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Dio normatics [MBD517]
Internal-II

UI Ans

Smith-Waterman algorithm: The algorithm is used to

determine similar regions

between two sequences of nucleic acid or protein

sequences. The algorithm compares segments of all

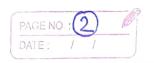
possible lengths and optimizes the similarity measure

Smith-Waterman algorithm align two sequences by mismatches/matches, insertions and deletions. Both insertions and deletions are the operations that introduce gaps, which are represented by dashes.

Performance:

- Wosst-Case: O(mn)
- · Worst case : O(mn)

Space Complexity



Algorithm: 1 Initialize the materix: (NXM) The top sow one left column are filled with zoro. 2) If the sub-alignment score become negative, restort the search 3) Fill the scoring matering using 11 = max max 9 = { H: 5 - W 3 max & Hink - Wk3 H. + & (9-, b;) (15i <n, 155 <m) 9 Traceback: Starting at the highest score in at a mateix cell that has a score of o traceback based on the source of each score recursively to generate the best local

alignment.

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Scole = 7

Ans Griven standard Newick format as

((A,B)C) (D,E)) F

A B C D E

(Phylogenetic Tree)

03 | Ans

Given Distance matrix:

→T-L:

-				The transmission of the last o	CONTRACTOR OF THE PARTY OF THE	
·	Species	A	В	C	D	
	B	3	-	-		
	C	6	5	_	-	
	$\mathcal{D}$	9	9	10	_	İ
	E	12	1)	13	9	1

Using UPGMA and TDM for above table we get
Min distance = 3 of Species A and B



## > T-2: By grouping A and B

-					
	Species	AB	C.	D	
	C	5.5			
	D	9	10		
	F	11.5	10	0	
-		1175	13	9	

Now min. distance is 5-5 of AB and C

-> T-3: By grouping AB and C

	ABC	D	-
D	9.5		-
E	12-25	9	-

## → T-4:

	ABC
$D\epsilon$	10-625

A B C D E

- Phylogenetic Tear

(A,B)C)(DE)

Q4) You have a file named "Protein\_seq.fasta" which contain 100 protein sequences. The header of the protein sequence is like that "gi|5524211|gb|AAD44166.1| cytochrome b [Elephas maximus maximus]". Then

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## 1 Writing a small Python code to identify the unique sequence?

```
[1]: from Bio import SeqIO
     import re
     fasta_filename='Protein_seq.fasta'
     fasta_seq=list()
     fasta_des=list()
     #loop to get sequence id, seq and description
     for record in SeqIO.parse(fasta_filename, "fasta"):
         fasta_seq.append(str(record.seq))
         fasta_des.append(repr(record.description))
     #create dataframe
     df = pd.DataFrame(data={'fasta_des':fasta_des,'fasta_seq':fasta_seq}, index = __
     →fasta seq)
     #remove duplicates and give unique sequences
     df = df[~df.index.duplicated()]
     #list of sequence
     unique_seq=list(df.fasta_seq)
     print("Total sequences are",len(fasta_seq))
     print("Total unique sequences are",len(unique_seq))
```

<IPython.core.display.Javascript object>

Total sequences are 100
Total unique sequences are 100

## 2 Using Python code how you separate the Species from that unique sequences?

```
[2]: #create list of unique sequences description
    seq_desc=list(df['fasta_des'])

def ExtractSpecies(seq_desc):
    species=list()
    for i in range(len(seq_desc)):
        species.append(re.findall(r"\[(.*?)\]", seq_desc[i])[0])
    s=set(species)
    return(s)

species=ExtractSpecies(seq_desc)
    print(species)
```

{'Staphylococcus', 'Bacillus cereus Q1', 'Ectocarpus sp. CCAP 1310/34', 'Hondaea fermentalgiana', 'Staphylococcus aureus', 'Lingulodinium polyedra'}