# metadamage(?) formats

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This document describe some of the internal formats used by the metadamage software. These are at the current time.

- .bdamage.gz
- $\bullet$  .lca
- .stat
- $\bullet$  .dfit.txt.gz
- .dfit.stat.txt.gz
- $\bullet$  .rlens.gz

## 1 bdamage format

bdamage files are files that contain counts of mismatchs conditional on strand and cycle (position within read). These are generated with metadamage lca or metadamage getdamage. The first 8 bytes magic number determines which bdamage version. If no magic number is present then version0 is assumed.

#### 1.1 version 0

First version of the bdamage file is a single bgzf compressed file. MAXLENGTH occurs once in the beginning of the file, followed by succesive blocks of data[1-8]. Block[3-5] indicates the actual mismatch counts for the forward which we have MAXLENGTH times. Block[6-8] indicates the actual mismatch counts for the reverse strand which will also occur MAXLENGTH times.

Col	Field	Type	Brief description
0	MAXLENGTH	int	Number of cycles
1	ID	int	Id for mismatch type <sup>1</sup>
2	NREADS	$size_t$	Number of reads used supporting the mismatch matrix
3	1	int[16]	mismatch rate for first cycle from the 5prime
4	i	int[16]	mismatch rate for the <i>i</i> 'th cycle from the 5prime
5	MAXLENGTH	int[16]	mismatch rate for the last cycle from the 5prime
6	1	int[16]	mismatch rate for first cycle from the 3prime
7	i	int[16]	mismatch rate for the <i>i</i> 'th cycle from the 3prime
8	MAXLENGTH	int[16]	mismatch rate for the last cycle from the 3prime

Table 1: Content of bdamage.gz file. Note¹ This is either the taxidID or the referenceID relative to the SAM/BAM header for single species resequencing projects or it is the *taxid* if output has been generated with metadamage lca 3) Order is given by AA,AC,AG,AT,CA,CC,CG,CT,GA,GC,GG,GT,TA,TC,TG,TT, with first base indicatting reference nucleotide and second base indicating observed nucleotide

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### 2 .lca

This section describes the test output generated by a metadamage lca subfunctionality and contains information at the readlevel regarding both taxonomic information and statistics pertaining usefull readinformation.

First line of the file begins with a hashtag followed by the actual command used for generating the file. Last entry of the line is again a hashtag followed by the git commit id which will serve as a primitive versioncontrol.

Each line consists of a number of items seperated by tabspace. First entry contains readID together with other information seperated by colon. After the first entry succesive blocks of the type taxid:name:"taxlevel" from the lca toward the root. The complete specification is seen in table below.

Col	Brief description	
1	readID	readID, this might contains colon
2	seq	The actual sequence
3	length(seq)	The length of the sequence
4	nAlignments	The number of alignments used for inferring the lca
5	gc-content	The GC content for the sequence
6	lca taxid	the taxomic id (integer)
7	lca taxid	the taxonomic name(string)
8	lca "taxlevel"	the taxonomic level
9	taxid	the taxomic id (integer)
10	taxid	the taxonomic name(string)
11	"taxlevel"	the taxonomic level

Table 2: Content of a .lca file. Note that 1) seperate between fields[1-6,8-9] is tab, readID might also contain colon. 2) the quotes around field[7,8] is intentional since taxlevels and names might contain spaces andor colons. 3) Number of tab seperated entries is consistent across different reads, since seperator between block 9-11 is semicolon. Seperator between 6-9,9-11 is colon

### 3 .stat

Very simple tabseperated flatfile

- 1. taxid
- 2. Number of supporting reads
- 3. Mean lengths of supporting reads
- 4. Variance of the lengths of the supporting reads
- 5. Mean gccontent of supporting reads
- 6. Variance of the gccontent of supporting reads
- 7. name of lca in quotes (if relevant, otherwise NA)
- 8. name of taxomic level of lca in quotes (if relevant, otherwise NA)

# 4 .dfit.txt.gz

The output format will be heavily dependent on the provided runmode and supplied parameters. It might contain the per file, the per reference or the per species estimate of damage.

Only the columns described in *showfits*  $\theta$  remains the same across the runmode, with the *showfits* 1 or *showfits* 2 describes the unique columns and order which is appended to the *showfits*  $\theta$  columns.

### Non-boostrapping optimization:

Column name	Brief description	
	-showfits 0	
id	identifier see paragraph for details	
А	Dfit statistic. Damage at position one, taking into account offset	
q	per cycle decrease	
c background substitution rate or noise baseline		
$\phi$	Variance between beta-binomial and binomial model	
Ilh	Likelihood for our MLE	
ncall	Number of optimization calls used for obtaining our MLE	
$\sigma_D$	Z value	
Zfit	significance	
	-show fits 1, i signifies position inferred from .bdamage.gz	
fwdxi	damage estimates forward strand	
fwdConfi	forward strand confidence interval $\pm$	
bwdxi	damage estimates backward strand	
bwdConfi	backward strand confidence interval $\pm$	
	-show fits 2, i signifies position inferred from .bdamage.gz	
fwKi	Number of deamination substitution $(C \rightarrow T)$ observations forward strand	
fwNi	Total number of C for forward strand	
fwdxi	Damage estimates forward strand	
fwfi	Calculated damage frequency $fwKi/fwNi$	
fwdConfi	forward strand confidence interval $\pm$	
bwKi	Number of deamination substitution ( $G \rightarrow A$ for ds, $C \rightarrow T$ for ss) observations forward strand	
bwNi	Total number of G for forward strand (ds) and C (ss)	
bwdxi	Damage estimates backward strand	
bwfi	Calculated damage frequency $fwKi/fwNi$	
bwdConfi	backward strand confidence interval $\pm$	

Table 3: Content of a .dfit.txt.gz file. Note that entry nine to 13 is repeated for each cycle first of the 5 $\acute{a}$ nd then from the 3: With the total number of times repeated is given by 1.1.

