# NFP:Network fingerprint - a knowledge-based characterization of biomedical networks

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#### 1 Introduction

Recent network fingerprint method based on GO knowledge and propagation clustering mentioned in [3] provide a novel representation of network differentiation known as biological spectra, or Network Fingerprint. This method was used to describe the relationship between multiple disease networks and its related pathways, and to visually compare and parse different diseases by generating a fingerprint overlay. Thus, we achieve the function of complex network comparison based on network fingerprint on the open scientific computing platform *RR*, and present *NFP*, an *Bioconductor* Bioconductor package for fingerprint-based network analyzing and comparison of systems. Driven by the research needs of customers, *NFP* provides a unified interface to three similarity clustering algorithms. In addition, *NFP* can also provide multiscale statistical analysis and visualization, access to the specific attributes of different disease networks.

This manual is a brief introduction to structure, functions and usage of *NFP* package. The NFP package provides a set of functions to support knowledge-based network fingerprint (NFP) framework. A biomedical network is characterized as a spectrum-like vector called "network fingerprint", which contains similarities to basic reference networks. This framework provides a more intuitive way to decipher molecular networks, especially for large-scale network comparisons and clustering analyses.

The three main features of NFP:

- Basic reference networks generation.
- Network comparison, which encompasses network merging, annotation and similarity scoring.
- Network standardization.

#### 1.1 Installation

NFP requires these packages: magrittr, igraph, plyr, ggplot2, apcluster, dplyr, stringr, graph and KEGGgraph. These should be installed automatically when you install NFP with biocLite() as follows:

```
## install release version of mmmet
source("http://bioconductor.org/biocLite.R")
biocLite("NFP")

##install the latest development version
useDevel()
biocLite("NFP")
```

Or users can install the latest development version from github. To install packages from GitHub, you first need install the devtools package on your system with install.packages("devtools"). Note that devtools sometimes needs some extra non-R software on your system – more specifically, an Rtools download for Windows or Xcode for OS X.

```
## install NFP from github
if (!require(devtools))
  install.packages("devtools")
devtools::install_github("yiluheihei/NFP")
```

After installation, the *NFP* is ready to load into the current workspace by the following codes to the current workspace by typing or pasting the following codes:

## 2 Analysis Pipeline: from Basic Reference Network Generation to Network Networkfingerprint Visualization

We will demonstrate go through an analysis pipeline to illustrate some of the main functions in *NFP*. This pipeline consists of several steps:

- 1. Basic Reference Network Generation: prepare the well-known biomedical netowks as the NFP framework reference networks. Several pathway databases have been developed for biological network research, e.g. KEGG, Reactome (www.reactome.org). All of this pathway databases is well-stuied and can be used as the basic reference networks of NFP.
- 2. Network fingerprint calculation: The similarity between two biomedical networks is calculated based on the following intition: grouping the nodes in the merged network into strongly inter-connected communities with high functional similarity score between intra-community nodes in different networks The functional similarity was measured based on GO [5]. And we employed affinity propagation (AP) [2] clustering algorithm to detect the aligned functional modules between the two networks to be compared.
- 3. Network fingerprint Visualization: Show the network fingerprint along all the reference networks. We could observe the differences of different biological networks' fingerprint intuitively from visualization.

#### 2.1 Generating well-studied basic reference networks

The basic idea of calculating the network fingerprint is to have the biomedical networks map to well-studied basic networks. KEGG PATHWAY is a collection of manually drawn pathway maps representing our knowledge on the molecular interaction and reaction networks. Since its first introduction in 1995, KEGG PATHWAY has been widely used as a reference knowledge base for understanding biological pathways and functions of cellular processes. The knowledge from KEGG has proven of great value by numerous work in a wide range of fields [1]. So by default, we take KEGG pathways as basic reference networks in *NFP* by default.

Function load\_KEGG\_refnet can be used to retrieve the KEGG pathway maps with KEGG API http://www.kegg.jp/kegg/rest/keggapi.h In KEGG pathways, only the pathways of the map are manual drawing, and to different organisms, the map reference helps the automatic generation of organism-specific pathway for each organism. The organism (e.g. organsim ="hsa") parameter indicate the organism name of KEGG pathway maps.

```
## donot run, retrive pathway maps from KEGG database may take several minutes,
## we have pre-stored this data in our package
## kegg_refnet <- load_KEGG_refnet(organism = 'hsa')</pre>
data(kegg_refnet)
# show the kegg reference networks
show(kegg_refnet)
## Basic networks of organism hsa
## 134 basic networks; classification into 4 groups:
##
                                        group_name net_num
## group1 Genetic Information Processing 22
## group2 Environmental Information Processing
                                                         28
                               Cellular Processes
                                                          15
## group3
                               Organismal Systems
## group4
                                                          69
## group1 RNA polymerase - Homo sapiens (human), ...
## group2 ABC transporters - Homo sapiens (human), ...
## group3 Endocvtosis - Homo sapiens (human), ...
## group4 Hematopoietic cell lineage - Homo sapiens (human), ...
```

We defined a new S4 class *NFPrefnet* to store the *NFP* reference networks. *NFP* also provides five kinds of methods for this S4 class:

- 1. net: Exact the basic reference networks of NFPRefnet.
- 2. group: Obtain the group information, group names, number and the size of each group, e.g. KEGG pathway database contains seven group pathway maps.

