_m</code> .
<code>...</code>	Options I might pass from other functions are dropped into arglist, used by <code>rpkm</code> (gene lengths) and <code>divide_seq</code> (genome, pattern to match, and annotation type).

**Value**

Dataframe of cpm/rpkm/whatever(counts)

**See Also**

**edgeR** **Biobase** [cpm](#)

## Examples

```
## Not run:
  converted_table = convert_counts(count_table, convert='cbcbcpm')

## End(Not run)
```

---

`create_combined_table` *Given a limma, edger, and deseq table, combine them into one.*

---

## Description

This combines the outputs from the various differential expression tools and formalizes some column names to make them a little more consistent.

## Usage

```
create_combined_table(li, ed, de, ba, table_name, annot_df = NULL,
  inverse = FALSE, include_basic = TRUE, fc_cutoff = 1, p_cutoff = 0.05)
```

## Arguments

<code>li</code>	Limma output table.
<code>ed</code>	Edger output table.
<code>de</code>	Deseq2 output table.
<code>ba</code>	Basic output table.
<code>table_name</code>	Name of the table to merge.
<code>annot_df</code>	Add some annotation information?
<code>inverse</code>	Invert the fold changes?
<code>include_basic</code>	Include the basic table?
<code>fc_cutoff</code>	Preferred logfoldchange cutoff.
<code>p_cutoff</code>	Preferred pvalue cutoff.

## Value

List containing a) Dataframe containing the merged limma/edger/deseq/basic tables, and b) A summary of how many genes were observed as up/down by output table.

---

create_expt	<i>Wrap bioconductor's expressionset to include some other extraneous information.</i>
-------------	----------------------------------------------------------------------------------------

---

## Description

It is worth noting that this function has a lot of logic used to find the count tables in the local filesystem. This logic has been superceded by simply adding a field to the .csv file called 'file'. create\_expt() will then just read that filename, it may be a full pathname or local to the cwd of the project.

## Usage

```
create_expt(metadata, gene_info = NULL, count_dataframe = NULL,
  sample_colors = NULL, title = NULL, notes = NULL,
  include_type = "all", include_gff = NULL, savefile = "expt",
  low_files = FALSE, ...)
```

## Arguments

metadata	Comma separated file (or excel) describing the samples with information like condition, batch, count_filename, etc.
gene_info	Annotation information describing the rows of the data set, this often comes from a call to import.gff() or biomaRt or organismdbi.
count_dataframe	If one does not wish to read the count tables from the filesystem, they may instead be fed as a data frame here.
sample_colors	List of colors by condition, if not provided it will generate its own colors using colorBrewer.
title	Provide a title for the expt?
notes	Additional notes?
include_type	I have usually assumed that all gff annotations should be used, but that is not always true, this allows one to limit to a specific annotation type.
include_gff	Gff file to help in sorting which features to keep.
savefile	Rdata filename prefix for saving the data of the resulting expt.
low_files	Explicitly lowercase the filenames when searching the filesystem?
...	More parameters are fun!

## Value

experiment an expressionset

## See Also

**Biobase** [pData](#) [fData](#) [exprs](#) [hpgl\\_read\\_files](#) [as.list.hash](#)

## Examples

```
## Not run:
new_experiment = create_expt("some_csv_file.csv", color_hash)
## Remember that this depends on an existing data structure of gene annotations.

## End(Not run)
```

---

default_norm	<i>Perform a default normalization of some data</i>
--------------	-----------------------------------------------------

---

## Description

This just calls `normalize_expt` with the most common arguments except `log2` transformation, but that may be appended with `'transform=log2'`, so I don't feel bad. Indeed, it will allow you to overwrite any arguments if you wish. In our work, the most common normalization is: `quantile(cpm(low-filter(data)))`.

## Usage

```
default_norm(expt, ...)
```

---

deparse_go_value	<i>Extract more easily readable information from a GOTERM datum.</i>
------------------	----------------------------------------------------------------------

---

## Description

The output from the GOTERM/GO.db functions is inconsistent, to put it nicely. This attempts to extract from that heterogeneous datatype something easily readable. Example: `Synonym()` might return any of the following: NA, NULL, "NA", "NULL", `c("NA",NA,"GO:00001")`, "GO:00002", `c("Some text",NA,NULL,"GO:00003")` This function will boil that down to 'not found', "", 'GO:00004', or "GO:0001, some text, GO:00004"

## Usage

```
deparse_go_value(value)
```

## Arguments

value	Result of <code>try(as.character(somefunction(GOTERM[id])), silent=TRUE)</code> . some-function would be 'Synonym' 'Secondary' 'Ontology', etc...
-------	---------------------------------------------------------------------------------------------------------------------------------------------------

## Value

something more sane (hopefully).



## Examples

```
## Not run:
## goterms = GOTERM[ids]
## sane_goterms = deparse_go_value(goterms)

## End(Not run)
```

---

deseq2_pairwise	<i>Set up model matrices contrasts and do pairwise comparisons of all conditions using DESeq2.</i>
-----------------	----------------------------------------------------------------------------------------------------

---

## Description

Invoking DESeq2 is confusing, this should help.

## Usage

```
deseq2_pairwise(input, conditions = NULL, batches = NULL,
  model_cond = TRUE, alt_model = NULL, extra_contrasts = NULL,
  model_intercept = FALSE, model_batch = TRUE, annot_df = NULL,
  force = FALSE, ...)
```

## Arguments

input	Dataframe/vector or expt class containing data, normalization state, etc.
conditions	Factor of conditions in the experiment.
batches	Factor of batches in the experiment.
model_cond	Is condition in the experimental model?
alt_model	Provide an arbitrary model here.
extra_contrasts	Provide extra contrasts here.
model_batch	Is batch in the experimental model?
annot_df	Include some annotation information in the results?
force	Force deseq to accept data which likely violates its assumptions.
...	triple dots! Options are passed to arglist.

## Value

List including the following information: run = the return from calling DESeq() denominators = list of denominators in the contrasts numerators = list of the numerators in the contrasts conditions = the list of conditions in the experiment coefficients = list of coefficients making the contrasts all\_tables = list of DE tables

## See Also

**DESeq2** [results](#) [estimateSizeFactors](#) [estimateDispersions](#) [nbinomWaldTest](#)

**Examples**

```
## Not run:
pretend = deseq2_pairwise(data, conditions, batches)

## End(Not run)
```

---

deseq\_coefficient\_scatter

*Plot out 2 coefficients with respect to one another from deseq2.*

---

**Description**

It can be nice to see a plot of two coefficients from a deseq2 comparison with respect to one another. This hopefully makes that easy.

**Usage**

```
deseq_coefficient_scatter(output, toptable = NULL, x = 1, y = 2,
  gvis_filename = NULL, gvis_trendline = TRUE, z = 1.5,
  tooltip_data = NULL, base_url = NULL, color_low = "#DD0000",
  color_high = "#7B9F35", ...)
```

**Arguments**

output	Set of pairwise comparisons provided by deseq_pairwise().
x	Name or number of the x-axis coefficient column to extract.
y	Name or number of the y-axis coefficient column to extract.
gvis_filename	Filename for plotting gvis interactive graphs of the data.
gvis_trendline	Add a trendline to the gvis plot?
tooltip_data	Dataframe of gene annotations to be used in the gvis plot.
base_url	When plotting interactive plots, have link-outs to this base url.

**Value**

Ggplot2 plot showing the relationship between the two coefficients.

**See Also**

[plot\\_linear\\_scatter deseq2\\_pairwise](#)

**Examples**

```
## Not run:
pretty = coefficient_scatter(deseq_data, x="wt", y="mut")

## End(Not run)
```

---

deseq_pairwise	<i>deseq_pairwise()</i> Because I can't be trusted to remember '2'.
----------------	---------------------------------------------------------------------

---

### Description

This calls `deseq2_pairwise(...)` because I am determined to forget typing `deseq2`.

### Usage

```
deseq_pairwise(...)
```

### Arguments

... I like cats.

### Value

stuff `deseq2_pairwise` results.

### See Also

[deseq2\\_pairwise](#)

---

divide_seq	<i>Express a data frame of counts as reads per pattern per million.</i>
------------	-------------------------------------------------------------------------

---

### Description

This uses a sequence pattern rather than length to normalize sequence. It is essentially fancy pants `rpk`.

### Usage

```
divide_seq(counts, genome = NULL, ...)
```

### Arguments

<code>counts</code>	Read count matrix.
<code>genome</code>	Genome to search (fasta/BSgenome).
...	Options I might pass from other functions are dropped into arglist.

### Value

The `RPseqM` counts

**See Also**[FaFile rpkm](#)**Examples**

```
## Not run:
cptam <- divide_seq(cont_table, fasta="mgas_5005.fasta.xz", gff="mgas_5005.gff.xz")

## End(Not run)
```

---

edger\_coefficient\_scatter

*Plot two coefficients with respect to one another from edgeR.*


---

**Description**

It can be nice to see a plot of two coefficients from a edger comparison with respect to one another. This hopefully makes that easy.

**Usage**

```
edger_coefficient_scatter(output, toptable = NULL, x = 1, y = 2,
  gvis_filename = NULL, gvis_trendline = TRUE, z = 1.5,
  tooltip_data = NULL, base_url = NULL, color_low = "#DD0000",
  color_high = "#7B9F35", ...)
```

**Arguments**

output	Set of pairwise comparisons provided by <code>edger_pairwise()</code> .
x	Name or number of the x-axis coefficient column to extract.
y	Name or number of the y-axis coefficient column to extract.
gvis_filename	Filename for plotting gvis interactive graphs of the data.
gvis_trendline	Add a trendline to the gvis plot?
tooltip_data	Dataframe of gene annotations to be used in the gvis plot.
base_url	Add a linkout to gvis plots to this base url.

**Value**

Ggplot2 plot showing the relationship between the two coefficients.

**See Also**[plot\\_linear\\_scatter](#) [edger\\_pairwise](#)

**Examples**

```
## Not run:
pretty = coefficient_scatter(limma_data, x="wt", y="mut")

## End(Not run)
```

---

edger_pairwise	<i>Set up a model matrix and set of contrasts to do pairwise comparisons using EdgeR.</i>
----------------	-------------------------------------------------------------------------------------------

---

**Description**

This function performs the set of possible pairwise comparisons using EdgeR.

**Usage**

```
edger_pairwise(input, conditions = NULL, batches = NULL,
               model_cond = TRUE, model_batch = TRUE, model_intercept = TRUE,
               alt_model = NULL, extra_contrasts = NULL, annot_df = NULL,
               force = FALSE, edger_method = "default", ...)
```

**Arguments**

input	Dataframe/vector or expt class containing data, normalization state, etc.
conditions	Factor of conditions in the experiment.
batches	Factor of batches in the experiment.
model_cond	Include condition in the experimental model?
model_batch	Include batch in the model? In most cases this is a good thing(tm).
model_intercept	Use cell means or intercept?
alt_model	Alternate experimental model to use?
extra_contrasts	Add some extra contrasts to add to the list of pairwise contrasts. This can be pretty neat, lets say one has conditions A,B,C,D,E and wants to do (C/B)/A and (E/D)/A or (E/D)/(C/B) then use this with a string like: "c_vs_b_ctrla = (C-B)-A, e_vs_d_ctrla = (E-D)-A, de_vs_cb = (E-D)-(C-B),"
annot_df	Annotation information to the data tables?
force	Force edgeR to accept inputs which it should not have to deal with.
...	The elipsis parameter is fed to write_edger() at the end.

**Value**

List including the following information: contrasts = The string representation of the contrasts performed. lrt = A list of the results from calling glmLRT(), one for each contrast. contrast\_list = The list of each call to makeContrasts() I do this to avoid running into the limit on # of contrasts addressable by topTags() all\_tables = a list of tables for the contrasts performed.

**See Also**

[edgeR](#) [topTags](#) [glmLRT](#) [make\\_pairwise\\_contrasts](#) [DGEList](#) [calcNormFactors](#) [estimateTagwiseDisp](#)  
[estimateCommonDisp](#) [estimateGLMCommonDisp](#) [estimateGLMTrendedDisp](#) [glmFit](#)

**Examples**

```
## Not run:
pretend = edger_pairwise(data, conditions, batches)

## End(Not run)
```

---

expt_subset	<i>Extract a subset of samples following some rule(s) from an experiment class.</i>
-------------	-------------------------------------------------------------------------------------

---

**Description**

Sometimes an experiment has too many parts to work with conveniently, this operation allows one to break it into smaller pieces.

**Usage**

```
expt_subset(expt, subset = NULL)
```

**Arguments**

expt	Expt chosen to extract a subset of data.
subset	Valid R expression which defines a subset of the design to keep.

**Value**

metadata Expt class which contains the smaller set of data.

**See Also**

**Biobase** [pData](#) [exprs](#) [fData](#)

**Examples**

```
## Not run:
smaller_expt = expt_subset(big_expt, "condition=='control'")
all_expt = expt_subset(expressionset, "") ## extracts everything

## End(Not run)
```

---

extract_go	<i>Extract a set of geneID to GOID mappings from a suitable data source.</i>
------------	------------------------------------------------------------------------------

---

### Description

Like extract\_lengths above, this is primarily intended to read gene ID and GO ID mappings from a OrgDb/OrganismDbi object.

### Usage

```
extract_go(db, metadf = NULL, keytype = "ENTREZID")
```

### Arguments

db	Data source containing mapping information.
metadf	Data frame containing extant information.
keytype	Keytype used for querying

### Value

Dataframe of 2 columns: geneID and goID.

---

extract_lengths	<i>Take gene/exon lengths from a suitable data source (gff/TxDb/OrganismDbi)</i>
-----------------	----------------------------------------------------------------------------------

---

### Description

Primarily goseq, but also other tools on occasion require a set of gene IDs and lengths. This function is responsible for pulling that data from either a gff, or TxDb/OrganismDbi.

### Usage

```
extract_lengths(db = NULL, gene_list = NULL,
  type = "GenomicFeatures::transcripts", id = "TXID",
  possible_types = c("GenomicFeatures::genes", "GenomicFeatures::cds",
    "GenomicFeatures::transcripts"), possible_ids = c("GENEID", "CDSID",
    "TXID"))
```

### Arguments

db	Object containing data, if it is a string then a filename is assumed to a gff file.
gene_list	Set of genes to query.
type	Function name used for extracting data from TxDb objects.
id	Column from the resulting data structure to extract gene IDs.
possible_types	Character list of types I have previously used.
possible_ids	Corresponding IDs for the above types.

**Value**

Dataframe containing 2 columns: ID, length

---

extract\_significant\_genes

*Extract the sets of genes which are significantly up/down regulated from the combined tables.*

---

**Description**

Given the output from `combine_de_tables()`, extract the genes in which we have the greatest likely interest, either because they have the largest fold changes, lowest p-values, fall outside a z-score, or are at the top/bottom of the ranked list.

**Usage**

```
extract_significant_genes(combined, according_to = "all", fc = 1,
  p = 0.05, z = NULL, n = NULL, p_type = "adj",
  excel = "excel/significant_genes.xlsx", csv = NULL)
```

**Arguments**

<code>combined</code>	Output from <code>combine_de_tables()</code> .
<code>according_to</code>	What tool(s) decide 'significant?' One may use the <code>deseq</code> , <code>edger</code> , <code>limma</code> , <code>basic</code> , <code>meta</code> , or <code>all</code> .
<code>fc</code>	Log fold change to define 'significant'.
<code>p</code>	(Adjusted)p-value to define 'significant'.
<code>z</code>	Z-score to define 'significant'.
<code>n</code>	Take the top/bottom-n genes.
<code>excel</code>	Write the results to this excel file, or <code>NULL</code> .

**Value**

The set of up-genes, down-genes, and numbers therein.

**See Also**

[combine\\_de\\_tables](#)



---

factor_rsquared	<i>Collect the <math>r^2</math> values from a linear model fitting between a singular value decomposition and factor.</i>
-----------------	---------------------------------------------------------------------------------------------------------------------------

---

**Description**

Collect the  $r^2$  values from a linear model fitting between a singular value decomposition and factor.

**Usage**

```
factor_rsquared(svd_v, fact, type = "factor")
```

**Arguments**

svd_v	$V^T V = I$ portion of a fast.svd call.
fact	Experimental factor from the original data.
type	Make this categorical or continuous with factor/continuous.

**Value**

The  $r^2$  values of the linear model as a percentage.

**See Also**

[fast.svd](#)

---

filter_counts	<i>Call various count filters.</i>
---------------	------------------------------------

---

**Description**

This calls the various filtering functions in genefilter along with suggestions made in our lab meetings; defaulting to the threshold based filter suggested by Hector.

**Usage**

```
filter_counts(count_table, filter = "cbcb", p = 0.01, A = 1, k = 1,  
  cv_min = 0.01, cv_max = 1000, thresh = 4, min_samples = 2, ...)
```

**Arguments**

count_table	Some counts to filter.
filter	Filtering method to apply (cbcb, pofa, kofa, cv right now).
p	Used by genefilter's pofa().
A	Also for pofa().
k	Used by genefilter's kofa().
cv_min	Used by genefilter's cv().
cv_max	Also used by cv().
thresh	Minimum threshold across samples for cbcb.
min_samples	Minimum number of samples for cbcb.
...	More options might be needed, especially if I fold cv/p/etc into ...

**Value**

Data frame of filtered counts.

**See Also**

**genefilter**

**Examples**

```
## Not run:
new <- filter_counts(old)

## End(Not run)
```

---

gather_goseq_genes	<i>Given a set of goseq data from simple_goseq(), make a list of genes represented in each ontology.</i>
--------------------	----------------------------------------------------------------------------------------------------------

---

**Description**

This function uses the GO2ALLEG data structure to reverse map ontology categories to a list of genes represented. It therefore assumes that the GO2ALLEG.rda data structure has been deposited in pwd(). This in turn may be generated by clusterProfilers buildGOMap() function if it doesn't exist. For some species it may also be auto-generated. With little work this can be made much more generic, and it probably should.

**Usage**

```
gather_goseq_genes(goseq_data, ontology = NULL, pval = 0.1,
  include_all = FALSE, ...)
```

**Arguments**

<code>goseq_data</code>	List of goseq specific results as generated by <code>simple_goseq()</code> .
<code>ontology</code>	Ontology to search (MF/BP/CC).
<code>pval</code>	Maximum accepted pvalue to include in the list of categories to cross reference.
<code>include_all</code>	Include all genes in the ontology search?
<code>...</code>	Extra options without a purpose just yet.

**Value**

Data frame of categories/genes.

**See Also**

[simple\\_goseq](#) [buildGOMap](#),

**Examples**

```
## Not run:
data = simple_goseq(de_genes=limma_output, lengths=annotation_df, goids=goids_df)
genes_in_cats = gather_genes(data, ont='BP')

## End(Not run)
```

---

<code>gbk2txdb</code>	<i>Given a genbank accession, make a txDb object along with sequences, etc.</i>
-----------------------	---------------------------------------------------------------------------------

---

**Description**

Let us admit it, sometimes biomart is a pain. It also does not have easily accessible data for microbes. Genbank does!

**Usage**

```
gbk2txdb(accession = "AE009949")
```

**Arguments**

<code>accession</code>	Accession to download and import
------------------------	----------------------------------

**Value**

List containing a txDb, sequences, and some other stuff which I haven't yet finalized.

---

gbk_annotations	<i>Extract some useful information from a gbk imported as a txDb.</i>
-----------------	-----------------------------------------------------------------------

---

**Description**

Maybe this should get pulled into the previous function?

**Usage**

```
gbk_annotations(gbr)
```

**Arguments**

gbr	TxDb object to poke at.
-----	-------------------------

**Value**

Granges data

---

genefilter_cv_counts	<i>Filter genes from a dataset outside a range of variance.</i>
----------------------	-----------------------------------------------------------------

---

**Description**

This function from genefilter removes genes surpassing a variance cutoff. It is not therefore a low-count filter per se.

**Usage**

```
genefilter_cv_counts(count_table, cv_min = 0.01, cv_max = 1000)
```

**Arguments**

count_table	Input data frame of counts by sample.
cv_min	Minimum coefficient of variance.
cv_max	Maximum coefficient of variance.

**Value**

Dataframe of counts without the high/low variance genes.

**See Also**

**genefilter** [kOverA](#) which this uses to decide what to keep.

## Examples

```
## Not run:  
filtered_table = genefilter_kofa_counts(count_table)  
  
## End(Not run)
```

---

genefilter\_kofa\_counts

*Filter low-count genes from a data set using genefilter's kOverA().*

---

## Description

This is the most similar to the function suggested by Hector I think.

## Usage

```
genefilter_kofa_counts(count_table, k = 1, A = 1)
```

## Arguments

count_table	Input data frame of counts by sample.
k	Minimum number of samples to have >A counts.
A	Minimum number of counts for each gene's sample in kOverA().

## Value

Dataframe of counts without the low-count genes.

## See Also

**genefilter** [kOverA](#) which this uses to decide what to keep.

## Examples

```
## Not run:  
filtered_table = genefilter_kofa_counts(count_table)  
  
## End(Not run)
```

---

`genefilter_pofa_counts`*Filter low-count genes from a data set using genefilter's pOverA().*

---

**Description**

I keep thinking this function is pofa... oh well. Of the various tools in genefilter, this one to me is the most intuitive. Take the ratio of counts/samples and make sure it is  $\geq$  a score.

**Usage**

```
genefilter_pofa_counts(count_table, p = 0.01, A = 100)
```

**Arguments**

<code>count_table</code>	Input data frame of counts by sample.
<code>p</code>	Minimum proportion of each gene's counts/sample to be greater than a minimum(A).
<code>A</code>	Minimum number of counts in the above proportion.

**Value**

Dataframe of counts without the low-count genes.

**See Also**

**genefilter** [pOverA](#) which this uses to decide what to keep.

**Examples**

```
## Not run:  
  filtered_table = genefilter_pofa_counts(count_table)  
  
## End(Not run)
```

---

`generate_gene_kegg_mapping`*Generate GENE/KEGG mapping.*

---

**Description**

This uses KEGGREST and related function `kegg_to_ensembl()` to associate genes to KEGG pathways.

**Usage**

```
generate_gene_kegg_mapping(pathways, org_abbreviation, verbose = FALSE)
```

**Arguments**

pathways	Vector of KEGG pathway IDs returned from call to keggLink() e.g. "path:mmu05134".
org_abbreviation	KEGG identifier for the species of interest (e.g. "hsa" for Homo sapiens).
verbose	talky talky?

**Value**

Df mapping kegg and gene IDs.

