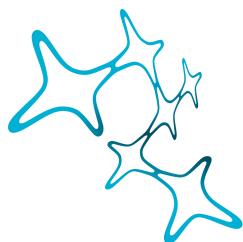


ABHISHEK MISHRA

NEURAL INFORMATION
PROCESSING IN THE
DROSOPHILA MOTION
VISION PATHWAY



Dissertation der Graduate School of Systemic Neurosciences der
Ludwig-Maximilians-Universität München

19th of September, 2022

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E-MAIL:

abhishek.mishra@bi.mpg.de

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First reviewer and supervisor
Prof. Dr. Alexander Borst

Second reviewer
Prof. Dr. Ruben Portugues

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SUMMARY

Detecting the direction of image motion is an essential component of visual computation. An individual photoreceptor, however, does not explicitly represent the direction in which the image is shifting. Comparing neighboring photoreceptor signals over time is used to extract directional motion information from the photoreceptor array in the circuit downstream. To implement direction selectivity, two opposing models have been proposed. In both models, one input line is asymmetrically delayed compared to the other, followed by a non-linear interaction between the two input lines. The Hassenstein-Reichardt (HR) model proposes an enhancement in the Preferred Direction (PD): the preferred side signal is delayed and then amplified by multiplying it with the other input signal. In contrast, the Barlow-Levick (BL) detector proposes a Null Direction (ND) suppression, whereby the null side signal is delayed and the other input is divided by it. The motion information is computed in parallel ON and OFF pathways. T₄ and T₅ are the first direction-selective neurons found in the ON and in the OFF pathway, respectively. Four subtypes of T₄ and T₅ cells exist each responding selectively to one of the four cardinal directions: front-to-back, back-to-front, upwards, and downwards, respectively.

In the first manuscript, we found that both preferred direction enhancement and null direction suppression are implemented in the dendrites of all four subtypes of both T₄ and T₅ cells to compute the direction of motion. We, therefore, propose a hybrid model combining both PD enhancement on the preferred side and ND suppression on the null side. This combined strategy ensures a high degree of direction selectivity already at the first stage of calculating motion direction.

Further processing, in addition to synaptic mechanisms on the dendrites of T₄ cells, can improve the direction selectivity of the T₄ cells' output signals. Such processing might involve: 1.) transformation from voltage to calcium, and 2.) from calcium to neurotransmitter release. In the second manuscript, we used *in vivo* two-photon imaging of genetically encoded voltage and calcium indicators, Arclight and GCaMP6f respectively, to measure responses in *Drosophila* direction-selective T₄ neurons. Comparison between Arclight and GCaMP6f signals revealed calcium signals to have a significantly higher direction selectivity compared to voltage signals. Using these recordings we built a model which transforms T₄ voltage responses into calcium responses. The model reproduced experimentally measured calcium responses across different visual stimuli using various temporal filtering steps and a stationary non-linearity. These findings provided a mechanistic underpinning of the voltage-to-calcium transformation and showed how this processing step, in addition to synaptic mechanisms on the dendrites of T₄ cells, enhances direction selectivity in the output signal of T₄ neurons.

The two manuscripts included in this thesis are presented chronologically. The first manuscript was published in a peer-reviewed journal, while the second manuscript is currently under review and is available as a preprint.

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1

INTRODUCTION

1.1 *Drosophila* AS A MODEL ORGANISM

Drosophila melanogaster is one of the most powerful model organisms available for the functional dissection of neural circuits. It allows for sophisticated *in vivo* neural manipulations that includes imaging, activation, and suppression of neural activity. The *Drosophila* research community has developed thousands of 'driver-lines' that can be used to express genes of interest in a neuron-specific manner (Pfeiffer *et al.* 2008). Additionally, *Drosophila* offers several practical advantages: fruit flies are small, have a short generation time of about 10 days, and are easy to grow in the lab.

The *Drosophila* brain is estimated to contain only about 200,000 neurons (Raji & Potter 2021) but produces behavior of rich complexity (Card & Dickinson 2008; Pavlou & Goodwin 2013; Ryu *et al.* 2022). In systems neuroscience, a common goal is to understand how the brain processes and extracts relevant information from the sensory inputs to produce behavior. *Drosophila* constitutes an ideal model organism to study the neural circuits and computations underlying behavior. Given some surprising parallels between how the fly and mammalian brains process information (Borst & Helmstaedter 2015), insights about the nervous system obtained in *Drosophila* often might be relevant for understanding the brain of other species (Bellen *et al.* 2010).

1.2 TOOLS FOR FUNCTIONAL DISSECTION OF *Drosophila* NEURAL CIRCUITS

To have a detailed understanding of how a neural circuit functions, the role each individual neuron plays in that particular circuit needs to be known. To achieve this, the following three types of manipulations can be performed on the given neuron: (i) record neuronal activity, (ii) activate the neuron, and (iii) silence the neuron. Fortunately, decades of research in *Drosophila* have provided multiple tools that allow for these manipulations in the choice of the neuron. The most important tool that enables to do this in a neuron-specific manner is the Gal4-UAS system (figure 1).

1.2.1 Targeting cell types: Gal4-UAS / LexA-lexAop

Following the discovery of transposable DNA sequences (P-elements) in the *Drosophila* genome (Rubin & Spradling 1982), the Gal4-UAS system was designed (Brand & Perrimon 1993). The Gal4-UAS system is a binary expression system consisting of two main components: the yeast transcriptional

factor Gal4 expressed in a specific pattern, and a reporter gene under the control of a upstream activation sequence (UAS) promoter that is silent in the absence of Gal4. The Gal4-UAS system involves crossing two fly lines: one called the 'driver-line', defines which neurons express the required effector gene; the other called 'reporter-line', defines what gene is expressed in the neurons defined by the driver line (figure 1).

Another independent binary transcriptional system that can be used is the LexA-lexAop system. This method is based on the bacterial DNA-binding operator lexAop and controlled by the expression of LexA. The LexA binds to and activates the lexA operator (lexAop). The LexA-lexAop system can be used in combination with the Gal4-UAS system to simultaneously express two different genes in two different neuronal populations.

Initially, the Gal4 fly lines were created by injecting randomly integrating P-elements transposons into the *Drosophila* embryos. However, the lack of control over specific insertion sites often resulted in broader expression patterns, making them unsuitable for circuit manipulations. Currently, DNA fragments with presumed enhancer activity are directly cloned for increased efficacy, and intersectional strategies, such as the split-Gal4 are used for increased specificity (Jenett *et al.* 2012; Pfeiffer *et al.* 2008). In this method, the coding region of the Gal4 is split into two units: (i) the Gal4 activating domain (AD), and (ii) the Gal4 DNA binding domain (DBD). The expression of the Gal4-AD is under the control of one enhancer and the expression of the Gal4-DBD is under the control of another enhancer. The DBD and AD proteins alone are not able to promote gene expression; only cells where both enhancers are active produce functional Gal4 protein. Therefore, only in cells containing both subunits, the gene of interest is expressed (Luan *et al.* 2006). Thousands of fly lines have been generated as a result, and nearly every type of fly neuron can be targeted with high specificity (Pfeiffer *et al.* 2010).

1.2.2 Measuring neural activity

ELECTROPHYSIOLOGY Electrophysiological recordings are used to measure neural activity by recording voltage or current changes across the neuronal membrane at a high temporal resolution. Depending on where the electrode is placed in relation to the cell, electrophysiological recordings can be classified into three main types: (1) extracellular recordings; (2) intracellular recordings; and (3) patch-clamp recordings. The large membrane voltage changes during an action potential causes local, temporary differences in potential in the extracellular space near the membrane of an active neuron. In extracellular recordings, an electrode is placed in the vicinity of the neuron to record these extracellular voltage changes. In insects, however, neurons often do not fire action potentials, but rather use graded potentials (Haag & Borst 1998). In the blowfly *Calliphora erythrocephala*, intracellular recordings using a sharp electrode were used to characterize the large lobula plate tangential cells (Hausen 1976; Krapp *et al.* 1998).

The small size of *Drosophila* neurons makes sharp electrode recordings difficult. The third variant, the whole-cell patch-clamp recordings were found

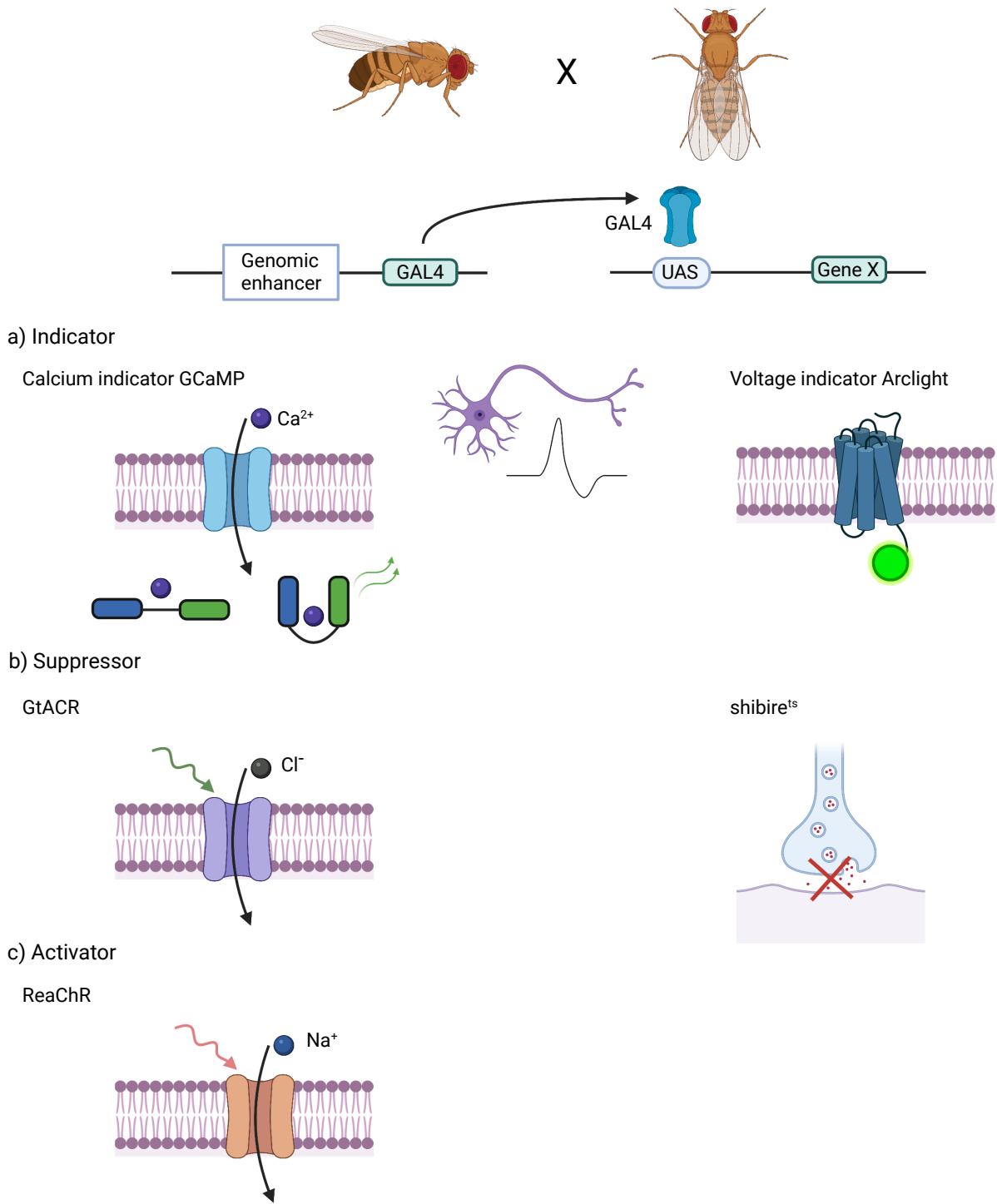


Figure 1: The Gal4-UAS system is used to express a gene of interest in a specific subset of neurons. (a) The calcium indicator is used to record neural activity using intracellular calcium concentration. The voltage indicator is used to optically record membrane potential changes in the neuron. (b) Neural activity can be suppressed by expressing light-sensitive chloride channels or by blocking synaptic transmission via the expression of temperature-sensitive *shibire^{ts}*. (c) Neurons can be activated via the expression of light-sensitive cation channels. (modified from Borst 2009)

to be better suited (Hamill *et al.* 1981). The patch-clamp method involves making tight contact with a tiny patch of the neuronal membrane with a glass micropipette. By briefly applying strong suction to the pipette, the membrane patch within it can be disrupted and the interior of the pipette can be continuous with the cytoplasm. In this configuration, electrical potentials and currents are measured from the entire cell, thus the method is called whole-cell recording. Several brain areas, including the visual (Behnia *et al.* 2014; Groschner *et al.* 2022; Gruntman *et al.* 2018; Joesch *et al.* 2008) and olfactory systems (Wilson *et al.* 2004), have been recorded *in vivo* using patch-clamp techniques in *Drosophila*.

TWO-PHOTON MICROSCOPY Although electrophysiology is widely used to record action potentials and sub-threshold changes in membrane potential, it has significant disadvantages. Electrodes must be inserted into the neurons or the brain tissue. This can cause cell or tissue damage. Additionally, only a limited number of neurons can be recorded simultaneously. With the advent of silicon probe technology, which allows multiple probes to be inserted into the brain (each with hundreds of contact points), a greater number of neurons and brain regions can be sampled using electrophysiology. However, these recordings are more invasive, more expensive, and have the same limitations as single-neuron electrode recordings. Optical probing of neural activity, especially combining calcium or voltage imaging with two-photon microscopy has become popular as an alternative to electrophysiology.

The invention of two-photon microscopy (Denk *et al.* 1990) has been one of the major breakthroughs in neuroscience. It allows high-sensitivity and high-resolution fluorescence detection in brain tissue *in vivo*. In two-photon microscopy, two low-energy near-infrared or infrared photons (usually from the same laser) cooperate to produce an electronic transition in a fluorescent molecule from the ground to the excited state. In other words, a fluorescent molecule can achieve a higher energy state either by absorbing a single photon from 455 nm light or by absorbing two photons simultaneously from a light of wavelength 910 nm. Compared to one-photon techniques, two-photon excitation provides several advantages for microscopy in scattering specimens like the brain (Denk *et al.* 1994; Svoboda & Yasuda 2006). First, compared to the visible wavelengths used in one-photon microscopy, the near-infrared or infrared excitation wavelengths used in two-photon microscopy penetrate the tissue better. This happens due to the reduced scattering and reduced absorption by endogenous chromophores. Second, the bleaching of fluorophores is reduced in two-photon imaging compared to one-photon imaging. Fluorophores lose their brightness when exposed to high-energy light. Light of higher wavelength used in two-photon microscopy carries less energy and hence, causes reduced bleaching. Third, the light above 900nm in the infrared region is beyond the spectral sensitivity of a fly's eye. Hence, the laser light won't interfere with the light used to create visual stimuli. Combining two-photon microscopy with a precise genetic expression of genetically encoded calcium indicators (GECIs) (Chen *et al.* 2013) has been extensively used to measure neural activity in *Drosophila* neuroscience.

TWO-PHOTON CALCIUM IMAGING *In vivo* two-photon calcium imaging is based on the principle that when neurons are sufficiently depolarized, intracellular calcium rise, which can be detected using GECIs that bind to calcium (figure 1a). GECIs typically consist of a calcium-binding domain - calmodulin, calmodulin-binding peptide M1₃, and a reporter element which is based on either a single fluorescent protein or two fluorescent proteins (Broussard *et al.* 2014). In the case of a single fluorescent protein for example in GCaMPs, calmodulin (CaM) binds to the M1₃ peptide in the presence of calcium. This coupling results in conformational changes in the fluorescent protein, resulting in a change in fluorescence intensity (Nagai *et al.* 2001).

Two-photon calcium imaging provides several advantages over electrophysiology. First, two-photon calcium imaging is less invasive. Second, it can be combined with genetic tools (for example, Gal4-UAS, LexA-lexAop), to precisely target and record from a specific subset of neurons. Third, two-photon calcium imaging allows recording from several neuronal compartments including soma, dendrites, axons, or single spines and boutons (Grienberger *et al.* 2022). In this thesis, I used GCaMP6f (Chen *et al.* 2013) in combination with two-photon microscopy for recording neural activity.

TWO-PHOTON VOLTAGE IMAGING Despite the many advantages that two-photon calcium imaging offer, there are some disadvantages. Calcium imaging does not reveal inhibitory, hyperpolarizing signals. Also, calcium imaging is limited on the temporal scale. It is possible to overcome the limitations inherent in calcium imaging with optical voltage imaging. The genetically encoded voltage indicators (GEVIs) consist of a voltage-sensing domain fused together with a fluorescent protein. The coupling of voltage sensing with optical output is achieved either via Förster resonance energy transfer (FRET) between fluorescent proteins (FPs) or by sensitizing a single fluorescent protein by circular permutation.

The voltage indicators produce weak optical signals compared to calcium indicators GCaMPx, which is why few system neuroscience studies have been conducted on them. However, the potential of GEVIs is very high, and therefore a lot of effort is being put into improving the existing GEVIs and also developing new ones. Due to the low signal amplitude, experiments with optical voltage indicators such as ASAP2f have been challenging (Yang *et al.* 2016). In this thesis, I used a fluorescence protein voltage sensor called Arclight (Jin *et al.* 2012). Arclight is based on the fusion of the voltage sensing domain of *Ciona intestinalis* voltage-sensitive phosphatase (Murata *et al.* 2005) and the fluorescent protein super ecliptic pHluorin with an A227D mutation. Arclight's fluorescence decreases with membrane depolarization and increases with membrane hyperpolarization. Arclight has been shown to robustly report both subthreshold events and action potentials in genetically targeted neurons in the intact *Drosophila* brain (Cao *et al.* 2013). I used Arclight in combination with two-photon imaging to record changes in the neuronal membrane potential.

1.2.3 Manipulating neural activity

The above-mentioned tools are suitable for measuring the neural activity and characterizing a neuron. It is necessary, however, to also activate or silence a neuron in order to investigate its functional contribution to the neural circuit.

SILENCING NEURONS There are several genetic tools that allow for silencing neurons in *Drosophila* (figure 1b). First, the cell death genes such as *reaper* (*rpr*) or *head involution defective* (*hid*) can be expressed to induce apoptosis and kill the neurons (P. Chen *et al.* 1996; Grether *et al.* 1995). Second, the synaptic output of neurons can be permanently blocked. The *tetanus toxin light chain* (*TNT*) cleaves the synaptic vesicle protein synaptobrevin and inhibits neurotransmitter exocytosis at chemical synapses (Sweeney *et al.* 1995). Third, the expression of *Kir2.1* – an inwardly rectified potassium channel, can cause neurons to constantly hyperpolarize, resulting in suppressed excitability (Johns *et al.* 1999). While using the Gal4-UAS system to express these effector proteins provides effective control over the functionality of the targeted neurons, their expression cannot be reversed and the precise timing of silencing the neurons cannot be determined. To overcome these limitations, the conditional effector proteins like *shibire^{ts}* & *GtACR*, which is activated by higher temperature and light respectively can be used.

The *Drosophila* gene *shibire* encodes the protein *dynamin*, which is involved in the process of endocytosis and is essential for vesicle recycling. The dominant-negative temperature-sensitive allele *shibire^{ts}* is defective in synaptic vesicle recycling at the restrictive temperature (>29°C). The reuptake of vesicles from the synaptic cleft, mediated by the GTPase dynamin, is still functional at a permissive temperature (~25°C). Thus, a rapid and reversible inhibition of synaptic transmission can be achieved by controlling the temperature of the specimen (Kitamoto 2001). As an alternative to temperature, light can be used to control the timing of silencing the neuron. Light-gated anion channel, *Guillardia theta* anion channel rhodopsins (*GtACR*) can be expressed in the neurons of interest. The application of light causes these channels to open, allowing an influx of chloride ions, thus causing the neurons to hyperpolarize. This is an extremely sensitive, precisely timed, and reversible method for manipulating neural activity (Govorunova *et al.* 2015; Mauss *et al.* 2017; Mohammad *et al.* 2017).

ACTIVATING NEURONS A second or complementary approach to probing the functional role of a neuron in a neural circuit is by activating the neuron (figure 1c). Temperature-sensitive channels can be used for activating the neurons. Flies naturally express the transient receptor potential cation channel *TrpA1* which is implicated in temperature detection (Hamada *et al.* 2008; Pulver *et al.* 2009). The expression of these channels in the neurons allows for temperature-mediated excitation. The light-gated optogenetic tools, however, allow for greater temporal control compared to the above-mentioned temperature-sensitive method. The light-gated cation channel *Channelrhodopsin* (*ChR*) was extracted from the green algae *Chlamydomonas reinhardtii* and expressed in *C. elegans* and mammalian hippocampal neurons (Boyden *et al.* 2005; Nagel *et al.* 2005). By expressing these light-gated

cation channels in the neurons of interest, a precise, reversible, and reliable excitation of the neurons can be achieved.

1.3 NEURAL COMMUNICATION

Camillo Golgi's silver-stain method made it possible for the first time, to visualize single neurons in tissue samples under the light microscope (1873). Santiago Ramón y Cajal in 1888 described the nervous system as a network of individual cells. About a decade later, in 1897, the term 'synapse', derived from the Greek word 'synapsis' (meaning 'conjunction'), was used to describe the connections between two neurons. Neurons form networks where they communicate via synapses. Two types of synapses exist 1) electrical synapses and 2) chemical synapses.

1.3.1 Electrical synapses

In electrical synapses, two cells are directly connected by a cluster of intercellular channels called gap junctions (Bennett & Zukin 2004). The gap junctions provide a conductive pathway for electrical current to spread between cells. Consequently, electrical currents underlying action potentials or graded potentials directly propagate to postsynaptic neurons, without additional delay. Since electrical signals propagate bidirectionally, signalling events generated in the postsynaptic cells can also spread to presynaptic cells. In *Drosophila*, electrical synapses are widely distributed throughout the nervous system and are essential to neuronal function (Ammer *et al.* 2022; Liu *et al.* 2016; Stebbings *et al.* 2002).

1.3.2 Chemical synapses

Neurons communicate mostly via chemical synapses (figure 2). When the presynaptic membrane is sufficiently depolarized, voltage-gated calcium channels open and allow Ca^{2+} to enter the cell (Luo 2020). Calcium entry leads to the fusion of synaptic vesicles with the membrane and the release of neurotransmitter molecules into the synaptic cleft (Chapman 2002). As neurotransmitters diffuse across the synaptic cleft, they bind to receptors in the postsynaptic membrane, causing the postsynaptic neuron to depolarize or hyperpolarize, thereby passing the information from pre- to postsynaptic neurons (Di Maio 2008).

1.3.3 Voltage-gated ion channels

Voltage-gated ion channels are transmembrane proteins that allow certain inorganic ions to cross cell membranes (figure 3). Generally, these channels consist of two distinct but functionally coupled transmembrane domains: the voltage sensing domain and the pore domain. The voltage sensing domain changes the conformation of the pore domain in response to changes

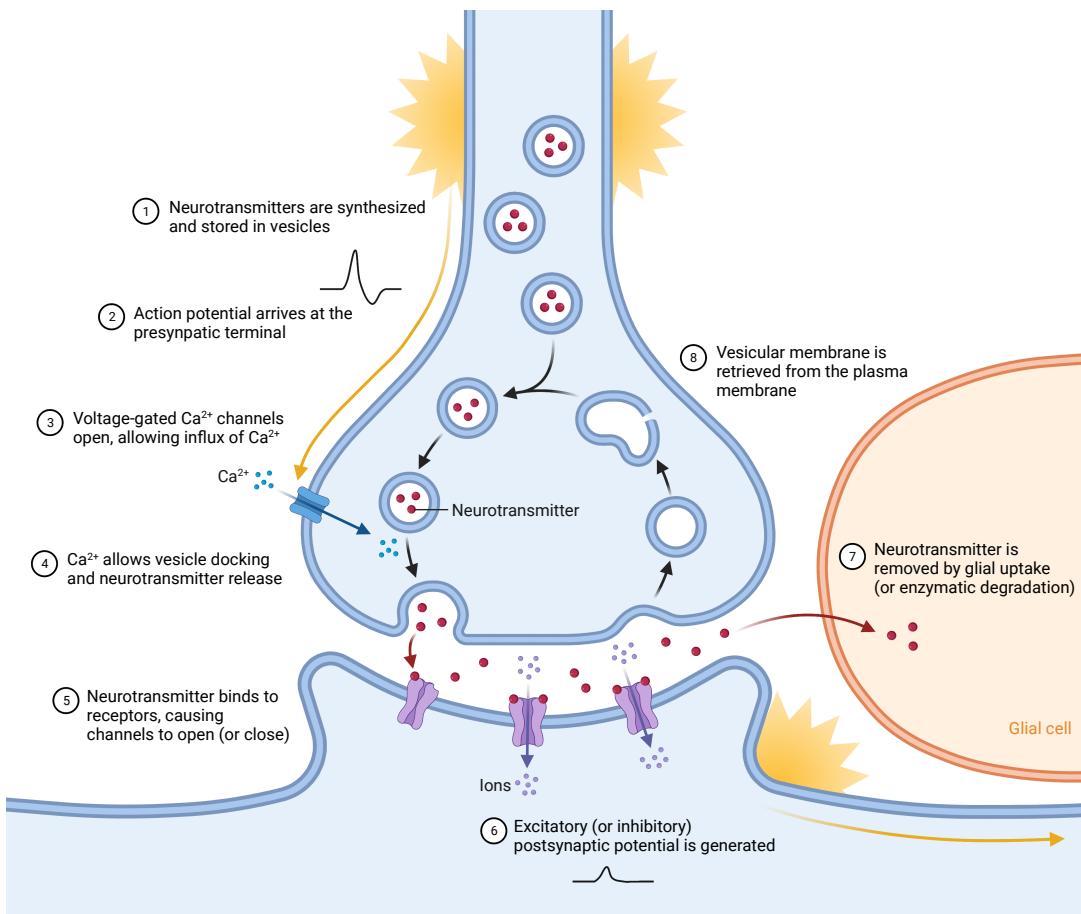


Figure 2: Chemical synapse: steps of synaptic transmission. (1) Synthesis and storage of neurotransmitters in the vesicles. (2) Depolarization in the presynaptic terminal causes (3) voltage-gated calcium channels to open and allow an influx of calcium ions. (4) High concentration of calcium ions triggers the fusion of neurotransmitters-filled vesicles with the presynaptic membrane and the release of neurotransmitters into the synaptic cleft. (5) Neurotransmitters released in the synaptic cleft bind to receptors in the postsynaptic membrane leading to (6) excitatory or inhibitory postsynaptic potential. (figure created with [Biorender.com](#))

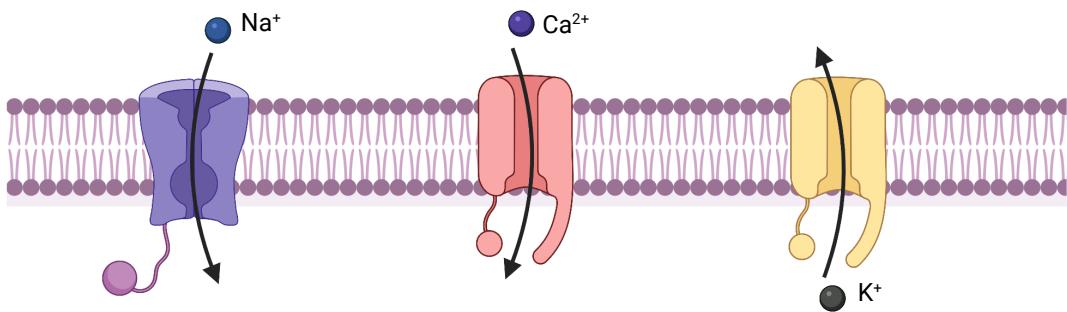


Figure 3: Voltage-gated ion channels: The sodium channels allow Na^+ ions to enter the cell. The calcium channels allow Ca^{2+} ions to enter the cell. The potassium channels allow efflux of the K^+ ions.

in transmembrane potential, allowing selected ions to flow down their electrochemical gradient.