3.subnet: Extract or replace parts of the NFP basic reference networks.

4.show: Display of NFPRefnet.

5.name: Extract the names of reference networks.

```
## group information of kegg reference networks
refnet_group <- group(kegg_refnet)</pre>
show(refnet_group)
## $name
## [1] "Genetic Information Processing"
## [2] "Environmental Information Processing"
## [3] "Cellular Processes"
## [4] "Organismal Systems"
##
## $num
## [1] 4
##
## $size
##
         Genetic Information Processing Environmental Information Processing
##
##
                                                             Organismal Systems
                      Cellular Processes
                                       15
## select goup 1 and 2, and subset this two groups
selected_group <- refnet_group$name[c(1,2)]</pre>
NFPnet <- subnet(kegg_refnet,selected_group)</pre>
```

```
## Basic networks of organism hsa
##
## 50 basic networks; classification into 2 groups:
## group_name net_num
## group1 Genetic Information Processing 22
## group2 Environmental Information Processing 28
## net_name
## group1 RNA polymerase - Homo sapiens (human), ...
## group2 ABC transporters - Homo sapiens (human), ...
```

Detailed instructions for this five methods refer to package function help.

Obviously users can also customize a NFPRefnet as a reference for computing network fingerprint. Users can refer to the documents of *NFPRefnet* about the composition details of this class. *graphite* [6] allow users to build *graphNEL* object from several pathway databases.

```
## Reactome human pathway maps
require(graphite)
human_pathway <- pathways("hsapiens", "kegg")</pre>
## just choose first two pathway maps for testing
p <- human_pathway[1:2]</pre>
show(p)
## KEGG pathways for hsapiens
## 2 entries, retrieved on 03-04-2015
g <- lapply(p, pathwayGraph)
show(g)
## $`Acute myeloid leukemia`
## A graphNEL graph with directed edges
## Number of Nodes = 57
## Number of Edges = 152
##
## $ Adherens junction
## A graphNEL graph with directed edges
## Number of Nodes = 70
## Number of Edges = 189
```

Then users can create their own customized NFPRefnet object as following:

```
group(customized_refnet)
## $name
## [1] "test group"
##
## $num
## [1] 1
##
## $size
## [1] 2
subnet(customized_refnet, 'test group', 1)
## Basic networks of organism hsa
##
## 1 basic networks; classification into 1 groups:
         group_name net_num
                                               net_name
## group1 test group 1 Acute myeloid leukemia, ...
```

#### 2.2 Network fingerprint calculation

NFP algorithm consists of three steps: merging network, nodes clustering and similarity scoring.

Network merging. The two networks to be compared are first merged into one. Given two networks G1 and G2, the merged network Gm is constructed by connecting each node between the G1 and G2 network. Two nodes corresponding to the same protein in the merged network are replaced by a single node that inherited all the interactions from the two individual nodes in the subsequent process.

Clustering in merged network. Grouping the nodes in the merged network into strongly inter-connected communities with high functional similarity score between intra-community nodes in different networks. We employed affinity propagation (AP) clustering algorithm to detect the aligned functional modules between the two networks to be compared. The nodes are grouped on the cluster based on nearest neighbor analysis.

Similarity scoring. The calculation of similarity score is processed in two steps: First, local similarity for each cluster and network similarity among cluster. Second, standardization: the original similarity score depends on the topological properties of query network to some extent. There is implicit bias of network fingerprint, because the outliers could be greatly distorted the relevant pattern presented in the network fingerprint. In order to eliminate the possible topological weight differences, the similarity calculation process of each node are standardized processing and the final network fingerprint facing to users is totally standardized. The standardization process is based on the random distribution of similarity scores. To the number of nodes, the number of edges and node degree, these three topological properties of random network for standardized estimate are consistent with the original network.

To not affect the results of standardization and improve the efficiency of fingerprint calculation, we set limit on the permutation time (the default is 100) of background network randomization in the standardization process. Users can also adjust randomization time of background network according to their own demands for the precision of network fingerprint.

NFP provides the calc\_sim\_score function for calculating the similarity score between two networks. As the similarity score is subjected to the size of the network, we use the maslov's method [4] to randomize a network while preserving the degree distribution. Thenperm parameter is added to calc\_sim\_score refers to the permutation times (the default is 100) of random network while calculating the similarity score mentioned above. We define a S4 class NFP in our package to store the calculation results of network fingerprints.