**See Also**

[keggLink](#)

**Examples**

```
## Not run:  
kegg_df <- generate_gene_kegg_mapping(path, org)  
  
## End(Not run)
```

---

```
generate_kegg_pathway_mapping
```

*Generate a KEGG PATHWAY / description mapping.*

---

**Description**

Make an easier to use df of KEGG -> descriptions using keggGet.

**Usage**

```
generate_kegg_pathway_mapping(pathways, verbose = FALSE)
```

**Arguments**

pathways	Vector of KEGG pathway identifiers.
verbose	talk talk?

**Value**

Data frame describing some kegg pathways

**See Also**[keggGet](#)**Examples**

```
mapping <- generate_kegg_pathway_mapping(c("hsa00040", "hsa00100"))
```

---

getEdgeWeights	<i>Plot the ontology DAG.</i>
----------------	-------------------------------

---

**Description**

This function was stolen from topgo in order to figure out where it was failing.

**Usage**

```
getEdgeWeights(graph)
```

**Arguments**

graph	Graph from topGO
-------	------------------

**Value**

Weights!

---

get_biomart_annotations	<i>Extract annotation information from biomart.</i>
-------------------------	-----------------------------------------------------

---

**Description**

Biomart is an amazing resource of information, but using it is a bit annoying. This function hopes to alleviate some common headaches.

**Usage**

```
get_biomart_annotations(species = "hsapiens", overwrite = FALSE,
  do_save = TRUE, host = "dec2015.archive.ensembl.org",
  trymart = "ENSEMBL_MART_ENSEMBL", include_lengths = TRUE)
```



**Arguments**

species	Choose a species.
overwrite	Overwrite an existing save file?
do_save	Create a savefile of annotations for future runs?
host	Ensembl hostname to use.
trymart	Biomart has become a circular dependency, this makes me sad, now to list the marts, you need to have a mart loaded...
include_lengths	Also perform a search on structural elements in the genome?

**Value**

Df of some (by default) human annotations.

**Examples**

```
## Not run:
tt = get_biomart_annotatons()

## End(Not run)
```

---

```
get_biomart_ontologies
```

*Extract gene ontology information from biomart.*

---

**Description**

I perceive that every time I go to acquire annotation data from biomart, they have changed something important and made it more difficult for me to find what I want. I recently found the \*.archive.ensembl.org, and so this function uses that to try to keep things predictable, if not consistent.

**Usage**

```
get_biomart_ontologies(species = "hsapiens", overwrite = FALSE,
  do_save = TRUE, host = "dec2015.archive.ensembl.org",
  trymart = "ENSEMBL_MART_ENSEMBL", secondtry = "_gene")
```

**Arguments**

species	Species to query.
overwrite	Overwrite existing savefile?
do_save	Create a savefile of the annotations? (if not false, then a filename.)
host	Ensembl hostname to use.
trymart	Default mart to try, newer marts use a different notation.
secondtry	The newer mart name.

**Value**

Df of geneIDs and GOIDs.

**See Also**

[getBM](#)

**Examples**

```
## Not run:  
tt = get_biomart_ontologies()  
  
## End(Not run)
```

---

get_eupath_config	<i>Grab some configuration data collated and used to make OrganismDbi/OrgDb/TxDb objects.</i>
-------------------	-----------------------------------------------------------------------------------------------

---

**Description**

This function uses some data copied into inst/ to decide some parameters used for generating the various packages generated here.

**Usage**

```
get_eupath_config(cfg = NULL)
```

**Arguments**

cfg	Optional data frame
-----	---------------------

**Value**

Dataframe of configuration data, a few columns are required, run it with no args to see which ones.

---

get_genelengths	<i>Grab gene lengths from a gff file.</i>
-----------------	-------------------------------------------

---

## Description

This function attempts to be robust to the differences in output from importing gff2/gff3 files. But it certainly isn't perfect.

## Usage

```
get_genelengths(gff, type = "gene", key = "ID", ...)
```

## Arguments

gff	Gff file with (hopefully) IDs and widths.
type	Annotation type to use (3rd column).
key	Identifier in the 10th column of the gff file to use.
...	Extra arguments likely for gff2df

## Value

Data frame of gene IDs and widths.

## See Also

**rtracklayer** [import.gff](#)

## Examples

```
## Not run:
tt = get_genelengths('reference/fun.gff.gz')
head(tt)
##           ID width
## 1  YAL069W   312
## 2  YAL069W   315
## 3  YAL069W     3
## 4 YAL068W-A   252
## 5 YAL068W-A   255
## 6 YAL068W-A     3

## End(Not run)
```

---

get_kegg_genes	<i>Extract the set of geneIDs matching pathways for a given species.</i>
----------------	--------------------------------------------------------------------------

---

### Description

This uses KEGGREST to extract the mappings for all genes for a species and pathway or 'all'. Because downloading them takes a while, it will save the results to kegg\_species.rda. When run interactively, it will give some information regarding the number of genes observed in each pathway.

### Usage

```
get_kegg_genes(pathway = "all", abbreviation = NULL,
               species = "leishmania major", savefile = NULL)
```

### Arguments

pathway	Either a single pathway kegg id or 'all'.
abbreviation	Optional 3 letter species kegg id.
species	Stringified species name used to extract the 3 letter abbreviation.
savefile	Filename to which to save the relevant data.

### Value

Dataframe of the various kegg data for each pathway, 1 row/gene.

### Examples

```
## Not run:
kegg_info <- get_kegg_genes(species="Canis familiaris")

## End(Not run)
```

---

get_kegg_sub	<i>Provide a set of simple substitutions to convert geneIDs from KEGG-&gt;TriTryDB</i>
--------------	----------------------------------------------------------------------------------------

---

### Description

This function should provide 2 character lists which, when applied sequentially, will result in a hopefully coherent set of mapped gene IDs matching the TriTryDB/KEGG specifications.

### Usage

```
get_kegg_sub(species = "lma")
```

**Arguments**

species                      3 letter abbreviation for a given kegg type

**Value**

2 character lists containing the patterns and replace arguments for gsub(), order matters!

---

get\_microbesonline\_annotation

*Skip the db and download all the text annotations for a given species.*

---

**Description**

Like I said, the microbesonline mysqldb is rather more complex than I prefer. This shortcuts that process and just grabs a tsv copy of everything and loads it into a dataframe.

**Usage**

```
get_microbesonline_annotation(ids = "160490", species = NULL)
```

**Arguments**

ids                          List of ids to query.  
species                      Species name(s) to use instead.

**Value**

List of dataframes with the annotation information.

---

get\_microbesonline\_ids

*Use the publicly available microbesonline mysql instance to get species ids.*

---

**Description**

The microbesonline mysql instance is more complex than I like. Their id system is reminiscent of KEGG's and similarly annoying. Though I haven't figured out how the tables interact, a query to get ids is simple enough.

**Usage**

```
get_microbesonline_ids(name, exact = FALSE)
```

**Arguments**

name	Text string containing some part of the species name of interest.
exact	Use an exact species name?

**Value**

Dataframe of ids and names.

---

```
get_microbesonline_name
```

*Use the publicly available microbesonline mysql instance to get species name(s).*

---

**Description**

The microbesonline mysql instance is more complex than I like. Their id system is reminiscent of KEGG's and similarly annoying. Though I haven't figured out how the tables interact, a query to get ids is simple enough.

**Usage**

```
get_microbesonline_name(id)
```

**Arguments**

id	Text string containing some part of the species name of interest.
----	-------------------------------------------------------------------

**Value**

Dataframe of ids and names.

---

```
get_model_adjust
```

*Extract some surrogate estimations from a raw data set using sva, ruv, and/or pca.*

---

**Description**

This applies the methodologies very nicely explained by Jeff Leek at <https://github.com/jtleek/svaseq/blob/master/recount.Rmd> and attempts to use them to acquire estimates which may be applied to an experimental model by either EdgeR, DESeq2, or limma. In addition, it modifies the count tables using these estimates so that one may play with the modified counts and view the changes (with PCA or heatmaps or whatever). Finally, it prints a couple of the plots shown by Leek in his document. In other words, this is entirely derivative of someone much smarter than me.

**Usage**

```
get_model_adjust(expt, estimate_type = "sva_supervised", surrogates = "be",
...)
```

**Arguments**

expt	Raw experiment object
estimate_type	One of sva_supervised, sva_unsupervised, ruv_empirical, ruv_supervised, ruv_residuals, or pca.
surrogates	Choose a method for getting the number of surrogates, be or leek.
...	Parameters fed to arglist.

**Value**

List including the adjustments for a model matrix, a modified count table, and 3 plots of the known batch, surrogates, and batch/surrogate.

---

get_ncbi_taxonid	<i>Use taxize to get ncbi taxon IDs</i>
------------------	-----------------------------------------

---

**Description**

taxize looks like it might be awesome, but it is also pretty annoying

**Usage**

```
get_ncbi_taxonid(species = "Leishmania major")
```

**Arguments**

species	Human readable species name
---------	-----------------------------

**Value**

potential NCBI taxon IDs

---

get_sig_genes	<i>Get a set of up/down differentially expressed genes.</i>
---------------	-------------------------------------------------------------

---

### Description

Take one or more criteria (fold change, rank order, (adj)p-value, z-score from median FC) and use them to extract the set of genes which are defined as 'differentially expressed.' If no criteria are provided, it arbitrarily chooses all genes outside of 1-z.

### Usage

```
get_sig_genes(table, n = NULL, z = NULL, fc = NULL, p = NULL,
              column = "logFC", fold = "plusminus", p_column = "adj.P.Val")
```

### Arguments

table	Table from limma/edger/deseq.
n	Rank-order top/bottom number of genes to take.
z	Number of z-scores >/< the median to take.
fc	Fold-change cutoff.
p	P-value cutoff.
column	Table's column used to distinguish top vs. bottom.
fold	Identifier reminding how to get the bottom portion of a fold-change (plusminus says to get the negative of the positive, otherwise 1/positive is taken). This effectively tells me if this is a log fold change or not.
p_column	Table's column containing (adjusted or not)p-values.

### Value

Subset of the up/down genes given the provided criteria.

---

gff2df	<i>Extract annotation information from a gff file into a df</i>
--------	-----------------------------------------------------------------

---

### Description

Try to make import.gff a little more robust; I acquire (hopefully) valid gff files from various sources: yeastgenome.org, microbesonline, tritrypdb, ucsc, ncbi. To my eyes, they all look like reasonably good gff3 files, but some of them must be loaded with import.gff2, import.gff3, etc. That is super annoying. Also, I pretty much always just do as.data.frame() when I get something valid from rtracklayer, so this does that for me, I have another function which returns the iranges etc. This function wraps import.gff/import.gff3/import.gff2 calls in try() because sometimes those functions fail in unpredictable ways.



**Usage**

```
gff2df(gff, type = NULL, id_col = "ID", second_id_col = "locus_tag",
      try = NULL)
```

**Arguments**

<code>gff</code>	Gff filename.
<code>type</code>	Subset the gff file for entries of a specific type.
<code>id_col</code>	Column in a successful import containing the IDs of interest.
<code>second_id_col</code>	Second column to check.
<code>try</code>	Give your own function call to use for importing.

**Value**

Dataframe of the annotation information found in the gff file.

**See Also**

**rtracklayer** [import.gff](#) [import.gff2](#) [import.gff3](#)

**Examples**

```
## Not run:
funkytown <- gff2df('reference/gff/saccharomyces_cerevsiae.gff.xz')

## End(Not run)
```

---

gff2irange

---

*Extract annotation information from a gff file into an irange object.*


---

**Description**

Try to make `import.gff` a little more robust; I acquire (hopefully) valid gff files from various sources: yeastgenome.org, microbesonline, tritrypdb, ucsc, ncbi. To my eyes, they all look like reasonably good gff3 files, but some of them must be loaded with `import.gff2`, `import.gff3`, etc. That is super annoying. Also, I pretty much always just do `as.data.frame()` when I get something valid from `rtracklayer`, so this does that for me, I have another function which returns the `iranges` etc. This function wraps `import.gff/import.gff3/import.gff2` calls in `try()` because sometimes those functions fail in unpredictable ways.

**Usage**

```
gff2irange(gff, type = NULL)
```

**Arguments**

<code>gff</code>	Gff filename.
<code>type</code>	Subset to extract.

**Details**

This is essentially `gff2df()`, but returns data suitable for `getSet()`

**Value**

Iranges! (useful for `getSeq()`.)

**See Also**

**rtracklayer** [gff2df](#) [getSeq](#)

**Examples**

```
## Not run:
library(BSgenome.Tcruzi.clbrener.all)
tc_clb_all <- BSgenome.Tcruzi.clbrener.all
cds_ranges <- gff2irange('reference/gff/tcruzi_clbrener.gff.xz', type='CDS')
cds_sequences <- Biostrings::getSeq(tc_clb_all, cds_ranges)

## End(Not run)
```

---

**godef**

*Get a go long-form definition from an id.*

---

**Description**

Sometimes it is nice to be able to read the full definition of some GO terms.

**Usage**

```
godef(go = "GO:0032432")
```

**Arguments**

<code>go</code>	GO ID, this may be a character or list (assuming the elements are goids).
-----------------	---------------------------------------------------------------------------

**Value**

Some text providing the long definition of each provided GO id.

**See Also**

**GOTermsAnnDbBimap**

### Examples

```
## Not run:
  godef("GO:0032432")
## > GO:0032432
## > "An assembly of actin filaments that are on the same axis but may be oriented with the
## > same or opposite polarities and may be packed with different levels of tightness."

## End(Not run)
```

---

golev

*Get a go level approximation from an ID.*

---

### Description

Sometimes it is useful to know how far up/down the ontology tree a given id resides. This attmepts to answer that question.

### Usage

```
golev(go)
```

### Arguments

go                      GO id, this may be a character or list (assuming the elements are goids).

### Value

Set of numbers corresponding to approximate tree positions of the GO ids.

### See Also

**GOTermsAnnDbBimap**

### Examples

```
## Not run:
  golev("GO:0032559")
## > 3

## End(Not run)
```

---

golevel	<i>Get a go level approximation from a set of IDs.</i>
---------	--------------------------------------------------------

---

**Description**

This just wraps golev() in mapply.

**Usage**

```
golevel(go = c("GO:0032559", "GO:0000001"))
```

**Arguments**

go                      Character list of IDs.

**Value**

Set pf approximate levels within the onlogy.

**See Also**

**GOTermsAnnDbBimap**

**Examples**

```
## Not run:
golevel(c("GO:0032559", "GO:0000001"))
## > 3 4

## End(Not run)
```

---

golevel_df	<i>Extract a dataframe of golevels using getGOLevel() from clusterProfiler.</i>
------------	---------------------------------------------------------------------------------

---

**Description**

This function is way faster than my previous iterative golevel function. That is not to say it is very fast, so it saves the result to ontlevel.rda for future lookups.

**Usage**

```
golevel_df(ont = "MF", savefile = "ontlevel.rda")
```

**Arguments**

ont                    the ontology to recurse.  
 savefile            a file to save the results for future lookups.

**Value**

golevels a dataframe of goids<->highest level

---

goont	<i>Get a go ontology name from an ID.</i>
-------	-------------------------------------------

---

**Description**

Get a go ontology name from an ID.

**Usage**

```
goont(go = c("GO:0032432", "GO:0032433"))
```

**Arguments**

go                    GO id, this may be a character or list (assuming the elements are goids).

**Value**

The set of ontology IDs associated with the GO ids, thus 'MF' or 'BP' or 'CC'.

**See Also**

**GOTermsAnnDbBimap**

**Examples**

```
## Not run:
goont(c("GO:0032432", "GO:0032433"))
## > GO:0032432 GO:0032433
## > "CC" "CC"

## End(Not run)
```

---

<code>gosec</code>	<i>Get a GO secondary ID from an id.</i>
--------------------	------------------------------------------

---

**Description**

Unfortunately, GOTERM's returns for secondary IDs are not consistent, so this function has to have a whole bunch of logic to handle the various outputs.

**Usage**

```
gosec(go = "GO:0032432")
```

**Arguments**

<code>go</code>	GO ID, this may be a character or list(assuming the elements, not names, are goids).
-----------------	--------------------------------------------------------------------------------------

**Value**

Some text comprising the secondary GO id(s).

**See Also**

**GOTermsAnnDbBimap**

**Examples**

```
## Not run:
gosec("GO:0032432")
## > GO:0032432
## > "GO:0000141" "GO:0030482"

## End(Not run)
```

---

<code>goseq_table</code>	<i>Enhance the goseq table of gene ontology information.</i>
--------------------------	--------------------------------------------------------------

---

**Description**

While goseq has some nice functionality, the table of outputs it provides is somewhat lacking. This attempts to increase that with some extra helpful data like ontology categories, definitions, etc.

**Usage**

```
goseq_table(df, file = NULL)
```

**Arguments**

df	Dataframe of ontology information. This is intended to be the output from goseq including information like numbers/category, GOids, etc. It requires a column 'category' which contains: GO:000001 and such.
file	Csv file to which to write the table.

**Value**

Ontology table with annotation information included.

**See Also**

**goseq**

**Examples**

```
## Not run:
annotated_go = goseq_table(go_ids)
head(annotated_go, n=1)
## >      category numDEInCat numInCat over_represented_pvalue
## > 571 GO:0006364          9      26      4.655108e-08
## >      under_represented_pvalue      qvalue ontology
## > 571      1.0000000 6.731286e-05      BP
## >      term
## > 571      rRNA processing
## >      synonym
## > 571      "35S primary transcript processing, GO:0006365"
## >      secondary definition
## > 571 GO:0006365 Any process involved in the conversion of a primary ribosomal
##      RNA (rRNA) transcript into one or more mature rRNA molecules.

## End(Not run)
```

---

goseq\_trees

---

*Make fun trees a la topgo from goseq data.*


---

**Description**

This seeks to force goseq data into a format suitable for topGO and then use its tree plotting function to make it possible to see significantly increased ontology trees.

**Usage**

```
goseq_trees(de_genes, godata, goid_map = "reference/go/id2go.map",
  score_limit = 0.01, goids_df = NULL, overwrite = FALSE,
  selector = "topDiffGenes", pval_column = "adj.P.Val")
```

**Arguments**

de_genes	Some differentially expressed genes.
godata	Data from goseq.
goid_map	File to save go id mapping.
score_limit	Score limit for the coloring.
goids_df	Mapping of IDs to GO in the Ramigo expected format.
overwrite	Overwrite the trees?
selector	Function for choosing genes.
pval_column	Column to acquire pvalues.

**Value**

A plot!

**See Also**

**Ramigo**

---

gostats_kegg	<i>Use gostats() against kegg pathways.</i>
--------------	---------------------------------------------

---

**Description**

This sets up a GSEABase analysis using KEGG pathways rather than gene ontologies. Does this even work? I don't think I have ever tested it yet. oh, it sort of does, maybe if I export it I will rembmer it.

**Usage**

```
gostats_kegg(organism = "Homo sapiens", pathdb = "org.Hs.egPATH",
             godb = "org.Hs.egGO")
```

**Arguments**

organism	The organism used to make the KEGG frame, human readable no taxonomic.
pathdb	Name of the pathway database for this organism.
godb	Name of the ontology database for this organism.

**Value**

Results from hyperGTest using the KEGG pathways.



---

`gostats_trees`*Take gostats data and print it on a tree as topGO does.*

---

## Description

This shoe horns gostats data into a format acceptable by topgo and uses it to print pretty ontology trees showing the over represented ontologies.

## Usage

```
gostats_trees(de_genes, mf_over, bp_over, cc_over, mf_under, bp_under, cc_under,
  goid_map = "reference/go/id2go.map", score_limit = 0.01,
  goids_df = NULL, overwrite = FALSE, selector = "topDiffGenes",
  pval_column = "adj.P.Val")
```

## Arguments

<code>de_genes</code>	Some differentially expressed genes.
<code>mf_over</code>	Mfover data.
<code>bp_over</code>	Bpover data.
<code>cc_over</code>	Ccover data.
<code>mf_under</code>	Mfunder data.
<code>bp_under</code>	Bpunder data.
<code>cc_under</code>	Ccunder expression data.
<code>goid_map</code>	Mapping of IDs to GO in the Ramigo expected format.
<code>score_limit</code>	Maximum score to include as 'significant'.
<code>goids_df</code>	Dataframe of available goids (used to generate <code>goid_map</code> ).
<code>overwrite</code>	Overwrite the <code>goid_map</code> ?
<code>selector</code>	Function to choose differentially expressed genes in the data.
<code>pval_column</code>	Column in the data to be used to extract pvalue scores.

## Value

plots! Trees! oh my!

## See Also

**topGO**

---

 gosyn

*Get a go synonym from an ID.*


---

### Description

I think I will need to do similar parsing of the output for this function as per `gosec()` In some cases this also returns stuff like `c("some text", "GO:someID")` versus "some other text" versus NULL versus NA. This function just goes a `mapply(gosn, go)`.

### Usage

```
gosyn(go = "GO:0000001")
```

### Arguments

`go` GO id, this may be a character or list(assuming the elements are goids).

### Value

Some text providing the synonyms for the given id(s).

### See Also

**GOTermsAnnDbBimap**

### Examples

```
## Not run:
text = gosyn("GO:0000001")
text
## > GO:000001
## > "mitochondrial inheritance"

## End(Not run)
```

---

 goterm

*Get a go term from ID.*


---

### Description

Get a go term from ID.

### Usage

```
goterm(go = "GO:0032559")
```

**Arguments**

go                      GO id or a list thereof, this may be a character or list(assuming the elements, not names, are goids).

**Value**

Some text containing the terms associated with GO id(s).

**See Also**

**GOTermsAnnDbBimap**

**Examples**

```
## Not run:
goterm("GO:0032559")
## > GO:0032559
## > "adenyl ribonucleotide binding"

## End(Not run)
```

---

gotest

*Test GO ids to see if they are useful.*

---

**Description**

This just wraps gotst in mapply.

**Usage**

```
gotest(go)
```

**Arguments**

go                      go IDs as characters.

**Value**

Some text

**See Also**

**GOTermsAnnDbBimap**

**Examples**

```
## Not run:
gotest("G0:0032559")
## > 1
gotest("G0:0923429034823904")
## > 0

## End(Not run)
```

graph\_metrics

*Make lots of graphs!***Description**

Plot out a set of metrics describing the state of an experiment including library sizes, # non-zero genes, heatmaps, boxplots, density plots, pca plots, standard median distance/correlation, and qq plots.