VOLTAGE-GATED CALCIUM CHANNELS As mentioned above, voltage-gated calcium channels mediate depolarization-induced calcium influx that drives the release of neurotransmitters. The $\alpha 1$ -subunit of the voltage-gated calcium channels forms the ion-conducting pore, which makes it distinct from other calcium channels. Three families of genes encode $\alpha 1$ subunits. *Drosophila* genome has one $\alpha 1$ subunit gene in each family: $\alpha 1\text{D}$ (Ca_v1), cac (Ca_v2), and $\alpha 1\text{T}$ (Ca_v3) (King 2007; Littleton & Ganetzky 2000). In *Drosophila* antennal lobe projection neurons, cac (Ca_v2) type and $\alpha 1\text{T}$ (Ca_v3) type voltage-gated calcium channels are involved in sustained and transient calcium currents, respectively (Gu *et al.* 2009; Iniguez *et al.* 2013).

VOLTAGE-GATED SODIUM CHANNELS In neurons, voltage-gated sodium channels play a crucial role in the initiation and propagation of action potentials (Hodgkin & Huxley 1952). Sodium channels are activated and deactivated within milliseconds when the membrane is depolarized by a few millivolts. There are at least ten genes in mammals that encode these large membrane proteins. In contrast, *paralytic (para)* is the only voltage-gated sodium channel gene described in *Drosophila* (Piggott *et al.* 2019).

VOLTAGE-GATED POTASSIUM CHANNELS Voltage-gated potassium channels are transmembrane channels specific to potassium ions. They play a crucial role in repolarizing the depolarized cell to its resting membrane potential, after each action potential. Voltage-gated potassium channels are the most diverse family of voltage-gated ion channels in the human genome, with 40 members for α subunit grouped into 12 families (Gutman *et al.* 2005). The first voltage-gated potassium channel discovered in *Drosophila* was *Shaker* (Papazian *et al.* 1987). Afterwards, three additional *Shaker* like voltage-gated potassium genes were identified: *Shab*, *Shaw* and *Shal* (Covarrubias *et al.* 1991).

1.4 THE FLY VISUAL SYSTEM

As mentioned earlier, the *Drosophila melanogaster* nervous system consists of ~200,000 neurons (Raji & Potter 2021). Almost half of these neurons (~100,000 neurons) are dedicated to processing visual signals in the optic lobe of the fly brain. Unlike vertebrates, many invertebrate species have compound eyes, which are composed of multiple optical units called facets or ommatidia. Each compound eye contains around 750 ommatidia (Ready *et al.* 1976). The ommatidia are arranged in a regular lattice with a 5-degree inter-ommatidia angle (Land 1997). There are eight different photoreceptors in an ommatidium (R1-R8). A circular arrangement is formed by R1-R6 enclosing the central photoreceptors R7 and R8, which are stacked on top of one another. Due to this arrangement, the photoreceptors in an ommatidium sense incident light from slightly offset positions in space. With this configuration, six photoreceptors in six different adjacent ommatidia that possess identical optical axes project their axons to a single cartridge in the brain forming a so-called neuro-ommatidium (Strausfeld 1971). As a result of this neural superposition principle, the sensitivity increases without compromising the spatial resolution (Kirschfeld 1967).

PHOTOTRANSDUCTION The process of converting light energy into electrochemical signals is called phototransduction. This process occurs in rhabdomeres in the flies photoreceptors. In rhabdomeres, there are ~30,000 microvilli, each containing about 1000 photoactive molecules, called rhodopsin. The chromophore 3-hydroxy-11-cis-retinal is covalently bound to rhodopsin. Upon absorption of a photon by rhodopsin, the chromophore 3-hydroxy-11-cis-retinal is isomerized to all-trans-retinal, and the activated metarhodopsin state is formed. This activates a G-protein coupled cascade that results in the activation of phospholipase C (PLC). The PLC hydrolyzes phosphatidyl-inositol 4,5 bisphosphate (PIP₂) to diacylglycerol (DAG), inositol 1,4,5 trisphosphate (InsP₃), and a proton. Downstream to PLC, two light-sensitive channels (TRP and TRPL) are activated, allowing sodium and calcium to enter the cell and depolarize it. The photoreceptors upon activation release the inhibitory neurotransmitter histamine, thus inhibiting postsynaptic neurons (Hardie 1989; Hardie & Juusola 2015; Hardie & Raghu 2001).

1.4.1 The optic lobe

Following the photoreceptor layer in the retina, the fly's optic lobe consists of 4 layers of neuropils called the lamina, the medulla, the lobula, and the lobula plate. These neuropil layers are arranged in a columnar, retinotopic fashion with each column processing information from a small point in the visual space (figure 4a) (Fischbach & Dittrich 1989).

LAMINA The lamina is organized in an array of ~750 retinotopic columns (also called 'cartridges'). Each column corresponds to ~5° discrete sample of the visual world. The light-sensitive photoreceptors, R1-6 project their axons into each lamina column. Two other photoreceptors, R7 & R8 pass

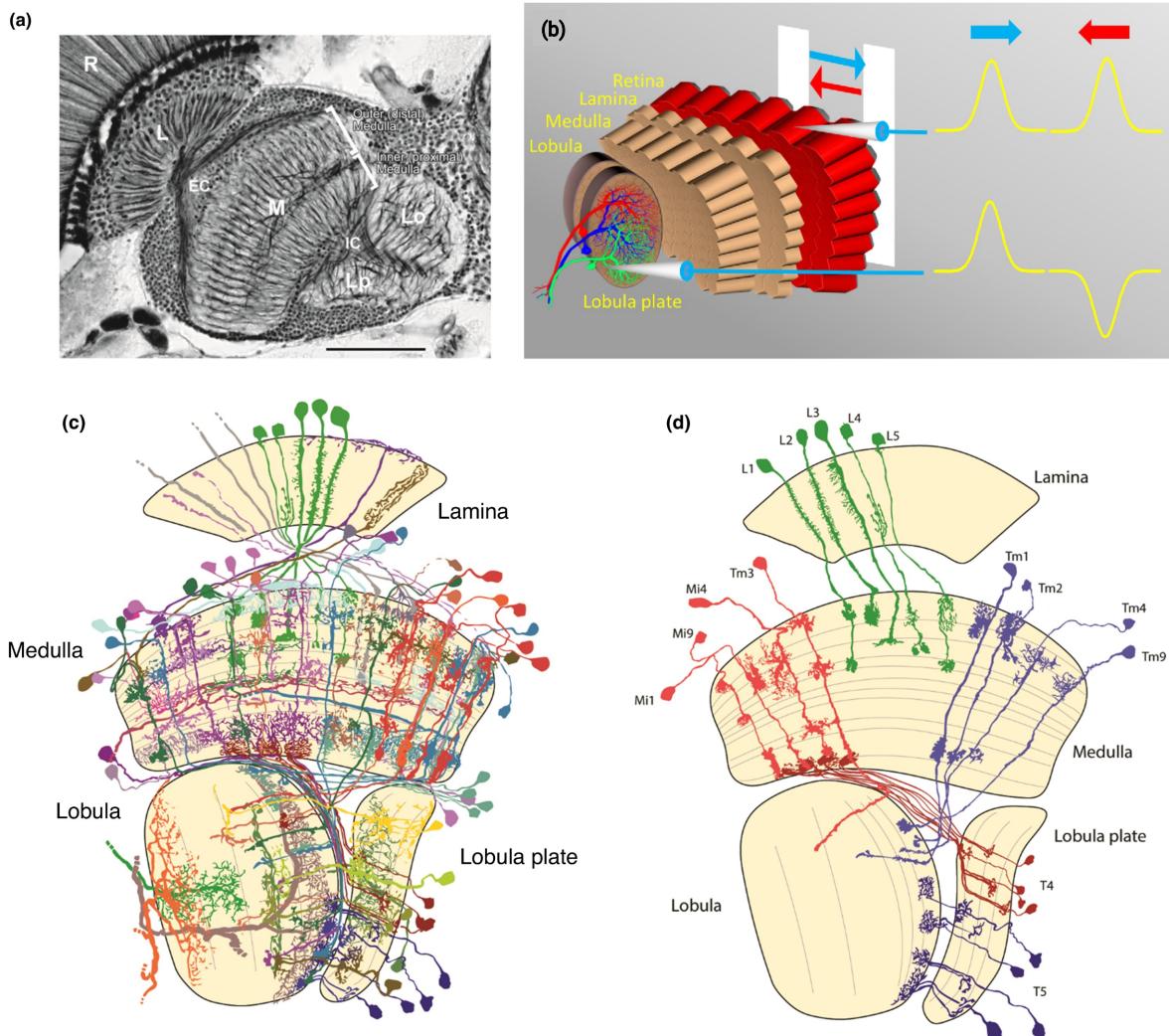


Figure 4: The fly optic lobe: (a) The horizontal cross-section of a reduced silver stain shows the columnar organization of the retina (R), lamina (L), external chiasm (EC), medulla (M), internal chiasm (IC), lobula (Lo), and lobula plate (Lp). Scale bar = 50 μm . Reproduced, with permission, from Takeuchi *et al.* 2008 (b) Schematic illustration of direction selectivity: moving a bar in front of a fly's eye leads to depolarization of photoreceptors every time, regardless of whether the bar moves to the right or left. It is a non-directional signal. A few synapses downstream, on the lobula plate tangential cells, signals are direction-selective: these cells depolarize during movement along one direction, i.e., their 'preferred' direction, and hyperpolarize during motion along the opposite direction, i.e., their 'null' direction. (c) An overview of all types of columnar cells in the *Drosophila* optic lobe. (Fischbach & Dittrich 1989) (d) Columnar cell types involved in the motion vision circuit. (Used with permission from Borst *et al.* 2020a,b)

through the lamina and synapse in specific layers of the medulla. Along with photoreceptor axons, the lamina includes 5 lamina output neurons (L₁-L₅), six putative feedback neurons (T₁, Lat, Law₁, Law₂, C₂, C₃), and one lamina intrinsic neuron (Lai). L₁, L₂, and L₃ cells receive the majority of their input from photoreceptors R₁-R₆. L₄ cells form reciprocal connections with L₂ and receive only a small number of input from photoreceptor R₆. L₅ receives input from L₂, L₄, and lamina interneurons (Rivera-Alba *et al.* 2011). The lamina columnar monopolar neurons, L₁-L₅ send their axonal projections into specific layers of the medulla. (Fischbach & Dittrich 1989; Tuthill *et al.* 2013). Lamina output neurons L₁ and L₂ are the primary input cells for motion vision (Zhu 2013).

MEDULLA Lamina cells send input projections to the medulla, the second neuropil in the optic lobe. There are ten synaptic layers (M₁ to M₁₀) in the medulla consisting of over 60 types of cells. The medulla is composed of hexagonal columns, similar to the lamina. In this way, the mapping between the lamina and medulla remains retinotopic. The fibers connecting the lamina to the medulla form a chiasm, in which posterior medulla cartridges receive input from anterior lamina cartridges. Two main classes of columnar interneurons are found in the medulla: around 10 types of medulla intrinsic neurons (Mi) and around 30 types of transmedullary neurons (Tm). Mi neurons connect different layers of the medulla to each other. Dendrites of Mi cells are located in the distal medulla layers, while axons are located in the proximal medulla layers. Tm neurons connect specific layers of the medulla to various layers in the lobula. Tm cells receive input in the distal medulla layers 1-5 from lamina monopolar cells and photoreceptors R₇ and R₈ and send their axonal projections to the lobula (Fischbach & Dittrich 1989; Takemura *et al.* 2011). In addition to these two types of neurons, the trans-medulla Y ('TmY') neurons connect specific layers of the medulla to various layers in the lobula and lobula plate.

LOBULA COMPLEX In the final stage of the visual processing in the optic lobe, the lobula complex consists of two neuropils: the lobula and the lobula plate. There are six layers in the lobula (Fischbach & Dittrich 1989). Lobula columnar (LC) neurons receive major inputs from the medulla and are the most prominent type of cell in the lobula. Multiple types of LC neurons span the entire visual field in a retinotopic manner (Otsuna & Ito 2006). In total, these neurons are divided into more than twenty distinct subtypes, each conveying information about a different visual feature (Wu *et al.* 2016). Lobula plates have four structurally distinct layers perpendicular to their columnar organization. A number of wide-field tangential cells are present in each layer (Strausfeld & Lee 1991). Dendritic trees of the tangential cells span large areas of the lobula plate, sometimes covering the entire layer. Thus, their receptive fields cover a large portion of the visual field. Two types of bushy T-cells T₄ and T₅ exist. Four subtypes of both T₄ and T₅ are found connecting the medulla and lobula respectively to the four layers of the lobula plate (Fischbach & Dittrich 1989).

1.4.2 Neural circuits underlying direction selectivity

Direction selectivity is the most important response characteristic of the lobula plate tangential cells. In response to a visual stimulus moving in their preferred direction, the cells depolarize. When the visual stimulus moves in the opposite direction (the null direction), the cells hyperpolarize. Two major types of lobula plate tangential cells have been described. The horizontal system (HS) cells respond preferentially to horizontal motion (Schnell *et al.* 2010), while vertical system (VS) cells respond preferentially to vertical motion (Joesch *et al.* 2008). Photoreceptor signals, in contrast, are not direction-selective, i.e. they display a similar response regardless of which direction the stimulus moves. Thus, a non-directional response at the photoreceptor level is transformed into a directional signal at the lobula plate tangential cell level (figure 4b). The lobula plate tangential cells, however, integrate signals over large parts of visual fields, i.e. they are not local motion detectors. Hence, the question arises: which cells are the local motion detectors?

Electrophysiological and two-photon calcium imaging experiments in the optic lobe over the years have revealed the following interesting results: (a) Visual processing in *Drosophila* occurs in two parallel processing pathways for luminance increment (ON) and luminance decrement (OFF) (Behnia *et al.* 2014; Eichner *et al.* 2011; Joesch *et al.* 2010, 2013; Shinomiya *et al.* 2014; Strother *et al.* 2014) (b) T₄ and T₅ are the first local motion detectors found in the *Drosophila* ON and OFF motion vision pathway respectively. Four sub-population of T_{4a-d} and T_{5a-d} cells tuned to the four cardinal directions and projecting to the four layers in the lobula plate is found within each column (Maisak *et al.* 2013).

PARALLEL ON AND OFF PROCESSING PATHWAYS In striking similarity to the mammalian retina (Masland 2012), visual processing in *Drosophila* occurs in two parallel ON and OFF processing pathways (Borst & Helmstaedter 2015). The ON pathway transmits information about luminance increments, while the OFF pathway transmits information about luminance decrements. The split into the ON and OFF pathways occurs in the lamina. The L₁ neurons provide inputs onto the ON pathway, while the L₂ neurons provide inputs onto the OFF pathway. When the output of L₁ neurons was genetically blocked, the downstream motion-sensitive lobula plate tangential cells no longer responded to ON motion. Blocking the output of L₂ neurons abolished the responses of the tangential cells to OFF motion (Joesch *et al.* 2010). In behavioral experiments, walking flies were unable to follow either ON or OFF motion when either L₁ or L₂ was blocked respectively (Clark *et al.* 2011; Maisak *et al.* 2013). The flies became completely motion-blind when both L₁ and L₂ were permanently hyperpolarized (via Kir2.1) (Bahl *et al.* 2013; Tuthill *et al.* 2013). These experiments together showed that the L₁ pathway specifically transmits information to the downstream ON motion detector, and the L₂ pathway specifically transmits information to the downstream OFF motion detector.

T₄ AND T₅ CELLS Based on previous studies (Buchner *et al.* 1984; Fischbach & Dittrich 1989), T₄ and T₅ were long thought to be the prime can-

didates for local motion detectors in the ON and OFF pathways respectively. However, due to its small size, it was difficult to do electrophysiological recordings from T₄ and T₅ cells (Douglass & Strausfeld 1996). This problem was solved using a combination of 2-photon imaging and a Gal4-UAS system to express GCaMP in T₄, and T₅ cells to record its neural activity in response to the ON and OFF stimuli. Stimulating the flies with visual motion in four cardinal directions (front-back, back-front, upwards, and downwards), Maisak *et al.* 2013 recorded direction-selective activity from T₄/T₅ cells. Four sub-populations of T_{4a-d} and T_{5a-d} cells tuned to the four cardinal directions and projecting to the four layers in the lobula plate were found within each column. Further, the T₄ cells were found to respond specifically to ON stimulus and the T₅ cells were found to respond specifically to OFF stimulus. Blocking T₄ and T₅ cells led to a complete loss of motion response in the lobula plate tangential cells (Schnell *et al.* 2012), and of the optomotor response of tethered walking flies (Bahl *et al.* 2013). Specific blocking of T₄ cells led to a reduction in LPTC and optomotor responses to ON stimulus selectively, while specific blocking of T₅ cells led to a reduction in LPTC and optomotor responses to OFF stimulus selectively. These results together show T₄ and T₅ cells to be the elementary motion detector for the ON and OFF pathways respectively (Maisak *et al.* 2013).

Having identified T₄ and T₅ cells as the elementary local motion detectors, the next question which arises is which cells provide synaptic inputs to T₄ and T₅ cells. Electron Microscopy (EM) studies (Shinomiya *et al.* 2019; Take-mura *et al.* 2017) provided the answer to this question. Shinomiya *et al.* 2019 used FIB-SEM (Focused Ion Beam Scanning Electron Microscopy) to record a volume of the optic lobe comprising seven columns of the medulla, lobula, and lobula plate. They identified all the different neuron types providing inputs to the T₄ and T₅ cells. T₄ cells receive input from Mi₁, Tm₃, Mi₄, Mi₉, C₃, CT₁, and TmY₁₅. T₅ cells receive input from Tm₁, Tm₂, Tm₄, Tm₉, CT₁, TmY₁₅, LT₃₃, and Tm₂₃. The T₄ and T₅ cells' dendrites span several columns along the preferred direction of the motion. The authors could also locate where the different cell types synapse onto the dendrites of T₄ and T₅. For example, a T_{4c} cell with the preferred direction of motion as upwards receives input from Mi₁, Tm₃, and TmY₁₅ in the central part of its dendrite, from Mi₉ and T_{4c} on the ventral part, and from Mi₄, C₃, and CT₁ on the dorsal part of its dendrite [figure 5 top]. T_{4d} cells with preferred direction as downwards receive input from Mi₁, Tm₃, and TmY₁₅ in the central part, from Mi₉ and T_{4d} on the dorsal part, and from Mi₄, C₃, CT₁ on the ventral part of its dendrite. In summary, all T₄ subtypes receive inputs from Mi₁, Tm₃, and TmY₁₅ in the central part, from Mi₉ on the preferred side (i.e. the side from which a preferred direction stimulus approaches), and from Mi₄, C₃, and CT₁ on the null side (i.e. the side from which a null direction stimulus approaches) of their dendrite. Similarly, all T₅ subtypes receive inputs from Tm₁, Tm₂, and Tm₄ on the central part, Tm₉ on the preferred side, and CT₁ on the null side of their dendrite [figure 5].

Most of these input elements have been characterised physiologically (Arenz *et al.* 2017; Behnia *et al.* 2014; Groschner *et al.* 2022; Meier & Borst 2019; Serbe *et al.* 2016; Strother *et al.* 2017). None of these cells were found to be direction-selective. Hence, we can conclude that the T₄ and T₅ cells are the elementary

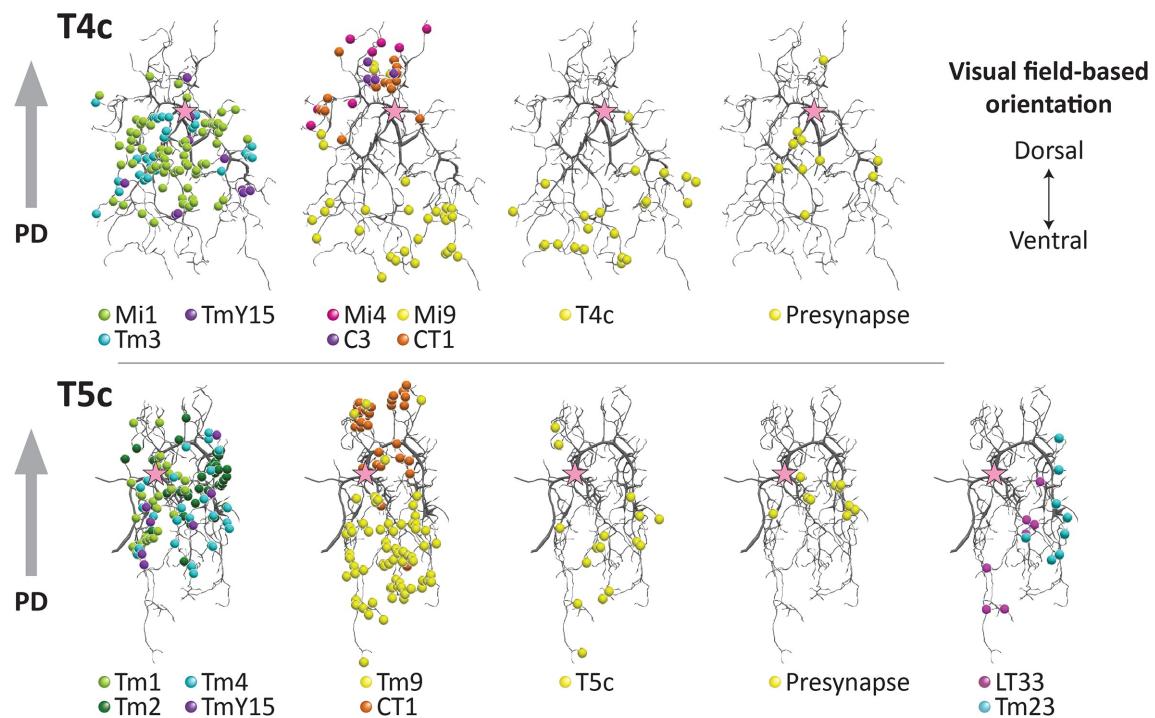


Figure 5: Synaptic sites distributed over T4 and T5 dendritic arbors: An arbor of a T4c (top panels) or T5c (bottom panels) cell is shown with synapse positions plotted on the dendritic arbors. Unless shown as 'Presynapse', puncta are postsynaptic sites (input to T4/T5 cells). T4c and T5c detect upward motion, and other subtypes of T4 and T5 cells show similar distribution patterns (not shown). An arbor's first branch point is indicated by pink stars. (Used with permission from Shinomiya *et al.* 2019)

motion detector found in the ON and OFF pathway respectively, and thus represents an important processing stage where the direction is computed.

1.4.3 Neural algorithms underlying direction selectivity

Different models have been proposed to explain the neural computations involved in motion detection. In order to detect motion in a directionally selective manner, local motion detection mechanisms must meet certain minimum requirements (Borst & Egelhaaf [1989](#)):

1. Spatial offset: Motion is a vector that needs two points to be represented, so at least two spatially separated inputs are required.
2. Temporal asymmetry: There must be at least one input that is delayed. If not, the input signals arrive in the subsequent stage simultaneously independent of the stimulus direction.
3. Non-linear interaction: It is necessary to integrate the input signals nonlinearly at a subsequent stage of the process. In the absence of this, the detector's output would be equal for both directions on average.

Classically two opposing models have been proposed for the implementation of direction selectivity. Both these models use two input lines, where one of the input lines has been asymmetrically delayed compared to the other, followed by a non-linear interaction. The Hassenstein-Reichardt (HR) model proposes a Preferred Direction (PD) enhancement: the signal on the preferred side is delayed and is subsequently amplified using multiplication of the signal from the other input line (figure [6a](#)) (Hassenstein & Reichardt [1956](#)). The Barlow-Levick (BL) detector, however, proposes a Null Direction (ND) suppression: the signal on the null side is delayed and divides the signal from the other input resulting in suppression (figure [6b](#)) (H. Barlow & Levick [1965](#)). Haag *et al.* [2016](#) used apparent motion stimuli to show that both the mechanisms i.e. PD enhancement on the preferred side and ND suppression on the null side are used by T4c and T5c cells to produce a direction-selective response (figure [6c](#)). Is that special to upward-tuned T4c cells or is it general for all subtypes of T4 and T5 cells. In the first manuscript [2.1](#) (Haag *et al.* [2017](#)), we showed that all four subtypes of T4 and T5 indeed use both PD enhancement and ND suppression to produce direction-selective responses. Therefore, a new model combining both PD enhancement on the preferred side and ND suppression on the null side was proposed. The next important task is to identify the neural correlates implementing these mechanisms.

The model requires a fast input at the center, slow input providing excitation on the preferred side, and slow input providing suppression on the null side. Interestingly, from the anatomical and functional characterization of the input data discussed earlier, we could predict the input neurons for T4 providing these three kinds of inputs. Mi1 is a fast neuron providing input at the central part of the dendrite, thus a candidate for central fast input. Mi9 is a slow neuron providing input on the preferred side of the dendrite, hence a candidate for input on the preferred side. Mi4, C3, and CT1 are slow neurons providing input on the null side of the dendrite (Arenz *et al.* [2017](#)).