Simply, we choose the two pathway maps g as a query network, and a subset networks of kegg\_refnet as the reference networks. Then the NFP can be calculated as following:

```
## set g as the query network
query_net <- g
## a subset of kegg_refnet, select the head five networks of group 1, 2
group_names <- group(kegg_refnet)$name</pre>
sample_NFPRefnet <- subnet(kegg_refnet, group_names[1:2],list(1:5,1:5))</pre>
## In order to save calculating time, we take nperm = 10
NFP_score <- lapply(query_net, calc_sim_score, NFPnet = sample_NFPRefnet,
 nperm = 10)
## methods of NFP class
show(NFP_score[[1]])
## NFP object
##
## Number of ref networks = 10
## Group numbrer of networks = 2
## Permutation number = 10
## Number of clusters = 10
##
## Standardized NFP
## [1] -3.8891487 0.1044768 -2.1581520 2.3518583 -1.4389459 -1.5158342
## [7] 2.3890924 1.0312322 0.7505621 4.5948691
randomized_score <- perm_score(NFP_score[[1]])</pre>
cluster <- cluster_info(NFP_score[[1]])</pre>
```

#### 2.3 Network fingerprint visualization

NFP provides the plot\_NFP function to visualize the network fingerprint of a single query network.

```
plot(NFP_score[[1]])
```

#### **Session Information**

The version number of R and packages loaded for generating the vignette were:

```
## R version 3.2.2 (2015-08-14)
## Platform: x86_64-apple-darwin13.4.0 (64-bit)
## Running under: OS X 10.10.2 (Yosemite)
##
## locale:
## [1] C/zh_CN.UTF-8/zh_CN.UTF-8/C/zh_CN.UTF-8/zh_CN.UTF-8
##
## attached base packages:
## [1] parallel stats graphics grDevices utils datasets methods
## [8] base
##
## other attached packages:
## [1] graphite_1.14.1 BiocGenerics_0.14.0 NFP_0.99.0
## [4] graph_1.46.0 knitr_1.11
##
```

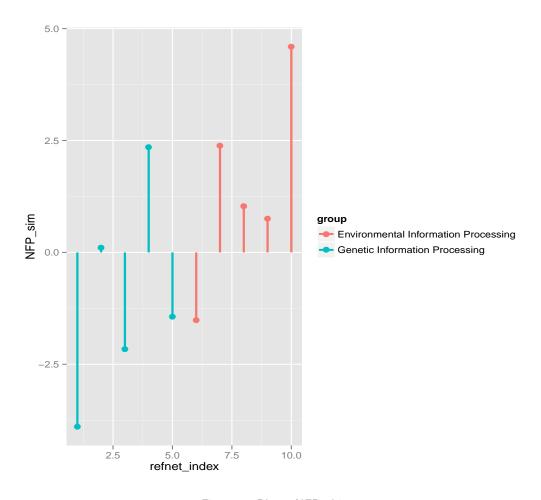


Figure 1: Plot a NFP object

```
## loaded via a namespace (and not attached):
   [1] Rcpp_0.12.1
                                                  plyr_1.8.3
##
                             formatR_1.2
   [4] highr_0.5
                             GenomeInfoDb_1.4.3
                                                  tools_3.2.2
##
   [7] digest_0.6.8
                             evaluate_0.7.2
                                                  RSQLite_1.0.0
  [10] gtable_0.1.2
                             lattice_0.20-33
                                                  Matrix_1.2-2
  [13] KEGGgraph_1.26.0
                             igraph_1.0.1
                                                  DBI_0.3.1
##
  [16] proto_0.3-10
                             apcluster_1.4.1
                                                  dplyr_0.4.3
   [19] stringr_1.0.0
                             IRanges_2.2.7
                                                  S4Vectors_0.6.6
   [22] stats4_3.2.2
                             grid_3.2.2
                                                   Biobase_2.28.0
  [25] R6_2.1.1
                             AnnotationDbi_1.30.1 XML_3.98-1.1
## [28] ggplot2_1.0.1
                             reshape2_1.4.1
                                                  magrittr_1.5
## [31] scales_0.3.0
                             MASS_7.3-43
                                                   assertthat_0.1
## [34] BiocStyle_1.6.0
                             colorspace_1.2-6
                                                  labeling_0.3
## [37] stringi_0.5-5
                             munsell_0.4.2
```

### Cleanup

This is a cleanup step for the vignette on Windows; typically not needed for users.

```
allCon <- showConnections()
socketCon <- as.integer(rownames(allCon)[allCon[, "class"] == "sockconn"])
sapply(socketCon, function(ii) close.connection(getConnection(ii)))
## list()</pre>
```

#### References

- [1] Kanehisa M, Araki M, Goto S, et al. KEGG for linking genomes to life and the environment[J]. Nucleic acids research, 2008, 36(suppl 1): D480-D484.
- [2] Frey B J, Dueck D. Clustering by passing messages between data points[J]. science, 2007, 315(5814): 972-976.
- [3] Cui X, He H, He F, et al. Network fingerprint: a knowledge-based characterization of biomedical networks[J]. Scientific reports, 2015, 5.
- [4] Maslov S, Sneppen K. Specificity and stability in topology of protein networks[J]. Science, 2002, 296(5569): 910-913.
- [5] Ashburner M, Ball C A, Blake J A, et al. Gene Ontology: tool for the unification of biology[J]. Nature genetics, 2000, 25(1): 25-29.
- [6] Sales G, Calura E, Cavalieri D, et al. graphite-a Bioconductor package to convert pathway topology to gene network[J]. BMC bioinformatics, 2012, 13(1): 20.