**Usage**

```
graph_metrics(expt, cormethod = "pearson", distmethod = "euclidean",
  title_suffix = NULL, qq = NULL, ma = NULL, ...)
```

**Arguments**

expt	an expt to process
cormethod	the correlation test for heatmaps.
distmethod	define the distance metric for heatmaps.
title_suffix	text to add to the titles of the plots.
qq	include qq plots
ma	include pairwise ma plots
...	extra parameters optionally fed to the various plots

**Value**

a loooong list of plots including the following: nonzero = a ggplot2 plot of the non-zero genes vs library size libsize = a ggplot2 bar plot of the library sizes boxplot = a ggplot2 boxplot of the raw data corheat = a recordPlot()ed pairwise correlation heatmap of the raw data smc = a recordPlot()ed view of the standard median pairwise correlation of the raw data disheat = a recordPlot()ed pairwise euclidean distance heatmap of the raw data smd = a recordPlot()ed view of the standard median pairwise distance of the raw data pcaplot = a recordPlot()ed PCA plot of the raw samples pcatable = a table describing the relative contribution of condition/batch of the raw data pcares = a table describing the relative contribution of condition/batch of the raw data pcavar = a table describing the variance of the raw data qq = a recordPlotted() view comparing the quantile/quantiles between the mean of all data and every raw sample density = a ggplot2 view of the density of each raw sample (this is complementary but more fun than a boxplot)

**See Also**

**Biobase** **ggplot2** **grDevices** **gplots** **exprs** **hpgl\_norm** **plot\_nonzero** **plot\_libsize** **plot\_boxplot** **plot\_corheat**  
**plot\_sm** **plot\_disheat** **plot\_pca** **plot\_qq\_all** **plot\_pairwise\_ma**

**Examples**

```
## Not run:
toomany_plots <- graph_metrics(expt)
toomany_plots$pcaplot
norm <- normalize_expt(expt, convert="cpm", batch=TRUE, filter_low=TRUE,
                      transform="log2", norm="rle")
holy_asscrackers <- graph_metrics(norm, qq=TRUE, ma=TRUE)
## good luck, you are going to be waiting a while for the ma plots to print!

## End(Not run)
```

heatmap.3

*a minor change to heatmap.2 makes heatmap.3***Description**

heatmap.2 is the devil.

**Usage**

```
heatmap.3(x, Rowv = TRUE, Colv = if (symm) "Rowv" else TRUE,
  distfun = dist, hclustfun = hclust, dendrogram = c("both", "row",
  "column", "none"), reorderfun = function(d, w) reorder(d, w),
  symm = FALSE, scale = c("none", "row", "column"), na.rm = TRUE,
  revC = identical(Colv, "Rowv"), add.expr, breaks, symbreaks = min(x < 0,
  na.rm = TRUE) || scale != "none", col = "heat.colors", colsep, rowsep,
  sepcolor = "white", sepwidth = c(0.05, 0.05), cellnote, notecex = 1,
  notecol = "cyan", na.color = par("bg"), trace = c("column", "row",
  "both", "none"), tracecol = "cyan", hline = median(breaks),
  vline = median(breaks), linecol = tracecol, margins = c(5, 5),
  ColSideColors, RowSideColors, cexRow = 0.2 + 1/log10(nr), cexCol = 0.2 +
  1/log10(nc), labRow = NULL, labCol = NULL, srtRow = NULL,
  srtCol = NULL, adjRow = c(0, NA), adjCol = c(NA, 0), offsetRow = 0.5,
  offsetCol = 0.5, key = TRUE, keysize = 1.5,
  density.info = c("histogram", "density", "none"), denscol = tracecol,
  symkey = min(x < 0, na.rm = TRUE) || symbreaks, densadj = 0.25,
  key.title = NULL, key.xlab = NULL, key.ylab = NULL,
  key.xtickfun = NULL, key.ytickfun = NULL, key.par = list(),
  main = NULL, xlab = NULL, ylab = NULL, lmat = NULL, lhei = NULL,
  lwid = NULL, extrafun = NULL, linewidth = 1, ...)
```

**Arguments**

x	data
Rowv	add rows?
Colv	add columns?
distfun	distance function to use
hclustfun	clustering function to use
dendrogram	which axes to put trees on
reorderfun	reorder the rows/columns?
symm	symmetrical?
scale	add the scale?
na.rm	remove nas from the data?
revC	reverse the columns?
add.expr	no clue
breaks	also no clue
symbreaks	still no clue
col	colors!
colsep	column separator
rowsep	row separator
sepcolor	color to put between columns/rows
sepwidth	how much to separate
cellnote	mur?
notecex	size of the notes
notecol	color of the notes
na.color	a parameter call to bg
trace	do a trace for rows/columns?
tracecol	color of the trace
hline	the hline
vline	the vline
linecol	the line color
margins	margins are good
ColSideColors	colors for the columns as annotation
RowSideColors	colors for the rows as annotation
cexRow	row size
cexCol	column size
labRow	hmmmm
labCol	still dont know
srtRow	srt the row?

srtCol	srt the column?
adjRow	adj the row?
adjCol	adj the column?
offsetRow	how far to place the text from the row
offsetCol	how far to place the text from the column
key	add a key?
keysize	if so, how big?
density.info	for the key, what information to add
denscol	tracecol hmm ok
symkey	I like keys
densadj	adj the dens?
key.title	title for the key
key.xlab	text for the x axis of the key
key.ylab	text for the y axis of the key
key.xtickfun	add text to the ticks of the key x axis
key.ytickfun	add text to the ticks of the key y axis
key.par	parameters for the key
main	the main title of the plot
xlab	main x label
ylab	main y label
lmat	the lmat
lhei	the lhei
lwid	the lwid
extrafun	I do enjoy me some extra fun
linewidth	the width of lines
...	because this function did not already have enough options

**Value**

a heatmap!

---

hpgltools

*hpgltools: a suite of tools to make our analyses easier*


---

## Description

This provides a series of helpers for working with sequencing data

## Details

It falls under a few main topics

- Data exploration, look for trends in sequencing data and identify batch effects or skewed distributions.
- Differential expression analyses, use DESeq2/limma/EdgeR in a hopefully robust and flexible fashion.
- Ontology analyses, use goseq/clusterProfiler/topGO/GOStats/gProfiler in hopefully robust ways.
- Perform some simple TnSeq analyses.

To see examples of this inaction, check out the vignettes: `browseVignettes(package = 'hpgltools')`

---

hpgl\_arescore

*Implement the arescan function in R*


---

## Description

This function was taken almost verbatim from AREScore() in SeqTools Available at: <https://github.com/lianos/seqtools.git> At least on my computer I could not make that implementation work So I rewrapped its apply() calls and am now hoping to extend its logic a little to make it more sensitive and get rid of some of the spurious parameters or at least make them more transparent.

## Usage

```
hpgl_arescore(x, basal = 1, overlapping = 1.5, d1.3 = 0.75, d4.6 = 0.4,
             d7.9 = 0.2, within.AU = 0.3, aub.min.length = 10,
             aub.p.to.start = 0.8, aub.p.to.end = 0.55)
```

## Arguments

x	DNA/RNA StringSet containing the UTR sequences of interest
basal	I dunno.
overlapping	default=1.5
d1.3	default=0.75 These parameter names are so stupid, lets be realistic
d4.6	default=0.4



```

d7.9          default=0.2
within.AU     default=0.3
aub.min.length default=10
aub.p.to.start default=0.8
aub.p.to.end   default=0.55

```

## Details

Note that I did this two months ago and haven't touched it since...

## Value

a DataFrame of scores

## See Also

**IRanges Biostrings**

## Examples

```

## Not run:
## Extract all the genes from my genome, pull a static region 120nt following the stop
## and test them for potential ARE sequences.
## FIXME: There may be an error in this example, another version I have handles the +/- strand
## genes separately, I need to return to this and check if it is providing the 5' UTR for 1/2
## the genome, which would be unfortunate -- but the logic for testing remains the same.
are_candidates <- hpgl_arescore(genome)
utr_genes <- subset(lmajor_annotations, type == 'gene')
threep <- GenomicRanges::GRanges(seqnames=Rle(utr_genes[,1]),
                                ranges=IRanges(utr_genes[,3], end=(utr_genes[,3] + 120)),
                                strand=Rle(utr_genes[,5]),
                                name=Rle(utr_genes[,10]))
threep_seqstrings <- Biostrings::getSeq(lm, threep)
are_test <- hpgltools::hpgl_arescore(x=threep_seqstrings)
are_genes <- rownames(are_test[ which(are_test$score > 0), ])

## End(Not run)

```

---

hpgl\_combatMod

*A modified version of comBatMod.*


---

## Description

This is a hack of Kwame Okrah's `combatMod` to make it not fail on corner-cases. This was mostly copy/pasted from <https://github.com/kokrah/cbcbSEQ/blob/master/R/transform.R>

## Usage

```
hpgl_combatMod(dat, batch, mod, noScale = TRUE, prior.plots = FALSE, ...)
```

**Arguments**

<code>dat</code>	Df to modify.
<code>batch</code>	Factor of batches.
<code>mod</code>	Factor of conditions.
<code>noScale</code>	The normal 'scale' option squishes the data too much, so this defaults to TRUE.
<code>prior.plots</code>	Print out prior plots?
<code>...</code>	Extra options are passed to <code>arglist</code>

**Value**

Df of batch corrected data

**See Also**

`sva` [ComBat](#)

**Examples**

```
## Not run:
df_new = hpgl_combatMod(df, batches, model)

## End(Not run)
```

---

### hpgl\_cor

---

*Wrap cor() to include robust correlations.*

---

**Description**

Take `covRob`'s robust correlation coefficient and add it to the set of correlations available when one calls `cor()`.

**Usage**

```
hpgl_cor(df, method = "pearson", ...)
```

**Arguments**

<code>df</code>	Data frame to test.
<code>method</code>	Correlation method to use. Includes <code>pearson</code> , <code>spearman</code> , <code>kendal</code> , <code>robust</code> .
<code>...</code>	Other options to pass to <code>stats::cor()</code> .

**Value**

Some fun correlation statistics.

**See Also****robust** [cor](#) [cov](#) [covRob](#)**Examples**

```
## Not run:
hpgl_cor(df=df)
hpgl_cor(df=df, method="robust")

## End(Not run)
```

---

hpgl\_enrich.internal    *A minor hack in the clusterProfiler function 'enrich.internal'.*

---

**Description**

I do not remember any longer why, but enrichGO errors out in ways which do not always make sense, this was written to alleviate that problem. I believe I sent a diff to the clusterProfiler author but did not hear back and so added this function.

**Usage**

```
hpgl_enrich.internal(gene, organism, pvalueCutoff = 1,
  pAdjustMethod = "fdr", ont, minGSSize = 2, qvalueCutoff = 0.2,
  readable = FALSE, universe = NULL)
```

**Arguments**

gene	Differentially expressed genes.
organism	Pull ensembl annotations if this is a supported species.
pvalueCutoff	P-value cutoff.
pAdjustMethod	P-adjust method.
ont	Molecular function, Biological process, or Cellular component?
minGSSize	Minimum gs size?
qvalueCutoff	Maximum allowed q-value.
readable	Set the readable flag for the DOSE object?
universe	Universe of genes to score significance against.

**Value**

Some clusterProfiler data.

**See Also****clusterProfiler**

---

hpgl\_enrichGO*A minor hack in the clusterProfiler function 'enrichGO'.*

---

## Description

I do not remember any longer why, but enrichGO errors out in ways which do not always make sense, this was written to alleviate that problem. I believe I sent a diff to the clusterProfiler author but did not hear back and so added this function.

## Usage

```
hpgl_enrichGO(gene, organism = "human", ont = "MF", pvalueCutoff = 0.05,  
  pAdjustMethod = "BH", universe, qvalueCutoff = 0.2, minGSSize = 2,  
  readable = FALSE)
```

## Arguments

gene	Some differentially expressed genes.
organism	if used will cause this to pull the ensG annotations.
ont	Molecular function, Biological process, or Cellular component?
pvalueCutoff	P-value cutoff.
pAdjustMethod	P-value adjustment.
universe	Gene universe to use.
qvalueCutoff	Maximum qvalue before adding.
minGSSize	Smallest ontology group size allowed.
readable	Set the readable tag on the returned object?

## Value

Some clusterProfiler data.

## See Also

**clusterProfiler**

---

hpgl_Gff2GeneTable	<i>A near copy-paste of clusterProfiler's readGff().</i>
--------------------	----------------------------------------------------------

---

**Description**

There is a redundant merge in the original code which caused my invocations to use up all the memory on my machine.

**Usage**

```
hpgl_Gff2GeneTable(gffFile, compress = TRUE, split = "=")
```

**Arguments**

gffFile	Gff file of annotations.
compress	Compress the results?
split	Splitter when reading gff files to extract annotation information.

**Value**

geneTable.rda file of gene attributes.

---

hpgl_GOplot	<i>A minor hack of the topGO GOplot function.</i>
-------------	---------------------------------------------------

---

**Description**

This allows me to change the line widths from the default.

**Usage**

```
hpgl_GOplot(dag, sigNodes, dag.name = "GO terms", edgeTypes = TRUE,  
  nodeShape.type = c("box", "circle", "ellipse", "plaintext")[3],  
  genNodes = NULL, wantedNodes = NULL, showEdges = TRUE,  
  useFullNames = TRUE, oldSigNodes = NULL, nodeInfo = NULL,  
  maxchars = 30)
```

**Arguments**

dag	DAG tree of ontologies.
sigNodes	Set of significant ontologies (with p-values).
dag.name	Name for the graph.
edgeTypes	Types of the edges for graphviz.
nodeShape.type	Shapes on the tree.

genNodes	Generate the nodes?
wantedNodes	Subset of the ontologies to plot.
showEdges	Show the arrows?
useFullNames	Full names of the ontologies (they can get long).
oldSigNodes	I dunno.
nodeInfo	Hmm.
maxchars	Maximum characters per line inside the shapes.

**Value**

Topgo plot!

---

hpgl_GroupDensity	<i>A hack of topGO's groupDensity()</i>
-------------------	-----------------------------------------

---

**Description**

This just adds a couple wrappers to avoid errors in groupDensity.

**Usage**

```
hpgl_GroupDensity(object, whichGO, ranks = TRUE, rm.one = FALSE)
```

**Arguments**

object	TopGO enrichment object.
whichGO	Individual ontology group to compare against.
ranks	Rank order the set of ontologies?
rm.one	Remove pvalue=1 groups?

**Value**

plot of group densities.

---

hpgl_log2cpm	<i>Converts count matrix to log2 counts-per-million reads.</i>
--------------	----------------------------------------------------------------

---

**Description**

Based on the method used by limma as described in the Law et al. (2014) voom paper.

**Usage**

```
hpgl_log2cpm(counts, lib.size = NULL)
```

**Arguments**

counts	Read count matrix.
lib.size	Library size.

**Value**

log2-CPM read count matrix.

**See Also**

**edgeR**

**Examples**

```
## Not run:  
l2cpm <- hpgl_log2cpm(counts)  
  
## End(Not run)
```

---

hpgl_norm	<i>Normalize a dataframe/expt, express it, and/or transform it</i>
-----------	--------------------------------------------------------------------

---

**Description**

There are many possible options to this function. Refer to `normalize_expt()` for a more complete list.

**Usage**

```
hpgl_norm(data, ...)
```

**Arguments**

data	Some data as a df/expt/whatever.
...	I should put all those other options here

**Value**

edgeR's DGEList expression of a count table. This seems to me to be the easiest to deal with.

**See Also**

[cpm](#) [rpkm](#) [hpgl\\_rpkm](#) [DESeqDataSetFromMatrix](#) [estimateSizeFactors](#) [DGEList](#) [calcNormFactors](#)

**Examples**

```
## Not run:
df_raw = hpgl_norm(expt=expt) ## Only performs low-count filtering
df_raw = hpgl_norm(df=a_df, design=a_design) ## Same, but using a df
df_ql2rpkm = hpgl_norm(expt=expt, norm='quant', transform='log2',
                      convert='rpkm') ## Quantile, log2, rpkm
count_table = df_ql2rpkm$counts

## End(Not run)
```

---

hpgl\_pathview

---

*Print some data onto KEGG pathways.*


---

**Description**

KEGGREST and pathview provide neat functions for coloring molecular pathways with arbitrary data. Unfortunately they are somewhat evil to use. This attempts to alleviate that.

**Usage**

```
hpgl_pathview(path_data, indir = "pathview_in", outdir = "pathview",
             pathway = "all", species = "lma", from_list = NULL, to_list = NULL,
             suffix = "_colored", filenames = "id", fc_column = "limma_logfc",
             format = "png", verbose = TRUE)
```

**Arguments**

path_data	Some differentially expressed genes.
indir	Directory into which the unmodified kegg images will be downloaded (or already exist).
outdir	Directory which will contain the colored images.
pathway	Perform the coloring for a specific pathway?
species	Kegg identifier for the species of interest.
from_list	Regex to help in renaming KEGG categories/gene names from one format to another.
to_list	Regex to help in renaming KEGG categories/gene names from one format to another.
suffix	Add a suffix to the completed, colored files.
filenames	Name the final files by id or name?



**Value**

A list of some information for every KEGG pathway downloaded/examined. This information includes: a. The filename of the final image for each pathway. b. The number of genes which were found in each pathway image. c. The number of genes in the 'up' category d. The number of genes in the 'down' category

**See Also**

**Ramigo pathview**

**Examples**

```
## Not run:
thy_el_comp2_path = hpgl_pathview(thy_el_comp2_kegg, species="spz", indir="pathview_in",
                                outdir="kegg_thy_el_comp2", string_from="_Spy",
                                string_to="_Spy_", filenames="pathname")

## End(Not run)
```

---

hpgl\_qshrink

*A hacked copy of Kwame's qsmooth/qstats code.*


---

**Description**

I made a couple small changes to Kwame's qstats() function to make it not fail when on corner-cases. I sent him a diff, but haven't checked to see if it was useful yet.

**Usage**

```
hpgl_qshrink(data = NULL, groups = NULL, refType = "mean",
             groupLoc = "mean", window = 99, groupCol = NULL, plot = TRUE, ...)
```

**Arguments**

data	Count table to modify
groups	Factor of the experimental conditions
refType	Method for grouping conditions
groupLoc	Method for grouping groups
window	Window, for looking!
groupCol	Column to define conditions
plot	Plot the quantiles?
...	More options

**Value**

New data frame of normalized counts

**See Also****qsmooth****Examples**

```
## Not run:  
df <- hpgl_qshrink(data)  
  
## End(Not run)
```

---

hpgl\_qstats*A hacked copy of Kwame's qsmooth/qstats code.*

---

**Description**

I made a couple small changes to Kwame's qstats() function to make it not fail when on corner-cases. I sent him a diff, but haven't checked to see if it was useful yet.

**Usage**

```
hpgl_qstats(data, groups, refType = "mean", groupLoc = "mean",  
            window = 99)
```

**Arguments**

data	Initial count data
groups	Experimental conditions as a factor.
refType	Method to separate groups, mean or median.
groupLoc	I don't remember what this is for.
window	Window for basking!

**Value**

Some new data.

**Examples**

```
## Not run:  
qstatted <- hpgl_qstats(data, conditions)  
  
## End(Not run)
```

---

hpgl_read_files	<i>Read a bunch of count tables and create a usable data frame from them.</i>
-----------------	-------------------------------------------------------------------------------

---

## Description

It is worth noting that this function has some logic intended for the elsayed lab's data storage structure. It shouldn't interfere with other usages, but it attempts to take into account different ways the data might be stored.

## Usage

```
hpgl_read_files(ids, files, header = FALSE, include_summary_rows = FALSE,  
  suffix = NULL, ...)
```

## Arguments

ids	List of experimental ids.
files	List of files to read.
header	Whether or not the count tables include a header row.
include_summary_rows	Whether HTSeq summary rows should be included.
suffix	Optional suffix to add to the filenames when reading them.
...	More options for happy time!

## Value

Data frame of count tables.

## See Also

[create\\_expt](#)

## Examples

```
## Not run:  
count_tables = hpgl_read_files(as.character(sample_ids), as.character(count_filenames))  
  
## End(Not run)
```

---

hpgl_rpkm	<i>Reads/(kilobase(gene) * million reads)</i>
-----------	-----------------------------------------------

---

### Description

Express a data frame of counts as reads per kilobase(gene) per million(library). This function wraps EdgeR's rpkm in an attempt to make sure that the required gene lengths get sent along.

### Usage

```
hpgl_rpkm(df, ...)
```

### Arguments

df	Data frame of counts, alternately an edgeR DGEList.
...	extra options including annotations for defining gene lengths.

### Value

Data frame of counts expressed as rpkm.

### See Also

**edgeR** and [cpm rpkm](#)

### Examples

```
## Not run:
rpkm_df = hpgl_rpkm(df, annotations=gene_annotations)

## End(Not run)
```

---

hpgl_voom	<i>A slight modification of limma's voom().</i>
-----------	-------------------------------------------------

---

### Description

Estimate mean-variance relationship between samples and generate 'observational-level weights' in preparation for linear modeling RNAseq data. This particular implementation was primarily scabbed from cbcSEQ, but changes the mean-variance plot slightly and attempts to handle corner cases where the sample design is confounded by setting the coefficient to 1 for those samples rather than throwing an unhelpful error. Also, the Elist output gets a 'plot' slot which contains the plot rather than just printing it.

**Usage**

```
hpgl_voom(dataframe, model = NULL, libsize = NULL, stupid = FALSE,
  logged = FALSE, converted = FALSE)
```

**Arguments**

dataframe	Dataframe of sample counts which have been normalized and log transformed.
model	Experimental model defining batches/conditions/etc.
libsize	Size of the libraries (usually provided by edgeR).
stupid	Cheat when the resulting matrix is not solvable?
logged	Is the input data is known to be logged?
converted	Is the input data is known to be cpm converted?

**Value**

EList containing the following information: E = The normalized data weights = The weights of said data design = The resulting design lib.size = The size in pseudocounts of the library plot = A ggplot of the mean/variance trend with a blue loess fit and red trend fit

**See Also**

[voom lmFit](#)

**Examples**

```
## Not run:
funkytown = hpgl_voom(samples, model)

## End(Not run)
```

---

kegg_get_orgn	<i>Search KEGG identifiers for a given species name.</i>
---------------	----------------------------------------------------------

---

**Description**

KEGG identifiers do not always make sense. For example, how am I supposed to remember that Leishmania major is lmj? This takes in a human readable string and finds the KEGG identifiers that match it.

**Usage**

```
kegg_get_orgn(species = "Leishmania", short = TRUE)
```

**Arguments**

species	Search string (Something like 'Homo sapiens').
short	Only pull the orgid?