#### Boostrapping optimization:

Providing -nbootstrap > 1 numerical optimizations of the binomial distribution will also be conducted using bootstrapping methods

Column name	Brief description		
	-showfits 0		
id	identifier see paragraph for details		
А	Dfit statistic. Damage at position one, taking into account offset		
q	per cycle decrease		
С	background substitution rate or noise baseline		
$\phi$	Variance between beta-binomial and binomial model		
llh	Likelihood for our MLE		
ncall	Number of optimization calls used for obtaining our MLE		
$\sigma_D$	Z value		
Zfit	significance		
$A_b$	Dfit statistic from bootstrap estimate. Damage at position one, taking into account offset		
q_b	per cycle decrease from bootstrap estimate		
c_b	background substitution rate or noise baseline from bootstrap estimate		
$\phi_{ extsf{L}}b$	Variance between beta-binomial and binomial model from bootstrap estimate		
$A_CI_I$	Lower bound of CI for A estimate calculated from all bootstrap values		
$A_CI_h$	Upper bound of CI for A estimate calculated from all bootstrap values		
$q_CI_I$	Lower bound of CI for q estimate calculated from all bootstrap values		
$q_CI_h$	Upper bound of CI for q estimate calculated from all bootstrap values		
c_CI_I	Lower bound of CI for c estimate calculated from all bootstrap values		
c_CI_h	Upper bound of CI for c estimate calculated from all bootstrap values		
$\phi_{ extsf{L}}$ CI $_{ extsf{L}}$	Lower bound of CI for $\phi$ estimate calculated from all bootstrap values		
$\phi_{ extsf{L}}$ Cl_h	Upper bound of CI for $\phi$ estimate calculated from all bootstrap values		
	-showfits 1, $i$ signifies position inferred from $.bdamage.gz$		
fwdxi	damage estimates forward strand		
fwdConfi	forward strand confidence interval $\pm$		
bwdxi	damage estimates backward strand		
bwdConfi	backward strand confidence interval $\pm$		
	-show fits 2, i signifies position inferred from .bdamage.gz		
fwKi	Number of deamination substitution $(C \rightarrow T)$ observations forward strand		
fwNi	Total number of C for forward strand		
fwdxi	Damage estimates forward strand		
fwfi	Calculated damage frequency $fwKi/fwNi$		
fwdConfi	forward strand confidence interval $\pm$		
bwKi	Number of deamination substitution ( $G \rightarrow A$ for ds, $C \rightarrow T$ for ss) observations forward strand		
bwNi	Total number of G for forward strand (ds) and C (ss)		
bwdxi	Damage estimates backward strand		
bwfi	Calculated damage frequency $fwKi/fwNi$		
$\phantom{aaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaa$	backward strand confidence interval $\pm$		

Table 4: Content of a .dfit.txt.gz file. Note that entry nine to 13 is repeated for each cycle first of the 5ánd then from the 3: With the total number of times repeated is given by 1.1.

Depending on which parameters and runmode (local, global or lca) that was supplied to both *get-damage* and *dfit*, the content of the id will be different. If a bamfile is supplied (with –bam) then each line will be the information associated with the different refids in the bam file and the id will be the referenceids from the bam file. The case scenario for this would be either obtaining perchromosome estimates of damage or per reference damage which could be relevant for metagenomic studies. If user are computing the damagesignal in the context of the lca. Then the id column will contain the taxid. If -names has been supplied to the dfit program, then the id column will the taxid: *scientific name*. If -nodes has not been defined the dfit.txt.gz will only contain information for the observed references.

If -nodes has been defined the program will aggregate the summary statitics for the internal nodes.

# 5 .dfit.stat.txt.gz

Depending on runmode and parameters supplied to the program. It will contain the per file, the per reference or the per species estimate of the statistic. This format extends section 3 to include the internal nodes.

### 6 .boot.stat.txt.gz

Providing dfit command with nbootstrap > 1 and doboot 1 the boostrapping values (id,A\_b,q\_b,c\_b, $\phi_-$ b) are stored in separate file

### 7 .rlens.gz

Readlength distribution. Distribution is count of alignments of specific readlengths. Depending on runmode there might be multiple groups as taxid/refs and there will be group specific distributions. Distributions for different groups are split by newlines. First entry on each line is the identifer (chromosomename,taxid). The remaining entries are the number of times we have observed an alignment of length (columnnumber). Notice that the first 30 column of counts is likely to be zero since reads shorter than 30basepairs are normally discarded.