Using two-photon voltage imaging in T5 neurons, Wienecke *et al.* 2018 showed that linear spatial summation is sufficient for the emergence of direction selectivity in T5 cells and that the preferred direction enhancement and null direction suppression in the calcium signal can arise from the non-linear voltage-calcium transformation. Using whole-cell recordings of T4, Gruntman *et al.* 2018 found that directional selectivity arises from simple integration of spatially offset fast excitatory and slow inhibitory inputs, thereby suppressing responses to nonpreferred motion directions. Groschner *et al.* 2022 recorded membrane potentials in T4 cells using whole-cell patch clamp recordings and in addition to the spatially offset inhibition, also found evidence for the preferred direction enhancement and a mechanism for multiplication like nonlinearity in T4 cells. In addition to synaptic mechanisms on the dendrites of T4 cells, further processing in the T4 neurons can enhance the direction selectivity of the output signals. In the second manuscript 2.2, we explored the transformation of voltage to calcium in T4c cells. We found that the voltage to calcium transformation in T4c neurons enhances their direction selectivity: calcium signals in T4c cells have a significantly higher direction selectivity and tuning compared to membrane voltage across different stimuli conditions.

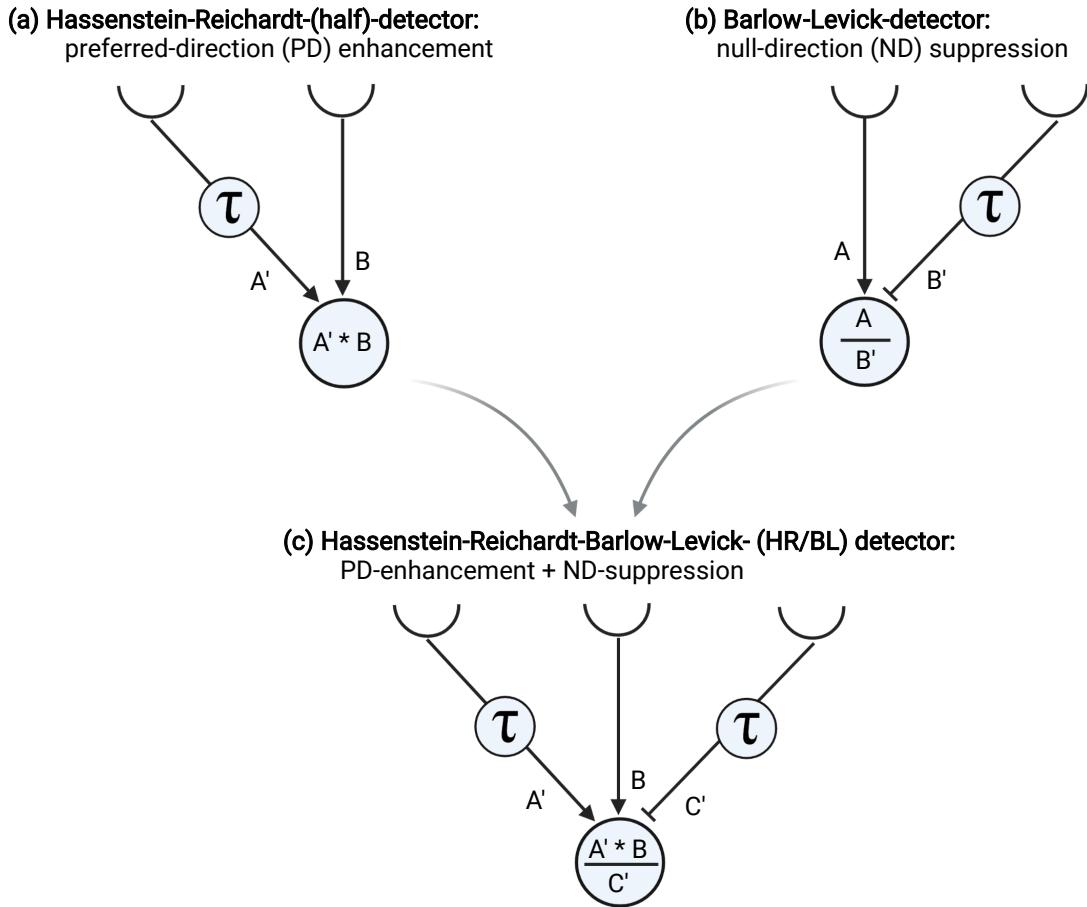


Figure 6: Models for motion detection: (a) The Hassenstein-Reichardt (HR) correlator (half-detector shown here) consists of two arms. Motion in the preferred direction (PD) causes two signals from neighboring photoreceptors to coincide due to a delay (τ) on the first arm. There is an enhancement in PD resulting from a multiplicative non-linearity. (b) The Barlow-Levick (BL) detector has the delay on the opposite arm, and the non-linearity is inhibitory, resulting in a null-direction (ND) suppression. (c) Hybrid detector consisting of one HR unit and one BL unit: Three points in space are sampled. There is a time delay (τ) on the outer two arms. The input signals from detector arms A and B are multiplied and divided by the signal from detector arm C in the following stage. Consequently, the signal in the preferred direction is enhanced and the signal in the null direction is suppressed.

2 | PUBLICATIONS

2.1 A COMMON DIRECTIONAL TUNING MECHANISM OF *Drosophila* MOTION-SENSING NEURONS IN THE ON AND IN THE OFF PATHWAY

ABSTRACT In the fruit fly optic lobe, T₄ and T₅ cells represent the first direction-selective neurons, with T₄ cells responding selectively to moving brightness increments (ON) and T₅ cells to brightness decrements (OFF). Both T₄ and T₅ cells comprise four subtypes with directional tuning to one of the four cardinal directions. We had previously found that upward-sensitive T₄ cells implement both preferred direction enhancement and null direction suppression (Haag et al., 2016). Here, we asked whether this mechanism generalizes to OFF-selective T₅ cells and to all four subtypes of both cell classes. We found that all four subtypes of both T₄ and T₅ cells implement both mechanisms, that is preferred direction enhancement and null direction inhibition, on opposing sides of their receptive fields. This gives rise to the high degree of direction selectivity observed in both T₄ and T₅ cells within each subpopulation.

AUTHORS Juergen Haag, Abhishek Mishra and Alexander Borst

CONTRIBUTIONS Juergen Haag, Conceptualization, Data curation, Software, Investigation, Visualization, Writing, review and editing; Abhishek Mishra, Investigation; Alexander Borst, Conceptualization, Funding acquisition, Writing original draft, Project administration, Writing, review and editing



A common directional tuning mechanism of *Drosophila* motion-sensing neurons in the ON and in the OFF pathway

Juergen Haag*, Abhishek Mishra, Alexander Borst

Max-Planck-Institute of Neurobiology, Martinsried, Germany

Abstract In the fruit fly optic lobe, T4 and T5 cells represent the first direction-selective neurons, with T4 cells responding selectively to moving brightness increments (ON) and T5 cells to brightness decrements (OFF). Both T4 and T5 cells comprise four subtypes with directional tuning to one of the four cardinal directions. We had previously found that upward-sensitive T4 cells implement both preferred direction enhancement and null direction suppression (Haag et al., 2016). Here, we asked whether this mechanism generalizes to OFF-selective T5 cells and to all four subtypes of both cell classes. We found that all four subtypes of both T4 and T5 cells implement both mechanisms, that is preferred direction enhancement and null direction inhibition, on opposing sides of their receptive fields. This gives rise to the high degree of direction selectivity observed in both T4 and T5 cells within each subpopulation.

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Introduction

The direction of visual motion is crucial for fundamental behaviors such as mate detection, prey capture, predator avoidance and visual navigation. This important visual cue, however, is not explicitly encoded at the output of a single photoreceptor but rather has to be computed by subsequent neural circuits. In order to extract local, directional information from moving images, mainly two competing algorithmic models of motion detectors have been proposed (*Figure 1a,b*). Both models implement a delay-and-compare mechanism where two input signals from neighboring image pixels interact in a nonlinear way after one of them has been delayed with respect to the other. This leads to an output that is larger for motion along one, the so-called ‘preferred’ direction than for the opposite, the so-called ‘null’ direction. Both models differ, however, by the type of non-linearity employed and the location of the delay. In the Hassenstein-Reichardt detector (*Figure 1a*), the delay is on the preferred side, that is where a preferred direction stimulus is entering the receptive field of the detector, and the non-linearity is excitatory. This leads to an enhancement of signals moving in the preferred direction (Hassenstein and Reichardt, 1956). In the Barlow-Levick detector (*Figure 1b*), the delay is on the null side, that is where a null direction stimulus is entering the receptive field of the detector, and the nonlinearity is inhibitory. This leads to a suppression of signals moving in the null direction (Barlow and Levick, 1965). While the predictions of both models concerning the responses to smooth grating motion are identical, apparent motion stimuli lend themselves well to discriminate between them (Egelhaaf and Borst, 1992; Eichner et al., 2011). Instead of moving an object smoothly across the image plane, an apparent motion stimulus consists of a bright or dark bar or spot that is abruptly jumped from one location to an adjacent one. Comparing the responses of directional neurons to the sequence with the sum of the responses to each individual stimulus presentation (‘linear expectation’) allows one to calculate the nonlinear response component as the difference between the sequence response and the linear expectation. If this nonlinear response component is positive for sequences along the preferred direction, and zero for

*For correspondence:
haag@neuro.mpg.de

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sequences along the null direction, a preferred direction enhancement is at work, supporting the Hassenstein-Reichardt model (*Figure 1a*). If the nonlinear response component is zero for sequences along the preferred direction and negative for sequences along the null direction, a null direction suppression is at work, supporting the Barlow-Levick model (*Figure 1b*). In the following, we will apply this approach in order to investigate which of the two mechanisms is at work in primary motion-sensitive neurons of the fruit fly *Drosophila*.

In *Drosophila*, visual signals are processed in the optic lobe, a brain area comprised of the lamina, medulla, lobula, and lobula plate, each arranged in a columnar, retinotopic fashion (for review, see: *Borst, 2014; Behnia and Desplan, 2015*). In striking parallel to the vertebrate retina (*Borst and Helmstaedter, 2015*), the direction of visual motion is computed within the optic lobe separately in parallel ON and OFF motion pathways (*Joesch et al., 2010; Reiff et al., 2010; Eichner et al., 2011; Joesch et al., 2013*). Anatomically, these two pathways split at the level of the lamina (*Bausenwein et al., 1992; Rister et al., 2007*) and lead, via a set of various intrinsic medulla and transmedulla interneurons, onto the dendrites of T4 and T5 cells, respectively. First described by Golgi staining (*Cajal and Sanchez, 1915; Strausfeld, 1976; Strausfeld and Lee, 1991; Fischbach and Dittrich, 1989*), T4 cells extend their dendrites in the most proximal layer of the medulla, while the dendrites of T5 cells are located in the inner-most layer of the lobula. There exist generally four T4 and 4 T5 cells per column (*Mauss et al., 2014*). The four subtypes of T4 cells respond selectively to brightness increments moving along one of the four cardinal directions, the four subtypes of T5 cells selectively to brightness decrements moving along the same four cardinal directions as T4 cells (*Maisak et al., 2013*). According to their preferred direction, T4 and T5 cells project into one of the four lobula plate layers (layer 1, most frontal: front-to-back; layer 2: back-to-front; layer 3: upward; layer 4, most posterior: downward; *Maisak et al., 2013*). There, T4 and T5 cells provide direct excitatory cholinergic input onto the dendrites of wide-field, motion-sensitive tangential cells as well as onto glutamatergic lobula plate interneurons that inhibit wide-field tangential cells in the adjacent layer (*Mauss et al., 2014; Mauss et al., 2015*). Through this circuit arrangement, lobula plate tangential cells depolarize to motion in their preferred direction (PD) and hyperpolarize in response to motion in the opposite or null direction (ND) (*Joesch et al., 2008; Schnell et al., 2010*). With T4 and T5 cells blocked, tangential cells lose all their direction selectivity (*Schnell et al., 2012*) and flies become completely motion-blind (*Bahl et al., 2013; Schilling and Borst, 2015*). This suggests that T4 and T5 cells are the elementary motion detectors and carry all directional information in the fly brain. Electrophysiological (*Behnia et al., 2014*), optical voltage (*Yang et al., 2016*) and Calcium recordings (*Meier et al., 2014; Serbe et al., 2016; Arenz et al., 2017; Strother et al., 2014; Strother et al., 2017*) from presynaptic medulla neurons revealed that none of them is directionally selective. Therefore, T4 and T5 cells are the first neurons in the visual processing chain that respond to visual motion in a direction selective manner.

Previous studies analyzed the mechanism underlying direction selectivity in T4 and T5 cells, yet arrived at different conclusions. Using apparent motion stimuli, one study found preferred direction enhancement to account for directional responses in T4 cells (*Fisher et al., 2015*). For T5 cells, the authors reported both enhancement for preferred and suppression for null direction sequences, but attributed the latter to circuit adaptation and not to the mechanism generating direction selectivity. The authors concluded that the dominant interaction producing direction selective responses in both T4 and T5 cells is a nonlinear signal amplification (*Fisher et al., 2015*). This conflicts with another report where spatio-temporal receptive fields of T5 cells were measured using white noise stimulation and reverse correlation. Based on ON and OFF subfields tilted in the space-time plane, T5 cells were concluded to incorporate both preferred direction enhancement and null direction suppression (*Leong et al., 2016*). This interpretation, however, suffers from a possible confusion of ON and OFF receptive subfields of T5 input neurons with the mechanism generating direction selectivity within T5 cells themselves. Addressing the same question, we recently applied apparent motion stimuli to one class of T4 cells that have upward as their preferred direction and, thus, project to layer 3 of the lobula plate. Using a telescopic stimulation technique to place the stimulus precisely onto the hexagonal lattice of the fly's eye (*Kirschfeld, 1967; Braitenberg, 1967; Franceschini, 1975; Schuling et al., 1989*), layer 3 T4 cells turned out to implement both mechanisms within different

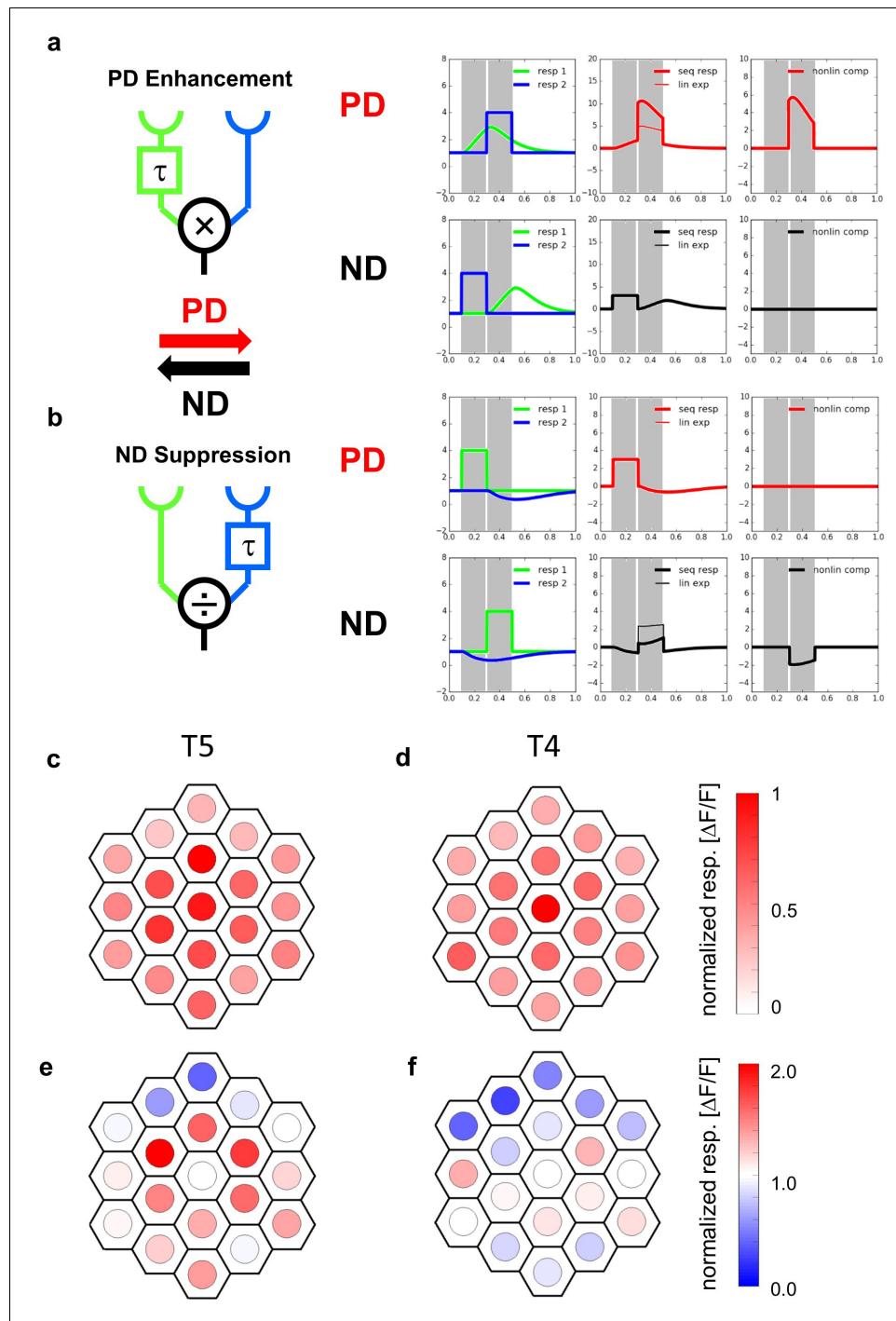


Figure 1. Receptive fields and responses to apparent motion stimuli of T5-cells. (a) The Hassenstein-Reichardt model incorporates PD enhancement only, realized by a multiplication. Left: Responses to individual light pulses ('Flicker') delivered at the two different positions. The responses are shifted according to the stimulus sequence used for the subsequent apparent motion stimuli. Middle: Responses of the model to apparent motion stimuli in preferred (upper row) and null direction (lower row, thick line = measured response, thin line = linear expectation, that is sum of responses to the single

Figure 1 continued on next page

Figure 1 continued

light pulses). Right: Nonlinear response component defined as the difference between measured response and linear expectation. (b) same as (a) but for a Barlow-Levick model. This model incorporates ND-suppression only, realized by a division. (c) Average responses of five T5 cells to flicker stimuli (stimulus size: 5 degree) delivered to different optical columns. In order to average the responses of different flies, the response patterns were aligned and normalized with respect to the maximum response (central column) and shown in a false color code. (d) Same as (c) but for T4-cells. Data represent the mean of 10 T4-cells from 10 flies (from [Haag et al., 2016](#)). (e) Responses of T5 cells to stimuli presented to the central column and simultaneously to one of the columns of the two surrounding rings. As in c, the responses of different flies were aligned with respect to the column eliciting the maximum response when stimulated individually and normalized to it. Depending on the location, simultaneous stimulation of a second column led to either a suppressed (blue colors) or an enhanced (red colors) response compared to the exclusive stimulation of the central column. The suppression is stronger on the null side of the T5 cells. Data represent the mean of 6 T5 cells from 6 different flies. (f) Same as e) but for T4-cells. Data represent the mean of T4-cells from 8 flies (from [Haag et al., 2016](#)).

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The following figure supplements are available for figure 1:

Figure supplement 1.

DOI: <https://doi.org/10.7554/eLife.29044.003>

Figure supplement 2. Responses of T4-cells and T5 cells to stimuli presented to the central column and simultaneously to one of the columns of the two surrounding rings.

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parts of their receptive field ([Haag et al., 2016](#)): While preferred direction enhancement was found to be dominant within the ventral part of the receptive field, a null direction suppression was significant in the dorsal part of their receptive field ([Haag et al., 2016](#)).

To resolve the conflicting evidences mentioned above and to test whether T5 cells are using the same or a different mechanism to compute the direction of motion as do T4 cells, we used the same strategy as in our previous account ([Haag et al., 2016](#)) and applied it to investigate the mechanism underlying direction selective responses of both T4 and T5 cells of all four directional tuning subtypes.

Results

In a first set of experiments, we used the same driver line as in our previous study ([Haag et al., 2016](#)) expressing the Calcium indicator GCaMP6m ([Chen et al., 2013](#)) in both T4 and T5 cells projecting to layer 3 of the lobula plate and, hence, having upward motion as their preferred direction. Since T5 cells are known to be OFF sensitive, we used dark spots on a bright background projected onto the raster of optical columns via a telescope to stimulate the cells and recorded the fluorescence changes in the lobula plate. We started by measuring the flicker responses of T5 cells to optical stimulation of 19 individual columns, forming two rings surrounding a central column. In [Figure 1c](#), the responses of five T5 cells were averaged and are shown in false color code overlaid on the columnar raster. T5 cells responded maximally to the stimulation of the central column, with about 50–100% amplitude to stimulation of the surrounding columns and about 20–50% to the next outer ring. An individual example trace and statistical evaluation of the responses are shown in [Figure 1—figure supplement 1](#). Compared to T4 cells ([Figure 1d](#), data replotted from [Haag et al., 2016](#)), the receptive field of T5 cells turned out to be broader with a somewhat stronger sensitivity within the surrounding columns. To explore spatial interactions within the receptive field of T5-cells, we stimulated the central column, simultaneously with one of surrounding columns. The results ([Figure 1e](#)) indicate a strong suppression of the response in the dorsal part of the receptive field compared to when the central column was stimulated alone, similar to what was found previously for T4 cells ([Figure 1f](#), data replotted from [Haag et al., 2016](#)).

Experiments performed on layer 3 T4 cells with apparent motion stimuli revealed different mechanisms of direction selectivity in different parts of the receptive field ([Haag et al., 2016](#)): Two-pulse apparent motion stimuli in the dorsal part of the receptive field led to a null direction suppression, apparent motion stimuli in the ventral part evoked preferred direction enhancement. We asked whether we could find this spatial arrangement of null direction suppression and preferred direction enhancement in T5 cells as well. In order to measure that, we presented OFF stimuli to four neighboring columns along the dorsal-ventral axis ([Figure 2a,b](#)). The columns were chosen in relation to

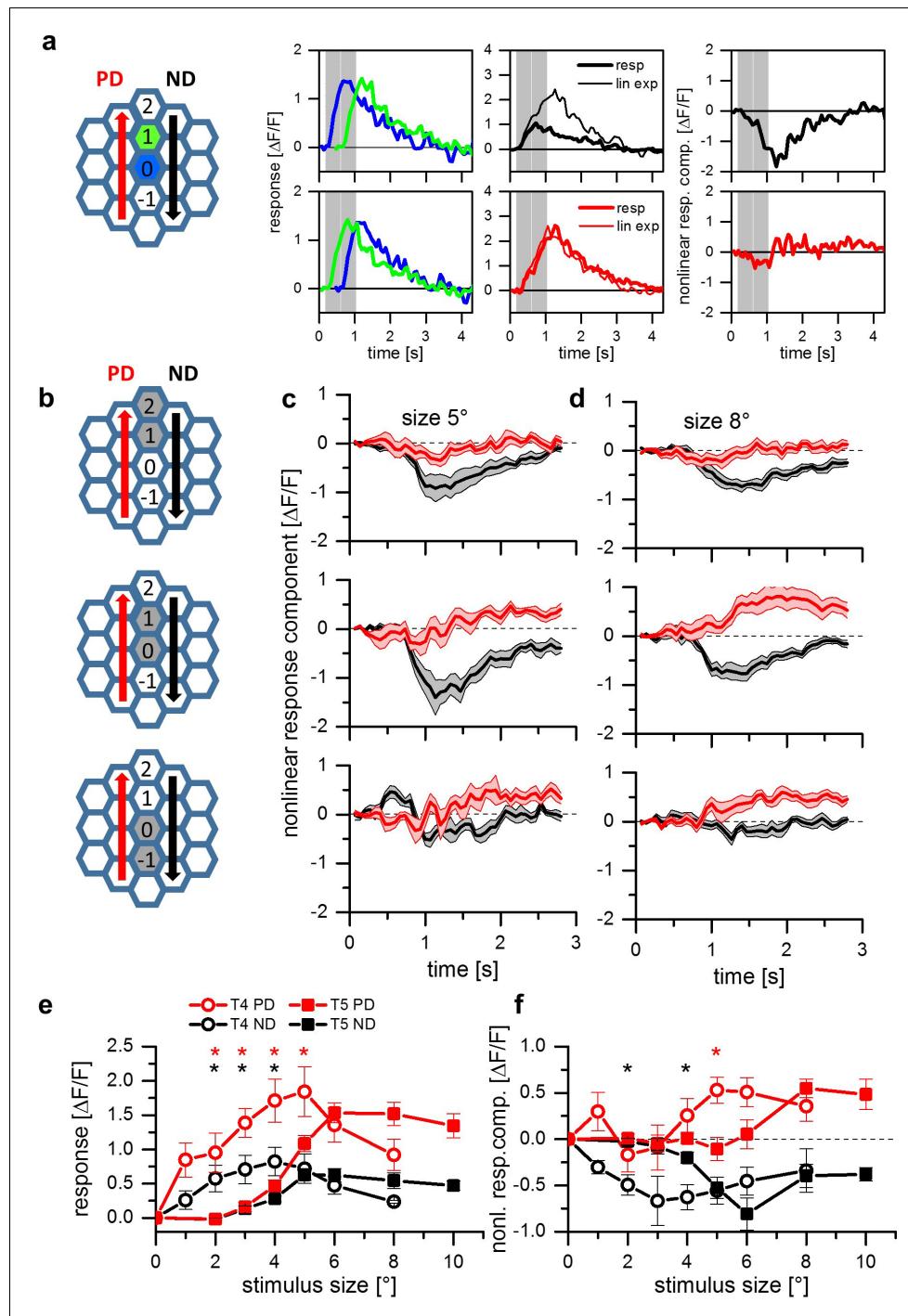


Figure 2. Apparent motion stimuli between adjacent cartridges. (a) Response of a single T5 cell recorded in a single sweep to two-step apparent motion stimuli. The schematic to the left shows the position of the two stimuli (blue and green shading). Left: Responses to individual light pulses ('Flicker') delivered at the two different positions. The responses are shifted in time according to the stimulus sequence used for the subsequent apparent motion stimuli. Middle: Responses of T5 to apparent motion stimuli in preferred and null direction (thick line = measured response, thin

Figure 2 continued

line = linear expectation, that is sum of responses to the single light pulses). Right: Nonlinear response component defined as the difference between measured sequence response and linear expectation. The responses are the mean obtained from $n = 3$ stimulus repetitions. (b) Two-step apparent motion stimuli were shown at three different position in the receptive field of T5 cells. The stimulus consisted of light off pulses positioned on one column for 472 ms, immediately followed by a light off pulse for 472 ms to the upper, neighboring cartridge. The same stimuli were repeated along the opposite direction. (c) Nonlinear response component, that is the difference between sequence response and the sum of the responses to the individual pulses, as a function of time for a stimulus size of 5 degree. Apparent motion stimuli delivered to the upper two cartridges resulted in a null direction suppression and no preferred direction enhancement. Apparent motion stimuli in the lower cartridges did not lead to a deviation from the linear expectation. For all three stimuli no preferred direction enhancement could be found. Data represent the mean \pm SEM in 6 T5-cells measured in 6 different flies. (d) Same as d, but with a stimulus size of 8 degree. In contrast to the results for a smaller stimulus size, we found preferred direction enhancement for stimulation of the lower and the central pair of columns. Data shows the mean \pm SEM in 10 T5-cells measured in 8 different flies. (e) Responses of T4 (open circles) and T5 cells (closed squares) to apparent motion stimuli in preferred (red colors) and null direction (black colors) between the two central columns 0 and 1. Compared to the responses of T4, T5 responses to two-pulse sequences along the preferred (PD) and the null (ND) direction are shifted to larger stimulus sizes. Data represent the mean values \pm SEM of 10 T5 cells measured in 5 flies and of 7 T4 cells in 4 different flies, respectively. Black asterisks represent statistically significant (*t*-test, p -value < 0.05) differences for null-direction responses of T4-cells and T5-cells, red asterisks for preferred direction responses. (f) Nonlinear response components of T4 and T5 cells. Same dataset as in **Figure 1e**.