**Value**

Data frame of possible KEGG identifier codes, genome ID numbers, species, and phylogenetic classifications.

**See Also**

**RCurl**

**Examples**

```
## Not run:
  fun = kegg_get_orgn('Canis')
## >      Tid      orgid      species      phylogeny
## > 17 T01007   cfa Canis familiaris (dog) Eukaryotes;Animals;Vertebrates;Mammals

## End(Not run)
```

---

kegg_to_ensembl	<i>Maps KEGG identifiers to ENSEMBL gene ids.</i>
-----------------	---------------------------------------------------

---

**Description**

Takes a list of KEGG gene identifiers and returns a list of ENSEMBL ids corresponding to those genes.

**Usage**

```
kegg_to_ensembl(kegg_ids)
```

**Arguments**

kegg\_ids            List of KEGG identifiers to be mapped.

**Value**

Ensembl IDs as a character list.

**See Also**

[keggGet](#)

**Examples**

```
## Not run:
ensembl_list <- kegg_to_ensembl("a")

## End(Not run)
```

---

limma\_coefficient\_scatter

*Plot out 2 coefficients with respect to one another from limma.*


---

## Description

It can be nice to see a plot of two coefficients from a limma comparison with respect to one another. This hopefully makes that easy.

## Usage

```
limma_coefficient_scatter(output, toptable = NULL, x = 1, y = 2,
  gvis_filename = NULL, gvis_trendline = TRUE, z = 1.5,
  tooltip_data = NULL, base_url = NULL, color_low = "#DD0000",
  color_high = "#7B9F35", ...)
```

## Arguments

output	Set of pairwise comparisons provided by limma_pairwise().
toptable	Use this to get up/downs and color them on the scatter plot.
x	Name or number of the x-axis coefficient column to extract.
y	Name or number of the y-axis coefficient column to extract
gvis_filename	Filename for plotting gvis interactive graphs of the data.
gvis_trendline	Add a trendline to the gvis plot?
z	How far from the median to color the plot red and green.
tooltip_data	Dataframe of gene annotations to be used in the gvis plot.
base_url	Basename for gvis plots.
color_low	Color for the ups.
color_high	Color for the downs.
...	More parameters to make you happy!

## Value

Ggplot2 plot showing the relationship between the two coefficients.

## See Also

[plot\\_linear\\_scatter limma\\_pairwise](#)

## Examples

```
## Not run:
pretty = coefficient_scatter(limma_data, x="wt", y="mut")

## End(Not run)
```

---

limma_pairwise	<i>Set up a model matrix and set of contrasts for pairwise comparisons using voom/limma.</i>
----------------	----------------------------------------------------------------------------------------------

---

## Description

Creates the set of all possible contrasts and performs them using voom/limma.

## Usage

```
limma_pairwise(input, conditions = NULL, batches = NULL,
  model_cond = TRUE, model_batch = TRUE, model_intercept = TRUE,
  extra_contrasts = NULL, alt_model = NULL, libsize = NULL,
  annot_df = NULL, ...)
```

## Arguments

input	Dataframe/vector or expt class containing count tables, normalization state, etc.
conditions	Factor of conditions in the experiment.
batches	Factor of batches in the experiment.
model_cond	Include condition in the model?
model_batch	Include batch in the model? This is hopefully TRUE.
model_intercept	Perform a cell-means or intercept model? A little more difficult for me to understand. I have tested and get the same answer either way.
extra_contrasts	Some extra contrasts to add to the list. This can be pretty neat, lets say one has conditions A,B,C,D,E and wants to do (C/B)/A and (E/D)/A or (E/D)/(C/B) then use this with a string like: "c_vs_b_ctrla = (C-B)-A, e_vs_d_ctrla = (E-D)-A, de_vs_cb = (E-D)-(C-B),"
alt_model	Separate model matrix instead of the normal condition/batch.
libsize	I've recently figured out that libsize is far more important than I previously realized. Play with it here.
annot_df	Data frame for annotations.
...	Use the elipsis parameter to feed options to write_limma().

## Value

List including the following information: macb = the mashing together of condition/batch so you can look at it macb\_model = The result of calling model.matrix(~0 + macb) macb\_fit = The result of calling lmFit(data, macb\_model) voom\_result = The result from voom() voom\_design = The design from voom (redundant from voom\_result, but convenient) macb\_table = A table of the number of times each condition/batch pairing happens cond\_table = A table of the number of times each condition appears (the denominator for the identities) batch\_table = How many times each



batch appears identities = The list of strings defining each condition by itself all\_pairwise = The list of strings defining all the pairwise contrasts contrast\_string = The string making up the makeContrasts() call pairwise\_fits = The result from calling contrasts.fit() pairwise\_comparisons = The result from eBayes() limma\_result = The result from calling write\_limma()

## See Also

[write\\_limma](#)

## Examples

```
## Not run:
pretend = balanced_pairwise(data, conditions, batches)

## End(Not run)
```

---

limma_scatter	<i>Plot arbitrary data from limma as a scatter plot.</i>
---------------	----------------------------------------------------------

---

## Description

Extract the adjusted abundances for the two conditions used in the pairw

## Usage

```
limma_scatter(all_pairwise_result, first_table = 1, first_column = "logFC",
              second_table = 2, second_column = "logFC", type = "linear_scatter", ...)
```

## Arguments

all_pairwise_result	Result from calling balanced_pairwise().
first_table	First table from all_pairwise_result\$limma_result to look at (may be a name or number).
first_column	Name of the column to plot from the first table.
second_table	Second table inside all_pairwise_result\$limma_result (name or number).
second_column	Column to compare against.
type	Type of scatter plot (linear model, distance, vanilla).
...	Use the elipsis to feed options to the html graphs.

## Value

plot\_linear\_scatter() set of plots comparing the chosen columns. If you forget to specify tables to compare, it will try the first vs the second.

**See Also**

[plot\\_linear\\_scatter](#) [topTable](#)

**Examples**

```
## Not run:
compare_logFC = limma_scatter(all_pairwise, first_table="wild_type", second_column="mutant",
                             first_table="AveExpr", second_column="AveExpr")
compare_B = limma_scatter(all_pairwise, first_column="B", second_column="B")

## End(Not run)
```

---

loadme

*Load a backup rdata file*

---

**Description**

I often use R over a sshfs connection, sometimes with significant latency, and I want to be able to save/load my R sessions relatively quickly. Thus this function uses my backup directory to load its R environment.

**Usage**

```
loadme(directory = "savefiles", filename = "Rdata.rda.xz")
```

**Arguments**

dir                      Directory containing the RData.rda.xz file.

**Value**

a bigger global environment

**See Also**

[load](#) [save](#)

**Examples**

```
## Not run:
## loadme()

## End(Not run)
```

---

load_annotations	<i>Load organism annotation data (parasite).</i>
------------------	--------------------------------------------------

---

## Description

Creates a dataframe gene and transcript information for a given set of gene ids using the OrganismDbi interface.

## Usage

```
load_annotations(orgdb, gene_ids = NULL, include_go = FALSE,  
  keytype = "ENSEMBL", fields = NULL, sum_exons = FALSE)
```

## Arguments

orgdb	OrganismDb instance.
gene_ids	Gene identifiers for retrieving annotations.
keytype	mmm the key type used?
fields	Columns included in the output.
sum_exons	Perform a sum of the exons in the data set?

## Value

Table of geneids, chromosomes, descriptions, strands, types, and lengths.

## See Also

[select](#)

## Examples

```
## Not run:  
one_gene <- load_parasite_annotations(org, c("LmJF.01.0010"))  
  
## End(Not run)
```

---

load_go_terms	<i>Retrieve GO terms associated with a set of genes.</i>
---------------	----------------------------------------------------------

---

### Description

AnnotationDbi provides a reasonably complete set of GO mappings between gene ID and ontologies. This will extract that table for a given set of gene IDs.

### Usage

```
load_go_terms(orgdb, gene_ids, keytype = "ENSEMBL")
```

### Arguments

orgdb	OrganismDb instance.
gene_ids	Identifiers of the genes to retrieve annotations.
keytype	the mysterious keytype returns yet again to haunt my dreams

### Value

Data frame of gene IDs, go terms, and names.

### See Also

[select](#)

### Examples

```
## Not run:
go_terms <- load_go_terms(org, c("a","b"))

## End(Not run)
```

---

load_host_annotations	<i>Load organism annotation data (mouse/human).</i>
-----------------------	-----------------------------------------------------

---

### Description

Creates a dataframe gene and transcript information for a given set of gene ids using the OrganismDbi interface.

### Usage

```
load_host_annotations(orgdb, gene_ids = NULL, keytype = "ENSEMBL",
  fields = c("TXCHROM", "GENENAME", "TXSTRAND", "TXSTART", "TXEND"),
  biomaht_dataset = "hsapiens_gene_ensembl")
```

**Arguments**

orgdb	OrganismDb instance.
gene_ids	Gene identifiers for retrieving annotations.
keytype	a, umm keytype? I need to properly read this code.
fields	Columns to include in the output.
biomart_dataset	Name of the biomaRt dataset to query for gene type.

**Value**

a table of gene information

**See Also**

[select](#)

**Examples**

```
## Not run:
host <- load_host_annotations(org, c("a", "b"))

## End(Not run)
```

---

load_kegg_mapping	<i>Creates a gene/KEGG mapping dataframe.</i>
-------------------	-----------------------------------------------

---

**Description**

In much the same way AnnotationDbi provides GO data, it also provides KEGG data.

**Usage**

```
load_kegg_mapping(orgdb, gene_ids, keytype = "ENSEMBL")
```

**Arguments**

orgdb	OrganismDb instance.
gene_ids	Identifiers of the genes to retrieve annotations.
keytype	the keytype, damn I really need to read this code

**Value**

Df of kegg mappings

**See Also**

[select](#)

## Examples

```
## Not run:
kegg_data <- load_kegg_mapping(org, c("a","b"))

## End(Not run)
```

---

load_kegg_pathways	<i>Creates a KEGG pathway/description mapping dataframe.</i>
--------------------	--------------------------------------------------------------

---

## Description

Use AnnotationDbi to map descriptions of KEGG pathways to gene IDs.

## Usage

```
load_kegg_pathways(orgdb, gene_ids, keytype = "ENSEMBL")
```

## Arguments

orgdb	OrganismDb instance.
gene_ids	Identifiers of the genes to retrieve annotations.
keytype	as per the previous functions, I don't know what this does yet

## Value

Character list of pathways.

## See Also

[select](#)

## Examples

```
## Not run:
pathnames <- load_kegg_pathways(org, c("a","b","c"))

## End(Not run)
```

---

makeSVD	<i>this a function scabbed from Hector and Kwame's cbcSEQ</i>
---------	---------------------------------------------------------------

---

**Description**

It just does fast.svd of a matrix comprised of the matrix - rowMeans(matrix)

**Usage**

```
makeSVD(data)
```

**Arguments**

data	A data frame to decompose
------	---------------------------

**Value**

a list containing the s,v,u from fast.svd

**See Also**

**corpcor** [fast.svd](#)

**Examples**

```
## Not run:  
svd = makeSVD(data)  
  
## End(Not run)
```

---

make_exempladata	<i>Small hack of limma's exampleData() to allow for arbitrary data set sizes.</i>
------------------	-----------------------------------------------------------------------------------

---

**Description**

exampleData has a set number of genes/samples it creates. This relaxes that restriction.

**Usage**

```
make_exempladata(ngenes = 1000, columns = 5)
```

**Arguments**

ngenes	How many genes in the fictional data set?
columns	How many samples in this data set?

**Value**

Matrix of pretend counts.

**See Also**

**limma**

**Examples**

```
## Not run:  
pretend = make_exempladata()  
  
## End(Not run)
```

---

make_id2gomap	<i>Make a go mapping from IDs in a format suitable for topGO.</i>
---------------	-------------------------------------------------------------------

---

**Description**

When using a non-supported organism, one must write out mappings in the format expected by topgo. This handles that process and gives a summary of the new table.

**Usage**

```
make_id2gomap(goid_map = "reference/go/id2go.map", goids_df = NULL,  
              overwrite = FALSE)
```

**Arguments**

goid_map	TopGO mapping file.
goids_df	If there is no goid_map, create it with this data frame.
overwrite	Rewrite the mapping file?

**Value**

Summary of the new goid table.



---

make_organismdbi	Create an OrganismDbi for a species at the TriTrypDb
------------------	------------------------------------------------------

---

## Description

OrganismDbi instances are pretty neat, they pull together OrgDb and TxDb. With any luck, this function provides the ability to pull together all the data from the TriTrypDb, GO.db, and KEGGREST in order to accomplish these peculiar tasks.

## Usage

```
make_organismdbi(id = "lmajor_friedlin", cfg = NULL,  
  output_dir = "organdb", ...)
```

## Arguments

id	Unique tritrypdb identifier.
cfg	A configuration dataframe, when null it will be replaced by reading a csv file in inst/extdata.
output_dir	The directory into which to put the various intermediate files, including downloads from the TriTrypdb, the created OrgDb and TxDb instances, and the final OrganismDbi.
...	Extra arguments including a boolean for whether to include kegg.

## Value

A path, some data files, and a kitty!

## See Also

**AnnotationForge**

## Examples

```
## Not run:  
  crazytown <- make_organismdbi() ## wait a loong time  
  
## End(Not run)
```

---

make_orgdb	<i>Make an orgDb object from some information provided by make_orgdb_info()</i>
------------	---------------------------------------------------------------------------------

---

### Description

An orgDb object should provide some useful annotation data including fun stuff like gene ontology, kegg, etc. In the case of the species at the TriTrypDb, much of this information is available in the species .txt file. This function takes that data and collates it into the final orgDb objects using AnnotationForge. It then makes some attempts to ensure that the resulting material created in the filesystem conforms to specifications which allow one to have multiple strains, etc. Finally, if everything goes according to plan, it calls devtools::install() and installs the resulting package.

### Usage

```
make_orgdb(orgdb_info, id = "lmajor_friedlin", cfg = NULL, kegg = TRUE,
  output_dir = "organismdbi", ...)
```

### Arguments

orgdb_info	List of data frames generated by make_orgdb_info()
id	Human readable species identifier, keys off the cfg data frame.
cfg	Configuration data extracted either from inst/eupath_configuration.csv or provided by the user.
kegg	Attempt adding kegg data?
output_dir	Base output directory for the resulting packages.

### Value

List of the resulting package name(s) and whether they installed.

---

make_orgdb_info	<i>Generate the (large) set of data frames required to make functional OrgDb/TxDb/OrganismDbi objects.</i>
-----------------	------------------------------------------------------------------------------------------------------------

---

### Description

This function should probably be split into a few more pieces as it is pretty unwieldy at the moment.

### Usage

```
make_orgdb_info(gff, txt, kegg = TRUE)
```

**Arguments**

gff	File to read gff annotations from.
txt	File to read txt annotations from.
kegg	Boolean deciding whether to try for KEGG data.

**Value**

List containing gene information (likely from the txt file), chromosome information (gff file), gene types (gff file), gene ontology information, and potentially kegg information.

---

`make_pairwise_contrasts`

*Run makeContrasts() with all pairwise comparisons.*

---

**Description**

In order to have uniformly consistent pairwise contrasts, I decided to avoid potential human errors(sic) by having a function generate all contrasts.

**Usage**

```
make_pairwise_contrasts(model, conditions, do_identities = TRUE,  
  do_pairwise = TRUE, extra_contrasts = NULL)
```

**Arguments**

model	Model describing the conditions/batches/etc in the experiment.
conditions	Factor of conditions in the experiment.
do_identities	Include all the identity strings? Limma can use this information while edgeR can not.
do_pairwise	Include all pairwise strings? This shouldn't need to be set to FALSE, but just in case.
extra_contrasts	Optional string of extra contrasts to include.

**Value**

List including the following information: all\_pairwise\_contrasts = the result from makeContrasts(...) identities = the string identifying each condition alone all\_pairwise = the string identifying each pairwise comparison alone contrast\_string = the string passed to R to call makeContrasts(...) names = the names given to the identities/contrasts

**See Also**

[makeContrasts](#)

Examples

```
## Not run:
pretend = make_pairwise_contrasts(model, conditions)

## End(Not run)
```

---

make_report	<i>Make a knitr report with some defaults set a priori.</i>
-------------	-------------------------------------------------------------

---

Description

I keep forgetting to set appropriate options for knitr. This tries to set them.

Usage

```
make_report(name = "report", type = "pdf")
```

Arguments

name	Name the document!
type	Html or pdf reports?

Value

Dated report file.

See Also

**knitr rmarkdown knitrBootstrap**

---

make_tooltips	<i>Create a simple df from a gff which contains tooltips usable in google-Vis graphs.</i>
---------------	-------------------------------------------------------------------------------------------

---

Description

The tooltip column is a handy proxy for more thorough anontations information when it would otherwise be too troublesome to acquire.

Usage

```
make_tooltips(annotations, desc_col = "description", type = "gene",
  id_col = "ID", ...)
```

**Arguments**

annotations	Either a gff file or annotation data frame (which likely came from a gff file.).
desc_col	Gff column from which to gather data.
type	Gff type to use as the master key.
id_col	Which annotation column to cross reference against?
...	Extra arguments dropped into arglist.

**Value**

Df of tooltip information or name of a gff file.

**See Also**

**googleVis** [gff2df](#)

**Examples**

```
## Not run:
tooltips <- make_tooltips('reference/gff/saccharomyces_cerevisiae.gff.gz')

## End(Not run)
```

---

**make\_txdb**

*Create a TxDb object given data provided by make\_orgdb\_info()*

---

**Description**

Much like make\_orgdb() above, this uses the same data to generate a TxDb object.

**Usage**

```
make_txdb(orgdb_info, cfg, gff = NULL, output_dir = "organismdbi", ...)
```

**Arguments**

orgdb_info	List of data frames generated by make_orgdb_info().
cfg	Configuration data frame as per make_orgdb.
gff	File to read

**Value**

List of the resulting txDb package and whether it installed.

---

median_by_factor	<i>Create a data frame of the medians of rows by a given factor in the data.</i>
------------------	----------------------------------------------------------------------------------

---

### Description

This assumes of course that (like expressionsets) there are separate columns for each replicate of the conditions. This will just iterate through the levels of a factor describing the columns, extract them, calculate the median, and add that as a new column in a separate data frame.

### Usage

```
median_by_factor(data, fact)
```

### Arguments

data	Data frame, presumably of counts.
fact	Factor describing the columns in the data.

### Value

Data frame of the medians.

### Examples

```
## Not run:
compressed = hpgltools::median_by_factor(data, experiment$condition)

## End(Not run)
```

---

model_test	<i>Make sure a given experimental factor and design will play together.</i>
------------	-----------------------------------------------------------------------------

---

### Description

Have you ever wanted to set up a differential expression analysis and after minutes of the computer churning away it errors out with some weird error about rank? Then this is the function for you!

### Usage

```
model_test(design, goal = "condition", factors = NULL, ...)
```

**Arguments**

design	Dataframe describing the design of the experiment.
goal	Experimental factor you actually want to learn about.
factors	Experimental factors you rather wish would just go away.
...	I might decide to add more options from other functions.

**Value**

List of booleans telling if the factors + goal will work.

---

my_identifyAUBlocks	<i>copy/paste the function from SeqTools and figure out where it falls on its ass.</i>
---------------------	----------------------------------------------------------------------------------------

---

**Description**

Yeah, I do not remember what I changed in this function.

**Usage**

```
my_identifyAUBlocks(x, min.length = 20, p.to.start = 0.8, p.to.end = 0.55)
```

**Arguments**

x	Sequence object
min.length	I dunno.
p.to.start	P to start of course
p.to.end	The p to end – wtf who makes names like this?

**Value**

a list of IRanges which contain a bunch of As and Us.

---

normalize_counts	<i>Perform a simple normalization of a count table.</i>
------------------	---------------------------------------------------------

---

**Description**

This provides shortcut interfaces for normalization functions from `deseq2`/`edgeR` and friends.

**Usage**

```
normalize_counts(data, design = NULL, norm = "raw", ...)
```

**Arguments**

data	Matrix of count data.
design	Dataframe describing the experimental design. (conditions/batches/etc)
norm	Normalization to perform: 'sfqlquantlqsmoothltmmlupperquartileltmmlrle' I keep wishy-washing on whether design is a required argument.
...	More arguments might be necessary.

**Value**

Dataframe of normalized(counts)

**See Also**

**edgeR** **limma** **DESeq2**

**Examples**

```
## Not run:
norm_table = normalize_counts(count_table, design=design, norm='qsmooth')

## End(Not run)
```

---

normalize_expt	<i>Normalize the data of an expt object. Save the original data, and note what was done.</i>
----------------	----------------------------------------------------------------------------------------------

---

**Description**

It is the responsibility of `normalize_expt()` to perform any arbitrary normalizations desired as well as to ensure that the data integrity is maintained. In order to do this, it writes the actions performed in `expt$state` and saves the intermediate steps of the normalization in `expt$intermediate_counts`. Furthermore, it should tell you every step of the normalization process, from count filtering, to normalization, conversion, transformation, and batch correction.



**Usage**

```
normalize_expt(expt, transform = "raw", norm = "raw", convert = "raw",
  batch = "raw", filter = FALSE, annotations = NULL, fasta = NULL,
  entry_type = "gene", use_original = FALSE, batch1 = "batch",
  batch2 = NULL, batch_step = 5, low_to_zero = FALSE, thresh = 2,
  min_samples = 2, p = 0.01, A = 1, k = 1, cv_min = 0.01,
  cv_max = 1000, ...)
```

**Arguments**

expt	Original expt.
transform	Transformation desired, usually log2.
norm	How to normalize the data? (raw, quant, sf, upperquartile, tmm, rle)
convert	Conversion to perform? (raw, cpm, rpkm, cp_seq_m)
batch	Batch effect removal tool to use? (limma sva fsva ruv etc)
filter	Filter out low/undesired features? (cbcb, pofa, kofa, others?)
annotations	Used for rpkm – probably not needed as this is in fData now.
fasta	Fasta file for cp_seq_m counting of oligos.
entry_type	For getting genelengths by feature type (rpkm or cp_seq_m).
use_original	Use the backup data in the expt class?
batch1	Experimental factor to extract first.
batch2	Second factor to remove (only with limma's removebatcheffect()).
batch_step	From step 1-5, when should batch correction be applied?
low_to_zero	When log transforming, change low numbers (< 0) to 0 to avoid NaN?
thresh	Used by cbcb_lowfilter().
min_samples	Also used by cbcb_lowfilter().
p	Used by genefilter's pofa().
A	Also used by genefilter's pofa().
k	Used by genefilter's kofa().
cv_min	Used by genefilter's cv().
cv_max	Also used by genefilter's cv().
...	more options

**Value**

Expt object with normalized data and the original data saved as 'original\_expressionset'

**See Also**

**genefilter limma sva edgeR DESeq2**

**Examples**

```
## Not run:
normed <- normalize_expt(exp, transform='log2', norm='rle', convert='cpm',
                        batch='raw', filter='pofa')
normed_batch <- normalize_expt(exp, transform='log2', norm='rle', convert='cpm',
                              batch='sva', filter='pofa')

## End(Not run)
```

---

orgdb_idmap	<i>Load organism annotation data (mouse/human).</i>
-------------	-----------------------------------------------------

---

**Description**

Creates a dataframe gene and transcript information for a given set of gene ids using the OrganismDbi interface.

**Usage**

```
orgdb_idmap(orgdb, gene_ids = NULL, mapto = c("ensembl"),
            keytype = "geneid")
```

**Arguments**

orgdb	OrganismDb instance.
gene_ids	Gene identifiers for retrieving annotations.
mapto	Key to map the IDs against.

**Value**

a table of gene information

**See Also**

[select](#)

**Examples**

```
## Not run:
host <- load_host_annotations(org, c("a", "b"))

## End(Not run)
```

---

parse_gene_go_terms	<i>TriTrypDB gene information table GO term parser</i>
---------------------	--------------------------------------------------------

---

**Description**

TriTrypDB gene information table GO term parser

**Usage**

```
parse_gene_go_terms(filepath, verbose = FALSE)
```

**Arguments**

filepath	Location of TriTrypDB gene information table.
verbose	Whether or not to enable verbose output.

**Value**

Returns a dataframe where each line includes a gene/GO terms pair along with some addition information about the GO term. Note that because each gene may have multiple GO terms, a single gene ID may appear on multiple lines.

**Author(s)**

Keith Hughitt

---

parse_gene_info_table	<i>TriTrypDB gene information table parser</i>
-----------------------	------------------------------------------------

---

**Description**

An example input file is the *T. brucei* Lister427 gene information table available at: [http://tritrypdb.org/common/downloads/5.0\\_TbruceiLister427Gene.txt](http://tritrypdb.org/common/downloads/5.0_TbruceiLister427Gene.txt)

**Usage**

```
parse_gene_info_table(file, verbose = FALSE)
```

**Arguments**

verbose	Whether or not to enable verbose output.
filepath	Location of TriTrypDB gene information table.

**Value**

Returns a dataframe of gene info.



**Arguments**

filepath	Location of TriTrypDB gene information table.
verbose	Whether or not to enable verbose output.

**Value**

Returns a dataframe where each line includes a gene/domain pairs.

**Author(s)**

Keith Hughitt

---

pattern\_count\_genome    *Find how many times a given pattern occurs in every gene of a genome.*

---

**Description**

There are times when knowing how many times a given string appears in a genome/CDS is helpful. This function provides that information and is primarily used by cp\_seq\_m().

**Usage**

```
pattern_count_genome(fasta, gff = NULL, pattern = "TA", type = "gene",  
  key = "locus_tag")
```

**Arguments**

fasta	Genome sequence.
gff	Gff of annotation information from which to acquire CDS (if not provided it will just query the entire genome).
pattern	What to search for? This was used for tnseq and TA is the mariner insertion point.
type	Column to use in the gff file.
key	What type of entry of the gff file to key from?

**Value**

Data frame of gene names and number of times the pattern appears/gene.

**See Also**

**Biostrings** **Rsamtools** [PDict](#) [FaFile](#)

**Examples**

```
## Not run:
num_pattern = pattern_count_genome('mgas_5005.fasta', 'mgas_5005.gff')

## End(Not run)
```

---

pca_highscores	<i>Get the highest/lowest scoring genes for every principle component.</i>
----------------	----------------------------------------------------------------------------

---

**Description**

This function uses princomp to acquire a principle component biplot for some data and extracts a dataframe of the top n genes for each component by score.

**Usage**

```
pca_highscores(df = NULL, conditions = NULL, batches = NULL, n = 20)
```

**Arguments**

df	a dataframe of (pseudo)counts
conditions	a factor or character of conditions in the experiment.
batches	a factor or character of batches in the experiment.
n	the number of genes to extract.

**Value**

a list including the princomp biplot, histogram, and tables of top/bottom n scored genes with their scores by component.

**See Also**

[princomp](#)

**Examples**

```
## Not run:
information = pca_highscores(df=df, conditions=cond, batches=bat)
information$pca_bitplot ## oo pretty

## End(Not run)
```

---

pca_information	<i>Gather information about principle components.</i>
-----------------	-------------------------------------------------------

---

## Description

Calculate some information useful for generating PCA plots. `pca_information` seeks to gather together interesting information to make principle component analyses easier, including: the results from `(fast.)svd`, a table of the  $r^2$  values, a table of the variances in the data, coordinates used to make a `pca` plot for an arbitrarily large set of PCs, correlations and `fstats` between experimental factors and the PCs, and heatmaps describing these relationships. Finally, it will provide a plot showing how much of the variance is provided by the top-n genes and (optionally) the set of all PCA plots with respect to one another. (PCx vs. PCy)

## Usage

```
pca_information(expt_data, expt_design = NULL, expt_factors = c("condition",
  "batch"), num_components = NULL, plot_pcas = FALSE)
```

## Arguments

<code>expt_data</code>	the data to analyze (usually <code>exprs(somedataset)</code> ).
<code>expt_design</code>	a dataframe describing the experimental design, containing columns with useful information like the conditions, batches, number of cells, whatever..
<code>expt_factors</code>	a character list of experimental conditions to query for $R^2$ against the <code>fast.svd</code> of the data.
<code>num_components</code>	a number of principle components to compare the design factors against. If left null, it will query the same number of components as factors asked for.
<code>plot_pcas</code>	plot the set of PCA plots for every pair of PCs queried.

## Value

a list of fun `pca` information: `svd_u/d/v`: The u/d/v parameters from `fast.svd` `rsquared_table`: A table of the `rsquared` values between each factor and principle component `pca_variance`: A table of the `pca` variances `pca_data`: Coordinates for a `pca` plot `pca_cor`: A table of the correlations between the factors and principle components `anova_fstats`: the sum of the residuals with the factor vs without (manually calculated) `anova_f`: The result from performing `anova(withfactor, withoutfactor)`, the F slot `anova_p`: The p-value calculated from the `anova()` call `anova_sums`: The RSS value from the above `anova()` call `cor_heatmap`: A heatmap from `recordPlot()` describing `pca_cor`.

## Warning

This function has gotten too damn big and needs to be split up.

## See Also

[fast.svd](#), [lm](#)

**Examples**

```
## Not run:
pca_info = pca_information(exprs(some_expt$expressionset), some_design, "all")
pca_info

## End(Not run)
```

---

pcRes	<i>Compute variance of each principal component and how they correlate with batch and cond</i>
-------	------------------------------------------------------------------------------------------------

---

**Description**

This was copy/pasted from cbcSEQ <https://github.com/kokrah/cbcSEQ/blob/master/R/explore.R>

**Usage**

```
pcRes(v, d, condition = NULL, batch = NULL)
```

**Arguments**

v	from makeSVD
d	from makeSVD
condition	factor describing experiment
batch	factor describing batch

**Value**

A dataframe containig variance, cum. variance, cond.R-sqrd, batch.R-sqrd

---

pct_all_kegg	<i>Extract the percent differentially expressed genes for all KEGG pathways.</i>
--------------	----------------------------------------------------------------------------------

---

**Description**

KEGGgraph provides some interesting functionality for mapping KEGGids and examining the pieces. This attempts to use that in order to evaluate how many 'significant' genes are in a given pathway.

**Usage**

```
pct_all_kegg(all_ids, sig_ids, organism = "dme", pathways = "all",
  pathdir = "kegg_pathways", verbose = FALSE, ...)
```



**Arguments**

all_ids	Set of all gene IDs in a given analysis.
sig_ids	Set of significant gene IDs.
organism	KEGG organism identifier.
pathways	What pathways to look at?
pathdir	Directory into which to copy downloaded pathway files.
...	Options I might pass from other functions are dropped into arglist.

**Value**

Dataframe including the filenames, percentages, nodes included, and differential nodes.

**See Also**

**KEGGgraph KEGGREST**

---

pct_kegg_diff	<i>Extract the percent differentially expressed genes in a given KEGG pathway.</i>
---------------	------------------------------------------------------------------------------------

---

**Description**

KEGGgraph provides some interesting functionality for mapping KEGGids and examining the pieces. This attempts to use that in order to evaluate how many 'significant' genes are in a given pathway.

**Usage**

```
pct_kegg_diff(all_ids, sig_ids, pathway = "00500", organism = "dme",
  pathdir = "kegg_pathways", ...)
```

**Arguments**

all_ids	Set of all gene IDs in a given analysis.
sig_ids	Set of significant gene IDs.
pathway	Numeric pathway identifier.
organism	KEGG organism identifier.
pathdir	Directory into which to copy downloaded pathway files.
...	Options I might pass from other functions are dropped into arglist.

**Value**

Percent genes/pathway deemed significant.

**See Also**

**KEGGgraph KEGGREST**

---

pkg_cleaner	<i>Packages generated by make_organismdbi(), make_orgdb(), and make_txdb() are entirely too fragile. This attempts to fix some of the common problems therein.</i>
-------------	--------------------------------------------------------------------------------------------------------------------------------------------------------------------

---

### Description

OrganismDbi instances are pretty neat, they pull together OrgDb and TxDb. With any luck, this function provides the ability to pull together all the data from the TriTrypDb, GO.db, and KEGREST in order to accomplish these peculiar tasks.

### Usage

```
pkg_cleaner(path, removal = "-like", replace = "")
```

### Arguments

path	Location for the original Db/Dbi instance.
removal	String to remove from the instance.
replace	What to replace removal with, when necessary.

### Value

A new OrgDb/TxDb/OrganismDbi

### Examples

```
## Not run:
crazytown <- make_organismdbi() ## wait a loong time

## End(Not run)
```

---

plot_batchsv	<i>Make a dotplot of known batches vs. SVs.</i>
--------------	-------------------------------------------------

---

### Description

This should make a quick df of the factors and surrogates and plot them. Maybe it should be folded into plot\_svfactor? Hmm, I think first I will write this and see if it is better.

### Usage

```
plot_batchsv(expt, sv, batch_column = "batch", factor_type = "factor")
```

**Arguments**

expt	Experiment from which to acquire the design, counts, etc.
svs	Set of surrogate variable estimations from sva/svg or batch estimates.
batch_column	Which experimental design column to use?
factor_type	This may be a factor or range, it is intended to plot a scatterplot if it is a range, a dotplot if a factor.

**Examples**

```
## Not run:
estimate_vs_snps <- plot_batchsv(start, surrogate_estimate, "snpcategory")

## End(Not run)
```

---

plot\_bcv

*Steal edgeR's plotBCV() and make it a ggplot2.*


---

**Description**

This was written primarily to understand what that function is doing in edgeR.

**Usage**

```
plot_bcv(data)
```

**Arguments**

data	A dataframe/expt/exprs with count data
------	----------------------------------------

**Value**

a plot! of the BCV a la ggplot2.

**See Also**

**edgeR** [plotBCV](#)

**Examples**

```
## Not run:
bcv <- plot_bcv(expt)
summary(bcv$data)
bcv$plot

## End(Not run)
```

---

plot\_boxplot

---

*Make a ggplot boxplot of a set of samples.*


---

## Description

Boxplots and density plots provide complementary views of data distributions. The general idea is that if the box for one sample is significantly shifted from the others, then it is likely an outlier in the same way a density plot shifted is an outlier.

## Usage

```
plot_boxplot(data, colors = NULL, names = NULL, title = NULL,
             scale = NULL, ...)
```

## Arguments

data	Expt or data frame set of samples.
colors	Color scheme, if not provided will make its own.
names	Another version of the sample names for printing.
title	A title!
scale	Whether to log scale the y-axis.
...	More parameters are more fun!

## Value

Ggplot2 boxplot of the samples. Each boxplot contains the following information: a centered line describing the median value of counts of all genes in the sample, a box around the line describing the inner-quartiles around the median (quartiles 2 and 3 for those who are counting), a vertical line above/below the box which shows 1.5x the inner quartile range (a common metric of the non-outliers), and single dots for each gene which is outside that range. A single dot is transparent.

## See Also

**ggplot2** [reshape2](#) [geom\\_boxplot](#) [melt](#) [scale\\_x\\_discrete](#)

## Examples

```
## Not run:
a_boxplot <- plot_boxplot(expt)
a_boxplot ## ooo pretty boxplot look at the lines

## End(Not run)
```

---

plot_corheat	<i>Make a heatmap.3 description of the correlation between samples.</i>
--------------	-------------------------------------------------------------------------

---

## Description

Given a set of count tables and design, this will calculate the pairwise correlations and plot them as a heatmap. It attempts to standardize the inputs and eventual output.

## Usage

```
plot_corheat(expt_data, expt_colors = NULL, expt_design = NULL,  
             method = "pearson", expt_names = NULL, batch_row = "batch",  
             title = NULL, ...)
```

## Arguments

expt_data	Dataframe, expt, or expressionset to work with.
expt_colors	Color scheme for the samples, not needed if this is an expt.
expt_design	Design matrix describing the experiment, not needed if this is an expt.
method	Correlation statistic to use. (pearson, spearman, kendall, robust).
expt_names	Alternate names to use for the samples.
batch_row	Name of the design row used for 'batch' column colors.
title	Title for the plot.
...	More options are wonderful!

## Value

Gplots heatmap describing describing how the samples are clustering vis a vis pairwise correlation.

## See Also

[hpgl\\_cor](#) [brewer.pal](#) [recordPlot](#)

## Examples

```
## corheat_plot = hpgl_corheat(expt=expt, method="robust")  
## corheat_plot
```

---

plot_density	Create a density plot, showing the distribution of each column of data.
--------------	-------------------------------------------------------------------------

---

## Description

Density plots and boxplots are cousins and provide very similar views of data distributions. Some people like one, some the other. I think they are both colorful and fun!

## Usage

```
plot_density(data, colors = NULL, sample_names = NULL,  
             position = "identity", fill = NULL, title = NULL, scale = NULL,  
             colors_by = "condition")
```

## Arguments

data	Expt, expressionset, or data frame.
colors	Color scheme to use.
sample_names	Names of the samples.
position	How to place the lines, either let them overlap (identity), or stack them.
fill	Fill the distributions? This might make the plot unreasonably colorful.
title	Title for the plot.
scale	Plot on the log scale?
colors_by	Factor for coloring the lines

## Value

Ggplot2 density plot!

## See Also

**ggplot2** [geom\\_density](#)

## Examples

```
## Not run:  
funkytown <- plot_density(data)  
  
## End(Not run)
```

---

plot_disheat	<i>Make a heatmap.3 description of the distances (euclidean by default) between samples.</i>
--------------	----------------------------------------------------------------------------------------------

---

### Description

Given a set of count tables and design, this will calculate the pairwise distances and plot them as a heatmap. It attempts to standardize the inputs and eventual output.

### Usage

```
plot_disheat(expt_data, expt_colors = NULL, expt_design = NULL,  
             method = "euclidean", expt_names = NULL, batch_row = "batch",  
             title = NULL, ...)
```

### Arguments

expt_data	Dataframe, expt, or expressionset to work with.
expt_colors	Color scheme (not needed if an expt is provided).
expt_design	Design matrix (not needed if an expt is provided).
method	Distance metric to use.
expt_names	Alternate names to use for the samples.
batch_row	Name of the design row used for 'batch' column colors.
title	Title for the plot.
...	More parameters!

### Value

a recordPlot() heatmap describing the distance between samples.

### See Also

[brewer.pal](#) [heatmap.2](#) [recordPlot](#)

### Examples

```
## Not run:  
disheat_plot = plot_disheat(expt=expt, method="euclidean")  
disheat_plot  
  
## End(Not run)
```

---

plot_dist_scatter	<i>Make a scatter plot between two sets of numbers with a cheesy distance metric and some statistics of the two sets.</i>
-------------------	---------------------------------------------------------------------------------------------------------------------------

---

## Description

The distance metric should be codified and made more intelligent. Currently it creates a dataframe of distances which are absolute distances from each axis, multiplied by each other, summed by axis, then normalized against the maximum.

## Usage

```
plot_dist_scatter(df, tooltip_data = NULL, gvis_filename = NULL, size = 2)
```

## Arguments

df	Dataframe likely containing two columns.
tooltip_data	Df of tooltip information for gvis graphs.
gvis_filename	Filename to write a fancy html graph.
size	Size of the dots.

## Value

Ggplot2 scatter plot. This plot provides a "bird's eye" view of two data sets. This plot assumes the two data structures are not correlated, and so it calculates the median/mad of each axis and uses these to calculate a stupid, home-grown distance metric away from both medians. This distance metric is used to color dots which are presumed the therefore be interesting because they are far from 'normal.' This will make a fun clicky googleVis graph if requested.

## See Also

**ggplot2** [plot\\_gvis\\_scatter](#) [geom\\_point](#) [plot\\_linear\\_scatter](#)

## Examples

```
## Not run:
dist_scatter(lotsofnumbers_intwo_columns, tooltip_data=tooltip_dataframe,
             gvis_filename="html/fun_scatterplot.html")

## End(Not run)
```



---

plot_essentiality	<i>Plot the essentiality of a library as per DeJesus et al.</i>
-------------------	-----------------------------------------------------------------

---

**Description**

This provides a plot of the essentiality metrics 'zbar' with respect to gene.

**Usage**

```
plot_essentiality(file)
```

**Arguments**

file	a file created using the perl script 'essentiality_tas.pl'
------	------------------------------------------------------------

**Value**

A couple of plots

---

plot_goseq_pval	<i>Make a pvalue plot from goseq data.</i>
-----------------	--------------------------------------------

---

**Description**

With minor changes, it is possible to push the goseq results into a clusterProfiler-ish pvalue plot. This handles those changes and returns the ggplot results.

**Usage**

```
plot_goseq_pval(goterms, wrapped_width = 20, cutoff = 0.1, n = 10,  
  mincat = 10, level = NULL)
```

**Arguments**

goterms	Some data from goseq!
wrapped_width	Number of characters before wrapping to help legibility.
cutoff	Pvalue cutoff for the plot.
n	How many groups to include?
mincat	Minimum size of the category for inclusion.
level	Levels of the ontology tree to use.

**Value**

Plots!

**See Also**

[goseq](#) [clusterProfiler](#) [plot\\_ontpval](#)

---

plot_gostats_pval	<i>Make a pvalue plot similar to that from clusterprofiler from gostats data.</i>
-------------------	-----------------------------------------------------------------------------------

---

**Description**

clusterprofiler provides beautiful plots describing significantly overrepresented categories. This function attempts to expand the repertoire of data available to them to include data from gostats. The pval\_plot function upon which this is based now has a bunch of new helpers now that I understand how the ontology trees work better, this should take advantage of that, but currently does not.

**Usage**

```
plot_gostats_pval(gs_result, wrapped_width = 20, cutoff = 0.1, n = 12,  
  group_minsize = 5)
```

**Arguments**

gs_result	Ontology search results.
wrapped_width	Make the text large enough to read.
cutoff	What is the maximum pvalue allowed?
n	How many groups to include in the plot?
group_minsize	Minimum group size before inclusion.

**Value**

Plots!

**See Also**

[clusterProfiler](#) [plot\\_ontpval](#)

---

plot_gprofiler_pval	<i>Make a pvalue plot from gprofiler data.</i>
---------------------	------------------------------------------------

---

### Description

The p-value plots from clusterProfiler are pretty, this sets the gprofiler data into a format suitable for plotting in that fashion and returns the resulting plots of significant ontologies.

### Usage

```
plot_gprofiler_pval(gp_result, wrapped_width = 20, cutoff = 0.1, n = 12,  
  group_minsize = 5, ...)
```

### Arguments

gp_result	Some data from gProfiler.
wrapped_width	Maximum width of the text names.
cutoff	P-value cutoff for the plots.
n	Maximum number of ontologies to include.
group_minsize	Minimum ontology group size to include.
...	Options I might pass from other functions are dropped into arglist.

### Value

List of MF/BP/CC pvalue plots.

### See Also

**topgo** clusterProfiler

---

plot_gvis_ma	<i>Make an html version of an MA plot: M(log ratio of conditions) / A(mean average).</i>
--------------	------------------------------------------------------------------------------------------

---

### Description

A fun snippet from wikipedia: "In many microarray gene expression experiments, an underlying assumption is that most of the genes would not see any change in their expression therefore the majority of the points on the y-axis (M) would be located at 0, since Log(1) is 0. If this is not the case, then a normalization method such as LOESS should be applied to the data before statistical analysis. If the median line is not straight, the data should be normalized.

**Usage**

```
plot_gvis_ma(counts, degenes, tooltip_data = NULL,
             filename = "html/gvis_ma_plot.html", base_url = "", ...)
```

**Arguments**

counts	Df of counts which have been normalized counts by sample-type, which is to say the output from voom/voomMod/hppl_voom().
degenes	Df from toptable or its friends containing p-values.
tooltip_data	Df of tooltip information (gene names, etc).
filename	Filename to write a fancy html graph.
base_url	String with a basename used for generating URLs for clicking dots on the graph.
...	more options are more options!

**Value**

NULL, but along the way an html file is generated which contains a googleVis MA plot. See plot\_ma() for details.

**See Also**

[plot\\_ma](#)

**Examples**

```
## Not run:
plot_gvis_ma(voomed_data, toptable_data, filename="html/fun_ma_plot.html",
             base_url="http://yeastgenome.org/accession?")

## End(Not run)
```

---

plot_gvis_scatter	<i>Make an html version of a scatter plot.</i>
-------------------	------------------------------------------------

---

**Description**

Given an arbitrary scatter plot, we can make it pretty and javascript-tacular using this function.

**Usage**

```
plot_gvis_scatter(df, tooltip_data = NULL,
                 filename = "html/gvis_scatter.html", base_url = "", trendline = NULL)
```

**Arguments**

df	Df of two columns to compare.
tooltip_data	Df of tooltip information for gvis graphs.
filename	Filename to write a fancy html graph.
base_url	Url to send click events which will be suffixed with the gene name.
trendline	Add a trendline?

**Value**

NULL, but along the way an html file is generated which contains a googleVis scatter plot. See `plot_scatter()` for details.