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the column that elicited the strongest response in each cell (central column 0). We then tested T5 cells with two consecutive light OFF pulses of 472 ms duration in immediate succession. Each light pulse was positioned on one of two neighboring columns resulting in three stimulus sequences (**Figure 2b**). To extract the nonlinear response component, we subtracted the sum of the responses to the individual stimuli from the response to the apparent motion sequence. Example traces from an individual experiment stimulating columns 0 and 1 are shown in **Figure 2a**. Using the same spot diameter as in our previous account, that is 5 degree, we found null direction suppression for stimulation of the central (column 0 and 1) and the dorsal (column 1 and 2) pairs, and only a slight, if any, sign of preferred direction enhancement for stimulation of the ventral pair (column -1 and 0) (**Figure 2c**). This changed when we enlarged the spot size from 5 to 8 degree. Now, in addition to null direction suppression for the central and dorsal stimulus pairs, preferred direction enhancement for the ventral and central stimulus pairs was observed (**Figure 2d**). This result mirrors our previous finding for T4 cells where both null direction suppression and preferred direction enhancement was found to account for direction selectivity (Haag et al., 2016). In further agreement with T4 cells, these two mechanisms are spatially separated, with null direction suppression on the null and preferred direction enhancement on the preferred side of the receptive field (Haag et al., 2016).

The above experiments indicate a different dependence of null direction suppression and preferred direction enhancement on the diameter of the stimulus spot in T5 cells. To measure this dependence in a gradual way, we again used apparent motion stimuli and varied the size of the stimuli from 1 to 10 degree. Since stimuli centered on the central column pair (0 and 1) resulted in both types of nonlinearity (**Figure 2d**, middle graph), we presented apparent motion stimuli with different stimulus sizes to these central columns only. To compare the results of T5 cells with the ones of T4 cells, the stimulus set consisted of either bright pulses on a dark background (for T4 cells) or dark pulses on a bright background (for T5 cells). **Figure 2e** shows the responses of T4 (circle symbols) and T5 (square symbols) to apparent motion stimuli as a function of the stimulus size for preferred (PD, red traces) and null (ND, black traces) direction sequences. For both directions of motion, T4 cells respond to smaller stimuli than T5 cells. The strongest response in T4 can be found for stimulus sizes of 4 to 5 deg. For stimulus sizes beyond these values, the responses of T4 cells decline. In contrast, T5 cells only start responding at these stimulus sizes and plateau for larger values. When instead of the response the nonlinear response component is plotted as a function of the stimulus size (**Figure 2f**, same symbol and color code as in **Figure 2e**), both preferred direction enhancement and null direction suppression become apparent, with both curves shifted to larger stimulus sizes for T5 cells. Furthermore, for both cell types, null direction suppression peaks at smaller stimulus sizes than preferred direction enhancement.

The results presented so far point towards a common mechanism for T4 and T5 cells underlying direction selectivity. However, the experiments on T4 and T5 cells were confined to those that terminate in layer 3 of the lobula plate. In order to investigate whether the properties described above

generalize to T4 and T5 cells of all four tuning categories, we next used a fly line expressing GCaMP5 in T4 and T5 cells projecting to all four layers. Similar to the experiments shown in **Figure 2c and d**, two-step apparent motion stimuli consisting of ON-ON pulses of 5 degree diameter for T4 cells as well of OFF-OFF pulses of 8 degree diameter for T5 cells were presented in three adjacent pairs of columns aligned to the column that elicited the strongest flicker response (**Figure 3a**). As before, the stimulus protocol consisted in the presentation of individual stimulus pulses for the calculation of the linear expectation as well as in the presentation of the two-pulse sequences, to measure the sequence response. From the latter, the linear expectation was subtracted to obtain the nonlinear response component. The time traces of these nonlinear response components are shown in **Figure 3b and c**. For T4 (**Figure 3b**) and T5 (**Figure 3c**) cells projecting to all layers, we found both preferred direction and null direction suppression, with a spatial separation that follows the same pattern: on the preferred side of the receptive field, a clear preferred direction enhancement was observed without any null direction suppression (**Figure 3b and c**, left column). In the center of the receptive field, both preferred direction enhancement and null direction suppression prevailed (**Figure 3b and c**, center column). On the null side of the receptive field, only null direction suppression was detectable (**Figure 3b and c**, right column). To investigate possible differences between T4 and T5 cells and between cells with different directional tuning, we performed a 3-way ANOVA test. Choosing a significance level of $p=0.05$, no significant differences were found, neither between T4 and T5 cells, nor between the neurons projecting to the four different layers. In **Figure 3d** (T4 cells) and **3e** (T5 cells), the nonlinear response components are shown as averaged between 1 and 2 s of the time courses shown above, as well as averaged across the cells from all four layers. On these data, two-sided t-tests were performed between T4 and T5 cell responses for each individual stimulus condition. Choosing again a significance level of $p=0.05$, no differences were found between T4 and T5 cell responses for 5 out of 6 stimulus conditions. Only the response amplitude of T4 cells to null direction stimulus sequences from column 2 to 1 was found to be significantly smaller than the respective value of T5 cells.

The results from apparent motion experiments reported so far suggest a common directional tuning mechanism for T4 and T5 cells for all cardinal directions. One, thus, would expect identical high degrees of direction selectivity in response to moving gratings mechanism within the different layers of the lobula plate. To test this directly, we used the same fly line as above expressing in both T4 and T5 cells of all four layers and presented grating motion along all four cardinal directions on a screen. For each pixel, we first calculated the vector sum of the responses and represented the vector angle in false color. The resulting image from one example fly is shown in **Figure 4a**. Clearly, the preferred direction is extremely homogeneous with little variation within each layer. We repeated such experiments in five different flies and determined the distribution of all preferred directions obtained from the whole data set. The histogram (**Figure 4b**) reveals exactly four sharp peaks separated by 90 degrees, corresponding to the preferred directions of T4 and T5 cells within each of the four layers. This transition from the hexagonal coordinates of the fly eye to Cartesian coordinates is likely to occur on the dendrites of T4 and T5 cells by their sampling from appropriately grouped columns ([Takemura et al., 2017](#)). From the same data set, we calculated a direction selectivity index for each pixel within each layer as the difference of the responses to preferred and null direction, divided by the sum of the responses. We then determined the mean direction selectivity for each layer from each fly and averaged the resulting values across the different experiments. The results reveal an extremely high degree of direction selectivity of about 0.8 that is almost identical within each layer (**Figure 4c**). To measure direction selectivity separately for T4 and T5 cells, we stimulated the flies with ON and OFF edges instead of gratings and obtained similar values of about 0.8 on average (**Figure 4d**).

Discussion

Having analyzed the mechanisms underlying direction selectivity of all four subtypes of both T4 and T5 cells, we found a common scheme that pertains to all of these cells: regardless of the directional tuning and the contrast preference for ON or OFF stimuli, elementary motion-sensitive neurons in *Drosophila* implement a preferred direction enhancement on the preferred side and a null direction suppression of input signals on the null side of their receptive field.

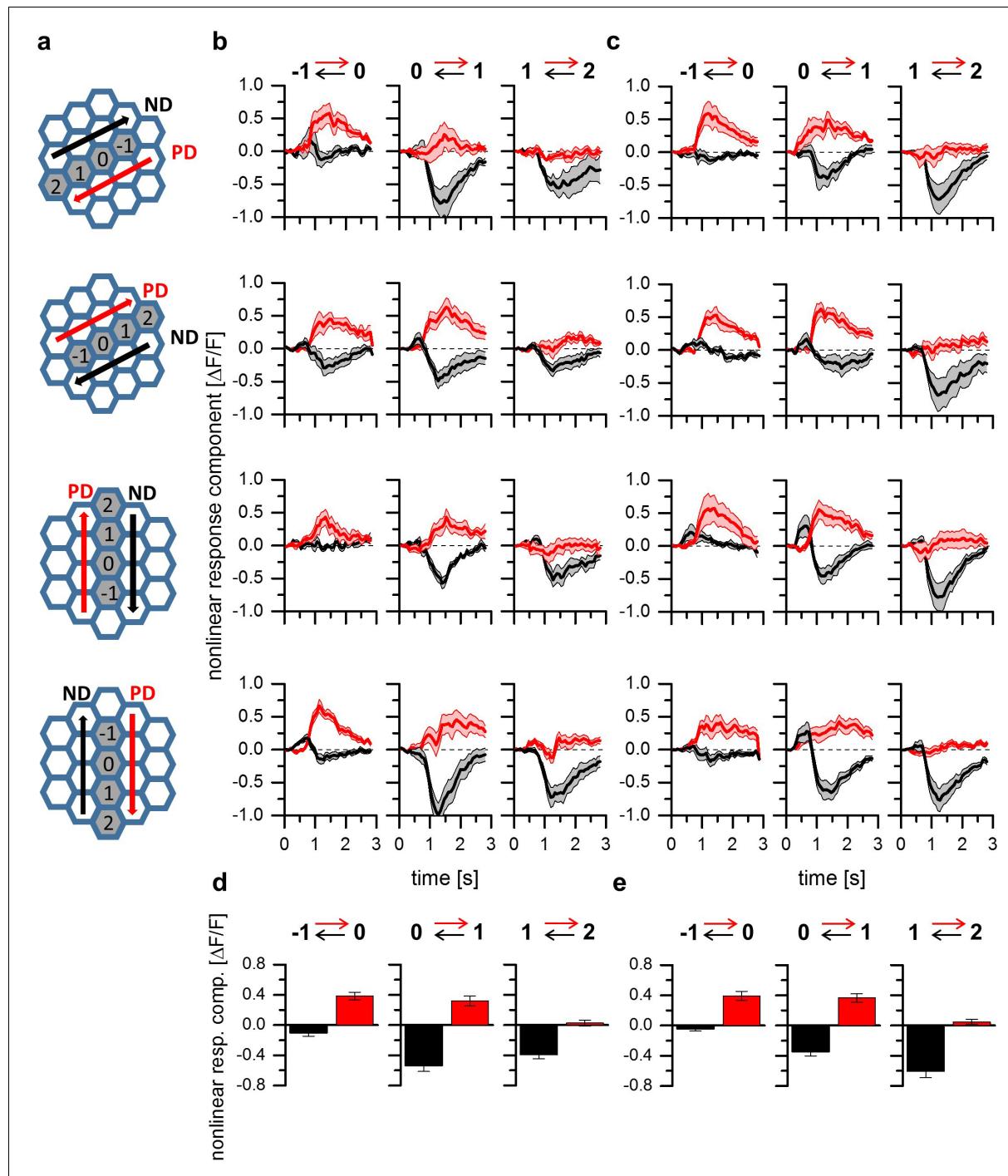


Figure 3. A common mechanism for direction selectivity in all four subtypes of T4 and T5 cells. **(a)** Pictograms indicating the stimulus positions and the preferred and null-direction of the respective layer. **(b)** Nonlinear response components of T4-cells to apparent motion stimuli in different layers of the lobula plate. For T4 cells projecting to all four layers, preferred direction enhancement and null direction suppression are found to be spatially distributed within the receptive field such that enhancement is found on the preferred side while suppression is predominant on the null side of the Figure 3 continued on next page

Figure 3 continued

receptive field. Data represent the mean \pm SEM of 6, 8, 7 and 9 T4 cells (from layer 1–4). (c) Nonlinear response components of T5 cells to apparent motion stimuli in different layers of the lobula plate. Data represent the mean \pm SEM of 8, 5, 6 and 13 T5 cells (from layer 1–4).

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ON and OFF pathways seemed to have adapted to the asymmetry of luminance distributions found in the real world. Consequently, functional differences between ON and OFF pathways have been described in the mammalian retina and in flies as well (Ratliff et al., 2010; Clark et al., 2014; Baden et al., 2016; Leonhardt et al., 2016). In fly motion vision, our finding of a common mechanism for T4 and T5 cells suggests the above mentioned asymmetries to rely on quantitative instead of qualitative differences, such as different time-constants used by the ON and the OFF pathway (Leonhardt et al., 2016; Arenz et al., 2017). One difference between T4 and T5 cells found in this

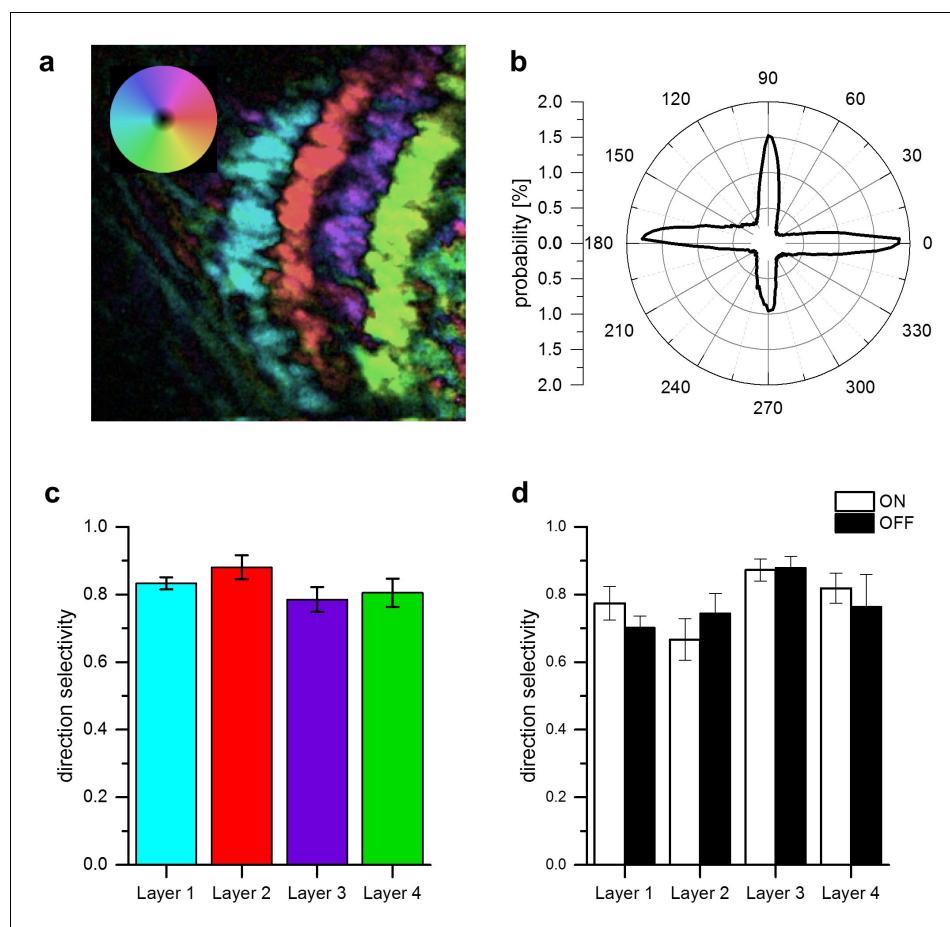


Figure 4. Directional tuning and selectivity of T4 and T5 cells. (a) Example of directional tuning to grating motion as determined by the vector sum of responses to grating motion along four cardinal directions. All neurons within each layer have almost identical preferred directions. (b) Histogram of preferred directions within all four layers. Clear peaks appear at the four cardinal directions. Data were obtained from 5 different flies. (c) Direction selectivity within each layer, as defined by the difference between the preferred and null direction responses, divided by the sum. Data represent the mean \pm SEM obtained from 5 flies (same data set as in b). (d) Same as c, but flies were stimulated by ON and OFF edges, respectively. Data represent the mean \pm SEM obtained from 3 flies.

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study relates to the dependence of the directional motion signal on the spot size in the apparent motion paradigm (**Figure 2e,f**). Given a half-width of the photoreceptor acceptance angle of approximately 5 degree in *Drosophila* (Götz, 1965), any stimulus is spatially low-pass filtered by a Gaussian with 5 degree full width at half maximum. Accordingly, enlarging the spot size will have two different effects: first, it will lead to an increasing peak intensity at the column where the stimulus spot is centered on, and second, it will lead to an increasing activation of neurons in neighboring columns. Which of these two effects is responsible for the higher threshold of T5 cells compared to T4 cells, and whether the sensitivity difference is in the input neurons or in T4/T5 cells themselves, cannot be decided by the present study.

In any case, our finding readily explains the high degree of direction selectivity found already at the processing stage where direction-selective signals first arise: neither a signal enhancement for preferred direction sequences nor a signal suppression for null direction sequences by itself would lead to such a strong direction selectivity as observed experimentally with large signals for preferred direction motion and zero responses for null direction motion (Maisak et al., 2013; Fisher et al., 2015). In analogy to the results presented for layer 3 T4 cells (Figure 5 in Haag et al., 2016), the responses of all T4 and T5 cells can be captured in algorithmic terms by a common mechanism, using a delayed, low-pass filtered input on the preferred side enhancing a fast, central input, with the result being suppressed by again a low-pass filtered input on the null side.

At the next processing stage, that is at the level of lobula plate tangential cells, the signals of oppositely tuned T4 and T5 cells become subtracted via inhibitory lobula plate interneurons (Mauss et al., 2015). This process, in a way, replicates the action of null direction suppression implemented on the dendrites of T4 and T5 cells. Since both mechanisms, that is the combination of preferred direction enhancement and null direction suppression on the dendrites of T4 and T5 cells as well as the subtraction of oppositely tuned T4 and T5 cells on the dendrites of tangential cells lead to high degree of direction selectivity at the output of the system, one might ask about the functional advantage of such a dual strategy. This question can be answered by either blocking null direction suppression on the T4/T5 cells dendrite or blocking the inhibitory lobula plate interneurons. The latter experiment has indeed been done, and the results revealed a loss of flow-field specificity of the tangential cells, due to the lack of inhibition caused by the non-matching part of the optic flow field (Mauss et al., 2015). For the converse situation, no experimental data exist so far and one has to rely on computer simulations (Haag et al., 2016, Figure 5). They suggest that a high direction selectivity is retained in tangential cells. This high degree of direction selectivity, however, rests on the relatively small differences between large, but poorly tuned signals and, thus, would be highly prone to noise. Improving the direction-tuning already at the level of T4 and T5 cells by the additional null-direction suppression should, therefore, increase the system's robustness to noise.

Our results open the door to the next level question about its neural implementation. Here, a recent connectomic study identified the major interneurons providing synaptic input to T4 cells as well as their placement on the dendrite (Takemura et al., 2017). Takemura and colleagues describe 4 columnar cell types, that is Mi1, Mi4, Mi9 and Tm3 as the major input elements to T4 cells, in addition to columnar cell types C3, TmY15, a wide-field neuron CT1 and other T4 cells with identical preferred direction. Columnar input neurons contact the T4 cell dendrite in a way that depends on the direction tuning subtype: while Mi9 synapses are clustered on the preferred side of the dendrite, Mi1 and Tm3 synapse on the central part and Mi4 are found predominantly on the null side. Most interestingly, the dynamic response properties of these different types of T4 input neurons match their position on the dendrite to suggest a specific function in the detector model discussed above: Mi9 and Mi4 indeed exhibit the temporal low-pass properties postulated for the inputs on the preferred and the null side, while Mi1 and Tm3 display fast band-pass properties needed for the central input (Arenz et al., 2017). This proposed correspondence needs to be tested by blocking individual input cell types and measuring the resulting effect on direction selectivity in T4 cells. Specifically, one would expect to abolish preferred direction enhancement when blocking Mi9, while blocking Mi4 should lead to a loss of null direction suppression. It is, however, important to stress that the effect of such blocking experiments is expected to be quite specific and directly observable only in directional responses of T4 cells: due to further network processing involving a subtraction of T4 cell signals with opposite directional tuning at the level of tangential cells (see previous paragraph, and Mauss et al., 2015), the effect might be far more subtle when downstream cells or behavior are used as a read-out (Strother et al., 2017). Nevertheless, blocking the central inputs Mi1 and Tm3

while recording from tangential cells revealed that Mi1 cells are absolutely essential for proper functioning of the ON pathway under all stimulus conditions tested, while blocking Tm3 only led to a loss of sensitivity for high edge velocities (Ammer et al., 2015). With respect to the polarity of the synapses of the various T4 input neurons, the correspondence outlined above predicts that Mi1 and Tm3 are excitatory while Mi4 should be inhibitory. In line with this, recent studies suggest a cholinergic phenotype in Mi1 and Tm3 (Pankova and Borst, 2017; Takemura et al., 2017) and a GABAergic one in Mi4 (Takemura et al., 2017). Seemingly in contrast to an enhancing action of Mi9 postulated above, this cell was found to be OFF sensitive (Arenz et al., 2017). However, Mi9 turned out to be immune-positive for the vesicular Glutamate reporter VGluT (Takemura et al., 2017). Together with the inhibitory action of Glutamate via the GluCl channel, well documented for other neurons of the Drosophila CNS (Liu and Wilson, 2013; Mauss et al., 2014; Mauss et al., 2015), this raises the possibility that Mi9 enhances the input from Mi1 and Tm3 onto T4 by a release from inhibition.

As for T4 cells, the major input neurons to T5 cells were identified by an EM study (Shinomiya et al., 2014). There, trans-medulla neurons Tm1, Tm2, Tm4, and Tm9 were found to make up for about 80% of all input synapses to T5 cell dendrites. However, the exact placement of the different inputs on the dendrite and, hence, the relative position of their receptive fields could not be determined by this report. As for their dynamic properties, only one of the cell types, Tm9, reveals low-pass characteristics, while all others (Tm1, Tm2 and Tm4) can be described as band-pass filters with different time-constants (Meier et al., 2014; Serbe et al., 2016; Arenz et al., 2017). In analogy of the arrangement of input neurons of T4 cells, Arenz et al., 2017 found that placing the two slowest cells (Tm1 and Tm9) on the outer arms and the fast Tm2 cell on the central arm of the three-input detector gives rise to a motion detector that fits real T5 cells both with respect to their high degree of direction selectivity and their temporal tuning optimum. Since, in contrast to T4 cells, the position of these interneurons on the dendrite of T5 cells is less well known so far, no prediction can be made whether blocking of Tm1 should lead to a loss of preferred direction enhancement and blocking of Tm9 to a loss of null direction suppression, or the other way round. Therefore, as is the case with T4 cells, further experiments are needed to determine which cell is playing which role in the functional context of preferred direction enhancement and null direction inhibition determined by the present study.

In summary, thus, we have found a common, uniform mechanism of direction selectivity for T4 and T5 cells that consists of combination of preferred direction enhancement and null direction suppression in different location of their receptive field, precisely related to their directional tuning. Mapping the different input neurons to T4 and T5 cells to their specific function in this context represents the next step of the analysis. The major challenge for future experiments will then consist in understanding the biophysical mechanisms underlying enhancement and suppression. Here, different ideas have been discussed in the past (Torre and Poggio, 1978; Koch and Poggio, 1992; Gabiani et al., 2002), and the different thresholds for preferred direction enhancement and null direction suppression described above might be an important result to decide between the various possibilities. These can now be tested at the molecular level using genome editing techniques available in *Drosophila* (Venken et al., 2011; Zhang et al., 2014; Fisher et al., 2017; Pankova and Borst, 2017).

Materials and methods

Flies

(*Drosophila melanogaster*) were raised at 25°C and 60% humidity on a 12 hr light/12 hr dark cycle on standard cornmeal agar medium. For calcium imaging of T5 cells, flies were used expressing the genetically-encoded calcium indicator GCaMP6m (Chen et al., 2013) in T4/T5 neurons with axon terminals predominantly in layer 3 of the lobula plate (w^- ; Sp/cyo; VT50384-lexA, lexAop-GCaMP6m/TM6b). For the imaging experiments of T4 and T5 cells in the four layers of the lobula plate we used flies expressing the calcium indicator GCaMP5 in both T4 and T5 cells in all layers of the lobula plate (w^- ; $+/+$; UAS-GCaMP5, R42F06-GAL4/UAS-GCaMP5, R42F06-GAL4).

Calcium imaging

Fly surgery was performed, and the neuronal activity was measured from the left optical lobe on a custom-built 2-photon microscope (*Denk et al., 1990*) as previously described (*Haag et al., 2016*). Images were acquired at a resolution of 64×64 pixels and at a frame rate of 15 Hz with the ScanImage software (*Pologruto et al., 2003*) in Matlab.

Optical stimulation

Stimulation with a telescopic stimulus device was similar to that used in our previous study (*Haag et al., 2016*). For the experiments shown in *Figure 4*, a regular stimulus display was used as described in (*Arenz et al., 2017*). The gratings had a spatial wavelength of 30 deg, a contrast of 100%, a mean luminance of 34 cd/m^2 and was moving along one of the four cardinal directions at 30 deg/sec.