**See Also**

[gvisScatterChart](#)

**Examples**

```
## Not run:
  gvis_scatter(a_dataframe_twocolumns, filename="html/fun_scatter_plot.html",
              base_url="http://yeastgenome.org/accession?")

## End(Not run)
```

---

plot_gvis_volcano	<i>Make an html version of an volcano plot.</i>
-------------------	-------------------------------------------------

---

**Description**

Volcano plots provide some visual clues regarding the success of a given contrast. For our data, it has the  $-\log_{10}(\text{pvalue})$  on the y-axis and fold-change on the x. Here is a neat snippet from wikipedia describing them generally: "The concept of volcano plot can be generalized to other applications, where the x-axis is related to a measure of the strength of a statistical signal, and y-axis is related to a measure of the statistical significance of the signal."

**Usage**

```
plot_gvis_volcano(toptable_data, fc_cutoff = 0.8, p_cutoff = 0.05,
  tooltip_data = NULL, filename = "html/gvis_vol_plot.html",
  base_url = "", ...)
```

**Arguments**

toptable_data	Df of toptable() data.
fc_cutoff	Fold change cutoff.
p_cutoff	Maximum p value to allow.
tooltip_data	Df of tooltip information.
filename	Filename to write a fancy html graph.
base_url	String with a basename used for generating URLs for clicking dots on the graph.
...	more options

**Value**

NULL, but along the way an html file is generated which contains a googleVis volcano plot.

**See Also**

[plot\\_volcano](#)

**Examples**

```
## Not run:
plot_gvis_volcano(voomed_data, toptable_data, filename="html/fun_ma_plot.html",
                  base_url="http://yeastgenome.org/accession?")

## End(Not run)
```

---

plot_heatmap	<i>Make a heatmap.3 plot, does the work for plot_disheat and plot_corheat.</i>
--------------	--------------------------------------------------------------------------------

---

**Description**

This does what is says on the tin. Sets the colors for correlation or distance heatmaps, handles the calculation of the relevant metrics, and plots the heatmap.

**Usage**

```
plot_heatmap(expt_data, expt_colors = NULL, expt_design = NULL,
             method = "pearson", expt_names = NULL, type = "correlation",
             batch_row = "batch", title = NULL, ...)
```

**Arguments**

expt_data	Dataframe, expt, or expressionset to work with.
expt_colors	Color scheme for the samples.
expt_design	Design matrix describing the experiment vis a vis conditions and batches.
method	Distance or correlation metric to use.
expt_names	Alternate names to use for the samples.
type	Defines the use of correlation, distance, or sample heatmap.
batch_row	Name of the design row used for 'batch' column colors.
title	Title for the plot.
...	I like ellipses!

**Value**

a recordPlot() heatmap describing the distance between samples.

**See Also**

[brewer.pal recordPlot](#)

---

plot_histogram	<i>Make a pretty histogram of something.</i>
----------------	----------------------------------------------

---

**Description**

A shortcut to make a ggplot2 histogram which makes an attempt to set reasonable bin widths and set the scale to log if that seems a good idea.

**Usage**

```
plot_histogram(df, binwidth = NULL, log = FALSE, bins = 500,
  fillcolor = "darkgrey", color = "black")
```

**Arguments**

df	Dataframe of lots of pretty numbers.
binwidth	Width of the bins for the histogram.
log	Replot on the log scale?
bins	Number of bins for the histogram.
fillcolor	Change the fill colors of the plotted elements?
color	Change the color of the lines of the plotted elements?

**Value**

Ggplot histogram.

**See Also**

[geom\\_histogram](#) [geom\\_density](#)

**Examples**

```
## Not run:
kittytime = plot_histogram(df)

## End(Not run)
```

---

plot_libsize	<i>Make a ggplot graph of library sizes.</i>
--------------	----------------------------------------------

---

**Description**

It is often useful to have a quick view of which samples have more/fewer reads. This does that and maintains one's favorite color scheme and tries to make it pretty!

**Usage**

```
plot_libsize(data, colors = NULL, names = NULL, text = TRUE,
             title = NULL, yscale = NULL, ...)
```

**Arguments**

data	Expt, dataframe, or expressionset of samples.
colors	Color scheme if the data is not an expt.
names	Alternate names for the x-axis.
text	Add the numeric values inside the top of the bars of the plot?
title	Title for the plot.
yscale	Whether or not to log10 the y-axis.
...	More parameters for your good time!

**Value**

a ggplot2 bar plot of every sample's size

**See Also**

[geom\\_bar](#) [geom\\_text](#) [prettyNum](#) [scale\\_y\\_log10](#)

**Examples**

```
## Not run:
libsize_plot = plot_libsize(expt=expt)
libsize_plot ## ooo pretty bargraph

## End(Not run)
```



---

plot_linear_scatter	<i>Make a scatter plot between two groups with a linear model superimposed and some supporting statistics.</i>
---------------------	----------------------------------------------------------------------------------------------------------------

---

## Description

Make a scatter plot between two groups with a linear model superimposed and some supporting statistics.

## Usage

```
plot_linear_scatter(df, tooltip_data = NULL, gvis_filename = NULL,
  cormethod = "pearson", size = 2, loess = FALSE, identity = FALSE,
  gvis_trendline = NULL, first = NULL, second = NULL, base_url = NULL,
  pretty_colors = TRUE, color_high = NULL, color_low = NULL, ...)
```

## Arguments

df	Dataframe likely containing two columns.
tooltip_data	Df of tooltip information for gvis graphs.
gvis_filename	Filename to write a fancy html graph.
cormethod	What type of correlation to check?
size	Size of the dots on the plot.
loess	Add a loess estimation?
identity	Add the identity line?
gvis_trendline	Add a trendline to the gvis plot? There are a couple possible types, I think linear is the most common.
first	First column to plot.
second	Second column to plot.
base_url	Base url to add to the plot.
pretty_colors	Colors!
color_high	Chosen color for points significantly above the mean.
color_low	Chosen color for points significantly below the mean.
...	Extra args likely used for choosing significant genes.

## Value

List including a ggplot2 scatter plot and some histograms. This plot provides a "bird's eye" view of two data sets. This plot assumes a (potential) linear correlation between the data, so it calculates the correlation between them. It then calculates and plots a robust linear model of the data using an 'SMDM' estimator (which I don't remember how to describe, just that the document I was reading said it is good). The median/mad of each axis is calculated and plotted as well. The distance from the linear model is finally used to color the dots on the plot. Histograms of each axis are plotted separately and then together under a single cdf to allow tests of distribution similarity. This will make a fun clicky googleVis graph if requested.

See Also

[lmRob weights plot\\_histogram](#)

Examples

```
## Not run:
plot_linear_scatter(lotsofnnumbers_intwo_columns, tooltip_data=tooltip_dataframe,
                    gvis_filename="html/fun_scatterplot.html")

## End(Not run)
```

---

plot_ma	<i>Make a pretty MA plot from the output of voom/limma/eBayes/toptable.</i>
---------	-----------------------------------------------------------------------------

---

Description

Make a pretty MA plot from the output of voom/limma/eBayes/toptable.

Usage

```
plot_ma(counts, de_genes, pval_cutoff = 0.05, alpha = 0.4,
        logfc_cutoff = 1, pval = "adjpval", size = 2, tooltip_data = NULL,
        gvis_filename = NULL, ...)
```

Arguments

counts	Df of linear-modelling, normalized counts by sample-type, which is to say the output from voom/voomMod/hppl_voom().
de_genes	Df from toptable or its friends containing p-values.
pval_cutoff	Cutoff defining significant from not.
alpha	How transparent to make the dots.
logfc_cutoff	Fold change cutoff.
pval	Name of the pvalue column to use for cutoffs.
size	How big are the dots?
tooltip_data	Df of tooltip information for gvis.
gvis_filename	Filename to write a fancy html graph.
...	More options for you!

Value

Ggplot2 MA scatter plot. This is defined as the rowmeans of the normalized counts by type across all sample types on the x-axis, and the log fold change between conditions on the y-axis. Dots are colored depending on if they are 'significant.' This will make a fun clicky googleVis graph if requested.

**See Also**

[plot\\_gvis\\_ma](#) [toptable](#) [voom](#) [hpgl\\_voom](#) [lmFit](#) [makeContrasts](#) [contrasts.fit](#)

**Examples**

```
## Not run:
plot_ma(voomed_data, toptable_data, gvis_filename="html/fun_ma_plot.html")
## Currently this assumes that a variant of toptable was used which
## gives adjusted p-values. This is not always the case and I should
## check for that, but I have not yet.

## End(Not run)
```

---

plot\_ma\_de

---

*Make a pretty MA plot from a DE tool (limma/deseq/edger/basic)*


---

**Description**

Make a pretty MA plot from a DE tool (limma/deseq/edger/basic)

**Usage**

```
plot_ma_de(table, expr_col = "logCPM", fc_col = "logFC", p_col = "qvalue",
  pval_cutoff = 0.05, alpha = 0.4, logfc_cutoff = 1, size = 2,
  tooltip_data = NULL, gvis_filename = NULL, ...)
```

**Arguments**

table	Df of linear-modelling, normalized counts by sample-type,
expr_col	Column showing the average expression across genes.
fc_col	Column showing the logFC for each gene.
p_col	Column containing the relevant p-values.
alpha	How transparent to make the dots.
logfc_cutoff	Fold change cutoff.
size	How big are the dots?
tooltip_data	Df of tooltip information for gvis.
gvis_filename	Filename to write a fancy html graph.
...	More options for you!
pval	Name of the pvalue column to use for cutoffs.

**Value**

Ggplot2 MA scatter plot. This is defined as the rowmeans of the normalized counts by type across all sample types on the x-axis, and the log fold change between conditions on the y-axis. Dots are colored depending on if they are 'significant.' This will make a fun clicky googleVis graph if requested.

**See Also**

[plot\\_gvis\\_ma](#) [toptable](#) [voom](#) [hpgl\\_voom](#) [lmFit](#) [makeContrasts](#) [contrasts.fit](#)

**Examples**

```
## Not run:
plot_ma(voomed_data, toptable_data, gvis_filename="html/fun_ma_plot.html")
## Currently this assumes that a variant of toptable was used which
## gives adjusted p-values. This is not always the case and I should
## check for that, but I have not yet.

## End(Not run)
```

---

plot_multihistogram	<i>Make a pretty histogram of multiple datasets.</i>
---------------------	------------------------------------------------------

---

**Description**

If there are multiple data sets, it might be useful to plot them on a histogram together and look at the t.test results between distributions.

**Usage**

```
plot_multihistogram(data, log = FALSE, binwidth = NULL, bins = NULL)
```

**Arguments**

data	Dataframe of lots of pretty numbers, this also accepts lists.
log	Plot the data on the log scale?
binwidth	Set a static bin width with an unknown # of bins? If neither of these are provided, then bins is set to 500, if both are provided, then bins wins.
bins	Set a static # of bins of an unknown width?

**Value**

List of the ggplot histogram and some statistics describing the distributions.

**See Also**

[pairwise.t.test](#) [ddply](#)

**Examples**

```
## Not run:
kittytime = plot_multihistogram(df)

## End(Not run)
```

---

plot_multiplot	<i>Make a grid of plots.</i>
----------------	------------------------------

---

**Description**

Make a grid of plots.

**Usage**

```
plot_multiplot(plots, file, cols = NULL, layout = NULL)
```

**Arguments**

plots	a list of plots
file	a file to write to
cols	the number of columns in the grid
layout	set the layout specifically

**Value**

a multiplot!

---

plot_nonzero	<i>Make a ggplot graph of the number of non-zero genes by sample.</i>
--------------	-----------------------------------------------------------------------

---

**Description**

This puts the number of genes with > 0 hits on the y-axis and CPM on the x-axis. Made by Ramzi Temanni <temanni at umd dot edu>.

**Usage**

```
plot_nonzero(data, design = NULL, colors = NULL, labels = NULL,  
             title = NULL, ...)
```

**Arguments**

data	Expt, expressionset, or dataframe.
design	Eesign matrix.
colors	Color scheme.
labels	How do you want to label the graph? 'fancy' will use directlabels() to try to match the labels with the positions without overlapping anything else will just stick them on a 45' offset next to the graphed point.
title	Add a title?
...	rawr!

**Value**

a ggplot2 plot of the number of non-zero genes with respect to each library’s CPM.

**See Also**

[geom\\_point](#) [geom\\_dl](#)

**Examples**

```
## Not run:
nonzero_plot = plot_nonzero(expt=expt)
nonzero_plot  ## ooo pretty

## End(Not run)
```

---

plot_num_siggenes	<i>Given a DE table with fold changes and p-values, show how ‘significant’ changes with changing cutoffs.</i>
-------------------	---------------------------------------------------------------------------------------------------------------

---

**Description**

Sometimes one might want to know how many genes are deemed significant while shifting the bars which define significant. This provides that metrics as a set of tables of numbers of significant up/down genes when p-value is held constant, as well as number when fold-change is held constant.

**Usage**

```
plot_num_siggenes(table, p_column = "limma_adjp", fc_column = "limma_logfc",
  bins = 100, constant_p = 0.05, constant_fc = 0)
```

**Arguments**

table	DE table to examine.
p_column	Column in the DE table defining the changing p-value cutoff.
fc_column	Column in the DE table defining the changing +/- log fold change.
bins	Number of incremental changes in p-value/FC to examine.
constant_p	When plotting changing FC, where should the p-value be held?
constant_fc	When plotting changing p, where should the FC be held?

**Value**

Plots and dataframes describing the changing definition of ‘significant.’

---

plot_ontpval	<i>Make a pvalue plot from a df of IDs, scores, and p-values.</i>
--------------	-------------------------------------------------------------------

---

**Description**

This function seeks to make generating pretty pvalue plots as shown by clusterprofiler easier.

**Usage**

```
plot_ontpval(df, ontology = "MF")
```

**Arguments**

df	Some data from topgo/goseq/clusterprofiler.
ontology	Ontology to plot (MF,BP,CC).

**Value**

Ggplot2 plot of pvalues vs. ontology.

**See Also**

[goseq](#) [ggplot2](#)

---

plot_pairwise_ma	<i>Plot all pairwise MA plots in an experiment.</i>
------------------	-----------------------------------------------------

---

**Description**

Use affy's ma.plot() on every pair of columns in a data set to help diagnose problematic samples.

**Usage**

```
plot_pairwise_ma(data, log = NULL, ...)
```

**Arguments**

data	Expt expressionset or data frame.
log	Is the data in log format?
...	Options are good and passed to arglist().

**Value**

List of affy::maplots

**See Also**[ma.plot](#)**Examples**

```
## Not run:
ma_plots = plot_pairwise_ma(expt=some_expt)

## End(Not run)
```

plot\_pca

*Make a ggplot PCA plot describing the samples' clustering.***Description**

Make a ggplot PCA plot describing the samples' clustering.

**Usage**

```
plot_pca(data, design = NULL, plot_colors = NULL, plot_labels = NULL,
         plot_title = NULL, plot_size = 5, size_column = NULL, ...)
```

**Arguments**

data	an expt set of samples.
design	a design matrix and.
plot_colors	a color scheme.
plot_labels	add labels? Also, what type? FALSE, "default", or "fancy".
plot_title	a title for the plot.
plot_size	size for the glyphs on the plot.
size_column	use an experimental factor to size the glyphs of the plot
...	arglist from elipsis!

**Value**

a list containing the following: `pca` = the result of `fast.svd()` `plot` = `ggplot2` `pca_plot` describing the principle component analysis of the samples. `table` = a table of the PCA plot data `res` = a table of the PCA `res` data `variance` = a table of the PCA plot variance This makes use of `cbcbSEQ` and prints the table of variance by component.

**See Also**[makeSVD](#), [geom\\_dl](#) [plot\\_pcs](#)



**Examples**

```
## Not run:
pca_plot = plot_pca(expt=expt)
pca_plot

## End(Not run)
```

---

plot_pcfactor	<i>make a dotplot of some categorised factors and a set of principle components.</i>
---------------	--------------------------------------------------------------------------------------

---

**Description**

This should make a quick df of the factors and PCs and plot them.

**Usage**

```
plot_pcfactor(pc_df, expt, exp_factor = "condition", component = "PC1")
```

**Arguments**

pc_df	Df of principle components.
expt	Expt containing counts, metadata, etc.
exp_factor	Experimental factor to compare against.
component	Which principal component to compare against?

**Examples**

```
## Not run:
estimate_vs_pcs <- plot_pcfactor(pcs, times)

## End(Not run)
```

---

plot_pcs	<i>A quick and dirty PCA plotter of arbitrary components against one another.</i>
----------	-----------------------------------------------------------------------------------

---

**Description**

A quick and dirty PCA plotter of arbitrary components against one another.

**Usage**

```
plot_pcs(pca_data, first = "PC1", second = "PC2", variances = NULL,
  design = NULL, plot_title = NULL, plot_labels = NULL, plot_size = 5,
  size_column = NULL, ...)
```

**Arguments**

pca_data	a dataframe of principle components PC1 .. PCN with any other arbitrary information.
first	principle component PCx to put on the x axis.
second	principle component PCy to put on the y axis.
variances	a list of the percent variance explained by each component.
design	the experimental design with condition batch factors.
plot_title	a title for the plot.
plot_labels	a parameter for the labels on the plot.
plot_size	The size of the dots on the plot
size_column	an experimental factor to use for sizing the glyphs
...	extra arguments dropped into arglist

**Value**

a ggplot2 PCA plot

**See Also**

[ggplot2](#) [geom\\_dl](#)

**Examples**

```
## Not run:
pca_plot = plot_pcs(pca_data, first="PC2", second="PC4", design=expt$design)

## End(Not run)
```

---

plot_qq_all	<i>Quantile/quantile comparison of the mean of all samples vs. each sample.</i>
-------------	---------------------------------------------------------------------------------

---

**Description**

This allows one to visualize all individual data columns against the mean of all columns of data in order to see if any one is significantly different than the cloud.

**Usage**

```
plot_qq_all(data, labels = "short")
```

**Arguments**

data	Expressionset, expt, or dataframe of samples.
labels	What kind of labels to print?

**Value**

List containing: logs = a recordPlot() of the pairwise log qq plots. ratios = a recordPlot() of the pairwise ratio qq plots. means = a table of the median values of all the summaries of the qq plots.

---

plot_qq_all_pairwise	<i>Perform qq plots of every column against every other column of a dataset.</i>
----------------------	----------------------------------------------------------------------------------

---

**Description**

This function is stupid, don't use it. It makes more sense to just use plot\_qq, however I am not quite read to delete this function yet.

**Usage**

```
plot_qq_all_pairwise(data)
```

**Arguments**

data	Dataframe to perform pairwise qqplots with.
------	---------------------------------------------

**Value**

List containing the recordPlot() output of the ratios, logs, and means among samples.

---

plot_qq_plot	<i>Perform a qqplot between two columns of a matrix.</i>
--------------	----------------------------------------------------------

---

**Description**

Given two columns of data, how well do the distributions match one another? The answer to that question may be visualized through a qq plot!

**Usage**

```
plot_qq_plot(data, x = 1, y = 2, labels = TRUE)
```

**Arguments**

data	Data frame/expt/expressionset.
x	First column to compare.
y	Second column to compare.
labels	Include the labels?

**Value**

a list of the logs, ratios, and mean between the plots as ggplots.

---

plot_sample_heatmap	<i>Make a heatmap.3 description of the similarity of the genes among samples.</i>
---------------------	-----------------------------------------------------------------------------------

---

### Description

Sometimes you just want to see how the genes of an experiment are related to each other. This can handle that. These heatmap functions should probably be replaced with neatmaps or heatplus or whatever it is, as the annotation dataframes in them are pretty awesome.

### Usage

```
plot_sample_heatmap(data, colors = NULL, design = NULL, names = NULL,
  title = NULL, Rowv = TRUE, ...)
```

### Arguments

data	Expt/expressionset/dataframe set of samples.
colors	Color scheme of the samples (not needed if input is an expt).
design	Design matrix describing the experiment (gotten for free if an expt).
names	Alternate samples names.
title	Title of the plot!
Rowv	Reorder the rows by expression?
...	More parameters for a good time!

### Value

a recordPlot() heatmap describing the samples.

### See Also

[brewer.pal](#) [recordPlot](#)

---

plot_scatter	<i>Make a pretty scatter plot between two sets of numbers.</i>
--------------	----------------------------------------------------------------

---

### Description

This function tries to supplement a normal scatterplot with some information describing the relationship between the columns of data plotted.

### Usage

```
plot_scatter(df, tooltip_data = NULL, color = "black",
  gvis_filename = NULL, size = 2)
```

**Arguments**

df	Dataframe likely containing two columns.
tooltip_data	Df of tooltip information for gvis.
color	Color of the dots on the graph.
gvis_filename	Filename to write a fancy html graph.
size	Size of the dots on the graph.

**Value**

Ggplot2 scatter plot.

**See Also**

[plot\\_gvis\\_scatter](#) [geom\\_point](#) [plot\\_linear\\_scatter](#)

**Examples**

```
## Not run:
plot_scatter(lotsofnnumbers_intwo_columns, tooltip_data=tooltip_dataframe,
             gvis_filename="html/fun_scatterplot.html")

## End(Not run)
```

---

plot_sm	<i>Make an R plot of the standard median correlation or distance among samples.</i>
---------	-------------------------------------------------------------------------------------

---

**Description**

This was written by a mix of Kwame Okrah <kokrah at gmail dot com>, Laura Dillon <dillonl at umd dot edu>, and Hector Corrada Bravo <hcorrada at umd dot edu> I reimplemented it using ggplot2 and tried to make it a little more flexible. The general idea is to take the pairwise correlations/distances of the samples, then take the medians, and plot them.

**Usage**

```
plot_sm(data, colors = NULL, method = "pearson", names = NULL,
        title = NULL, ...)
```

**Arguments**

data	Expt, expressionset, or data frame.
colors	Color scheme if data is not an expt.
method	Correlation or distance method to use.
names	Use pretty names for the samples?
title	Title for the graph.
...	More parameters to make you happy!

**Value**

ggplot of the standard median something among the samples. This will also write to an open device. The resulting plot measures the median correlation of each sample among its peers. It notes 1.5\* the interquartile range among the samples and makes a horizontal line at that correlation coefficient. Any sample which falls below this line is considered for removal because it is much less similar to all of its peers.