Experimental protocol

In order to discriminate between T4 and T5 cells we stimulated single optical columns with bright pulses on a dark background. The cells were selected based on their response to light-on stimuli. While T4 cells respond to the onset of a light pulse, the T5 cells respond to the light-off. For the experiments the stimuli consisted either of dark pulses on a bright background (T5 cells) or bright pulses on a dark background (T4 cells). The pulses had a duration of 472 ms. At each position, three stimulus presentations were delivered. The resulting responses were averaged and the peak of the averaged response was taken. Apparent motion stimuli consisted of consecutive light stimuli to two neighboring cartridges. The second stimulus was presented right after the first turned off, resulting in a delay from onset to onset of 472 ms.

Data analysis

was performed offline using custom-written routines in Matlab. Regions of interests (ROIs) were selected by hand of the lobula plate. Time courses of relative fluorescence changes ($\Delta F/F$) were calculated from the raw imaging sequence. Responses to the stimulus were baseline-subtracted, averaged across repetitions, and quantified as the peak responses over the stimulus epochs. Those responses were averaged across experiments. Where indicated, responses were normalized to the maximum average response before averaging. For the apparent motion experiments, non-linear response components were calculated as the differences of the time-courses of the responses to the apparent motion stimuli and the sum of the appropriately time-shifted responses to flicker stimuli at the corresponding positions.

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Alexander Borst: Reviewing editor, eLife. The other authors declare that no competing interests exist.

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Author contributions

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Author ORCIDs

Juergen Haag  <https://orcid.org/0000-0002-6535-0103>
Abhishek Mishra  <http://orcid.org/0000-0002-1933-1251>
Alexander Borst  <http://orcid.org/0000-0001-5537-8973>

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Additional files

Supplementary files

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2.2 VOLTAGE TO CALCIUM TRANSFORMATION ENHANCES DIRECTION SELECTIVITY IN *Drosophila* T4 NEURONS

ABSTRACT A critical step in neural information processing is the transformation of membrane voltage into calcium signals leading to transmitter release. However, the effect of voltage to calcium transformation on neural responses to different sensory stimuli is not well understood. Here, we use *in vivo* two-photon imaging of genetically encoded voltage and calcium indicators, Arclight and GCaMP6f respectively, to measure responses in *Drosophila* direction-selective T4 neurons. Comparison between Arclight and GCaMP6f signals revealed calcium signals to have a significantly higher direction selectivity compared to voltage signals. Using these recordings we build a model which transforms T4 voltage responses to calcium responses. The model reproduces experimentally measured calcium responses across different visual stimuli using different temporal filtering steps and a stationary non-linearity. These findings provide a mechanistic underpinning of the voltage-to-calcium transformation and show how this processing step, in addition to synaptic mechanisms on the dendrites of T4 cells, enhances direction selectivity in the output signal of T4 neurons.

AUTHORS Abhishek Mishra, Alexander Borst, and Juergen Haag

CONTRIBUTIONS Abhishek Mishra, Conceptualization, Investigation, Data curation, Software, Visualization, Writing original draft, review and editing; Juergen Haag, Conceptualization, Data curation, Software, Investigation, review and editing; Alexander Borst, Conceptualization, Funding acquisition, Project administration, Review and editing

Voltage to Calcium Transformation Enhances Direction Selectivity in *Drosophila* T4 neurons

Abhishek Mishra^{1,2}, Alexander Borst^{1,2}, Juergen Haag¹

⁵ Max Planck Institute for Biological Intelligence (in foundation), Martinsried, Germany;

⁶ Graduate School of Systemic Neurosciences, LMU Munich, Martinsried, Germany

Abstract A critical step in neural information processing is the transformation of membrane voltage into calcium signals leading to transmitter release. However, the effect of voltage to calcium transformation on neural responses to different sensory stimuli is not well understood. Here, we use *in vivo* two-photon imaging of genetically encoded voltage and calcium indicators, Arclight and GCaMP6f respectively, to measure responses in *Drosophila* direction-selective T4 neurons. Comparison between Arclight and GCaMP6f signals revealed calcium signals to have a significantly higher direction selectivity compared to voltage signals. Using these recordings we build a model which transforms T4 voltage responses to calcium responses. The model reproduces experimentally measured calcium responses across different visual stimuli using different temporal filtering steps and a stationary non-linearity. These findings provide a mechanistic underpinning of the voltage-to-calcium transformation and show how this processing step, in addition to synaptic mechanisms on the dendrites of T4 cells, enhances direction selectivity in the output signal of T4 neurons.

Introduction

In order to guide animal behavior, neurons perform a wide range of computations. Neurons encode information via graded changes in membrane potential or action potential frequency. Mostly they communicate via chemical synapses which requires the release of neurotransmitters. When the presynaptic membrane is sufficiently depolarized, voltage-gated calcium channels open and allow Ca^{2+} to enter the cell (Luo 2020). Calcium entry leads to the fusion of synaptic vesicles with the membrane and release of neurotransmitter molecules into the synaptic cleft (Chapman 2002). As neurotransmitters diffuse across the synaptic cleft, they bind to receptors in the postsynaptic membrane, causing postsynaptic neuron to depolarize or hyperpolarize, passing the information from pre to postsynaptic neurons (Di Maio 2008). Voltage to calcium transformation therefore represents one crucial step in neural information processing and neural computation.

A classic example of neural computation is how *Drosophila* neurons compute the direction of visual motion (Borst *et al.* 2020). In *Drosophila*, visual information is processed in parallel ON (contrast increments) and OFF (contrast decrements) pathways (Joesch *et al.* 2010; Eichner *et al.* 2011). Direction selectivity emerges three synapses downstream of photoreceptors, in T4 and T5 for ON and OFF pathways respectively. Four subtypes of T4 and T5 cells exist, each responding selectively to one of the four cardinal directions (Maisak *et al.* 2013). Amazingly, right at the first stage where direction selectivity emerges, T4 and T5 cells exhibit a high degree of direction selectivity, with no responses to null direction stimuli. This statement is, however, based on calcium

41 recordings. Whole-cell patch clamp recordings show a somewhat different picture: While preferred
42 direction stimuli also lead to large membrane depolarizations, edges or gratings moving along
43 the null directions elicit smaller but significant responses as well (Wienecke *et al.* 2018; Groschner
44 *et al.* 2022). This hints to an additional processing step where voltage signals are transformed
45 into calcium signals that increases direction selectivity of the cells. In order to study this step
46 systematically, we recorded both voltage and calcium signals in response to a large stimulus set
47 that includes gratings and edges moving along various directions at different speeds and contrasts.
48 Using these data, we build a model that captures the transformation from voltage to calcium by a
49 few linear and non-linear processing steps.

50 Results

51 We first expressed the genetically encoded calcium indicator GCaMP6f (Chen *et al.* 2013) in T4 cells
52 projecting to layer 3 of the lobula plate. These cells have upward motion as their preferred direction
53 (PD) and downward motion as their null direction (ND). We also expressed the genetically encoded
54 voltage indicator Arclight (Jin *et al.* 2012) using the same driver line. Arclight's fluorescence decreases
55 with membrane depolarization and increases with membrane hyperpolarization. To compare the
56 voltage and calcium signals, we recorded the neural activity in T4c cells dendrites in medulla layer 10
57 in response to the same set of stimuli using 2-photon microscopy (Denk *et al.* 1990). The complete
58 stimuli set included square-wave gratings of 30° spatial wavelength moving in 12 different directions,
59 and ON edges moving in PD and ND, at four different speeds ($15^{\circ}s^{-1}$, $30^{\circ}s^{-1}$, $60^{\circ}s^{-1}$, $120^{\circ}s^{-1}$) and four
60 different contrasts (10%, 20%, 50%, 100%).

61 In a first set of experiments, we measured the voltage and calcium signals in response to
62 gratings moving in PD and ND at four different speeds (figure 1A). As the grating stimuli consists
63 of alternate bright and dark bars moving in a certain direction, there was a modulation in the
64 Arclight (black traces) and GCaMP6f (red traces) responses to it. The GCaMP6f responses showed
65 modulations only for slower speeds, while Arclight responses revealed modulations also for faster
66 speeds. The response amplitudes were much higher for GCaMP6f ($\approx 2.0\Delta F/F$) compared to Arclight
67 ($\approx -0.06\Delta F/F$). The peak responses (maximum $\Delta F/F$) decreased with increasing stimulus speed
68 both for GCaMP6f and Arclight (figure 1B). To understand if voltage to calcium transformation
69 affects direction selectivity in T4 cells, we compared the responses to gratings moving in PD and
70 ND. GCaMP6f responses in ND were negligible compared to its responses in PD, while for Arclight
71 responses in ND were quite visible. We quantified the direction selectivity using a direction selectivity
72 index (DSI) calculated as the difference of the peak responses to preferred and null direction, divided
73 by the sum of the peak responses (Materials and Methods equation (1)). The results revealed a high
74 degree of direction selectivity of ≈ 0.8 for GCaMP6f at slower velocities, compared to a direction
75 selectivity of ≈ 0.4 for Arclight (figure 1E). For both GCaMP6f and Arclight signals, direction selectivity
76 decreased with increasing velocity.

77 Next, instead of gratings, we used moving bright edges with all other stimulus parameters
78 remaining the same (figure 1C). As the edge moves upward on the screen, it crosses the receptive
79 field of T4c neurons ($\approx 15^{\circ}$) only once. Hence, there was only a single peak in the response. The
80 peak response decreased with increasing stimulus speed for GCaMP6f, while the peak response
81 remained almost constant for Arclight throughout all speeds (figure 1D). When comparing edge
82 responses moving along preferred and null directions, GCaMP6f showed negligible responses in
83 null direction while Arclight revealed considerable responses to null direction stimuli. The direction
84 selectivity index was again much higher for GCaMP6f compared to Arclight (figure 1F). Together
85 these results show that GCaMP6f signals have a high level of direction selectivity compared to
86 Arclight signals, both for grating and edge stimuli.

87 The stimulus strength was further varied by changing the contrast between bright and dark
88 bars for gratings and between moving edge and background for edge stimuli. We measured
89 Arclight and GCaMP6f responses to gratings moving at $30^{\circ}s^{-1}$ at four different contrasts (figure 2A).
90 Increasing contrast resulted in an increase in response for both Arclight and GCaMP6f. GCaMP6f

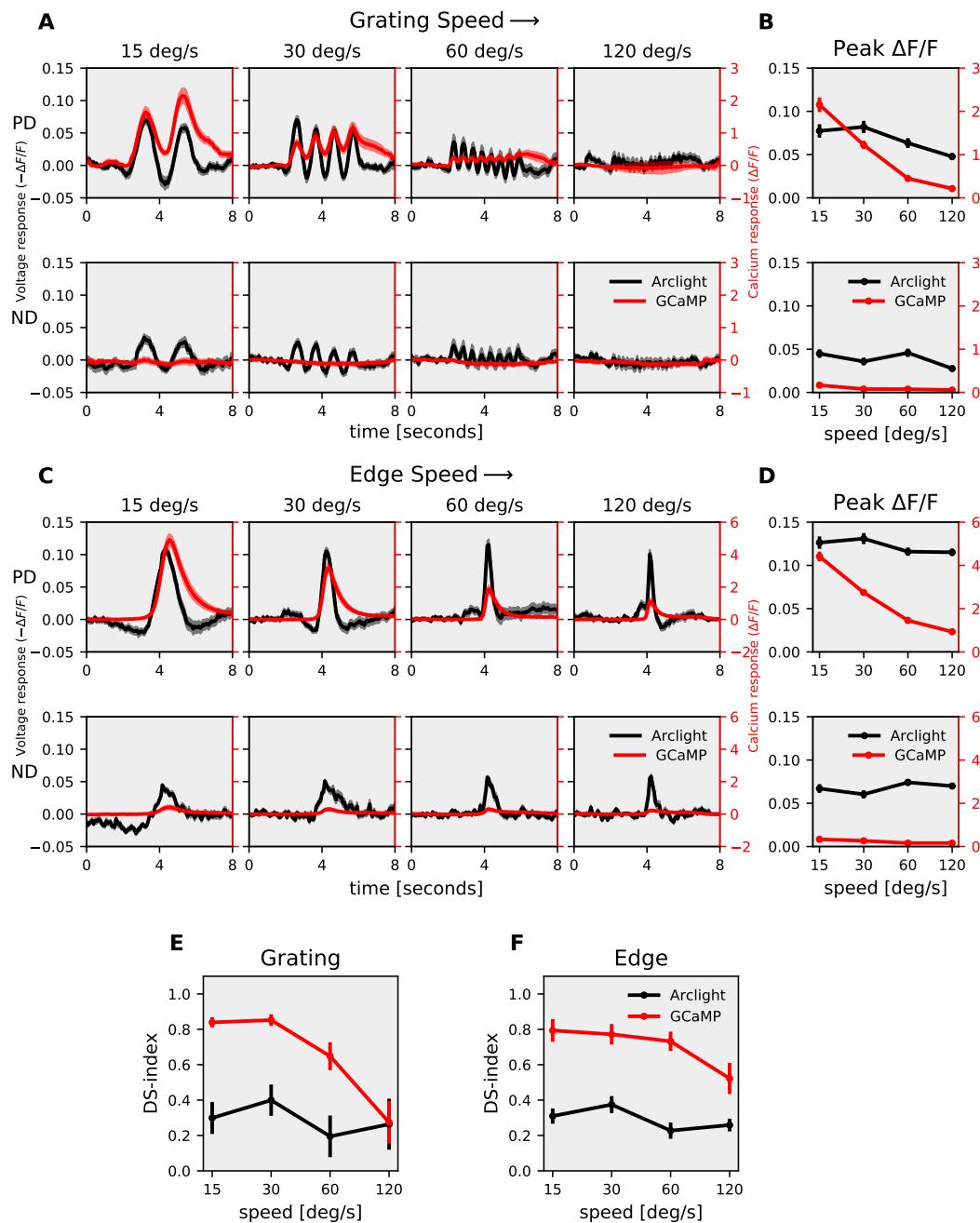


Figure 1. T4c speed dependence : (A) T4c Arclight (black) and GCaMP6f (red) responses to grating moving in PD (top row) and ND (bottom row) at 4 different speeds. The plots have twin y-axis. The left y-axis of the plot represents voltage responses i.e. changes in Arclight fluorescence ($-\Delta F/F$) and the right y-axis of the plot represents calcium responses i.e. changes in GCaMP6f fluorescence ($\Delta F/F$) (B) T4c peak responses to grating moving in PD (top) and ND (bottom) at 4 different speeds. ($n = 20$ ROIs from $N = 10$ flies for Arclight, $n = 18$, $N = 9$ for GCaMP6f) (C) T4c Arclight (black) and GCaMP6f (red) responses to ON-edge moving in PD (top row) and ND (bottom row) at 4 different speeds. ($n = 29$, $N = 10$ for Arclight, $n = 17$, $N = 4$ for GCaMP6f) (E) Direction selectivity index (DSI) calculated as difference of peak responses in PD and ND divided by the sum of peak responses for grating. (F) Direction selectivity index (DSI) for ON-edge. All data shows the mean \pm SEM. PD: preferred direction, ND: null direction.

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signals were modulated at the temporal frequency of the grating but showed an additional rise over time. This slow increase was not observed in Arclight signals. We also measured Arclight and GCaMP6f responses to ON edges moving at the same speed of 30°s^{-1} but having different contrasts (figure 2C). The peak response (maximum $\Delta F/F$) increased with increasing contrast (figure 2D). Similar to previous experiments, the direction selectivity index was much higher for GCaMP6f (≈ 0.9) compared to that for Arclight (≈ 0.4) (figure 2E,F).

In the results presented so far we compared responses for two directions only, i.e. along the preferred (upward) and along the null direction (downward). We next extended the comparison to motion along 12 directions, from 0° to 360° in steps of 30° . For this comparison, we determined the normalized peak responses of Arclight and GCaMP6f signals to gratings moving in 12 directions at 4 different speeds and 4 different contrasts, respectively (figure 3A, B). The directional tuning was much sharper for GCaMP6f compared to Arclight. To quantify this we calculated the directional tuning index L_{dir} (Mazurek *et al.* 2014) for each speed and each contrast as the vector sum of the peak responses divided by the sum of all individual vector magnitudes (Materials and Methods equation (2)). In general, the directional tuning indices again were much higher for GCaMP6f (≈ 0.6) compared to that of Arclight (≈ 0.2) (figure 3C, D). Together these results show that GCaMP6f signals have a higher degree of directional tuning across different speeds and contrasts than Arclight.

How does the voltage to calcium transformation lead to calcium signals with significantly higher directional tuning compared to voltage signals? To address this question, we constructed an algorithmic model (figure 4) which takes Arclight signals as inputs and outputs GCaMP signal. In order to find the optimal parameter values, we first defined an error function. For each stimulus condition, the error was calculated as the sum of the squared difference between the model and experimental data at each time-point (Materials and Methods equation (3)). There were a total of 112 stimulus conditions: gratings speed (48), gratings contrast (48), edge speed (8) and edge contrast (8). The total error amounted to the sum of errors across all stimulus conditions (Materials and Methods equation (4)). We defined the model error as the total error divided by the power of the data (Materials and Methods equation (5)). We then found the optimal parameters values of the model that correspond to the minimum total error using Python SciPy optimize minimize function (Virtanen *et al.* 2020).

We started with a simple model (figure 4A). The model first passes the Arclight signal through a high-pass filter. The high-pass filter brings the input Arclight signal closer to the actual voltage signal by removing the slowly fluctuating Arclight indicator dynamics. This is followed by a threshold, assuming that the voltage changes below a certain threshold does not affect the calcium level in the cell. Now, few experimental observations which we took into consideration for building up the model further were as follows : First, the GCaMP6f response to gratings showed modulations only for slower speeds, whereas Arclight response had modulations even at faster speeds (figure 1A). This suggests that the GCaMP6f signal is a low-pass filtered version of the Arclight signal. In the simple model, we used a single low-pass filter followed by a gain and time-shift. Multiplication with a gain factor was required since GCaMP6f signals have a much higher magnitude compared to Arclight. Arclight and GCaMP6f responses were recorded from cells in different flies with different receptive fields, therefore the responses had different phases, and a time-shift was necessary to align the signals. However, the simple model with single low-pass filter could not reproduce responses across all stimuli. The model error for the complete dataset fit for the simple model was around 34%. Specifically, the simple model failed to suppress the ND-responses and to reproduce the edge responses. The directional tuning index L_{dir} was much smaller for the simple model compared to the experimental data (figure 5E,F). Second, the GCaMP6f responses in addition to modulation also had a steady rise over time whereas Arclight signal only had modulations (figure 1A, 2A). For reproducing the edge responses and modulation in grating responses, the model needed a low-pass filter with a small time constant. However to simulate the steady rise in the grating signal, a low-pass filter with a large time constant was necessary. Hence, we combined the output of two low-pass filters. Summing up the low-pass filter outputs did not lead to much improvement.

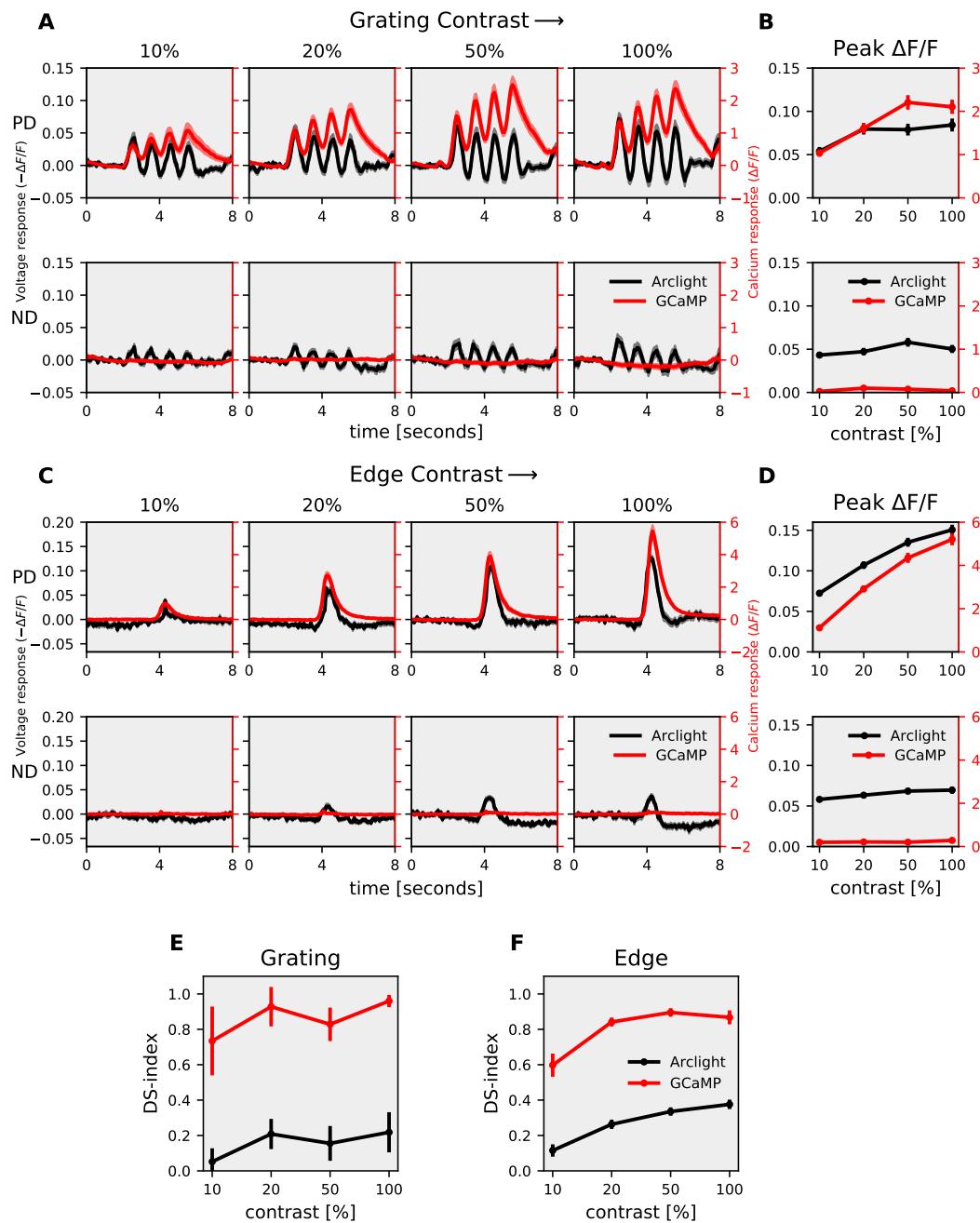


Figure 2. T4c contrast dependence : (A) T4c Arclight (black) and GCaMP6f (red) responses to grating moving in PD (top row) and ND (bottom row) at 4 different contrasts. The left y-axis of the plot represents voltage responses i.e. changes in Arclight fluorescence ($-\Delta F/F$) and the right y-axis of the plot represents calcium responses i.e. changes in GCaMP6f fluorescence ($\Delta F/F$) (B) T4c peak responses to grating moving in PD (top) and ND (bottom) at 4 different contrasts. (n = 23 ROIs from N = 11 flies for Arclight, n = 22, N = 9 for GCaMP6f) (C) T4c Arclight (black) and GCaMP6f (red) responses to ON-edge moving in PD (top row) and ND (bottom row) at 4 different contrasts. (D) T4c peak responses to ON-edge moving in PD and ND at 4 different contrasts. (n = 36, N = 5 for Arclight, n = 41, N = 7 for GCaMP6f) (E) Direction selectivity index (DSI) calculated as difference of peak responses in PD and ND divided by the sum of peak responses for grating. (F) Direction Selectivity Index (DSI) for ON-edge. All data shows the mean ± SEM.

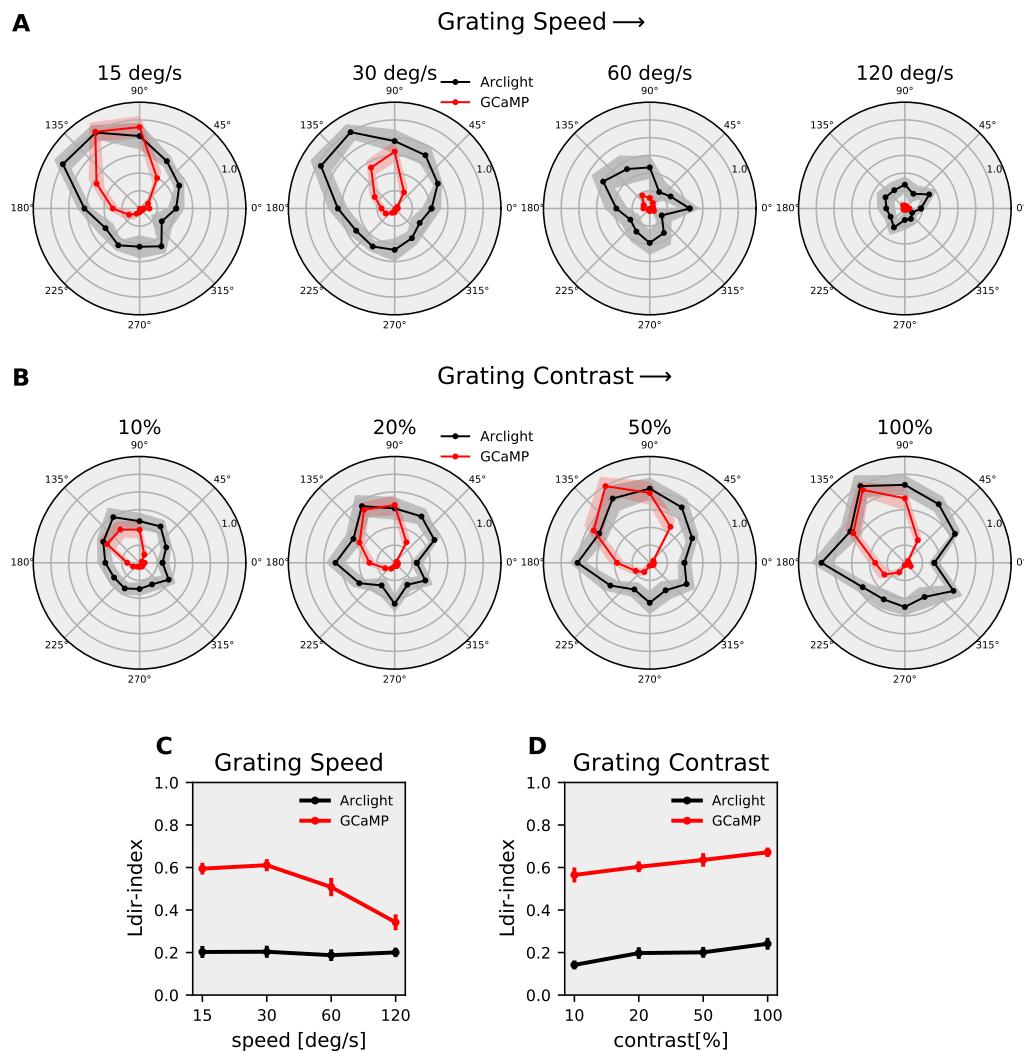


Figure 3. T4c direction tuning : (A) T4c Arclight (black) and GCaMP6f (red) normalized peak responses to grating moving in 12 directions at 4 different speeds. ($n = 20$ ROIs from $N = 10$ flies for Arclight, $n = 18$, $N = 9$ for GCaMP6f) (B) T4c Arclight (black) and GCaMP6f (red) normalized peak responses to grating moving in 12 directions at 4 different contrasts. ($n = 23$, $N = 11$ for Arclight, $n = 22$, $N = 9$ for GCaMP6f) (C) The directional tuning index L_{dir} for grating moving at 4 different speeds. The directional tuning index is calculated as the vector sum of the peak responses divided by the sum of all individual vector magnitudes. (D) The directional tuning index for grating at 4 different contrasts. All data shows the mean \pm SEM measured in 5 different flies.

A Simple Model



B Multiplicative Model

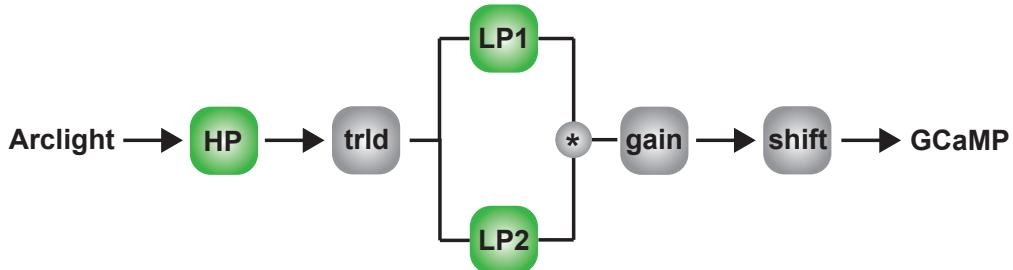


Figure 4. Models for voltage to calcium transformation : (A) Simple model consisting of High-Pass filter (HP), threshold (trld), Low-Pass filter (LP), gain and shift. (B) Multiplicative model combining output of two low-pass filters via multiplication.

142 However, combining both outputs from the low-pass filters with a multiplication led to significant
 143 decrease in the error. The model error for the multiplicative model (figure 4B) then was only at
 144 around 20%.

145 The multiplicative model thus has in total 6 parameters - high-pass filter time constant, threshold,
 146 low-pass filter 1 time constant, low-pass filter 2 time constant, gain and shift. The multiplicative
 147 model was able to reproduce calcium signals across different visual stimuli (figure 5). It could
 148 reproduce both the modulation as well as slow rise in the GCaMP6f signal in response to gratings
 149 (figure 5A). The multiplicative model could also reproduce the ON edge speed tuning responses
 150 across different speeds (figure 5C,D). The directional tuning index L_{dir} were similar for multiplicative
 151 model and experimental data across slower speeds and all contrasts (figure 5E,F).

152 Is the slow rise in GCaMP6f signals over time due to the properties of T4 cells or due to the
 153 properties of GCaMP6f? To answer this question we used a faster version of the calcium indicator
 154 GCaMP8f (Zhang et al. 2020). GCaMP8f was expressed in T4c cells using the same driver line. The
 155 experiments were repeated using grating stimuli in 12 directions at 4 speeds and ON edges moving
 156 in PD and ND. T4c cells GCaMP8f responses were similar to GCaMP6f responses but faster. As
 157 with GCaMP6f, GCaMP8f signals had modulation and slow rise over time. We further compared
 158 the model parameters values for GCaMP6f data fit and GCaMP8f data fit (figure 6). The model
 159 parameters were similar, but with time constants having smaller values for GCaMP8f as it is a faster
 160 indicator. Therefore, the slow rise in the calcium signal is not due to the properties of GCaMP6f
 161 indicator.

162 To reproduce the calcium responses for direction-selective T4c cells under all stimuli conditions,
 163 a multiplicative model was required. For non-direction-selective cells, what does the voltage to
 164 calcium transformation look like, and is the simple model able to replicate the calcium response
 165 for these cells? In order to answer this question, we expressed Arclight & GCaMP6f in medulla
 166 neurons Mi1 & Tm3 cells, which are both non-direction-selective. Mi1 and Tm3 are pre-synaptic to
 167 T4 cells and have ON-center receptive field (Behnia et al. 2014; Arenz et al. 2017). We measured
 168 Mi1, Tm3 Arclight (black) and GCaMP6f (red) responses to gratings moving at 4 different speeds
 169 and to gratings moving at 4 different contrasts (figure 7). The gratings were moved in only one

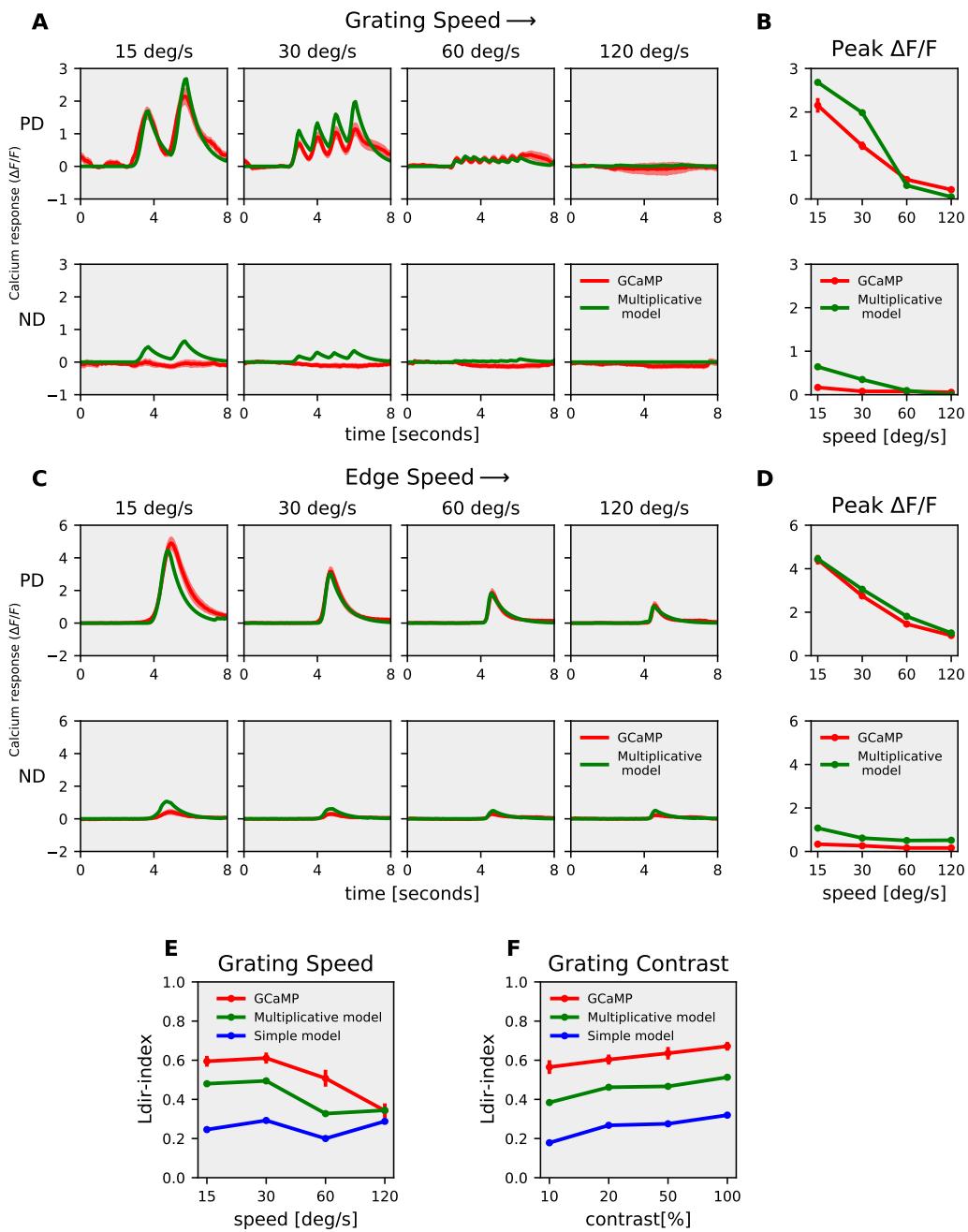


Figure 5. Model responses : (A) T4c GCaMP6f (red) and multiplicative model (green) responses to grating moving in PD (top row) and ND (bottom) at 4 different speeds. (B) T4c GCaMP6f and model peak responses to grating moving in PD (top) and ND(bottom) at 4 different speeds. (C) T4c GCaMP6f (red) and multiplicative model (green) responses to ON-edge moving in PD (top row) and ND (bottom row) at 4 different speeds. (D) T4c GCaMP6f and model peak responses to ON-edge moving in PD (top) and ND (bottom) at 4 different speeds. (E, F) The directional tuning index L_{dir} for GCaMP6f (red), multiplicative (green) and simple (blue) model for grating moving in 12 directions at 4 different speeds and at 4 different contrasts respectively.

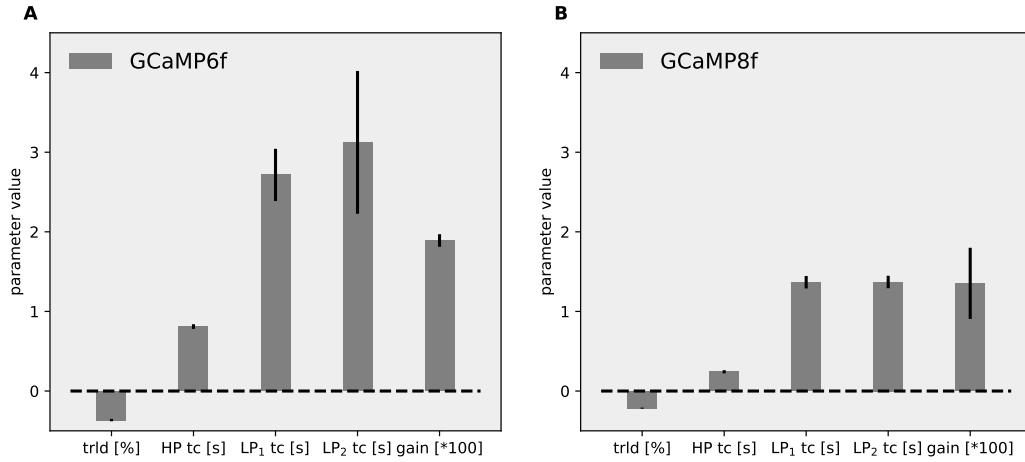


Figure 6. Model parameters for GCaMP6f (A) and GCaMP8f (B): Data shows mean \pm SD for optimal parameters for the multiplicative model. The data were fit for grating moving in 12 directions and 4 speeds, and for ON-edge moving in PD and ND at 4 speeds. trld : threshold, HP : High Pass, LP : Low Pass, tc : Time constant

170 direction, since the direction does not affect non-direction-selective cells' responses. Contrary to T4,
 171 Mi1 GCaMP6f responses had only modulation without a slow increase over time (figure 7A). Tm3
 172 GCaMP responses did not increase over time, and showed only modulation for gratings moving at
 173 $15^{\circ}s^{-1}$. For gratings moving at $30^{\circ}s^{-1}$ and $60^{\circ}s^{-1}$, there was an increase in Tm3 GCaMP6f response
 174 over time, but the Arclight response also already had a slow increment over time (figure 7A). Similar
 175 to T4, the peak response for Mi1 and Tm3 decreased with an increase in speed and increased
 176 with an increase in contrast (figure 7B, D). Together, these results show that voltage to calcium
 177 transformation causes GCaMP6f response increment over time only for direction-selective T4 cells
 178 and not for non-direction-selective Mi1 and Tm3 cells.

179 Next, we used the model described in figure 4 to reproduce Mi1 and Tm3 calcium responses
 180 using their Arclight responses. As discussed earlier, the simple model (figure 4A) with single
 181 low-pass filter was not able to reproduce T4 calcium responses across all stimuli. However, for
 182 Mi1 and Tm3, the simple model with a single low-pass filter was able to reproduce the calcium
 183 responses across all stimuli conditions (figure 8). The model also accurately replicated the speed
 184 and contrast tuning for Mi1 and Tm3 (figure 8B, D). We further compared the model error for
 185 simple and multiplicative model for Mi1, Tm3 and T4c data (figure 9). The model error for Mi1 and
 186 Tm3 for simple model was $\approx 6.5\%$ and $\approx 5.9\%$ respectively compared to $\approx 11.9\%$ and $\approx 7\%$ for the
 187 multiplicative model. Thus, the simple model already performed well for Mi1 and Tm3 dataset,
 188 and changing to multiplicative model did not improve the performance. For the T4c dataset the
 189 model error was $\approx 34\%$ and $\approx 21\%$ for the simple and multiplicative model respectively. Hence, the
 190 multiplicative model with two low-pass filters performed better for T4c dataset whereas for Mi1 and
 191 Tm3 the Simple model with single low-pass filter was sufficient to reproduce the calcium responses.
 192 This suggests that voltage-to-calcium transformation is more complex for direction-selective cell T4
 193 than for the non-direction-selective cells Mi1 and Tm3.

194 Discussion

195 Neuronal signaling and information processing involves the transformation of membrane voltage
 196 into calcium signals, which lead to transmitter release. Computations can occur at different stages in
 197 the signalling cascade: 1.) dendritic integration and processing of voltage signals. 2.) transformation
 198 from voltage to calcium and 3.) between calcium and neurotransmitter release. In this study, we

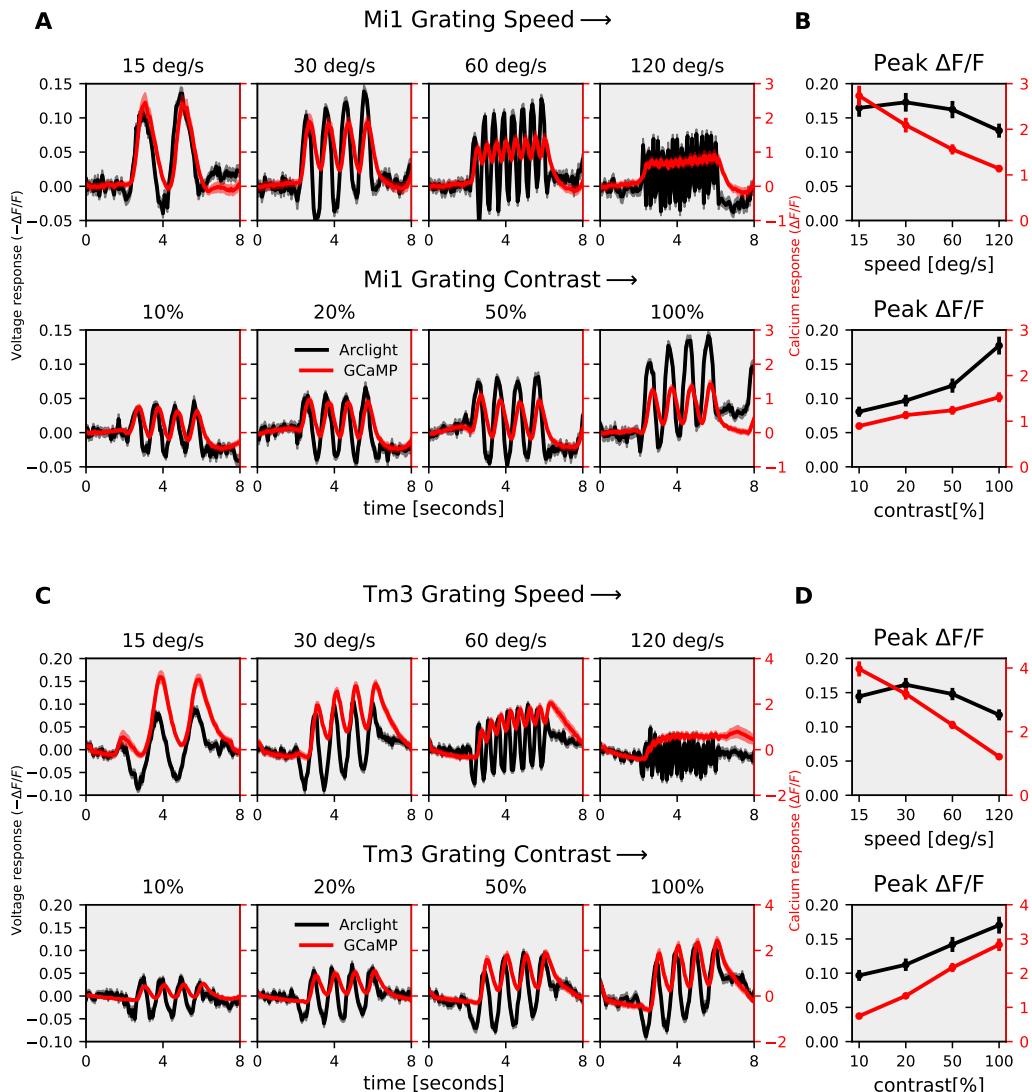


Figure 7. Mi1, Tm3 speed and contrast dependence : (A) Mi1 Arclight (black) and GCaMP6f (red) responses to grating moving at 4 different speeds (top row) and 4 different contrasts (bottom row). The left y-axis of the plot represents voltage responses i.e. changes in Arclight fluorescence ($-\Delta F/F$) and the right y-axis of the plot represents calcium responses i.e. changes in GCaMP6f fluorescence ($\Delta F/F$) (B) Mi1 peak responses to grating moving at 4 different speeds ($n = 24$ ROIs from $N = 5$ flies for Arclight, $n = 19$, $N = 8$ for GCaMP) and 4 different contrasts ($n = 24$, $N = 5$ for Arclight, $n = 22$, $N = 8$ for GCaMP). (C) Tm3 Arclight (black) and GCaMP6f (red) responses to grating moving at 4 different speeds (top row) and 4 different contrasts (bottom row). (D) Tm3 peak responses to grating moving at 4 different speeds ($n = 52$, $N = 5$ for Arclight, $n = 37$, $N = 4$ for GCaMP) and 4 different contrasts ($n = 35$, $N = 5$ for Arclight, $n = 36$, $N = 4$ for GCaMP). All data shows the mean \pm SEM.

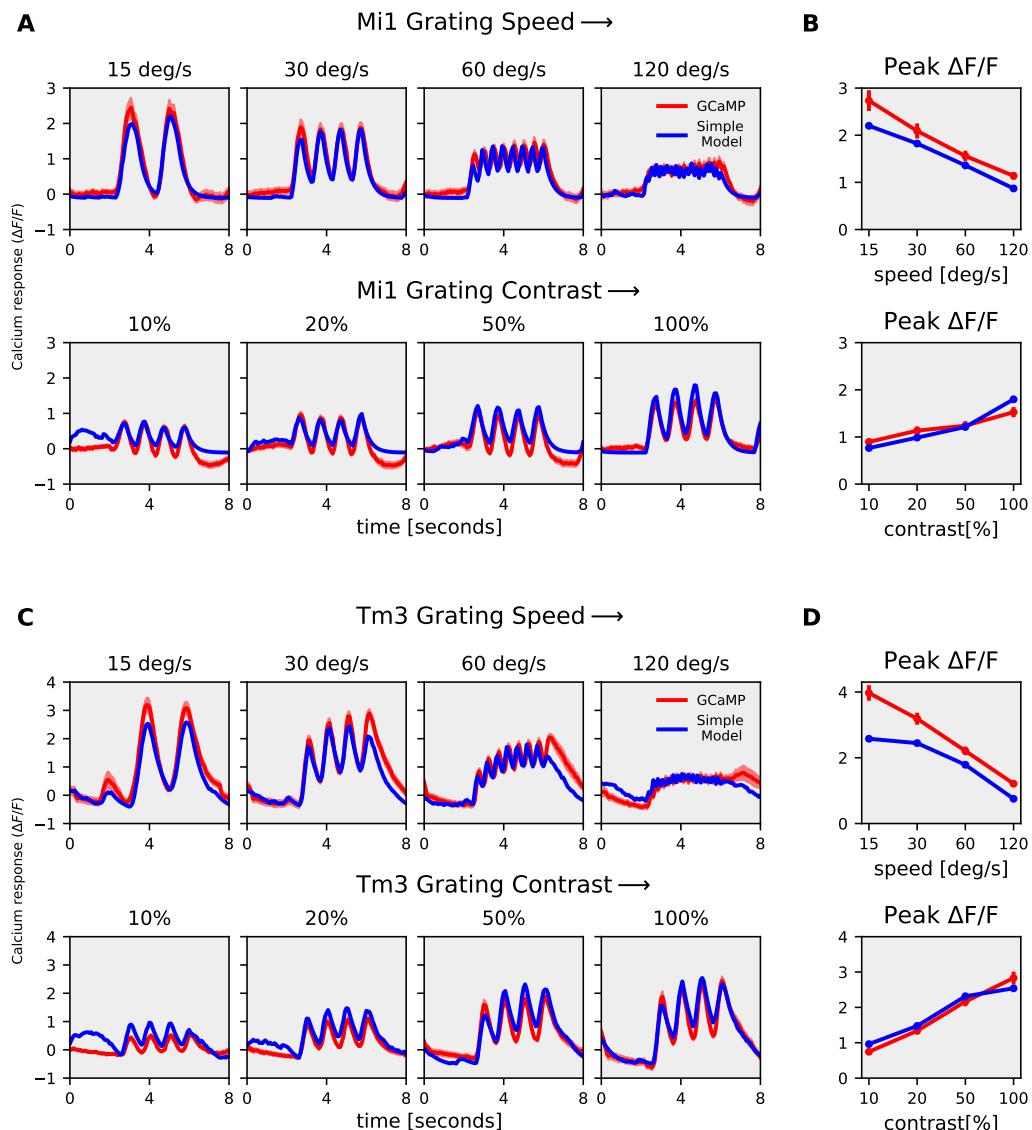


Figure 8. Mi1, Tm3 Simple model responses : (A) Mi1 GCaMP6f (red) and simple model (blue) responses to gratings moving at 4 different speeds (top row) and to gratings moving at 4 different contrasts (bottom row). (B) Mi1 GCaMP6f and model peak responses to gratings moving at 4 different speeds (top) and 4 different contrasts (bottom). (C) Tm3 GCaMP6f (red) and simple model (blue) responses to gratings moving at 4 different speeds (top row) and to gratings moving at 4 different contrasts (bottom row). (D) Tm3 GCaMP6f and model peak responses to gratings moving at 4 different speeds (top) and 4 different contrasts (bottom).

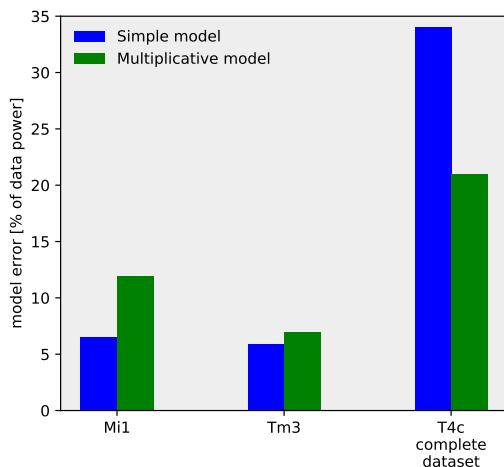


Figure 9. Model error for the simple and multiplicative model : The model error for the simple model (blue) and multiplicative model (green). Mi1 and Tm3 dataset consists of gratings at 4 different speeds and contrast moving in a single direction. T4c complete dataset consists of gratings moving in 12 different directions, and ON edge moving in PD, ND at 4 different speeds and contrasts i.e. a total of 112 stimuli conditions.

199 explored the transformation of voltage to calcium in T4-cells, the first direction-selective neurons in
200 the *Drosophila* ON motion vision pathway.

201 We found that the voltage to calcium transformation in T4c neurons enhances their direction
202 selectivity: calcium signals in T4c cells have a significantly higher direction selectivity and tuning
203 compared to membrane voltage across different stimuli conditions (figure 1-3). The direction
204 selectivity index for calcium signals compared with voltage signals for a few stimuli conditions was
205 previously found to be higher in a study in T5 cells using ASAP2f as an optical voltage indicator
206 (Wienecke *et al.* 2018). As calcium is required for neurotransmitter release, this is expected to
207 increase the direction selectivity of T4/T5 cells' output signals. In the lobula plate, T4/T5 cells
208 provide inputs onto large lobula plate tangential cells that are depolarized during preferred and
209 hyperpolarized during null direction motion (Mauss *et al.* 2014). For example, vertical system
210 (VS) cells with dendrites in layer 4 receive direct excitatory inputs from downward tuned T4d/T5d
211 neurons causing depolarization during motion in the downward preferred direction. These VS cells
212 also receive indirect inhibitory inputs from upward tuned T4c/T5c neurons via glutamatergic LPi3-4
213 neurons projecting from layer 3 to layer 4 causing hyperpolarization in VS cells during motion in the
214 upward null direction. Upon silencing LPi3-4 neurons' synaptic output via tetanus toxin, VS neurons
215 depolarization response in the preferred direction did not change, but the null direction response
216 was absent (Mauss *et al.* 2015). This suggests T4/T5 do not release any transmitter in response
217 to null direction motion, which matches our findings for the calcium responses. Thus, voltage to
218 calcium transformation increases direction selectivity in T4/T5 cells and this enhances direction
219 selectivity in downstream neurons.

220 Electrophysiology has been the most frequently used method to measure the membrane
221 potential changes in neurons. However, due to the small size of neurons in the optic lobe, single-cell
222 electrophysiological recordings of these neurons have been difficult. Genetically encoded voltage
223 indicators (GEVIs) have evolved as powerful tools for recording changes in neuronal membrane
224 potentials (Yang *et al.* 2016). Optical methods of monitoring brain activity are appealing because
225 they allow simultaneous, noninvasive monitoring of activity in many individual neurons. We used
226 a fluorescence protein (FP) voltage sensor called Arclight (Jin *et al.* 2012). Arclight is based on the
227 fusion of the voltage-sensing domain of *Ciona intestinalis* voltage-sensitive phosphatase (Murata
228 *et al.* 2005) and the fluorescent protein super eYFP with an A227D mutation. Arclight

229 has been shown to robustly report both subthreshold events and action potentials in genetically
230 targeted neurons in the intact *Drosophila* brain (Cao *et al.* 2013).