**See Also**

[hpgl\\_cor](#) [rowMedians](#) [quantile](#) [diff](#) [recordPlot](#)

**Examples**

```
## Not run:
  smc_plot = hpgl_smc(expt=expt)

## End(Not run)
```

---

plot\_spirograph

*Make spirographs!*

---

**Description**

Taken (with modifications) from: <http://menugget.blogspot.com/2012/12/spirograph-with-r.html#more>  
 A positive value for 'B' will result in a epitrochoid, while a negative value will result in a hypotrochoid.

**Usage**

```
plot_spirograph(radius_a = 1, radius_b = -4, dist_bc = -2,
  revolutions = 158, increments = 3160, center_a = list(x = 0, y = 0))
```

**Arguments**

radius_a	The radius of the primary circle.
radius_b	The radius of the circle travelling around a.
dist_bc	A point relative to the center of 'b' which rotates with the turning of 'b'.
revolutions	How many revolutions to perform in the plot
increments	The number of radial increments to be calculated per revolution
center_a	The position of the center of 'a'.

**Value**

something which I don't yet know.

---

plot_svfactor	<i>Make a dotplot of some categorised factors and a set of SVs (for other factors).</i>
---------------	-----------------------------------------------------------------------------------------

---

### Description

This should make a quick df of the factors and surrogates and plot them.

### Usage

```
plot_svfactor(expt, svest, chosen_factor = "snpcategory",
              factor_type = "factor")
```

### Arguments

expt	Experiment from which to acquire the design, counts, etc.
svest	Set of surrogate variable estimations from sva/svg or batch estimates.
chosen_factor	Factor to compare against.
factor_type	This may be a factor or range, it is intended to plot a scatterplot if it is a range, a dotplot if a factor.

### Examples

```
## Not run:
estimate_vs_snps <- plot_svfactor(start, surrogate_estimate, "snpcategory")

## End(Not run)
```

---

plot_topgo_densities	<i>Plot the density of categories vs. the possibilities of all categories.</i>
----------------------	--------------------------------------------------------------------------------

---

### Description

This can make a large number of plots.

### Usage

```
plot_topgo_densities(godata, table)
```

### Arguments

godata	Result from topgo.
table	Table of genes.

---

plot_topgo_pval	<i>Make a pvalue plot from topgo data.</i>
-----------------	--------------------------------------------

---

### Description

The p-value plots from clusterProfiler are pretty, this sets the topgo data into a format suitable for plotting in that fashion and returns the resulting plots of significant ontologies.

### Usage

```
plot_topgo_pval(topgo, wrapped_width = 20, cutoff = 0.1, n = 12,
  type = "fisher")
```

### Arguments

topgo	Some data from topgo!
wrapped_width	Maximum width of the text names.
cutoff	P-value cutoff for the plots.
n	Maximum number of ontologies to include.
type	Type of score to use.

### Value

List of MF/BP/CC pvalue plots.

### See Also

**topgo** clusterProfiler

---

plot_volcano	<i>Make a pretty Volcano plot!</i>
--------------	------------------------------------

---

### Description

Volcano plots and MA plots provide quick and easy methods to view the set of (in)significantly differentially expressed genes. In the case of a volcano plot, it places the  $-\log_{10}$  of the p-value estimate on the y-axis and the fold-change between conditions on the x-axis. Here is a neat snippet from wikipedia: "The concept of volcano plot can be generalized to other applications, where the x-axis is related to a measure of the strength of a statistical signal, and y-axis is related to a measure of the statistical significance of the signal."

### Usage

```
plot_volcano(toptable_data, tooltip_data = NULL, gvis_filename = NULL,
  fc_cutoff = 0.8, p_cutoff = 0.05, size = 2, alpha = 0.6,
  xaxis_column = "logFC", yaxis_column = "P.Value", modify_y = TRUE, ...)
```



**Arguments**

<code>toptable_data</code>	Dataframe from limma's toptable which includes log(fold change) and an adjusted p-value.
<code>tooltip_data</code>	Df of tooltip information for gvis.
<code>gvis_filename</code>	Filename to write a fancy html graph.
<code>fc_cutoff</code>	Cutoff defining the minimum/maximum fold change for interesting. This is log, so I went with +/- 0.8 mostly arbitrarily as the default.
<code>p_cutoff</code>	Cutoff defining significant from not.
<code>size</code>	How big are the dots?
<code>alpha</code>	How transparent to make the dots.
<code>xaxis_column</code>	Column from the data to use on the x axis (logFC)
<code>yaxis_column</code>	Column from the data to use on the y axis (p-value)
<code>...</code>	I love parameters!

**Value**

Ggplot2 volcano scatter plot. This is defined as the  $-\log_{10}(\text{p-value})$  with respect to log(fold change). The cutoff values are delineated with lines and mark the boundaries between 'significant' and not. This will make a fun clicky googleVis graph if requested.

**See Also**

[plot\\_gvis\\_ma toptable voom hpgl\\_voom lmFit makeContrasts contrasts.fit](#)

**Examples**

```
## Not run:
  plot_volcano(toptable_data, gvis_filename="html/fun_ma_plot.html")
## Currently this assumes that a variant of toptable was used which
## gives adjusted p-values. This is not always the case and I should
## check for that, but I have not yet.

## End(Not run)
```

---

pp

*png() shortcut*

---

**Description**

I hate remembering my options for png()

**Usage**

```
pp(file)
```

**Arguments**

file                    a filename to write

**Value**

a png with height=width=9 inches and a high resolution

---

print_ups_downs	<i>Reprint the output from extract_significant_genes().</i>
-----------------	-------------------------------------------------------------

---

**Description**

I found myself needing to reprint these excel sheets because I added some new information. This shortcuts that process for me.

**Usage**

```
print_ups_downs(upsdowns, wb = NULL, excel = "excel/significant_genes.xlsx",
  csv = NULL, according = "limma", summary_count = 1)
```

**Arguments**

upsdowns              Output from extract\_significant\_genes().  
wb                    Workbook object to use for writing, or start a new one.  
excel                  Filename for writing the data.  
according            Use limma, deseq, or edger for defining 'significant'.  
summary\_count        For spacing sequential tables one after another.

**Value**

Return from write\_xls.

**See Also**

[combine\\_de\\_tables](#)

---

read_metadata	<i>Given a table of meta data, read it in for use by create_expt().</i>
---------------	-------------------------------------------------------------------------

---

**Description**

Reads an experimental design in a few different formats in preparation for creating an expt.

**Usage**

```
read_metadata(file, ...)
```

**Arguments**

file	Csv/xls file to read.
...	Arguments for arglist, used by sep, header and similar read.csv/read.table parameters.

**Value**

Df of metadata.

---

require.auto	<i>Automatic loading and/or installing of packages.</i>
--------------	---------------------------------------------------------

---

**Description**

Load a library, install it first if necessary.

**Usage**

```
require.auto(lib, update = FALSE)
```

**Arguments**

lib	String name of a library to check/install.
update	Update packages?

**Details**

This was taken from: <http://sbamin.com/2012/11/05/tips-for-working-in-r-automatically-install-missing-package/>

**Value**

0 or 1, whether a package was installed or not.

**See Also**

[biocLite install.packages](#)

**Examples**

```
## Not run:  
require.auto("ggplot2")  
  
## End(Not run)
```

---

saveme

*Make a backup rdata file for future reference*

---

**Description**

I often use R over a sshfs connection, sometimes with significant latency, and I want to be able to save/load my R sessions relatively quickly. Thus this function uses pxz to compress the R session maximally and relatively fast. This assumes you have pxz installed and  $\geq 4$  CPUs.

**Usage**

```
saveme(directory = "savefiles", backups = 4, filename = "Rdata.rda.xz")
```

**Arguments**

directory	Directory to save the Rdata file.
backups	How many revisions?

**Value**

Command string used to save the global environment.

**See Also**

[save pipe](#)

**Examples**

```
## Not run:  
## saveme()  
  
## End(Not run)
```

---

semantic\_copynumber\_filter

*Remove multicopy genes from up/down gene expression lists.*


---

### Description

In our parasite data, there are a few gene types which are consistently obnoxious. Multi-gene families primarily where the coding sequences are divergent, but the UTRs nearly identical. For these genes, our sequence based removal methods fail and so this just excludes them by name.

### Usage

```
semantic_copynumber_filter(de_list, max_copies = 2, semantic = c("mucin",
  "sialidase", "RHS", "MASP", "DGF"), semantic_column = "1.tooltip")
```

### Arguments

de_list	List of sets of genes deemed significantly up/down with a column expressing approximate count numbers.
max_copies	Keep only those genes with <= n putative copies.
semantic	Set of strings with gene names to exclude.
semantic_column	Column in the DE table used to find the semantic strings for removal.

### Value

Smaller list of up/down genes.

---

sequence\_attributes     *Gather some simple sequence attributes.*


---

### Description

This extends the logic of the pattern searching in pattern\_count\_genome() to search on some other attributes.

### Usage

```
sequence_attributes(fasta, gff = NULL, type = "gene", key = "locus_tag")
```

### Arguments

fasta	Genome encoded as a fasta file.
gff	Optional gff of annotations (if not provided it will just ask the whole genome).
type	Column of the gff file to use.
key	What type of entry of the gff file to key from?

**Value**

List of data frames containing gc/at/gt/ac contents.

**See Also**

**Biostrings** **Rsamtools** [letterFrequency](#) [FaFile](#)

**Examples**

```
## Not run:
num_pattern = sequence_attributes('mgas_5005.fasta', 'mgas_5005.gff')

## End(Not run)
```

---

set_expt_batch	<i>Change the batches of an expt.</i>
----------------	---------------------------------------

---

**Description**

When exploring differential analyses, it might be useful to play with the conditions/batches of the experiment. Use this to make that easier.

**Usage**

```
set_expt_batch(expt, fact, ids = NULL, ...)
```

**Arguments**

- expt            Expt to modify.
- ids            Specific samples to change.
- ...            Extra options are like spinach.
- factor        Batches to replace.

**Value**

The original expt with some new metadata.

**Examples**

```
## Not run:
expt = set_expt_batch(big_expt, factor=c(some,stuff,here))

## End(Not run)
```

---

set_expt_colors	<i>Change the colors of an expt!</i>
-----------------	--------------------------------------

---

## Description

After fiddling with conditions/batches, one might want to change the colors.

When exploring differential analyses, it might be useful to play with the conditions/batches of the experiment. Use this to make that easier.

## Usage

```
set_expt_colors(expt, colors = TRUE, chosen_palette = "Dark2")
```

```
set_expt_colors(expt, colors = TRUE, chosen_palette = "Dark2")
```

## Arguments

expt	Expt to modify.
colors	New color list.
ids	Specific ids to change.
...	Arguments passed along (likely colors)
expt	Expt to modify
colors	colors to replace

## Value

expt Send back the expt with some new metadata

expt Send back the expt with some new metadata

## Examples

```
## Not run:
expt = set_expt_colors(big_expt) ## This will call rcolorbrewer again

## End(Not run)
## Not run:
expt = set_expt_batch(big_expt, factor=c(some,stuff,here))

## End(Not run)
```

---

set_expt_condition	<i>Change the condition of an expt</i>
--------------------	----------------------------------------

---

### Description

When exploring differential analyses, it might be useful to play with the conditions/batches of the experiment. Use this to make that easier.

### Usage

```
set_expt_condition(expt, fact, ids = NULL, ...)
```

### Arguments

expt	Expt to modify
factor	Conditions to replace
colors	Reset the set of colors (Give a factor if you want to choose your own).

### Value

expt Send back the expt with some new metadata

### Examples

```
## Not run:
expt = set_expt_condition(big_expt, factor=c(some,stuff,here))

## End(Not run)
```

---

set_expt_factors	<i>Change the factors (condition and batch) of an expt</i>
------------------	------------------------------------------------------------

---

### Description

When exploring differential analyses, it might be useful to play with the conditions/batches of the experiment. Use this to make that easier.

### Usage

```
set_expt_factors(expt, condition = NULL, batch = NULL, ids = NULL, ...)
```

### Arguments

expt	Expt to modify
condition	New condition factor
batch	New batch factor
...	Arguments passed along (likely colors)



**Value**

expt Send back the expt with some new metadata

**Examples**

```
## Not run:
expt = set_expt_factors(big_expt, condition="column", batch="another_column")

## End(Not run)
```

---

sillydist

---

*Calculate a simplistic distance function of a point against two axes.*


---

**Description**

Sillydist provides a distance of any point vs. the axes of a plot. This just takes the abs(distances) of each point to the axes, normalizes them against the largest point on the axes, multiplies the result, and normalizes against the max of all point.

**Usage**

```
sillydist(firstterm, secondterm, firstaxis = 0, secondaxis = 0)
```

**Arguments**

firstterm	X-values of the points.
secondterm	Y-values of the points.
firstaxis	X-value of the vertical axis.
secondaxis	Y-value of the second axis.

**Value**

Dataframe of the distances.

**See Also**

**ggplot2**

**Examples**

```
## Not run:
mydist <- sillydist(df[,1], df[,2], first_median, second_median)
first_vs_second <- ggplot2::ggplot(df, ggplot2::aes_string(x="first", y="second"),
                                environment=hpgl_env) +
  ggplot2::xlab(paste("Expression of", df_x_axis)) +
  ggplot2::ylab(paste("Expression of", df_y_axis)) +
  ggplot2::geom_vline(color="grey", xintercept=(first_median - first_mad), size=line_size) +
  ggplot2::geom_vline(color="grey", xintercept=(first_median + first_mad), size=line_size) +
```

```

ggplot2::geom_vline(color="darkgrey", xintercept=first_median, size=line_size) +
ggplot2::geom_hline(color="grey", yintercept=(second_median - second_mad), size=line_size) +
ggplot2::geom_hline(color="grey", yintercept=(second_median + second_mad), size=line_size) +
ggplot2::geom_hline(color="darkgrey", yintercept=second_median, size=line_size) +
ggplot2::geom_point(colour=grDevices::hsv(mydist$dist, 1, mydist$dist),
                    alpha=0.6, size=size) +
ggplot2::theme(legend.position="none")
first_vs_second ## dots get colored according to how far they are from the medians
## replace first_median, second_median with 0,0 for the axes

## End(Not run)

```

---

## simple\_clusterprofiler

*Perform the array of analyses in the 2016-04 version of clusterProfiler*

---

### Description

The new version of clusterProfiler has a bunch of new toys. However, it is more stringent in terms of input in that it now explicitly expects to receive annotation data in terms of a orgdb object. This is mostly advantageous, but will probably cause some changes in the other ontology functions in the near future. This function is an initial pass at making something similar to my previous 'simple\_clusterprofiler()' but using these new toys.

### Usage

```

simple_clusterprofiler(sig_genes, all_genes, orgdb = "org.Dm.eg.db",
  orgdb_from = "FLYBASE", orgdb_to = c("ENSEMBL", "SYMBOL", "ENTREZID"),
  go_level = 3, pcutoff = 0.05, qcutoff = 0.1, fc_column = "logFC",
  permutations = 100, min_groupsize = 5, kegg_prefix = "Dmel_",
  kegg_organism = "dme", kegg_id_column = "FLYBASECG", categories = 12)

```

### Arguments

sig_genes	Dataframe of genes deemed 'significant.'
orgdb	Name of the orgDb used for gathering annotation data.
orgdb_from	Name of a key in the orgdb used to cross reference to entrez IDs.
orgdb_to	List of keys to grab from the orgdb for cross referencing ontologies.
go_level	How deep into the ontology tree should this dive for over expressed categories.
pcutoff	P-value cutoff for 'significant' analyses.
qcutoff	Q-value cutoff for 'significant' analyses.
fc_column	When extracting vectors of all genes, what column should be used?
permutations	How many permutations for GSEA-ish analyses?
min_groupsize	What is the minimum ontology group's size?
kegg_prefix	Many KEGG ids need a prefix before they will cross reference.

kegg_organism	Choose the 3 letter KEGG organism name here.
kegg_id_column	Column in the orgdb to use for cross referencing to KEGG.
categories	How many categories should be plotted in bar/dot plots?
universe	Dataframe of all genes in the analysis, primarily for gse analyses.

---

```
simple_clusterprofiler_old
```

*Perform a simplified clusterProfiler analysis.*

---

## Description

I like clusterProfiler quite a lot, but making it work for non-standard species is a bit of a chore. This attempts to alleviate some of those headaches and cover some corner cases where it fails.

## Usage

```
simple_clusterprofiler_old(de_genes, goids_df = NULL, golevel = 4,
  pcutoff = 0.1, fold_changes = NULL, include_cnetplots = FALSE,
  showcategory = 12, universe = NULL, species = "undef", gff = NULL,
  wrapped_width = 20, method = "Wallenius", padjust = "BH", ...)
```

## Arguments

de_genes	Data frame of differentially expressed genes, it must contain an ID column.
goids_df	df containing mappings of genes to goids in the format expected by build-GOmap().
golevel	Relative level in the tree for printing p-value plots, higher is more specific.
pcutoff	(Adj)p-value cutoff to define 'significant'.
fold_changes	Df of fold changes for the DE genes.
include_cnetplots	Cnetplots often have too many glyphs to read, so by default they are not included, however on occasion they are fairly interesting to look at.
showcategory	How many categories to show in p-value plots? Too many and they become illegible.
universe	Gene universe to use.
species	Name of the species to use if supported, jibberish otherwise.
gff	Gff file to generate the universe of genes.
wrapped_width	Width of ontology names in the pvalue plots, too long and the bars disappear, too short and the words run into the lines above.
method	Method for calculating p-values.
padjust	Method for adjusting the p-values.
...	More options, passed to arglist.

**Value**

List including the following: mf\_interesting: A table of the interesting molecular function groups  
 bp\_interesting: A table of the interesting biological process groups cc\_interesting: A table of the interesting cellular component groups mf\_pvals: A histogram of the molecular function p-values  
 bp\_pvals: Ditto, biological process cc\_pvals: And cellular component... mf\_enriched: A table of the enriched molecular function groups by adjusted p-value. bp\_enriched: yep, you guessed it  
 cc\_enriched: cellular component, too mf\_all/bp\_all/cc\_all: A table of all go categories observed (mf/bp/cc respectively) mfp\_plot/bpp\_plot/ccp\_plot: ggplot2 p-value bar plots describing the over represented groups mf\_cnetplot/bp\_cnetplot/cc\_cnetplot: clusterProfiler cnetplots mf\_group\_barplot/bp\_group\_barplot/cc\_group\_barplot: The group barplots from clusterProfiler

**Examples**

```
## Not run:
up_cluster = simple_clusterprofiler(mga2_ll_thy_top, goids=goids, gff="reference/genome/gas.gff")
## > Some chatter while it runs
## tail(head(up_cluster$bp_interesting, n=10), n=1)
## > ID ont GeneRatio BgRatio pvalue p.adjust qvalue
## > 10 GO:0009311 BP 5/195 10/1262 0.01089364 0.01089364 0.1272835
## > geneID Count
## > 10 M5005_Spy1632/M5005_Spy1637/M5005_Spy1635/M5005_Spy1636/M5005_Spy1638 5
## > Description
## > 10 oligosaccharide metabolic process

## End(Not run)
```

---

simple\_comparison

---

*Perform a simple experimental/control comparison.*


---

**Description**

This is a function written primarily to provide examples for how to use limma. It does the following:

1. Makes a model matrix using condition/batch
2. Optionally uses sva's combat (from cbcSEQ)
3. Runs voom/lmfit
4. Sets the first element of the design to "changed" and the second to "control".
5. Performs a makeContrasts() of changed - control.
6. Fits them
7. Makes histograms of the two elements of the contrast, cor.tests() them, makes a histogram of the p-values, ma-plot, volcano-plot, writes out the results in an excel sheet, pulls the up/down significant and p-value significant (maybe this should be replaced with write\_limma()?)
8. And returns a list containing these data and plots.

Currently this assumes that a variant of toptable was used which gives adjusted p-values. This is not always the case and I should check for that, but I have not yet.

**Usage**

```
simple_comparison(subset, workbook = "simple_comparison.xls",
  sheet = "simple_comparison", basename = NA, batch = TRUE,
  combat = FALSE, combat_noscale = TRUE, pvalue_cutoff = 0.05,
  logfc_cutoff = 0.6, tooltip_data = NULL, ...)
```

**Arguments**

subset	Experimental subset with two conditions to compare.
workbook	Excel workbook to which to write.
sheet	Excel worksheet to which to write.
basename	Url to which to send click evens in clicky volcano/ma plots.
batch	Whether or not to include batch in limma's model.
combat	Whether or not to use combatMod().
combat_noscale	Whether or not to include combat_noscale (makes combat a little less heavy-handed).
pvalue_cutoff	P-value definition of 'significant.'
logfc_cutoff	Fold-change cutoff of significance. 0.6 on the low end and therefore 1.6 on the high.
tooltip_data	Text descriptions of genes if one wants google graphs.
...	More parameters!

**Value**

A list containing the following pieces: amean\_histogram = a histogram of the mean values between the two conditions coef\_amean\_cor = a correlation test between the mean values and coefficients (this should be a p-value of 1) coefficient\_scatter = a scatter plot of condition 2 on the y axis and condition 1 on x coefficient\_x = a histogram of the x axis coefficient\_y = a histogram of the y axis coefficient\_both = a histogram of both coefficient\_lm = a description of the line described by  $y = \text{slope}(y/x) + b$  where coefficient\_lmsummary = the r-squared and such information for the linear model coefficient\_weights = the weights against the linear model, higher weights mean closer to the line comparisons = the result from eBayes() contrasts = the result from contrasts.fit() contrast\_histogram = a histogram of the coefficients downsignificant = a subset from toptable() of the 'down-regulated' genes ( $< 1$  Z from the mean) fit = the result from lmFit(voom\_result) ma\_plot = an ma plot using the voom\$E data and p-values psignificant = a subset from toptable() of all genes with p-values  $\leq$  pvalue\_cutoff pvalue\_histogram = a histogram of all the p-values table = everything from toptable() upsignificant = a subset from toptable() of 'up-regulated' genes ( $> 1$  Z from the mean) volcano\_plot = a volcano plot of x/y voom\_data = the result from calling voom() voom\_plot = a plot from voom(), redundant with voom\_data

**See Also**

[plot\\_gvis\\_ma](#) [toptable](#) [voom](#) [hpgl\\_voom](#) [lmFit](#) [makeContrasts](#) [contrasts.fit](#)

**Examples**

```
## Not run:
model = model.matrix(~ 0 + subset$conditions)
simple_comparison(subset, model)

## End(Not run)
```

---

simple_cp_enricher	<i>Generic enrichment using clusterProfiler.</i>
--------------------	--------------------------------------------------

---

**Description**

culsterProfiler::enricher provides a quick and easy enrichment analysis given a set of significant genes and a data frame which connects each gene to a category.