231 We built a model to capture voltage to calcium transformation in T4c, Mi1, and Tm3 cells. A
232 simple model with a single low-pass filter was able to reproduce calcium responses in non-direction-
233 selective Mi1 and Tm3 cells (figure 8), whereas a more complex model combining the output of two
234 low-pass filters via a multiplication was required to reproduce T4c calcium responses (figure 5). The
235 direction selectivity for the simple model signals for T4c was lower compared to the multiplicative
236 model. This suggests that voltage-calcium transformation in Mi1 and Tm3 cells is different from
237 those in T4c cells.

238 The time constants for the two low-pass filters were identical for the multiplicative model for
239 T4c data fit. Thus, these two low-pass filters in the multiplicative model could also be replaced by a
240 single low-pass filter followed by a quadratic non-linearity. An exponent of close to 2 (exact value:
241 2.2) was found in the parameter search for a model with a single low-pass filter followed by an
242 exponential nonlinearity.

243 Differential expression of voltage-gated calcium channels in these cells could explain the dif-
244 ferent voltage to calcium transformation. Voltage-gated calcium channels mediate depolarization-
245 induced calcium influx that drives the release of neurotransmitters. The α 1-subunit of the voltage-
246 gated calcium channels forms the ion-conducting pore, which makes it distinct from other calcium
247 channels. Three families of genes encode α 1 subunits. *Drosophila* genome has one α 1 subunit
248 gene in each family: α 1D (Ca_v 1), cac (Ca_v 2), and α 1T (Ca_v 3) (Littleton & Ganetzky 2000; King 2007).
249 In *Drosophila* antennal lobe projection neurons, cac (Ca_v 2) type and α 1T (Ca_v 3) type voltage-gated
250 calcium channels are involved in sustained and transient calcium currents, respectively (Gu *et al.*
251 2009; Iniguez *et al.* 2013). According to a RNA-sequencing study (Davis *et al.* 2020), α 1T (Ca_v 3)
252 mRNA have higher expression in Mi1 (2050.16 Transcripts per Million (TPM)) compared to T4 (686.68
253 TPM) and Tm3 (336.45 TPM). While cac (Ca_v 2) mRNA have higher expression in T4 (1298.53 TPM)
254 compared to Mi1 (986.25 TPM) and Tm3 (817.61 TPM). Different expression of voltage-gated calcium
255 channels could cause different voltage to calcium transformations in non-direction selective and
256 direction-selective cells. In addition to dendritic integration of postsynaptic voltages, the specific
257 voltage-to-calcium transformation described in this study provides an important processing step
258 that enhances direction selectivity in the output signal of motion-sensing neurons of the fly.

259 Materials and Methods

260 Flies

261 Flies (*Drosophila melanogaster*) were raised at 25°C and 60% humidity on a 12 hour light/12 hour dark
262 cycle on standard cornmeal agar medium. For calcium imaging experiments, genetically-encoded
263 calcium indicator GCaMP6f (Chen *et al.* 2013) was expressed in T4 neurons with axon terminals
264 predominantly in layer 3 of the lobula plate. Similarly for voltage imaging experiments, genetically-
265 encoded voltage indicator Arclight (Jin *et al.* 2012) was expressed in T4 layer 3 neurons. The flies
266 genotype were as follows :

- 267 1. T4c>GCaMP6f : w+ ; VT15785-Gal4AD / UAS-GCaMP6f; VT50384-Gal4DBD / UAS-GCaMP6f
- 268 2. T4c>Arclight : w+ ; VT15785-Gal4AD / UAS-Arclight; VT50384-Gal4DBD / +

269 For Mi1 and Tm3 experiments, the flies genotype were as follows :

- 270 1. Mi1>GCaMP6f : w+ ; R19F01-Gal4AD / UAS-GCaMP6f; R71D01-Gal4DBD / UAS-GCaMP6f
- 271 2. Mi1>Arclight : w+ ; R19F01-Gal4AD / UAS-Arclight; R71D01-Gal4DBD / +
- 272 3. Tm3>GCaMP6f : w+ ; R13E12-Gal4AD / UAS-GCaMP6f; R59C10-Gal4DBD / UAS-GCaMP6f
- 273 4. Tm3>Arclight : w+ ; R13E12-Gal4AD / UAS-Arclight; R59C10-Gal4DBD / +

274 Calcium & voltage imaging

275 For imaging experiments, fly surgeries were performed as previously described (Maisak *et al.* 2013).
276 Briefly, flies were anaesthetized with CO₂ or on ice, fixed with their backs, legs and wings to a

277 Plexiglas holder with back of the head exposed to a recording chamber filled with fly external
278 solution. The cuticula at the back of the head on one side of the brain was cut away with a fine
279 hypodermic needle and removed together with air sacks covering the underlying optic lobe. The
280 neuronal activity was then measured from the optic lobe with a custom-built 2-photon microscope
281 as previously described (Maisak *et al.* 2013). Images were acquired at 64 x 64 pixels resolution and
282 frame rate 13 Hz with the Scanimate software in Matlab (Pologruto *et al.* 2003).

283 **Visual stimulation**

284 For the study of visual responses of T4c cells, visual stimuli were presented on a custom-built
285 projector-based arena as described in (Arenz *et al.* 2017). In brief : Two micro-projectors (TI DLP
286 Lightcrafter 3000) were used to project stimuli onto the back of an opaque cylindrical screen
287 covering 180° in azimuth and 105° in elevation of the fly's visual field. To increase the refresh rate
288 from 60 Hz to 180 Hz (at 8 bit color depth), projectors were programmed to use only green LED
289 (OSRAM L CG H9RN) which emits light between 500 nm to 600 nm wavelength. Two long-pass filters
290 (Thorlabs FEL0550 and FGL550) were placed in front of each projector to restrict the stimulus light
291 to wavelengths above 550 nm. This prevents overlap between fluorescence signal and arena light
292 spectra. To allow only fluorescence emission spectrum to be detected, a band-pass filter (Brightline
293 520/35) was placed in-front of the photomultiplier. Stimuli were rendered using custom written
294 software in Python 2.7.

295 **Stimuli**

296 Stimuli were presented with 3-5 repetitions per experiment in a randomized fashion. To measure
297 the directional and speed tuning, square-wave gratings with a spatial wavelength of 30° spanning
298 the full extent of the stimulus arena were used. The gratings were moved in 12 different directions
299 from 0° – 360° at 4 different speeds (15°s⁻¹, 30°s⁻¹, 60°s⁻¹, 120°s⁻¹). Similarly, to measure direction
300 and contrast tuning, square-wave gratings with a spatial wavelength of 30° spanning the full extent
301 of the stimulus arena were used. The gratings moved at a speed of 30°s⁻¹ in 12 different directions
302 at 4 different contrasts (10%, 20%, 50%, 100%). Edge responses were measured using ON edge i.e.
303 bright edge moving on a dark background with full contrast. The ON edge moved in preferred
304 direction (upward) or null direction (downward) at 4 different speeds (15°s⁻¹, 30°s⁻¹, 60°s⁻¹, 120°s⁻¹).

305 **Data analysis**

306 Data analysis was performed using custom-written routines in Matlab and Python 2.7, 3.7. Images
307 were automatically registered using horizontal and vertical translations to correct for the movement
308 of brain. Fluorescence changes ($\Delta F/F$) were then calculated using a standard baseline algorithm
309 (Jia *et al.* 2011). Regions of interest (ROIs) were drawn on the average raw image manually by hand
310 in the medulla layer M10 for signals from T4 dendrites. Averaging the fluorescence change over this
311 ROI in space resulted in a ($\Delta F/F$) time course. Voltage imaging with Arclight and calcium imaging
312 with GCaMP6f and GCaMP8f were performed and analysed using same settings.

313 The direction selectivity was evaluated using a direction selectivity index (DSI) calculated as the
314 difference of the peak responses to preferred and null direction, divided by the sum of the peak
315 responses:

$$DSI = \frac{PD_{peak} - ND_{peak}}{PD_{peak} + ND_{peak}} \quad (1)$$

316 In the above measurement, only the difference in response between the two opposing directions of
317 motion is quantified. To take into account all 12 directions of motion, we calculated the directional
318 tuning index:

$$L_{dir} = \left| \frac{\sum_{\varphi} \vec{v}(\varphi)}{\sum_{\varphi} |\vec{v}(\varphi)|} \right| \quad (2)$$

319 where $\vec{v}(\varphi)$ is a vector proportionally scaled with the mean response and points in the direction
320 corresponding to the direction of motion given by the rotation angle φ of the stimulus (Mazurek

321 *et al.* 2014).

322 Model simulations

323 Custom-written Python3.7 scripts were used to simulate the models (figure 4). To calculate the
324 optimal parameter values, we first defined an error function. For each stimulus condition (s_i), the
325 error was calculated as:

$$326 \quad error(s_i) = \sum_{t=0}^{t=N} (model(s_i, t) - data(s_i, t))^2 \quad (3)$$

326 The model took as input Arclight data across all 112 different stimuli conditions. Next, we summed
327 the error for all stimuli conditions:

$$328 \quad total\ error = \sum_{i=1}^{i=112} error(s_i) \quad (4)$$

328 The model parameters were initialized with random values within the defined parameter bounds.
329 Python SciPy optimize minimize function then used the L-BFGS-B (Limited Broyden Fletcher Goldfarb
330 Shanno) algorithm to find the parameter values corresponding to the minimum total error. A total
331 of 300 runs were performed, and the parameter values that corresponded to the run with the
332 lowest error were used to produce the final output signals. To compare the model performances,
333 we calculated the model error as:

$$334 \quad model\ error [\% of data power] = \frac{total\ error}{\sum_{i=1}^{i=112} (data(s_i))^2} * 100 \quad (5)$$

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3

DISCUSSION

In manuscript 2.1, we found that the elementary motion-sensitive neurons T₄ and T₅ in *Drosophila* regardless of their directional tuning and contrast preferences for ON or OFF stimuli implemented a preferred direction (PD) enhancement on the preferred side of their receptive field and a null direction (ND) suppression on the null side. This combination of PD enhancement and ND suppression increases the direction selectivity in T₄ and T₅ cells, the first cells where the direction selectivity emerges. In T₄ cells, whole-cell patch clamp recordings revealed that while the preferred direction stimuli led to large membrane depolarizations, the null direction stimuli also evoked small, but significant responses (Groschner *et al.* 2022). The calcium recordings, however, had a large response to stimuli moving in the preferred direction and almost no response to stimuli moving in the null direction (Fisher *et al.* 2015; Maisak *et al.* 2013). In manuscript 2.2, we showed that the voltage-to-calcium transformation in addition to the synaptic mechanisms on the dendrites of the T₄ neurons enhances the direction selectivity in the T₄ neurons: the calcium signals in T₄ cells had a significantly higher direction selectivity compared to the voltage signals, thus making T₄ output signals more direction-selective.

3.1 CELLULAR IMPLEMENTATION OF PD ENHANCEMENT AND ND SUPPRESSION IN THE T₄ AND T₅ NEURONS

How are PD enhancement and ND suppression implemented in T₄ and T₅ cells? Through decades of research, a complete connectome of motion vision circuitry has been assembled (Shinomiya *et al.* 2019, 2014; Takemura *et al.* 2008, 2017). A great deal of detail has been provided on the functional response properties (Arenz *et al.* 2017; Drews *et al.* 2020; Serbe *et al.* 2016; Strother *et al.* 2017) and transmitter phenotypes (Davis *et al.* 2020; Pankova & Borst 2017; Richter *et al.* 2018; Takemura *et al.* 2017) of the inputs to T₄/T₅ cells. Furthermore, it is also possible to visualize the endogenous expression of receptor proteins in T₄ and T₅ neurons. The subcellular distributions of the Acetylcholine (ACh) receptor subunit D α 7, the GABA receptor subunit Rdl, and the glutamate-gated chloride channel GluCl α have been studied in T₄ and T₅ cells (Fendl *et al.* 2020).

In the T₄ dendrites, GluCl α receptors and the glutamatergic input Mi9 synapses are located on the distal part (the preferred side) of the dendrite (figure 7c, e). The ACh D α 7 receptors and the cholinergic inputs Mi1 and Tm3 synapses are located at the center of the T₄ dendrites. The GABA Rdl receptors and the GABAergic inputs Mi4, C₃, and CT1 synapses are located on the proximal (the null side) of the T₄ dendrites. The presence of the glutamate-gated chloride channels makes glutamate input from Mi9

inhibitory. Mi9 has an OFF center receptive field. Mi9 maintains an active state in the dark that abruptly ends when light stimulates its receptive field. The preferred direction enhancement or the multiplication-like nonlinearity arises from the coincidence of cholinergic excitation and release from glutamatergic inhibition (Groschner *et al.* 2022). The PD enhancement on the T₄ dendrites is thus achieved by multiplying the release from Mi9 inhibitory glutamate input on the distal arm with an excitatory cholinergic input of Mi1 in the center. This 'multiplicative disinhibition' represents the opposite of divisive inhibition. GABAergic inhibitory inputs on the null sides Mi₄, C₃, and CT₁ provide the divisive inhibition or the null direction suppression. Why are there multiple cells providing input on the null side and the specific contributions of each of these cells on the null side remain to be investigated.

There is still uncertainty regarding how PD enhancement and ND suppression are achieved in T₅ dendrites. In the T₅ dendrites, the ACh D α ₇ receptors and the cholinergic Tm₁, Tm₂, and Tm₄ inputs synapses are located in the center of the dendrites (figure 7d, f). The GABA Rdl receptors and the GABAergic input CT₁ synapses are located on the proximal part (the null side) of the dendrites. However, the GluCl α receptors are absent in the case of T₅ cells. The T₅ dendrites on the distal part (the preferred side) receive input from cholinergic neuron Tm₉ and also express ACh receptors. Hence, the implementation of PD enhancement in the case of T₅ cells is most likely different from the implementation in the T₄ cells. In addition, while 3 neurons (Mi₄, C₃, and CT₁) provide GABAergic input to the null side of T₄ dendrites, the T₅ neurons receive inhibitory input on the null side only from CT₁ neurons. As a result of these differences, the implementation of ND suppression in T₅ cells may also be different in comparison to the T₄ cells. These are some of the questions that need to be investigated further.

One possible approach to discern the individual contribution of the GABAergic input neurons Mi₄, C₃, and CT₁ is to silence one of these neurons, while simultaneously recording neural activity from the T₄ neurons. However, such blocking experiments come together with their own challenges. One such challenge is that the current blocking techniques are not connection-specific. In other words, silencing one type of cell will silence synaptic transmission to all of its postsynaptic targets. Especially in the fly visual system, where almost all of the medulla neurons are highly interconnected, this can be problematic (Takemura *et al.* 2017). For example, Mi₄ and Mi₉ cells have very strong reciprocal connections. Hence, silencing Mi₄ neurons would also affect the neural activity in the Mi₉ neurons. The effect thus observed downstream in T₄ neurons is difficult to be attributed solely to Mi₄. It is therefore possible to cause second-order effects if one cell type is blocked, resulting in a profound effect on the circuit overall. The development of connection-specific blockers could provide a solution to such confounding effects. Secondly, it is difficult to determine the effectiveness of a block. It is hard to say whether blocked cells have no effect or if the block was ineffective when there is no effect or negative results. Thirdly, does the stimulus being used test the distinct contribution of the cell to the response? If no effect or phenotype is observed and the block is effective, using another stimulus may result in an effect.

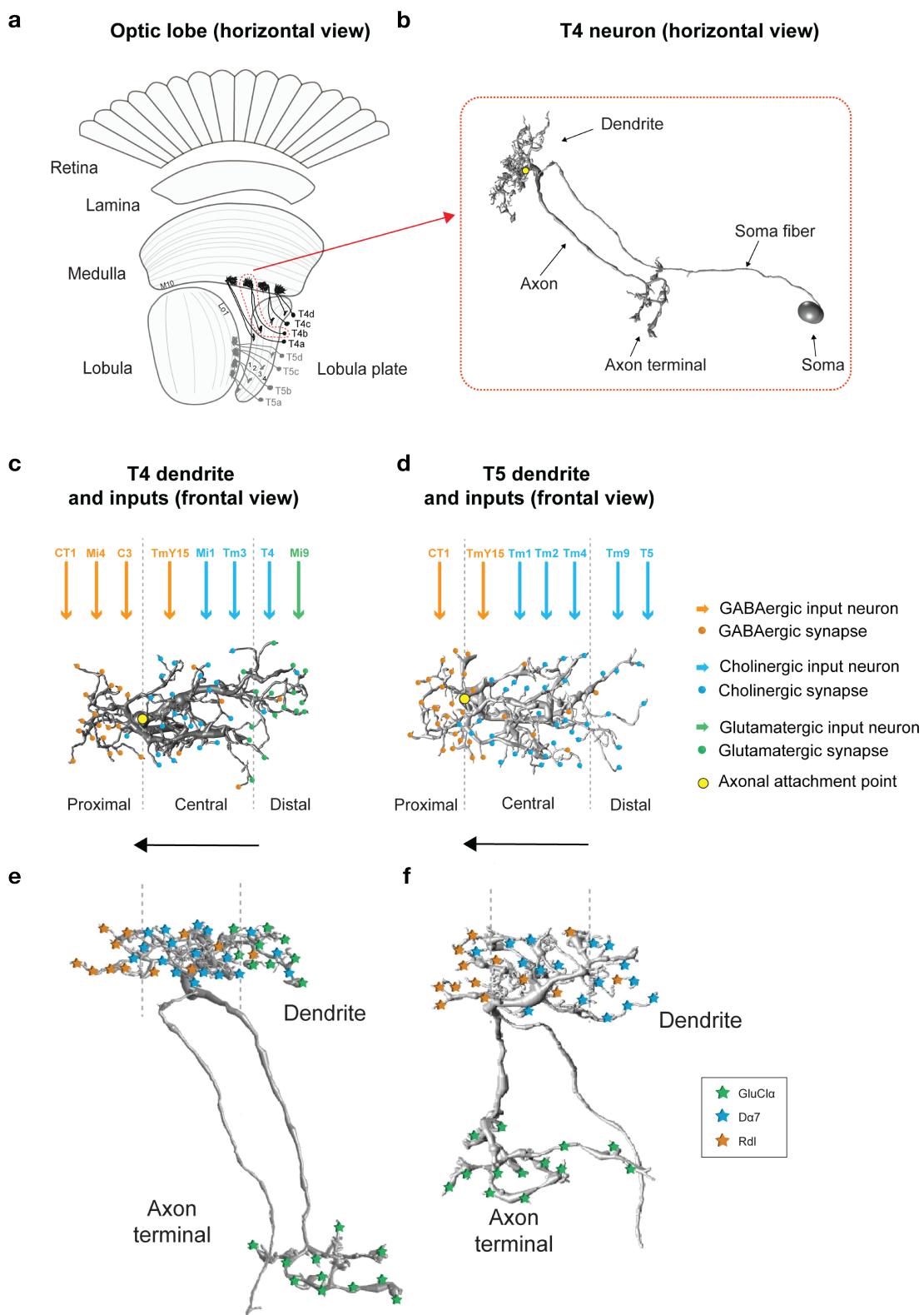


Figure 7: Distribution of the presynaptic partners, input synapses and receptors on the T4 and T5 dendrites: (a) A horizontal view of the optic lobe shows the retina, lamina, medulla, lobula, and lobula plate. Dendrites of T4 (darker gray) are found in layer 10 of the medulla, and those of T5 (lighter gray) are found in layer 1 of the lobula. (b) An EM-reconstructed T4 neuron showing the location of the dendrite, axon, and cell body. (c) An illustration of an individual T4 dendrite and its distribution of input synapses (frontal view). In this illustration, the dendrite points to the right, against its preferred right-to-left orientation (indicated by an arrow). (d) Same as in c, but for T5 cells. (e) Anatomic distribution of glutamate-gated chloride channels GluCl α , acetylcholine receptors D α 7, and GABA receptors Rdl on T4 dendrites. (f) Same as in e, but for T5 cells. (Used and modified with permission from (Fendl *et al.* 2020))

3.2 THE MECHANISM FOR THE TEMPORAL DELAY IN T4 AND T5 INPUT NEURONS

Most major cells presynaptic to T4 and T5 have been characterized in detail in terms of their temporal response properties. Generally, inputs can be classified into two classes with respect to their temporal properties: transient band-pass filters and slow sustained low-pass filters (Arenz *et al.* 2017; Behnia *et al.* 2014; Serbe *et al.* 2016). In the ON pathway, while Mi9 and Mi4 show characteristics of a low-pass filter, Mi1 and Tm3 show characteristics of a band-pass filter. In the OFF pathway, Tm1, Tm2, and Tm4 revealed a band-pass characteristic, and Tm9 showed low-pass filter characteristics. The mechanisms underlying their diverse temporal properties, however, remain unknown. It is possible to implement a delay either intracellularly or synaptically. Simple mechanisms such as passive dendritic filtering can be used by cells. A neurite's length, diameter, and membrane resistance, for example, determine its conduction velocity. The time constant of a neuron, which describes its passive low-pass filtering properties, is linearly related to its input resistance. Therefore, slower medulla neurons may simply have higher resistances.

In addition to the cellular passive properties, voltage-gated ion channels can also delay electrical signals or render them more transient (Destexhe 1999). As an example, depolarizing synaptic inputs activate A-type potassium channels, but they quickly deactivate. The brief increase in potassium conductance prevents the membrane from reaching a threshold, causing a delay (Groschner *et al.* 2018). In the lateral geniculate nucleus of guinea pigs, a transient A-type potassium current has been hypothesized to explain the delayed visual response (McCormick 1991).

Synaptic transmission can have an important impact on a circuit's temporal dynamics in addition to cell-intrinsic mechanisms. The process of neurotransmitter release, diffusion, and binding to a receptor already imposes a delay of 2-3 milliseconds during chemical synaptic transmission. Additionally, postsynaptic receptor properties contribute to temporal filtering. Direct current can flow through ionotropic receptors, but metabotropic receptors initiate a second messenger cascade, which activates ionic conductances after a delay typically of tens to hundreds of milliseconds. All of these mechanisms are plausible since T4 and T5 neurons express a wide array of ionotropic and metabotropic neurotransmitter receptors and voltage-gated ion channels.

Mi9 and Tm9 with low-pass filter characteristics receive their major input from lamina cell L3, which exhibits slow temporal characteristics (Silies *et al.* 2013). L1 provides a major input to the faster cells Mi1 and Tm3 (Takemura *et al.* 2017). There are already fast band-pass characteristics in L1 (Clark *et al.* 2011; Drews *et al.* 2020; Reiff *et al.* 2010). Therefore, it is possible that medulla neurons inherit the temporal properties of their lamina inputs. Thus, the delay mechanism associated with ON and OFF motion detectors may be implemented between photoreceptors and lamina cells at the first synapse. In this scenario, motion blindness should be the result of flies with dysfunctional L3 cells. However, blocking synaptic transmission from L3 does not

significantly affect fly optomotor activity (Bahl *et al.* 2015; Silies *et al.* 2013; Tuthill *et al.* 2013). The temporal filtering properties of a given neuron in the fly medulla are likely determined by several biophysical mechanisms.

3.3 CIRCUITS DOWNSTREAM OF T4 AND T5 CELLS

How do downstream circuits use the motion direction information computed by the T4/T5 neurons in the optic lobe to guide fly behavior? The crucial roles of T4 and T5 cells in visually guided behaviors have been revealed through several studies inhibiting synaptic transmission from these cells. For fly optomotor behavior, T4 and T5 cells are required. Blocking the synaptic output of these cells led to motion blindness in flies (Bahl *et al.* 2013). Figure-ground discrimination (Fenk *et al.* 2014), avoidance of expanding stimuli, as well as landing responses (Schilling & Borst 2015) involve T4 and T5 cells. In what ways are direction-selective signals passed from T4/T5 cells to the central brain and to the motor areas? Lobula plate tangential cells (LPTCs) provide the most direct link between motion-sensitive T4/T5 neurons and motor circuits.

In the lobula plate, T4 and T5 neurons provide excitatory inputs onto LPTCs and lobula plate intrinsic neurons (Lpi). In addition to abolishing the depolarization of LPTCs when stimulated along their preferred direction, blocking the output of T4 and T5 cells also affects the hyperpolarization when stimulated along their null direction (Schnell *et al.* 2012). Additionally, LPTCs respond with a fast excitation, followed by inhibition when T4 and T5 are optically activated (Mauss *et al.* 2015). These results suggest that T4 and T5 cells provide LPTCs with both direct stimulation and indirect inhibition from adjacent lobula plate layers. Since LPi neurons bi-stratify in layer-specific ways, dendrites from one subtype reside exclusively in one layer, and axons in the neighboring layer, LPi are perfect candidates for this task (figure 8). T4/T5 cells in one layer provide feedforward glutamatergic inhibitory input via LPi via glutamate-gated chloride channels GluCl α to LPTC dendrites in the adjacent layer. The LPis are thus essential for increasing flow-field selectivity since it subtracts motion-opponent signals (Mauss *et al.* 2015).

Dendritic trees of the LPTCs span large areas of the lobula plate, sometimes even covering the entire layer. They, therefore, cover a large portion of the visual field with their receptive fields. There are two major groups of *Drosophila*'s LPTCs, based on their response characteristics: The horizontal system (HS) and the vertical system (VS) cells. When presented with motion in a preferred direction (PD), LPTCs depolarize, while when presented with motion in a null direction (ND), they hyperpolarize. The T4 and T5 excitatory inputs from one lobula plate layer are integrated and subtracted from the adjacent layer. Dendrite location determines the preferred direction. As all preferred directions are combined onto the dendritic tree, so-called flow fields emerge that describe the optimal pattern of motion resulting from ego-motion. Thus, LPTCs are specialized filters that detect optical flow patterns resulting from self-motion (Krapp & Hengstenberg 1996). During the tethered flight, unilateral activation of HS cells induces yaw-turning responses

of the head and wings (Haikala *et al.* 2013). VS and HS synaptic output blocks impair the head optomotor response severely but have a weaker impact on wing steering (Kim *et al.* 2017). Accordingly, *Drosophila* optomotor responses may be controlled by LPTCs.