**Usage**

```
simple_cp_enricher(sig_genes, de_table, goids_df = NULL)
```

**Arguments**

sig_genes	Set of 'significant' genes as a table.
de_table	All genes from the original analysis.
goids_df	Dataframe of GO->ID matching the gene names of sig_genes to GO categories.

**Value**

Table of 'enriched' categories.

---

simple_filter_counts	<i>Filter low-count genes from a data set only using a simple threshold and number of samples.</i>
----------------------	----------------------------------------------------------------------------------------------------

---

**Description**

This was a function written by Kwame Okrah and perhaps also Laura Dillon to remove low-count genes. It drops genes based on a threshold and number of samples.

**Usage**

```
simple_filter_counts(count_table, threshold = 2, min_samples = 2)
```

**Arguments**

count_table	Data frame of (pseudo)counts by sample.
threshold	Lower threshold of counts for each gene.
min_samples	Minimum number of samples.

**Value**

Dataframe of counts without the low-count genes.

**Examples**

```
## Not run:
filtered_table <- simple_filter_counts(count_table)

## End(Not run)
```

---

simple_gadem	<i>run the rGADEM suite</i>
--------------	-----------------------------

---

**Description**

This is another function I started but never had cause to finish for the test sequences it works though

**Usage**

```
simple_gadem(inputfile, genome = "BSgenome.Hsapiens.UCSC.hs19", ...)
```

**Arguments**

inputfile	Fasta or bed file containing sequences to search.
genome	BSgenome to read.
...	Parameters for plotting the gadem result.

---

simple_goseq	<i>Perform a simplified goseq analysis.</i>
--------------	---------------------------------------------

---

**Description**

goseq can be pretty difficult to get set up for non-supported organisms. This attempts to make that process a bit simpler as well as give some standard outputs which should be similar to those returned by clusterprofiler/topgo/gostats/gprofiler.

**Usage**

```
simple_goseq(de_genes, go_db, length_db, doplot = TRUE, adjust = 0.1,
  pvalue = 0.1, qvalue = 0.1, length_keytype = "transcripts",
  go_keytype = "ENTREZID", goseq_method = "Wallenius",
  padjust_method = "BH", bioc_length_db = "ensGene", ...)
```

**Arguments**

de_genes	Data frame of differentially expressed genes, containing IDs etc.
go_db	Database of go to gene mappings (OrgDb/OrganismDb)
length_db	Database of gene lengths (gff/TxDb)
doplot	Include pwf plots?
adjust	Minimum adjusted pvalue for 'significant.'
pvalue	Minimum pvalue for 'significant.'
qvalue	Minimum qvalue for 'significant.'
length_keytype	Keytype to provide to extract lengths
go_keytype	Keytype to provide to extract go IDs
goseq_method	Statistical test for goseq to use.
padjust_method	Which method to use to adjust the pvalues.
bioc_length_db	Source of gene lengths?
...	Extra parameters which I do not recall

**Value**

Big list including: the pwd:pwf function, alldata:the godata dataframe, pvalue\_histogram:p-value histograms, godata\_interesting:the ontology information of the enhanced groups, term\_table:the goterms with some information about them, mf\_subset:a plot of the MF enhanced groups, mfp\_plot:the pvalues of the MF group, bp\_subset:a plot of the BP enhanced groups, bpp\_plot, cc\_subset, and ccp\_plot

**See Also**

[goseq](#) [goseq nullp](#)

---

simple_gostats	<i>Simplification function for gostats, in the same vein as those written for clusterProfiler, goseq, and topGO.</i>
----------------	----------------------------------------------------------------------------------------------------------------------

---

**Description**

GOstats has a couple interesting peculiarities: Chief among them: the gene IDs must be integers. As a result, I am going to have this function take a gff file in order to get the go ids and gene ids on the same page.

**Usage**

```
simple_gostats(de_genes, gff, goids_df, universe_merge = "id",
  second_merge_try = "locus_tag", species = "fun", pcutoff = 0.1,
  direction = "over", conditional = FALSE, categorysize = NULL,
  gff_type = "cds", ...)
```



**Arguments**

de_genes	Input list of differentially expressed genes.
gff	Annotation information for this genome.
goids_df	Set of GOids, as before in the format ID/GO.
universe_merge	Column from which to create the universe of genes.
second_merge_try	If the first universe merge fails, try this.
species	Genbank organism to use.
pcutoff	Pvalue cutoff for deciding significant.
direction	Under or over represented categories.
conditional	Perform a conditional search?
categorysize	Category size below which to not include groups.
gff_type	Gff column to use for creating the universe.
...	More parameters!

**Value**

List of returns from GSEABase, Category, etc.

**See Also**

**GSEABase Category**

---

simple_gprofiler	<i>Run searches against the web service g:Profiler.</i>
------------------	---------------------------------------------------------

---

**Description**

Thank you Ginger for showing me your thesis, gProfiler is pretty cool!

**Usage**

```
simple_gprofiler(de_genes, species = "hsapiens", first_col = "logFC",
  second_col = "limma_logfc", do_go = TRUE, do_kegg = TRUE,
  do_reactome = TRUE, do_mi = TRUE, do_tf = TRUE, do_corum = TRUE,
  do_hp = TRUE)
```

**Arguments**

de_genes	guess!
species	an organism supported by gprofiler
first_col	where to search for the order of 'significant' first
second_col	if that fails, try some where else.
do_go	Perform GO search?
do_kegg	Perform KEGG search?
do_reactome	Perform reactome search?
do_mi	Do miRNA search?
do_tf	Search for transcription factors?
do_corum	Do corum search?
do_hp	Do the hp search?

**Value**

a list of results for go, kegg, reactome, and a few more.

---

simple_topgo	<i>Perform a simplified topgo analysis.</i>
--------------	---------------------------------------------

---

**Description**

This will attempt to make it easier to run topgo on a set of genes.

**Usage**

```
simple_topgo(de_genes, goid_map = "id2go.map", goids_df = NULL,
  pvals = NULL, limitby = "fisher", limit = 0.1, signodes = 100,
  sigforall = TRUE, numchar = 300, selector = "topDiffGenes",
  pval_column = "adj.P.Val", overwrite = FALSE, densities = FALSE,
  pval_plots = TRUE, ...)
```

**Arguments**

de_genes	Data frame of differentially expressed genes, containing IDs any other columns.
goid_map	File containing mappings of genes to goids in the format expected by topgo.
goids_df	Data frame of the goids which may be used to make the goid_map.
pvals	Set of pvalues in the DE data which may be used to improve the topgo results.
limitby	Test to index the results by.
limit	Ontology pvalue to use as the lower limit.
signodes	I don't remember right now.
sigforall	Provide the significance for all nodes?

numchar	Character limit for the table of results.
selector	Function name for choosing genes to include.
pval_column	Column from which to acquire scores.
overwrite	Yeah I do not remember this one either.
densities	Densities, yeah, the densities...
pval_plots	Include pvalue plots of the results a la clusterprofiler?
...	Other options which I do not remember right now!

**Value**

Big list including the various outputs from topgo

---

sm	<i>Silence, m...</i>
----	----------------------

---

**Description**

Some libraries/functions just won't shut up. Ergo, silence, peasant! This is a simpler silence peasant.

**Usage**

```
sm(...)
```

**Arguments**

...                      Some code to shut up.

**Value**

Whatever the code would have returned.

---

subset_ontology_search	<i>Perform ontology searches on up/down subsets of differential expression data.</i>
------------------------	--------------------------------------------------------------------------------------

---

**Description**

In the same way all\_pairwise() attempts to simplify using multiple DE tools, this function seeks to make it easier to extract subsets of differentially expressed data and pass them to goseq, clusterProfiler, topGO, GOSTats, and gProfiler.

**Usage**

```
subset_ontology_search(changed_counts, doplot = TRUE, do_goseq = TRUE,
  do_cluster = TRUE, do_topgo = TRUE, do_gostats = TRUE,
  do_gprofiler = TRUE, according_to = "limma", ...)
```

**Arguments**

changed_counts	List of changed counts as ups and downs.
doplot	Include plots in the results?
do_goseq	Perform goseq search?
do_cluster	Perform clusterprofiler search?
do_topgo	Perform topgo search?
do_gostats	Perform gostats search?
do_gprofiler	Do a gprofiler search?
according_to	If results from multiple DE tools were passed, which one defines 'significant'?
...	Extra arguments!

**Value**

List of ontology search results, up and down for each contrast.

---

sum_exons	<i>Given a data frame of exon counts and annotation information, sum the exons.</i>
-----------	-------------------------------------------------------------------------------------

---

**Description**

This function will merge a count table to an annotation table by the child column. It will then sum all rows of exons by parent gene and sum the widths of the exons. Finally it will return a list containing a df of gene lengths and summed counts.

**Usage**

```
sum_exons(data, gff = NULL, annotdf = NULL, parent = "Parent",
  child = "row.names")
```

**Arguments**

data	Count tables of exons.
gff	Gff filename.
annotdf	Dataframe of annotations (probably from gff2df).
parent	Column from the annotations with the gene names.
child	Column from the annotations with the exon names.

**Value**

List of 2 data frames, counts and lengths by summed exons.

**See Also**

**rtracklayer**

**Examples**

```
## Not run:  
summed <- sum_exons(counts, gff='reference/xenopus_laevis.gff.xz')  
  
## End(Not run)
```

---

s\_p

*Silence, peasant!*

---

**Description**

Some libraries/functions just won't shut up. Ergo, silence, peasant! This function uses 2 invocations of `capture.output` and a `try(silent=TRUE)` to capture the strings of the outputs from the given expression in 'output', and the messages in 'message'. The result of the expression goes into 'result.' If there is an error in the expression, it is returned as a try-error object which may therefore be inspected as needed.

**Usage**

```
s_p(code)
```

**Arguments**

code                      Some code to shut up.

**Value**

List of the output log, message log, and result of the expression.

---

tnseq_saturation	<i>Make a plot and some simple numbers about tnseq saturation</i>
------------------	-------------------------------------------------------------------

---

**Description**

This function takes as input a tab separated file from essentiality\_tas.pl This is a perl script written to read a bam alignment of tnseq reads against a genome and count how many hits were observed on every TA in the given genome. It furthermore has some logic to tell the difference between reads which were observed on the forward vs. reverse strand as well as reads which appear to be on both strands (eg. they start and end with 'TA').

**Usage**

```
tnseq_saturation(file)
```

**Arguments**

file	a file created using the perl script 'essentiality_tas.pl'
------	------------------------------------------------------------

**Value**

A plot and some numbers

---

topDiffGenes	<i>A very simple selector of strong scoring genes (by p-value)</i>
--------------	--------------------------------------------------------------------

---

**Description**

This function was provided in the topGO documentation, but not defined. It was copied/pasted here. I have ideas for including up/down expression but have so far deemed them not needed because I am feeding topGO already explicit lists of genes which are up/down/whatever. But it still is likely to be useful to be able to further subset the data.

**Usage**

```
topDiffGenes(allScore)
```

**Arguments**

allScore	The scores of the genes
----------	-------------------------

---

topgo_tables	<i>Make pretty tables out of topGO data</i>
--------------	---------------------------------------------

---

### Description

The topgo function GenTable is neat, but it needs some simplification to not be obnoxious.

### Usage

```
topgo_tables(result, limit = 0.1, limitby = "fisher", numchar = 300,  
             orderby = "classic", ranksof = "classic")
```

### Arguments

result	Topgo result.
limit	Pvalue limit defining 'significant'.
limitby	Type of test to perform.
numchar	How many characters to allow in the description?
orderby	Which of the available columns to order the table by?
ranksof	Which of the available columns are used to rank the data?

---

topgo_trees	<i>Print trees from topGO.</i>
-------------	--------------------------------

---

### Description

The tree printing functionality of topGO is pretty cool, but difficult to get set correctly.

### Usage

```
topgo_trees(tg, score_limit = 0.01, sigforall = TRUE,  
            do_mf_fisher_tree = TRUE, do_bp_fisher_tree = TRUE,  
            do_cc_fisher_tree = TRUE, do_mf_ks_tree = FALSE, do_bp_ks_tree = FALSE,  
            do_cc_ks_tree = FALSE, do_mf_el_tree = FALSE, do_bp_el_tree = FALSE,  
            do_cc_el_tree = FALSE, do_mf_weight_tree = FALSE,  
            do_bp_weight_tree = FALSE, do_cc_weight_tree = FALSE)
```

**Arguments**

tg	Data from simple_topgo().
score_limit	Score limit to decide whether to add to the tree.
sigforall	Add scores to the tree?
do_mf_fisher_tree	Add the fisher score molecular function tree?
do_bp_fisher_tree	Add the fisher biological process tree?
do_cc_fisher_tree	Add the fisher cellular component tree?
do_mf_ks_tree	Add the ks molecular function tree?
do_bp_ks_tree	Add the ks biological process tree?
do_cc_ks_tree	Add the ks cellular component tree?
do_mf_el_tree	Add the el molecular function tree?
do_bp_el_tree	Add the el biological process tree?
do_cc_el_tree	Add the el cellular component tree?
do_mf_weight_tree	Add the weight mf tree?
do_bp_weight_tree	Add the bp weighted tree?
do_cc_weight_tree	Add the guess

**Value**

Big list including the various outputs from topgo.

---

transform_counts	<i>Perform a simple transformation of a count table (log2)</i>
------------------	----------------------------------------------------------------

---

**Description**

the add argument is only important if the data was previously cpm'd because that does a +1, thus this will avoid a double+1 on the data.

**Usage**

```
transform_counts(count_table, transform = "raw", base = NULL, ...)
```

**Arguments**

count_table	A matrix of count data
transform	A type of transformation to perform: log2/log10/log
base	for other log scales
...	Options I might pass from other functions are dropped into arglist.



**Value**

dataframe of logx(counts)

**Examples**

```
## Not run:
filtered_table = transform_counts(count_table, transform='log2', converted='cpm')

## End(Not run)
```

---

translate\_ids\_querymany

*Use mygene's queryMany to translate gene ID types*

---

**Description**

Juggling between entrez, ensembl, etc can be quite a hassel. This hopes to make it easier.

**Usage**

```
translate_ids_querymany(queries, from = "ensembl", to = "entrez",
  species = "human")
```

**Arguments**

queries	Gene IDs to translate.
from	Database to translate IDs from.
to	Database to translate IDs into.
species	Human readable species for translation (Eg. 'human' instead of 'hsapiens'.)

**Value**

Df of translated IDs/accessions

**See Also**

[queryMany](#)

**Examples**

```
## Not run:
data <- translate_ids_querymany(genes)

## End(Not run)
```

---

trityp_downloads	<i>Download the various data files from <a href="http://tritypdb.org/">http://tritypdb.org/</a></i>
------------------	-----------------------------------------------------------------------------------------------------

---

**Description**

The tritypdb nicely makes their downloads standardized!

**Usage**

```
trityp_downloads(version = "27", species = "lmajor", strain = "friedlin",
  dl_dir = "organdb/trityp", quiet = TRUE)
```

**Arguments**

version	What version of the tritypdb to use?
species	Human readable species to use.
strain	Strain of the given species to download.
dl_dir	Directory into which to download the various files.
quiet	Print download progress?

---

tryCatch.W.E	<i>tryCatch both warnings and errors</i>
--------------	------------------------------------------

---

**Description**

We want to catch \*and\* save both errors and warnings, and in the case of a warning, also keep the computed result.

**Usage**

```
tryCatch.W.E(expr)
```

**Arguments**

expr	an expression to try
------	----------------------

**Details**

This was taken from:<http://r.789695.n4.nabble.com/How-to-catch-both-warnings-and-errors-td3073597.html>  
and <http://tolstoy.newcastle.edu.au/R/help/04/06/0217.html>

**Value**

a list with 'value' and 'warning', where 'value' may be an error caught.

**Author(s)**

Martin Maechler

---

u\_plot*Plot the rank order svd\$u elements to get a view of how much the first genes contribute to the total variance by PC.*

---

**Description**

Plot the rank order svd\$u elements to get a view of how much the first genes contribute to the total variance by PC.

**Usage**

```
u_plot(plotted_us)
```

**Arguments**

plotted\_us      a list of svd\$u elements

**Value**

a recordPlot() plot showing the first 3 PCs by rank-order svd\$u.

---

varpart*Use variancePartition to gather some plots about a troubling dataset*

---

**Description**

variancePartition is the newest toy introduced by Hector

**Usage**

```
varpart(expt, factors = c("condition", "batch"), cpus = 6, genes = 20)
```

**Arguments**

expt	Some data
factors	Character list of columns in the experiment design to query
cpus	Number cpus to use
genes	Number of genes to count

**Value**

partitions List of plots and variance data frames

---

write_goseq_data	<i>Make a pretty table of goseq data in excel.</i>
------------------	----------------------------------------------------

---

### Description

It is my intention to make a function like this for each ontology tool in my repertoire

### Usage

```
write_goseq_data(goseq, file = "excel/goseq.xlsx", pval = 0.1,
  add_plots = TRUE)
```

### Arguments

goseq	A set of results from simple_goseq().
file	An excel file to which to write some pretty results.
pval	Choose a cutoff for reporting by p-value.
add_plots	Include some pvalue plots in the excel output?

### Value

The result from openxlsx

---

write_go_xls	<i>Write gene ontology tables for excel</i>
--------------	---------------------------------------------

---

### Description

Combine the results from goseq, cluster profiler, topgo, and gostats and drop them into excel Hopefully with a relatively consistent look.

### Usage

```
write_go_xls(goseq, cluster, topgo, gostats, gprofiler,
  file = "excel/merged_go", dated = TRUE, n = 30, overwritefile = TRUE)
```

### Arguments

goseq	The goseq result from simple_goseq()
cluster	The result from simple_clusterprofiler()
topgo	Guess
gostats	Yep, ditto
file	the file to save the results.
dated	date the excel file
n	the number of ontology categories to include in each table.
overwritefile	overwrite an existing excel file

**Value**

the list of ontology information

---

write\_gprofiler\_data    *Write some excel results from a gprofiler search.*

---

**Description**

Gprofiler is pretty awesome. This function will attempt to write its results to an excel file.

**Usage**

```
write_gprofiler_data(gprofiler_result, wb = NULL,
  excel = "excel/gprofiler_result.xlsx", add_plots = TRUE, ...)
```

**Arguments**

gprofiler_result	The result from simple_gprofiler().
wb	Optional workbook object, if you wish to append to an existing workbook.
excel	Excel file to which to write.
add_plots	Add some pvalue plots?

---

write\_limma            *Writes out the results of a limma search using toptable().*

---

**Description**

However, this will do a couple of things to make one's life easier: 1. Make a list of the output, one element for each comparison of the contrast matrix 2. Write out the toptable() output for them in separate .csv files and/or sheets in excel 3. Since I have been using qvalues a lot for other stuff, add a column for them.

**Usage**

```
write_limma(data, adjust = "fdr", n = 0, coef = NULL,
  workbook = "excel/limma.xls", excel = FALSE, csv = FALSE,
  annot_df = NULL)
```

**Arguments**

data	Output from eBayes().
adjust	Pvalue adjustment chosen.
n	Number of entries to report, 0 says do them all.
coef	Which coefficients/contrasts to report, NULL says do them all.
workbook	Excel filename into which to write the data.
excel	Write an excel workbook?
csv	Write out csv files of the tables?
annot_df	Optional data frame including annotation information to include with the tables.

**Value**

List of data frames comprising the toptable output for each coefficient, I also added a qvalue entry to these toptable() outputs.

**See Also**

[toptable](#) [write\\_xls](#)

**Examples**

```
## Not run:
finished_comparison = eBayes(limma_output)
data_list = write_limma(finished_comparison, workbook="excel/limma_output.xls")

## End(Not run)
```

---

write\_subset\_ontologies

*Write gene ontology tables for data subsets*

---

**Description**

Given a set of ontology results, this attempts to write them to an excel workbook in a consistent and relatively easy-to-read fashion.

**Usage**

```
write_subset_ontologies(kept_ontology, outfile = "excel/subset_go",
  dated = TRUE, n = NULL, overwritefile = TRUE, add_plots = TRUE,
  table_style = "TableStyleMedium9", ...)
```

**Arguments**

kept\_ontology    A result from subset\_ontology\_search()  
 outfile          Workbook to which to write.  
 dated            Append the year-month-day-hour to the workbook.  
 n                How many ontology categories to write for each search  
 overwritefile    Overwrite an existing workbook?  
 add\_plots        Add the various p-value plots to the end of each sheet?  
 table\_style      The chosen table style for excel  
 ...              some extra parameters

**Value**

a set of excel sheet/coordinates

**Examples**

```
## Not run:
all_contrasts <- all_pairwise(expt, model_batch=TRUE)
keepers <- list(bob = ('numerator','denominator'))
kept <- combine_de_tables(all_contrasts, keepers=keepers)
changed <- extract_significant_genes(kept)
kept_ontologies <- subset_ontology_search(changed, lengths=gene_lengths,
                                           goids=goids, gff=gff, gff_type='gene')
go_writer <- write_subset_ontologies(kept_ontologies)

## End(Not run)
```

---

write\_xls

---

*Write a dataframe to an excel spreadsheet sheet.*


---

**Description**

I like to give folks data in any format they prefer, even though I sort of hate excel. Most people I work with use it, so therefore I do too. This function has been through many iterations, first using XLConnect, then xlsx, and now openxlsx. Hopefully this will not change again.

**Usage**

```
write_xls(data, wb = NULL, sheet = "first", rownames = TRUE,
          start_row = 1, start_col = 1, ...)
```

**Arguments**

data	Data frame to print.
wb	Workbook to which to write.
sheet	Name of the sheet to write.
start_row	First row of the sheet to write. Useful if writing multiple tables.
start_col	First column to write.
...	Set of extra arguments given to openxlsx.

**Value**

List containing the sheet and workbook written as well as the bottom-right coordinates of the last row/column written to the worksheet.

**See Also**

**openxlsx** [writeDataTable](#)

**Examples**

```
## Not run:
xls_coords <- write_xls(dataframe, sheet="hpgl_data")
xls_coords <- write_xls(another_df, sheet="hpgl_data", start_row=xls_coords$end_col)

## End(Not run)
```

---

ymxb_print	<i>Print a model as <math>y = mx + b</math> just like in grade school!</i>
------------	----------------------------------------------------------------------------

---

**Description**

Because, why not!?

**Usage**

```
ymxb_print(model)
```

**Arguments**

model	Model to print from glm/lm/robustbase.
-------	----------------------------------------

**Value**

a string representation of that model.



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