3.4 THE DIRECTION SELECTIVITY OF T4 OUTPUT SIGNALS

We showed that the voltage to calcium transformation in T4c neurons enhances their direction selectivity. The calcium signals in T4c cells had a significantly higher direction selectivity and tuning than the membrane voltage across different stimuli conditions. As calcium is required for neurotransmitter release, this is expected to increase the direction selectivity of T4 cells' output signals.

In order to determine the direction selectivity of T4 cells' output signals, one can record the neurons that are postsynaptic to T4 cells. As discussed, the horizontal system (HS) cells with dendrites in layer 1 depolarize during front-to-back motion and hyperpolarize during back-to-front motion, while HS cells in layer 2 display the opposite tuning, while vertical system (VS) cells with dendrites mainly in layer 4 depolarize primarily during downward motion and hyperpolarize during upward motion (Hopp *et al.* 2014; Schnell *et al.* 2010; Scott *et al.* 2002). Monosynaptic, excitatory, and cholinergic connections exist between T4/T5 and tangential cells (Mauss *et al.* 2014). T4/T5 cells are responsible for depolarizing tangential cells during preferred direction motion. Tangential cells hyperpolarise in the null direction in response to inhibition from Lpi neurons, which receive excitatory input from T4/T5 neurons in the opposite layer. Upon silencing the LPi3-4 neurons' synaptic output via tetanus toxin, the VS neurons' depolarization response in the preferred direction did not change, but the null direction response was absent (figure 9) (Mauss *et al.* 2015). This suggests T4/T5 do not release any transmitter in response to the null direction motion, which matches our findings for the calcium responses. Thus, the voltage to calcium transformation increases direction selectivity in T4/T5 cells and this enhances direction selectivity in the downstream neurons.

3.5 NEURONAL CALCIUM SIGNALING

In every eukaryotic cell, calcium (Ca^{2+}) regulates the most important activities. Neurons depend on it for the transmission of the depolarizing signal and for synaptic activity. A variety of neuronal processes including long-term potentiation of synaptic transmission or depression of synaptic transmission are controlled by Ca^{2+} signals. As a result, neurons have developed extensive and intricate calcium signaling pathways (Brini *et al.* 2014). Plasma membrane receptors and voltage-dependent ion channels facilitate calcium influx into neurons. Calcium is also released from intracellular stores, such as the endoplasmic reticulum, by intracellular channels. As Ca^{2+} is essential

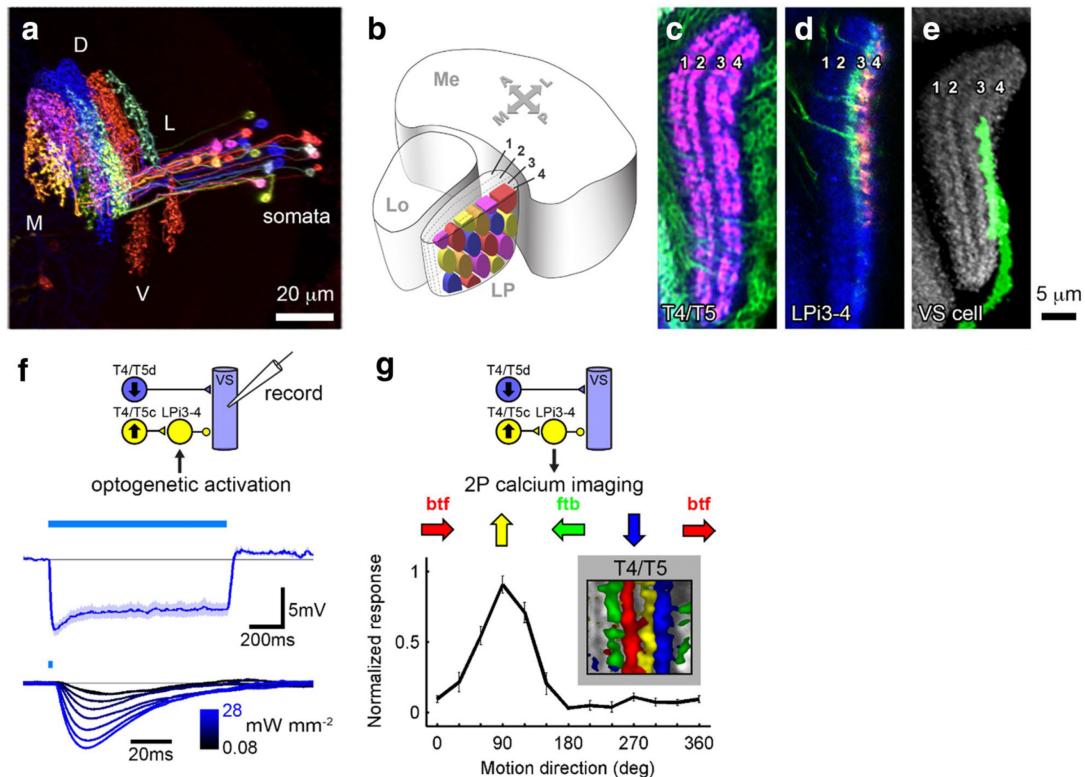


Figure 8: Lobula plate intrinsic neurons (LPis): (a) Multi-color flip-out showing several LPi neurons in the lobula plate. D = dorsal, V = ventral, L = lateral, M = medial. (b) Dendritic fields of adjacent LPi neurons shown schematically. (c) The horizontal cross-section of the lobula plate shows GFP-expressing T4/T5 cells (green) and synaptotagmin-HA (red). (d) LPi neurons expressing GFP (green) and presynaptic synaptotagmin-HA (sytHA, red). Presynaptic specializations are restricted to layer 4 only, even though neurons ramify in layers 3 and 4. (e) GFP staining of a VS cell dendrite in layer 4 of the lobula plate. (f) Patch-clamp recordings from VS cells and optogenetic stimulation of LPi cells to study the synaptic connection between LPi and VS cells. A sustained hyperpolarizing potential is evoked by 1 s light stimulation of LPi neurons (upper recording trace). Light pulses (2 ms) are delivered in different strengths (bottom traces). (g) Direction-selectivity of Lpi cells. Visual activity in LPi3-4 neurons measured by calcium imaging in response to gratings moving in different directions. (Used with permission from (Borst *et al.* 2020b))

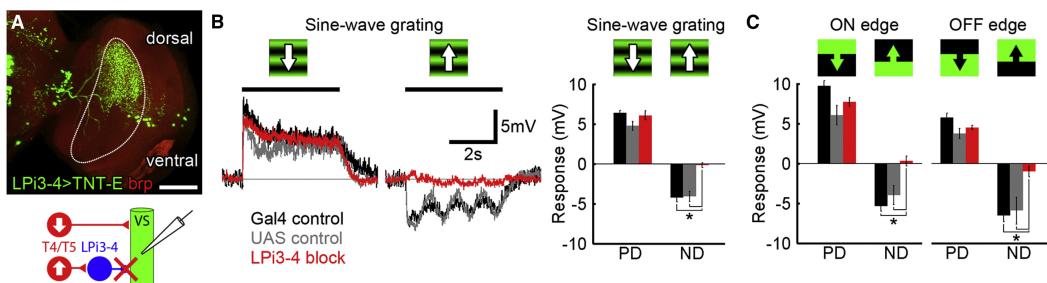


Figure 9: Tangential cells receive null direction responses from LPi neurons: (a) In LPi3-4 neurons, the tetanus toxin light chain suppresses synaptic release. The schematic below illustrates the experimental approach used to measure whole-cell voltages from VS cells in order to investigate LPi3-4 cell function. (b) Control flies respond to sine-wave gratings moving down (preferred direction [PD]) or up (null direction [ND]) by depolarizing or hyperpolarizing respectively. Hyperpolarizing responses to ND motion are selectively abolished in LPi3-4 block flies. (c) In LPi3-4 block flies, VS cell responses to moving ON and OFF edges are similarly affected with ND responses. (Used with permission from (Mauss *et al.* 2015))

for cellular signaling, its background concentration within cells must be low enough to allow it to be significantly altered without consuming excessive energy.

There are three major groups of plasma membrane Ca^{2+} channels based on their mechanisms of opening: voltage-gated Ca^{2+} channels, receptor-operated Ca^{2+} channels (ROC), and store-operated Ca^{2+} entry channels (SOC), which are activated when the cellular Ca^{2+} stores are empty. There are five distinct subunits ($\alpha 1$, $\alpha 2$, β , γ , δ) encoded by different genes in the voltage-gated Ca^{2+} channels. They are divided into three subfamilies, namely, Cav1, Cav2, and Cav3 depending on the type of $\alpha 1$ pore-forming subunit. According to the physiological and pharmacological properties of the type of current they carry, they can further be classified into six classes L, N, P, Q, R, and T. The $\alpha 1$ subunit consists of four repeat domains (I-IV), each with six transmembrane segments (S1-S6). Within the pore-containing subunit, the S4 segments contain some positively charged residues that act as voltage sensors. The associated $\alpha 2$, β , γ , and δ subunits have supplementary functions, including the control of channel expression and the modulation of current kinetics (Catterall 2000; Hofmann *et al.* 1999). In skeletal and cardiac muscle, the Cav1 subfamily mediates L-type currents and initiates excitation-contraction coupling. As a result of its activity in neuronal cells, Ca^{2+} transients are generated in dendrites and cell bodies, which in turn regulate processes like secretion and gene expression. Synaptic transmission, neurotransmitter release, and dendritic Ca^{2+} transients are mainly initiated by Cav2 channels, which generate N-, P/Q-, and R-type currents. Cav3 subfamily members are responsible for the T-type current, which is important for pacemaking in cardiac myocytes and repetitive action potential firing in the thalamus (Catterall 2011).

Extracellular ligands, such as neurotransmitters, activate receptor-operated Ca^{2+} channels (ROC). In mammals, L-Glutamate stimulates two classes of receptors: ionotropic receptors (iGluRs) and metabotropic receptors (mGluRs). The two principal types of ionotropic glutamate receptors are N-methyl-

d-aspartate sensitive receptors (NMDARs) and Alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid-sensitive receptors (AMPARs). In the mammalian central nervous system, AMPARs transmit fast excitatory synaptic signals and are permeable to Na^+ and K^+ , and may be permeable to Ca^{2+} ions. NMDARs are permeable to both Na^+ and Ca^{2+} . Compared to AMPARs, NMDARs respond more slowly to glutamate. Not only glutamate is required for their activation (ligand-gating), but also membrane depolarization (voltage dependence) to remove Mg^{2+} that normally blocks them. The coincidence detection process that opens NMDAR channels is critical in learning and memory (Miyashita *et al.* 2012).

Store-operated Ca^{2+} entry channels (SOC) are activated when Ca^{2+} is released from the endoplasmic reticulum. Originally discovered in non-excitable cells, they are now being discovered in skeletal muscle and neurons as well. It was originally proposed that store-operated Ca^{2+} entry ensured the replenishment of intracellular stores after Ca^{2+} was released (Putney Jr 1986). There is evidence that Ca^{2+} influx through this pathway may directly signal targets located close to sites of Ca^{2+} entry, thus initiating specific signaling pathways (Feske 2011). A TRP channel is a type of channel that can either regulate intracellular Ca^{2+} concentration directly by acting as a Ca^{2+} entry pathway or indirectly by triggering voltage-dependent ion channel activation when cells are depolarized.

Considering the special importance of calcium signaling in neuronal function, the voltage to calcium transformation we studied in our second manuscript with a focus on direction selectivity may have a broader impact on other neuronal processes and should be investigated further.

3.6 DIFFERENTIAL EXPRESSION OF VOLTAGE-GATED CALCIUM CHANNELS

In manuscript 2.2, we built a model to capture the voltage to calcium transformation in T4c, Mi1, and Tm3 cells. A simple model with a single low-pass filter was able to reproduce the calcium responses in non-direction-selective Mi1 and Tm3 cells, whereas a more complex model combining the output of two low-pass filters via a multiplication was required to reproduce T4c calcium responses. The direction selectivity for the simple model signals for T4c was lower compared to the multiplicative model. This suggests that voltage-calcium transformation in Mi1 and Tm3 cells is different from these in T4c cells.

Differential expression of voltage-gated calcium channels in these cells could explain the different voltage to calcium transformation. The voltage-gated calcium channels mediate depolarization-induced calcium influx that drives the release of neurotransmitters. The $\alpha 1$ -subunit of the voltage-gated calcium channels form the ion-conducting pore, which makes it distinct from other calcium channels. Three families of genes encode $\alpha 1$ subunits. *Drosophila* genome has one $\alpha 1$ subunit gene in each family: $\alpha 1D$ (Ca_v1), cac (Ca_v2), and $\alpha 1T$ (Ca_v3) (King 2007; Littleton & Ganetzky 2000). In *Drosophila* antennal lobe projection neurons, cac (Ca_v2) type and $\alpha 1T$ (Ca_v3)

type voltage-gated calcium channels are involved in sustained and transient calcium currents, respectively (Gu *et al.* 2009; Iniguez *et al.* 2013). According to a RNA-sequencing study (Davis *et al.* 2020), $\alpha 1T$ (Ca_v3) mRNA have higher expression in Mi1 (2050.16 Transcripts per Million (TPM)) compared to T4 (686.68 TPM) and Tm3 (336.45 TPM). While cac (Ca_v2) mRNA have higher expression in T4 (1298.53 TPM) compared to Mi1 (986.25 TPM) and Tm3 (817.61 TPM). Different expressions of voltage-gated calcium channels could cause the different voltage to calcium transformations in non-direction selective and direction-selective cells.

3.7 COMPARISON BETWEEN THE ON AND OFF PATHWAYS IN THE FLY OPTIC LOBE AND THE MOUSE RETINA

Among the most striking similarities between the retina and the fly optic lobe is the early splitting of pathways into ON and OFF channels (figure 10). This allows for more efficient encoding of visual stimuli (Gjorgjieva *et al.* 2014). In the vertebrate retina, this splitting takes place right at the photoreceptor-bipolar synapse, whereas in the fly, it occurs one synapse later.

The photoreceptors of the mouse retina hyperpolarize in response to light, while in darkness they release glutamate onto their postsynaptic partners, the bipolar cells. The split between the ON and OFF pathways occurs at the synaptic level between photoreceptors and bipolar cells, resulting in the ON- and OFF-responsive bipolar cells. In the ON bipolar cells, the metabotropic inhibitory glutamate receptor mGluR6 causes a sign inversion and the ON channel is formed (Masu *et al.* 1995). The OFF bipolar cells, however, express ionotropic AMPA receptors that depolarize when glutamate binds (Euler *et al.* 2014). As in the fly optic lobe, there are fast and slow bipolar cells, similar to the medulla and transmedulla neurons.

In the fly, the split into the ON and OFF pathways occurs at the level of lamina cells. Vertebrates don't seem to have any equivalent to the lamina. The *Drosophila* photoreceptors depolarize under light and release histamine, which in turn inhibits lamina neurons via histamine-gated chloride channels (Hardie 1989). The cholinergic lamina neurons L2-L5 transmit photoreceptor signals to the medulla and transmedulla neurons. In the ON channel, L1 is the main input, while in the OFF channel, L2 is the main input (Joesch *et al.* 2010). The glutamatergic L1 neurons inhibit postsynaptic Mi1 and Tm3 neurons via the glutamate-gated chloride channel GluCl α , implementing a sign inversion and creating an ON channel. Thus, the photoreceptors depolarize in response to the light, inhibiting L1 neurons, thereby disinhibiting Mi1 and Tm3 neurons, creating the ON-responses. Both GluCl α and Rdl receptors are involved in this multi-synaptic sign inversion in the ON pathway (Molina-Obando *et al.* 2019). Both mouse and fly visual systems exhibit sign inversion in the ON pathway as a result of glutamatergic, inhibitory signaling. Fly uses the GluCl α channel, which is unique to the invertebrates, instead of the mGluR6 receptor, which causes inhibition in the mouse retina.

Direction-selective T₄/T₅ cells in the flies are comparable to the starburst amacrine cells (SACs) in mammals and the lobula plate tangential cells are comparable to direction-selective ganglion cells (figure 10). The direction-selective retinal ganglion cells (DSGCs) were the first direction-selective cells to be described in the mammalian retina (H. B. Barlow & Hill 1963). Their four subtypes respond to movement in one of the four cardinal directions, similar to the elementary motion detectors in the fly (T₄/T₅ neurons) (Elstrott *et al.* 2008). Pharmacology and ablation experiments suggest that GABAergic starburst amacrine cells (SACs) are necessary for direction-selective responses in retinal ganglion cells (Yoshida *et al.* 2001). It is interesting to note that starburst amacrine cells are already direction-selective themselves in a centrifugal manner (Euler *et al.* 2002). Dendrites of these cells protrude radially, and they respond preferentially to stimuli from the base to the tip of the cell. The SACs, in turn, enable DSGCs to be direction selective by inhibiting the null side of their dendrites with asymmetric GABAergic inhibition (Briggman *et al.* 2011). How do the SACs become direction-selective? There are several hypotheses and lines of evidence about how bipolar cells providing excitatory glutamatergic input to both cell types shape their direction-selective responses. The starburst amacrine cells which respond to stimuli moving from the soma to the dendritic tips receive input from different types of bipolar cells, including those with fast and slow temporal dynamics (Baden *et al.* 2013). The different types of bipolar cells also exhibit space-time wiring specificity with starburst amacrine cells: slow bipolar cells wire with starburst amacrine cells proximally, whereas fast bipolar cells wire with the starburst amacrine cells distally (Kim *et al.* 2014). Thus, direction selectivity in flies and mammals may arise by similar mechanisms.

3.8 THE FUNCTION OF THE VISUAL CIRCUIT DURING NATURAL BEHAVIOR

In this thesis, experiments were conducted on tethered flies whose movement is severely restricted. How do motion circuits operate during unrestrained behavior? State-dependent modulations are observed in the activity and tuning properties of visual circuits in mice and flies (Maimon 2011). During tethered flight or walking, tangential cells in the fly's lobula plate shift their temporal frequency tuning optimum towards higher frequencies (Chiappe *et al.* 2010; Jung *et al.* 2011; Maimon *et al.* 2010). In the ON motion vision pathway, the medulla neurons modulate their baseline calcium level according to their behavioral state, and octopaminergic neurons are needed to process fast-moving visual stimuli appropriately (Strother *et al.* 2018). Chlordimeform (CDM), an octopamine agonist, shifted the temporal tuning optima for T₄ and T₅ cells and all input elements towards higher frequencies (Arenz *et al.* 2017). Both mammalian and fly visual systems are affected by the general behavior state of the animal early on in the circuit and only a few synapses from photoreceptors.

A free-moving or flying fly experiences not only visual cues but also proprioceptive cues through its antennae and halteres (Mamiya *et al.* 2011;

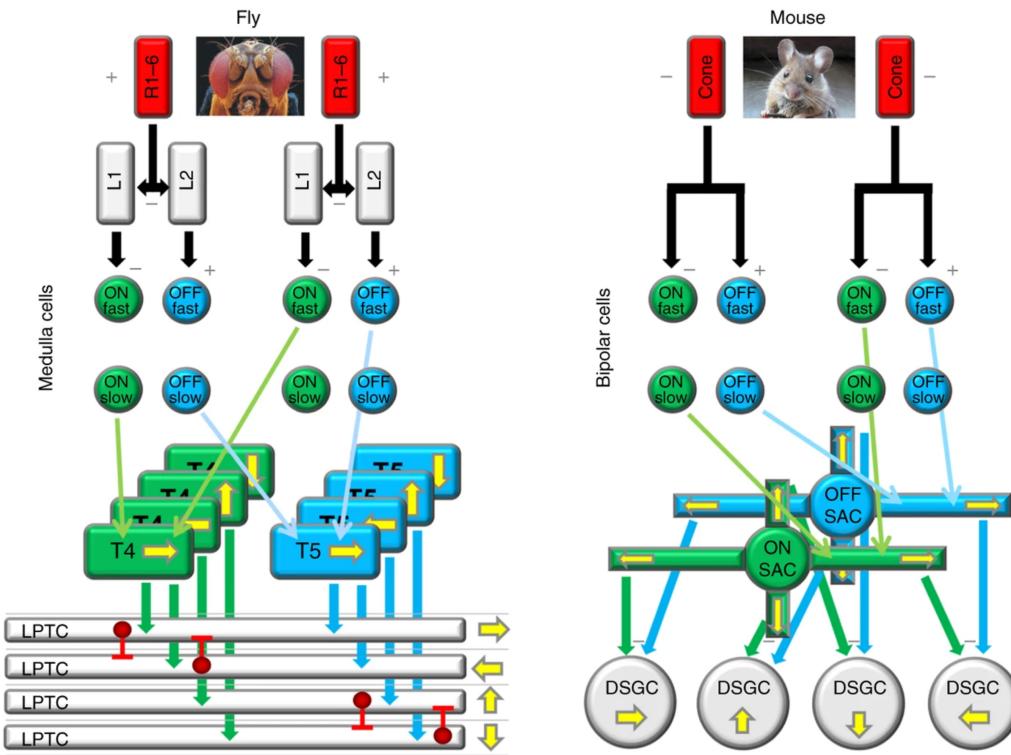


Figure 10: Fly and mouse motion detection circuits: In the fly, the photoreceptors connect via sign-inverting synapses to the lamina monopolar cells L1 and L2, the entry to the ON and OFF pathway, respectively. The mouse retina lacks this additional layer of lamina cells and splits the signal directly between ON and OFF bipolar cells via two types of glutamate receptors. The T4 (ON) and T5 (OFF) neurons in the fly optic lobe and the ON and OFF SACs in the mouse retina are the first stages of direction-selective cells. Lobula plate tangential cells (LPTCs) in the fly and ON-OFF direction-selective ganglion cells (DSGCs) in the mouse integrate direction-selective information from these two pathways. (Used with permission from Borst & Helmstaedter 2015)

Sandeman & Markl 1980). Multiple cues are combined in higher multimodal circuits, usually in a non-linear way (Haag *et al.* 2010; Huston & Krapp 2009). As a result, the activity of visually responsive neurons during tethered flight might be completely different from that during free flight. Flies perform complex maneuvers during free flight, often involving multiple axes of rotation and translation. In a restrained environment, these maneuvers are hard or impossible to repeat. In order to fully understand the function of a visual circuit, it is ideally best to study it in its natural state. In larger animals that can carry head-mounted microscopes, head-stages, or fiber optics, this is easier, but in fruit flies, it is extremely challenging. Over the past years, substantial progress has been made toward achieving this goal. Fruit flies can be tracked online in 2D and 3D with high precision. An optical laser can be used to target the fly for thermogenetic or optogenetic activation of nerve cells using this information (Bath *et al.* 2014; Stowers *et al.* 2014; Straw *et al.* 2011). It is therefore possible to manipulate the activity of a subset of neurons when a fly performs a specific behavioral action or experiences a visual stimulus. Efforts are being made to perform functional imaging in freely walking flies (Grover *et al.* 2016). The combined use of these promising tools can give us a better understanding of how individual nerve cells and visual circuits operate under natural conditions.

3.9 CONCLUSION

In the course of this work, I investigated neural computation in the *Drosophila* motion vision pathway. Together with my co-authors, we showed that both the preferred direction enhancement and null direction suppression are implemented in all four subtypes of T4 and T5 cells. Already at the first stage of direction selectivity computation, this combined strategy ensures a high degree of direction selectivity. Additionally, we showed that the voltage-to-calcium transformations further enhance direction selectivity in the output signals of T4 cells in addition to the synaptic mechanisms at the dendrites. We built a model to transform voltage signals into calcium signals. The model was more complex for the direction-selective T4 cells compared to non-direction selective cells Mi1 and Tm3. Future work will focus on the comparison of voltage-gated calcium channels in these neurons which might lead to the observed differences in the voltage-to-calcium transformations.

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CURRICULUM VITAE

EDUCATION

- 2017–present: PhD student at Graduate School for Systemic Neurosciences (LMU Munich)/Max-Planck-Institute for Neurobiology, Germany; supervised by Prof. Dr. Alexander Borst
- 2011–2016: B.Tech/M.Tech in Biotechnology and Biochemical Engineering at the Indian Institute of Technology (IIT) Kharagpur, India

RESEARCH INTERNSHIPS

- 2015: University of British Columbia, Vancouver, Canada
- 2014: University of Alberta, Edmonton, Canada
- 2013: Indian Institute of Science, Bangalore, India

MAJOR AWARDS/FELLOWSHIPS

- Graduate school of systemic neurosciences (GSN-LMU) fast-track Ph.D. fellowship 2016
- MITACS Globalink fellowship, Canada 2015
- University of Alberta research experience fellowship 2014
- Indian Academy of Sciences summer research fellowship 2013

PERSONAL INFORMATION

- Birthday: 20th April 1992
- Nationality: Indian
- Languages: Hindi (native), Nepali (native), English (fluent), German (beginner)

PUBLICATIONS

- Haag, J., **Mishra, A.**, and Borst, A. “A common directional tuning mechanism of *Drosophila* motion-sensing neurons in the ON and in the OFF pathway” *eLife* 6, e29044 (2017)

- **Mishra, A.**, Borst, A., and Haag, J. "Voltage to calcium transformation enhances direction selectivity in *Drosophila* T₄ neurons" *bioRxiv* (2022)

AFFIDAVIT

I hereby confirm that the dissertation **Neural information processing in the *Drosophila* motion vision pathway** is the result of my own work and that I have only used sources or materials listed and specified in the dissertation.

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München, den
Munich, Date

.....
Unterschrift
Signature

CONTRIBUTIONS

- Juergen Haag, **Abhishek Mishra** and Alexander Borst (2017). A common directional tuning mechanism of *Drosophila* motion-sensing neurons in the ON and in the OFF pathway. *eLife* 6:e29044.
 - Juergen Haag, Conceptualization, Data curation, Software, Investigation, Visualization, Writing, review and editing; Abhishek Mishra, Investigation; Alexander Borst, Conceptualization, Funding acquisition, Writing original draft, Project administration, Writing, review and editing
- **Abhishek Mishra**, Alexander Borst, and Juergen Haag (2022). Voltage to Calcium Transformation Enhances Direction Selectivity in *Drosophila* T₄ neurons. *bioRxiv*.
 - Abhishek Mishra, Conceptualization, Investigation, Data curation, Software, Visualization, Writing original draft, review and editing; Juergen Haag, Conceptualization, Data curation, Software, Investigation, review and editing; Alexander Borst, Conceptualization, Funding acquisition, Project administration, Review and editing

.....
Abhishek Mishra

.....
Juergen Haag
Co-author

.....
Prof. Dr. Alexander Borst
Supervisor